

Article

The Phyloperiodic Approach Removes the “Cryptic Species” and Puts forward Multilevel Organismal Diversity

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Abstract: The notion of the “cryptic species” has recently become an important agenda in biodiversity research. In this study, we show, by applying a periodic-like morphological and molecular framework to the nudibranch genus *Cadlina* from the world’s least explored locations in the Kuril Islands in the northwestern Pacific, including a description of six new species, that the term “cryptic species” should be removed from biodiversity research terminology. We explicitly show that different species of this complex have various degrees of molecular phylogenetic distances and morphological distinctness, revealing a truly multilevel system of fine-scale differences and similarities. Therefore, to designate any of these species as “cryptic” or “non-cryptic” would be completely arbitrary, non-operational, and generally meaningless. By this, we finally strongly propose to remove the notion of “crypticity” and the term “cryptic species” (in the sense of an “indistinguishable species”) from the arsenal of modern biology, including phylogeny and taxonomy. The importance of fine-scale species differentiation in the multilevel framework is shown for addressing conservation and global warming challenges. Six new species are named after scientists who have not always received the honours they deserve, including two women who did not receive their respective Nobel Prizes.

Keywords: fine-scale morphology; molecular analysis; molluscs; multilevel organismal diversity approach



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1. Introduction

Recently, a number of significant contradictions in the use of the term “cryptic species” have been clearly shown [1–9]. Despite this, the term “cryptic species” is still widely used in biodiversity studies [10–28]. It is important to note that the apparent “crypticity” has been indicated for the widest range of groups of organisms, including plants, fungi, animals, and unicellular and multicellular groups, as exemplified by the references above.

In this study, we ultimately show, applying a periodic-like morphological and molecular framework (which has recently been coined as phyloperiodic [29]) to the nudibranch genus *Cadlina* from one of the least explored and hard-to-reach places in the world (the Kuril Islands in the north-western Pacific [30,31]), that the term “cryptic species” should be excluded from the terminology of biodiversity studies. The present *Cadlina* case, where even phylogenetically distant species exhibit a seemingly “almost indistinguishable” pale body with yellow spots and lines [32], is particularly relevant to show that simply the combination of molecular and morphological data, which has previously been applied under various terms, including “integrative taxonomy” [33,34], is not enough and more consistent approaches need to be developed.

Although the periodic approach has not been in common usage in biological taxonomy previously, compelling interdisciplinary evidence has accumulated over the past two decades showing that periodic processes influence the origin of the biological diversity at the genomic [35–37], cellular [38], and morphological levels, including chromatic patterns [39,40]. Thus, the basis of any taxonomic diversity is not only “tree-like” phylogenetic

patterns as such, but also periodic processes at different organismal levels, united by the process of ontogenesis [29]. Recently, it has also been shown that the common phylogenetic “lineage approach” is insufficient for understanding biodiversity [41].

The presence of an apparently huge number of “cryptic species” [42–44], which presumably cannot be assessed and described other than using exclusively genetic data [45–47], greatly hamper the description of biological diversity. It also poses a particular challenge for practitioners, including conservationists, because most of them do not have the ability to conduct molecular analysis in the field. The phyloperiodic approach is particularly relevant for the study of “hidden diversity” and is applied here to facilitate the identification of the particular species within a species complex. It involves the building of phyloperiodic tables according to the central biological periodic process—ontogenesis, which underlies and unites all biological levels from molecular to macromorphological—and therefore provides not just artificially formed, but biologically well founded sets of regularities, “established rules” for the further construction of tables.

Thus, the phyloperiodic approach captures and compares organism-based shared regularities (phyloperiods) at any ontogenetic level (both large-scale, such as earlier ontogenetic patterns, or small-scale, including chromatic variants) with phylogeny-based molecular “lineages” [29]. The molecular phylogenetic data are placed in vertical columns (groups, taxa) and diagnostic morphological data (in a broad ontogenetic sense) are arranged in horizontal rows (periods) in an orderly manner (see below for more details). Due to the multilevel basis of any organisms (=ontogeneses), it is possible to construct many different phyloperiodic tables based on different traits, including commonly used diagnostic morphological characters in a given group.

This study does not simply focus on the “narrow taxonomic group”, but instead uses relevant particular material in order to show that the basic terminology and methodology in the widely important field of biological diversity still needs significant improvement.

2. Materials and Methods

2.1. Sampling

Specimens for this study were collected from the Kuril Islands (NW Pacific) in the course of the two recent expeditions (2021, 2022). Specimens were photographed underwater and measured in the laboratory. Specimens were placed in magnesium sulphate (7%) before preservation in 96% ethanol. The specimens were deposited in the Zoological Museum of Lomonosov Moscow State University (ZMMU).

2.2. Nomenclatural Acts

The electronic version of this article in Portable Document Format (PDF) represents a published work according to the International Commission on Zoological Nomenclature (ICZN), and hence the new names contained in the electronic version are effectively published under that Code from the electronic edition alone. This published work and the nomenclatural acts it contains have been registered in ZooBank, the online registration system for the ICZN. The ZooBank LSIDs (Life Science Identifiers) can be resolved and the associated information viewed through any standard web browser by appending the LSID to the prefix <http://zoobank.org/>. The LSID for this publication is: [urn:lsid:zoobank.org:pub:B33A0BCE-F0A2-4370-A25D-78821AB80BB2].

2.3. Morphological Analysis

The external and internal morphology was studied using a stereomicroscope and digital cameras (Nikon D-810 and Nikon D-80). An amount of 10% sodium hypochlorite solution was used to extract the radula and the jaws. The dried radulae and jaws were coated with gold or palladium and examined using scanning electron microscopes, SEM (CamScan Series II, JSM 6380, and QuattroS). Reproductive systems were examined using a stereomicroscope.

2.4. Molecular Analysis

Thirty-seven specimens of the genus *Cadlina* were newly sequenced for the mitochondrial genes cytochrome c oxidase subunit I (COI), 16S rRNA, and the nuclear 28S rDNA (C1-C2 domain). The DNA extraction procedure, primers, PCR amplification options, and sequencing have been previously described in detail [4,32]. The PCR amplification options are presented in the Supplementary Materials, Table S1. DNA sequences of both strands were obtained using the ABI PRISM1Big-Dye™ Terminator v. 3.1. on an automated DNA sequencer (Applied Biosystems Prism 3700). All new sequences were deposited in GenBank. Moreover, molecular data of *Cadlina* and *Aldisa* species were obtained from GenBank. GenBank accession numbers and references for all sequences used in this study are presented in the Supplementary Materials, Table S2 (newly sequenced data highlighted in bold). Separate analyses were conducted for COI (658 bp), 16S (457 bp), 28S (331 bp), and the concatenated dataset (1446 bp). Sequences were aligned with the MAFFT algorithm [48].

Evolutionary models for each dataset were selected using MEGA11 [49]. The HKY+I+G model was chosen for the COI dataset and for the combined full dataset, the HKY+G model for the 16S dataset, and the GTR model for the 28S dataset. Two different methods of phylogenetic analyses, Bayesian Inference (BI) and Maximum Likelihood (ML), were used. Bayesian estimation of posterior probability was performed in MrBayes 3.2 [50]. Four Markov chains were sampled at intervals of 500 generations; analysis was started with random starting trees and 10,000,000 generations. Maximum Likelihood-based phylogeny inference was performed in RAxML 7.2.8 [51] with bootstrap in 1000 pseudo-replications. Final phylogenetic tree images were rendered in FigTree 1.4.2 (<http://tree.bio.ed.ac.uk>, accessed on 3 February 2023). The Assemble Species by Automatic Partitioning (ASAP) [52] analysis (available at <https://bioinfo.mnhn.fr/abi/public/asap/asapweb.html>, accessed on 3 February 2023) was performed for the COI dataset separately using the models proposed by Jukes–Cantor and Kimura.

The program MEGA11 [49] was used to calculate the uncorrected p-distances. The haplotype networks for the COI molecular data were reconstructed using Population Analysis with Reticulate Trees (PopART, <http://popart.otago.ac.nz>, accessed on 3 February 2023) with the TCS network method.

3. Results

3.1. Molecular Phylogeny

In this molecular study, a total of 90 specimens from the genus *Cadlina* were examined combining 102 novel sequences with 133 sequences from GenBank. Taxon sampling including the outgroup species from the genus *Aldisa* was used according to previous results [32]. Bayesian Inference (BI) and Maximum Likelihood (ML) analyses based on the combined dataset for the mitochondrial COI and 16S and the nuclear 28S genes yielded the same results (Figure 1). All *Cadlina* taxa are clustered in a highly supported monophyletic clade (PP = 1, BS = 100).

The ASAP analysis (asap-score 1.0) of the COI dataset run with two different models revealed 23 lineages: *C. bellburnellae* sp. nov., *C. flavomaculata*, *C. franklinae* sp. nov., *C. jannanicholsae*, *C. japonica*, *C. kamchatica*, *C. klasmalmbergi*, *C. koltzovi* sp. nov., *C. koreana*, *C. laevis*, *C. lomonosovi* sp. nov., *C. luarna*, *C. luteomarginata*, *C. modesta*, *C. paninae*, *C. pellucida*, *C. rumia*, *C. sparsa*, *C. sylviaearleae*, *C. umiushi*, *C. vavilovi* sp. nov., *C. vinogradovi* sp. nov., and *Cadlina* sp. from South Africa.

All six new species from the Kuril Islands formed separate distinct clades [*C. bellburnellae* sp. nov. (PP = 1, BS = 100), *C. franklinae* sp. nov. (PP = 1, BS = 100), *C. koltzovi* sp. nov. (PP = 1, BS = 99), *C. lomonosovi* sp. nov. (PP = 1, BS = 96), *C. vavilovi* sp. nov., and *C. vinogradovi* sp. nov. (PP = 1, BS = 100)] and clustered in a high supported monophyletic clade (PP = 1, BS = 81) together with four species from the North Pacific Ocean [*C. kamchatica* (PP = 1, BS = 100), *C. koreana* (PP = 1, BS = 100), *C. paninae* (PP = 1, BS = 100), and *C. umiushi* (PP = 1, BS = 92)], and one species from the North Atlantic [*C. laevis* (PP = 1, BS = 100)]. Unresolved evolutionary ties were revealed for these species. Whereas *C. koltzovi* sp. nov. and

C. koreana showed close evolutionary relationships, as well as another group *C. franklinae* sp. nov. and *C. paninae*.

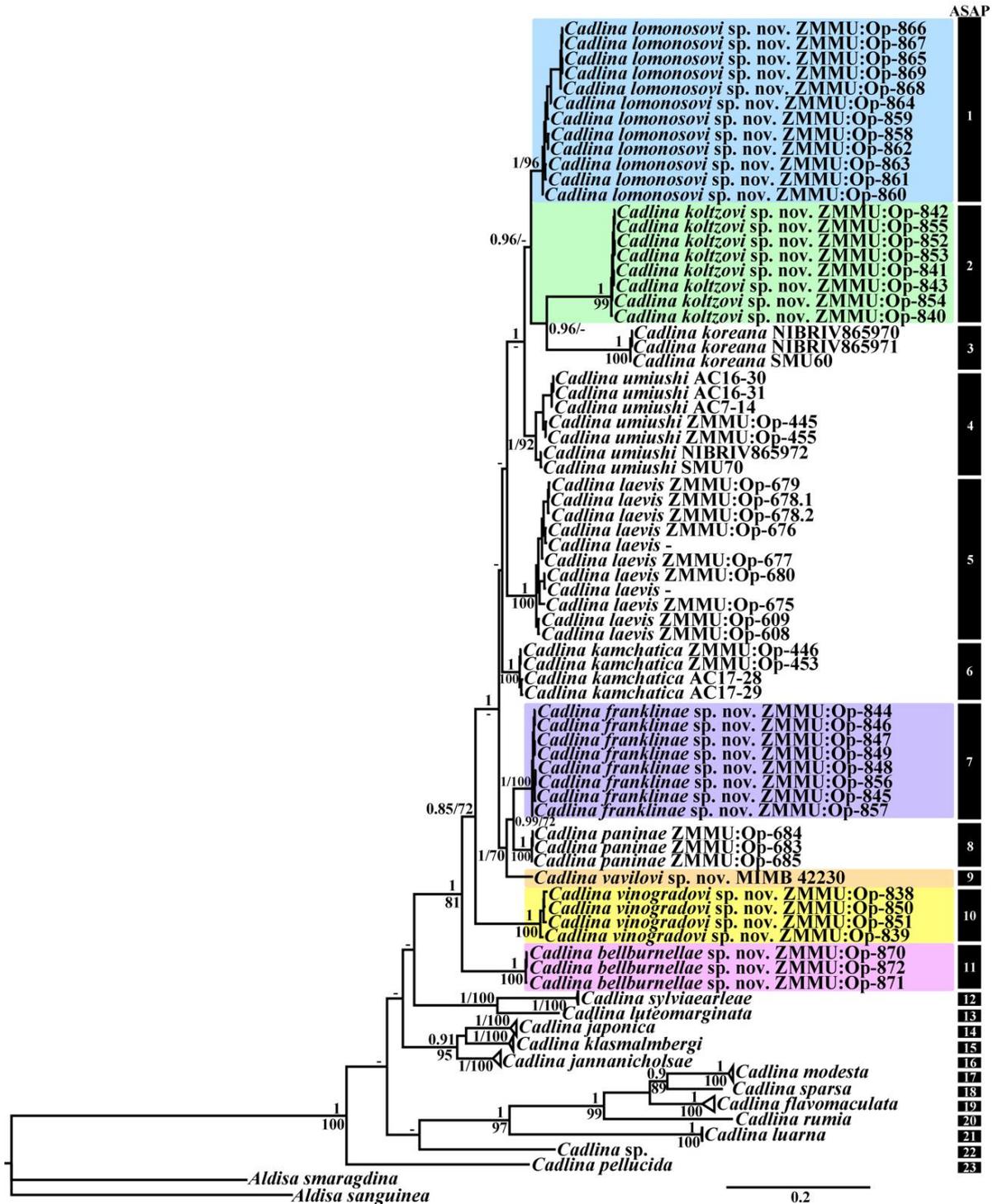


Figure 1. Phylogenetic tree of the genus *Cadlina* based on concatenated molecular data (COI + 16S + 28S) represented by Bayesian Inference. The numbers above branches represent posterior probabilities from Bayesian Inference. The numbers below branches indicate bootstrap values for Maximum Likelihood. Summary of species delimitation results are noted by numbered clusters from the ASAP analyses for the cytochrome c oxidase subunit I (COI) dataset.

The results based on COI molecular data obtained by PopART showed a network of haplotypes that clearly clustered into 11 groups (Figure 2A). The haplotype network results show close evolutionary relationships between *C. franklinae* sp. nov., *C. paninae*, *C. bellburnellae* sp. nov., and *C. vavilovi* sp. nov.

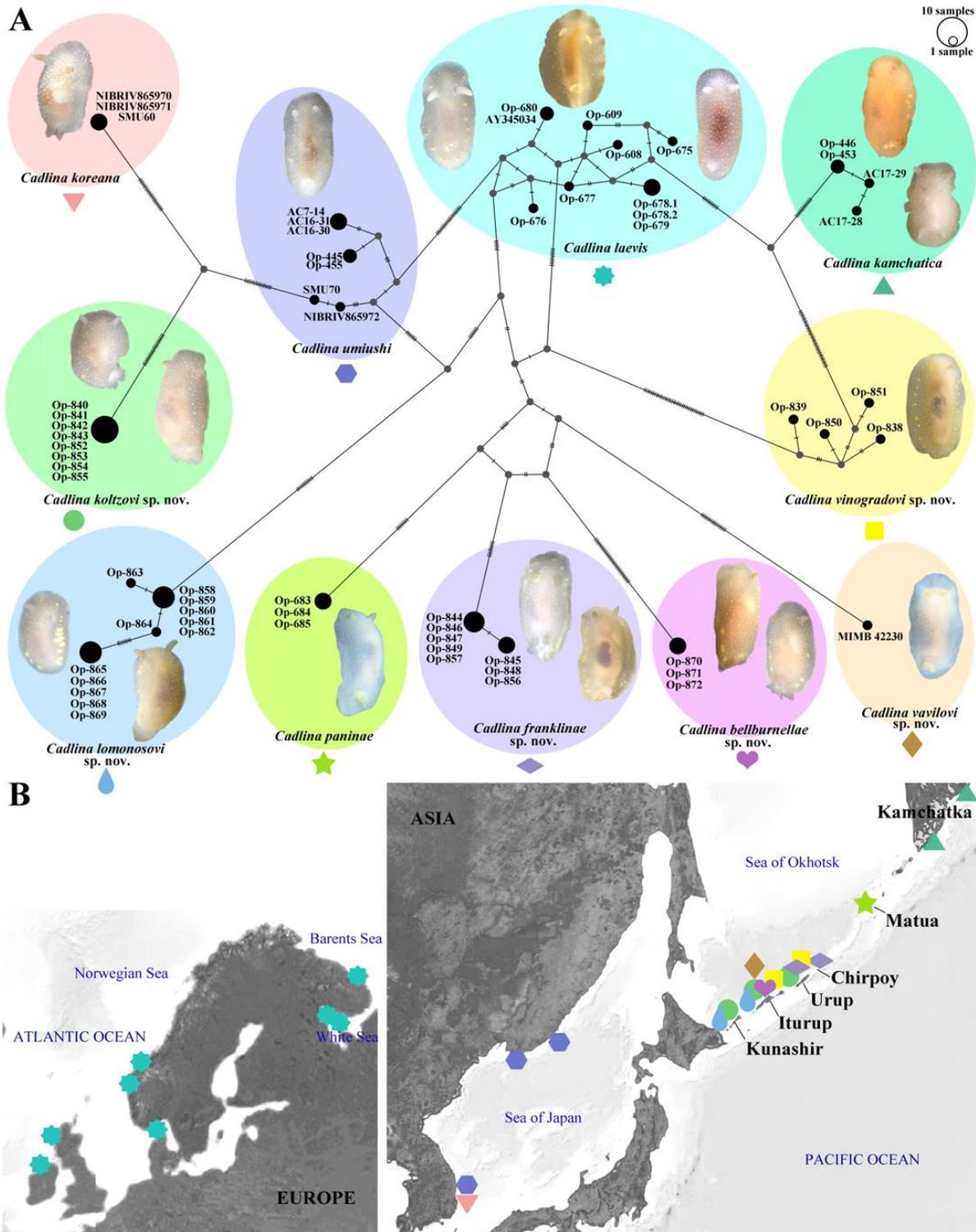


Figure 2. (A) The haplotype network is based on cytochrome c oxidase subunit I (COI) molecular data and shows genetic mutations occurring within eleven *Cadlina* species. (B) Coloured symbols placed on the map denote the *Cadlina* species distribution used in the above analysis.

