

Article

An Electrochemical Microsensor Based on a AuNPs-Modified Microband Array Electrode for Phosphate Determination in Fresh Water Samples [†]

Fangfang Wang ^{1,2}, Jianhua Tong ^{1,*}, Yang Li ¹, Chao Bian ¹, Jizhou Sun ¹ and Shanhong Xia ¹

¹ State Key Laboratory of Transducer Technology, Institute of Electronics, Chinese Academy of Sciences, Beijing 100190, China; E-Mails: wffang_89@163.com (F.W.); math101@163.com (Y.L.); cbian@mail.ie.ac.cn (C.B.); sunjizhou628@163.com (J.S.); shxia@mail.ie.ac.cn (S.X.)

² University of Chinese Academy of Sciences, Beijing 100190, China

[†] This paper is an extended version of the paper entitled “Electrochemical Microsensor Based on Gold Microband Array Electrode for Phosphate Determination in Freshwater”, presented at CSMN 2014, Chengdu, China, 31 August 2014.

* Author to whom correspondence should be addressed; E-Mail: jhtong@mail.ie.ac.cn; Tel./Fax: +86-10-5888-7187.

External Editor: W. Rudolf Seitz

Received: 29 October 2014; in revised form: 2 December 2014 / Accepted: 5 December 2014 / Published: 19 December 2014

Abstract: This work describes the fabrication, characterization, and application of a gold microband array electrode (MAE) for the determination of phosphate in fresh water samples. The working principle of this MAE is based on the reduction of a molybdophosphate complex using the linear sweep voltammetric (LSV) method. The calibration of this microsensor was performed with standard phosphate solutions prepared with KH_2PO_4 and pH adjusted to 1.0. The microsensor consists of a platinum counter electrode, a gold MAE as working electrode, and an Ag/AgCl electrode as reference electrode. The microelectrode chips were fabricated by the Micro Electro-Mechanical System (MEMS) technique. To improve the sensitivity, gold nanoparticles (AuNPs) were electrodeposited on the working electrode. With a linear range from 0.02 to 0.50 mg P/L, the sensitivity of the unmodified microsensor is 2.40 μA per (mg P/L) ($R^2 = 0.99$) and that of the AuNPs-modified microsensor is 7.66 μA per (mg P/L) ($R^2 = 0.99$). The experimental

results showed that AuNPs-modified microelectrode had better sensitivity and a larger current response than the unmodified microelectrode.

Keywords: electrochemical; microband array electrode; gold nanoparticles; phosphate; fresh water samples

1. Introduction

Phosphorus is an essential element for organisms in the aqueous environment. Total phosphorus (TP), defined as the sum of orthophosphate, condensed phosphates (pyro-, meta- and polyphosphates) and organically bound phosphorus, plays a key role in the water environment. Excessive TP causes the phenomenon of eutrophication and algae bloom bursts, and reduces dissolved oxygen in lakes and reservoirs [1].

Most TP determination methods include two steps: the first step is a digestion process which turns all forms of phosphorus into orthophosphate, and the second step is the detection process which obtains the concentration of TP by detecting the concentration of orthophosphate in the digested samples. The common techniques of determination of orthophosphate mainly include optical methods [2,3] and electrochemical methods [4]. Optical methods, such as spectrophotometric and chromatographic methods, involve the treatment of the sample with an acidic molybdate solution to produce a phosphomolybdate complex which is further reduced by ascorbic acid to give an intensely colored product. They are prone to interference, unstable or erratic measurements and a lack of selectivity due to the fact that some reagents can produce emission with more than one analyte [5]. Electrochemical methods provide a promising approach to decrease the response time and have less interference from dissolved silicon or turbidity. Additionally, electrochemistry also has the advantages of long lifetime, high precision, low detection limit and good reproducibility. Electrochemical phosphate sensors include biosensors (such as enzyme electrode [6,7] and plant tissue electrodes [8–10]), ion-selective electrodes [11,12] and metal electrodes [13,14]. The determination of phosphate has been performed by voltammetry with carbon paste electrodes [15], gold microdisk electrodes [16], and glassy carbon electrodes [17], and amperometric procedures are also reported [18,19]. Several studies on gold electrodes based on molybdophosphate complex with an electrochemical method for phosphate detection have already been published, however, the current response is very low under the selected voltage using the chronoamperometry method [20–22].

