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**Abstract:** Impoundments can drastically change the physical and biological characteristics of fluvial systems. Changes in the physical characteristics, such as reductions in flow, increased sediment deposition, and increased surface area, often influence the system's biological components, including plant, macroinvertebrate, and fish assemblages. In addition to having direct effects on impounded waterbodies, impoundments can also have wide-ranging effects at the watershed scale, particularly on upstream tributary streams. The purpose of this study was to assess the magnitude of these effects. We analyzed historical data from 26 streams distributed across five sub-basins in the Bluff Hills region of the Yazoo Basin, MS, USA. All five major tributary rivers in this region are impounded by large (11,240–26,143 hectares) reservoirs for flood control. We compared fish assemblages in streams located upstream and downstream of the four reservoirs using PERMANOVA, and contrary to expectations, we found no significant differences between the upstream and downstream assemblages. We explore several possible explanations for this discrepancy and suggest that stream assemblage response to impoundment may be nuanced by the regional species pool, the history of stream conditions in the watershed, and the resistance of the streams to periodic disturbances.

Keywords: impoundments; community ecology; stream ecology

# 1. Introduction

Because of its unique biogeographic history, the southeastern United States is an epicenter of fish biodiversity in North America [1,2]. Yet, many streams in the region have experienced significant habitat degradation due to anthropogenic watershed alterations, including deforestation, channelization, and dam construction [3,4]. Dams are commonly used to control the flow regimes of rivers, relying on both small impoundments and large flood control reservoirs to mitigate flooding caused by the extensive precipitation from the southeastern United States receives annually. Over 17,000 dams and reservoirs have been constructed in the Tennessee, Lower Mississippi, and South Atlantic Gulf River basins [5,6]. Reservoirs may influence stream fish assemblages through changes in habitat, hydrology, and isolation effects, although disentangling reservoir effects from other factors that influence patterns in fish assemblage structure remains a challenge.

Reservoirs change stream conditions throughout impounded watersheds. Immediately upstream of the impoundment, the rivers and tributaries gain lentic properties: flow is reduced or eliminated, width and depth increase, and suspended sediment settles out of the water column. Stream habitat in the lower reaches of the tributary streams can be converted to wetland habitat as fine sediment aggrades and riparian vegetation spreads into the channel, leading to high production of zooplankton and phytoplankton. Downstream of the impoundment, the sequence of flooding events often becomes decoupled from the natural flow regime due to reservoir releases that, depending on how water storage and the outlet are engineered and managed, may alter temporal flows and physicochemical properties



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of the water such as turbidity, temperature, and dissolved oxygen levels [7]. Because the outlet of the dam is constrained within a defined bed, the river downstream of the reservoir is often channelized. The decrease in riverbed elevation caused by channelization can cause headcuts (i.e., downcutting of the stream bed in an upstream direction) that degrade tributaries [8–10]. As streams become incised, instream habitat progressively changes due to cycles of scouring and aggradation [9,11,12]. As the stream banks become steeper, connection to the floodplain and associated wetlands is reduced, which can also have implications for the biotic community [13]. All these physical changes can drastically change the type, amount, and quality of aquatic habitat available and fish assemblage diversity and composition.

In addition to changes in habitat, reservoirs can play a direct role in altering fish assemblages by influencing the connectivity between populations. Winston et al. [14] reported four native fluvial specialist cyprinids absent upstream of a dam on the Red River in Oklahoma despite their presence elsewhere in the drainage. They conjectured that as the intermittent streams above the dam desiccated during dry seasons, the cyprinid species migrated into the reservoir, where they became naïve prey to piscivores, which are often abundant in reservoirs. The loss of longitudinal connectivity caused by the dam prohibited recolonization by downstream populations. Species with drifting larvae may also be adversely affected by fragmented habitat, as some may require large reaches of free-flowing river habitat and be unable to persist when their larvae or eggs drift into a reservoir and are preyed on or settle on the substrate prematurely. This shift in fish assemblage has been supported by multiple studies that have reported extirpations, higher representation of generalists, and shifts in fish assemblages upstream of reservoirs [15–21]. Isolation effects and community shifts have also been documented for other aquatic species, including crayfish [22].

We investigated the degree to which fish assemblages in the Yazoo Basin, a major tributary to the Mississippi River, had been impacted by large flood control reservoirs. We expected that the reservoirs would have diminished fish diversity and altered assemblages by reducing network connectivity and changing habitat quality above and below the reservoirs. Specifically, we expected that fish assemblages in tributaries upstream of a reservoir would include a greater representation of tolerant lentic generalists, especially piscivorous species common in reservoirs in southeastern North America, while the assemblages in tributaries downstream from the influence of the tailrace of the dam would retain a fluvial assemblage complete with a greater emphasis on intolerant stream specialists. Moreover, we expected species diversity to be lower above the reservoirs as occasional droughts, siltation, or changes in other environmental conditions following over half a century of impoundment may have produced localized extirpations due to the lack of recolonization from downstream reaches blocked by dams.

#### 2. Materials and Methods

#### 2.1. Study Area

The study was conducted in streams in the Bluff Hill region of the eastern Yazoo Basin in north Mississippi, USA (Figure 1). This region consists of six subbasins that discharge into the Yazoo River. Five of the rivers in these six subbasins (the Coldwater, Little Tallahatchie, Yocona, Skuna, and Yalobusha) are impounded by four flood-control reservoirs (the Arkabutla, Sardis, Enid, and Grenada reservoirs) built between 1938 and 1956 and ranging in size from 4800 to 14,500 ha. These reservoirs are not equipped with fish passage facilities. The annual discharge of the five rivers upstream of the impoundments ranges from 17 to 51 m<sup>3</sup>s<sup>-1</sup> [23]. Streams in the region have a long history of erosion and sedimentation due to the highly erodible loess soils and the conversion of the native hickory-oak forests into agricultural lands in the mid to late 1800s [24,25]. Federal programs were introduced in the 1940s to reduce overland erosion and have been largely successful, and other programs were initiated in the 1980s to reduce instream erosion [26,27]. The regional species pool in the study streams includes over 50 fish species, including two endemics, the

Yazoo Shiner *Notropis rafinesquei* and the Yazoo Darter *Etheostoma raneyi* [28,29]. Commercial fishing in the Yazoo Basin only occurs downstream of the study area, and to our knowledge, the four reservoirs are not enrolled in a fish stocking program.