3.2. Practical Guidelines Proposing How to Perform a Taxonomic Study in the Molecular Era

The following guidelines have been proposed recently [7,53], and we follow them in this study with a little modification: (1) make a selection of a taxonomic group and specimens; (2) conduct a morphological study in a given group, e.g., SEM of previously used diagnostic characters, or application of other methods if available; (3) ensure that ontogenetic information is taken into account during taxonomic assessment (see more details in [29]); (4) perform a relevant bibliographic study of original sources, rather than simply referencing a taxonomic data base; (5) conduct a molecular study of taxonomically checked specimens with appropriate genetic markers; (6) compare the results of morphological (step 2) and taxonomic studies (steps 3, 4) with the molecular (step 5) results; (7) in case inconsistencies are found between commonly used diagnostic characters in a given group and results of molecular phylogeny, respective features should be presented in the phyloperiodic framework, which will enable their detailed comparison and further search for fine-scale differences for each of the closely related taxa; (8) in case difficult to distinguish variants (patterns) are present among the same parallel rows of the phyloperiodic tables, a relevant presentation of diagnostic features should be conducted in order to reveal fine-scale differences between related taxa; (9) a complete study at a given time and using available research possibilities for a given group of taxa should result in fine-scale taxonomic diagnoses for all closely related species in a given taxa/complex (including new taxa); and (10) test the established framework by further investigation of a given group with new materials and data.

3.3. Phyloperiodic Framework for Recognition of Diagnosable Characters in Species Complexes

In the present study, we followed steps 1–6 above using both morphological and molecular data and found that the taxonomic complex represents several apparently difficult-to-distinguish sets of diagnostic characters when the conventional taxonomic comparison format did not work properly. Thus, the phyloperiodic arrangements of several taxonomically reliable characters were used as the next step. The method of the periodic-like presentation includes both tree-like patterns (phylogeny) and ontogenetic (morphological in a broad developmental sense [54]) periodic patterns and was therefore coined phyloperiodic [29]. Accordingly, the general construction of phyloperiodic tables depends, on the one hand, on molecular phylogenetic data ordering the vertical columns (Figures 3–5), and data derived from the ontogenetic model [32,54], which, in turn, orders the horizontal rows (Figures 3–5). It is important to note that in order to use ontogenetic data in a phyloperiodic framework, it is not necessary to obtain ontogenetic data for all species, but it is necessary to formulate an appropriate ontogenetic model for a given high-ranking group (e.g., for the order Doridida in the present case, see more details on the model formulation in [29,32]).

The proposed scheme of chromatic patterns has an ontogenetic basis since it has been shown in *Cadlina*, as well as other disparate members of the nudibranch order Doridida, that during the earlier post-larval ontogenetic development the darker colouration is absent and distinct spots, yellow lines, or darker yellowish or brownish colouration (if any) appear only towards later ontogenetic periods [32,55]. Because any ontogeny encompasses all structures and processes of the organisms (not only those that occurred at the embryonal period), this allows the use of any features for an ontogenetic model, regardless of whether they are earlier embryonal inter-taxon shared nodes (phylotypic periods in a narrow sense) [29,54] or later juvenile–subadult acquisitions of various origin (e.g., chromatic variants, small-scale phyloperiods of the later stages), including potential environmental, food, and other influences. In this regard, the resulting morphology of a particular organism, which has formed during ontogeny and includes both shared and unique features, is important, and any trait of both genetic or epigenetic origin can potentially be represented in phyloperiodic tables.

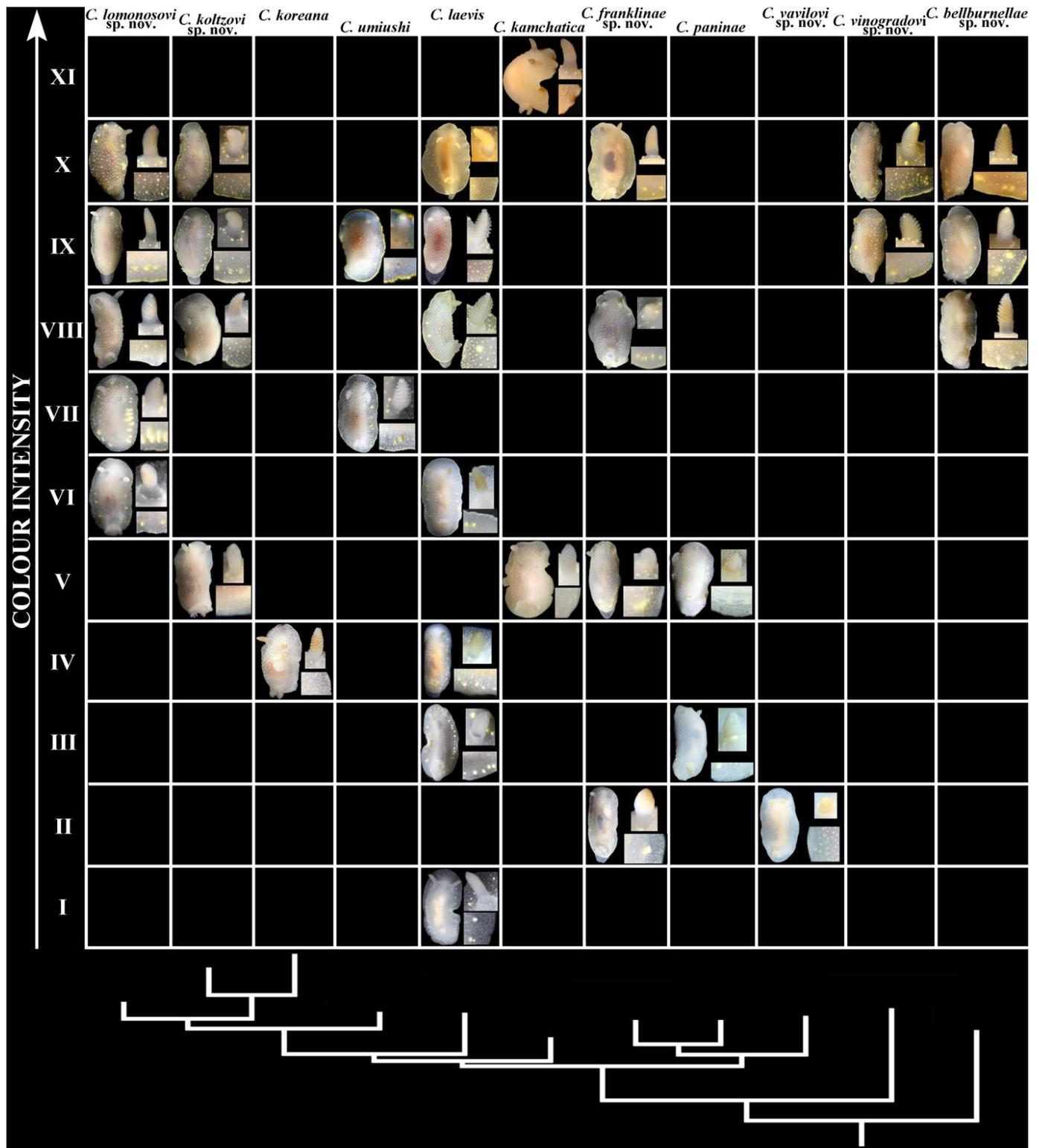


Figure 3. Phyloperiodic presentation of the external chromatic patterns (including dorsal views and details of gills and rhinophores) among species *Cadlina* represented by the horizontal rows. The vertical columns (taxa) are ordered by the molecular phylogenetic tree. Eleven main periods (horizontal rows, Roman numerals) are presented. Spotless body/colourless forms are at the bottom and forms with a maximal number of spots and colour intensity are at the top. Empty cells imply non-observed forms. See text for details.

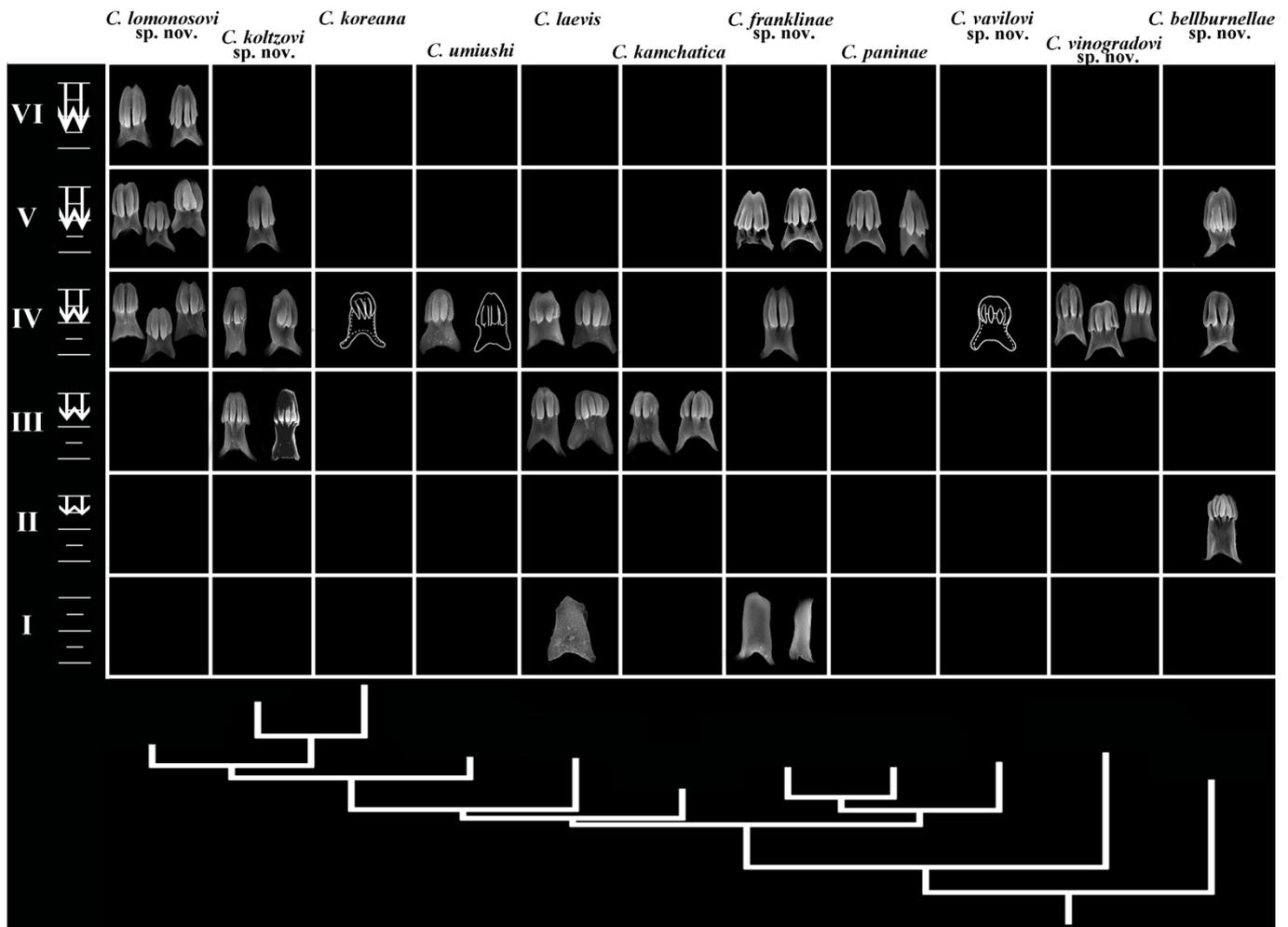


Figure 4. Phyloperiodic presentation of the central teeth of the radula among species *Cadlina*, represented by the horizontal rows. The vertical columns (groups, taxa) are ordered by the molecular phylogenetic tree. Six main periods (horizontal rows, Roman numerals) are presented following the direction from non-denticulate and relatively low teeth to higher teeth, where denticles occupy an increasingly greater part of the teeth's length. Empty cells imply non-observed forms. See text for details.

The central radular teeth and copulative spines, which range from simple non-cuspidate or low cuspidate teeth or rounded plate-like spines to more elongated and elaborated ones, also correspond to the ontogenetic model [32,55]. Therefore, for taxonomic purposes, we present here a phyloperiodic framework for three main sets of commonly used *Cadlina* diagnostic characters in three different organ systems, including external patterns (colouration, gills, and rhinophoral lamellae patterns), radular details (central teeth have been proven to be the best diagnostic feature in the radula of *Cadlina*) in the digestive system, and details of copulative spines in the reproductive system [32]. Only specimens for which molecular data have been obtained (Figures 3–5) are included in the phyloperiodic tables to ensure that any morphological features can be matched with the molecular phylogenetic tree (Figure 1). Under this approach, diagnostic characters in respective periods are presented not as strict diagnoses, but as a complex range (Figures 3–5). The premise of the phyloperiodic approach is precisely that the traditional “rigid” diagnosis will be gradually replaced by multidimensional “soft” or “complex” diagnoses that therefore go beyond the current conventional taxonomic rules and contribute to the formation of future biological nomenclature. The construction of the phyloperiodic

tables and the placement of some characters in a certain cell already indicate potential “complex” diagnoses.

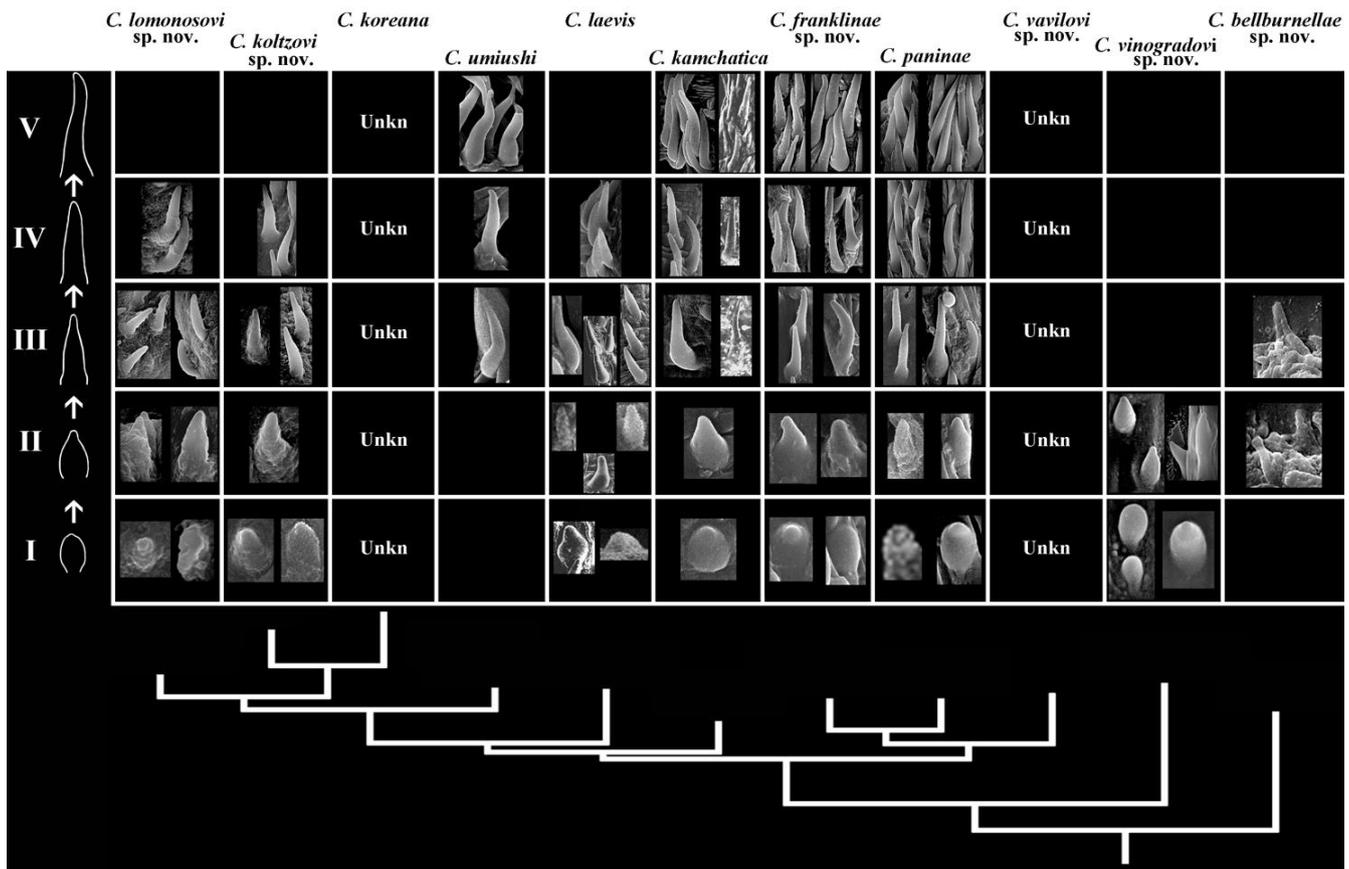


Figure 5. Phyloperiodic presentation of the copulative spines among species *Cadlina*, represented by the horizontal rows. The vertical columns (groups, taxa) are ordered by the molecular phylogenetic tree. Five main periods (horizontal rows, Roman numerals) are presented following the direction from short, oval rounded or irregular plates to the elongated spines. Empty cells imply non-observed/unknown forms. See text for details.