Nanomaterials have been extensively investigated in electrochemistry for sensor fabrication, due to their small size effects, surface effects and quantum tunneling effects. Gold nanoparticles (AuNPs) have good biocompatibility. AuNPs can be prepared by chemical synthesis or electrodeposition techniques. Electrodeposition is rapid, cheap and easily controllable method. The modes of electrodeposition mainly include chronoamperometry [23] and cyclic voltammetry [24]. Chronoamperometry has the advantage of controlling the morphology and the density of the resulting AuNPs [25].

In this work, we use a three-electrode-system to detect phosphate using linear sweep voltammetry. A gold MAE is developed as working electrode by the MEMS technique and AuNPs were

electrodeposited on this gold MAE. SEM images and cyclic voltammtry responses in sulphuric acid were determined for the unmodified and AuNPs modified gold microelectrode. The working principle of this MAE is based on the reduction of the molybdophosphate complex using a linear sweep voltammetric method. The current of the second reduction peak was selected to record the phosphate response. From the experimental results, we can see that the AuNPs-modified microsensor has a good linearity for phosphate. The AuNPs-modified gold microelectrode provides a much larger current response and better sensitivity than the corresponding unmodified gold microelectrode. The characteristics of this AuNPs-modified microsensor could meet the requirements for surface water monitoring and classification.

2. Experimental Section

2.1. Reagents and Equipments

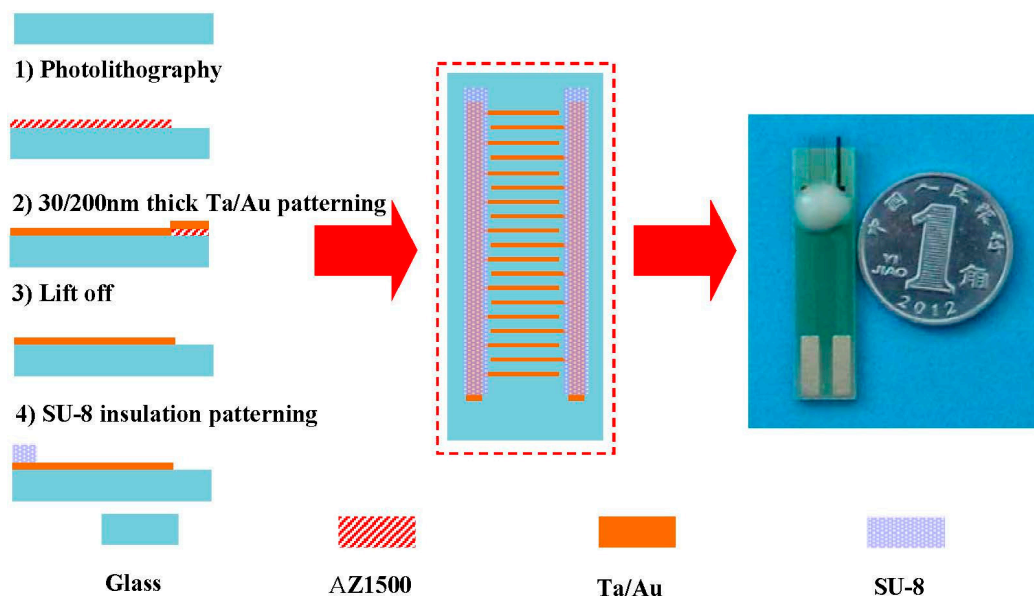
Phosphate standard solutions were prepared with potassium dihydrogen phosphate (KH_2PO_4) (Sinopharm Chemical Reagent, Beijing, China) diluted to different concentrations using ultra-pure water. The molybdate sodium solution (20 mM) was prepared with molybdate sodium (Sinopharm Chemical Reagent, Beijing, China) (Na_2MoO_4) dissolved in ultra-pure water. The detected solutions were mixtures of phosphate standard solution and molybdate sodium solutions in a volumetric proportion of 1:1. Gold deposition solution (2 mM) was prepared with chloroauric acid (Sinopharm Chemical Reagent, Beijing, China) (HAuCl_4) dissolved in 0.5 mol/L sulphuric acid (Sinopharm Chemical Reagent, Beijing, China) (H_2SO_4). The oxidant of digestion was 40 g/L potassium persulfate (Sinopharm Chemical Reagent, Beijing, China) ($\text{K}_2\text{S}_2\text{O}_8$) which was dissolved in ultra-pure water and kept in a brown glass bottle. All the detected solutions were adjusted to pH = 1.0 with 98% sulfuric acid (Sinopharm Chemical Reagent, Beijing, China), and kept in polypropylene bottles. All reagents used were of analytical grade. Calibration and measurements of the sensor were carried out in a 15 mL polypropylene bottle with a three-electrode-system which consists of a platinum plate as counter electrode, a gold MAE as working electrode, and an Ag/AgCl electrode as reference electrode. The electrodeposition of AuNPs on the gold microband array electrode and all the electrical signal generation and detection processes were carried out in the gold deposition solution using a three-electrode system with a Reference-600 electrochemical analyser (Gamry Instruments, PA, USA).