**Figure 1.** Map of the eastern Yazoo River Basin in northwest Mississippi, USA. Sites sampled by Arkansas State University in 1999–2000, coded by color. Upstream sites are gray, while downstream sites are black.

## 2.2. Site Selection

We included sites upstream and downstream of the study reservoirs in most subbasins (Figure 1). To bolster the representation of the downstream fish assemblages, we also included sites from two neighboring unimpounded subbasins, the Tallahatchie River and Upper Yazoo River, that contain tributary streams that flow directly into the Yazoo River downstream of the four reservoirs. Sampling sites were further selected such that (1) no two sites were on the same stream, (2) if grade control structures installed to minimize instream incision [10] were present, sampling sites were established at least 2.5 km downstream

of the structures, and (3) sites were located at least 50 m upstream of a road crossing to avoid potential confounding effects associated with road construction. In all, 26 sites were included. Fourteen of the sites were located on tributary streams 1–81 km upstream of the four reservoirs. Twelve sites were located on streams that joined the mainstem rivers at least 3 km downstream of the reservoir tailwaters and positioned 2–31 km above the confluence of the tributary and the river discharging from the reservoir.

No site descriptions were available with this archival dataset. However, in a recent study conducted in the same streams, although not at the same sites, Faucheux [30] reported that streams average 8.3 m in wetted width and 0.7 m in maximum water depth. The substrates are approximately 60% sand and clay, 10% hard clay, 10% silt, and 20% gravel, cobble, and riprap. There were no obvious differences in sites upstream or downstream of the reservoirs. Moreover, for each sampling site in this study, we estimated the catchment area using the StreamStats program [31]. Catchment area indexes stream size and discharge as precipitation quantities are consistent across the region, and for the study sites, catchment area averaged 111.4 sqkm (1.9–556.8 sqkm).

### 2.3. Fish Sampling

Fish collections consisted of a 200-m backpack electrofishing pass, with a target electrofishing time of 20 min (mean = 23 min, SD = 14 min). Sampling during June—September 1999 and 2000 (i.e., two sampling seasons) coincided with baseflow conditions. At each site, specific conductance was measured before sampling, and the voltage on the backpack unit was adjusted to maintain a relatively constant power. Sampling was conducted in a zig-zag pattern in an upstream direction and covered all types of habitats present. Two netters accompanied the backpack electrofisher to retrieve fish affected by the electric field. After a sample was completed, fish larger than 10 cm in total length were identified by species and returned to the stream. Smaller fish were anesthetized in a solution of MS-222, preserved in 10% buffered formalin, and transported to a lab for identification.

Backpack electrofishing is one of the most commonly used methods for characterizing fish assemblages in streams, although it can be biased by differences in detection probabilities across sites and species [32,33]. Even though capture efficiency tends to increase with fish size, which can result in under-sampling smaller-bodied species [32,34,35], electrofishing typically captures more species compared to other single-gear stream sampling methods [36]. Backpack electrofishing is particularly effective for collecting centrarchid species that seek cover in large woody debris [33]. However, benthic fish (e.g., darters, *Ethesosthoma* spp.) have lower capture efficiency due to their small size and lack of a gaseous swim bladder that prevents them from surfacing when stunned. Conversely, minnows (Cyprinidae) may be under-sampled due to the difficulty of thoroughly netting large schools and the reduced effectiveness of electrical fields on small-bodied fish [34,37]. These difficulties can bias fish collections by preferentially selecting larger species. It is likely that collection biases affected our characterization of the diversity and composition of fish assemblages, but it is less likely that they affected our upstream versus downstream comparisons because sampling was standardized among all sites.

#### 2.4. Analysis

The analysis was designed to assess the potential effects of the reservoirs on stream fish assemblages while accounting for differences in river subbasin and stream size. Fish data were standardized as catch per unit of effort (CPE), defined as the number of individuals of a given taxon collected divided by the time needed to complete sampling of the stream reach. We applied a multivariate analysis of covariance to test if species CPEs (multiple continuous variables) differed between tributaries upstream and downstream of the reservoirs (categorical variable), while controlling for subbasin (random categorical variable to control for potential differences in species pool across the region) and catchment size (continuous variable to account for differences in stream size). Subbasins in the region could potentially support different fish in each subbasin assemblage, and catchment size influences discharge and stream volume, as well as a multitude of other accompanying physical stream characteristics that shape fish assemblages at any particular site [38,39]. The catchment size for each site was obtained using the StreamStats program [31] and log<sub>10</sub>-transformed to linearize catchment areas that tend to increase exponentially. The multivariate analysis of covariance was run with a permutation MANCOVA (PERMANCOVA) applied to an among-sites similarity matrix computed with the Bray-Curtis similarity index implemented on the transformed CPE values. Species CPE were fourth root transformed to reduce right-skewness. Non-metric multidimensional scaling (NMDS) was also applied to the resemblance matrix to interpret graphically the results of the PERMANCOVA.

