

Article

A Highly Selective and Sensitive Fluorescent Chemosensor for Detecting Al^{3+} Ion in Aqueous Solution and Plant Systems

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Abstract: The solubilized form of aluminum, Al^{3+} , is present under acid soil conditions and toxic to both animals and plants. Detecting and quantifying Al^{3+} is vital for both chemistry and biology. A new Schiff-based fluorescent turn-on sensor (probe L) for the selective detection of the Al^{3+} ion was synthesized by coupling 2-hydroxy-1-naphthaldehyde and 2-aminoisindoline-1,3-dione, and the structure was characterized by nuclear magnetic resonance spectra. The probe L exhibited an excellent selective and sensitive response to the Al^{3+} ion over other metal ions in DMSO- H_2O (1:9 v/v). Fluorescence quantification revealed that probe L was promising for the detection and accumulation of Al^{3+} . Treating rice seedlings with Al^{3+} at 25–200 μM inhibited their growth. Al^{3+} treatment produced reactive oxygen species in rice roots. Practical applications of the fluorescent probe for the quantification of Al^{3+} in water samples and rice seedlings are demonstrated. Detecting the Al^{3+} ion with the probe L is easy and a potential alternative to existing analytical methods. The method can be used for detecting the Al^{3+} content of aqueous solution and plant systems. The novel fluorescent probe L has good potential for monitoring Al^{3+} content in the environment and biological systems.

Keywords: aluminum; rice seedlings; fluorescence

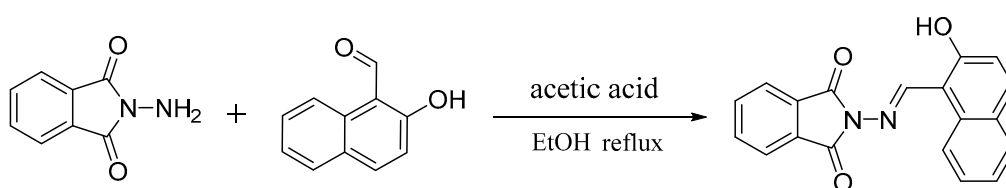
1. Introduction

Aluminum (Al) is the third most abundant element in the earth's crust. With acidic rain and industrial pollution, a trivalent cation of Al^{3+} is solubilized and widely exists in the environment. Approximately 40 to 50% of the world's arable soils are acidic, and the toxicity of Al is considered an important limiting factor of plant productivity on acidic soils [1–3]. Excessive Al content in contaminated soil leads to a decrease in soil microbial content and hampers plant growth [4]. Additionally, Al is widespread in food additives, pharmaceuticals, and storage/cooking utensils. Al^{3+} ions, the solubilized form of Al, enhance the peroxidation of phospholipids and proteins in cell membranes. Al^{3+} ions can move in the human body via food and water sources. Heavy metal intake by human populations through the food chain damages the nervous system and can induce Alzheimers and Parkinsons disease and amyotrophic lateral sclerosis [5–8].

Al in its ionic form, Al^{3+} , is found in most animal and plant tissues and in natural water. Detecting and quantifying Al^{3+} is crucial for controlling its concentration in the biosphere and its direct impact on human health. Various kinds of sensors have been reported for selective Al^{3+} detection [9–26]. However, these sensors are limited by their complicated synthesis, low water solubility, and lack of sensitivity. For practical applications, a water-soluble sensor for detecting Al^{3+} is still in great

demand. Recently, several fluorescent probes have been synthesized for sensing Al^{3+} in cells [27–30]. The use of fluorescent probes for biosensing and bioimaging the ion in animal cells has also been reported. However, these fluorescent probes have not yet been examined for their application in plant systems. The detection of metal ion by fluorescent probes in plant tissues has always been problematic because of interference from the intrinsic fluorescence of the cell wall and secondary metabolites such as tannins, alkaloids, and polyphenols [31,32].

In this study, we designed a fluorescent sensor, probe L (Scheme 1), that showed a selective colorimetric change toward Al^{3+} among the other tested metal ions. To demonstrate the potential of probe L for biological detection, we first analyzed the effect of Al^{3+} exposure on root elongation in rice (*Oryza sativa* L. cv. TN67). The toxicity and accumulation of Al^{3+} were evaluated in rice seedlings and we used probe L to quantify the Al^{3+} level in rice seedlings. The toxicity and accumulation of Al^{3+} inhibited cell elongation and cell division, which led to the stunted growth of roots. Al^{3+} treatment may decrease the uptake of water and nutrients and negatively affect plant growth.



Scheme 1. Synthesis of probe L.

2. Materials and Methods

2.1. Synthesis of Probe L

The synthesis of probe L was based on the following method (Scheme 1). 2-hydroxy-1-naphthaldehyde (0.278 g, 1.61 mmol) was added to a solution of 2-aminoisoindoline-1,3-dione (0.303 g, 1.87 mmol) in dry ethanol (15 mL) with two drops of acetic acid. The reaction mixture was refluxed overnight and then the solvent was removed under reduced pressure. The resulting solid was washed with cooled EtOH repeatedly to give probe L (0.469 g, 80%) as a yellow solid; melting point: 234.0 °C; ^1H NMR (300 MHz, $\text{DMSO}-d_6$) δ 11.95 (s, 1H), 10.07 (s, 1H), 8.62 (d, 1H, $J = 4.4$ Hz), 8.05 (d, 1H, $J = 4.4$ Hz), 7.99–7.90 (m, 5H), 7.66–7.61 (m, 1H), 7.47–7.42 (m, 1H), 7.28 (d, 1H, $J = 4.5$ Hz); ^{13}C NMR (75 MHz, $\text{DMSO}-d_6$) δ 164.33, 159.50, 159.26, 134.99, 131.77, 130.16, 129.04, 128.44, 127.99, 123.94, 123.54, 122.28, 118.67, 108.61; HRMS(EI): calculated for $\text{C}_{19}\text{H}_{12}\text{N}_2\text{O}_3$ ($M + \text{H}$), m/z 316.0848; resulting m/z 316.0839. (Figures S1–S3).