2.2. Electrode Fabrication

The microelectrode chips were fabricated by the MEMS technique. The fabrication process and a photograph of the gold electrodes are shown in Figure 1. The MAE electrode is based on an inter-digitated electrode which consists of two sets of 50 fingers. Each finger is 10 μm in width, 2000 μm in length, and the gap between two adjacent fingers is 50 μm . A 3-inch diameter and 0.5 mm thick glass substrate was first coated with a positive photoresist. After development of the photoresist, metal films consisting of Ta (300 Å) and Au (2000 Å) were deposited by sequential sputtering. The Au layer is used as working electrode and the Ta layer is used as a seed layer as well as an electrical connection between glass substrate and Au layer. The photoresist and the films were removed from the unwanted areas in an acetone bath. After the lift-off process, SU-8 negative photoresist was coated as

insulating layer. The microband array electrode was attached on PCB board for the electrical connection with the electrochemical work station. The modification of AuNPs was performed by the method of chronoamperometry at the potential of -0.3 V for 300 s [26]. Because the microband array electrode is the working electrode, the valid area of the microelectrode is 0.5 mm^2 .

Figure 1. The fabrication process and a photograph of a gold MAE (coin diameter 19 mm).



2.3. Measurement Procedure

The microsensor measurements were carried out with a three-electrode-system which consists of a platinum plate as counter electrode, a gold MAE as working electrode, and an Ag/AgCl electrode as reference electrode in polypropylene bottles. To ensure the stability and consistency of the phosphate determinations, the three electrodes were fixed in a triangular shaped fixture. The distance between counter electrode and working electrode is 1 cm, the distance between counter electrode and reference electrode is 0.5 cm, and the distance between working electrode and reference electrode is 0.8 cm. The microsensor was first cleaned in 0.01 mol/L sulfuric acid (H_2SO_4) solution using cyclic voltammetry (CV) for ten cycles. The CV analyses were performed using applied voltages between 0 and 1.5V with a scan rate of 50 mV/s. The cleaned microsensor was immersed in the detection mixture solutions. The detection was carried out using linear sweep voltammetry from 0.5 V to -0.1 V. The recorded results were the reduction response currents at the applied voltage of -0.07 V vs. the Ag/AgCl reference electrode for different concentration samples.