Research has suggested that analysis of assemblages organized by functional group rather than taxonomic group can highlight relationships between fish assemblages and environmental conditions [40], especially changes in stream conditions due to dams [41]. To assess our hypotheses regarding the higher CPE of tolerant, lentic generalists in the upstream tributaries and higher lotic specialists in the downstream tributaries, we repeated the multivariate PERMANCOVA and NMDS analyses outlined above but replaced the taxa CPE values with functional group descriptors including physicochemical tolerances, habitat preferences, and trophic guilds (Table 1). Tolerances (i.e., intolerant, moderate, tolerant) were assessed according to Meador and Carlisle [42], habitat preferences (generalist, lentic, lotic) according to Frimpong and Angermeier [43], and trophic guilds (detritivore, herbivore, invertivore, parasite, piscivore, planktivore) according to Goldstein and Simon [44]. For all three of these functional categorizations, species CPE values were summed across samples according to each category, and then category CPEs were standardized to percentage composition across each sample. These values were then square-root transformed to reduce right-skewness. For each functional categorization, an among-sites matrix was constructed with the Bray-Curtis similarity index. Log<sub>10</sub>-transformed catchment size was included as a covariate because species differences in trophic guilds, tolerances, and habitat preferences are expected to change with increasing stream size [45], but subbasin was not included as a covariate since analysis of functional guilds generalizes species' identity to their traits, which allows for community comparisons between areas that are geographically distant [46]. PRIMER-E version 7 software [45] was used for all analyses.

Because one of the primary ways reservoirs can affect stream fish assemblages is by acting as barriers to recolonization after stochastic extinction events, we hypothesized that tributaries upstream of the reservoir would have lower species richness than downstream tributaries. Species richness at each site was estimated using individual-based rarefaction or Chao 1 extrapolation as needed to facilitate comparisons between sites with unequal sample sizes [47]. As noted by Colwell et al. [48], extrapolation provides reliable estimates only up to roughly double the size of a sample, so we rarified or extrapolated all samples to a sample size of 50 fish (smallest catch was 27 fish). Rarefaction and extrapolation estimates were calculated using the *iNEXT* package in program *R* [49,50]. Potential differences in species richness among sites above and below reservoirs were assessed using a permutational analysis of covariance (PERANCOVA) applied to an among-site similarity matrix computed with Euclidean distance, with  $log_{10}$ -transformed catchment size as a covariate [51]. Subbasin was not included as a covariate in this analysis because previous surveys have indicated that the species pool does not differ in number. The PERANCOVA was applied using PRIMER-E version 7 software [52].

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Table 1. List of species caught in the 26 study sites in the Yazoo Basin, MS, USA. The values represent the number of sites where each species was detected. The percentage of catch was calculated using the total number of fish collected in the study. Asterisks (\*) denote migratory species. Migration habits and habitat preferences were assessed using Frimpong and Angermeier [43]. The trophic guild was assessed using Goldstein and Simon [44]. Tolerance was assessed using Meador and Carlisle [42].

Scientific Name	Common Name	Upstream Sites	Downstream Sites	Percentage of Catch	Habitat Preference	Trophic Guild	Tolerance
Ichthyomyzon castaneus	Chestnut Lamprey	1	0	0.18	lotic	parasite	intolerant
Atractosteus spatula	Alligator Gar	1	0	0.02	lentic	piscivore	tolerant
Lepisosteus oculatus	Spotted Gar	1	4	0.27	lentic	piscivore	tolerant
Dorosoma cepedianum	Gizzard Shad	2	0	0.31	generalist	herbivore	tolerant
Dorosoma petenense	Threadfin Shad	2	0	0.47	generalist	planktivore	tolerant
Campostoma anomalum	Central Stoneroller	0	2	0.07	lotic	herbivore	moderate
Cyprinella camura	Bluntface Shiner	6	10	11.02	lotic	invertivore	moderate
Cyprinella lutrensis	Red Shiner	1	2	1.36	generalist	invertivore	tolerant
Cyprinella venusta	Blacktail Shiner	9	9	9.44	lotic	invertivore	moderate
Cyprinus carpio	Common Carp	2	0	0.04	generalist	detritivore	tolerant
Luxilus chrysocephalus	Striped Shiner	2	4	2.68	lotic	invertivore	moderate
Lythrurus fumeus	Ribbon Shiner	0	1	0.02	lotic	invertivore	intolerant
Notemigonus crysoleucas	Golden Shiner	3	0	0.09	generalist	invertivore	moderate
Notropis ammophilus	Orangefin Shiner	2	3	0.94	lotic	invertivore	intolerant
Notropis atherinoides	Emerald Shiner	4	0	2.05	generalist	planktivore	tolerant
Notropis buchanani	Ghost Shiner	1	0	0.29	generalist	invertivore	tolerant
Notropis rafinesquei	Yazoo Shiner	1	3	4.60	lotic	invertivore	intolerant
Dimembalas mototus	Pugnose Minnow	1	0	0.04	generalist	detritivore	moderate
Pimephales notatus	Diuntnose Minnow	6	9	4.13	generalist	detritivore	tolerant
Pimephales olgitax	Graal Chub	5	5	0.47 E 29	generalist	invertivore	tolerant
Semotitus atromaculatus	Creek Chub	8	8	5.58	lotic	invertivore	tolerant
Erimyzon obiongus	Smallmouth Buffalo	5	4	0.76	ronoralist	invertivore	tolorant
Monostoria om the man	Coldon Rodhorso	0	4	0.20	generalist	invertivore	tolerant
Moxostoma poscilurum	Blacktail Rodhorse	1	0	0.02	lotic	dotritivoro	intolorant
Ameiurus malas	Black Bullboad	0	5	0.51	ronoralist	invertivore	tolorant
Ameiurus metalia	Vollow Bullboad	1	11	0.13	lotic	invertivore	tolorant
Ameiurus natulosus	Brown Bullhood	9	11	4.39	lontic	invertivore	tolerant
Ictalurus numetatus	Channel Catfish	1	1 7	0.02	conoralist	nivertivore	tolorant
Noturus ourinus	Tadpole Madtom	1	0	0.04	lotic	invertivore	tolerant
Noturus kildebrandi	Least Madtom	2	0	0.04	lotic	invertivore	intolerant
Noturus nocturnus	Freckled Madtom	2	0	0.09	lotic	invertivore	tolerant
Pulodictis olivaris	Flathead Catfish	0	2	0.20	lotic	piscivore	tolerant
Fsor americanus	Redfin Pickerel	1	0	0.02	lentic	piscivore	moderate
Anhredoderus savanus	Pirate Perch	4	0	1.03	lentic	invertivore	moderate
Lahidesthes sicculus	Brook Silverside	1	1	0.42	lentic	planktivore	tolerant
Fundulus chrysotus	Golden Topminnow	1	1	0.33	lentic	invertivore	moderate
	Blackstripe	-	_	0.00	Territe		incucrute
Fundulus notatus	Topminnow	1	7	1.96	lotic	invertivore	tolerant
Fundulus olivaceus	Topminnow	10	10	4.08	lotic	invertivore	moderate
Gambusia affinis	Mosquitofish	4	3	0.85	generalist	invertivore	tolerant
Morone chrysops *	White Bass	1	0	0.02	generalist	piscivore	tolerant
Lepomis cyanellus	Green Sunfish	12	11	11.40	generalist	invertivore	tolerant
Lepomis gulosus	Warmouth	4	1	0.18	generalist	invertivore	tolerant
Lepomis macrochirus	Bluegill	12	12	12.65	generalist	invertivore	tolerant
Lepomis megalotis	Longear Sunfish	4	10	8.48	generalist	invertivore	tolerant
Lepomis miniatus	Redspotted Sunfish	1	1	0.04	generalist	invertivore	moderate
Micropterus punctulatus	Spotted Bass	2	6	0.65	generalist	piscivore	tolerant
Micropterus salmoides	Largemouth Bass	6	9	3.86	generalist	piscivore	tolerant
Pomoxis annularis	White Crappie	2	0	0.11	lentic	piscivore	tolerant
Etheostoma artesiae	Redspotted Darter	1	2	0.11	lotic	invertivore	moderate
Etheostoma histrio	Harlequin Darter	1	0	0.04	lotic	invertivore	intolerant
Etheostoma lynceum	Brighteye Darter	0	2	0.13	lotic	invertivore	intolerant
Etheostoma parvipinne	Goldstripe Darter	1	1	0.07	lotic	invertivore	intolerant
Etheostoma swaini	Gulf Darter	1	1	0.13	lotic	invertivore	intolerant
Percina caprodes	Logperch	0	1	0.02	lotic	invertivore	moderate
Percina maculata	Blackside Darter	2	4	0.25	lotic	invertivore	tolerant
Percina sciera	River Darter	2	6	0.76	lotic	invertivore	moderate
Aplodinotus grunniens	Freshwater Drum	3	3	0.62	generalist	piscivore	tolerant