2.2. Plant Materials and Growth Conditions

Rice plants (*Oryza sativa* L. cv. TN67) were grown as described in [33] with minor modification. The seeds were incubated at 37 °C for 3 d in darkness. Ten germinated seeds with coleoptiles reaching 2 mm in length were transferred to a Petri dish (9 cm in diameter) with a filter paper. The rice seedlings were incubated with deionized water supplemented with $\text{Al}(\text{ClO}_4)_3$ (Sigma-Aldrich, St. Louis, MO, USA) from 25 to 200 μM , treated with Al^{3+} solution (10 mL in each Petri dish) for 10 d, then subcultured with fresh solution for another 10 d. Control seedlings were incubated without Al^{3+} . The rice seedlings were analyzed for root elongation and reactive oxygen species (ROS) production. For measuring dry weight and accumulation of Al^{3+} , samples were dried by incubation at 37 °C for 7 d. Data were obtained from at least three biological replicates.

2.3. Use of Probe L to Determine Al Concentration in Rice

Roots of 20-day-old rice seedlings were rinsed thoroughly with deionized water and oven-dried for 2 d at 37 °C. Ten rice seedlings were ground into fine powder and dissolved with 24× deionized water. The extract was mixed with an equal volume of chloroform and then centrifuged at 2000 g for 2 min. Al^{3+} content was determined in the supernatant (270 μL) by supplementation with the fluorescence probe L (30 μL) dissolved in 10% DMSO. Samples of 400 μM probe L were incubated at 2 °C for 20 min. The Al^{3+} -induced specific fluorescence was determined by fluorescence spectrometry (FP-8300, JASCO, Germany) at 375 nm excitation. Data are mean values from three independent biological replicates.

2.4. In Situ Detection of Reactive Oxygen Species (ROS)

Rice roots were treated with various concentrations of $\text{Al}(\text{ClO}_4)_3$ for 14 d. H_2O_2 levels were detected by treating roots with 3,3'-diaminobenzidine (DAB). To detect H_2O_2 , root samples were placed in a solution [10 mM 2-(N-morpholino)ethanesulfonic acid (MES) buffer, pH 3.8] of 1 mg mL^{-1} DAB overnight in darkness. Root samples were rinsed briefly with deionized water. Staining was observed by stereomicroscopy.

3. Results and Discussion

3.1. Absorption and Fluorescence Studies of Probe L Toward Various Metal Ions

We used by UV/vis and fluorescence measurements to investigate the chemosensor behavior of probe L with the following 16 metal ions (as perchlorate salts): Li^+ , Na^+ , K^+ , Ca^{2+} , Mg^{2+} , Mn^{2+} , Hg^{2+} , Fe^{2+} , Fe^{3+} , Co^{2+} , Ni^{2+} , Cu^{2+} , Pb^{2+} , Cd^{2+} , Zn^{2+} , and Al^{3+} in DMSO- H_2O (1:9 v/v). The solution of probe L itself showed two absorption bands at 320 nm and 370 nm, respectively (Figure 1). The addition of Fe^{3+} and Fe^{2+} to the solution enhanced the absorption band. In the presence of Cu^{2+} , the absorption spectrum of probe L was red-shifted, accompanied by a striking color change from dark green to black, which could be seen under UV light (Figure 2). In the presence of Al^{3+} , the intensity of the absorption spectrum of probe L was decreased, accompanied by a striking color change from dark green to blue, which could be detected under UV light (Figure 2). These changes in absorption bands indicated a partial interaction between the ions and probe L. As compared with the weak emission band of probe L alone, the addition of Al^{3+} generated an additional fluorescent-enhanced emission band at 472 nm (Figure 3). The enhanced efficiency at 472 nm was 11-fold greater than the control without Al^{3+} (Figure S4). The fluorescent enhancement may be attributed to the formation of a rigid system after binding with Al^{3+} .

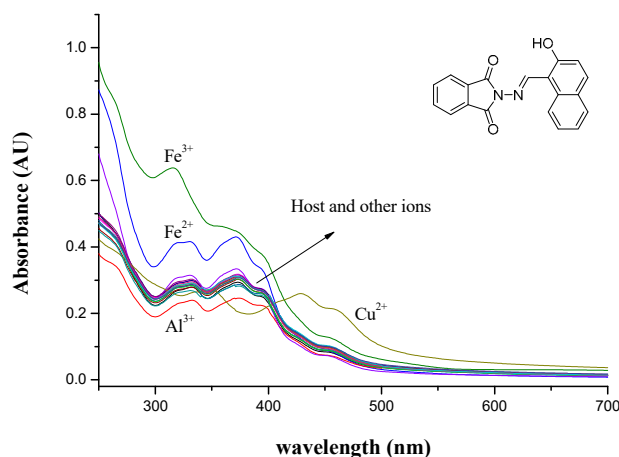


Figure 1. UV/vis spectra for probe L (40 μM) in DMSO- H_2O (1:9 v/v) with the addition of 5 equiv. of various metal ions.

