



Article

Formica picea and F. candida (Hymenoptera: Formicidae): Synonyms or Two Species?

Zoya A. Zhigulskaya ¹, Sergei V. Shekhovtsov ^{1,2,*}, Tatiana V. Poluboyarova ² and Daniil I. Berman ^{1,*}

- ¹ Institute of the Biological Problems of the North FEB RAS, Magadan 685000, Russia; aborigen@ibpn.ru
- ² Institute of Cytology and Genetics SB RAS, Novosibirsk 630090, Russia; tanita11@mail.ru
- * Correspondence: shekhovtsov@bionet.nsc.ru (S.V.S.); dberman@mail.ru (D.I.B.)

Abstract: Ants of the *«Formica picea—F. candida»* complex are widespread across Eurasia. However, it is still a matter of debate if it constitutes one or two species. In this study, we collected a sample of specimens from different parts of Eurasia, sequenced the mitochondrial *cox1* and *cytb* genes, as well as three nuclear loci—*wg*, *Top1*, and ITS2—and analyzed the available published data. We found this complex to contain a new, yet undescribed, taxon that has a large distribution in Siberia and East Asia. Thus, the *«Formica picea—F. candida»* complex consists of at least three taxa with distinct distributions.

Keywords: Formica picea; Formica candida; cryptic species; Northern Eurasia; cox1; cytb

1. Introduction

The taxonomic position of many widespread and abundant ant species remains uncertain [1,2]. Among them are the species of the «black bog ant» complex, *Formica picea* Nylander, 1846 and *F. candida* Smith, 1878. Members of this complex are found from West Europe to Kamchatka, as well as from the valleys of northern rivers to Tibet (Figure 1).

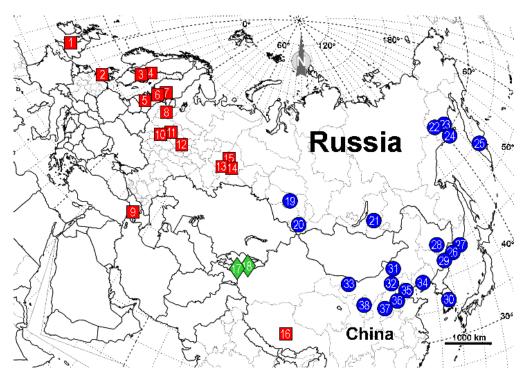


Figure 1. Specimens of the *«F. picea—F. candida»* complex used in this study. Red squares, *F. picea*; green diamonds, *F. candida*; blue circles, *Formica* sp. Specimen numbers refer to Table 1.



Citation: Zhigulskaya, Z.A.; Shekhovtsov, S.V.; Poluboyarova, T.V.; Berman, D.I. *Formica picea* and *F. candida* (Hymenoptera: Formicidae): Synonyms or Two Species? *Diversity* **2022**, *14*, 613. https://doi.org/ 10.3390/d14080613

Academic Editor: Michael Wink

Received: 30 June 2022 Accepted: 26 July 2022 Published: 29 July 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

Diversity **2022**, 14, 613 2 of 9

Table 1. Specimens used in this study.

