



# **Review Recent Trends in Chemical Sensors for Detecting Toxic Materials**

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Abstract: Industrial development has led to the widespread production of toxic materials, including carcinogenic, mutagenic, and toxic chemicals. Even with strict management and control measures, such materials still pose threats to human health. Therefore, convenient chemical sensors are required for toxic chemical monitoring, such as optical, electrochemical, nanomaterial-based, and biological-system-based sensors. Many existing and new chemical sensors have been developed, as well as new methods based on novel technologies for detecting toxic materials. The emergence of material sciences and advanced technologies for fabrication and signal-transducing processes has led to substantial improvements in the sensing elements for target recognition and signal-transducing elements for reporting interactions between targets and sensing elements. Many excellent reviews have effectively summarized the general principles and applications of different types of chemical sensors. Therefore, this review focuses on chemical sensor advancements in terms of the sensing and signal-transducing elements, as well as more recent achievements in chemical sensors for toxic materials.

**Keywords:** chemical optical sensor; electrochemical sensor; nanomaterial; biosensor; transcription factor; toxic material



**Citation:** Kim, Y.; Jeon, Y.; Na, M.; Hwang, S.-J.; Yoon, Y. Recent Trends in Chemical Sensors for Detecting Toxic Materials. *Sensors* **2024**, *24*, 431. https://doi.org/10.3390/s24020431

Academic Editors: Marijo Buzuk, Maša Buljac and Nives Vladislavić

Received: 7 December 2023 Revised: 3 January 2024 Accepted: 9 January 2024 Published: 10 January 2024



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## 1. Introduction

Anthropogenic activities, including industrial development and manufacturing, are responsible for various types of environmental pollution caused by the release of toxic chemicals [1]. These chemicals include airborne toxic chemicals, medicines, heavy metals, and the byproducts of anthropogenic activities. Although often generated for the good of humankind, failure to effectively manage or control such materials threatens both the environment and human health [2]. The release of harmful chemicals (such as phenols, polycyclic aromatic hydrocarbons, herbicides, insecticides, and nitroxides) into the environment can disrupt the balance between ecosystems and human well-being [3,4]. Therefore, tools and methods for detecting and monitoring toxic materials are crucial for protecting human health. As we live in an era of abundant material development, it is also important to characterize and assess the risks of newly developed materials. Many countries, including South Korea, have implemented laws such as "The Act on Registration and Evaluation of Chemicals" to document the chemical and physical properties of these materials [5]. Although these systems are strictly controlled by national authorities, toxic materials must also be thoroughly monitored to avoid potential threats to human health.

Traditionally, environmental monitoring of toxic chemicals has been performed using instrument-based analyses, such as spectrophotometry, high-performance liquid chromatography, gas chromatography, and mass spectrometry [6–8]. However, despite the superior sensitivity and precision of these tools, demand has increased for other methods that can compensate for the disadvantages of instrument-based analyses, such as high instrument and time costs. Chemical sensors represent a rapid and simple alternative method of target detection. As such, multiple types of chemical sensors with various applications have been developed using the novel techniques of different research fields.

Chemical sensors transform chemical information, ranging from the concentration of a specific sample component to the total composition, into an analytically useful signals [9]. In other words, chemical sensors systems can recognize and report signals originating from the chemical reaction of an analyte or changes in its chemical and physical properties. The IUPAC Commission defines chemical sensors as follows: "analytical chemical sensors are miniaturized transducers that selectively and reversibly respond to chemical compounds or ions and yield electrical signals which depend on the concentration" [10]. Janata et al. documented the definitions and types of existing chemical sensors [11]. Chemical sensing involves a data collection process that identifies the chemical components in a system by measuring the chemical and physical property changes induced by the interaction between targets and sensing platforms. Based on this unique interaction, diverse data acquisition techniques have been applied to develop many types of chemical sensor, including thermal, electrochemical, potentiometric, and optical techniques [12–14]. Although traditionally classified according to the method of data transduction, such as electrochemical responses, optical responses, electrical signals, optical signals, changes in electrical properties, and mass changes, chemical sensors can also be classified according to the materials used for the sensing devices; for example, nanostructure-based sensors, carbon nanotube-based sensors, biological system-based sensors, and graphene-based sensors [15–19]. Since the chemical sensors were designed using a combination of sensing and signal transducing elements, the types of chemical sensors could be diversified enormously. In this regard, it is not meaningful to classify chemical sensors simply by their signal transducing/translating type or by their sensing elements.

Many existing review articles have effectively summarized the principles and mechanisms of various types of existing chemical sensors, including nanostructured materialbased sensors, carbon nanotube sensors, graphene-based sensors, nanomaterial-based sensors, and biological system-based sensors, which are also categorized as electrochemical and optical chemical sensors [12,15,20–23]. Biosensors that use biomolecules such as enzymes, proteins, and living cells as composites for sensing and transducing signals represent a subclass of chemical sensors [10,24]. As a review of all chemical sensors would be impractical because of their substantial diversity and the continual development of new biosensor techniques, this review focuses on recent trends in chemical sensors used to detect toxic chemicals and discusses recent achievements in novel chemical sensor technologies. First, we discuss the applications and prospects of chemical sensors and introduce different types of chemical sensors for detecting toxic chemicals. We then discuss recent findings and applications of chemical sensors for toxic chemical monitoring, including transcription factor (TF)-based biosensors.

#### 2. Overview of Chemical Sensors for Detecting Toxic Materials

Chemical sensors are divided into various subclasses based on the materials used for the sensing elements and the types of output signal. Nonetheless, the basic principles and working mechanisms of chemical sensors are similar in terms of the major components, i.e., sensing elements and signal-transducing elements. The basic components of chemical sensors are shown in Figure 1. Chemical sensors comprise target sensing and signaltransducing element. Output type is also a significant factor in the classification of chemical sensors. Although chemical sensors are classically divided into optical, electrochemical, and thermal sensors based on the type of output signal, classification based on the sensing element has become more complicated. Chemical sensors that employ biomolecules as components are now categorized as biosensors. Owing to advances in materials science and fabrication technologies, new types of sensing element have recently been developed. Several types of chemical sensors detect the same target, and some chemical sensors can detect several targets via the sensing elements. For example, Balaji et al. reported that an optical sensor employing nanostructured cages as the sensing element can detect various metal ions including Sb, Hg, Pb, and Cd by modifying the cage structures [25]. In addition, methyl parathion, a component of pesticides, can be detected using electrochemical sensors that employ nanostructure-based sensing elements [26,27]. Therefore, the performance of chemical sensors can be determined using the target selectivity of the sensing elements and the sensitivity of the signal-transducing elements. From this perspective, new chemical sensors can be developed by combining sensing and signal-transducing elements.



Figure 1. Types of chemical sensors based on sensing and transducing elements.

The purpose of chemical sensors is to detect targets, including gases, chemicals, biomolecules, and cells, for diagnosis, monitoring, or determining production efficiency. However, considering the abundance of toxic materials released into the environment, the major application of chemical sensors is the detection of harmful materials that adversely affect human health. Among the many types of chemical sensors reported in different research fields, we focus on three types: optical, electrochemical, and biological system-based sensors.

## 3. Optical Chemical Sensors

Optical sensors are a group of chemical sensors that employ electromagnetic radiation as the energy source to induce transduction signals in the presence of targets. As described above, optical chemical sensors comprise target sensing (recognition) and signaltransducing elements; however, signal transduction is based on various optical principles, such as absorbance, emission, reflectance, and fluorescence. Therefore, direct spectroscopic methods such as UV spectroscopy and Raman spectroscopy are also considered optical sensors. In this review, according to the definition of chemical sensors, we include only sensors that employ changes in electromagnetic radiation as signal-transducing elements. Because the principles and working mechanisms of optical sensors have been discussed in many excellent reviews, we focus here on their applications for detecting toxic materials and recent progress in optical chemical sensors [14,28–32].

## 3.1. Fiber Optic Chemical Sensors

Optical fibers are the most widely investigated platform for optical chemical biosensors. Optical fibers are used as sensing elements to measure physical properties such as fluorescence, absorption, and reflectance. Although the different subclasses of fiber-optic chemical sensors have the same working principle, the target specificity and selectivity are determined by the fabrication of optical fibers. Fiber-optic chemical sensors (FOCS) represent an area of active research that has led to several new chemical sensors [14,33–35]. The optical fiber technology was coupled with surface plasmon resonance (SPR) to generate different novel fiber-optic sensors. Since the characteristics of SPR were varied by optic structures and metal fabrication on fiber optics, the novel SPR-based, fiber-optic sensors have been developed. Boruah et al. reported a SPR-based, fiber-optic sensor for monitoring lead in water samples [36]. They used U-shaped optic fiber fabrication with chitosan and glutathione as sensing elements and detected ppb ranges of lead ions employing SPR as a transducing element. In this regard, it was speculated that the new chemical sensors would be obtained by coupling new sensing elements and signal transducing elements if the appropriate sensing elements were developed. Additionally, advanced techniques such as microfluidic fabrication, lossy mode resonance, and nanoparticle fabrication have emerged to enhance the sensitivity and selectivity of FOCS [37–39]. However, new types of optical chemical sensors continue to emerge that combine various types of sensing and transducing elements. Table 1 lists existing types of optical chemical sensors according to their sensing element and transducer type.

Type of Transducer	Sensing Element	Target	LOD/Detection Ranges	Optical Response	Ref.
LMR-based	Indium tin oxide NPs	Hydrogen gas	-	LMR	[40]
refractometer	Zinc oxide nanorods	Sulfide gas	-		[41]
SPR based-optic fiber	Graphene film	Streptavidin	-	Reflective index	[35]
	Chitosan-optic fiber	Pb(II)	1–7 ppb		[36]
In-fiber optofluidic device	mPOF	Minocycline	100 ppb	Chemilumin.	[42]
		$C_{11}(II)$ Ni(II)	0.29 ppm,		
Microfluidic device	Chemicals	Cr(VI)	0.33 ppm,	Colorimetric	[43]
			0.35 ppm		
	Zinc microparticles	Nitrate	19 µM	Colorimetric	[44]
	Berthelot reaction	Ammonia	-	Absorbance	[45]
	AuNPs	Hg(II)	-	Colorimetric	[46]
Microfluidic capillary waveguide	Griess reagents	Nitrite	7 ppb	Colorimetric	[47]
Naked eyes/ UV-Vis spec.	Nanostructured cages	Sb(III), Hg(II), Pb(II)	33.7 nM, 6.34 nM, 2.38 nM	Absorbance	[25]
Naked eyes	DNA hybridized AuNPs	Mercury ions (Hg <sup>2+</sup> )	0.5 mM	Colorimetric	[48]
Fluorescence spectroscopy	SWCNTs	DNA sequences	4.0 nM	Fluorescence	[49]
Chemiluminescence					
analyzer	CDs	Phenol	0.76 mM	Fluorescence	[50]
Fluorescence	wsNP-CDs	Trinitrophenol	23 µM	Fluorescence	[51]
spectroscopy		*			

Table 1. List of optical chemical sensors classified by type of sensing element and transducer.