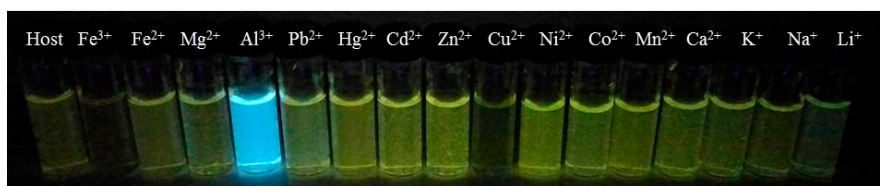


Figure 2. Color changes by UV light for probe L (40 μ M) with the addition of 5 equiv. of various metal ions.

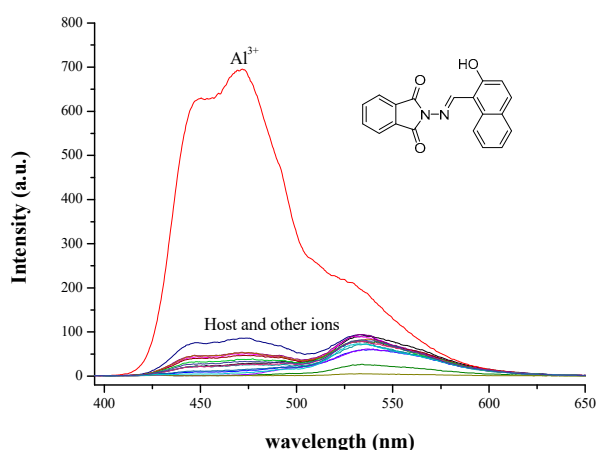


Figure 3. Fluorescence emission spectra for probe L (40 μ M) in the presence of 5 equiv. of various metal ions in DMSO-H₂O (1:9 *v/v*).

3.2. Fluorescence Titration and Binding Studies

To further investigate the chemosensing properties of probe L, probe L was fluorescence-titrated with Al³⁺. The changes in fluorescence spectra for probe L as a function of Al³⁺ content are shown in Figure 4. With increasing amounts of Al³⁺ added to a solution of probe L in DMSO-H₂O (1:9 *v/v*), the emission band at 530 nm gradually increased, and concomitantly a new emission band appeared that peaked at 472 nm. A Job plot indicated a 1:1 stoichiometric complexation of probe L with Al³⁺ (Figure S5). Additionally, the formation of a 1:1 complex between probe L and Al³⁺ was confirmed by the appearance of a peak at *m/z* 393, assignable to [probe L + Al³⁺ + 3H₂O – 4H⁺] in the ESI/MS (Figure S6). From the fluorescent titration profiles, the association constant for probe L-Al³⁺ in DMSO-H₂O (1:9 *v/v*) was determined as 3×10^7 M^{−1} by a Hill plot (Figure S7). By using the abovementioned fluorescence titration results, the detection limit for Al³⁺ was estimated as 1 ppb.

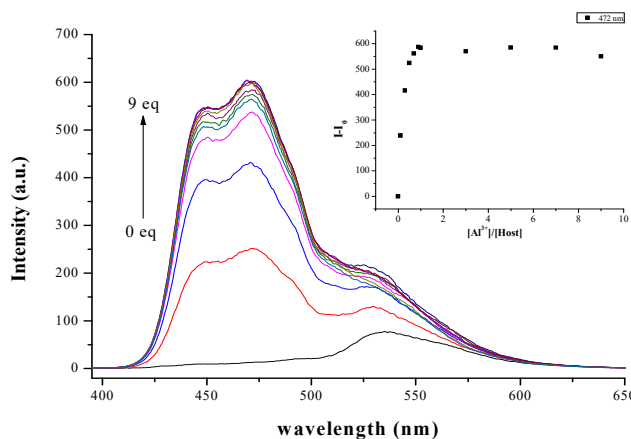


Figure 4. Fluorescence emission spectra for probe L (40 μ M) in DMSO-H₂O (1:9 *v/v*) with increasing concentrations of Al³⁺. Inset: intensity (at 540 nm) for probe L as a function of Al³⁺ equiv.

3.3. Application in Water

In order to determine the optimal pH condition of the probe L in the water, we examined the effect of pH on the emission bands of the probe L–Al³⁺ complex in DMSO–H₂O (1:9 *v/v*) solution. The fluorescence intensity showed a significant emission band (Figure S8) accompanied by a marked color change from dark green to light blue (Figure S9), which could be detected under UV light in the pH range 5–10. Thus, probe L could be applied for the analysis of environmental aqueous samples with a relatively wide pH range.

The practical application of probe L (40 μ M) for selective sensing of Al³⁺ (5 equiv.) in different sources of water (DMSO: H₂O = 1 : 9 *v/v*) was demonstrated (Figure 5). Al(ClO₄)₃ was first dissolved in water of the above source, followed by the addition of probe L to each water sample. All water samples containing Al³⁺ showed a clear color change from dark green to light blue. The result indicated that probe L can detect Al³⁺ in different water sources.