Location	Species	cox1	cytb
	Formica picea		
1	United Kingdom: Cors Goch National Nature Reserve, S. Wales	-	AY786145 (as F. candida) [3]
2	Denmark: N55.83 E12.56	LT977411 [4]	-
3	Sweden: Trajmossen, Brattforsheden	-	AY786144 (as F. candida) [3]
4	Sweden: Varmland	-	JX170886 [5]
5	Estonia: Torma	-	AY786147 (as F. candida) [3]
6	Finland: Nylandia, Raasepori, Harpar Stortraesket	MZ607401 * [6]	-
7	Finland: Helsinki, Vihti	-	AY786146 * (as F. candida) [3]
8	Russia: Novgorod oblast	-	AY786148 (as F. candida) [3]
9	Armenia: Aragaz mtn, N40.43203 E44.23642	ON220876	ON228281
10	Russia: Moscow oblast, Odintsovo district, raised bog, moss tussock, N55.733 E36.850	ON220886-87	ON228278-79
11	Russia: Moscow oblast, collected by G. Dlussky in 2000	-	AY786149 (as F. candida) [3]
12	Russia: Nizhniy, Novgorod oblast, Kerzhenskiy Natural Reserve, Vishenskoye swamp, N56.467 E44.500	ON220875	ON228280
13	Russia: Sverdlovsk oblast, Revda	-	AY786151 (as F. candida) [3]
14	Russia: Sverdlovsk oblast, Yekaterinburg	-	AY786150 (as <i>F. candida</i>) [3]
15	Russia: Perm oblast, 30 km SSW of Kachkanar	-	AY786152 (as F. candida) [3]
16	China: Quinghai, Tibet	-	AY786157 (as F. candida) [3]
	Formica candida		
17	Kyrgyzstan: Alai Valley	-	AY786154 **, AY786153 [3], JX170887 [5]
18	Kyrgyzstan: Tian-Shan	-	AY786155-AY786156 [3]
	Formica sp.		
19	Russia: Novosibirsk oblast, Iskitim district, Listvyanskiy town, N54.450 E83.483	ON220885	-
20	Russia: Altai Republic, Ongudai town, N50.733 E86.133	ON220880	ON228273
21	Russia: Buryatia Republic, Kizhinga town, N51.84639 E109.91278	ON220883-84	ON228270, ON228274
22	Russia: Magadan oblast, 15 km from Yagodnoye town, bank of Sokhatina river, N62.517 E149.617	ON220878	ON228277
23	Russia: Magadan oblast, 16 km from Yagodnoye town, bank of Detrin river, N62.750 E150.467	ON220879	ON228271
24	Russia: Magadan oblast, Arman town, high floodplain, N59.667 E150.117	ON220872	ON228275
25	Russia, Kamchatka oblast, N53.19 E158.48	LT977377 (as F. candida) [4]	-
26	Russia: Primorye krai, Lazo town, Lazovka river, N43.38056 E133.89778	ON220881	ON228276
27	Russia: Primorye krai, Sinii Utes town, N43.06222 E131.35472	ON220882	ON228272
28	China: Harbin, HelongJiang	KX665031-32 (as <i>F. candida</i>) [7]	-
			·

Diversity 2022, 14, 613 3 of 9

Table 1. Cont.

Location	Species	cox1	cytb
29	China: Yanbian, Jilin	KX665026-27 (as F. candida) [7]	-
30	South Korea	MT800217 (as <i>F. candida</i>) [8]	-
31	Xilin Gol League, Inner Mongolia	KX665022, KX665024 (as F. candida) [7]	-
32	Ulanqab, Inner Mongolia	KX665023, KX665025 (as F. candida) [7]	-
33	Helan Mountains, Inner Mongolia	KX665019-21 (as F. candida) [7]	-
34	Qinglong, Hebei	KX665028 (as F. candida) [7]	-
35	China: Hebei	HQ619704, HQ619710 (as F. candida) [9]	HQ651081, HQ651074 (as F. candida) [9]
36	Xiaowutai Mountain, Hebei	KX665029-30 (as F. candida) [7]	-
37	Zhong-Tiao Mountains, Shanxi	KX665033 (as F. candida) [7]	-
38	Liupan Mountains, Ningxia	KX665034 (as F. candida) [7]	
	Outgroups		
	F. lemani Belarus: Vitebsk oblast, Braslav district, bank of Uklya lake, N55.60001 E27.333918	ON220877	ON228284
	F. lemani Magadan, Arman town high river terrace, N59.867 E150.567	ON220872	ON228283
	F. lemani China	-	HQ651082, HQ651086 [9]
	F. lemani Magadan	-	JX170882 [10]
	F. gagatoides China: Ningxia	-	HQ651073 [9]
	F. gagatoides China	KX665069, KX665071, KX665072 [7], HQ619703	-
	F. gagatoides Canada	JN291934	-
	F. gagatoides Norway	LT977396, LT977398 [4]	-
	F. gagatoides Russia: Magadan oblast, Arman town, N59.667 E150.117	ON220874	ON228282

Locations refer to Figure 1. Sequences obtained in this study shown in bold; *, terra typica; **, type specimen. If an accession was deposited under another species name, this is acknowledged in parentheses; references to published articles are given whenever possible.