This method has been successfully applied to three distantly related groups of nudibranchs [7,29,56]. The shape of the jaw elements [32], traditionally used in taxonomy, is shown here as similar in all species and is therefore not included here in the phyloperiodic table, despite the fact that some small-scale differences could potentially be discovered later. The presentation of the general schemes of reproductive systems required drawings, which may include subjective elements; therefore, we did not use them for the purposes of the phyloperiodic approach. In addition, not all specimens may represent equally similar reproductive systems due to varying degrees of maturity and subsequent preservation. However, we carefully checked the reproductive systems and documented all differences in a more conventional taxonomic way. The molecular phylogenetic component of the phyloperiodic approach helps to clearly confirm the presence of periodic patterns in morphological characters in every available specimen of each species. The vertical columns based on molecular data (Figures 3–5) show existing or potentially existing similar morphological characters in each corresponding horizontal row, which is the essence of the periodic approach in science [36,57]. The conjunction of morphology in the broad ontogenetic sense [54] and molecular data thus brings forward truly biological, multilevel dimensions for both traditional and “phylogenetic” systematics.

Building the phyloperiodic rows of chromatic patterns. Similar chromatic patterns within each potential species were aligned using calibration by the degree of light to dark surface pigmentation and transparency of body tissue, including the presence of yellow spots, yellow notal lines, and brownish/dark pigmentation. The present scheme is a further elaboration of a similar scheme that has been used on a practical basis [7].

These similar forms establish horizontal rows of similar-looking specimens within the species (Figure 3). In total, the following eleven chromatic patterns (horizontal rows I–XI) are recognized:

- I Body white, yellow spots absent, brownish/dark colouration absent, dispersed white spots may be present, yellow notal line absent, light yellow subepidermal glands distinguishable, and rhinophores and gills white;
- II Body white, yellow spots absent, brownish/dark colouration absent, dispersed white spots may be present, yellow notal line absent, yellow subepidermal glands barely distinguishable, and rhinophores and gills light yellow;
- III Body white, yellow spots absent to hardly distinguishable, dispersed white spots may be present, brownish/dark colouration absent, yellow notal line absent, yellow subepidermal glands distinguishable, and rhinophores and gills white to light yellow;
- IV Body white to creamy, yellow spots absent, dispersed white spots may be present, brownish/dark colouration absent, yellow notal line absent, yellow subepidermal glands distinguishable, and rhinophores and gills yellow to light brownish;
- V Body white to creamy, faint yellow spots may be present, dispersed white spots may be present, brownish/dark colouration absent, yellow notal line absent, yellow subepidermal glands barely distinguishable, and rhinophores and gills white to light yellow;
- VI Body white to creamy, faint yellow spots present, dispersed white spots may be present, brownish/dark colouration absent, yellow notal line partially present, dispersed, yellow subepidermal glands distinguishable, and rhinophores and gills yellow to light brownish;
- VII Body white to creamy, distinct yellow spots present, white spots barely distinguishable, brownish/dark colouration absent, yellow notal line present, moderately distinct, yellow subepidermal glands distinguishable, and rhinophores and gills white to light brownish;
- VIII Body white to creamy and light brownish, distinct yellow spots present, white spots may form distinct opaque white covering, brownish/dark colouration absent, yellow notal line present, yellow subepidermal glands distinguishable, and rhinophores and gills yellow to light brownish;
- IX Body white to brownish, distinct yellow spots present, white spots barely distinguishable, brownish/dark colouration completely absent, yellow notal line distinct, yellow subepidermal glands distinguishable, and rhinophores and gills white to light yellow;
- X Body with patchy brownish to uniform yellowish/dark colouration, white spots barely distinguishable, numerous yellow spots present, yellow notal line distinct, yellow subepidermal glands distinguishable, and rhinophores and gills yellow to brownish;
- XI Body with almost completely brownish/dark colouration, white and yellow spots, yellow notal line or subepidermal glands barely distinguishable, and rhinophores and gills brownish.

Building the phyloperiodic rows of the central teeth of the radula. Juveniles have no denticles on the central (rachidian) teeth, and teeth are generally low [32,58]. Therefore, rows here are built following the direction from non-denticulate and relatively low teeth to higher teeth, where denticles occupy an increasingly larger part of the teeth's length (Figure 4). Six main phyloperiods are presented as follows:

- I—central teeth without denticles; II—denticles on central teeth occupy ca. $\frac{1}{4}$ or are shorter than $\frac{1}{4}$ of the teeth height; III—denticles on central teeth are longer than $\frac{1}{4}$ but shorter than $\frac{1}{2}$ of the teeth height; IV—denticles on central teeth occupy ca. $\frac{1}{2}$ of the teeth

height; V—denticles on central teeth are longer than $\frac{1}{2}$ but shorter than $\frac{3}{4}$ of the teeth height; and VI—denticles on central teeth occupy ca. $\frac{3}{4}$ or are longer than $\frac{3}{4}$ of the teeth height.

Building the phyloperiodic rows of copulative spines. Juveniles have no elaborated reproductive system [55] or spines of the copulative apparatus; therefore, further development should first involve short spines and then longer spines. The rows were built using direction from short, oval rounded or irregular plates to the elongated spines (Figure 5): I—short, oval to irregularly shaped, and top part not elongated; II—short, oval to irregularly shaped, and top part elongated; III—relatively short, straight, conical, and top part slightly elongated; IV—elongated, partly curved, conical, and top part rather rounded; and V—elongated, significantly convoluted, conical, and top part rather rounded.

3.4. Systematics

Class Gastropoda Cuvier, 1795

Family Cadlinidae Bergh, 1891

Genus *Cadlina* Bergh, 1879

***Cadlina bellburnellae* sp. nov.**

(Figures 3, 4, 5, 6 and 7A)

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Etymology. This species is named after Jocelyn Bell Burnell, who was one of the main contributors to the discovery of pulsars, but did not receive the recognition of a Nobel Prize in Physics in 1974.

Type materials. Holotype ZMMU Op-870, length 14 mm live, Kuril Islands, Iturup Island, 29 August 2022, depth 15–20 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-871, length 20 mm live, Kuril Islands, Iturup Island, 30 August 2022, depth 15–20 m, coll. T. A. Korshunova, A.V. Martynov. Paratype ZMMU Op-872, length 15 mm live, Kuril Islands, Iturup Island, 3 September 2022, depth 15–20 m, coll. T.A. Korshunova, A.V. Martynov.

External morphology. The notum is broad, and rounded in front and posteriorly. The rhinophores are long and retracted into raised soft sheaths bearing small tubercles (Figure 6(A1,B1–B3,C1,C2)); 9–12 rhinophoral lamellae. The notum is covered with medium- to small-sized, rounded or irregular tubercles (Figure 6(A1,A9,B1,B3,B8,C1,C11,C12)). The spicules form a sparse network in the notum. Eight to ten multipinnate gills united by a common membrane form a circle around the anus. The gills are retractable into the common gill cavity. The border of the gill cavity is moderately raised with a tuberculated edge (Figure 6). The oral veil is small, trapezoid, with oblique notched lateral sides (Figure 6(A2,B2,B3,C2)). The foot is broad, anteriorly rounded, and slightly thickened to form a double edge. The foot appears entirely (Figure 6(A2,B2,C2)); posteriorly, it sometimes projects slightly from notum in crawling animals, forming a rounded tail.

Colour. The notum is partly semi-transparent or almost completely opaque, from white to pale creamy to light yellow and brownish (Figure 6(A1,A2,B1–B3,C1,C2)). The gills are somewhat paler or darker than the notum with a patchy light-yellow pigment. The rhinophores are slightly darker than the notum. The digestive gland is black brownish to brownish; the stomach and intestine are partly visible through the notum dorsally (Figure 6(A1,B1,C1)). A whitish area (parts of the reproductive and digestive systems) may shine through the notum in front of the stomach. The internal organs can also be almost completely invisible. A range of 3–8 subepidermal yellow glands are well conspicuous dorsally and less conspicuous ventrally (some of them are placed close to each other) from each body side (Figure 6(A1,B1,B3,C1)). A yellow line appears around the notum (sometimes is less distinct) and yellow pigment in the dorsal tubercles is well defined (Figure 6(A1,A2,B1–B3,C1,C2)). The yellow line around the foot is somewhat more dispersed but also distinct.

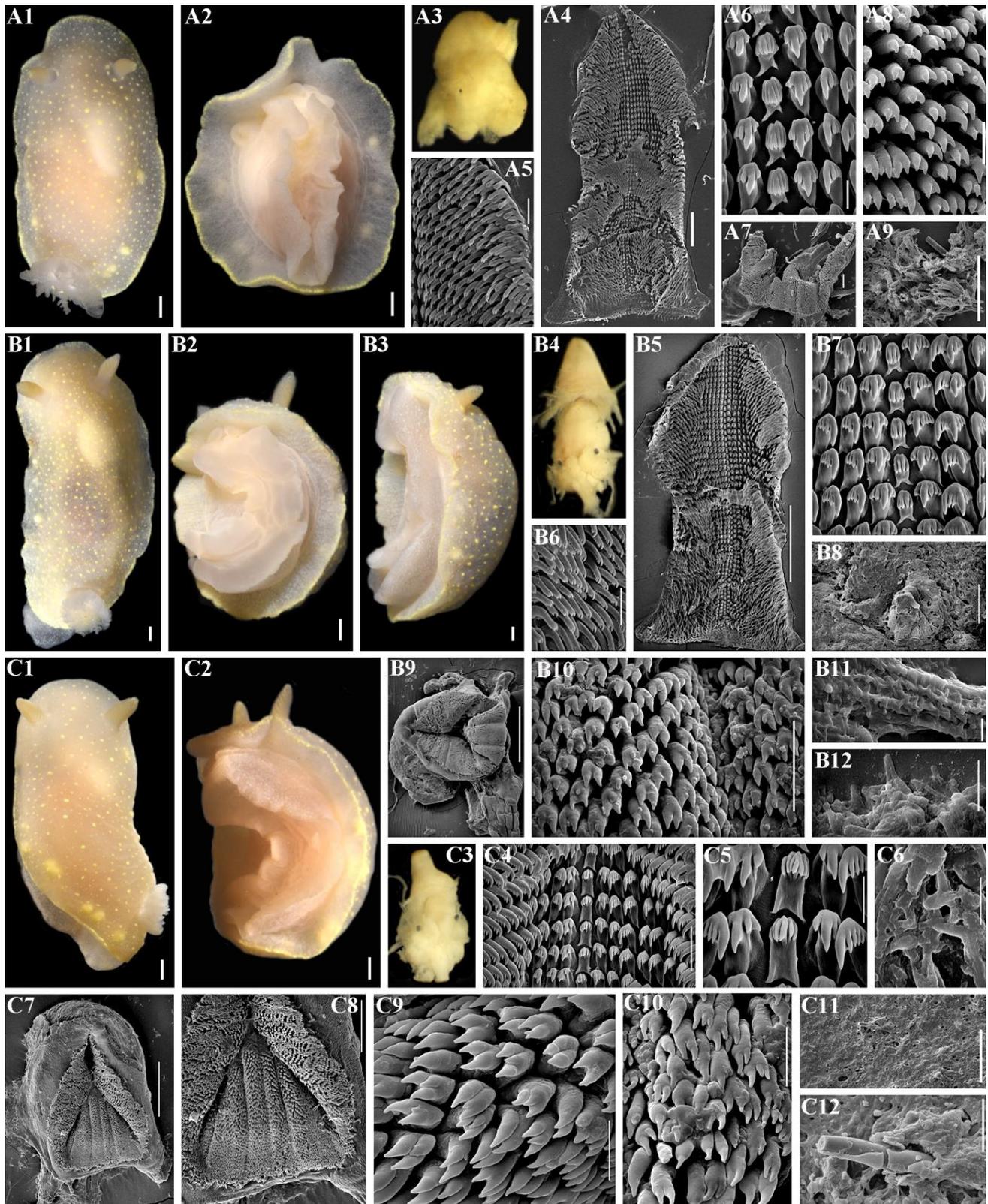


Figure 6. *Cadlina bellburnellae* sp. nov., external and internal features. (A) Holotype ZMMU Op-870, length 14 mm live, Iturup Island. (A1) Dorsal view. (A2) Ventral view. (A3) Buccal bulb, light microscopy (LM). (A4) Complete radula, scanning electron microscopy (SEM), scale bar 200 µm. (A5) Outer lateral teeth, SEM, 50 µm. (A6) Central teeth and first lateral teeth, SEM, 20 µm. (A7) Labial

cuticle, SEM, 100 μm . (A8) Elements of labial cuticle, SEM, 10 μm . (A9) Spicules of notum, SEM, 500 μm . (B) Paratype ZMMU Op-871, length 20 mm live, Iturup Island. (B1) Dorsal view. (B2) Ventral view. (B3) Lateral view. (B4) Buccal bulb, LM. (B5) Complete radula, SEM, 400 μm . (B6) outer lateral teeth, SEM, 50 μm . (B7) Central teeth and first lateral teeth, SEM, 40 μm . (B8) Notum, SEM, 100 μm . (B9) Complete labial cuticle, SEM, 300 μm . (B10) Elements of labial cuticle, SEM, 20 μm . (B11) Copulative spines, SEM, 5 μm . (B12) Copulative spines, SEM, 10 μm . (C) Paratype ZMMU Op-872, length 15 mm live. (C1) Dorsal view. (C2) Ventrolateral view. (C3) Buccal bulb, LM. (C4) Central part of radula, SEM, 50 μm . (C5) Enlarged central part of radula, central teeth and first lateral teeth, SEM, 20 μm . (C6) Copulative spines, SEM, 10 μm . (C7) Complete labial cuticle, SEM, 200 μm . (C8) Enlarged labial cuticle, SEM, 100 μm . (C9) Elements of labial cuticle, SEM, 10 μm . (C10) Elements of labial cuticle, SEM, 10 μm . (C11) Notum, SEM, 100 μm . (C12) Spicules of notum, SEM, 20 μm . Scale bars for all living specimens are 1 mm. Photos: Tatiana Korshunova and Alexander Martynov.

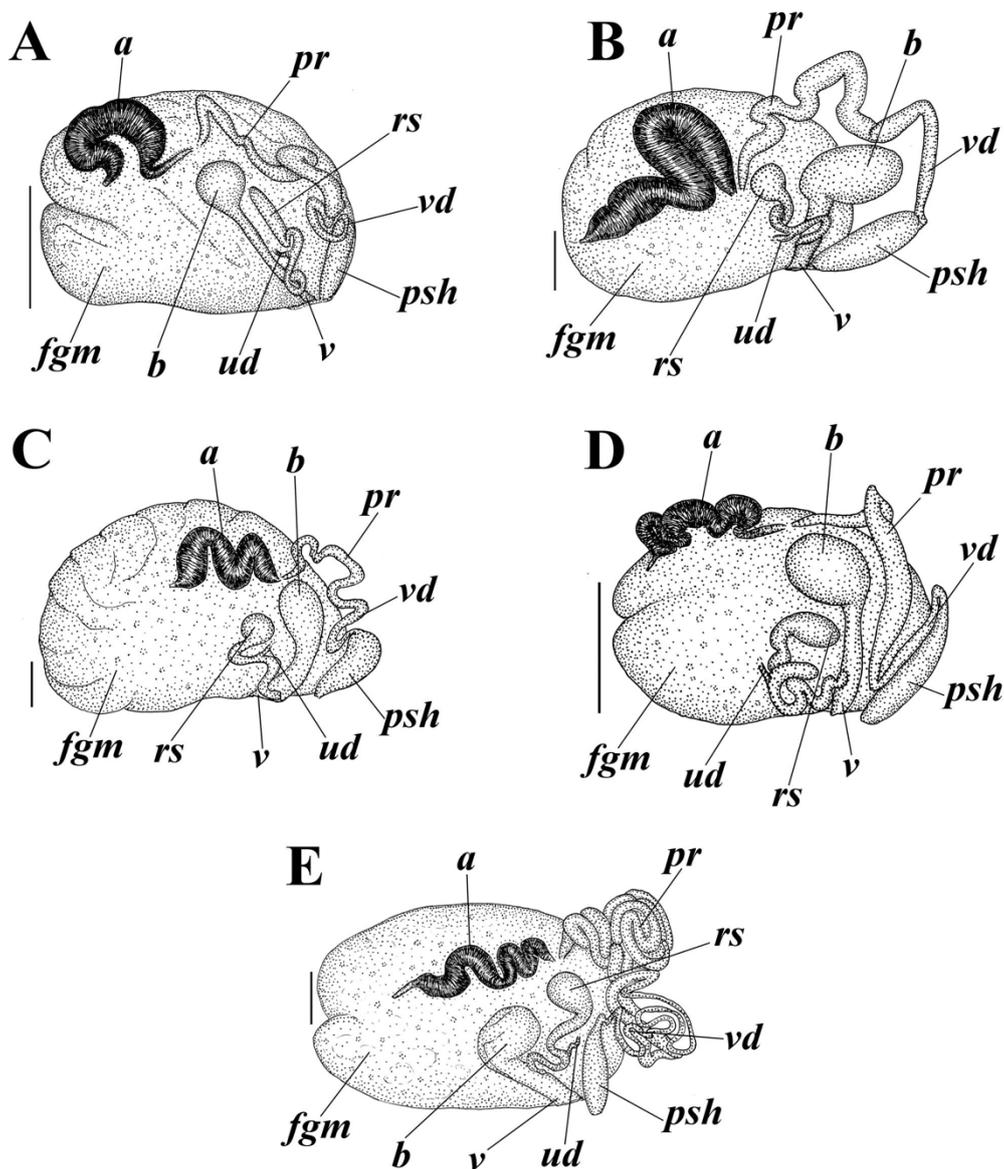


Figure 7. Reproductive systems of the genus *Cadlina*. (A) *Cadlina bellburnellae* sp. nov. (B) *Cadlina franklinae* sp. nov. (C) *Cadlina koltzovi* sp. nov. (D) *Cadlina lomonosovi* sp. nov. (E) *Cadlina vinogradovi* sp. nov. Abbreviations: a, ampulla; b, bursa; fgm, female gland mass; rs, receptaculum seminis; pr, prostate; psh, penial sheath; ud, uterine duct; v, vaginal duct; and vd, vas deferens. Scale bars are 1 mm. Drawings: Tatiana Korshunova.