Note: LMR, lossy mode resonance; SPR, surface plasmon resonance; mPOF, microstructured polymer optic fiber; AuNPs, gold nanoparticles; SWCNTs, single-walled carbon nanotubes; wsNP-CDs, water-soluble nitrogen and phosphorous-doped carbon dots.

#### 3.2. Microfluidic System-Based Optical Chemical Sensors

Among the new chemical sensors, biochip- and microfluidic-based optical chemical sensors can rapidly and conveniently detect and monitor various environmentally toxic materials [52,53]. Since microfluidic devices handle micron-scale samples using microfluidic channels, it has the advantage of requiring fewer samples and having faster analytic processes. Although the structure of fiber-optic microfluidic sensors differs from that of typical FOCS, they can also target toxic materials such as antibiotics, bisphenol A, and various pharmaceuticals with comparable detection limits. Similar to other chemical sensors, microfluidic devices would act as sensing platforms by integrating sensing elements into microfluidic devices. Recently, Li et al. fabricated a microfluidic chip-based, fiber-optic sensor for detecting the antibiotic minocycline [42], which comprised an optical fiber with a microstructured polymer, fabricated using polymethyl methacrylate and polystyrene, to generate an in-fiber optofluidic chemiluminescence device for minocycline detection. A luminol solution was added to report changes in chemiluminescence signals against minocycline concentrations. The sensor exhibited a minocycline detection limit of 100 ppb, which is comparable to that of high-performance liquid chromatography-based analysis [54]. Wang et al. has reported heavy-metal detecting sensors based on a paper-based microfluidic device [43]. They generated heavy metal monitoring microfluidic-based sensors by fabricating metal selective chromogenic reagents on paper-based microfluidic devices. And, it showed 0.29 ppm, 0.33 ppm, and 0.35 ppm detection limits for Cu(II), Ni(II), and Cr(VI), respectively. Consequently, it was emphasized that the fusion of scientific technologies from different research fields makes chemical sensors more advanced. Besides the microfluidic sensors based on the optic fiber mentioned here, many FOCS have been coupled with microfluidic systems for the detection of toxic materials [47,55,56].

FOCS are classified via their target sensing elements, sensing materials, and detection principles [55,56]. As shown in Figure 1, the target sensing element is used to identify chemical compounds, functionalized nanomaterials, aptamers, antibodies, proteins, and even cells. The target sensing elements are coupled with sensing materials such as nanoparticles, luminol, graphene oxide, carbon nanotubes (CNTs), and microfluidic devices, which transduce changes to the signals [57]. Therefore, the targets of chemical sensors are determined by the specificity and selectivity of the sensing elements. As such, it is critical to develop and investigate new sensing elements to expand the range of sensor targets. Once target-specific sensing elements are prepared, they can be coupled with different techniques to generate various types of chemical sensors.

## 3.3. Nanoparticle-Based Optical Chemical Sensors

Optical chemical sensors are based on changes in electromagnetic radiation. Among the many detection principles related to optical properties, colorimetric and fluorometric changes are suitable for the detection of toxic materials. Chemical sensors based on these principles must be equipped with a source for excitation and a detector [58,59]. However, samples must include species that exhibit intrinsic fluorescence. Thus, it is critical to endow samples with fluorescent properties for colorimetric and fluorometric measurements. Gold nanoparticles (AuNPs) are suitable sensor materials because they exhibit size-dependent colorimetric and fluorescent properties [60,61].

He et al. reported that rhodamine B functionalized AuNPs can be used in microfluidic devices to detect Hg ions [46]. Because AuNPs show different colorimetric changes according to their functionalization, size variation, and dispersion rate, AuNP-based chemical sensors have been developed to detect various targets, such as ribonucleotides, ascorbic acid, Cr, Pb, and glucose [62]. Although AuNPs are employed as signal-producing elements in chemical sensors, the target selectivity is determined using their functionalization/fabrication. For example, cysteine-capped AuNPs have been used for ethyl parathion and malathion detection [63,64], and DNA hybridization on AuNPs has been used for Hg ion detection [48]. Consequently, new functionalization (fabrication) technologies for AuNPs have accelerated the development of AuNP-based chemical sensors and expanded the range of targets, not only heavy metal ions but also various other chemicals [65–67]. The type of transducer used in these chemical sensors is related to the target sensitivity. For example, the sensitivity of AuNP-based chemical sensors toward Hg ions was enhanced from 5  $\mu$ M to 0.002  $\mu$ M of the detection limit by employing a surface-enhanced Raman as the transducer [68]. Developing chemical sensor based on metal nanoparticles including AuNPs and AgNPs is a state-of-the-art multidisciplinary science; thus, it is not reasonable to classify them just by the type of transducers. Rather, it would be better to put them into nanomaterial-based sensors, discussed in Section 3.4.

#### 3.4. Nanomaterial-Based Optical Chemical Sensors

The rapid development of new materials and technologies in different research fields has diversified the range of materials and fabrication methods available for sensing elements. Notably, techniques for the fabrication of nanomaterials and nanostructures have opened the door to new sensing elements that can recognize various targets [69,70]. For

example, metal nanoparticles, CNTs, metal oxides (quantum dots), and metal organic frameworks are current areas of active research in nanomaterials [70–72]. Sensor applications and target selectivity are determined by the type of fabrication and conjugation of nanomaterials. For example, carbon nanomaterials (CNMs) have been used to construct carbon dots, CNTs, graphene, and carbon black, all of which have different properties and are employed in various chemical sensors as sensing elements [73,74]. Carbon dots have been used to develop sensors for detecting cations and anions as well as small molecules and drugs with surface modifications [75,76]. CNTs have also been used as sensing elements with various modifications to detect gaseous analytes such as NH3, NO2, and CO, as well as biomolecules such as boronic acid and glucose [77,78].

Recently, optical sensors based on nanostructured cage material were developed by Balaji et al. for detecting toxic metal ions [25]. Heavy metal ions are typically determined using atomic absorption and emission spectroscopy. However, methods with greater sensitivity and simplicity are required because traditional methods are expensive and complicated. Nanomaterial-cage-based chemical sensors comprise optical-sensor-based cubic Fm3m cage monoliths with different fabrications as sensing elements that use colorimetric changes for signal transduction. Briefly, cubic Fm3m cage monoliths were designed as platforms, and dithizone, TMPyP ( $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$ -tetrakis(1-methylpyridinium-4-yl) porphinep-toluenesulfonate), pyrogallol red, and tetraphenyl porphine tetrasulfonic acid were used as cages for detecting Pb, Cd, Sb, and Hg, respectively. The greatest advantage of the nanomaterial-cage-based sensor is that it enables target detection with a high quantification limit by the naked eye.

In addition to the studies mentioned here, many review papers have focused not only on specific nanomaterial-based chemical sensors but also on their working mechanisms, target selectivity upon fabrication, and applications [69,70,79]. Although some aspects of nanomaterial-based chemical sensors are discussed in this review, it should be noted that all chemical sensors share similarities in their basic principles, and the potential of chemical sensors continues to expand along with technological advances.

## 4. Electrochemical Sensors

Electrochemical sensors are an important subclass of chemical sensors that use electrodes as signal-transducing elements [12,80]. Electrochemical sensors can be further divided into potentiometric sensors, voltametric sensors, and conductimetric sensors. These sensors measure the changes in a potential signal caused by an ion-recognition event, whereby the potential between two electrodes causes the oxidation (or reduction) of an electroactive species, with the resistance representing the signal-transducing element [81,82]. Electrochemical sensors measure changes in electrochemical properties induced by the interactions between sensing elements and targets and are further classified based on the type of sensing element, which includes nanomaterials, biomolecules, CNTs, graphene, and optical chemical sensors. Advances in the fields of materials sciences, electrical engineering, device fabrication, and physical chemistry have continually improved the signal transducers and sensing elements of electrochemical sensors. Table 2 lists the types of transducers, sensing elements, and targets of electrochemical sensors.

Table 2. List of electrochemical sensors classified by type of sensing element and transducer.

Type of Transducer	Sensing Element	Target	LOD	Electrochemical Response	Ref.
SPCE	WS2/MWCNTs-OH	2,4,6-trichlorophenol	-	Cyclic voltammetry	[83]
		bisphenol AF	-		
		PSNP	-		
Electrochemical analyzer	LIG	4-nitrophenol	95 nM	Cyclic voltammetry	[84]
Metal electrode	Cu <sub>2</sub> O-rGO	NÔ <sub>2</sub>	50 ppb	Resistance	[85]
	Graphene flake	CO <sub>2</sub>	-	Resistance	[86]
GCE	MWCNTs/CuO-Au	4-aminophenol	0.105 μM	Differential pulse	[87]
		Acetaminophen	0.016 µM	voltammetry	

Type of Transducer	Sensing Element	Target	LOD	Electrochemical Response	Ref.
	AuNPs/DNA AuNPs/CNTs-ErGO	DNA Hydrazine	0.78 fmol 0.065 μM	Cyclic voltammetry	[88] [89]
	AgNPs	Pendimethalin Ethyl parathion	36 nmol/L 40 nmol/L	Square-wave adsorptive Stripping voltammetry	[90]
Metal electrode	cauliflower-shaped ZnO	Picric acid Nitrophenol	0.078 mM	Current voltage technique	[91]
	TiO <sub>2</sub> -CNTs/Pt	$H_2O_2$	0.016 µM	Cyclic voltammetry	[92]

Table 2. Cont.

Note: SPCE, screen-printed carbon electrode; WS2/MWCNTs-OH, tungsten disulfide nanosheets/hydroxylated multi-walled carbon nanotubes; PSNP, polystyrene nanoplastics; LIG, laser-induced graphene; rGO, reduced graphene oxide; GCE, glassy carbon electrode; CNTs-ErGO, carbon nanotube electrochemically reduced graphene oxide composite film.

As many toxic materials are released from anthropogenic activities, heavy metal(loid)s (such as mercury, lead, cadmium, arsenic), chemicals (such as phenolic compounds, nitroaromatics, organophosphorus), and pesticides are increasingly found in environmental systems. Therefore, electrochemical sensors play a pivotal role in avoiding the impact of toxic materials by enabling constant monitoring via simple and fast analytical tools.

#### 4.1. CNM-Based Electrochemical Sensors

Electrochemical sensors are defined as chemical sensors that employ potentiometric, amperometric, and conductometric changes as transduced signals, affording them superior sensitivity to optical chemical sensors. Nonetheless, electrochemical sensors are similar to optical chemical sensors in terms of sensing element advances and diversity. Recent advances in nanomaterial science have provided various materials for sensing elements, including metal nanoparticles (NPs) and CNMs such as carbon dots, CNTs, and graphene [26,93,94]. The integration of fabrication technologies has also enabled the use of nanostructured materials as sensing elements in chemical sensors [95,96].