Morphological variations within the complex were studied by Dlussky [11] based on extensive collections. Dlussky [11] concluded that there is no ground to split it into several species. Later on, Bolton [12] suggested that the name *F. candida* has the priority over *F. picea*; this was picked up by some—but not all—of the scientists. Seifert [13] suggested that *F. picea* and *F. candida* are two separate species, based on the features of pubescence and some morphometric parameters. According to Seifert [13], the distribution of *F. picea* includes Europe, the Caucasus, and the West Siberian Lowlands, and that of *F. candida* is in Central Asia, the Altai Mountains, and East Siberia. The habitats of *F. picea* include the bogs and European mountains above the forest belt, while *F. candida* lives in diverse habitats, including river floodplains and dry steppes of Asia.

However, Zakharov et al. [14] refuted Seifert's [13] conclusion, stating that the variation of pubescence within the European populations of *F. picea* is higher than the proposed

Diversity 2022, 14, 613 4 of 9

differences between the species. They also stated that there is a smooth gradient of morphological variation from Europe to the Far East. Based on this, the authors returned to treating *F. picea* and *F. candida* as synonyms.

Molecular genetic analysis is increasingly used to resolve problematic questions in ant taxonomy, due to its high resolution and its potential to detect cryptic taxa. [15] There were several molecular genetic attempts to resolve the relationships in the *«F. picea—F. candida»*. Goropashnaya [3] performed a phylogenetic analysis based on the mitochondrial *cytb* gene. She demonstrated that the complex is split into two clades: one restricted to Europe and West Siberia and the other, to Central Asia. Later Goropashnaya et al. referred to these clades as a pair of species [5]. Antonov and Bukin [16] studied a sample of *cytb* sequences of specimens from several regions of the Palearctic identified as *F. candida* and *F. picea*. They concluded that genetic distances within *F. candida* are higher than those between the two species. Important datasets were also obtained by Chen et al. [7] and Schär et al. [4], based on the mitochondrial *cox1* gene.

Therefore, the status of the *«F. picea—F. candida»* species complex remains uncertain. The problem with molecular data is that very similar sequences can be deposited under different species names depending on the authors' viewpoint on this matter. Moreover, some authors sequenced the *cytb* gene, and the others sequenced *cox1*, so the resulting samples cannot be compared directly.

We collected a sample of the *«F. picea—F. candida»* complex specimens from the Northern Palearctic. Moreover, we studied the diversity, geographic and habitat distribution, and ecology of its population from the north of the Russian Far East [17]. The most important specimens were identified by Dlussky and Radchenko, who are specialists in the genus *Formica*; both consider this complex a single species (Radchenko: [18], personal communication). Here, we attempt to resolve the issue on the *«F. picea—F. candida»* complex using molecular analysis, based on the mitochondrial *cox1* and *cytb* genes, as well as three nuclear loci, with the integration of the available GenBank sequences obtained by other authors.

2. Materials and Methods

We collected a set of *Formica* specimens from Eurasia (Figure 1, Table 1). DNA was isolated from single ethanol-fixed worker ants using the commercial silica columns (BioSilica, Novosibirsk, Russia), as described in [4]. A fragment of the mitochondrial cytochrome c oxidase subunit 1 (*cox1*) gene was amplified using the universal primers LCO1490m (5'-TACTC-AACAA-ATCAC-AAAGA-TATTG-G-3'; modified from [10]) and HCO2198 (5'-TAAAC-TTCAG-GGTGA-CCAAA-AAATC-A-3'; [19]). A part of the cytochrome b (*cytb*) was amplified using the primers Fcbl-F (5'-ACCCT-CACCT-GTAAA-TATTT-CTT-3') and Fcbl-R (GGAAT-AGATC-GTAAA-ATTGC-AT-3') designed in this study. PCR was performed using the Biomaster HS-Taq PCR Mix (Biolabmix, Novosibirsk, Russia).