3. Results and Discussion

3.1. Electrochemical Response Mechanism

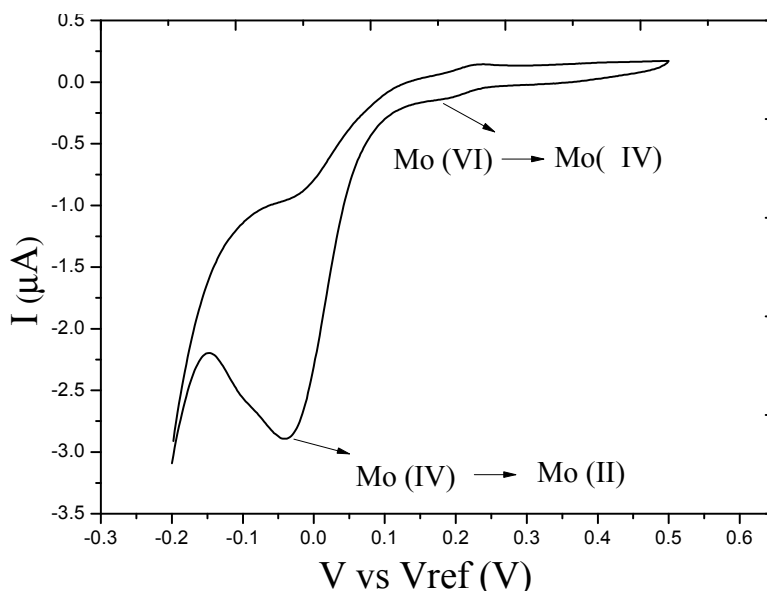
Phosphate is not an electroactive substance, therefore the electrochemical detection of phosphate is based on the formation of a complex formed by its reaction with molybdate in an acidic solution to form a Keggin anion ($\text{PMo}_{12}\text{O}_{40}^{3-}$) according to Reaction (1). In order to ensure that all the phosphates present are involved in the complexation reaction, the concentration of molybdate used in this work is

20 mM, which is in large excess over the reaction stoichiometry ($\sim 10 \mu\text{M}$) [20]. The pH is adjusted to 1.0 with sulfuric acid.



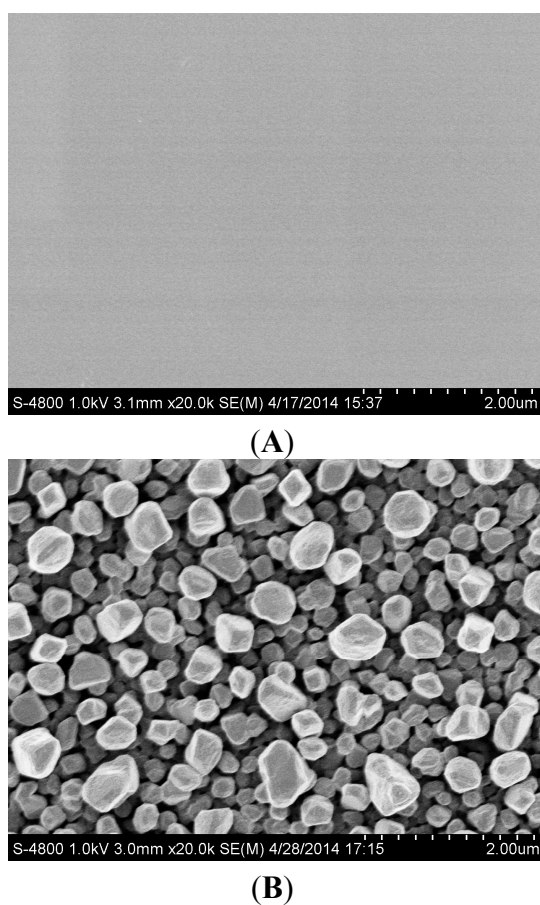
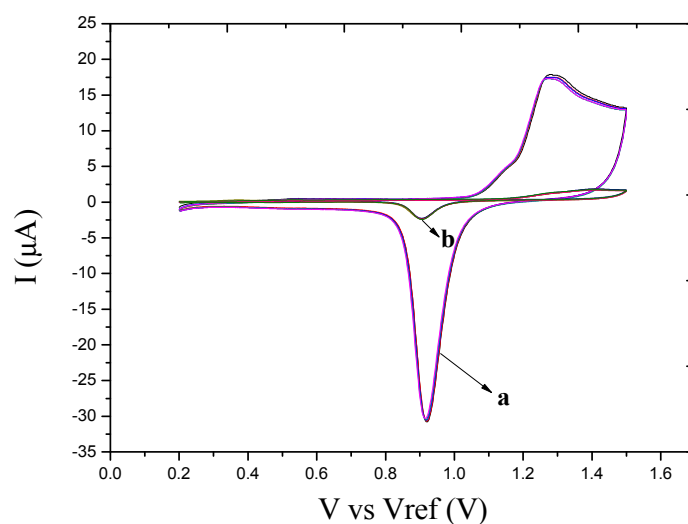
For the CV responses of the molybdophosphate complex, there are two reduction peaks and two oxidation peaks in one cycle. The first reduction peak indicates that Mo (VI) is reduced to Mo (IV), and the second reduction peak is produced by the reduction from Mo (IV) to Mo (II). Figure 2 shows the cyclic voltammetric responses of the molybdophosphate complex with a concentration of 0.40 mg P/L. The peak current value of the reduction wave is related to the concentration of phosphate. Comparing the two reduction peaks, the second peak is much sharper than the first peak. Therefore, to avoid the influence of background current, the phosphate detection voltage was chosen as -0.07 V vs. Ag/AgCl reference electrode, which is at the second reduction peak.