## 3. Results

Overall, 4483 fish, representing 58 species, were collected from the 26 study sites (Table 1). Except for White Bass *Morone chrysops* and Smallmouth Buffalo *Ictiobus bubalus*, the assemblage was composed mostly of non-migratory fishes (Table 1). The average number of fish collected per site was 172 individuals (SD = 126), representing 13 species. One upstream site was removed from further analysis because the laboratory identification data were missing, leading to an extremely low sample size (N = 2 fish). Roughly 40% of the species collected were uncommon and detected in a few locations. Relative to habitat preferences, 43% of the species were lotic, 14% lentic, and 43% generalists. Concerning tolerance, 57% of species were tolerant, 26% were moderately tolerant, and 17% were intolerant. Most of the species were either invertivores (69%) or piscivores (17%).

Contrary to expectations influenced by our literature review, there were no significant differences between the composition of fish assemblages upstream and downstream of reservoirs (Pseudo F = 1.5, p = 0.16). Although the fish assemblage did change with catchment size (Pseudo F = 3.4, p < 0.01), there were no significant differences attributable to subbasin (Pseudo F = 1.01, p = 0.45), nor was the interaction between subbasin and sample location relative to upstream or downstream from the reservoir statistically significant (Pseudo F = 1.1, p = 0.36). The lack of segregation between upstream and downstream sites is made apparent by the large overlap of fish composition among sites illustrated with the NMDS plot (Figure 2). Although the upstream sites seemed more dissimilar as a set (i.e., greater dispersion in Figure 2), their distribution overlapped entirely with the distribution of the downstream sites. Eight species found in the downstream sites were not captured from the upstream sites. Three of these 27 species were present at more than two sites (Table 1). The majority of species present at many sites were common species found both upstream and downstream of the reservoirs.



**Figure 2.** Non-metric multidimensional scaling plot of catch per effort data in streams of the eastern Yazoo Basin in North Mississippi, USA. Sites upstream and downstream of dams are coded by color. Grey symbols represent upstream sites, while black symbols represent downstream sites.

No differences were detected between upstream and downstream sites for physicochemical tolerances (Pseudo F = 0.96, p = 0.37) or habitat preferences (Pseudo F = 0.66, p = 0.48), and similar to the species composition, sites above the reservoirs tend to have more variability in ordination space (Figure 3a,c). However, a marginal difference was apparent when fish were grouped according to trophic guild (Pseudo F = 2.8, p = 0.04). The trophic guild ordination (Figure 3b) displays some separation of upstream from downstream sites mostly due to a higher representation of planktivores, herbivores, and detritivores in upstream sites while downstream sites were dominated by insectivores. In general, both upstream and downstream sites were dominated by moderately tolerant to tolerant lotic specialists and by generalists, although a few upstream sites had a greater percentage of planktivores, herbivores, and detritivores.



**Figure 3.** Non-metric multidimensional scaling plots showing site similarity based on species tolerances (**a**), species trophic levels (**b**), and species habitat preferences (**c**) for fish in streams of the eastern Yazoo Basin in North Mississippi, USA.

The species richness analysis concurred with the species composition analysis. Overall, species richness was not markedly different in sites upstream or downstream from the reservoirs (Pseudo F = 0.62, p = 0.45). Species richness estimated at 50 individuals ranged from 7 to 14 species for the majority of sites (Figure 4), but in general upstream sites showed

more variability. The confidence intervals for all estimates were quite narrow (typically within two species) indicating relatively good fits for both the rarefaction and extrapolation estimates [48].