As for optical chemical sensors, CNMs are widely used as sensing elements in electrochemical sensors [97,98]. Carbon nanostructures used for electrochemical sensors include fullerenes, CNTs, graphene, nanocones, and carbon dots, which are classified in terms of their size and shape. In general, nanostructured carbon is applied to electrodes via a fabrication process to detect targets such as pesticides, toxic chemicals, and heavy metal ions. Recently, Wu et al. reported CNT-based electrochemical sensors for measuring the cytotoxicity of 2,4,6-trichlorophenol, bisphenol AF, and polystyrene nanoplastics [83]. The electrodes were modified with tungsten disulfide nanosheets/hydroxylated multi-walled CNTs (WS2/MWCNTs-OH) for enhanced sensitivity. A recent study reported the development of laser-induced graphene printed on polyimide films for 4-nitrophenol detection in water [84]. The presence of 4-nitrophenol induced a cyclic voltammetry response that corresponded to the concentration of 4-nitrophenol. In addition, several studies and review papers have been written on CNM-based sensors [97,99,100]. Owing to their diverse intrinsic electronic and optical properties, chemical versatility, and stability, CNMs are attracting increasing research attention, with new findings on CNM-based chemical sensors also reported.

#### 4.2. NP-Based Electrochemical Sensors

Recently, the use of NPs has increased in a variety of fields, including environmental, pharmaceutical, medical, and material sciences, as well as the cosmetics industry [101–104]. The wide application of NPs is related to their novel properties, which depend on the shape and size of the particles [105]. Thus, methods of synthesizing NPs represent a key research area [106,107]. Typically, NPs are divided into metal-based, metal-oxide-based, and carbon-based NPs [108]. Because the application of CNMs to electrochemical sensors

was addressed in the previous section, only electrochemical sensors employing metal- and metal-oxide-based NPs are discussed in this section.

Among the metal-NP-based electrochemical sensors, AuNPs have received the most attention. Castañeda et al. reported AuNP-based electrochemical sensing of DNA [88], and Zhao et al. reported hydrazine detection using an AuNPs/CNTs-ErGO electrochemically reduced graphene oxide composite film [89]. In both cases, the AuNPs were employed as sensing elements to recognize targets. Silver nanoparticles (AgNPs) have also been used in electrochemical sensors [108]. Similar to AuNPs, AgNPs were fabricated on the electrodes of electrochemical sensors to serve as target sensors. AgNP-based electrochemical sensors targeting DNA, chemicals, and pesticides have been developed with variations in electrode types and other modifications [90,109].

Similar to NPs, metal oxides have been applied in various fields because of their novel characteristics, which are determined by their shape and size [110,111]. Among the metal oxides, the use of zinc oxide (ZnO) for sensing toxic materials has been extensively discussed in review articles [70,112–114]. ZnO is inexpensive and can be used to fabricate various nanostructures in a relatively simple manner. The shapes of ZnO nanostructures range from 1-D to 3-D, with each shape exhibiting distinct properties. For example, a thin layer of ZnO on the sensor surface enhances the electrochemical response to Hg by facilitating the migration of electrons between the redox-active analytes [115]. Moreover, Ibrahim et al. synthesized cauliflower-shaped ZnO and used it to modify electrodes for the detection of picric acid [91]. Many other types of ZnO have also been reported and employed for electrode modification. Unlike other nanostructured materials, ZnO plays a role in both target selectivity and transduced-signal amplification.

The versatility of nanomaterials, including metal NPs and metal oxides, has been accepted and proven in different research fields, which has accelerated research into their development and application. Advances in nanomaterials science have also provided significant benefits to many different research fields and industries, with their potential as components of chemical sensors being particularly important. Although metal NPs act to enhance the transducing signals through their integration with electrodes, they can also function as sensing elements with improvements to the chemical modification and surface fabrication of NPs. Although it is difficult to endow NPs with target selectivity, studies have achieved target sensitivity by conjugating NPs with organic ligands as well as biological molecules, such as chitosan, DNA, and proteins [116–118]. If NPs were conjugated with biomolecules and employed as sensing elements in chemical sensors, they could be classified as biosensors. In this regard, we discussed the application of biomolecule conjugated NPs further in Section 5 with biosensors.

## 5. Biosensors

Biosensors targeting biomolecules of interest are usually considered a subclass of chemical sensors because they use the same target sensing and transduction methods [119]. As shown in Figure 2, biosensors are divided into two categories based on their ability to transduce signals into digitized values. One category includes biosensors that integrate sensing and transducing elements with analytical devices, whereas the other requires additional instruments to read the transduced signals. The former type employs biomolecules and biochemical or biological mechanisms as sensing elements and recognition systems and are, therefore, considered to be conventional biosensors [120]. The latter are biosensing systems based on transcription factors (TFs), such as whole-cell biosensors, which require equipment to read the transduced signals. In this regard, various chemical sensors that use biomolecules, such as DNA, enzymes, and antibodies, for target recognition are considered biosensors [121–123]. In contrast, TF-based biosensors are not fully accepted as chemical sensors, despite target recognition occurring through the transduction of enzymatic activities and fluorescence. Nonetheless, TF-based biosensors are included in this review because the sensing systems can be chemical sensors equipped with sensing and signal-transducing elements.



Figure 2. The types of biosensors based on ability to transduce signals into digitized values.

#### 5.1. Optical Biosensors and Electrochemical Biosensors

As discussed above, optical and electrochemical sensors are classified according to the method of transducing signals indicating the interaction between targets and sensing elements. In general, optical biosensors are based on technology that detects evanescentwave changes caused by the interaction between targets and sensing elements incorporated with biological molecules [124,125]. Surface plasmon resonance, optical waveguides, optical resonators, and optical fibers have all been used as transducers to measure changes in the reflective index, fluorescence, Raman scattering, and optical absorption upon target recognition [126]. Electrochemical biosensors also employ biomolecules as sensing elements and electrodes to measure electrochemical properties such as voltametric, potentiometric, and electrometric changes as transduced signals [127–129]. Although different technologies have been employed for biosensors, the range of targets is determined by the sensing elements coupled with various biomolecules, such as antibodies, enzymes, DNA, aptamers, and cells. As biomolecules are more specific and selective to targets than other materials, biomolecule-coupled chemical sensors are a key area of research. Since the targets of biomolecules are diverse such as chemicals, heavy metals, chemicals, other proteins, and cells, the detecting ranges of targets were expanded. Therefore, the type and integration method of biomolecules used in chemical sensors are critical considerations. Table 3 lists the different types of biosensors according to their sensing and transducing elements.

<b>Table 3.</b> List of biosensors classified by type of sensing element and transduc
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Type of Transducer	Sensing Element	Target	LOD	Response	Ref.
Fluorescence spec.	ChE-SWCNT	Pesticides Heavy metals	-	NIR fluorescence	[130]
Optic fiber GCE Au electrode Potentiostat	Toluene monooxygenase Urease-polyaniline DNA-Cu <sub>2</sub> O@NCs DNA-ZnO NPs	Toluene Urea Hg(II) Yellow fever virus	3 μM 0.1 mM 0.15 nM 0.01 μM	Absorbance Cyclic Voltammetry Cyclic voltammetry	[131] [132] [133] [134]
Fluorescence spec. SPR	Aptamer-CDs/GO Aptamer-Biotin	Pseudomonas aeruginosa	9 CFU/mL 10 CFU/mL	Fluorescence Reflective index	[135] [136]
TFs-based biosensors	ArsR CueR ZntR TbuT MntR MobR TetR mphR	As(III), As(V) Cu(II) Pb(II), Hg(II), Cd(II) BTEX Mn(II) 3-hydroxybenzoate Tetracycline erythromycin	$\begin{array}{c} 10 \ \mu g/L \\ 10 \ nM \\ - \\ 0.24 \pm 0.22 \ \mu M \\ 0.01 \ \mu M \\ 2 \ mM \\ 1.25 \ \mu M \\ 50 \ \mu M \end{array}$	Fluorescence	[137,138] [139] [139,140] [141] [142] [143]
	BenR	benzoate	1 nM		[144]

Note: SWCNT, single-walled carbon nanotubes; Cu<sub>2</sub>O@NCs, cuprous oxide and nanochitosan composites; CDs, carbon dots; GO, graphene oxide; SPR, surface plasmon resonance; BTEX, benzene, toluene, ethylbenzene, and xylene.

#### 5.2. Enzyme-Based Biosensors

Because enzymes have specific targets, they are attractive for use as sensing elements in chemical sensors. The role of enzymes in chemical sensors is to sense targets and transduce signals based on their activities [14,145], with enzymes such as hydrolases, oxidoreductases, and transferases all used as bioreceptors for sensing targets. Enzyme activities induce optical or electrochemical changes and can be used to detect heavy metals, pharmaceuticals, and phenolics [146].

For example, cholinesterase is widely used as a sensing element to detect toxic materials such as pesticides, heavy metals, and toxins because its activity is inhibited by toxic materials [147]. Recently, Loewenthal et al. used acetylcholinesterase (AChE) coupled with a near-infrared, fluorescent, single-walled carbon nanotube optical sensor. AChE releases thiocholines from acetylthiocholine, increasing near-infrared fluorescence and, thereby, detecting AChE inhibitors by measuring the decrease in signals [130,148]. Another example of an enzyme used in chemical sensors is toluene monooxygenase for toluene detection [131]. This enzyme recognizes and degrades toluene, resulting in the consumption of oxygen, then induces changes in the phosphorescence intensity, which act as transduced signals.

The greatest advantage of enzymes is their target selectivity and enzymatic activity. Enzymes are integrated into chemical sensors to detect targets and produce output signals induced by catalytic activity [149]. However, the stability of enzymes would be an obstacle to enlarging their application because the enzymatic activity is dependent on the stable structure of enzymes. Thus, it would be necessary to consider the stability of enzymes during the integration on the components of sensors. Nonetheless, the advantageous aspects of enzymes accelerate the application of enzyme-based biosensors not only for the detection of toxic materials but also for biomedical analysis [150,151]. As biomolecules such as specific proteins and metabolites indicate certain diseases, enzymes that recognize these molecules have been employed as sensing elements in chemical sensors for biomedical analysis.

## 5.3. Biomolecule-Based Biosensors

Biomolecules such as DNA, peptides, antibodies, and aptamers have been employed as target-sensing elements in chemical sensors [121,152,153]. Target-specific biomolecules are integrated into sensing devices as sensing elements. Changes in the chemical properties induced by the interaction with targets are then transduced by various transducing elements. The basic components are the same as those of other chemical sensors; however, the biomolecules are used for sensing the targets. This type of sensor can also be classed as optical or electrochemical sensors according to the type of signal transducer.

