



A Novel "Off-On" Fluorescent Probe based on Carbon Nitride Nanoribbons for Detection of Citrate Anion and Live Cell Imaging

Yanling Hu, Dongliang Yang, Chen Yang, Ning Feng, Zhouwei Shao, Lei Zhang, Xiaodong Wang, Lixing Weng, Zhimin Luo, Lianhui Wang



Figure S1. (a) SEM image, (b) XPS survey spectrum, (c) XRD pattern and (d) FTIR spectrum of the bulk C₃N₄.

Figure S1a shows two-dimensional bulk shape of C₃N₄. The XPS survey spectrum indicates that the bulk product contains carbon (283 eV) and nitrogen (397 eV) (Figure S1b). XRD pattern of bulk C₃N₄ (Figure S1c) shows two distinct diffraction peaks at 13.2 and 27.2°, identified as (100) due to the in-plane structural packing feature and (002) due to interlayer stacking of pi-conjugated layers [1,2]. The FTIR spectrum (Figure S1d) presents broad peaks between 3000 and 3400 cm⁻¹ which are associated with the stretching vibrations of N-H groups [3]. Several strong bands of bulk C₃N₄ at 1237, 1321, 1415, 1566 and 1640 cm⁻¹ belong to the typical stretching modes of CN heterocycles [4]. The peak at 808 cm⁻¹ corresponds to the breathing mode of *s*-triazine [3,5].



Figure S2. XPS survey spectrum of C₃N₄ nanoribbons.



Figure S3. C 1s XPS spectrum of C₃N₄ nanoribbons.



Figure S4. FTIR spectrum of C_3N_4 nanoribbons.



Figure S5. Effect of the pH value on the PL intensity of C₃N₄ nanoribbons.



Figure S6. The fluorescence responses of C₃N₄ nanoribbons to various metal ions (Cu²⁺, Al³⁺, Ba²⁺, Co²⁺, Ag⁺, Fe³⁺, K⁺, Li⁺, Mg²⁺, Mn²⁺, Na⁺, Ni²⁺, Pb²⁺, Sn²⁺ and Zn²⁺) at a concentration of 100 μ M in aqueous solution.



Figure S7. The fluorescent changes of $Cu^{2+}-C_3N_4$ nanoribbon complex as a function of interaction time after addition of $C_6H_5O7^{3-}$ (1 mM). The fluorescence intensities were monitored at 415 nm.



Figure S8. The value of fluorescent enhancement (I/I₀) of Cu²⁺-C₃N₄ nanoribbon complex after the addition of C₆H₅O₇³, formic acid, sodium acetate and propionic acid (1 mM). I₀ and I are the fluorescence intensities of Cu²⁺-C₃N₄ nanoribbon complex at 415 nm in the absence and presence of different anions, respectively.



Figure S9. (a) Effect of the pH value on the fluorescence responses of Cu²⁺-C₃N₄ nanoribbon complex after addition of C₆H₅O₇³⁻ (1 mM). (b) Fluorescence responses of Cu²⁺-C₃N₄ nanoribbon complex upon addition of C₆H₅O₇³⁻ and metal ions (10 μ M) mixture. (c) Fluorescence responses of Cu²⁺-C₃N₄ nanoribbon complex upon addition of C₆H₅O₇³⁻ and biological molecule (10 μ M) mixture. (d) Z-scan images of living HeLa cell that preincubated with 1 mM C₆H₅O₇³⁻ for 12 h and then stained with Cu²⁺-C₃N₄ nanoribbon complex for 4 h.



Figure S10. (a) Hydrodynamic size of C_3N_4 nanoribbons. (b) Hydrodynamic size of $Cu^{2+}-C_3N_4$ nanoribbon complex. (c) Hydrodynamic size of C_3N_4 nanoribbons after the $C_6H_5O_7^{3-}$ was added into $Cu^{2+}-C_3N_4$ nanoribbon solution.



Figure S11. The fluorescence responses of C₃N₄ nanoribbons in aqueous solution upon addition of different anions (Br, C₆H₅O₇³⁻, Cl⁻, CN⁻, F⁻, H₂PO₄⁻, HCO₃⁻, I⁻, NO₃⁻, OH⁻, CH₃COO⁻, and SO₄²⁻) (final concentration: 1 mM).



Figure S12. Cell viability of HeLa cells incubated with various concentrations of C₃N₄ nanoribbons (grey) or Cu²⁺-C₃N₄ nanoribbon complex (black) for 24 h.

Materials	Detection limit	Linear range	Response time	Reference
Coumarin	0.10M	0105M	neoponeo unio	[6]
Coumann	0.19 μινι	0.1-0.5 µM	-	[0]
Rhodamine	25 nM	0.1-50 μΜ	1 min	[7]
Diketoprrrolopyrrole	0.18 µM	0–40 µM	-	<u>[8]</u>
CdTe quantum dots	60 nM	0.67–133 μM	5 min	<u>[9]</u>
Boronate derived	10 nM	0~950 nM	15 min	[10]
Carbon nitride	0.78 μM	1–400 µM	20 s	This work

Table S1. Comparison of fluorescent citrate sensors.

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