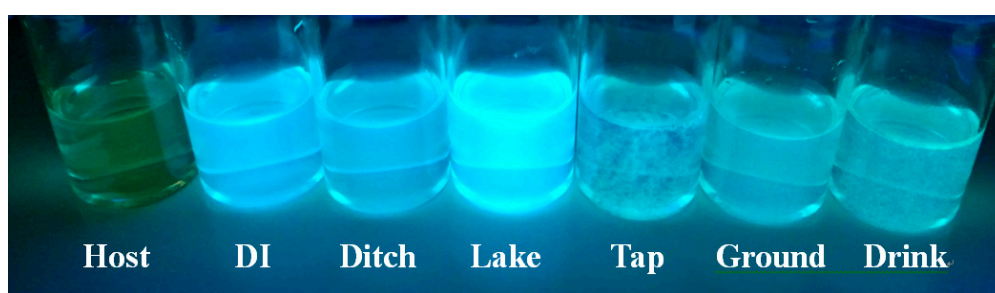


Figure 5. Color changes by UV light for probe L (40 μ M) in different sources of water (DMSO : H₂O = 1 : 9 *v/v*) containing 5 equiv. of Al³⁺.

Testing paper detection is an inexpensive approach to detect the presence of an analyte. Toward this goal, we first coated filter paper with probe L by soaking in a DMSO–H₂O (1:9 *v/v*) solution followed by air drying. Al³⁺ at different concentrations was added to this probe L-coated test paper: the color of the paper changed from dark green to light blue instantly (Figure 6), caused by the interaction of probe L and Al³⁺, therefore implying that the test paper specifically recognized Al³⁺. The detection limit of Al³⁺ with this probe L-coated test paper was 1.0×10^{-5} M.

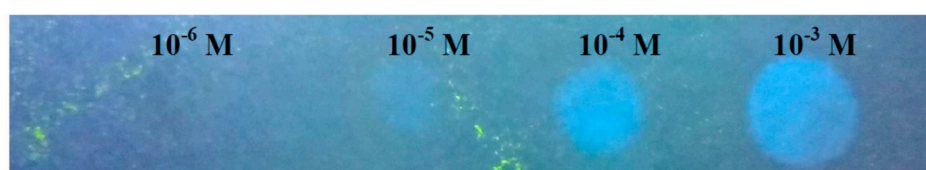


Figure 6. The color changes of probe L observed by UV light upon the addition of Al³⁺ at different concentrations in the filter paper.

3.4. Effect of Al³⁺ Stress on Root Elongation and Oxidative Burst in Rice

The inhibition of root growth is the primary response of the plant exposed to heavy metals. We analyzed the effect of Al³⁺ exposure on root elongation to evaluate Al³⁺ toxicity in rice seedlings. After exposing rice seedlings to Al³⁺ for 14 d, root elongation was measured. At 25 and 50 μ M, Al³⁺ significantly reduced root elongation as compared with control conditions (Figure 7A). The root elongation decreased with increasing Al³⁺ concentration. At 100 μ M Al³⁺, root elongation was reduced to about 50% of the control conditions. At 200 μ M Al³⁺, root growth was completely inhibited. Al³⁺ at 200 μ M inhibited root elongation by approximately 80%. We determined production of ROS such as H₂O₂ by treating roots with DAB. Treatment with 25 and 50 μ M Al³⁺ induced more ROS production compared with control plants (Figure 7B). The ROS level was greater with 100 and 200 μ M Al³⁺ than in controls (Figure 7B). Accordingly, the dry weight of roots was significantly decreased with 100 and

200 μM Al^{3+} (Figure 7C). It was reported that treatment of 200 μM perchlorate had subtle effects on root length and root weight of rice plants [34]. With treatment of 400 μM perchlorate, most of the rice varieties exhibited noticeable toxicity symptoms such as wilting, abscission, and even necrosis [34]. In our study, root weight was markedly inhibited by treatment with 25 μM $\text{Al}(\text{ClO}_4)_3$ (Figure 7C). It is suggested that Al^{3+} resulted in the heavy metal stress in the rice plants. Thus, Al^{3+} -induced ROS production may inhibit growth in rice seedlings (Figure 8B,C). Thereafter, we treated rice seedlings with 25–200 μM Al^{3+} for the quantification of Al^{3+} by the colorimetric fluorescent probe.

