## **Supplemental Section**

## Planar interdigitated aptasensor for flow-through detection of Listeria spp. In hydroponic lettuce growth media

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Table S1. Materials and design dimensions for Pt-IME fabrication.			
Design Element	Dimensions		
Substrate	4 inch SiO <sub>2</sub> with 300 nm thermal oxide layer		
Active area	0.81 cm <sup>2</sup>		
Electrode width	25 µm		
Electrode gap	Variable: 15, 25, 50, 100 $\mu m$ as noted		
Bonding pads	2mm X 2mm		
Ti layer thickness	15 nm		
Pt layer thickness	100 nm		

Table C4 Materials and design din . . .. . : . .. . 

Table S2. Design characteristics of IME with various spacing and measured physical features using Dektak profilometer.

Design Feature	Design Specifications	Actual Dimensions	Percent difference
gap	25 µm	12 to 15	40 to 52
gap	50 μm	35 to 40	20 to 30
gap	100 μm	82 to 87	13 to 18
width	25 µm	22 to 27	8 to 12
Ti/Pt metal layer			
thickness	110 nm	110 to 115	0 to 5

**Table S3**. Summary of electrochemical characterization using ferrocyanide as the redox
 probe. Values are shown as mean  $\pm$  standard deviation (n= 3 independent replicates).

Electrode gap [μm]	Electroactive surface area (ESA) [cm²]	Sensitivity toward H₂O₂ [µA mM⁻¹]	HET Constant [cm s <sup>-1</sup> X10 <sup>-4</sup> ]	Current Density [µA mM <sup>-1</sup> cm <sup>-2</sup> ]
25	$0.04_{a}\pm0.01$	COv	$7.9_{\text{a}}\pm6.4$	COv
50	$0.14_{\rm b}\pm0.02$	$21.3_{\text{a}}\pm0.1$	$34.6_{\rm b}\pm9.1$	$149_{\text{a}}\pm20$
100	$0.11 \text{c} \pm 0.02$	$5.4_{\text{b}}\pm0.1$	$44.2{\scriptstyle b}\pm10.2$	$75_{b}\pm2$

a,b,c means within a column which are not followed by a common subscript are significantly different (p<0.05).

COv = data not stable due to charge overflow

Average ESA was calculated using the mean of oxidation and reduction peaks for three replicate Pt-IMEs.

gap	S*Cs	C*C <sub>C</sub>	H*C <sub>H</sub>	ESA*C <sub>ESA</sub>	I*CI	Z*Cz	Z*C <sub>Ef</sub>	Cumulative
spacing								score
25	2.6	14.9	1.6	4.0	7.1	4.0	9.9	44.0
50	2.6	6.0	6.9	14.0	20.6	2.4	8.8	61.2
100	0.7	3.0	8.8	11.0	4.4	1.1	7.0	36.0

**Table S4.** Portfolio analysis for IME with various gap spacing. Weighting factors are normalized for a maximum score of 100.

 $C_{\rm S}$  = weighting factor for sensitivity toward H<sub>2</sub>O<sub>2</sub> [ $\mu$ A<sup>-1</sup> mM] (0.125);

 $C_c$  = weighting factor for current density [ $\mu A^{-1} m M^{-1} cm^2$ ] (0.040);

 $C_{\rm H}$  = weighting factor for HET constant [cm<sup>-1</sup> s<sup>1</sup> X10<sup>4</sup>] (0.200);

 $C_{ESA}$  = weighting factor for electroactive surface area [cm<sup>-2</sup>] (0.30);

 $C_l$  = weighting factor for peak current [ $\mu A^{-1}$ ] (0.029);

 $C_Z$  = weighting factor for impedance at 1 Hz [ $\Omega^1$ ] (0.025); and

 $C_{Ef}$  = weighting factor for electric field [m V<sup>-1</sup>] (0.003);

**Table S5**. Cleaning electrodes with Piranha solution. Protocol for cleaning and maximum current shown.

			Maximum	
Piranha		scan rate	current	solution
cleaning time	electrode configuration	[mV/s]	[µA]	color
Virgin	3 electrode & sensing not connected	50	183.3	white
10 min	4 electrode w/0 checking	50	400.0	
10 min	4 electrode w/0 checking	50	186.3	white
10 min	the 4-electrode box	50	187.3	yellow
	3 electrode w/ dbl layer			•
10 min	ON	100	225.5	yellow
	3 electrode w/ dbl layer			-
30 min	ON	50	176.5	white
	4 electrode w/0 checking			
30 min	the 4-electrode box	50	182.1	yellow
	3 electrode w/ dbl layer			•
30 min	ON	100	222.8	yellow



**Figure S1**. General design schematic for platinum interdigitated microelectrodes (Pt-IME). Gap spacing (S) design values of 25, 50, and 100  $\mu$ m were used in this study.



**Figure S2**. IME incorporated into particle flow trap for continuous analysis. **A**) Photograph of nylon particle trap with stainless steel filter screen (304 micron, grade 50 mesh). The dimensions of the trap are 3" L x 1.89" W x 2.67" H, and **B**) Photograph of Pt-IME in the particle trap.



**Figure S3**. The gap size of 50 µm electrode array with Dektak profilometer measurement.



**Figure S4. A)** IME model output for various gap spacing (COMSOL). **B)** Comparison of measured and predicted capacitance for various gap spacing in buffer.



Figure S5. A) Estimation of cell constant and electrode spacing for IME B) Olthius plot C) Simulation of electrical field at the surface of IME.



Figure S6. Representative plots of electrochemical characterization fo Ti/Pt IME with different gap spacing. Panels are organized as follows: (A-C) gap spacing of 25 μm, (D-F) gap spacing of 100 μm, (G-I) gap spacing of 100 μm. Top row-Cyclic voltammograms in 4mM K<sub>3</sub>FeCN<sub>6</sub> at room temperature (pH=7.1). Middle row- Randles-Sevcik plots for oxidative and reductive peak current. Bottom row-Nicholson plots for determination of k<sup>0</sup>. Average data for all IMEs is shown in Table S3.



**Figure S7**. Representative DCPA for Pt-IME with 50 (**A**, **B**) and 100 (**C**, **D**)  $\mu$ m gap spacing. Average data for all IMEs is shown in Table S3.



Figure S8: Cartoon representation of secondary structure predicted using mfold. (left) Potential repulsion between base pairs when multiple aptamers are in mirror conformation. (right) Potential hydrogen bonding between the upper stem loop structure when multiple aptamers are in ordered conformation. The bond strength of the thiolmetal (≈40 kcal/mol)at the base tether is significantly higher than the H bonds (≈2 kcal/mol) near the upper stem loop, indicating that any H bonding in the ordered conformation is likely reversible. This dynamic interaction likely plays a role in the measured electrochemical behavior.



**Figure S9**: Cartoon representation of ferricyanide redox probe near the surface of Pt-IME with no aptamers. The redox probe orients near the electrode surface and undergoes oxidation to ferrocyanide via a single potassium ion (a one electron exchange reaction). Water hydrates electrolyte and redox probe within the dielectric layer.



**Figure S10**. Cartoon representation of redox probe used to measure electrochemical behavior. Nucleobase are represented by spheres according to the color legend (drawing not to scale). Near the surface, no nucleotide interactions occur due to base pair repulsion and the 2D tethering of aptamers is assumed to be locally ordered as depicted in the cartoon.



Figure S11. A) Photograph of hydroponic system with Pt-IME. B) Pt-IME incorporated into particle flow trap for continuous analysis.



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