Unlike enzymes, the biomolecules such as DNA, peptides, antibodies, and aptamers possess more versatile natures to modulate target selectivity and sensitivity. In this regard, those biomolecules were actively investigated as sensing elements for chemical sensors. With advances in nanomaterial fabrication technologies, various materials have been conjugated onto nanomaterials. In addition to chemicals and functional groups, biomolecules, such as DNA, antibodies, and aptamers, have been conjugated as ligands onto nanomaterials [123,154]. For example, Liu et al. developed mercury-sensing electrochemical sensors by integrating DNA strands onto cuprous oxide/nanochitosan composites [133]. Interfacial changes on the surface of the electrode caused by DNA-Hg interactions were measured using electrochemical impedance spectroscopy, which showed a nanomolar range of sensitivity. Aptamers, also known as chemical antibodies, are versatile materials used as sensing elements in chemical sensors. Aptamers are single-stranded DNA/RNA oligonucleotides with the advantages of low production costs, versatile applications, and easy modification and operation [155]. Wang et al. and Hu et al. have reported aptamer-based biosensors for monitoring *Pseudomonas aeruginosa* with 10 CFU/mL of detection limit using CDs and biotin integrated aptamers, respectively [135,136]. To facilitate the use of aptamers in chemical sensors, techniques for conjugating aptamers with nanomaterials have rapidly improved. Consequently, aptamer-conjugated NPs, CNMs, and quantum dots have been employed as elements in various chemical sensors [156-158]. For example, aptamer-based

chemical sensors were applied to detect heavy metals, cancer cells, and various proteins according to the specificity of the aptamers. In addition to the biomolecules mentioned here, many other chemical sensors employ biomolecules as components, such as antibodies, proteins, chitosan, and carbohydrates. By integrating these biomolecules into sensing elements, biosensors can be used to detect a wide range of targets, including toxic gases, heavy metals, phenolic compounds, proteins, and even cancer cells.

Researchers have also reviewed chemical sensors with antibody-, aptamer-, and DNAbased sensors, emphasizing the huge potential of integrating biomolecules with nanomaterials, including CNTs, various NPs, quantum dots, and different types of electrodes for detecting toxic materials [121,159,160].

#### 5.4. TF-Based Biosensors

Unlike the chemical sensors discussed above, TF-based biosensors do not transduce signals to digitized values but still comprise sensing and signal transduction elements [161,162]. The sensing elements are TFs, and targets interacting with the TFs are turned on or off via the transcription of genes. The target–TF interaction is indicated by gene expression; therefore, the gene expression level is the signal output [163,164]. For this reason, TF-based biosensors are often called bioreporters and are coupled with additional instruments such as UV/Vis spectroscopy and fluorescence spectroscopy.

Among the TF-based biosensors, whole-cell-based biosensors have been intensively investigated. In these biosensors, the cells are used as sensor platforms to serve as both sensing and signal-transducing elements [165]. Typically, the TFs are target-sensing elements and genes that encode enzymes or fluorescent proteins as signal-transducing elements. As TFs are involved in external stimuli, cells possessing genetically engineered TFs and reporter genes are responsible for specific stimuli. Thus, specific toxic materialsensing biosensors have been developed by employing TFs that respond to toxic materials as sensing elements. TF-based biosensors target a wide variety of materials, including biomolecules and cellular metabolites, but the targets are restricted to toxic materials in this review. The TFs and their corresponding toxic materials are listed in Table 3. For example, Escherichia coli contains operons responsive to arsenic and manganese, which are regulated by ArsR and MntR, respectively. When reporter genes such as egfp are inserted under the promoter regions of operons, E. coli cells serve as sensors to detect arsenic and manganese based on changes in fluorescent signals [142,166]. In addition, the applications of whole-cell biosensors can be increased by integrating cells with electrochemical devices. Because the cells produce electrochemically active materials upon exposure to toxic materials, the signal changes indicate the presence of toxic materials in samples. However, as the target recognition processes occur inside cells, targets impermeable to cells are not detected by whole-cell-based biosensors. To overcome this disadvantage, cell-free sensing systems that use a mixture of components are required for transcription and translation [167,168]. Alam et al. and colleagues have reported a cell-free, TFs-based sensor system named ROSALIND (RNA output sensors activated by ligand induction) [143]. The same TFs used as sensing elements for whole cell-based biosensors are employed as sensing elements. The interaction between target TFs induces the production of RNA reporting signals by forming a complex with fluorescent chemicals. Thus, it avoids the issue of cell permeability of the target. Nonetheless, the cell free system has disadvantages such as the purification of TFs and preparation of components for transcription and translation.

TF-based biosensors are a subclass of chemical sensors that include both sensing and signal-transducing elements. Therefore, similar to the chemical sensors discussed above, diverse targets and improved applications and performance can be achieved by enhancing the sensing elements and signal-transducing technologies. In addition, the target selectivity and specificity of TF-based biosensors could be modulated via genetic engineering of TFs. Although it is challenging to modulate the target interaction of TFs via genetic engineering, the advance in protein modelling and computational analysis makes the process more accurate and efficient. In this way, the performance of TF-based biosensor can be enhanced,

and the targets can also be diversified from existing genetic systems. Because biosensors have many advantages over other chemical sensors, especially in terms of their target specificity and selectivity, efforts should be made to further improve biosensors. Moreover, advances in diverse scientific fields can be used to enhance the performance and application of both chemical sensors and biosensors because both sensors share sensing and signaltransducing elements. Consequently, the rapid development of scientific technologies related to chemical sensors will help protect humans from exposure to toxic materials in the environment.

#### 6. Conclusions and Future Prospects

Advances in various industrial fields have increased the production of toxic materials. Although systems for the control and monitoring of toxic materials have been established in many countries, attempts to reduce the threat to human health have been hindered by the rapid increase in toxic materials. It is impossible to identify all toxic materials, but fast and convenient detection methods are vital for effective monitoring. Moreover, technologies must be developed to respond to and monitor newly generated toxic materials. Chemical sensors are commonly used to detect toxic materials and can, therefore, address these concerns. As described in this review, chemical sensors have undergone rapid improvements, and their applications have increased substantially following advances in different scientific fields. The basic structure of all types of chemical sensors is the same, comprising sensing and signal-transducing elements; however, these elements vary widely between sensor types. Although the chemical sensors discussed here were classified just by types of sensing elements and signal transducing elements, it would not be meaningful these days. To enhance the performance of chemical sensors, it is critical to integrate the interdisciplinary sciences as well as to upgrade sensing and signal transducing elements. In addition, the application fields of chemical sensors have been expended rapidly along with the advances in sensing and signal-transducing elements. Recently, various chemical sensors have been applied to medical sciences to diagnose diseases by monitoring biological markers and pathogenic markers [169,170]. Meanwhile, it has been also reported that whole-cell-based biosensors were used to monitor the toxicity and genotoxicity of harmful materials [171,172]. In this way, the chemical sensors could contribute to secure and improve the human health.

Consequently, the future prospects of chemical sensors not only depend on improving sensing elements and integrating them with signal-transducing elements but also integrating advanced technologies. Although this review only discusses a small number of chemical sensors, we focus on the similarity of chemical sensors in terms of their basic principles and future goals. As such, this review provides a unique perspective on chemical sensors and contributes to their continued development.

**Author Contributions:** Conceptualization, Y.K., Y.J. and Y.Y.; writing—original draft preparation, Y.K., Y.J., M.N. and S.-J.H.; writing—review and editing, Y.K., S.-J.H. and Y.Y.; visualization, Y.K. and Y.J.; supervision, Y.Y. All authors have read and agreed to the published version of the manuscript.

**Funding:** This paper was supported by Konkuk University Researcher Fund in 2023 (to S.-J.H.) and the National Research Foundation of Korea funded by the Ministry of Science, ICT, and Future Planning (2021R1F1A1056635 to Y.Y.).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data are contained within the article.

Conflicts of Interest: The authors declare no conflicts of interest.