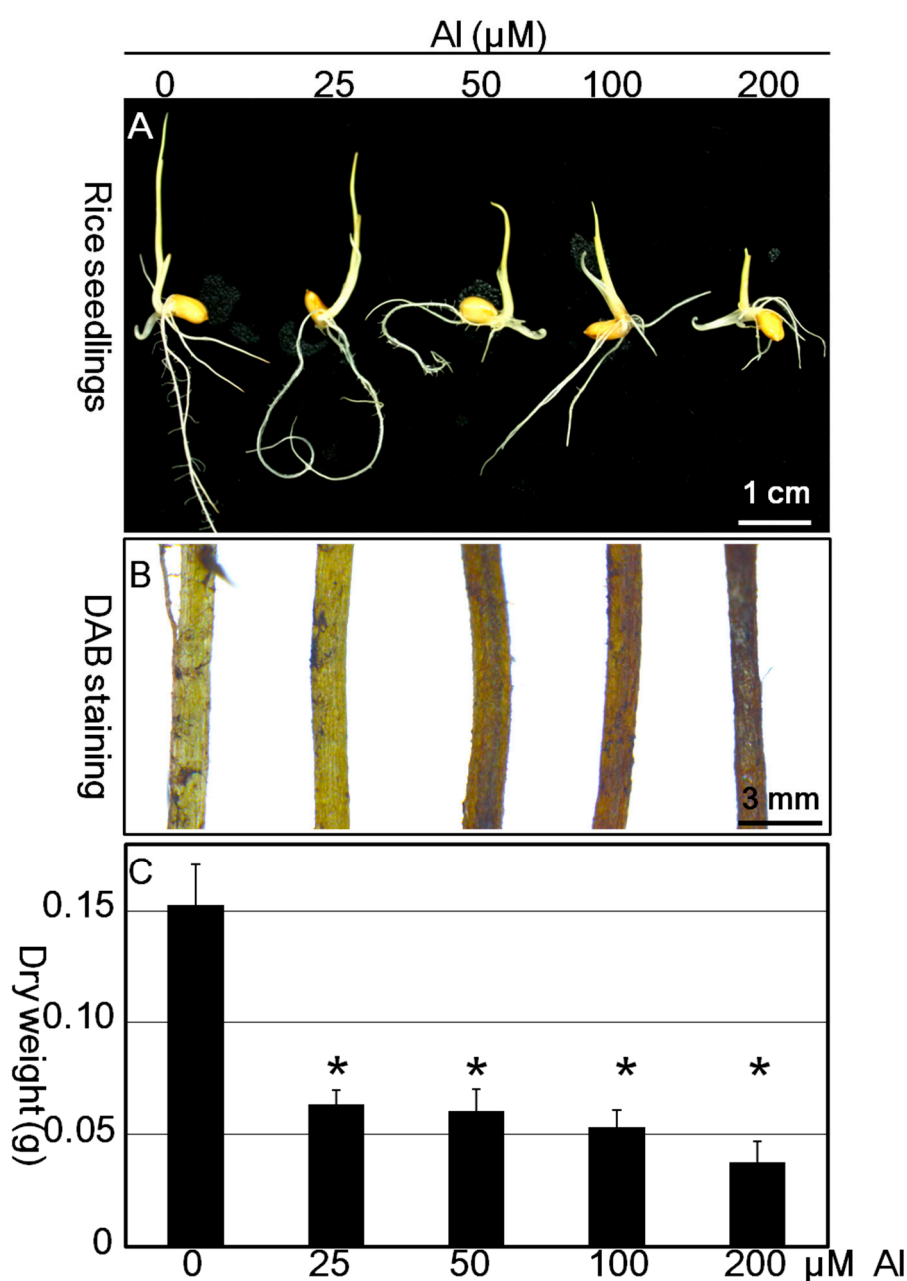


Figure 7. Growth and production of reactive oxygen species (ROS) in rice seedlings with Al treatment. (A) Rice seedlings were treated with different concentrations of $\text{Al}(\text{ClO}_4)_3$ (0, 25, 50, 100, and 200 μM) for 20 d, and the morphology of rice seedlings was photographed. (B) Al-induced ROS accumulation in rice roots. Root samples treated with Al^{3+} were stained with 3, 3'-diaminobenzidine (DAB) for determining H_2O_2 . (C) Dry weight of rice seedlings with treatment of Al. Data represent the mean \pm SD of three independent experiments. Results are the mean \pm SE (n = 10). * Significantly different from the control at $P < 0.05$ by paired t test.