**Figure 4.** Sites upstream and downstream of dams are coded by color. Grey curves represent upstream sites, while black curves represent downstream sites. Solid lines represent rarefied estimates of fish species richness per number of individuals in the sample, while dashed lines represent estimates extrapolated using the Chao 1 estimator. Points on the curve represent the sampled values. The *Y*-axis denotes species richness; *X*-axis denotes number of individuals in a sample.

#### 4. Discussion

We did not observe a compelling difference in tributary fish assemblages upstream or downstream of impoundments within our study region. This result represents an anomaly in light of the published literature [14–21]. We offer three explanations to account for this anomaly: (1) the history of land use in the Yazoo Basin, (2) the size of the tributaries investigated, and (3) a fish assemblage consisting mostly of non-migratory species. We consider each of these explanations below and argue that the effects of impoundments on fish assemblages may be region-specific. We suggest that the anomaly we observed can assist in refining expectations about fish assemblages and stream fish conservation in impounded river basins.

The Yazoo Basin, originally a hickory-oak hardwood forest, was cleared and converted to agriculture starting in the mid-1800s. By 1940, over 60% of the forests in the basin had been cleared [53]. Deforestation has resulted in widespread soil loss and erosion, which has degraded the quality of the streams in the region. By 1900, many streams were completely aggraded with sand and silt [26]. In response, local communities dredged and channelized the streams [54]. These instream alterations initiated a cycle of channelization, incision, and aggradation that has left a legacy on area streams, despite federal programs to reduce overland erosion beginning in the 1940s and instream erosion beginning in the 1980s [54]. Based on this history, it is possible that species richness was reduced and the fish assemblage was transformed and homogenized before faunal surveys were implemented and before the reservoirs were constructed, in such a way that any effect of the reservoirs on the fish assemblages are overshadowed by the "ghost of land use past" [55]. This explanation is supported by the dominance of tolerant fish species captured during our study (Table 1). Sixty percent of the species are considered tolerant. Only nine species

captured in this study, all classified as intolerant, are generally considered indicators of healthy streams [56]. The representation of tolerant species in our study falls outside the range reported in national assessments. Barbour et al. [57] reported tolerances for 266 species, of which 10% were tolerant, 62% were moderately tolerant, and 28% were intolerant. Similarly, Meador and Carlisle [42] reported that in a sample of 105 species in streams across the U.S., 24% were tolerant, 60% were moderately tolerant, and 16% were intolerant. Considering these two reports, historical changes to landscapes in the Yazoo Basin could have shifted stream fish assemblages towards tolerant and generalist species resilient to impounding.

Most of the studies reporting changes in fish assemblages above reservoirs have been conducted in small basins supporting small reservoirs impounding intermittent or loworder influents [14,18,20,21]. The 4800–14,500 ha reservoirs included in this study had larger tributaries, many of them perennial, which may provide adequate refuge to fish assemblages during droughts or other physicochemical disturbances. This hypothesis is supported by estimates of species richness that are similar between upstream and downstream areas. As additional reinforcement for this argument, Adams and Warren [58] studied the recolonization rate for Yazoo Basin streams that became desiccated during an extreme drought (occurrence <1 in 50 years). All but two of the streams included in our study have a catchment area that is over an order of magnitude larger than the largest catchment area of their desiccated sites, suggesting that the streams included in this study rarely, if ever, become desiccated. Therefore, the fragmentation caused by dams in the Yazoo Basin may not noticeably degrade fish assemblages, as the larger tributaries above the reservoirs may provide sufficient populations for recolonization after rare desiccation events at headwater sites. As additional support for this idea, a recent study [59] also independently tested the influence of Sardis Reservoir (one of the reservoirs included in this analysis) on fish assemblages in low-order headwater streams, and no association was detected between fish assemblage and proximity to Sardis Reservoir. Our study did include a few low-order tributaries that drained directly into a reservoir; in fact, the three lowest richness estimates from the species accumulation curves in Figure 4 represent sites on streams that drain directly into Arkabutla Reservoir. It is possible that these tributaries show isolation effects from periodic extirpations and may be contributing to the higher variation in dissimilarity between upstream tributaries, but that the signal from these smaller tributaries is being overridden by the larger tributaries we included. This suggests that isolation effects, rather than the widespread effects of impoundment, may be limited by the resistance of streams to periodic disturbances.

Dams acting as barriers to potamodromous migrations are another major mechanism that can cause fish assemblage changes in impounded systems. Aadland et al. [60] documented the near total absence of potamodromous fish upstream of dams that were not equipped for fish passage in the Red River of the North, Canada-United States. These findings are supported by similar studies of migratory fishes in Brazil and Puerto Rico [19,61–63]. In our study region, the species pool included only two migratory species, white bass and smallmouth buffalo [29]. Conceivably, a mostly non-migratory fish assemblage may not be conspicuously affected by the fragmentation created by reservoirs. Alternatively, potamodromous species still common in the Mississippi River basin (e.g., Blue Sucker *Cycleptus elongatus*) may have dwindled in the region over the half century the subbasins have been impounded, removing the portion of the assemblages that would normally distinguish fish assemblages in reaches upstream and downstream from reservoirs.

The only apparent difference between sites upstream and downstream from the reservoirs was a marginally significant higher representation of planktivores, herbivores, and detritivores in upstream sites. All but two of the species that formed these guilds were classified as lentic or generalist and commonly occur in reservoirs in southeastern North America. Their representation was irregular across sites, with different species represented at different sites, and when present, they generally occurred in low numbers. Given their low representation and abundance, these common reservoir species did not

have a major influence on the species composition or habitat preference analyses, yet they were highlighted by the trophic guild analysis. Hoeinghaus et al. [40] similarly found that analysis of taxonomic descriptions highlighted only regional geographic patterns, while functional groups described patterns associated with environmental conditions irrespective of geography. Thus, while taxonomic identities (e.g., species) are generally suitable for representing aquatic assemblages and implementing conservation measures targeting taxa, our study supports using functional classifications to examine patterns over regional scales where the species pool may differ over subbasins [45,64].