## References

- 1. Ukaogo, P.O.; Ewuzie, U.; Onwuka, C.V. Environmental pollution: Causes, effects, and the remedies. In *Microorganisms for Sustainable Environment and Health*; Elsevier: Amsterdam, The Netherlands, 2020; pp. 419–429.
- Ajibade, F.O.; Adelodun, B.; Lasisi, K.H.; Fadare, O.O.; Ajibade, T.F.; Nwogwu, N.A.; Sulaymon, I.D.; Ugya, A.Y.; Wang, H.C.; Wang, A. Environmental pollution and their socioeconomic impacts. In *Microbe Mediated Remediation of Environmental Contaminants*; Elsevier: Amsterdam, The Netherlands, 2021; pp. 321–354.
- 3. Young, S.; Balluz, L.; Malilay, J. Natural and technologic hazardous material releases during and after natural disasters: A review. *Sci. Total Environ.* **2004**, 322, 3–20. [CrossRef]
- 4. Corn, M. Handbook of Hazardous Materials; Academic Press: New York, NY, USA, 2012.
- 5. Ha, S.; Seidle, T.; Lim, K.-M. Act on the Registration and Evaluation of Chemicals (K-REACH) and replacement, reduction or refinement best practices. *Environ. Health Toxicol.* **2016**, *31*, e2016026. [CrossRef]
- 6. Rouessac, F.; Rouessac, A. Chemical Analysis: Modern Instrumentation Methods and Techniques; John Wiley & Sons: Hoboken, NJ, USA, 2022.
- Chen, H.; Zheng, J.; Zhang, X.; Luo, M.; Wang, Z.; Qiao, X. Surface desorption atmospheric pressure chemical ionization mass spectrometry for direct ambient sample analysis without toxic chemical contamination. *J. Mass Spectrom.* 2007, 42, 1045–1056. [CrossRef]
- 8. Leary, P.E.; Kammrath, B.W.; Lattman, K.J.; Beals, G.L. Deploying portable gas chromatography–mass spectrometry (GC-MS) to military users for the identification of toxic chemical agents in theater. *Appl. Spectrosc.* **2019**, *73*, 841–858. [CrossRef]
- 9. Hulanicki, A.; Glab, S.; Ingman, F. Chemical sensors: Definitions and classification. Pure Appl. Chem. 1991, 63, 1247–1250. [CrossRef]
- 10. Cammann, K.; Lemke, U.; Rohen, A.; Sander, J.; Wilken, H.; Winter, B. Chemical sensors and biosensors—Principles and applications. *Angew. Chem. Int. Ed. Engl.* **1991**, *30*, 516–539. [CrossRef]
- 11. Janata, J. Principles of Chemical Sensors; Springer Science & Business Media: Berlin/Heidelberg, Germany, 2010.
- 12. Privett, B.J.; Shin, J.H.; Schoenfisch, M.H. Electrochemical sensors. Anal. Chem. 2010, 82, 4723–4741. [CrossRef] [PubMed]
- 13. Karker, N.; Dharmalingam, G.; Carpenter, M.A. Thermal energy harvesting plasmonic based chemical sensors. *ACS Nano* **2014**, *8*, 10953–10962. [CrossRef]
- 14. Wolfbeis, O.S. Fiber-optic chemical sensors and biosensors. Anal. Chem. 2006, 78, 3859–3874. [CrossRef] [PubMed]
- Huang, X.-J.; Choi, Y.-K. Chemical sensors based on nanostructured materials. *Sens. Actuators B Chem.* 2007, 122, 659–671. [CrossRef]
  Jimenez-Cadena, G.; Riu, J.; Rius, F.X. Gas sensors based on nanostructured materials. *Analyst* 2007, 132, 1083–1099. [CrossRef]
- [PubMed] 17. Liu, Y.; Dong, X.; Chen, P. Biological and chemical sensors based on graphene materials. *Chem. Soc. Rev.* **2012**, *41*, 2283–2307.
- 17. Liu, Y.; Dong, X.; Chen, P. Biological and chemical sensors based on graphene materials. *Chem. Soc. Rev.* **2012**, *41*, 2283–2307. [CrossRef]
- 18. Kauffman, D.R.; Star, A. Carbon nanotube gas and vapor sensors. Angew. Chem. Int. Ed. 2008, 47, 6550–6570. [CrossRef] [PubMed]
- 19. Borisov, S.M.; Wolfbeis, O.S. Optical biosensors. *Chem. Rev.* 2008, 108, 423–461. [CrossRef] [PubMed]
- 20. Meyyappan, M. Carbon nanotube-based chemical sensors. *Small* **2016**, *12*, 2118–2129. [CrossRef] [PubMed]
- 21. Yavari, F.; Koratkar, N. Graphene-based chemical sensors. J. Phys. Chem. Lett. 2012, 3, 1746–1753. [CrossRef] [PubMed]
- 22. El-Ansary, A.; Faddah, L.M. Nanoparticles as biochemical sensors. *Nanotechnol. Sci. Appl.* 2010, 3, 65–76. [CrossRef]
- 23. Ispas, C.R.; Crivat, G.; Andreescu, S. Recent developments in enzyme-based biosensors for biomedical analysis. *Anal. Lett.* **2012**, 45, 168–186. [CrossRef]
- Ronkainen, N.J.; Halsall, H.B.; Heineman, W.R. Electrochemical biosensors. Chem. Soc. Rev. 2010, 39, 1747–1763. [CrossRef] [PubMed]
- 25. Balaji, T.; El-Safty, S.A.; Matsunaga, H.; Hanaoka, T.; Mizukami, F. Optical sensors based on nanostructured cage materials for the detection of toxic metal ions. *Angew. Chem.* 2006, 118, 7360–7366. [CrossRef]
- Karimi-Maleh, H.; Darabi, R.; Baghayeri, M.; Karimi, F.; Fu, L.; Rouhi, J.; Niculina, D.E.; Gündüz, E.S.; Dragoi, E. Recent developments in carbon nanomaterials-based electrochemical sensors for methyl parathion detection. *J. Food Meas. Charact.* 2023, 17, 5371–5389. [CrossRef]
- 27. Dong, J.; Wang, X.; Qiao, F.; Liu, P.; Ai, S. Highly sensitive electrochemical stripping analysis of methyl parathion at MWCNTs– CeO<sub>2</sub>–Au nanocomposite modified electrode. *Sens. Actuators B Chem.* **2013**, *186*, 774–780. [CrossRef]
- 28. Qazi, H.H.; Mohammad, A.B.b.; Akram, M. Recent progress in optical chemical sensors. Sensors 2012, 12, 16522–16556. [CrossRef]
- 29. McDonagh, C.; Burke, C.S.; MacCraith, B.D. Optical chemical sensors. *Chem. Rev.* 2008, 108, 400–422. [CrossRef] [PubMed]
- 30. Wolfbeis, O.S. Materials for fluorescence-based optical chemical sensors. J. Mater. Chem. 2005, 15, 2657–2669. [CrossRef]
- Fakayode, S.O.; Lisse, C.; Medawala, W.; Brady, P.N.; Bwambok, D.K.; Anum, D.; Alonge, T.; Taylor, M.E.; Baker, G.A.; Mehari, T.F. Fluorescent chemical sensors: Applications in analytical, environmental, forensic, pharmaceutical, biological, and biomedical sample measurement, and clinical diagnosis. *Appl. Spectrosc. Rev.* 2023, 59, 1–89. [CrossRef]
- 32. Singh, A.K.; Mittal, S.; Das, M.; Saharia, A.; Tiwari, M. Optical biosensors: A decade in review. *Alex. Eng. J.* **2023**, 67, 673–691. [CrossRef]
- 33. Lee, B.; Roh, S.; Park, J. Current status of micro-and nano-structured optical fiber sensors. *Opt. Fiber Technol.* **2009**, *15*, 209–221. [CrossRef]
- Lin, J. Recent development and applications of optical and fiber-optic pH sensors. *TrAC Trends Anal. Chem.* 2000, 19, 541–552. [CrossRef]

- 35. Kim, J.A.; Hwang, T.; Dugasani, S.R.; Amin, R.; Kulkarni, A.; Park, S.H.; Kim, T. Graphene based fiber optic surface plasmon resonance for bio-chemical sensor applications. *Sens. Actuators B Chem.* **2013**, *187*, 426–433. [CrossRef]
- Boruah, B.S.; Biswas, R. An optical fiber based surface plasmon resonance technique for sensing of lead ions: A toxic water pollutant. Opt. Fiber Technol. 2018, 46, 152–156. [CrossRef]
- 37. Ozcariz, A.; Ruiz-Zamarreno, C.; Arregui, F.J. A comprehensive review: Materials for the fabrication of optical fiber refractometers based on lossy mode resonance. *Sensors* **2020**, *20*, 1972. [CrossRef]
- Ju, S.; Nguyen, V.L.; Watekar, P.R.; Kim, B.H.; Jeong, C.; Boo, S.; Kim, C.J.; Han, W.-T. Fabrication and optical characteristics of a novel optical fiber doped with the Au nanoparticles. *J. Nanosci. Nanotechnol.* 2006, *6*, 3555–3558. [CrossRef] [PubMed]
- Hartmann, D.M.; Nevill, J.T.; Pettigrew, K.I.; Votaw, G.; Kung, P.-J.; Crenshaw, H.C. A low-cost, manufacturable method for fabricating capillary and optical fiber interconnects for microfluidic devices. *Lab A Chip* 2008, *8*, 609–616. [CrossRef] [PubMed]
- 40. Mishra, S.K.; Usha, S.P.; Gupta, B.D. A lossy mode resonance-based fiber optic hydrogen gas sensor for room temperature using coatings of ITO thin film and nanoparticles. *Meas. Sci. Technol.* **2016**, *27*, 045103. [CrossRef]
- 41. Usha, S.P.; Gupta, B.D. Performance analysis of zinc oxide-implemented lossy mode resonance-based optical fiber refractive index sensor utilizing thin film/nanostructure. *Appl. Opt.* **2017**, *56*, 5716–5725. [CrossRef]
- Li, Z.; Yang, X.; Teng, P.; Kong, D.; Gao, S.; Liu, Z.; Yang, J.; Gao, D.; Luo, M.; Wen, X. Determination of the antibiotic minocycline by integrated optofluidic microstructured polymer optical fiber chemiluminescence. *Instrum. Sci. Technol.* 2021, 49, 571–584. [CrossRef]
- 43. Wang, H.; Li, Y.-j.; Wei, J.-f.; Xu, J.-r.; Wang, Y.-h.; Zheng, G.-x. Paper-based three-dimensional microfluidic device for monitoring of heavy metals with a camera cell phone. *Anal. Bioanal. Chem.* **2014**, *406*, 2799–2807. [CrossRef]
- 44. Jayawardane, B.M.; Wei, S.; McKelvie, I.D.; Kolev, S.D. Microfluidic paper-based analytical device for the determination of nitrite and nitrate. *Anal. Chem.* **2014**, *86*, 7274–7279. [CrossRef]
- 45. Park, J.-S.; Park, K.-B.; Shin, K.-S.; Park, H.-D.; Kim, M.-C.; Kim, J.-R.; Park, S.-J.; Song, Y.-H. Design, fabrication and characterization of an integrated micro ammonia analysis system (IMAAS) with microreactor and in-plane type optical detector based on the Berthelot reaction. *Sens. Actuators B Chem.* **2006**, *117*, 516–522. [CrossRef]
- 46. He, S.; Li, D.; Zhu, C.; Song, S.; Wang, L.; Long, Y.; Fan, C. Design of a gold nanoprobe for rapid and portable mercury detection with the naked eye. *Chem. Commun.* **2008**, *40*, 4885–4887. [CrossRef] [PubMed]
- Xiong, Y.; Wang, C.-J.; Tao, T.; Duan, M.; Fang, S.-W.; Zheng, M. A miniaturized fiber-optic colorimetric sensor for nitrite determination by coupling with a microfluidic capillary waveguide. *Anal. Bioanal. Chem.* 2016, 408, 3413–3423. [CrossRef] [PubMed]
- Kanayama, N.; Takarada, T.; Maeda, M. Rapid naked-eye detection of mercury ions based on non-crosslinking aggregation of double-stranded DNA-carrying gold nanoparticles. *Chem. Commun.* 2011, 47, 2077–2079. [CrossRef] [PubMed]
- Yang, R.; Jin, J.; Chen, Y.; Shao, N.; Kang, H.; Xiao, Z.; Tang, Z.; Wu, Y.; Zhu, Z.; Tan, W. Carbon nanotube-quenched fluorescent oligonucleotides: Probes that fluoresce upon hybridization. J. Am. Chem. Soc. 2008, 130, 8351–8358. [CrossRef] [PubMed]
- Xue, H.; Yan, Y.; Hou, Y.; Li, G.; Hao, C. Novel carbon quantum dots for fluorescent detection of phenol and insights into the mechanism. *New J. Chem.* 2018, 42, 11485–11492. [CrossRef]
- Babar, D.G.; Garje, S.S. Nitrogen and phosphorus co-doped carbon dots for selective detection of nitro explosives. ACS Omega 2020, 5, 2710–2717. [CrossRef] [PubMed]
- 52. Yuan, Y.; Jia, H.; Xu, D.; Wang, J. Novel method in emerging environmental contaminants detection: Fiber optic sensors based on microfluidic chips. *Sci. Total Environ.* **2023**, *857*, 159563. [CrossRef] [PubMed]
- 53. Wu, J.; Zheng, G.; Lee, L.M. Optical imaging techniques in microfluidics and their applications. *Lab A Chip* **2012**, *12*, 3566–3575. [CrossRef]
- 54. Orti, V.; Audran, M.; Gibert, P.; Bougard, G.; Bressolle, F. High-performance liquid chromatographic assay for minocycline in human plasma and parotid saliva. *J. Chromatogr. B Biomed. Sci. Appl.* 2000, 738, 357–365. [CrossRef]
- 55. Jaywant, S.A.; Arif, K.M. A comprehensive review of microfluidic water quality monitoring sensors. *Sensors* **2019**, *19*, 4781. [CrossRef]
- 56. Yin, M.-j.; Gu, B.; An, Q.-F.; Yang, C.; Guan, Y.L.; Yong, K.-T. Recent development of fiber-optic chemical sensors and biosensors: Mechanisms, materials, micro/nano-fabrications and applications. *Coord. Chem. Rev.* **2018**, *376*, 348–392. [CrossRef]
- 57. Kuswandi, B.; Huskens, J.; Verboom, W. Optical sensing systems for microfluidic devices: A review. *Anal. Chim. Acta* 2007, 601, 141–155. [CrossRef]
- 58. Zhou, Y.; Zhang, J.F.; Yoon, J. Fluorescence and colorimetric chemosensors for fluoride-ion detection. *Chem. Rev.* **2014**, *114*, 5511–5571. [CrossRef] [PubMed]
- 59. Chemchem, M.; Chemchem, A.; Aydıner, B.; Seferoğlu, Z. Recent advances in colorimetric and fluorometric sensing of neurotransmitters by organic scaffolds. *Eur. J. Med. Chem.* **2022**, 244, 114820. [CrossRef] [PubMed]
- 60. Montes-García, V.; Squillaci, M.A.; Diez-Castellnou, M.; Ong, Q.K.; Stellacci, F.; Samori, P. Chemical sensing with Au and Ag nanoparticles. *Chem. Soc. Rev.* 2021, *50*, 1269–1304. [CrossRef] [PubMed]
- 61. Wang, C.; Yu, C. Detection of chemical pollutants in water using gold nanoparticles as sensors: A review. *Rev. Anal. Chem.* **2013**, 32, 1–14. [CrossRef]
- 62. Cho, H.H.; Jung, D.H.; Heo, J.H.; Lee, C.Y.; Jeong, S.Y.; Lee, J.H. Gold Nanoparticles as Exquisite Colorimetric Transducers for Water Pollutant Detection. *ACS Appl. Mater. Interfaces* **2023**, *15*, 19785–19806. [CrossRef] [PubMed]
- 63. Bala, R.; Sharma, R.K.; Wangoo, N. Highly sensitive colorimetric detection of ethyl parathion using gold nanoprobes. *Sens. Actuators B Chem.* **2015**, *210*, 425–430. [CrossRef]