Buccal bulb and oral tube. The buccal bulb is relatively short, similar in length to the oral tube (Figure 6(A3,B4,C3)). The salivary glands are relatively long and narrow.

Jaws. The jaws take the form of a rounded labial disk covered by a yellowish-brown cuticle bearing rod-shaped labial elements with double- to triple-pointed, hook-shaped tips (Figure 6(A7,A8,B9,B10,C7–C10)).

Radula. Radular formula ca. 65–80 × 35–40.1.35–40. The central tooth is moderately low to relatively high and bears up to nine cusps (Figure 6(A6,B7,C4,C5)). The inner lateral teeth (including unique teeth with two main cusps and double sets of denticles on lateral edges) have up to five denticles on the outer edge, and up to three on the inner edge (Figure 6(A4,A6,B5,B7,C4,C5)). The middle and outer teeth are comb-shaped, bearing up to ca. seven, which are concentrated at the top of the teeth, making the shape of the denticles quite peculiar (Figure 6(A5,B6)).

Reproductive system. The ampulla comprises at least two thickened compartments (Figure 7A, a). The ampulla bifurcates into moderately long vas deferens and oviduct. The uterine duct emerges some distance from the female gland mass. The prostatic part of the vas deferens is long, narrow, and only moderately distinct (Figure 7A, pr). The prostate transits to narrow vas deferens (Figure 7A, vd), which considerably widens towards the penial sheath that encloses the evertable ejaculatory duct (Figure 7A, psh). The copulative (penial) spines are conical and relatively short according to available data (Figures 5 and 6(B11,B12,C6)). The vagina narrows (Figure 7A, v) and enters a medium-sized rounded bursa copulatrix (Figure 7A, b). The uterine duct is short and narrow (Figure 7A, ud); it begins from the female gland mass and then enters near the base of a relatively large elongate receptaculum seminis (Figure 7A, rs).

Habitat. The species inhabits shallow waters with rocky and stony substrates at depths of c. 15–20 m.

Distribution. Currently, the species is only known to occur in the coastal waters of Iturup Island (Figure 2B); however, potential distribution includes at least the neighbouring Kuril Islands and Hokkaido Island.

Remarks. The species exhibits known chromatic patterns VIII–X (Figure 3), central teeth patterns II, IV, and V (Figure 4), and copulative spine patterns II and III (Figure 5). Due to the combination of a significant number of yellow spots, the distinct to the less distinct yellow line around the notum, the moderately low to relatively high (in that case only pattern II is known) central radular teeth, the moderately long distal part of vas deferens, the relatively short but elongated copulative spines (more elongated ones can be potentially discovered), and the molecular phylogenetic data (Figure 1), this new species *Cadlina bellburnellae* sp. nov. is distinguished from all known [32] and herein-described species of *Cadlina* (Figures 3–5 and 7). The intragroup distance in *C. bellburnellae* sp. nov. is 0% for the COI. The lowest COI intergroup distance of 6.53% is found between *C. bellburnellae* sp. nov. and *C. paninae* (Table 1). See also the details in Section 4.

Table 1. Uncorrected *p*-distances for COI (%) estimated within (highlighted in bold) and between species of the genus *Cadlina*.

Distances within Species	<i>C. bellburnellae</i> sp. nov.	<i>C. franklinae</i> sp. nov.	<i>C. koltzovi</i> sp. nov.	<i>C. lomonosovi</i> sp. nov.	<i>C. varilovi</i> sp. nov.	<i>C. vinogradovi</i> sp. nov.
<i>C. bellburnellae</i> sp. nov.	0	7.90–8.05	8.05–8.21	7.45–7.75	8.99	8.81–9.57
<i>C. franklinae</i> sp. nov.	0–0.30	-	7.14–7.45	5.93–6.23	4.82–4.98	8.66–9.57
<i>C. koltzovi</i> sp. nov.	0–0.15	8.05–8.21	-	6.38–7.14	8.83	10.18–11.09

Table 1. Cont.

	Distances within Species	<i>C. bellburnellae</i> sp. nov.	<i>C. franklinae</i> sp. nov.	<i>C. koltzovi</i> sp. nov.	<i>C. lomonosovi</i> sp. nov.	<i>C. vavilovi</i> sp. nov.	<i>C. vinogradovi</i> sp. nov.
<i>C. lomonosovi</i> sp. nov.	0–1.52	7.45–7.75	5.93–6.23	6.38–7.14	-	6.10–6.42	8.81–9.73
<i>C. vavilovi</i> sp. nov.	-	8.99	4.82–4.98	8.83	6.10–6.42	-	8.83–9.31
<i>C. vinogradovi</i> sp. nov.	0.61–1.06	8.81–9.57	8.66–9.57	10.18–11.09	8.81–9.73	8.83–9.31	-
<i>C. flavomaculata</i>	0.85	13.61–13.98	16.33–16.72	13.95–14.13	15.48–16.41	17.34–17.52	15.65–16.57
<i>C. jannanicholsae</i>	0.17–0.91	10.49–10.83	11.40–12.18	10.64–11.25	10.33–11.40	12.52–12.86	11.85–12.61
<i>C. japonica</i>	0–0.78	11.41–12.01	11.41–12.46	11.88–12.77	11.09–12.61	13.32–13.48	11.88–12.77
<i>C. kamchatica</i>	0–0.30	7.90–8.21	4.56–5.02	7.45–7.90	5.62–6.08	5.30–5.62	8.05–8.97
<i>C. klasmalmbergi</i>	0–0.30	11.85–12.19	12.46–12.96	12.31–12.77	11.85–12.61	14.45–14.61	12.77–13.73
<i>C. koreana</i>	0	8.13	8.28–8.44	6.72–6.88	6.41–7.03	9.47	10.00–10.63
<i>C. laevis</i>	0–1.37	7.14–7.90	4.41–5.02	6.69–7.45	4.56–5.32	5.14–5.78	7.75–8.68
<i>C. luarna</i>	0	13.68	16.11–16.26	13.68–13.83	15.81–16.41	17.34	15.35–16.26
<i>C. luteomarginata</i>	-	11.57	13.24–13.39	11.26–11.42	10.50–11.57	13.32	12.48–13.24
<i>C. modesta</i>	0–0.61	14.29–14.89	15.96–16.57	14.59–15.35	15.35–16.72	17.34–17.98	16.57–17.48
<i>C. paninae</i>	0	6.53	2.74–2.89	7.14–7.29	5.02–5.17	4.82	8.36–9.12
<i>C. pellucida</i>	-	11.51	1.83–11.99	11.04–11.20	10.73–11.51	13.68	11.83–12.78
<i>C. rumia</i>	-	13.83	15.35–15.50	13.83–13.98	14.13–14.44	16.05	15.05–15.65
<i>C. sp.</i> (Africa)	-	12.94	14.92–14.76	14.46–14.61	13.70–14.46	17.01	12.94–13.24
<i>C. sparsa</i>	-	14.74	17.63–17.78	15.05–15.20	15.96–16.72	17.98	14.35–15.81
<i>C. sylviaearleae</i>	0	14.29	14.29–14.44	13.37–13.53	12.46–12.77	16.05	14.74–15.35
<i>C. umiushi</i>	0–1.56	6.88–7.60	5.47–6.38	5.63–6.99	3.28–4.10	5.46–6.58	7.97–9.42

***Cadlina franklinae* sp. nov.**

Figures 3–5, Figures 7B and 8

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Etymology. This species is named after Rosalind Franklin, co-author of the DNA discovery, who died early (radiation during the study of DNA is considered a factor in her death), not having received the Nobel Prize together with Crick and Watson.

Type materials. Holotype ZMMU Op-845, length 39 mm live, Kuril Islands, Urup Island, 27 August 2021, depth 15–20 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-844, length 36 mm live, Kuril Islands, Urup Island, 2 September 2021, depth 10–20 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-846, length 34 mm, Kuril Islands, Chirpoy Island, 24 August 2021, depth 15–20 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-847, 19 mm live, Kuril Islands, Urup Island, 2 September 2021, depth 20–30 m, coll. T. A. Korshunova, A.V. Martynov. Paratype ZMMU Op-848, length 17 mm live, Kuril Islands, Urup Island, 27 August 2021, depth 15–20 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-849, 24 mm, Kuril Island, Urup Island, 2 September 2021, depth 20–30 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-856, length 15 mm live, Kuril Islands, Urup Island, 1 September 2022, depth 15–20 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-857, length 14 mm live, Kuril Islands, Urup Island, 1 September 2022, depth 15–20 m, coll. T.A. Korshunova, A.V. Martynov.

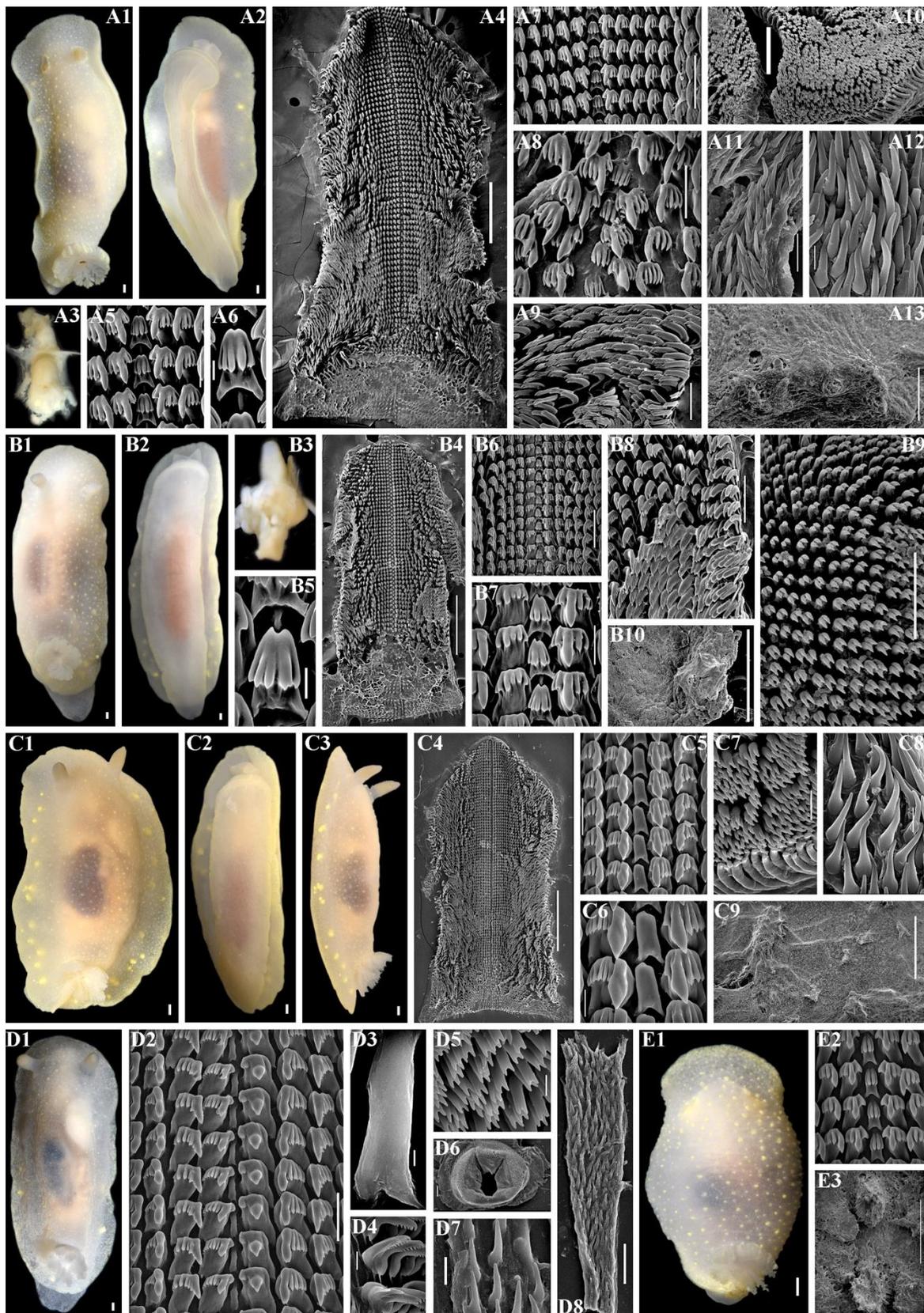


Figure 8. *Cadlina franklinae* sp. nov., external and internal features. (A) Holotype ZMMU Op-845, length 39 mm live, Urup Island. (A1) Dorsal view. (A2) Ventral view. (A3) Buccal bulb, light microscopy (LM). (A4) Complete radula, scanning electron microscopy (SEM), scale bar 500 μ m.

(A5) Central teeth and first lateral teeth, SEM, 40 μm . (A6) Central tooth, SEM, 10 μm . (A7) Central part of radula, SEM, 100 μm . (A8) Central part of radula, SEM, 100 μm . (A9) Outer lateral teeth, SEM, 100 μm . (A10) Labial cuticle, SEM, 100 μm . (A11) Copulative spines, SEM, 30 μm . (A12) Copulative spines, SEM, 10 μm . (A13) Notum and spicules, SEM, 100 μm . (B) Paratype ZMMU Op-844, length 36 mm live, Urup Island. (B1) Dorsal view. (B2) Ventral view. (B3) Buccal bulb, LM. (B4) Complete radula, SEM, 500 μm . (B5) Central tooth, SEM, 10 μm . (B6) Central part of radula, SEM, 100 μm . (B7) Central teeth and first lateral teeth, SEM, 30 μm . (B8) Outer lateral teeth, SEM, 100 μm . (B9) Elements of labial cuticle, SEM, 50 μm . (B10) Notum, SEM, 500 μm . (C) Paratype ZMMU Op-849, length 24 mm live, Urup Island. (C1) Dorsal view. (C2) Ventral view. (C3) Lateral view. (C4) Complete radula, SEM, 500 μm . (C5) Central part of radula, SEM, 50 μm . (C6) Central teeth and first lateral teeth, SEM, 20 μm . (C7) Elements of labial cuticle, SEM, 20 μm . (C8) Copulative spines, SEM, 10 μm . (C9) Notum, SEM, 500 μm . (D) Paratype ZMMU Op-846, length 34 mm, Chirpoy Island. (D1) Dorsal view. (D2) Central part of radula, SEM, 50 μm . (D3) Central tooth, SEM, 2 μm . (D4) Outer lateral teeth, SEM, 20 μm . (D5) Elements of labial cuticle, SEM, 10 μm . (D6) Complete labial cuticle, SEM, 200 μm . (D7) Copulative spines, SEM, 10 μm . (D8) Copulative spines, SEM, 50 μm . (E) Paratype ZMMU Op-857, length 14 mm live, Urup Island. (E1) Dorsal view. (E2) Central part of radula, SEM, 20 μm . (E3) Notum, SEM, 300 μm . Scale bars for all living specimens are 1 mm. Photos: Tatiana Korshunova and Alexander Martynov.