Figure 2. Cyclic voltammetric responses of molybdophosphate complex with an unmodified microelectrode.



3.2. Electrochemical Characterization of AuNPs-Modified Microelectrode

The surface morphology of the electrode was investigated by scanning electronic microscope (SEM). The SEM images of MAE (Figure 3) revealed that the unmodified microelectrode (Figure 3A) showed a two-dimensional flat morphology while the AuNPs-modified electrode (Figure 3B) had a three-dimensional structure and the average diameter of AuNPs was about 500 nm. The cyclic voltammetry responses in 0.01M sulphuric acid are presented in Figure 4. The uniform reduction peaks show that the microsensor was cleaned well and in stable state. The reduction peak current of the modified electrode was 15 times that of the unmodified electrode. This difference results from the mechanism of growth of the three-dimensional crystals.

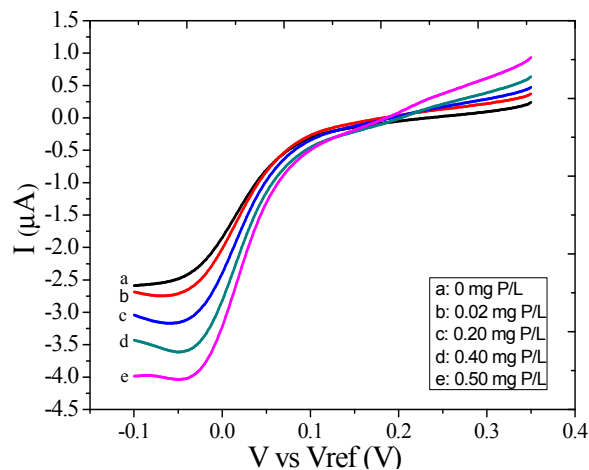
Figure 3. SEM images of bare (A) and AuNPs-modified MAE (B).**Figure 4.** Cyclic voltammetric response in 0.01 M sulfuric acid. **a**: AuNPs-modified microelectrode; **b**: unmodified microelectrode.

3.3. Electrochemical Response and Phosphate Determination

The determination of phosphate was carried out using linear sweep voltammetry from 0.5 V to -0.1 V. The LSV responses of phosphate were recorded with different concentrations from 0.02 to 0.50 mg P/L. The current responses of the unmodified microelectrode are shown in Figure 5. We can see that the

response current decreases with the decrease of voltage and the second reduction peak currents decrease with the increase of phosphate concentration.

Figure 5. LSV responses of different concentrations of phosphate.