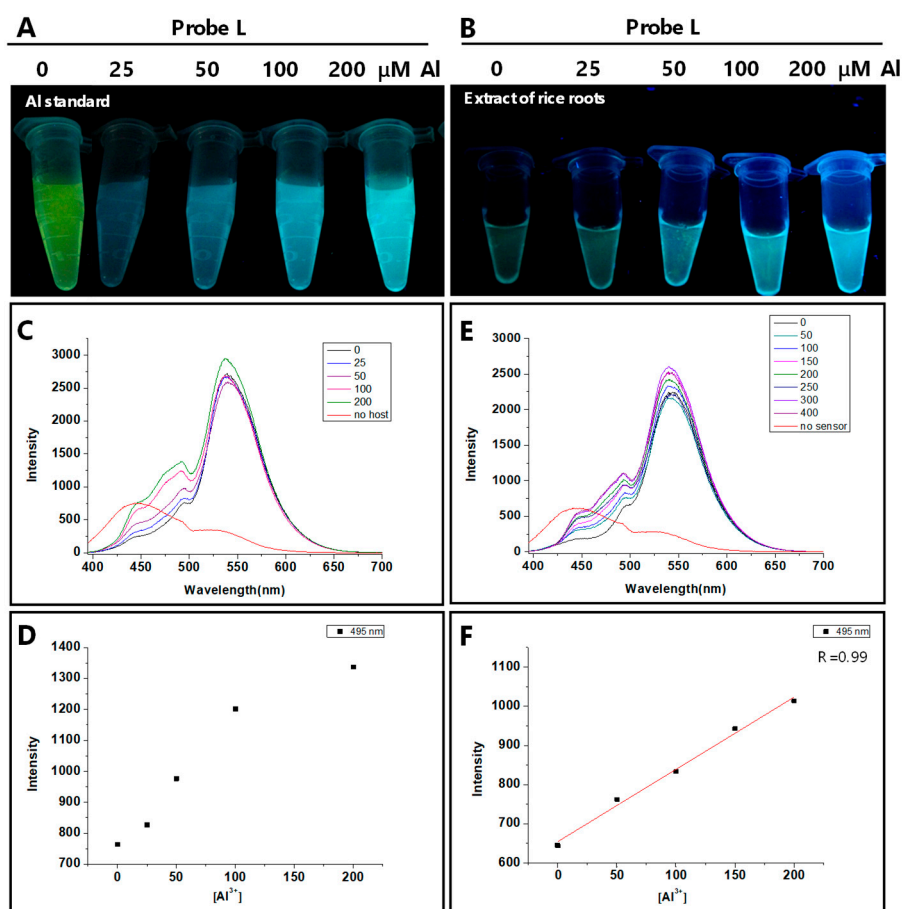


Figure 8. Quantitative determination of Al concentrations in rice seedlings. (A) Fluorescence probe L with the addition of various concentrations of $\text{Al}(\text{ClO}_4)_3$ solution (0, 25, 50, 100, and 200 μM) was excited by a handheld UV lamp with 345 nm excitation. The blue emission was photographed immediately in the dark. (B) Fluorescence probe L with the addition of rice extracts on treatment with various concentrations of Al^{3+} . The samples were excited by UV, and blue emission was photographed. (C) Measurement of fluorescence intensity of probe L at 495 nm with the addition of rice extracts on treatment with various concentrations of Al^{3+} . All data were measured after 20 min. (D) Fluorescence values corresponding to each treatment in (C) were plotted. (E) Measurement of fluorescence intensity of probe L with increasing concentrations of Al^{3+} standards (0, 50, 100, 150, 200, and 300 μM) dissolved in control seedling extracts. (F) A linear calibration curve was constructed on the basis of fluorescence intensity and Al^{3+} standards. Three replicates were performed with nearly identical results.

3.5. Practical Applications of Probe L for Quantifying Al Content in Rice Seedlings

With the excellent response of probe L to Al^{3+} in aqueous solution, we investigated its ability to detect Al^{3+} content in rice seedlings. The intensity of blue fluorescence was increased with increasing Al^{3+} solution standard from 25 to 200 μM (Figure 8A). Extracts of Al^{3+} -treated rice seedlings were incubated with the fluorescent probe. Extracts with 50 μM Al^{3+} showed mild fluorescence. The maximal fluorescence intensity was observed with 200 μM Al^{3+} (Figure 8B). The intensity of fluorescence was further quantified by fluorescence spectroscopy. Fluorescence emission spectra were measured at 495 nm (Figure 8C). Fluorescence intensity was increased with increasing Al^{3+} concentration from 25 to 100 μM (Figure 8D). The highest levels of Al^{3+} occurred with 200 μM Al^{3+} . The stationary phase of Al^{3+} accumulation in rice seedlings may be due to reaching its saturation level (Figure 8D). Therefore, Al^{3+} accumulation in rice was associated with Al^{3+} concentration, with greater accumulation at 200 μM Al^{3+} .

An Al^{3+} standard curve was constructed by linear regression with the Al^{3+} calibration standards in control seedling extracts (Figure 8E). The standard curve showed an acceptable regression line with concentration range 0 to 300 μM (Figure 8F). We then determined Al^{3+} content in 25, 50, 100, and 200 μM -treated samples. The level of heavy metals detected in rice seedlings was 98, 200, 353, and 446 μM , respectively. The differences in Al^{3+} toxicity (Figure 7) may be related to the accumulation in rice (Figure 8). This result agrees with the ratiometric fluorescence changes observed in aqueous solutions. Therefore, the fluorescence probe can be used for the quantification of Al^{3+} content in plants.

4. Conclusions

We developed a new fluorescent probe, probe L, for detecting Al^{3+} content. The probe exhibited rapid, highly selective and sensitive detection of Al^{3+} content, with distinct changes in fluorescent emission. Fluorescent quantification demonstrated that probe L could be used to detect plant samples with heavy metal Al^{3+} pollution. This study provides a promising fluorescent probe for the detection and accumulation of Al^{3+} that could be an easy potential alternative to existing analytical methods for investigating Al^{3+} content in plants.

Supplementary Materials: The following are available online at <http://www.mdpi.com/1424-8220/19/3/623/s1>, Figure S1: ^1H NMR ($\text{DMSO}-d_6$) spectra for probe L, Figure S2: ^{13}C NMR ($\text{DMSO}-d_6$) spectra for probe L, Figure S3: High-resolution mass spectrometry data for probe L, Figure S4: Fluorescence intensity at 536 nm ($\lambda_{\text{ex.}} = 320 \text{ nm}$) for probe L (40 μM) in the presence of 5 equiv. of various cations in $\text{DMSO}-\text{H}_2\text{O}$ (1:9 v/v), Figure S5: Job plot of probe L and Al^{3+} , Figure S6: ESI mass spectra [(probe L + Al^{3+} + DMSO + ClO_4^-) + 2Na^+], Figure S7: Hill plot, Figure S8: Fluorescence emission intensity for probe L (40 μM) in the presence and absence of Al^{3+} (5 equiv) at different pH, Figure S9: Color changes by UV light for probe L with the addition of 5 equiv. of Al^{3+} at different pH value.