External morphology. The notum is broad, and rounded in front and posteriorly. The rhinophores are long and retracted into raised soft almost smooth or bearing very small tubercle sheaths (Figure 8(A1,B1,C1,C3,D1,E1)); 10–16 rhinophoral lamellae. The notum is almost smooth or covered with very small, indistinct tubercles (Figure 8(A1,A13,B1,B10,C1,C3,C9,D1,E1,E3)). The spicules form a sparse network in the notum. Seven to nine multipinnate gills united by a common membrane form a circle around the anus. The gills are retractable into a common gill cavity. The border of the gill cavity is moderately raised with a tuberculated edge (Figure 8). The oral veil is small, trapezoid, with oblique notched lateral sides (Figure 8(A2,B2,C2)). The foot is broad, anteriorly rounded and slightly thickened to form a double edge. The foot appears entirely (Figure 8(A2,B2,C2)); posteriorly, it sometimes projects slightly from the notum in crawling animals, forming a rounded tail.

Colour. The notum is commonly white, and additionally may have patchy covers with opaque white dust, partly semi-transparent, more rarely more towards a yellowish uniform colour (Figure 8(A1,B1,C1,C3,D1,E1)). The gills are somewhat paler, darker, or almost the same colour as the notum, without or with a patchy yellow pigment. The rhinophores are slightly darker than notum. The digestive gland is black and brownish; the stomach and intestine are partly visible through the notum dorsally (Figure 8(A1,B1,C1,C3,D1,E1)). The whitish to pinkish area (parts of the reproductive and digestive systems) may shine through the notum in front and behind the stomach. The internal organs can also be almost completely invisible. A range of 2–15 (sometimes 19 if smaller ones are counted) subepidermal yellow glands are barely distinguishable to conspicuous dorsally and ventrally from each body side. The yellow line around the notum is more commonly absent or very indistinct; only rarely is it more distinct. A yellow pigment appears dorsally or within small-sized tubercles (Figure 8(A1,B1,C1,C3,D1,E1)). The yellow lines around the foot are commonly absent or very weak but can also be more distinct.

Buccal bulb and oral tube. The buccal bulb is relatively short, similar in length to the oral tube (Figure 8(A3,B3)). The salivary glands are relatively long and narrow.

Jaws. The jaws take the form of a rounded labial disk covered by a yellowish-brown cuticle bearing rod-shaped labial elements with double-pointed hook-shaped tips (Figure 8(A10,B9,C7,D5,D6)).

Radula. Radular formula ca. 80–120 \times 30–50.1.30–50. The central tooth is moderately low, bears up to six cusps, and is rarely completely devoid of cusps (Figure 8(A5–A8,B5–B7,C5,C6,D2,D3,E2)). The inner lateral teeth have up to eight denticles on the outer edge, and up to two on the

inner edge (Figure 8(A4,A5,A7,A8,B4,B6,B7,C4,C5,C6,D2,E2)). The middle and outer teeth are comb-shaped, bearing up to 25 denticles (Figure 8(A4,A5,A9,B4,B8,C4,D4)).

Reproductive system. The ampulla comprises at least three thickened compartments (Figure 7B, a). The ampulla bifurcates into moderately long vas deferens and oviduct. The uterine duct emerges some distance from the female gland mass. The prostatic part of the vas deferens is long, narrow, and only moderately distinct (Figure 7B, pr). The prostate transits to narrow vas deferens (Figure 7B, vd), which considerably widens towards the penial sheath that encloses the evertible ejaculatory duct (Figure 7B, psh). The copulative (penial) spines are commonly elongated and convoluted, and some shorter ones may present in addition (Figures 5 and 8(A11,A12,C8,D7,D8)). The vagina narrows (Figure 7B, v) and enters a relatively large oval bursa copulatrix (Figure 7B, b). The uterine duct is short and narrow (Figure 7B, ud); it begins from the female gland mass and then enters near the base of the pear-shaped receptaculum seminis (Figure 7B, rs).

Habitat. The species inhabits shallow waters with rocky and stony substrates at depths of c. 15–30 m.

Distribution. Currently, the species is only known to occur in the coastal waters of the Urup and Chirpoy Islands (Figure 2B); however, potential distribution includes at least the neighbouring Kuril Islands.

Remarks. The species exhibits known chromatic patterns II, V, VIII, and X (Figure 3), central teeth patterns I, IV, and V (Figure 4), and copulative spine patterns I–V (Figure 5). Due to the combination of the moderate number of yellow spots, the usual absence of a distinct yellow line around the notum, the notum commonly with opaque white sprinkled small spots, the relatively low central radular teeth, the moderate distal part of vas deferens and commonly elongate copulative spines, and the molecular phylogenetic data (Figure 1), this new species *Cadlina franklinae* sp. nov. is distinguished from all known [32] and herein-described species of *Cadlina* (Figures 3, 4, 5 and 7B). The intragroup distances in *C. franklinae* sp. nov. are 0–0.30% for the COI. The lowest COI intergroup distance of 2.74% is found between *C. franklinae* sp. nov. and *C. paninae* (Table 1). See also the details in Section 4.

***Cadlina koltzovi* sp. nov.**

Figures 3, 4, 5, 7C and 9

zoobank no.: urn:lsid:zoobank.org:act: 31006AD5-3604-4574-8A4B-66E277DA5441

Etymology. This species is named after Nikolai Koltzov, founder of the Institute of Experimental Biology (which has now transitioned to the Institute of Developmental Biology RAS), who pioneered the idea of a cytoskeleton and anticipated many concepts of modern genetics (including the structure of DNA and epigenetics), but died unexpectedly following persecution after refusal to support false testimonies against Nikolai Vavilov.

Type materials. Holotype ZMMU Op-842, length 24 mm live, Kuril Islands, Urup Island, 23 August 2021, depth 10–20, coll. T.A. Korshunova, A. V. Martynov. Paratype ZMMU Op-840, length 17 mm live, Kuril Islands, Urup Island, 2 September 2021, depth 10–40 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-841, length 15 mm live, Kuril Islands, Urup Island, 30 August 2021, depth 10–20 m, coll. T.A. Korshunova, A. V. Martynov. Paratype ZMMU Op-843, length 18 mm live, Kuril Island, Urup Island, 23 August 2021, coll. T. A. Korshunova, A.V. Martynov. Paratype ZMMU Op-852, length 17 mm live, Kuril Islands, Kunashir Island, 29 August 2022, depth 5–10 m, coll. T.A. Korshunova, A. V. Martynov. Paratype ZMMU Op-853, length 22 mm live, Kuril Islands, Iturup Island, 4 September 2022, depth 10–15 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-854, length 25 mm live, Kuril Island, Iturup Island, 4 September 2022, depth 10–15 m, coll. T.A. Korshunova, A.V. Martynov. ZMMU Op-855, length 24 mm live, Kuril Islands, Iturup Island, 4 September 2022, depth 10–15 m, coll. T.A. Korshunova, A.V. Martynov.

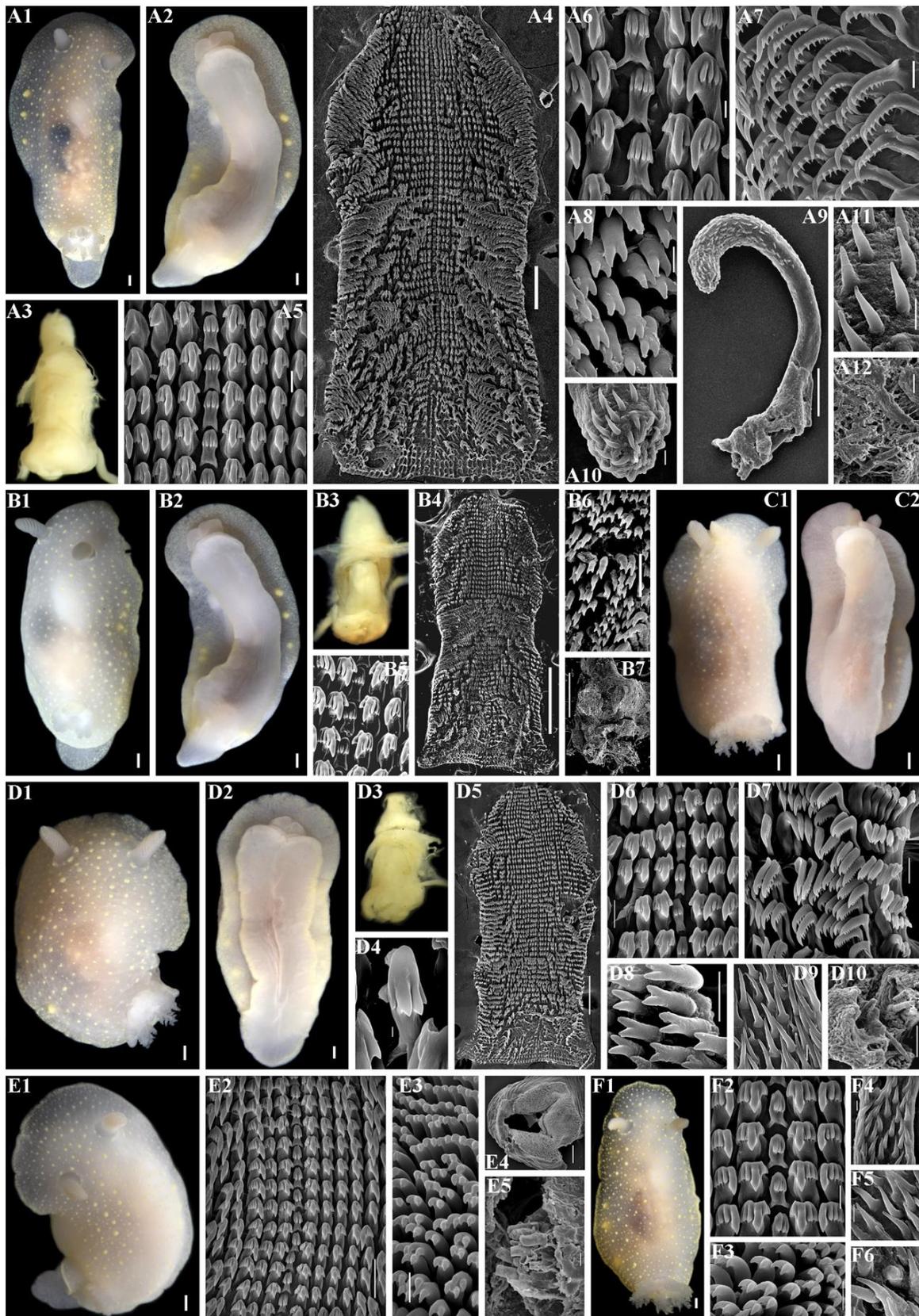


Figure 9. *Cadlina koltzovi* sp. nov., external and internal features. (A) Holotype ZMMU Op-842, length 24 mm live, Urup Island. (A1) Dorsal view. (A2) Ventral view. (A3) Buccal bulb, light microscopy (LM). (A4) Complete radula, scanning electron microscopy (SEM), scale bar 200 μ m.

(A5) Central part of radula, SEM, 20 μ m. (A6) Central teeth and first lateral teeth, SEM, 10 μ m. (A7) Outer lateral teeth, SEM, 20 μ m. (A8) Elements of labial cuticle, SEM, 5 μ m. (A9) Everted copulative apparatus, SEM, 100 μ m. (A10) Copulative spines, SEM, 10 μ m. (A11) Copulative spines, SEM, 10 μ m. (A12) Notum, SEM, 50 μ m. (B) Paratype ZMMU Op-840, length 17 mm live, Urup Island. (B1) Dorsal view. (B2) Ventral view. (B3) Buccal bulb, LM. (B4) Complete radula, SEM, 500 μ m. (B5) Central teeth and first lateral teeth, SEM, 30 μ m. (B6) Elements of labial cuticle, SEM, 20 μ m. (B7) Notum, SEM, 500 μ m. (C) Paratype ZMMU Op-841, length 15 mm live, Urup Island. (C1) Dorsal view. (C2) Ventral view. (D) Paratype ZMMU Op-843, length 18 mm live, Urup Island. (D1) Dorsal view. (D2) Ventral view. (D3) Buccal bulb, LM. (D4) Central tooth, SEM, 3 μ m. (D5) Complete radula, SEM, 300 μ m. (D6) Central part of radula, SEM, 30 μ m. (D7) Outer lateral teeth, SEM, 30 μ m. (D8) Elements of labial cuticle, SEM, 10 μ m. (D9) Copulative spines, SEM, 10 μ m. (D10) Notum and spicules, SEM, 100 μ m. (E) Paratype ZMMU Op-852, length 17 mm live, Kunashir Island. (E1) Dorsal view. (E2) Central part of radula, SEM, 50 μ m. (E3) Elements of labial cuticle, SEM, 10 μ m. (E4) Complete labial cuticle, SEM, 200 μ m. (E5) Spicules of notum, SEM, 20 μ m. (F) Paratype ZMMU Op-853, length 22 mm live, Iturup Island. (F1) Dorsal view. (F2) Central part of radula, SEM, 20 μ m. (F3) Elements of labial cuticle, SEM, 10 μ m. (F4) Copulative spines, SEM, 10 μ m. (F5) Copulative spines, SEM, 10 μ m. (F6) Copulative spines, SEM, 3 μ m. Scale bars for all living specimens are 1 mm. Photos: Tatiana Korshunova and Alexander Martynov.

External morphology. The notum is broad, and rounded in front and posteriorly. The rhinophores are long and retracted into raised soft almost smooth or bearing very small tubercle sheaths (Figure 9(A1,B1,C1,D1,E1,F1)); 8–13 rhinophoral lamellae. The notum is almost smooth or covered with very small, indistinct tubercles (Figure 9(A1,A12,B1,B7,C1,D1,D10,E1,E5,F1)). The spicules form a sparse network in the notum. Seven to nine multipinnate gills united by a common membrane form a circle around the anus. The gills are retractable into a common gill cavity. The border of the gill cavity is moderately raised with a tuberculated edge (Figure 9(A1,B1,C1,D1,E1,F1)). The oral veil is small, trapezoid, with oblique notched lateral sides (Figure 9(A2,B2,C2,D2)). The foot is broad, anteriorly rounded, and slightly thickened to form a double edge. The foot appears entirely (Figure 9(A2,B2,C2,D2)); posteriorly, it sometimes projects slightly from the notum in crawling animals, forming a rounded tail.

Colour. The notum is partly semi-transparent to more opaque, white to creamy, and patchy brownish to uniform yellowish in colour (Figure 9(A1,A2,B1,B2,C1,C2,D1,D2,E1,F1)). The gills are almost the same colour as the notum, without or with a patchy yellow pigment. The rhinophores are slightly darker than the notum. The digestive gland is black brownish to creamy; the stomach and intestine are partly visible through the notum dorsally (Figure 9(A1,B1,C1,D1,E1,F1)). The whitish area (parts of the reproductive and digestive systems) may shine through the notum in front and behind the stomach. The internal organs can also be almost completely invisible. A range of 3–12 subepidermal yellow glands are barely distinguishable to conspicuous dorsally and ventrally from each body side. The yellow line around the notum is usually absent or dispersed, or, rarely, present (Figure 9(A1,A2,B1,B2,C1,C2,D1,D2,E1,F1)). A yellow pigment appears dorsally or within small- to medium-sized tubercles (Figure 9(A1,B1,C1,D1,E1,F1)). The yellow line around the foot is absent.

Buccal bulb and oral tube. The buccal bulb is relatively short, similar in length to the oral tube (Figure 9(A3,B3,D3)). The salivary glands are relatively long and narrow.

Jaws. The jaws take the form of a rounded labial disk covered by a yellowish-brown cuticle bearing rod-shaped labial elements with double hook-shaped tips (Figure 9(A8,B6,D8,E3,E4,F3)).

Radula. Radular formula ca. 60–75 \times 25–45.1.25–45. The central tooth is moderately high and bears up to seven cusps (Figure 9(A5,A6,B5,D4,D6,E2,F2)). The inner lateral teeth have up to five denticles on the outer edge, and up to three on the inner edge (Figure 9(A4–A6,B4,B5,D5–D7,E2,F2)). The middle and outer teeth are comb-shaped, bearing up to 12 denticles (Figure 9(A4,A7,D5,D7)).

Reproductive system. The ampulla comprises at least three thickened compartments (Figure 7C, a). The ampulla bifurcates into moderately long vas deferens and oviduct. The uterine duct emerges some distance from the female gland mass. The prostatic part of the vas deferens is long, narrow, and not distinct (Figure 7C, pr). The prostate transits to narrow vas deferens (Figure 7C, vd), which considerably widens towards the penial sheath that encloses the evertable ejaculatory duct (Figure 7C, psh). The copulative (penial) spines are commonly elongated and curved, and some shorter ones may present in addition (Figures 5 and 9(A9–A11,D9,F4–F6)). The vagina narrows (Figure 7C, v) and enters a medium-sized oval bursa copulatrix (Figure 7C, b). The uterine duct is short and narrow (Figure 7C, ud); it begins from the female gland mass and then enters near the base of the pear-shaped receptaculum seminis (Figure 7C, rs).

Habitat. The species inhabits shallow waters with rocky and stony substrates at depths of c. 5–40 m.

Distribution. Currently, the species is only known to occur in the coastal waters of the Urup, Iturup, and Kunashir Islands (Figure 2B); however, potential distribution includes at least the neighbouring Kuril Islands and Hokkaido Island.

Remarks. The species exhibits known chromatic patterns V, VIII–X (Figure 3), central teeth patterns III–V (Figure 4), and copulative spine patterns I–IV (Figure 5). Due to the combination of a moderate number of yellow spots, the presence or absence of a distinct yellow line around the notum, the relatively high central radular teeth, the moderate distal part of vas deferens and commonly elongate copulative spines, and the molecular phylogenetic data (Figure 1), this new species *Cadlina koltzovi* sp. nov. is distinguished from all known [32] and herein-described species of *Cadlina* (Figures 3–5). The intragroup distances in *C. koltzovi* sp. nov. are 0–0.15% for the COI. The lowest COI intergroup distance of 5.63% is found between *C. koltzovi* sp. nov. and *C. umiushi* (Table 1). See also the details in Section 4.