The calibration curve in Figure 6(1) shows that the unmodified phosphate microsensor has a linear response to phosphate with a sensitivity of $2.40 \mu\text{A per (mg P/L)}$ ($R^2 = 0.99$) and the AuNPs-modified microsensor in Figure 6(2) has a sensitivity of $7.66 \mu\text{A per (mg P/L)}$ ($R^2 = 0.99$). It is found that the sensitivity of the microelectrode was enhanced three times after the AuNPs modification. The main reason of this phenomenon may be that the AuNPs modification led to a larger area-to-volume on the surface of the gold microelectrode. The larger effective surface may contain a larger number of active sites, which results in an increase of the current density on the surface of the working area, and an improvement of the sensitivity of the microelectrode. The microsensor has a linear range of $0.02\text{--}0.50 \text{ mg P/L}$. The limit of detection (LOD) was calculated as $3 (\text{SD}/S)$, where SD is the standard deviation of the current response of a blank sample, and S is the slope of the calibration curve. The values of LOD using the unmodified electrode and the AuNPs modified electrode are calculated as 12.5 and $41.0 \mu\text{g/L}$, respectively. Table 1 shows the differences between some different gold microsensors for phosphate determination. Compared with chronoamperometry for the detection of phosphate [21,22], this microsensor using LSV method has a much higher current response and less background current influence.

Figure 6. Calibration curve of phosphate standard solution measurement.

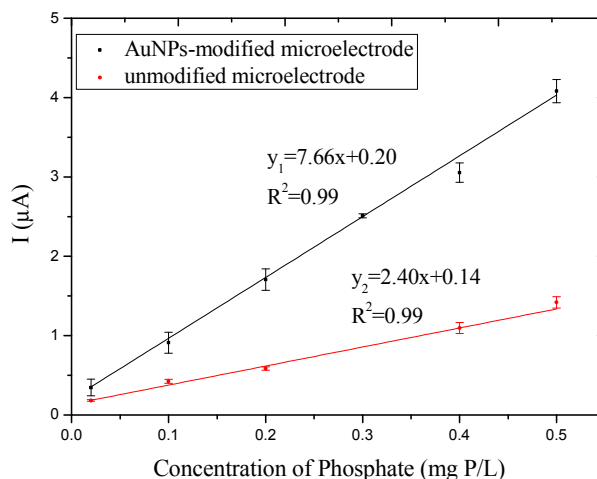


Table 1. The current response sensitivities of different gold microsensor for phosphate detection.

Electrode	Electrode Area (mm ²)	Current Response Sensitivity ($\mu\text{A}/\text{mm}^2 \cdot (\text{mg/L})^{-1}$)	Limit of Detection ($\mu\text{g/L}$)
Unmodified gold electrode in this paper	0.5	4.80	12.5
AuNPs-modified gold electrode in this paper	0.5	15.32	41.0
Jońca [20]	7.065	0.0787	3.72
Bai [21]	0.45	0.0471	20.0
Bai [22]	0.45	0.0638	3.72

3.4. Total Phosphate Determination

To examine the performance of the unmodified gold microelectrode and AuNPs-modified microelectrode for TP determination, a real water sample from the Wanquan River at Tsinghua University was tested. The digestion of this sample was performed using a Lianhua 5B-6P digestion device (Lianhua Tech. Co., Ltd., Lanzhou, China). First, a real sample (8 mL) and oxidant (1 mL) (see Section 2.1) were mixed and sealed in a reaction tube. Then the tube was put into the digestion device and heated at 120 °C for 30 min. After digestion, the sample was cooled to room temperature in a cold-water bath. As a traditional standard method for TP detection, a spectrophotometric method was used to determine the concentration of the real sample. Then, the digested sample was mixed with 20 mM molybdate sodium and detected by the LSV method using the unmodified micro electrode and AuNPs-modified microelectrode, respectively. The measurement results of the real sample are listed in Table 2.

Table 2. The measurement results of a real sample.