Three nuclear loci were amplified using the primers designed in this study: a fragment of the *wingless* (*wg*) gene, with FWg-F (5'-CGTGG-TCGGC-GATAA-TCTA-3') and FWg-R (5'-CACCA-CCACC-TCCTG-AGTCT-3'); topoisomerase 1 (Top1), with FTPO-F (5'-GAACC-ATTGC-CACCC-ATAGT-3') and FTPO-R (5'-AGCGC-CAGCT-TGTCA- ATAAA-3'); the ribosomal internal transcribed spacer (ITS2), with FITS2-Fw (5'-TCATT-AACGT-TCCGG-AGGTC-3') and FITS2-Rv (5'-TAAAA-TCGTT-GGCCC-TTACG-3').

The obtained DNA fragments were visualized by agarose gel electrophoresis; unincorporated primers and nucleotide phosphates were removed using the shrimp alkaline phosphatase / *E. coli* exonuclease I mix (New England Biolabs, Ipswich, MA, USA). Sanger sequencing was performed on a 3130xl DNA Analyzer (Applied Biosystems, Framingham, MA, USA) in the SB RAS Genomics Core Facility (ICBFM SB RAS, Novosibirsk, Russia) using both forward and reverse primers. The obtained sequences were deposited in GenBank under accession numbers ON228270-ON228284, ON220872-ON220887, and ON862896-ON862909. Sequences obtained by other authors were also used in this study (see Table 1 for GenBank accessions). This dataset includes the *cytb* sequence for the *F. candida* neotype (AY786154).

Diversity 2022, 14, 613 5 of 9

Phylogenetic trees were constructed using the Maximum Likelihood (ML) algorithm and Bayesian Inference, as described in [20]. ML trees were built in RAxML v. 8.2.12 (A. Stamatikis, Hedelberg, Germany) [21], using the GTR + I + G model, as suggested by MrModeltest v.2.0 (J. Nylander, Uppsala, Sweden) [22]. Bayesian analysis was made in MrBayes v. 3.4 (F. Ronquist et al., Stockholm, Sweden) [23] as two simultaneous independent analyses from different random starting trees. There were 20 million generations performed; 25% of the generations were discarded as burn-in.

3. Results

We sequenced fragments of the mitochondrial *cox1* and *cytb* loci for our sample (Table 1) and constructed phylogenetic trees. The final alignment for *cox1* contained 593 bp; for *cytb*, 693 bp. On the *cytb* tree (Figure 2) the *«F. picea—F. candida»* complex was split into two big clades. Nucleotide distances between them were as large as those among other Formica species. One of the clades included the specimens from Kyrgyzstan, Europe, the Urals, the Caucasus, and Tibet. The sequences from Kyrgyzstan that included the *F. candida* neotype fell into a well-supported subclade. The rest of the specimens (shown on Figure 2 as *F. picea*) formed a group with no statistical support. This group included *F. picea* (including a specimen from southern Finland, the *terra typica* of *F. picea*), as well as those submitted to GenBank as *F. candida*. The second clade (shown on Figure 2 as *Formica* sp.) contained the specimens from West and East Siberia, the Russian Far East, China, and Korea. For *cytb*, the average p-distances between *F. candida* and *F. picea* were 1.34%; between *F. picea* and *Formica* sp., 3.25%; between *F. candida* and *Formica* sp., 3.35%. The mean p-distance within *F. picea* was 0.81%; within *F. candida*, 0.06%; within *Formica* sp., 1.05%.

The tree constructed using the *cox1* gene (Figure 3) was built on more specimens than the *cytb* tree. However, there were no data for Kyrgyzstan, where the neotype of *F. candida* was described, or other regions of Central Asia, and there were few *F. picea* specimens. On the *cox1* tree, the *«F. picea—F. candida»* complex was also found to contain two clades: one corresponding to *F. picea* (including *terra typica*, i.e., the region where the original type specimens were collected) and the other to *Formica* sp. The average p-distances between *F. picea* and *Formica* sp. were 2.52%; within the species, they were 0.57 and 0.38%, respectively. We should also note that the *cox1* tree contained as many as three clades, with specimens identified as *F. gagatoides*.

The final alignment for the *wg* locus included 312 bp; for *Tpo1*, 717 bp; for ITS2, 678 bp. Sequences of wg and TPO of *F. picea*, *Formica* sp., and *F. gagatoides* were identical, except for some degenerate positions. In the ITS sequence, *F. picea* and *Formica* sp. could be distinguished by one A<>G substitution in position 216 of the alignment, while these two taxa differed from *F. gagatoides* by two substitutions and one short indel.