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References

- Kochian, L.V.; Hoekenga, O.A.; Pineros, M.A. How do crop plants tolerate acid soils? Mechanisms of aluminum tolerance and phosphorous efficiency. *Annu. Rev. Plant Biol.* **2004**, *55*, 459–493. [[CrossRef](#)] [[PubMed](#)]
- Fageria, N.K.; Carvalho, J.R.P. Influence of aluminum in nutrient solutions on chemical composition in upland rice cultivars. *Plant Soil* **1982**, *69*, 31–44. [[CrossRef](#)]
- Vasconcelos, S.S.; Jacob-Neto, J.; Rossiello, R.O.P. Differential root responses to aluminum stress among Brazilian rice genotypes. *J. Plant Nutr.* **2002**, *25*, 655–669. [[CrossRef](#)]
- Alvarez, E.; Fernandez-Marcos, M.L.; Monterroso, C.; Fernandez-Sanjurjo, M.J. Application of aluminium toxicity indices to soils under various forest species. *For. Ecol. Manag.* **2005**, *211*, 227–239. [[CrossRef](#)]
- Savory, J.; Ghribi, O.; Forbes, M.S.; Herman, M.M. Aluminium and neuronal cell injury: Inter-relationships between neurofilamentous arrays and apoptosis. *J. Inorg. Biochem.* **2001**, *87*, 15–19. [[CrossRef](#)]
- Walton, J.R. Aluminum in hippocampal neurons from humans with Alzheimer's disease. *Neurotoxicology* **2006**, *2*, 385–394. [[CrossRef](#)] [[PubMed](#)]
- Croom, J.; Taylor, I.L. Neuropeptide Y, peptide YY and aluminum in Alzheimer's disease: Is there an etiological relationship? *J. Inorg. Biochem.* **2001**, *87*, 51–56. [[CrossRef](#)]
- Perlmutter, J.S.; Tempel, L.W.; Black, K.J.; Parkinson, D.; Todd, R.D. MPTP induces dystonia and parkinsonism. Clues to the pathophysiology of dystonia. *Neurology* **1997**, *49*, 1432–1438. [[CrossRef](#)] [[PubMed](#)]

9. Hau, F.K.; He, X.M.; Lam, W.H.; Yam, V.W. Highly selective ion probe for Al^{3+} based on $\text{Au(I)} \dots \text{Au(I)}$ interactions in a bis-alkynyl calix[4]arene Au(I) isocyanide scaffold. *Chem. Commun.* **2011**, *47*, 8778–8780. [[CrossRef](#)]
10. Maity, D.; Govindaraju, T. Naphthaldehyde–Urea/Thiourea Conjugates as Turn-On Fluorescent Probes for Al^{3+} Based on Restricted C=N Isomerization. *Eur. J. Inorg. Chem.* **2011**, *36*, 5479–5489. [[CrossRef](#)]
11. Maity, D.; Govindaraju, T. Conformationally Constrained (Coumarin–Triazolyl–Bipyridyl) Click Fluoroionophore as a Selective Al^{3+} Sensor. *Inorg. Chem.* **2010**, *49*, 7229–7231. [[CrossRef](#)] [[PubMed](#)]
12. Lu, Y.; Huang, S.S.; Liu, Y.Y.; He, S.; Zhao, L.C.; Zeng, X.S. Highly Selective and Sensitive Fluorescent Turn-on Chemosensor for Al^{3+} Based on a Novel Photoinduced Electron Transfer Approach. *Org. Lett.* **2011**, *13*, 5274–5277. [[CrossRef](#)] [[PubMed](#)]
13. Jang, Y.K.; Nam, U.C.; Kwon, H.L.; Hwang, I.H.; Kim, C. A selective colorimetric and fluorescent chemosensor based-on naphthol for detection of Al^{3+} and Cu^{2+} . *Dyes Pigm.* **2013**, *99*, 6–13. [[CrossRef](#)]
14. Sahana, A.; Banerjee, A.; Lohar, S.; Sarkar, B.; Mukhopadhyay, S.K. Rhodamine-Based Fluorescent Probe for Al^{3+} through Time-Dependent PET–CHEF–FRET Processes and Its Cell Staining Application. *Inorg. Chem.* **2013**, *52*, 3627–3633. [[CrossRef](#)] [[PubMed](#)]
15. Kim, S.H.; Choi, H.S.; Kim, J.; Lee, S.J.; Quang, D.T.; Kim, J.S. Novel Optical/Electrochemical Selective 1,2,3-Triazole Ring-Appended Chemosensor for the Al^{3+} Ion. *Org. Lett.* **2010**, *12*, 560–563. [[CrossRef](#)] [[PubMed](#)]
16. Ramos, M.L.; Justino, L.L.G.; Salvador, A.I.N.; de Sousa, A.R.E.; Abreu, P.E.; Fonseca, S.M.; Burrows, H.D. NMR, DFT and luminescence studies of the complexation of Al(III) with 8-hydroxyquinoline-5-sulfonate. *Dalton Trans.* **2012**, *41*, 12478–12489. [[CrossRef](#)] [[PubMed](#)]
17. Jung, J.Y.; Han, S.J.; Chun, J.; Lee, C.; Yoon, J. New thiazolothiazole derivatives as fluorescent chemosensors for Cr^{3+} and Al^{3+} . *Dyes Pigm.* **2012**, *94*, 423–426. [[CrossRef](#)]
18. Han, T.Y.; Feng, X.; Tong, B.; Shi, J.B.; Chen, L.; Zhi, J.G.; Dong, Y.P. A novel “turn-on” fluorescent chemosensor for the selective detection of Al^{3+} based on aggregation-induced emission. *Chem. Commun.* **2012**, *48*, 416–418. [[CrossRef](#)]
19. Wang, F.; Nandhakumar, R.; Moon, J.H.; Kim, K.M.; Lee, J.Y.; Yoon, J. Ratiometric Fluorescent Chemosensor for Silver Ion at Physiological pH. *Inorg. Chem.* **2011**, *50*, 2240–2245. [[CrossRef](#)]
20. Mahato, P.; Saha, S.; Suresh, E.; Liddo, R.D.; Parnigotto, P.P.; Conconi, M.T.; Kesharwani, M.K.; Ganguly, B.; Das, A. Ratiometric Detection of Cr^{3+} and Hg^{2+} by a Naphthalimide-Rhodamine Based Fluorescent Probe. *Inorg. Chem.* **2012**, *51*, 1769–1777. [[CrossRef](#)]
21. Xuan, W.; Chen, C.; Cao, Y.; He, W.; Jiang, W.; Liu, K.; Wang, W. Rational design of a ratiometric fluorescent probe with a large emission shift for the facile detection of Hg^{2+} . *Chem. Commun.* **2012**, *48*, 7292–7294. [[CrossRef](#)] [[PubMed](#)]
22. Sun, X.; Wang, Y.-W.; Peng, Y. A Selective and Ratiometric Bifunctional Fluorescent Probe for Al^{3+} Ion and Proton. *Org. Lett.* **2012**, *14*, 3420–3423. [[CrossRef](#)] [[PubMed](#)]
23. Wang, Y.; Xiong, L.; Geng, F.; Zhang, F.; Xu, M. Design of a dual-signaling sensing system for fluorescent ratiometric detection of Al^{3+} ion based on the inner-filter effect. *Analyst* **2011**, *136*, 4809–4814. [[CrossRef](#)] [[PubMed](#)]
24. Goswami, S.; Paul, S.; Manna, A. A differentially selective chemosensor for a ratiometric response to Zn^{2+} and Al^{3+} in aqueous media with applications for molecular switches. *RSC Adv.* **2013**, *3*, 25079–25085. [[CrossRef](#)]
25. Maity, D.; Govindaraju, T. A differentially selective sensor with fluorescence turn-on response to Zn^{2+} and dual-mode ratiometric response to Al^{3+} in aqueous media. *Chem. Commun.* **2012**, *48*, 1039–1041. [[CrossRef](#)] [[PubMed](#)]
26. Goswami, S.; Aich, K.; Das, A.K.; Manna, A.; Das, S. A naphthalimide–quinolone based probe for selective, fluorescence ratiometric sensing of trivalent ions. *RSC Adv.* **2013**, *3*, 2412–2416. [[CrossRef](#)]
27. Sahana, A.; Banerjee, A.; Lohar, S.; Banik, A.; Mukhopadhyay, S.K.; Safin, D.A.; Babashkina, M.G.; Bolte, M.; Garcia, Y.; Das, D. FRET based tri-color emissive rhodamine–pyrene conjugate as an Al^{3+} selective colorimetric and fluorescence sensor for living cell imaging. *Dalton Trans.* **2013**, *42*, 13311–13314. [[CrossRef](#)]
28. Kumar, J.; Sarma, M.J.; Phukan, P.; Das, D.K. A new simple Schiff base fluorescence “on” sensor for Al^{3+} and its living cell imaging. *Dalton Trans.* **2015**, *44*, 4576–4581. [[CrossRef](#)] [[PubMed](#)]