Spectrophotometric Method (mg P/L)	Unmodified Microelectrode		AuNPs-Modified Microelectrode	
	Measurement Result (mg P/L)	Recovery (%)	Measurement Result (mg P/L)	Recovery (%)
0.105	0.110	104.76	0.131	124.76
	0.115	109.52	0.124	118.09
	0.114	108.57	0.111	105.71
average	0.113	107.62	0.122	116.19

4. Conclusions/Outlook

In this work, an electrochemical microsensor based on an AuNPs-modified gold microband array electrode was developed for the determination of phosphate in freshwater samples. The working principle of this microsensor is based on the reduction of molybdophosphate complex in pH = 1.0. Under the potential of −70 mV vs. Ag/AgCl reference electrode, the second reduction peak of molybdophosphate complex offers different response currents for different phosphate concentrations. The AuNPs-modified microsensor has a linear response to phosphate with a linear range of 0.02 ~ 0.50 mg P/L. The experimental results showed that AuNPs-modified microelectrode has better sensitivity and a larger current response than the corresponding unmodified microelectrode. In future studies, the sensitivity will

be improved by changing the dimensions and structure of the microband array electrode. This method could be applied in the determination of TP in real water samples.

Acknowledgments

The authors express our deep gratitude to the staff at the State Key Laboratory of Transducers Technology at the Institute of Electronics, Chinese Academy of Sciences for clean room and micro fabrication support. We acknowledge financial support from the National Natural Science Foundation (No. 61372053), and the National High Technology Research and Development Program (863 Program, No. 2012AA040506).

Author Contributions

F.W. conceived the method, performed the design, fabrication and testing of the device, and wrote the paper. J.T. supervised all the research work of F.W. Y.L. supplied help for the device fabrication. B.C. made contributions to the electrochemical analysis. J.S. offered instructions on the modification of the microelectrode. S.X. provided financial support for this work.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. McKelvie, I.D.; Peat, D.; Worsfold, P.J. Analytical perspective. Techniques for the quantification and speciation of phosphorus in natural waters. *Anal. Proc.* **1995**, *32*, 437–445.
2. Shyla, B.; Mahadevaiah; Nagendrappa, G. A simple spectrophotometric method for the determination of phosphate in soil, detergents, water, bone and food samples through the formation of phosphomolybdate complex followed by its reduction with thiourea. *Spectrochim. Acta Part A Mole Biomol. Spectrosc.* **2011**, *78*, 497–502.
3. Bulatov, A.V.; Tsapko, A.A.; Moskvina, L.N. Photometric Cyclic-Injection Determination of Phosphate and Silicate Ions Simultaneously Present in Aqueous Solutions. *J. Anal. Chem.* **2009**, *64*, 580–584.
4. Strickland, J.; Parsons, T. *Practical Handbook of Seawater Analysis*; Fisheries Board of Canada: Ottawa, ON, Canada, 1972; Volume 167, pp. 49–52.
5. Al Law, A.T.; Adeloju, S.B. Progress and recent advances in phosphate sensors: A review. *Talanta* **2013**, *114*, 191–203.
6. Guilbault, G.G.; Nanjo, M.A. Phosphate-selective electrode based on immobilized alkaline phosphatase and glucose oxidase. *Anal. Chim. Acta* **1975**, *78*, 69–80.
7. Menzel, C.; Lerch, T.; Scheper, T.; Schügerl, K. Development of biosensors based on an electrolyte isolator semiconductor (EIS)—capacitor structure and their application for process monitoring. Part I. Development of the biosensors and their characterization. *Anal. Chim. Acta* **1995**, *317*, 259–264.