### 5. Conclusions

Our rather different result suggests the need for refining expectations about similarities in fish assemblages upstream and downstream from impoundments. Typically, the expectation is that impoundments impact upstream fish assemblages through mechanisms such as changes in habitats and loss of longitudinal connectivity. Our study further suggests that the impacts of these mechanisms may not be universal, as the severity of the effects may be nuanced by the regional species pool, the history of stream conditions in the watershed, and the resistance of the streams to periodic disturbances. Additionally, this study highlights how examining stream fish assemblages from several organizational perspectives can give insight to different mechanisms working in the system.

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#### References

- McAllister, D.E.; Platania, S.P.; Schueler, F.W.; Baldwin, M.E.; Lee, D.S. Ichthyofaunal patterns on a geographic grid. In *The Zoogeography of North American Freshwater Fishes*; Hocutt, C.H., Wiley, E.O., Eds.; John Wiley and Sons: New York, NY, USA, 1986; pp. 17–52, ISBN 0-471-86419-6.
- Jelks, H.L.; Walsh, S.; Burkhead, N.M.; Contreras-Balderas, S.; Diaz-Pardo, E.; Hendrickson, D.A.; Lyons, J.; Mandrak, N.E.; McCormick, F.; Nelson, J.S.; et al. Conservation status of imperiled North American freshwater and diadromous fishes. *Fisheries* 2008, 33, 372–407. [CrossRef]
- 3. Warren, M.L., Jr.; Haag, W.R.; Adams, S.B. Forest linkages to diversity and abundance in lowland stream fish communities. In Proceedings of the Conference on Sustainability of Wetlands and Water Resources: How Well Can Riverine Wetlands Continue to Support Society into the 21st Century? Oxford, MS, USA, 23–25 May 2000; Holland, M.M., Warren, M.L., Jr., Stanturf, J.A., Eds.; U.S. Department of Agriculture, Forest Service, Southern Research Station: Ashville, NC, USA, 2002.
- Bennet, S.J.; Rhoton, F.E. Linking upstream channel instability to downstream degradation: Grenada Lake and the Skuna and Yalobusha river basins, Mississippi. *Ecohydrology* 2009, 2, 235–247. [CrossRef]
- Graf, W.L. Dam nation: A geographic census of American dams and their large-scale hydrologic impacts. *Water Resour. Res.* 1999, 35, 1305–1311. [CrossRef]