29. Lee, S.A.; You, G.R.; Choi, Y.W.; Jo, H.Y.; Kim, A.R.; Noh, I.; Kim, S.-J.; Kim, Y.; Kim, C. A new multifunctional Schiff base as a fluorescence sensor for Al^{3+} and a colorimetric sensor for CN^- in aqueous media: An application to bioimaging. *Dalton Trans.* **2014**, *43*, 6650–6659. [[CrossRef](#)]
30. Mahapatra, A.K.; Ali, S.S.; Maiti, K.; Manna, S.K.; Maji, R.; Mondal, S.; Uddin, M.R.; Mandal, S.; Sahoo, P. Aminomethylpyrene-based imino-phenols as primary fluorescence switch-on sensors for Al^{3+} in solution and in Vero cells and their complexes as secondary recognition ensembles toward pyrophosphate. *RSC Adv.* **2015**, *5*, 81203–81211. [[CrossRef](#)]
31. Ranatunge, I.; Adikary, S.; Dasanayake, P.; Fernando, C.D.; Soysa, P. Development of a rapid and simple method to remove polyphenols from plant extracts. *Int. J. Anal. Chem.* **2017**, *2017*, 7230145. [[CrossRef](#)] [[PubMed](#)]
32. Abd-Elsalam, K.; Bahkali, A.; Moslem, M.; Amin, O.E.; Niessen, L. An optimized protocol for DNA extraction from wheat seeds and Loop-Mediated Isothermal Amplification (LAMP) to detect *Fusarium graminearum* contamination of wheat grain. *Int. J. Mol. Sci.* **2011**, *12*, 3459–3472. [[CrossRef](#)] [[PubMed](#)]
33. Huang, T.L.; Nguyen, Q.T.T.; Fu, S.F.; Lin, C.Y.; Chen, Y.C.; Huang, H.J. Transcriptomic changes and signalling pathways induced by arsenic stress in rice roots. *Plant Mol. Biol.* **2012**, *80*, 587–608. [[CrossRef](#)]
34. Chen, G.K.; Li, X.B.; He, H.Z.; Li, H.S.; Zhang, Z.M. Varietal differences in the growth of rice seedlings exposed to perchlorate and their antioxidative defense mechanisms. *Environ. Toxicol. Chem.* **2015**, *34*, 1926–1933. [[CrossRef](#)] [[PubMed](#)]



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