***Cadlina lomonosovi* sp. nov.**

Figures 3, 4, 5, 7D and 10

zoobank no.: urn:lsid:zoobank.org:act: DA87EC75-2E17-45B3-8117-51DBF6960402

Etymology. This species is named after Mikhail Lomonosov, founder of the Moscow State University. Despite his fame, Lomonosov struggled with scientific bureaucrats and obscurantists in Russia throughout most of his career.

Type materials. Holotype ZMMU Op-868, length 17 mm live, Kuril Islands, Kunashir Island, 29 August 2022, depth 5–10 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-858, length 15 mm live, Kuril Islands, Iturup Island, 4 September 2022, depth 15–20 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-859, length 20 mm live, Kuril Islands, Iturup Island, 4 September 2022, depth 15–20 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-860, length 14 mm live, Kuril Islands, Iturup Island, 4 September 2022, depth 15–20 m, coll. T. A. Korshunova, A.V. Martynov. Paratype ZMMU Op-861, length 19 mm live, Kuril Islands, Iturup Island, 17 August 2022, depth 10–15 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-862, length 12 mm live, Kuril Islands, Iturup Island, 9 September 2022, depth 15–20 m, coll. T. A. Korshunova, A.V. Martynov. Paratype ZMMU Op-863, length 11 mm live, Kuril Islands, Iturup Island, 17 August 2022, depth 10–15 m, coll. T. A. Korshunova, A. V. Martynov. Paratype ZMMU Op-864, length 8 mm live, Kuril Islands, Iturup Islands, 5 September 2022, depth 10–15 m, coll. T. A. Korshunova, A.V. Martynov. Paratype ZMMU Op-865, length 10 mm live, Kuril Islands, Iturup Island, 30 August 2022, depth 15–20 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-866, length 24 mm live, Kuril Islands, Kunashir Island, 29 August 2022, depth 5–10 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-867, length 7 mm live, Kuril Islands, Iturup Island, 29 August 2022, depth 10–15 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-869, length 15 mm live, Kuril Islands, Kunashir Island, 29 August 2022, depth 5–10, coll. T. A. Korshunova, A.V. Martynov.

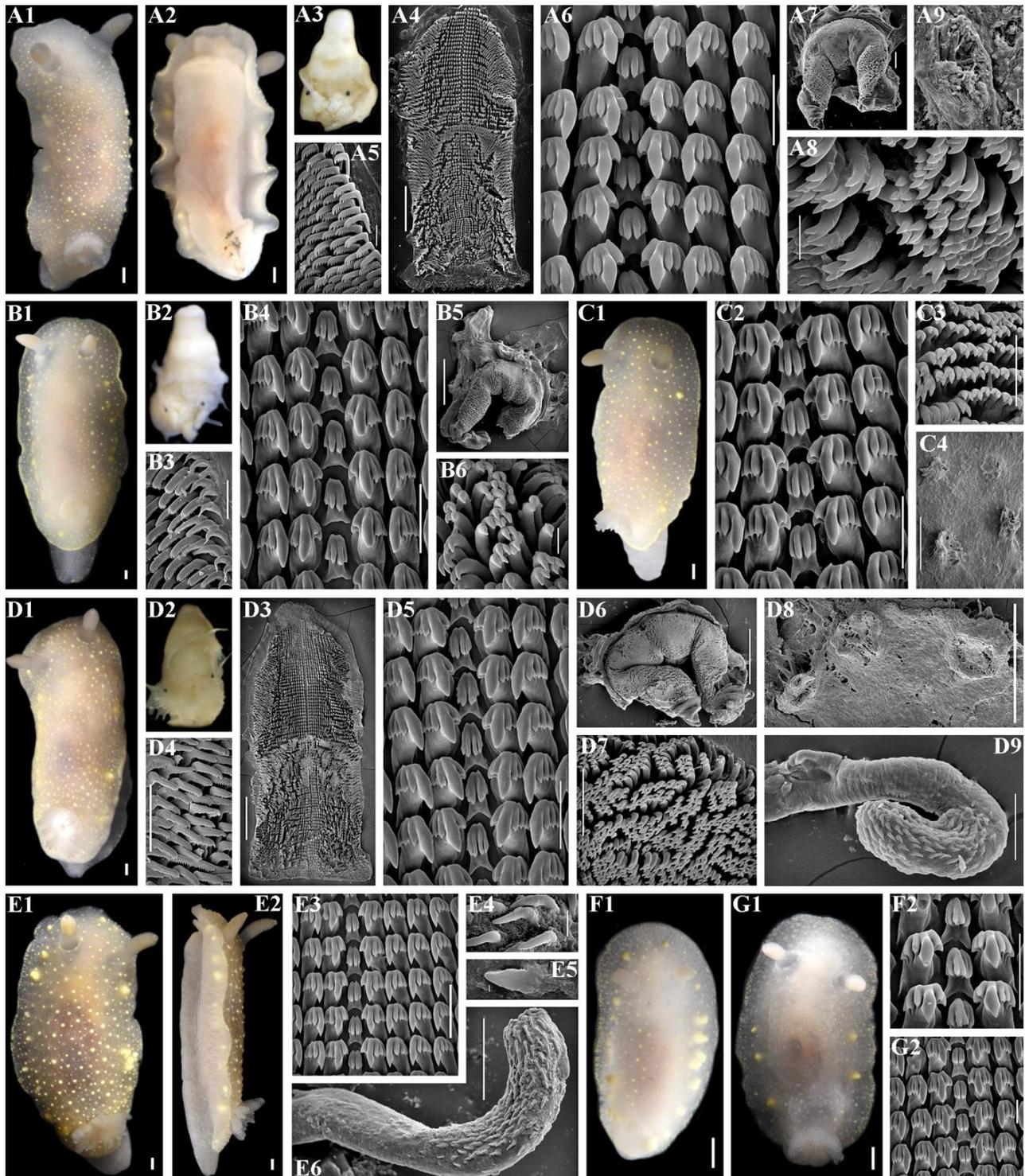


Figure 10. *Cadlina lomonosovi* sp. nov., external and internal features. (A) Holotype ZMMU Op-868, length 17 mm live, Kunashir Island. (A1) Dorsal view. (A2) Ventral view. (A3) Buccal bulb, light micro-copy (LM). (A4) Complete radula, scanning electron microscopy (SEM), scale bar, 300 μ m. (A5) Outer lateral teeth, SEM, 30 μ m. (A6) Central part of radula, 30 μ m. (A7) Complete labial cuticle, SEM, 100 μ m. (A8) Elements of labial cuticle, SEM, 10 μ m. (A9) Notum, SEM, 30 μ m. (B) Paratype ZMMU Op-866, length 24 mm live, Kunashir Island. (B1) Dorsal view. (B2) Buccal bulb, LM. (B3) Outer lateral teeth, SEM, 50 μ m. (B4) Central part of radula, SEM, 40 μ m. (B5) Complete labial cuticle, SEM,

500 µm. (B6) Elements of labial cuticle, SEM, 10 µm. (C) Paratype ZMMU Op-869, length 15 mm live, Kunashir Island. (C1) Dorsal view. (C2) Central part of radula, SEM, 30 µm. (C3) Elements of labial cuticle, SEM, 30 µm. (C4) Notum and spicules, SEM, 300 µm. (D) Paratype ZMMU Op-861, length 19 mm live, Iturup Island. (D1) Dorsal view. (D2) Buccal bulb, LM. (D3) Complete radula, SEM, 500 µm. (D4) Outer laterals, SEM, 100 µm. (D5) Central part of radula, SEM, 50 µm. (D6) Complete labial cuticle, SEM, 500 µm. (D7) Elements of labial cuticle, SEM, 50 µm. (D8) Notum and spicules, SEM, 500 µm. (D9) Everted copulative apparatus, SEM, 100 µm. (E) Paratype ZMMU Op-859, length 20 mm live, Iturup Island. (E1) Dorsal view. (E2) Lateral view. (E3) Central part of radula, SEM, 50 µm. (E4) Copulative spines, SEM, 10 µm. (E5) Copulative spines, SEM, 1 µm. (E6) Everted copulative apparatus, SEM, 100 µm. (F) Paratype ZMMU Op-864, length 8 mm live, Iturup Islands. (F1) Dorsal view. (F2) Central teeth and first lateral teeth, SEM, 30 µm. (G) Paratype ZMMU Op-863, length 11 mm live, Iturup Island. (G1) Dorsal view. (G2) Central part of the radula, SEM, 20 µm. Scale bars for all living specimens are 1 mm. Photos: Tatiana Korshunova and Alexander Martynov.

External morphology. The notum is broad, and rounded in front and posteriorly. The rhinophores are long and retracted into raised soft sheaths bearing small tubercles (Figure 10(A1,A2,B1,C1,D1,E1,E2,F1,G1)); 8–15 rhinophoral lamellae. The notum is covered with medium- to small-sized, rounded or irregular tubercles (Figure 10(A1,A9,B1,C1,C4,D1,D8,E1,E2,F1,G1)). The spicules form a sparse network in the notum. Eight to nine multipinnate gills united by a common membrane form a circle around the anus. The gills are retractable into a common gill cavity. The border of the gill cavity is moderately raised with a tuberculated edge (Figure 10(A1,B1,C1,D1,E1,E2,F1,G1)). The oral veil is small, trapezoid, with oblique notched lateral sides (Figure 10(A2,E2)). The foot is broad, anteriorly rounded, and slightly thickened to form a double edge. The foot appears entirely (Figure 10(A2,E2)); posteriorly, it sometimes projects slightly from the notum in crawling animals, forming a rounded tail.

Colour. The notum is partly semi-transparent or almost completely opaque, from white to pale creamy to light yellow and brownish (Figure 10(A1,A2,B1,C1,D1,E1,E2,F1,G1)). The gills are somewhat paler or darker than the notum with a patchy light-yellow pigment. The rhinophores are slightly darker than the notum. The digestive gland is black brownish to pinkish; the stomach and intestine are partly visible through the notum dorsally (Figure 10(A1,B1,C1,D1,E1,F1,G1)). A pinkish area (parts of the reproductive and digestive systems) may shine through the notum in front of the stomach. The internal organs can also be almost completely invisible. A range of 2–12 subepidermal yellow glands are well conspicuous dorsally and less conspicuous ventrally (some of them are placed close to each other) from each body side. The yellow lines around the notum and the yellow pigment in the dorsal tubercles are usually distinct and numerous (Figure 10(A1,A2,B1,C1,D1,E1)), although specimens without distinct yellow spots and dispersed notal lines are also present, which is consistent with the ontogenetic model since these specimens are commonly smaller in size (Figure 10(F1,G1)). The yellow line around the foot is dispersed and indistinct.

Buccal bulb and oral tube. The buccal bulb is relatively short, similar in length to the oral tube (Figure 10(A3,B2,D2)). The salivary glands are relatively long and narrow.

Jaws. The jaws take the form of a rounded labial disk covered by a yellowish-brown cuticle bearing rod-shaped labial elements with double-pointed hook-shaped tips (Figure 10(A7,A8,B6,C3,D6,D7)).

Radula. Radular formula ca. 100–110 × 30–50.1.30–50. The central tooth is moderately low and bears up to eight cusps (Figure 10(A6,B4,C2,D5,E3,F2,G2)). The inner lateral teeth have up to five denticles on the outer edge, and up to three on the inner edge (Figure 10(A4,A6,B4,C2,D3,D5,E3,F2,G2)). The middle and outer teeth are comb-shaped, bearing up to ca. 15 denticles (Figure 10(A4,A5,B3,D3,D4)).

Reproductive system. The ampulla comprises at least three thickened and convoluted compartments (Figure 7D, a). The ampulla bifurcates into moderately long vas deferens and oviduct. The uterine duct emerges some distance from the female gland mass (Figure 7D). The prostatic part of the vas deferens is long, narrow, and moderately distinct (Figure 7D, pr). The prostate transits to narrow vas deferens (Figure 7D, vd), which

considerably widens towards the penial sheath that encloses the evertable ejaculatory duct (Figure 7D, psh). The copulative (penial) spines are commonly elongated and curved, and some shorter ones may present in addition (Figures 5 and 10(D9,E4–E6)). The vagina narrows (Figure 7D, v) and enters a relatively large rounded bursa copulatrix (Figure 7D, b). The uterine duct is short and narrow (Figure 7D, ud); it begins from the female gland mass and then enters near the base of the oval receptaculum seminis (Figure 7D, rs).

Habitat. The species inhabits shallow waters with rocky and stony substrates at depths of c. 5–20 m.

Distribution. Currently, the species is only known to occur in the coastal waters of the Iturup and Kunashir Islands (Figure 2B); however, potential distribution includes at least the neighbouring Kuril Islands and Hokkaido Island.

Remarks. The species features known chromatic patterns VI–X (Figure 3), central teeth patterns IV–VI (Figure 4), and copulative spine patterns I–IV (Figure 5). Due to the combination of an insignificant to significant number of yellow spots, the distinct or less distinct but still present yellow line around the notum, the moderately low central radular teeth (in this case, pattern II is not known, only patterns IV–VI), the moderately long distal part of vas deferens, the commonly elongated copulative spines, and the molecular phylogenetic data (Figure 1), this new species *Cadlina lomonosovi* sp. nov. is distinguished from all known [32] and herein-described species of *Cadlina* (Figures 3–5). The intragroup distances in *C. lomonosovi* sp. nov. are 0–1.52% for the COI. The lowest COI intergroup distance of 3.28% is found between *C. lomonosovi* sp. nov. and *C. umiushi* (Table 1). See also the details in Section 4.

***Cadlina vavilovi* sp. nov.**

Figures 3–5 and 11

zoobank no.: urn:lsid:zoobank.org:act: C2A62183-1931-4F61-8CAA-381302E047D3

Etymology. This species is named after Nikolai Vavilov. Famous for his studies on cultivated plants, Vavilov was one of the advocates of the periodic approach in taxonomy. He died in Stalin's prison due to falsehood allegations made by agronomist Trofim Lysenko, whose work was initially supported by Nikolai Vavilov.

Type material. Holotype MIMB 42230, length 24 mm live, Kuril Islands, Iturup Island, 19 August 2019, depth 263–273 m, coll. unspecified.

External morphology. The notum is broad, and rounded in front and posteriorly. The rhinophores are retracted into raised soft sheaths bearing small tubercles (Figure 11(A1)); 14 rhinophoral lamellae. The notum is covered with small, indistinct tubercles (Figure 11(A1)). Eight multipinnate gills united by a common membrane form a circle around the anus. The gills are retractable into a common gill cavity. The border of the gill cavity is moderately raised with a tuberculated edge (Figure 11(A1)). The foot is broad.

Colour. The notum is partly semi-transparent, white, without a conspicuous yellow pigment (Figure 11(A1)). The gills are light creamy. The rhinophores are slightly darker than the notum. The digestive system is light brownish and pinkish, partly visible through the notum dorsally (Figure 11(A1)). The subepidermal glands are barely visible. The yellow line around the notum is absent. The yellow pigment dorsally or within small tubercles is barely distinguishable (Figure 11(A1)). The yellow line around the foot is absent.

Buccal bulb and oral tube. The buccal bulb and oral tube are present. The salivary glands are narrow.

Jaws. The jaws take the form of an oval labial disk covered by cuticle-bearing rod-shaped labial elements with double-hook-shaped tips (Figure 11(A4)).