- U.S. Army Corps of Engineers. National Inventory of Dams. 2018. Available online: https://nid.usace.army.mil (accessed on 18 August 2021).
- Hannan, H.H. Chemical modifications in reservoir-regulated streams. In *The Ecology of Regulated Streams*; Ward, J.V., Stanford, J.A., Eds.; Springer: Boston, MA, USA, 1979; pp. 75–94, ISBN 978-1-4684-8613-1.
- 8. Whitten, C.B.; Patrick, D.M. *Engineering Geology and Geomorphology of Stream-Bank Erosion*; Report 2, Yazoo River Basin Uplands, Mississippi; Geotechnical Laboratory, US Army Corps of Engineers: Washington, DC, USA, 1981.
- 9. Simon, A. A model of channel response in disturbed alluvial channels. Earth Surf. Process. Landf. 1989, 14, 11–26. [CrossRef]
- 10. Biedenharn, D.S.; Elliott, C.M.; Watson, C.C. *The WES Stream Investigation and Streambank Stabilization Handbook*; US Army Engineer Waterways Experiment Station: Vicksburg, MS, USA, 1997.
- 11. Schumm, S.A.; Harvey, M.D.; Watson, C.C. *Incised Channels: Morphology, Dynamics, and Control;* Water Resources Publications: Highlands Ranch, CO, USA, 1984; ISBN 978-1-887201-64-3.
- Simon, A. Gradation Processes and Channel Evolution in Modified West Tennessee Streams: Process, Response and Form; U.S. Geological Survey Professional Paper 1470: Washington, DC, USA, 1994.
- Junk, W.; Bayley, P.B.; Sparks, R.E. The flood pulse concept in river-floodplain systems. In Proceedings of the International Large River Symposium (LARS), Honey Harbour, ON, Canada, September 1989; Dodge, D.P., Ed.; Canadian Special Publication of Fisheries and Aquatic Sciences 106: Ottawa, ON, Canada, 1989; pp. 110–127.
- 14. Winston, M.R.; Taylor, C.M.; Pigg, J. Upstream extirpation of four minnow species due to damming of a prairie stream. *Trans. Am. Fish. Soc.* **1991**, *120*, 98–105. [CrossRef]
- 15. Herbert, M.E.; Gelwick, F.P. Spatial variation of headwater fish assemblages explained by hydrologic variability and upstream effects of impoundment. *Copeia* **2003**, 2003, 273–284. [CrossRef]
- Falke, J.A.; Gido, K.B. Effects of reservoir connectivity on stream fish assemblages in the Great Plains. *Can. J. Fish. Aquat. Sci.* 2006, 63, 480–493. [CrossRef]
- 17. Guenther, C.B.; Spacie, A. Changes in fish assemblage structure upstream of impoundments within the Upper Wabash River Basin, Indiana. *Trans. Am. Fish. Soc.* **2006**, *135*, 570–583. [CrossRef]
- 18. Kashiwagi, M.T.; Miranda, L.E. Influence of small impoundments on habitat and fish communities in headwater streams. *Southeast. Nat.* **2009**, *8*, 23–36. [CrossRef]
- 19. Sá-Oliveira, J.C.; Hawes, J.E.; Isaac-Nahuum, V.J.; Peres, C.A. Upstream and downstream responses of fish assemblages to an Eastern Amazonian hydroelectric dam. *Freshw. Biol.* **2015**, *60*, 2037–2050. [CrossRef]
- Hedden, S.C.; Renner, E.A.; Gido, K.B.; Hase, K.J. Impacts of small impoundments on an intermittent headwater stream community. *Southwest. Nat.* 2018, 63, 34–41. [CrossRef]
- Reuter, C.N.; Kaller, M.D.; Walsh, C.E.; Kelso, W.E. Fish assemblage response to altered dendritic connectivity in the Red River Basin, Central Louisiana. Am. Midl. Nat. 2019, 181, 63–80. [CrossRef]
- 22. Barnett, Z.C.; Adams, S.B. Review of dam effects on native and invasive crayfishes illustrates complex choices for conservation planning. *Front. Ecol. Evol.* **2021**, *8*, 621723. [CrossRef]
- U.S. Geological Survey. National Hydrography Dataset (ver. USGS National Hydrography Dataset Best Resolution (NHD) for Hydrologic Unit (HU) 4–0803 (published 202110102)). 2021. Available online: https://www.usgs.gov/core-science-systems/ ngp/national-hydrography/access-national-hydrography-products. (accessed on 5 January 2021).
- 24. Hilgard, E.W. Report on the Geology and Agriculture of the State of Mississippi; E. Barksdale State Printer: Jackson, MS, USA, 1860.
- Dabney, S.M.; Shields, F.D.; Binger, R.L.; Kuhnle, R.A.; Rigby, J.R. Watershed management for erosion and sedimentation control case study Goodwin Creek, Panola County. In *Advances in Soil Science, Soil Water and Agronomic Productivity*; Lal, R., Stewart, B.A., Eds.; Taylor and Francis: Boca Raton, FL, USA, 2012; pp. 539–568, ISBN 978-1439850794.
- 26. Williston, H.L. *The Yazoo-Little Tallahatchie Flood Prevention Project: A History of the Forest Service's Role;* Forgotten Books: London, UK, 1998; ISBN 978-0265065969.
- 27. Bledsoe, B.P.; Watson, C.C.; Biedenharn, D.S. Quantification of incised channel evolution and equilibrium. *J. Am. Water Resour. Assoc.* **2002**, *38*, 861–870. [CrossRef]
- Knight, S.S.; Cooper, C.M. Fishes of Otoucalofa Creek, Mississippi prior to major channel modifications. J. Miss. Acad. Sci. 1987, 32, 31–38.
- 29. Ross, S.T. Inland Fishes of Mississippi; University Press of Mississippi: Jackson, MS, USA, 2001; ISBN 1-57806-246-2.
- 30. Faucheux, N.M. Assessing the Legacy of Erosion and Flood Control Management Efforts on the Fish Assemblages and Physical Conditions of Yazoo Basin Bluff Hill Streams. Doctoral Dissertation, Mississippi State University, Mississippi State, MS, USA, 2022.
- 31. U.S. Geological Survey. The StreamStats Program v.4.3.11. 2020. Available online: http://streamstats.usgs.gov (accessed on 6 February 2020).
- Rabeni, C.F.; Lyons, J.; Mercado-Silva, N.; Patterson, J.T. Warmwater fish in wadeable streams. In *Standard Methods for Sampling North American Freshwater Fishes*; Bonar, S.A., Hubert, W.A., Willis, D.W., Eds.; American Fisheries Society: Bethesda, MD, USA, 2009; pp. 43–58, ISBN 978-1-934874-10-3.
- 33. Price, A.L.; Peterson, J.T. Estimation and modeling of electrofishing capture efficiency for fishes in wadeable warmwater streams. *N. Am. J. Fish. Manag.* **2010**, *30*, 481–498. [CrossRef]
- Mahon, R. Accuracy of catch-effort methods for estimating fish density and biomass in streams. *Environ. Biol. Fishes* 1980, 5, 343–360. [CrossRef]