- 64. Li, D.; Wang, S.; Wang, L.; Zhang, H.; Hu, J. A simple colorimetric probe based on anti-aggregation of AuNPs for rapid and sensitive detection of malathion in environmental samples. *Anal. Bioanal. Chem.* **2019**, *411*, 2645–2652. [CrossRef]
- 65. Khattab, T.A.; Abdelrahman, M.S. Advances in gold nanoparticles for optical detection of nerve agents. In *Sensing of Deadly Toxic Chemical Warfare Agents, Nerve Agent Simulants, and Their Toxicological Aspects*; Elsevier: Amsterdam, The Netherlands, 2023; pp. 111–131.
- 66. Sahu, B.; Kurrey, R.; Deb, M.K.; Khalkho, B.R.; Manikpuri, S. Recognition of malathion pesticides in agricultural samples by using α-CD functionalized gold nanoparticles as a colorimetric sensor. *Talanta* **2023**, *259*, 124526. [CrossRef]
- 67. Chatterjee, S.; Lou, X.-Y.; Liang, F.; Yang, Y.-W. Surface-functionalized gold and silver nanoparticles for colorimetric and fluorescent sensing of metal ions and biomolecules. *Coord. Chem. Rev.* **2022**, 459, 214461. [CrossRef]
- 68. Ding, X.; Kong, L.; Wang, J.; Fang, F.; Li, D.; Liu, J. Highly sensitive SERS detection of Hg<sup>2+</sup> ions in aqueous media using gold nanoparticles/graphene heterojunctions. *ACS Appl. Mater. Interfaces* **2013**, *5*, 7072–7078. [CrossRef] [PubMed]
- 69. Ullah, N.; Mansha, M.; Khan, I.; Qurashi, A. Nanomaterial-based optical chemical sensors for the detection of heavy metals in water: Recent advances and challenges. *TrAC Trends Anal. Chem.* **2018**, *100*, 155–166. [CrossRef]
- Chaudhary, S.; Umar, A.; Bhasin, K.; Baskoutas, S. Chemical sensing applications of ZnO nanomaterials. *Materials* 2018, 11, 287. [CrossRef] [PubMed]
- Jiang, H. Chemical preparation of graphene-based nanomaterials and their applications in chemical and biological sensors. *Small* 2011, 7, 2413–2427. [CrossRef] [PubMed]
- Liyanage, T.; Qamar, A.Z.; Slaughter, G. Application of nanomaterials for chemical and biological sensors: A review. *IEEE Sens. J.* 2020, 21, 12407–12425. [CrossRef]
- 73. Baptista, F.R.; Belhout, S.A.; Giordani, S.; Quinn, S.J. Recent developments in carbon nanomaterial sensors. *Chem. Soc. Rev.* 2015, 44, 4433–4453. [CrossRef]
- 74. Speranza, G. Carbon nanomaterials: Synthesis, functionalization and sensing applications. Nanomaterials 2021, 11, 967. [CrossRef]
- 75. Ehtesabi, H.; Amirfazli, M.; Massah, F.; Bagheri, Z. Application of functionalized carbon dots in detection, diagnostic, disease treatment, and desalination: A review. *Adv. Nat. Sci. Nanosci. Nanotechnol.* **2020**, *11*, 025017. [CrossRef]
- Chen, B.B.; Liu, M.L.; Li, C.M.; Huang, C.Z. Fluorescent carbon dots functionalization. *Adv. Colloid Interface Sci.* 2019, 270, 165–190. [CrossRef]
- Schroeder, V.; Savagatrup, S.; He, M.; Lin, S.; Swager, T.M. Carbon nanotube chemical sensors. *Chem. Rev.* 2018, 119, 599–663. [CrossRef] [PubMed]
- 78. Norizan, M.N.; Moklis, M.H.; Demon, S.Z.N.; Halim, N.A.; Samsuri, A.; Mohamad, I.S.; Knight, V.F.; Abdullah, N. Carbon nanotubes: Functionalisation and their application in chemical sensors. *RSC Adv.* **2020**, *10*, 43704–43732. [CrossRef] [PubMed]
- 79. Mamun, M.A.A.; Yuce, M.R. Recent progress in nanomaterial enabled chemical sensors for wearable environmental monitoring applications. *Adv. Funct. Mater.* 2020, *30*, 2005703. [CrossRef]
- 80. Bakker, E.; Telting-Diaz, M. Electrochemical sensors. Anal. Chem. 2002, 74, 2781–2800. [CrossRef] [PubMed]
- Hanrahan, G.; Patil, D.G.; Wang, J. Electrochemical sensors for environmental monitoring: Design, development and applications. J. Environ. Monit. 2004, 6, 657–664. [CrossRef] [PubMed]
- Stradiotto, N.R.; Yamanaka, H.; Zanoni, M.V.B. Electrochemical sensors: A powerful tool in analytical chemistry. J. Braz. Chem. Soc. 2003, 14, 159–173. [CrossRef]
- 83. Wu, G.; Zheng, H.; Xing, Y.; Wang, C.; Yuan, X.; Zhu, X. A sensitive electrochemical sensor for environmental toxicity monitoring based on tungsten disulfide nanosheets/hydroxylated carbon nanotubes nanocomposite. *Chemosphere* **2022**, *286*, 131602. [CrossRef]

84. Wanjari, V.P.; Duttagupta, S.P.; Singh, S.P. Dual Linear Range Laser-Induced Graphene-Based Sensor for 4-Nitrophenol Detection in Water. *ACS Appl. Nano Mater.* **2023**, *6*, 11351–11360. [CrossRef]

- 85. Zhu, X.; Zhou, Y.; Guo, Y.; Ren, H.; Gao, C. Nitrogen dioxide sensing based on multiple-morphology cuprous oxide mixed structures anchored on reduced graphene oxide nanosheets at room temperature. *Nanotechnology* **2019**, *30*, 455502. [CrossRef]
- Yoon, H.J.; Yang, J.H.; Zhou, Z.; Yang, S.S.; Cheng, M.M.-C. Carbon dioxide gas sensor using a graphene sheet. *Sens. Actuators B Chem.* 2011, 157, 310–313. [CrossRef]
- 87. Shaikshavali, P.; Reddy, T.M.; Palakollu, V.; Karpoormath, R.; Rao, Y.S.; Venkataprasad, G.; Gopal, T.V.; Gopal, P. Multi walled carbon nanotubes supported CuO-Au hybrid nanocomposite for the effective application towards the electrochemical determination of acetaminophen and 4-aminophenol. *Synth. Met.* **2019**, 252, 29–39. [CrossRef]
- 88. Castañeda, M.e.T.; Alegret, S.; Merkoci, A. Electrochemical sensing of DNA using gold nanoparticles. *Electroanal. Int. J. Devoted Fundam. Pract. Asp. Electroanal.* 2007, 19, 743–753. [CrossRef]
- 89. Zhao, Z.; Sun, Y.; Li, P.; Zhang, W.; Lian, K.; Hu, J.; Chen, Y. Preparation and characterization of AuNPs/CNTs-ErGO electrochemical sensors for highly sensitive detection of hydrazine. *Talanta* **2016**, *158*, 283–291. [CrossRef] [PubMed]
- 90. de Lima, C.A.; Santana, E.R.; Piovesan, J.V.; Spinelli, A. Silver nanoparticle-modified electrode for the determination of nitro compound-containing pesticides. *Anal. Bioanal. Chem.* **2016**, *408*, 2595–2606. [CrossRef] [PubMed]
- 91. Ibrahim, A.A.; Kumar, R.; Umar, A.; Kim, S.; Bumajdad, A.; Ansari, Z.; Baskoutas, S. Cauliflower-shaped ZnO nanomaterials for electrochemical sensing and photocatalytic applications. *Electrochim. Acta* **2016**, 222, 463–472. [CrossRef]
- 92. Frontera, P.; Malara, A.; Stelitano, S.; Leonardi, S.G.; Bonavita, A.; Fazio, E.; Antonucci, P.; Neri, G.; Neri, F.; Santangelo, S. Characterisation and H2O2 sensing properties of TiO2-CNTs/Pt electro-catalysts. *Mater. Chem. Phys.* **2016**, *170*, 129–137. [CrossRef]