8. Schubert, F.; Renneberg, R.; Scheller, F.W.; Kirstein, L. Plant tissue hybrid electrode for determination of phosphate and fluoride. *Anal. Chem.* **1984**, *56*, 1677–1682.
9. Linders, C.R.; Vincké, B.J.; Patriarche, G.J. Electrode hybride pour la détermination de phosphates et de polyphosphates. Application aux problèmes liés à l'environnement. *Anal. Lett.* **1985**, *18*, 2195–2208. (In French)
10. Campanella, L.; Cordatore, M.; Mazzei, F.; Tomassetti, M. Determination of inorganic phosphate in drug formulations and biological fluids using a plant tissue electrode. *J. Pharm. Biom. Anal.* **1990**, *8*, 711–716.
11. Zarinskii, V.A.; Shpigun, L.K.; Shkinev, V.M.; Spivakov, B.Y.; Trepalina, V.M.; Zolotov, Y.A. Electrochemical properties of liquid membranes based on dialkyltin (IV) compounds in phosphorus (V) solutions. *Anal. Chem. USSR* **1980**, *35*, 1376–1380.
12. Shkinev, V.M.; Spivakov, B.Y.; Vorob'eva, G.A.; Zolotov, Y.A. Dialkyltin salts as extractants in methods for the determination of arsenic and phosphorus. *Anal. Chim. Acta* **1985**, *167*, 145–160.
13. Xiao, D.; Yuan, H.; Li, J.; Yu, R. Surface-modified cobalt-based sensor as a phosphate-sensitive electrode. *Anal. Chem.* **1995**, *67*, 288–291.
14. Chen, Z.; Marco, D.R.; Alexander, P.W. Flow-injection potentiometric detection of phosphates using a metallic cobalt wire ion selective electrode. *Anal. Commun.* **1997**, *34*, 93–95.
15. Lu, G.; Wu, X. Studies on 1:12 phosphomolybdic heteropoly anion film modified carbon paste electrode. *Talanta* **1999**, *49*, 511–515.
16. Carpenter, N.; Hodgson, A.W.E.; Pletcher, D. Microelectrode procedures for the determination of silicate and phosphate in waters—Fundamental studies. *Electroanalysis* **1997**, *9*, 1311–1317.
17. Matsunaga, T.; Suzuki, T.; Tomoda, R. Photomicrobial sensors for selective determination of phosphate. *Enzyme Microb. Technol.* **1984**, *6*, 355–358.
18. Fogg, A.G.; Bsebsu, N.K. Differential-pulse voltammetric determination of phosphate as molybdovanadophosphate at a glass-carbon electrode and assessment of eluents for the flow-injection voltammetric determination of phosphate, silicon, arsenate and germanate. *Analyst* **1981**, *106*, 1288–1295.
19. Harden, S.M.; Nonidez, W.K. Determination of ortho-phosphate by flow-injection analysis with amperometric detection. *Anal. Chem.* **1984**, *56*, 2218–2223.
20. Jońca, J.; Fernández, V.; Thouron, D.; Paulmier, A.; Graco, M.; Garçon, V. Phosphate determination in seawater: Toward an autonomous electrochemical method. *Talanta* **2011**, *87*, 161–167.
21. Bai, Y.; Tong, J.; Bian, C.; Xia, S. An Electrochemical Microsensor Based on Molybdophosphate Complex for Fast Determination of Total Phosphorus in Water. In Proceedings of the 8th Annual IEEE International Conference on Nano/Micro Engineered and Molecular Systems (NEMS), Suzhou, China, 7–10 April 2013.
22. Bai, Y.; Tong, J.; Wang, J.; Bian, C.; Xia, S. Electrochemical microsensor based on gold nanoparticles modified electrode for total phosphorus determinations in water. *IET Nanobiotechnol.* **2014**, *8*, 31–36.
23. Etesami, M.; Mohamed, N. Catalytic application of gold nanoparticles electrodeposited by fast scan cyclic voltammetry to glycerol electrooxidation in alkaline electrolyte. *Int. J. Electrochem. Sci.* **2011**, *6*, 4676–4689.

24. Ma, Y.; Di, J.; Yan, X.; Zhao, M.; Lu, Z.; Tu, Y. Direct electrodeposition of gold nanoparticles on indium tin oxide surface and its application. *Biosens. Bioelectron.* **2009**, *24*, 1480–1483. Hezard, T.; Fajerweg, K.; Evrard, D.; Collière, V.; Behra, P.; Gros, P. Influence of the gold nanoparticles electrodeposition method on Hg (II) trace electrochemical detection. *Electrochim. Acta* **2012**, *73*, 15–22.
26. Sakai, N.; Fujiwara, Y.; Arai, M.; Yu, K.; Tatsuma, T. Electrodeposition of gold nanoparticles on ITO: Control of morphology and plasmon resonance-based absorption and scattering. *J. Electroanal. Chem.* **2009**, *628*, 7–15.

© 2014 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/4.0/>).