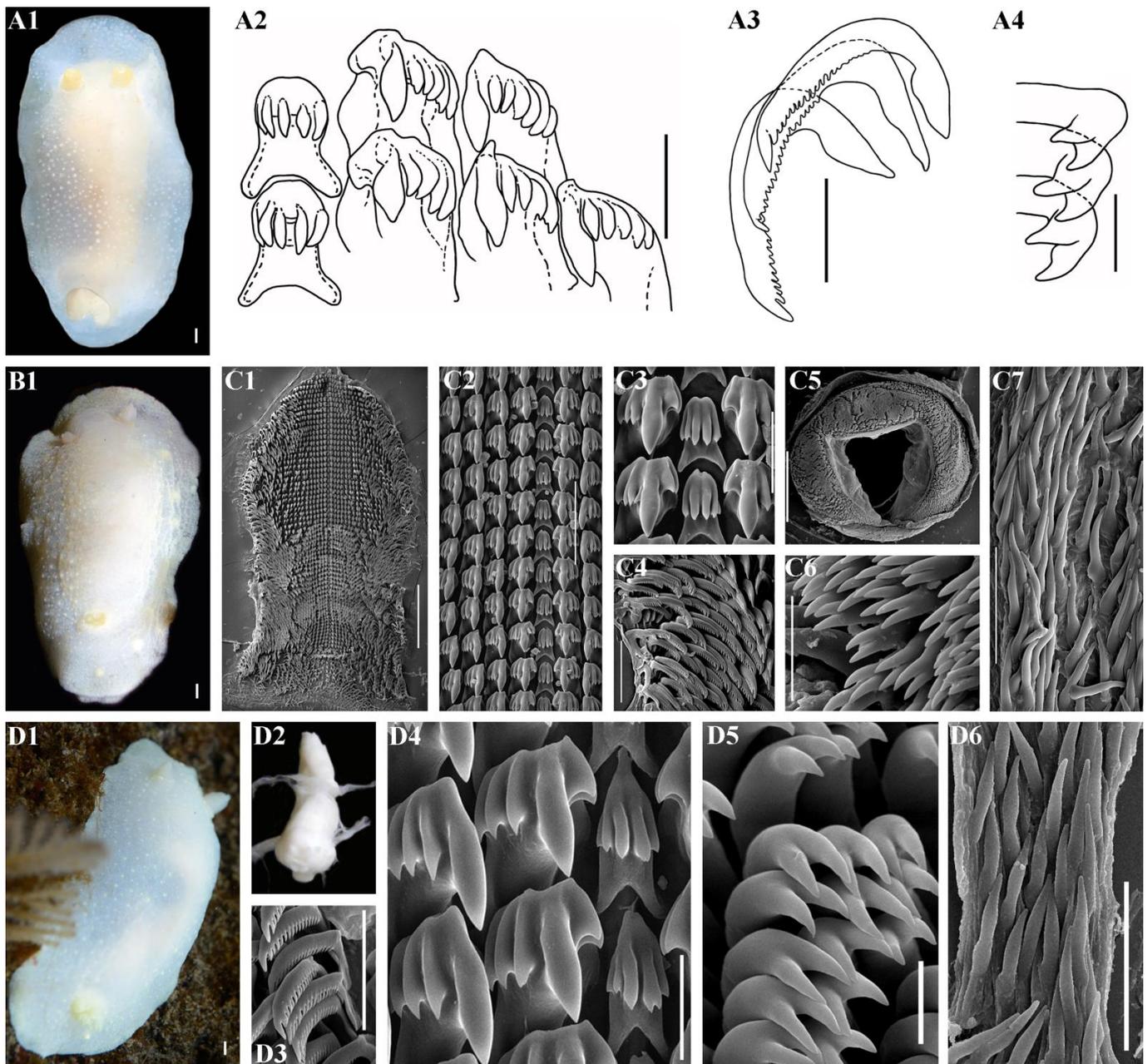


Figure 11. *Cadlina vavilovi* sp. nov. (A) and *Cadlina paninae* Korshunova et al., 2020 (B–D), external and internal features. (A) *Cadlina vavilovi* sp. nov., Holotype MIMB 42230, length 24 mm live, Iturup Island. (A1) Dorsal view, drawing. (A2) Central teeth and first lateral teeth, drawing, scale bar 30 μ m. (A3) Outer lateral teeth, drawing, 30 μ m. (A4) Elements of labial cuticle, drawing, 10 μ m (the drawings based on data [59]). (B–D) *Cadlina paninae* Korshunova et al., 2020. (B1) Holotype ZMMU Op-683, Matua Island, dorsal view. (C) Paratype ZMMU Op-684, Matua Island. (C1) Complete radula, scanning electron microscopy (SEM), 500 μ m. (C2) Central part of the radula, SEM, 100 μ m. (C3) Central teeth with first lateral teeth, SEM, 40 μ m. (C4) Outer lateral teeth, 100 μ m. (C5) Complete labial cuticle, SEM, 500 μ m. (C6) Elements of labial cuticle, SEM, 20 μ m. (C7) Copulative spines, SEM, 40 μ m. (D) Paratype ZMMU Op-685, Matua Island. (D1) Dorso-lateral view. (D2) Buccal bulb, light microscopy (LM). (D3) Outer lateral teeth, SEM, 100 μ m. (D4) Central teeth with first lateral teeth, SEM, 30 μ m. (D5) Elements of labial cuticle, SEM, 10 μ m. (D6) Copulative spines, SEM, 40 μ m. Scale bars for all living specimens are 1 mm. Photos and drawings: Tatiana Korshunova, Nadezhda Sanamyan, and Alexander Martynov.

Radula. Radular formula $79 \times 34.1.34$. The central tooth is moderately low and bears up to six cusps (Figure 11(A2)). The inner lateral teeth have up to six denticles on the outer edge, and up to three on the inner edge (Figure 11(A2)). The middle and outer teeth are comb-shaped, bearing up to 20 denticles (Figure 11(A3)).

Reproductive system. The ampulla is folded and broad; it bifurcates into moderately long vas deferens and oviduct. The prostate transits to narrow vas deferens that enclose the ejaculatory duct with copulative (penial) spines. The vagina is narrow, and the bursa copulatrix and receptaculum seminis are oval.

Habitat. The species inhabits relatively deeper waters with gravel sand substrates at depths of c. 263–273 m.

Distribution. Currently, the species is only known to occur in the coastal waters of Iturup Island (Figure 2B); however, potential distribution includes at least the neighbouring Kuril Islands.

Remarks. The species exhibits known chromatic pattern II and central teeth pattern IV. Due to the combination of indistinguishable yellow spots, the absence of the yellow line around the notum, the uniformly coloured ground notum, the relatively low central radular teeth, and the molecular phylogenetic data (Figures 1–5 and 11), this new species *Cadlina vavilovi* sp. nov. is distinguished from all known [32] and herein-described species of *Cadlina* (Figures 3–5). The species is mentioned as *Cadlina* sp. in [59]. The lowest COI intergroup distance of 4.82% is found between *C. vavilovi* sp. nov. and *C. franklinae* sp. nov. as well as between *C. vavilovi* sp. nov. and *C. paninae* (Table 1). Extended morphological data for *C. paninae* are provided here (Figures 3–5 and 11(B1,C1–C7,D1–D6)). See also the details in Section 4.

***Cadlina vinogradovi* sp. nov.**

Figures 3, 4, 5, 7E and 12

zoobank no.: urn:lsid:zoobank.org:act: 1C45ED81-E24B-487C-95C8-986D6C53ABF2

Etymology. This species is named after Dmitry Vinogradov, one of the first re-inventors of porcelain in Europe, who developed Russian hard-paste porcelain. Vinogradov died at the age of 38 without receiving an honour for his important contributions.

Type materials. Holotype ZMMU Op-839, length 60 mm live, Kuril Islands, Urup Island, 23 August 2021, depth 10–20 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-838, length 32 mm live, Kuril Islands, Urup Island, 2 September 2021, depth 10–40 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-850, length 33 mm live, Kuril Islands, Iturup Island, 5 September 2022, 10–15 m, coll. T.A. Korshunova, A.V. Martynov. Paratype ZMMU Op-851, length 12 mm live, Kuril Islands, Iturup Island, 3 September 2022, 20–30 m, coll. T.A. Korshunova, A.V. Martynov.

External morphology. The notum is broad, and rounded in front and posteriorly. The rhinophores are long and retracted into raised soft sheaths bearing small tubercles (Figure 12(A1,A3,A4,B1,C1,D1)); 12–20 rhinophoral lamellae. The notum is covered with very distinct, medium-sized, rounded or irregular tubercles (Figure 12(A1,A16,A3,A4,B1,C1,C7,D1,D7)). The spicules form a sparse network in the notum. Eight to nine multipinnate gills united by a common membrane form a circle around the anus. The gills are retractable into a common gill cavity. The border of the gill cavity is moderately raised with a tuberculated edge (Figure 12(A1,A3,A4,B1,C1,D1)). The oral veil is small, trapezoid, with oblique notched lateral sides (Figure 12(A2–A4)). The foot is broad, anteriorly rounded and slightly thickened to form a double edge. The foot appears entirely (Figure 12(A2–A4)); posteriorly, it sometimes projects slightly from the notum in crawling animals, forming a rounded tail.

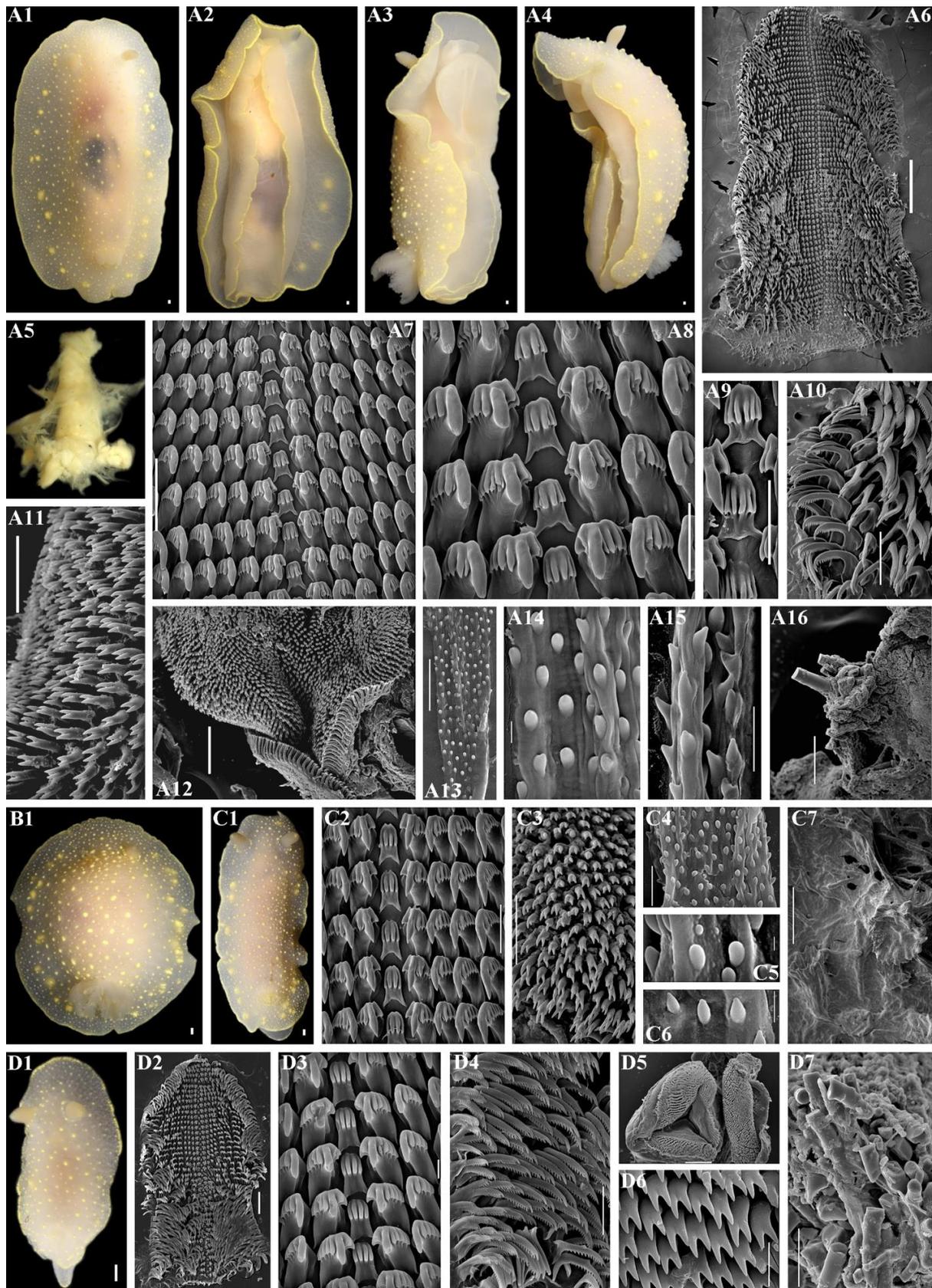


Figure 12. *Cadlina vinogradovi* sp. nov., external and internal features. (A) Holotype ZMMU Op-839, length 60 mm live, Urup Island. (A1) Dorsal view. (A2) Ventral view. (A3) Lateral view. (A4) Lateral view. (A5) Buccal bulb, light microscopy (LM). (A6) Complete radula, scanning electron microscopy

(SEM), scale bar, 500 μm . (A7) Central part of radula, SEM, 100 μm . (A8) Central teeth with first lateral teeth, SEM, 50 μm . (A9) Central teeth, SEM, 50 μm . (A10) Outer lateral teeth, SEM, 100 μm . (A11) Elements of labial cuticle, SEM, 40 μm . (A12) Labial cuticle, SEM, 100 μm . (A13) Copulative spines, SEM, 30 μm . (A14) Copulative spines, SEM, 3 μm . (A15) Copulative spines, SEM, 10 μm . (A16) Notum and spicules, SEM, 100 μm . (B1) Paratype ZMMU Op-838, length 32 mm live, Urup Island, dorsal view. (C) Paratype ZMMU Op-850, length 33 mm live, Iturup Island. (C1) Dorsal view. (C2) Central part of radula, SEM, 50 μm . (C3) Elements of labial cuticle, SEM, 40 μm . (C4) Copulative spines, SEM, 10 μm . (C5) Copulative spines, SEM, 1 μm . (C6) Copulative spines, SEM, 3 μm . (C7) Notum and spicules, SEM, 400 μm . (D) Paratype ZMMU Op-851, length 12 mm live, Iturup Island. (D1) Dorsal view. (D2) Complete radula, SEM, 100 μm . (D3) Central part of radula, SEM, 10 μm . (D4) Outer lateral teeth, SEM, 50 μm . (D5) Complete labial cuticle, SEM, 100 μm . (D6) Elements of labial cuticle, SEM, 10 μm . (D7) Spicules of notum, SEM, 50 μm . Scale bars for all living specimens are 1 mm. Photos: Tatiana Korshunova and Alexander Martynov.

Colour. The notum is partly semi-transparent or almost completely opaque, from white to pale creamy to light yellow and brownish (Figure 12(A1–A4,B1,C1,D1)). The gills are somewhat paler or darker than the notum with a patchy light-yellow pigment. The rhinophores are slightly darker than the notum. The digestive gland is black-to-brownish; the stomach and intestine are partly visible through the notum dorsally (Figure 12(A1,A3,A4,B1,C1,D1)). A pinkish area (parts of the reproductive and digestive systems) may shine through the notum in front of the stomach. The internal organs can also be almost completely invisible. A range of 14–23 subepidermal yellow glands are well conspicuous dorsally and less conspicuous ventrally (some of them are placed close to each other) from each body side. The yellow lines around the notum and the yellow pigment in the dorsal tubercles are well defined in both larger and smaller specimens (Figure 12(A1–A4,B1,C1,D1)). Together with the tubercles, the yellow spots form an extensive field of yellow pigmentation. The yellow line around the foot is somewhat more dispersed but also distinct.

Buccal bulb and oral tube. The buccal bulb is relatively short, similar in length to the oral tube (Figure 12(A5)). The salivary glands are relatively long and narrow.

Jaws. The jaws take the form of a rounded labial disk covered by a yellowish-brown cuticle bearing rod-shaped labial elements with double- to triple-pointed hook-shaped tips (Figure 12(A11,C3,D5,D6)).

Radula. Radular formula ca. 60–90 \times 20–40.1.20–40. The central tooth is moderately low and bears up to seven cusps (Figure 12(A7–A9,C2,D3)). The inner lateral teeth have up to 12 denticles on the outer edge, and up to four on the inner edge (Figure 12(A6–A8,C2,D2,D3)). The middle and outer teeth are comb-shaped, bearing up to ca. 40 denticles (Figure 12(A6,A10,D2,D4)).

Reproductive system. The ampulla comprises at least four thickened and convoluted compartments (Figure 7E, a). The ampulla bifurcates into very long vas deferens and oviduct. The uterine duct emerges some distance from the female gland mass. The prostatic part of the vas deferens is long, narrow, and only moderately distinct (Figure 7E, pr). The prostate transits to narrow vas deferens (Figure 7E, vd), which considerably widens towards the penial sheath that encloses the evertable ejaculatory duct (Figure 7E, psh). The copulative (penial) spines are significantly short, blunt, oval or slightly more triangular; elongated ones are absent (Figures 5 and 12(A13–A15,C4–C6)). The vagina narrows (Figure 7E, v) and enters a medium-sized rounded bursa copulatrix (Figure 7E, b). The uterine duct is short and narrow (Figure 7E, ud); it begins from the female gland mass and then enters near the base of the pear-shaped receptaculum seminis (Figure 7E, rs).

Habitat. The species inhabits shallow waters with rocky and stony substrates at depths of c. 10–40 m.

Distribution. Currently, the species is only known to occur in the coastal waters of the Urup and Iturup Islands (Figure 2B); however, potential distribution includes at least the neighbouring Kuril Islands and Hokkaido Island.

Remarks. The species exhibits known chromatic patterns IX–X (Figure 3), central teeth pattern IV (Figure 4), and copulative spine patterns I and II (Figure 5). Due to the combination of a significant number of yellow spots, the consistently present distinct yellow line around the notum, the moderately low central radular teeth, the very long vas deferens, the peculiar short, rounded, with occasionally partly elongated top, copulative spines (Figure 12(A13–A15,C4–C6)), and the molecular phylogenetic data (Figures 1 and 2), this new species *Cadlina vinogradovi* sp. nov. is distinguished from all known [32] and herein-described species of *Cadlina* (Figures 3–5). *Cadlina vinogradovi* sp. nov. has a remarkable pattern of short copulative spines in the adult state, whereas most other *Cadlina* species of this complex commonly have elaborated, elongated, curved spines, with only a few shorter spines in addition (Figure 5). In this regard, the copulative spines of *C. vinogradovi* sp. nov. could correspond to an earlier ontogenetic state (and, hence, evolutionary [54]) in other *Cadlina* species. The intragroup distances in *C. vinogradovi* sp. nov. are 0.61–1.06% for the COI. The lowest COI intergroup distance of 7.75% is found between *C. vinogradovi* sp. nov. and *C. laevis* from the North Atlantic (Table 1). See also the details in Section 4 below.