- Meskher, H.; Ragdi, T.; Thakur, A.K.; Ha, S.; Khelfaoui, I.; Sathyamurthy, R.; Sharshir, S.W.; Pandey, A.; Saidur, R.; Singh, P. A review on CNTs-based electrochemical sensors and biosensors: Unique properties and potential applications. *Crit. Rev. Anal. Chem.* 2023, 1–24. [CrossRef] [PubMed]
- 94. Fu, L.; Zheng, Y.; Li, X.; Liu, X.; Lin, C.-T.; Karimi-Maleh, H. Strategies and Applications of Graphene and Its Derivatives-Based Electrochemical Sensors in Cancer Diagnosis. *Molecules* **2023**, *28*, 6719. [CrossRef]
- 95. Zhu, C.; Yang, G.; Li, H.; Du, D.; Lin, Y. Electrochemical sensors and biosensors based on nanomaterials and nanostructures. *Anal. Chem.* **2015**, *87*, 230–249. [CrossRef]
- 96. Goud, K.Y.; Kailasa, S.K.; Kumar, V.; Tsang, Y.F.; Gobi, K.V.; Kim, K.-H. Progress on nanostructured electrochemical sensors and their recognition elements for detection of mycotoxins: A review. *Biosens. Bioelectron.* **2018**, *121*, 205–222. [CrossRef]
- 97. Power, A.C.; Gorey, B.; Chandra, S.; Chapman, J. Carbon nanomaterials and their application to electrochemical sensors: A review. *Nanotechnol. Rev.* **2018**, *7*, 19–41. [CrossRef]
- Yang, Y.; Yang, X.; Yang, Y.; Yuan, Q. Aptamer-functionalized carbon nanomaterials electrochemical sensors for detecting cancer relevant biomolecules. *Carbon* 2018, 129, 380–395. [CrossRef]
- 99. Deji, R.; Rahul; Choudhary, B.; Sharma, R.K. Role of Graphene-Based Materials in Gas Sensing Applications: From Synthesis to Device Fabrication. In *Handbook of Porous Carbon Materials*; Springer: Berlin/Heidelberg, Germany, 2023; pp. 493–518.
- 100. Krishna Perumal, P.; Chen, C.-w.; Giri, B.S.; Singhania, R.R.; Patel, A.K.; Dong, C.-D. Graphene-based functional electrochemical sensors for the detection of chlorpyrifos in water and food samples: A review. *J. Food Sci. Technol.* **2023**, 1–11. [CrossRef]
- Das, P.K.; Mohanty, C.; Purohit, G.K.; Mishra, S.; Palo, S. Nanoparticle assisted environmental remediation: Applications, toxicological implications and recommendations for a sustainable environment. *Environ. Nanotechnol. Monit. Manag.* 2022, 18, 100679.
- 102. Puri, A.; Loomis, K.; Smith, B.; Lee, J.-H.; Yavlovich, A.; Heldman, E.; Blumenthal, R. Lipid-based nanoparticles as pharmaceutical drug carriers: From concepts to clinic. *Crit. Rev. Ther. Drug Carr. Syst.* **2009**, *26*, 523–580. [CrossRef]
- 103. Murphy, M.; Ting, K.; Zhang, X.; Soo, C.; Zheng, Z. Current development of silver nanoparticle preparation, investigation, and application in the field of medicine. *J. Nanomater.* **2015**, 2015, 696918. [CrossRef]
- 104. Gajbhiye, S.; Sakharwade, S. Silver nanoparticles in cosmetics. J. Cosmet. Dermatol. Sci. Appl. 2016, 6, 48–53. [CrossRef]
- 105. Khan, I.; Saeed, K.; Khan, I. Nanoparticles: Properties, applications and toxicities. Arab. J. Chem. 2019, 12, 908–931. [CrossRef]
- 106. Sakamoto, M.; Fujistuka, M.; Majima, T. Light as a construction tool of metal nanoparticles: Synthesis and mechanism. J. Photochem. Photobiol. C Photochem. Rev. 2009, 10, 33–56. [CrossRef]
- 107. Sajid, M.; Płotka-Wasylka, J. Nanoparticles: Synthesis, characteristics, and applications in analytical and other sciences. *Microchem. J.* **2020**, *154*, 104623. [CrossRef]
- Zahran, M.; Khalifa, Z.; Zahran, M.A.-H.; Azzem, M.A. Recent advances in silver nanoparticle-based electrochemical sensors for determining organic pollutants in water: A review. *Mater. Adv.* 2021, 2, 7350–7365. [CrossRef]
- 109. Yari, A.; Saidikhah, M. Trithiane silver-nanoparticles-decorated polyaniline nanofibers as sensing element for electrochemical determination of Adenine and Guanine in DNA. *J. Electroanal. Chem.* **2016**, *783*, 288–294. [CrossRef]
- Liu, Q.; Zhang, A.; Wang, R.; Zhang, Q.; Cui, D. A review on metal-and metal oxide-based nanozymes: Properties, mechanisms, and applications. *Nano-Micro Lett.* 2021, 13, 154. [CrossRef] [PubMed]
- 111. Khan, M.; Tahir, M.N.; Adil, S.F.; Khan, H.U.; Siddiqui, M.R.H.; Al-Warthan, A.A.; Tremel, W. Graphene based metal and metal oxide nanocomposites: Synthesis, properties and their applications. *J. Mater. Chem. A* **2015**, *3*, 18753–18808. [CrossRef]
- 112. Ahmed, M.M.; Zhao, R.; Du, J.; Li, J. nanostructural ZnO-based electrochemical sensor for environmental application. *J. Electrochem. Soc.* 2022, 169, 020573. [CrossRef]
- 113. Beitollahi, H.; Tajik, S.; Nejad, F.G.; Safaei, M. Recent advances in ZnO nanostructure-based electrochemical sensors and biosensors. *J. Mater. Chem. B* 2020, *8*, 5826–5844. [CrossRef] [PubMed]
- 114. Shetti, N.P.; Bukkitgar, S.D.; Reddy, K.R.; Reddy, C.V.; Aminabhavi, T.M. ZnO-based nanostructured electrodes for electrochemical sensors and biosensors in biomedical applications. *Biosens. Bioelectron.* **2019**, *141*, 111417. [CrossRef]
- 115. Bhanjana, G.; Dilbaghi, N.; Kumar, R.; Kumar, S. Zinc oxide quantum dots as efficient electron mediator for ultrasensitive and selective electrochemical sensing of mercury. *Electrochim. Acta* 2015, *178*, 361–367. [CrossRef]
- 116. Hu, P.; Chen, L.; Kang, X.; Chen, S. Surface functionalization of metal nanoparticles by conjugated metal-ligand interfacial bonds: Impacts on intraparticle charge transfer. *Acc. Chem. Res.* **2016**, *49*, 2251–2260. [CrossRef]
- 117. Tran, P.H.; Duan, W.; Tran, T.T. Fucoidan-based nanostructures: A focus on its combination with chitosan and the surface functionalization of metallic nanoparticles for drug delivery. *Int. J. Pharm.* **2020**, *575*, 118956. [CrossRef]
- 118. Sapsford, K.E.; Algar, W.R.; Berti, L.; Gemmill, K.B.; Casey, B.J.; Oh, E.; Stewart, M.H.; Medintz, I.L. Functionalizing nanoparticles with biological molecules: Developing chemistries that facilitate nanotechnology. *Chem. Rev.* **2013**, *113*, 1904–2074. [CrossRef]
- Stetter, J.R.; Penrose, W.R.; Yao, S. Sensors, chemical sensors, electrochemical sensors, and ECS. J. Electrochem. Soc. 2003, 150, S11. [CrossRef]
- 120. Banica, F.-G. Chemical Sensors and Biosensors: Fundamentals and Applications; John Wiley & Sons: Hoboken, NJ, USA, 2012.
- Holford, T.R.; Davis, F.; Higson, S.P. Recent trends in antibody based sensors. *Biosens. Bioelectron.* 2012, 34, 12–24. [CrossRef]
  [PubMed]
- Yang, B.; Li, J.; Deng, H.; Zhang, L. Progress of mimetic enzymes and their applications in chemical sensors. *Crit. Rev. Anal. Chem.* 2016, 46, 469–481. [CrossRef] [PubMed]
- 123. Willner, I.; Willner, B. Biomolecule-based nanomaterials and nanostructures. Nano Lett. 2010, 10, 3805–3815. [CrossRef]