- 35. Wagner, M.D.; Schumann, D.A.; Smith, B.J. Gear effectiveness and size selectivity for five cryptic madtom species (*Noturus* spp.). *J. Appl. Ichthyol.* **2019**, *35*, 673–682. [CrossRef]
- 36. Poos, M.S.; Mandrak, N.E.; McLaughlin, R.L. The effectiveness of two common sampling methods for assessing imperiled freshwater fishes. *J. Fish Biol.* **2007**, *70*, 691–708. [CrossRef]
- Reid, S.M.; Yunker, G.; Jones, N.E. Evaluation of single-pass backpack electrofishing for stream fish community monitoring. *Fish. Manag. Ecol.* 2009, 16, 1–9. [CrossRef]
- 38. Rodriguez-Iturbe, I.; Rinaldo, A. *Fractal River Networks: Chance and Self-Organization;* Cambridge University Press: New York, NY, USA, 1997; ISBN 0-521-00405-5.
- 39. Altermatt, F. Diversity in riverine metacommunities: A network perspective. Aquat. Ecol. 2013, 47, 365–377. [CrossRef]
- Hoeinghaus, D.J.; Winemiller, K.O.; Birnbaum, J.S. Local and regional determinants of stream fish assemblage structure: Inferences based on taxonomic vs. functional groups. J. Biogeogr. 2007, 34, 324–338. [CrossRef]
- 41. Smith, S.C.F.; Meiners, S.J.; Hastings, R.P.; Thomas, T.; Columbo, R.E. Low-head dam impacts on habitat and the functional composition of fish communities. *River Res. Appl.* **2017**, *33*, 680–689. [CrossRef]
- 42. Meador, M.R.; Carlisle, D.M. Quantifying tolerance indicator values for common stream fish species of the United States. *Ecol. Indic.* **2007**, *7*, 329–338. [CrossRef]
- 43. Frimpong, E.A.; Angermeier, P.L. FishTraits: A database of ecological and life-history traits of freshwater fishes of the United States. *Fisheries* **2009**, *34*, 487–495. [CrossRef]
- Goldstein, R.M.; Simon, T.P. Toward a united definition of guild structure for feeding ecology of North American freshwater fishes. In Assessing the Sustainability and Biological Integrity of Water Resources Using Fish Communities; Simon, T.P., Ed.; CRC Press: Boca Raton, FL, USA, 1999; pp. 123–202, ISBN 978-0849340079.
- 45. Vannote, R.L.; Minshall, G.W.; Cummins, K.W.; Sedell, J.R.; Cushing, C.E. The river continuum concept. *Can. J. Fish. Aquat. Sci.* **1980**, *37*, 130–137. [CrossRef]
- Troia, M.J.; McManamay, R.A. Biogeographic classification of streams using fish community- and trait-environment relationships. Divers. Distrib. 2019, 26, 108–125. [CrossRef]
- 47. Chao, A.; Gotelli, N.J.; Hsieh, T.C.; Sander, E.L.; Ma, K.H.; Colwell, R.K.; Ellison, A.M. Rarefaction and extrapolation with Hill numbers: A framework for sampling and estimation in species diversity studies. *Ecol. Monogr.* **2014**, *84*, 45–67. [CrossRef]
- Colwell, R.K.; Chao, A.; Gotelli, N.J.; Lin, S.Y.; Mao, C.X.; Chazdon, R.L.; Longino, J.T. Models and estimators linking individualbased and sample-based rarefaction, extrapolation and comparison of assemblages. J. Plant Ecol. 2012, 5, 3–21. [CrossRef]
- Hsieh, T.C.; Ma, K.H.; Chao, A. iNEXT: An R package for rarefaction and extrapolation of species diversity (Hill numbers). *Methods Ecol. Evol.* 2016, 7, 1451–1456. [CrossRef]
- 50. R Core Team. *R: A Language and Environment for Statistical Computing;* R Foundation for Statistical Computing: Vienna, Austria, 2021.
- Anderson, M.J. Permutational Multivariate Analysis of Variance (PERMANOVA); Balakrishnan, N., Colton, T., Everitt, B., Piegorsch, W., Ruggeri, F., Teugels, J.L., Eds.; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2017; pp. 1–15. [CrossRef]
- 52. Clark, K.R.; Gorley, R.N. Primer v7: User Manual/Tutorial; Primer-E: Plymouth, UK, 2015.
- 53. U.S. Army Corps of Engineers. A comprehensive summary report on streambank erosion control demonstration projects constructed in eight watersheds of the Yazoo River basin. Appendix F. In *The Final Report to Congress: The Streambank Erosion Control Evaluation and Demonstration Act of 1974 Section 32, Public Law 93-251;* U.S. Army Corps of Engineers: Washington, DC, USA, 1981.
- 54. Leech, J.R.; Biedenharn, D.S. Sediment Management at the Watershed Level. Coastal and Hydraulics Engineering Technical Note ERDC/CHL CHETN-VII-12; US Army Corps of Engineers, Defense Technical Information Center: Vicksburg, MS, USA, 2012.
- Harding, J.S.; Benfield, E.F.; Bolstad, P.V.; Helfman, G.S.; Jones, E.B.D. III. Stream biodiversity: The ghost of land use past. *Proc. Natl. Acad. Sci. USA* 1998, 95, 14843–14847. [CrossRef] [PubMed]
- Shields, F.D.; Knight, S.S.; Cooper, C.M. Use of the index of biotic integrity to assess physical habitat degradation in warmwater streams. *Hydrobiologia* 1995, 312, 191–208. [CrossRef]
- Barbour, M.T.; Gerritsen, J.; Snyder, B.D.; Stribling, J.B. Rapid Bioassessment Protocols for Use in Streams and Wadeable Rivers: Periphyton, Benthic Macroinvertebrates, and Fish, 2nd ed.; EPA 841-B-99-002; U.S. Environmental Protection Agency, Office of Water: Washington, DC, USA, 1999.
- 58. Adams, S.B.; Warren, M.L. Recolonization by warmwater fishes and crayfishes after severe drought in Upper Coastal Plain hill streams. *Trans. Am. Fish. Soc.* 2005, 134, 1173–1192. [CrossRef]
- 59. Hubbell, J.P.; Schaefer, J.F.; Flood, P.; Warren, M.L., Jr.; Sterling, K.A. Fragmentation alters ecological gradients and headwater fish assemblage composition relative to land use in a dendritic river system. *Can. J. Fish. Aquat. Sci.* 2020, 77, 1281–1291. [CrossRef]
- Aadland, L.P.; Koel, T.M.; Franzin, W.G.; Stewart, K.W.; Nelson, P. Changes in fish assemblage structure of the Red River of the North. In *Historical Changes in Large River Fish Assemblages of the Americas*; Symposium 45; Rinne, J.R., Hughes, R.M., Calamussa, B., Eds.; American Fisheries Society: Bethesda, MD, USA, 2005; pp. 293–321, ISBN 978-1888569728.
- 61. Agostinho, A.A.; Pelicice, F.M.; Gomes, L.C. Dams and the fish fauna of the Neotropical region: Impacts and management related to diversity and fisheries. *Braz. J. Biol.* **2008**, *68*, 1119–1132. [CrossRef]
- 62. Cooney, P.B.; Kwak, T.J. Spatial extent and dynamics of dam impacts on tropical island freshwater fish assemblages. *Bioscience* **2013**, *63*, 176–190. [CrossRef]

- 63. Agostinho, A.A.; Gomes, L.C.; Santos, N.L.C.; Ortega, J.C.G.; Pelicice, F.M. Fish assemblages in Neotropical reservoirs: Colonization patterns, impacts and management. *Fish. Res.* **2016**, *173*, 26–36. [CrossRef]
- 64. Salmaso, N.; Naselli-Flores, L.; Padisák, J. Functional classifications and their application in phytoplankton ecology. *Freshw. Biol.* **2015**, *60*, 603–619. [CrossRef]

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