4. Discussion

There are several definitions of “cryptic species” that are widely circulated in the current literature. For example, Sáez et al. [42] indicated that “These molecular differences have been taken as evidence of reproductive isolation between morphologically indistinguishable (cryptic), or only a posteriori distinguished species (pseudocryptic)”. Bickford et al. [43] outlined that “We consider two or more species to be ‘cryptic’ if they are, or have been, classified as a single nominal species because they are at least superficially morphologically indistinguishable”. The latter definition is more straightforwardly emphasized in the glossary of the same work [43]: “Cryptic species: two or more distinct species that are erroneously classified (and hidden) under one species name”. Gill et al. [60] concluded that “Cryptic species are taxa that are morphologically indistinguishable and consequently often incorrectly considered as a single nominal species when in fact constituent taxa are genetically divergent and reproductively isolated from each other”.

It is immediately apparent that all these different authors strongly referred to an “indistinguishable” morphology as the main definition. Notably, while Bickford et al.’s [43] consideration of “cryptic species” was presented in a relatively balanced way (though still with a central reference to “indistinguishability”), instead, there has been a recent trend towards a too straightforward definition of the term “cryptic species”. For example, Walters et al. [61] define the term as “those that lack distinguishing morphological characters”; Kramina et al. [11] state that “Bickford et al. (2007) defined cryptic species as two or more species which are sufficiently distinct based on molecular or other evidence, but classified as a single nominal species, because they are at least superficially morphologically indistinguishable”; and in Yin et al. [28] writes that “. . . cryptic species are morphologically indistinguishable. . .”. The tendency to overuse the term “cryptic species” is notable in that studies highly critical of the “cryptic species concept” have been cited inaccurately, without mentioning the underlying criticism of the “cryptic concept” [61,62].

Recently, the combination of molecular analysis with a phyloperiodic framework as a set of practical methods [7,29,56] has been used in order to break with those notorious indications of the “impossibility of morphological distinguishing” in closely related species complexes [43]. This approach is considerably simpler in practical use than, for example, the complex, although otherwise very useful, method of geometric morphometrics [2,63] because commonly used taxonomic characters in a given group (which are usually declared “indistinguishable”) are presented explicitly, allowing fine morphological details to be searched by eye (Figures 3–5). Thus, such a phyloperiodic framework can be used by both scientists and various practitioners (for example, citizen scientists) when it is necessary to distinguish species of a given complex/group in the field. Although this is still a

complicated task, we have already successfully applied the phyloperiodic approach to another complex case: the dorids of the genus *Polycera* [7].

This is very evident in the case of *Cadlina*, when the putative “chaos” of external colouration (typically presented for nudibranchs [64]) represented in ordered rows (in combination with the molecular phylogeny, Figures 1 and 3–5), in most cases it reveals fine-scale differences, as well as the presence or absence of some particular patterns (variants) within a species (Figures 3–5). For example, *Cadlina vinogradovi* sp. nov. has a significant number of yellow spots on the notum (Figure 12(A1,B1,C1)) compared to other species of the complex (Figure 3), although partly similar in that respect to *Cadlina lomonosovi* sp. nov., which commonly includes significantly more pale variants (Figure 10(A1,B1,D1,F1,G1)), but also more intensively coloured (Figure 10(E1)). *Cadlina koltzovi* sp. nov. has a more translucent notum (Figure 9(A1,B1,C1,D1,E1,F1)) compared to the more opaque notum in *C. franklinae* sp. nov. (Figure 8(A1,B1,C1,D1,E1)). Moreover, there are clear differences between *C. vinogradovi* sp. nov., *C. koltzovi* sp. nov., *C. franklinae* sp. nov., *C. vavilovi* sp. nov., as well as in *Cadlina laevis*, *C. kamchatica*, *C. paninae*, and *C. umiushi* regarding the amount and distinctness of the yellow notal line (Figure 3). *Cadlina laevis*, a type species of the genus *Cadlina*, despite showing a wide range of chromatic patterns (I, III, IV, VI, and VIII–X, Figure 3), it only presumably overlaps with several North Pacific Ocean species because forms with distinct yellow notal line are rare [32] (Figure 3). Whereas in species like *C. vinogradovi* sp. nov., *C. lomonosovi* sp. nov., *C. bellburnellae* sp. nov., and *C. umiushi* instead, a distinct yellow notal line is very common (Figure 3). In its turn, *Cadlina lomonosovi* sp. nov. and *C. bellburnellae* sp. nov. are more similar to *C. umiushi*, but the latter species invariably has a very distinct yellow notal line, whereas *C. lomonosovi* sp. nov. and *Cadlina bellburnellae* sp. nov. may include forms with a less distinct notal yellow line, and also generally have more yellow spots on the notum than *C. umiushi* (Figure 3). *Cadlina franklinae* sp. nov. and *Cadlina paninae* so far have not been revealed as commonly having distinct yellow notal lines [32] (Figure 3). These apparently subjective qualitative characteristics can be objectively represented within a phyloperiodic framework (Figure 3) and can then significantly facilitate species identification.

We conclude that the incorporation of morphological characters in the PhyloPeriodic Tables is a practical method for detecting diagnosable characters within apparently “cryptic species”. A molecular phylogeny allows ordering the morphological characters in a multilevel framework [7,29,65] (Figures 3–5). While the resolved phylogeny provides supported hypotheses of monophyly (Figure 1), the use of the phyloperiodic framework provides a more rigorous study of the morphology (Figures 3–12). Thus, the periodic framework is not just a theoretical addition to existing practical methods, but also a key element for truly integrative study. Currently, in applying the controversial concept of “cryptic species”, a substantial part of the diversity remains unnamed [66], whereas the phyloperiodic approach and multilevel organismal diversity (MOD) method allow us to name each particular small unit of diversity. This is a very important practical achievement (including conservation biology) that immediately distinguishes the phyloperiodic and MOD approaches from the notion of “integrative taxonomy” since the latter does not necessarily recognize morphodiversity and does not necessarily allow all molecularly distinct groups to be named [34,44]. Even taking into account all biological irregularities, the phyloperiodic representation greatly streamlines any further potential variations that may be discovered and significantly facilitates species identification. These features are directly related to predictability, which is an essential characteristic of any periodic system [57].

Furthermore, naming as small units of biodiversity as possible using the phyloperiodic approach is very important for other fields such as biogeography and climatology, and directly relevant to a key recent issue such as global warming [67–69]. For example, in this study we discovered very interesting patterns associated with the overall distribution of sea temperatures: *Cadlina franklinae* sp. nov. is restricted so far only to the Urup and Chirpoy Islands, the Middle Kuril Islands (Figure 2), where at the depth of 10 m the mean annual temperature is +3.64 °C ... +5.64 °C [70], whereas *C. lomonosovi* sp. nov. sp. and

C. bellburnellae sp. nov. were found only at the most southern Kuril Islands, Iturup and Kunashir (Figure 2), where at the depth of 10 m the mean annual temperature is $+5.62\text{ }^{\circ}\text{C}$. . . $+8.88\text{ }^{\circ}\text{C}$ [70], thus, the average ca. is 1.5 times higher than in Urup Island. *Cadlina vinogradovi* and *Cadlina koltzovi* sp. nov. in turn occur both in cold water (Urup, Chirpoy) and temperate water (Iturup, Kunashir) in the Kuril Islands (Figure 2). We have detailed sampling for two years, so we can say with confidence that these patterns are not due to insufficient collection efforts since, given the diversity of locations around the islands and the number of specimens, we have never found *C. franklinae* sp. nov. further south than Urup Island, and we have never found *C. lomonosovi* sp. nov. and *C. bellburnellae* sp. nov. further north than the Iturup and Kunashir Islands. *Cadlina vavilovi* sp. nov. occurs at Iturup Island (Figure 2) in a deeper location, c. 250 m, with mean annual temperature at this depth considerably low, $+1.0\text{ }^{\circ}\text{C}$. . . $+1.93\text{ }^{\circ}\text{C}$ [70]. The $+12\text{ }^{\circ}\text{C}$ isotherm in the south direction was indicated precisely at the borders of the Urup and Iturup Islands [71].

Thus, the fine-scale morphological, molecular, and biogeographical differences in this species complex identified in the present study are in broad agreement with the well established geophysical data. All these environmental peculiarities, coupled with the direct development known in the related species *C. laevis* [32], may imply similar modes of development in other species in this complex and therefore limited dispersal ability. These processes may contribute to the formation of a significant amount of fine-scale species-level diversity. These interesting patterns can be used in planning conservation activities since the natural reserve on the Kuril Islands is planned to be expanded to all the islands [31]. Furthermore, due to the ongoing global warming, it will be important to check if the ranges of colder water species in this complex, such as *C. franklinae* sp. nov., will shift southward, whereas ranges of more temperate species, like *C. lomonosovi* sp. nov. sp. and *C. bellburnellae* sp. nov., will shift northward instead. Without the coherent morphological and molecular framework presented here using the phyloperiodic approach, species-related information of such importance to biogeography, climatology, and conservation would simply be lost in the “cryptic species” approach, which lumps all the diversity into a single “undistinguishable” assemblage.

Ultimately, the phyloperiodic framework removes the term “cryptic species”. In this study, we show this explicitly: within the uncovered diversity of the genus *Cadlina*, which is supposedly “completely cryptic”, we can indicate, for example, that *C. vinogradovi* sp. nov. readily distinguishes from all other new species described here (*C. koltzovi* sp. nov., *C. franklinae* sp. nov. sp., *C. vavilovi* sp. nov., *C. lomonosovi* sp. nov., and *C. bellburnellae* sp. nov.), as well as the previously described *Cadlina laevis*, *C. kamchatica* [32], *C. koreana* (external and radular drawings on Figures 2–4 for the latter species are based on [72]), *C. paninae*, and *C. umiushi* [32] based on the shape of copulative spines (Figures 5 and 12(A13–A15)). However, the radular morphology (central teeth) and external features of *C. vinogradovi* sp. nov. remain “cryptic” (in the currently accepted erroneous meaning) compared to the above-listed species (Figures 4 and 12(A8,C2,D3)). In turn, *Cadlina koltzovi* sp. nov. is well distinguished from all of the species listed, with the exception of *C. koreana*, by relatively high central teeth of radula (Figures 4 and 9(A5,A6,D6,F2)), but remains “cryptic” by the external features and copulative spines (Figures 3, 5 and 9(A1,B1,D1,E1,F1)). Several more species, like *Cadlina bellburnellae* sp. nov., may also include relatively high central teeth, but they differ from other species by the fine-scale details of the central teeth (Figure 4). The type species of the genus *Cadlina*, *C. laevis*, which also belongs to the studied clade (Figure 1), externally includes almost all colour varieties, which can be found in any other member of this clade (Figure 1), even such distinct species such as *C. umiushi* and *C. kamchatica* (Figure 3), and according to the radula and copulative spines, *C. laevis* is “cryptic” to the majority of these species except *Cadlina vinogradovi* sp. nov., *C. koltzovi* sp. nov., and *C. koreana* (Figures 3 and 4). *Cadlina franklinae* sp. nov., *C. paninae*, and *C. vavilovi* sp. nov. show more subtle differences in external, radular, and copulative spine morphology (Figures 3–5), and compared to some variants of *Cadlina laevis*, but not all of them (Figure 3). The North Atlantic *Cadlina laevis* clearly does not overlap the range of the central radular teeth and

copulative spines with the North Pacific Ocean *C. vinogradovi* sp. nov., *C. koltzovi* sp. nov., *C. vavilovi* sp. nov., *C. lomonosovi* sp. nov., and *C. bellburnellae* sp. nov. (Figures 4 and 5).

This series of comparisons can be continued, but it is already possible to point out a key consideration for the crucial problem of “cryptic species”. We can designate this complex as “cryptic”, but then some of these species exhibit distinct internal features (Figure 4, Figure 5, Figure 9(A5,A6,D6,F2) and Figure 12(A13–A15)). We may attempt to designate some of these species as “cryptic” by external features, e.g., *Cadlina franklinae* sp. nov. and *C. vavilovi* sp. nov., but the type species *Cadlina laevis* will include some “cryptic” specimens and some “non-cryptic” specimens (Figure 3). Then, we can designate several species in this complex as “cryptic” in terms of internal features, whereas *Cadlina vinogradovi* sp. nov. and *C. koltzovi* sp. nov. can be designated as “non-cryptic”. However, according to external features, *Cadlina vinogradovi* sp. nov. and *C. koltzovi* sp. nov. will remain “cryptic”, and *C. koltzovi* sp. nov. will also remain “cryptic” by radular morphology. In addition, most of the species newly described here, as well as *Cadlina umiushi*, can be externally considered as cryptic to *Cadlina luteomarginata* and *Cadlina klasmalmbergi*, which definitely belong to other clades and are easily distinguished by radular and reproductive characters [32]. Considering that different species from this complex have various degrees of molecular–phylogenetic distances (Figures 1 and 2) and morphological distinctness (Figures 4 and 5), a truly multilevel system of fine-scale differences and similarities is revealed (Figures 3–5). This is very instructive because when comparing the horizontal rows of the phyloperiodic tables IX–XI containing the larger, darker, and ontogenetically more advanced specimens, the fine-scale differences in the colouration patterns between most of the species become evident (Figure 3).

Therefore, to designate any of these species as “cryptic”, “pseudocryptic”, or non-cryptic would be completely arbitrary, non-operational, and generally meaningless. By this, we finally strongly propose that the concept of “crypticity” and the term “cryptic species” be removed from the arsenal of modern biology, including phylogeny and taxonomy. These terms no longer help to reveal the enormous biological diversity, but, on the contrary, significantly hinder its further exploration. Prospects for other groups of organisms, which do not always have such complex morphological features as nudibranchs, e.g., protists [42], should not be used as a justification of the rationale for the term “cryptic species”. This is also due to the fact that any statements about the “morphologically indistinguishable species” have no biological basis since there cannot be two completely identical biological organisms, even twins [5,7,73], so “cryptic twin species” should not be excluded from that strict biological rule. There is growing dissatisfaction with the concept of “cryptic species” in various studies of disparate organismal groups, such as plants and animals [3,6,8,26,74,75], and more recently species for which morphological data can be provided have been highlighted as “non-cryptic” [76]. There are also a number of studies [77–84] in which the more neutral term “hidden diversity” has already been used, which does not overlap semantically with “cryptic species” in its original meaning: “well camouflaged on a substrate and in an environment” [7,9,85–87]. However, there is also a confusing mixed use of both “cryptic” and “hidden” terms [88–92].

Rather than continuing to rely on the straightforward notion of “crypticity”, the proposed multilevel periodic-like framework is able to provide a reliable method for displaying and distinguishing even very subtle differences, both external and internal, within a given *Cadlina* species complex (Figures 1–12). The phyloperiodic approach makes it possible to perform a “granular”, fine-scale comparison of every specimen, whereas the “cryptic species approach” instead generally asserts that some species morphologically are completely “indistinguishable” [42]. This is consistent with the growing understanding that the conventional “species concept” represents a considerable overgeneralization and that more organism-focused approaches need to be implemented [29]. Potentially, the phyloperiodic approach can be expanded to more features, but this is definitely a task for the next studies, as even the current framework is quite complex to implement. If new material becomes available, future studies will test the framework developed here,

and, for example, more periodic rows and columns can be recognized. This approach can be universally applied to any other groups of organisms, using appropriate diagnostic characters in the ontogenetic framework of each given group in combination with molecular phylogenetic data. The periodic approach in various forms opens up a new avenue for the further development of various practical applications in biology and has begun to be applied, for example, in the fields of ecology [93,94] or biomolecules [95]. Significant efforts remain to be made in the taxonomy to ensure widespread acceptance of the periodic approach. This is especially important since systematics is the inevitable basis for all other areas of biology.

The traditional, still widely used, rigid taxonomic diagnoses need to be gradually replaced with the fine-scale multilevel presentation (Figures 3–5), and the most recent studies are in line with this emerging agenda [74,96]. The patterns of natural biodiversity have not been established according to pre-evolutionary, strict diagnosis-based “box-thinking”, but instead reflect the most complex, dynamic, constantly evolved biological organisms.

Thus, one of the most important messages of this study for any taxonomic, phylogenetic, and biodiversity studies is the following: without the phyloperiodic framework and multilevel understanding of biological diversity, all numerous and existing, fine-scale diversity units will be just considered as a chaotic “cryptic species”.

5. Conclusions

A morphologically difficult-to-distinguish species complex of dorid of the genus *Cadlina* was discovered in the little-studied Kuril Islands in the North Pacific Ocean and used as a model system to show persistent pitfalls in the common usage of the “cryptic species” concept.

Analysing the diagnostic characters among the newly described and related species in the phyloperiodic framework, it is clearly shown that dividing them into “cryptic” and “non-cryptic” taxa is completely arbitrary and meaningless.

Applying our novel data and analysing the current literature on the widely cited problem of “cryptic species” for a wide range of different groups of organisms, we strongly propose that the notion of “crypticity” and the term “cryptic species” be excluded from the arsenal of modern biology. This is because the concept misleadingly divides overwhelming biological diversity into “cryptic” and “non-cryptic” components and, rather than emphasizing the multilevel nature of organismal diversity, artificially reduces it to two simple, supposedly contrasting modes.

The implications of the present study are directly relevant to studying the diversity, phylogeny, biogeography, conservation, and many other aspects of any group of organisms.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/d16040220/s1>, Table S1. PCR amplification options. Table S2. GenBank accession numbers and references for all sequences used in this study.

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