- 124. Chen, C.; Wang, J. Optical biosensors: An exhaustive and comprehensive review. *Analyst* **2020**, *145*, 1605–1628. [CrossRef] [PubMed]
- 125. Uniyal, A.; Srivastava, G.; Pal, A.; Taya, S.; Muduli, A. Recent advances in optical biosensors for sensing applications: A review. *Plasmonics* **2023**, *18*, 735–750. [CrossRef]
- Kaur, B.; Kumar, S.; Kaushik, B.K. Recent advancements in optical biosensors for cancer detection. *Biosens. Bioelectron.* 2022, 197, 113805. [CrossRef]
- 127. Reddy, Y.V.M.; Shin, J.H.; Palakollu, V.N.; Sravani, B.; Choi, C.-H.; Park, K.; Kim, S.-K.; Madhavi, G.; Park, J.P.; Shetti, N.P. Strategies, advances, and challenges associated with the use of graphene-based nanocomposites for electrochemical biosensors. *Adv. Colloid Interface Sci.* 2022, 304, 102664. [CrossRef]
- Mohammadpour-Haratbar, A.; Zare, Y.; Rhee, K.Y. Electrochemical biosensors based on polymer nanocomposites for detecting breast cancer: Recent progress and future prospects. *Adv. Colloid Interface Sci.* 2022, 309, 102795. [CrossRef]
- Wang, S.; Liu, Y.; Zhu, A.; Tian, Y. In vivo electrochemical biosensors: Recent advances in molecular design, electrode materials, and electrochemical devices. *Anal. Chem.* 2023, 95, 388–406. [CrossRef]
- 130. Loewenthal, D.; Kamber, D.; Bisker, G. Monitoring the activity and inhibition of cholinesterase enzymes using single-walled carbon nanotube fluorescent sensors. *Anal. Chem.* **2022**, *94*, 14223–14231. [CrossRef] [PubMed]
- Zhong, Z.; Fritzsche, M.; Pieper, S.B.; Wood, T.K.; Lear, K.L.; Dandy, D.S.; Reardon, K.F. Fiber optic monooxygenase biosensor for toluene concentration measurement in aqueous samples. *Biosens. Bioelectron.* 2011, 26, 2407–2412. [CrossRef] [PubMed]
- 132. Atailia, S.; Baraket, A.; Rabai, S.; Benounis, M.; Jaffrezic, N.; Araar, H.; Naït-Bouda, A.; Boumaza, A.; Errachid, A.; Houhamdi, M. Electrochemical urea biosensor based on Proteus mirabilis urease immobilized over polyaniline PANi-Glassy carbon electrode. *Electroanalysis* 2023, 35, e202200502. [CrossRef]
- 133. Liu, S.; Kang, M.; Yan, F.; Peng, D.; Yang, Y.; He, L.; Wang, M.; Fang, S.; Zhang, Z. Electrochemical DNA biosensor based on microspheres of cuprous oxide and nano-chitosan for Hg (II) detection. *Electrochim. Acta* 2015, *160*, 64–73. [CrossRef]
- 134. Mehto, N.K.; Sharma, P.; Kumar, S.; Khanuja, M.; Rawal, R.; Narang, J. Towards papertronics based electrode decorated with zinc oxide nanoparticles for the detection of the yellow fever virus consensus sequence. *Process Biochem.* 2022, 123, 36–43. [CrossRef]
- 135. Wang, H.; Chi, Z.; Cong, Y.; Wang, Z.; Jiang, F.; Geng, J.; Zhang, P.; Ju, P.; Dong, Q.; Liu, C. Development of a fluorescence assay for highly sensitive detection of Pseudomonas aeruginosa based on an aptamer-carbon dots/graphene oxide system. *RSC Adv.* 2018, *8*, 32454–32460. [CrossRef]
- Hu, J.; Fu, K.; Bohn, P.W. Whole-cell Pseudomonas aeruginosa localized surface plasmon resonance aptasensor. *Anal. Chem.* 2018, 90, 2326–2332. [CrossRef]
- 137. Stocker, J.; Balluch, D.; Gsell, M.; Harms, H.; Feliciano, J.; Daunert, S.; Malik, K.A.; Van der Meer, J.R. Development of a set of simple bacterial biosensors for quantitative and rapid measurements of arsenite and arsenate in potable water. *Environ. Sci. Technol.* 2003, *37*, 4743–4750. [CrossRef]
- Yoon, Y.; Kim, S.; Chae, Y.; Jeong, S.-W.; An, Y.-J. Evaluation of bioavailable arsenic and remediation performance using a whole-cell bioreporter. *Sci. Total Environ.* 2016, 547, 125–131. [CrossRef]
- Riether, K.; Dollard, M.-A.; Billard, P. Assessment of heavy metal bioavailability using *Escherichia coli* zntAp::lux and copAp::luxbased biosensors. *Appl. Microbiol. Biotechnol.* 2001, 57, 712–716. [CrossRef] [PubMed]
- 140. Yoon, Y.; Kim, S.; Chae, Y.; Kang, Y.; Lee, Y.; Jeong, S.-W.; An, Y.-J. Use of tunable whole-cell bioreporters to assess bioavailable cadmium and remediation performance in soils. *PLoS ONE* **2016**, *11*, e0154506. [CrossRef] [PubMed]
- Tecon, R.; Beggah, S.; Czechowska, K.; Sentchilo, V.; Chronopoulou, P.-M.; McGenity, T.J.; van der Meer, J.R. Development of a multistrain bacterial bioreporter platform for the monitoring of hydrocarbon contaminants in marine environments. *Environ. Sci. Technol.* 2010, 44, 1049–1055. [CrossRef]
- 142. Jeon, Y.; Lee, Y.; Kim, Y.; Park, C.; Choi, H.; Jang, G.; Yoon, Y. Development of novel Escherichia coli cell-based biosensors to monitor Mn (II) in environmental systems. *Front. Microbiol.* **2022**, *13*, 1051926. [CrossRef] [PubMed]
- 143. Alam, K.K.; Jung, J.K.; Verosloff, M.S.; Clauer, P.R.; Lee, J.W.; Capdevila, D.A.; Pastén, P.A.; Giedroc, D.P.; Collins, J.J.; Lucks, J.B. Rapid, low-cost detection of water contaminants using regulated in vitro transcription. *BioRxiv* 2019, 619296.
- 144. Zhang, P.; Feng, H.; Yang, J.; Jiang, H.; Zhou, H.; Lu, Y. Detection of inorganic ions and organic molecules with cell-free biosensing systems. J. Biotechnol. 2019, 300, 78–86. [CrossRef] [PubMed]
- 145. Nigam, V.K.; Shukla, P. Enzyme based biosensors for detection of environmental pollutants-a review. J. Microbiol. Biotechnol. 2015, 25, 1773–1781. [CrossRef]
- 146. Coronado-Apodaca, K.G.; González-Meza, G.M.; Aguayo-Acosta, A.; Araújo, R.G.; Gonzalez-Gonzalez, R.B.; Oyervides-Muñoz, M.A.; Martínez-Ruiz, M.; Melchor-Martínez, E.M.; Barceló, D.; Parra-Saldívar, R.; et al. Immobilized Enzyme-based Novel Biosensing System for Recognition of Toxic Elements in the Aqueous Environment. *Top. Catal.* 2023, *66*, 606–624. [CrossRef]
- Pundir, C.S.; Chauhan, N. Acetylcholinesterase inhibition-based biosensors for pesticide determination: A review. *Anal. Biochem.* 2012, 429, 19–31. [CrossRef]
- 148. Andreescu, S.; Marty, J.-L. Twenty years research in cholinesterase biosensors: From basic research to practical applications. *Biomol. Eng.* **2006**, 23, 1–15. [CrossRef]
- Economou, A.; Karapetis, S.K.; Nikoleli, G.P.; Nikolelis, D.P.; Bratakou, S.; Varzakas, T.H. Enzyme-Based Sensors. *Adv. Food Diagn.* 2017, 231–250. [CrossRef]
- 150. Wilson, G.S.; Hu, Y. Enzyme-based biosensors for in vivo measurements. Chem. Rev. 2000, 100, 2693–2704. [CrossRef] [PubMed]

- Rocchitta, G.; Spanu, A.; Babudieri, S.; Latte, G.; Madeddu, G.; Galleri, G.; Nuvoli, S.; Bagella, P.; Demartis, M.I.; Fiore, V. Enzyme biosensors for biomedical applications: Strategies for safeguarding analytical performances in biological fluids. *Sensors* 2016, 16, 780. [CrossRef] [PubMed]
- 152. Shi, W.; Yu, X.; Zheng, Y.; Yu, J. DNA based chemical sensor for the detection of nitrogen dioxide enabled by organic field-effect transistor. *Sens. Actuators B Chem.* **2016**, 222, 1003–1011. [CrossRef]
- 153. Schoukroun-Barnes, L.R.; Macazo, F.C.; Gutierrez, B.; Lottermoser, J.; Liu, J.; White, R.J. Reagentless, structure-switching, electrochemical aptamer-based sensors. *Annu. Rev. Anal. Chem.* **2016**, *9*, 163–181. [CrossRef]
- 154. Xu, K.; Purahmad, M.; Brenneman, K.; Meshik, X.; Farid, S.; Poduri, S.; Pratap, P.; Abell, J.; Zhao, Y.; Nichols, B. Design and applications of nanomaterial-based and biomolecule-based nanodevices and nanosensors. *Des. Appl. Nanomater. Sens.* **2014**, *16*, 61–97.
- 155. Dunn, M.R.; Jimenez, R.M.; Chaput, J.C. Analysis of aptamer discovery and technology. Nat. Rev. Chem. 2017, 1, 0076. [CrossRef]
- 156. Chen, Z.; Xie, M.; Zhao, F.; Han, S. Application of nanomaterial modified aptamer-based electrochemical sensor in detection of heavy metal ions. *Foods* **2022**, *11*, 1404. [CrossRef]
- 157. Sargazi, S.; Simge, E.; Mobashar, A.; Gelen, S.S.; Rahdar, A.; Ebrahimi, N.; Hosseinikhah, S.M.; Bilal, M.; Kyzas, G.Z. Aptamerconjugated carbon-based nanomaterials for cancer and bacteria theranostics: A review. *Chem. Biol. Interact.* 2022, 361, 109964. [CrossRef]
- 158. Lou, B.; Liu, Y.; Shi, M.; Chen, J.; Li, K.; Tan, Y.; Chen, L.; Wu, Y.; Wang, T.; Liu, X. Aptamer-based biosensors for virus protein detection. *TrAC Trends Anal. Chem.* 2022, 157, 116738. [CrossRef]
- 159. Kadam, U.S.; Hong, J.C. Recent advances in aptameric biosensors designed to detect toxic contaminants from food, water, human fluids, and the environment. *Trends Environ. Anal. Chem.* **2022**, *36*, e00184. [CrossRef]
- Rahman, M.M.; Li, X.-B.; Lopa, N.S.; Ahn, S.J.; Lee, J.-J. Electrochemical DNA hybridization sensors based on conducting polymers. *Sensors* 2015, 15, 3801–3829. [CrossRef]
- 161. Pham, C.; Stogios, P.J.; Savchenko, A.; Mahadevan, R. Advances in engineering and optimization of transcription factor-based biosensors for plug-and-play small molecule detection. *Curr. Opin. Biotechnol.* **2022**, *76*, 102753. [CrossRef] [PubMed]
- 162. Liu, C.; Yu, H.; Zhang, B.; Liu, S.; Liu, C.-g.; Li, F.; Song, H. Engineering whole-cell microbial biosensors: Design principles and applications in monitoring and treatment of heavy metals and organic pollutants. *Biotechnol. Adv.* 2022, *60*, 108019. [CrossRef]
- 163. Mahr, R.; Frunzke, J. Transcription factor-based biosensors in biotechnology: Current state and future prospects. *Appl. Microbiol. Biotechnol.* **2016**, *100*, 79–90. [CrossRef] [PubMed]
- Mannan, A.A.; Liu, D.; Zhang, F.; Oyarzún, D.A. Fundamental design principles for transcription-factor-based metabolite biosensors. ACS Synth. Biol. 2017, 6, 1851–1859. [CrossRef] [PubMed]
- Harms, H.; Wells, M.C.; Van der Meer, J.R. Whole-cell living biosensors—Are they ready for environmental application? *Appl. Microbiol. Biotechnol.* 2006, 70, 273–280. [CrossRef] [PubMed]
- 166. Elcin, E.; Öktem, H. Whole-cell fluorescent bacterial bioreporter for arsenic detection in water. *Int. J. Environ. Sci. Technol.* **2019**, *16*, 5489–5500. [CrossRef]
- 167. Zhang, L.; Guo, W.; Lu, Y. Advances in cell-free biosensors: Principle, mechanism, and applications. *Biotechnol. J.* **2020**, *15*, 2000187. [CrossRef]
- 168. Jung, J.K.; Alam, K.K.; Verosloff, M.S.; Capdevila, D.A.; Desmau, M.; Clauer, P.R.; Lee, J.W.; Nguyen, P.Q.; Pastén, P.A.; Matiasek, S.J. Cell-free biosensors for rapid detection of water contaminants. *Nat. Biotechnol.* **2020**, *38*, 1451–1459. [CrossRef]
- 169. Tricoli, A.; Neri, G. Miniaturized bio-and chemical-sensors for point-of-care monitoring of chronic kidney diseases. *Sensors* 2018, 18, 942. [CrossRef]
- 170. Sharafeldin, M.; Davis, J.J. Point of care sensors for infectious pathogens. Anal. Chem. 2020, 93, 184–197. [CrossRef] [PubMed]
- 171. Sørensen, S.J.; Burmølle, M.; Hansen, L.H. Making bio-sense of toxicity: New developments in whole-cell biosensors. *Curr. Opin. Biotechnol.* 2006, *17*, 11–16. [CrossRef] [PubMed]
- 172. Jiang, B.; Li, G.; Xing, Y.; Zhang, D.; Jia, J.; Cui, Z.; Luan, X.; Tang, H. A whole-cell bioreporter assay for quantitative genotoxicity evaluation of environmental samples. *Chemosphere* **2017**, *184*, 384–392. [CrossRef] [PubMed]

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