Diversity 2022, 14, 613 6 of 9

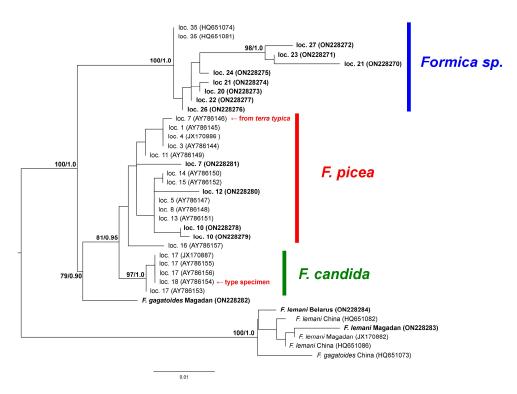


Figure 2. Phylogenetic tree constructed using *cytb* sequences. Numbers near branches indicate ML bootstrap support/Bayesian posterior probabilities. GenBank accessions given in parentheses; sequences obtained in this study are shown in bold.

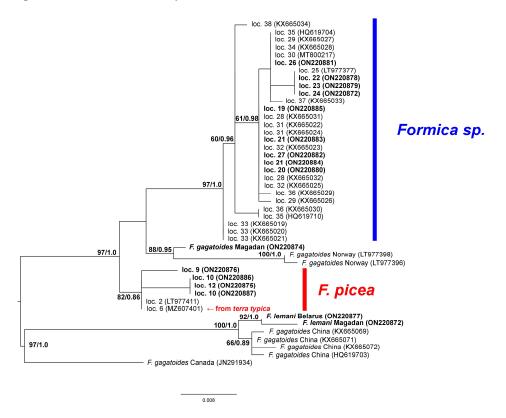


Figure 3. Phylogenetic tree constructed using *cox1* sequences. Numbers near branches indicate ML bootstrap support/Bayesian posterior probabilities. GenBank accessions given in parentheses; sequences obtained in this study are shown in bold.

Diversity 2022, 14, 613 7 of 9

4. Discussion

In this study, we analyzed the dataset of the *«F. picea—F. candida»* complex in order to find out if F. picea and F. candida are synonyms or two distinct species. The results were unexpected. The sample, indeed, contained two clades: one with F. candida and F. picea specimens and the other representing a previously unknown taxon (Figure 2). For mitochondrial sequences, genetic distances between these clades were as high as between the well-established species of the genus Formica. This is in line with earlier findings that recovered multiple cryptic species within the genus Formica [24,25], as well as other wellstudied genera, such as Pheidole [26,27], Myrmica [28,29], Lasius and Cardiocondyla [24], etc. For the nuclear sequences, however, the differences were not as high: wg and Tpo1 gene fragments were identical, and only one substitution distinguished F. picea from Formica sp. On the other hand, F. gagatoides was also very close to these taxa, differing from F. picea by two substitutions and one indel in ITS2, with two other genes being identical. It is well known that nuclear DNA has a lower substitution rate compared to the mitochondrial genome, and closely related ant species may have few distinctive positions or none at all. From the data obtained in [4], one can see that there are very few differences in the nuclear sequences of the whole F. picea/F. gagatoides/F. lemani/F. fusca/F. neorufarbis group. This is the usual situation that is found in ants, e.g., in the well-studied Formica rufa group, several species acknowledged by myrmecologists cannot be distinguished by nuclear markers and even show significant interspecific hybridization [30,31]. A similar case can also be observed in the genus *Proformica* [32]. Thus, we can conclude that the Asian *Formica* sp. is apparently distinct genetically from F. picea and F. candida, but it is unclear whether it should be regarded as a separate species or as a subspecies.

F. candida was initially described by Smith [33] based on a single specimen. This specimen was collected "On the road across the Pamir, from Sarikol to Panja", which is probably somewhere near the easternmost part of China, on the border with Tajikistan. According to [13], this specimen was lost, so he fixed a neotype from Kyrgyzstan. This location is quite remote (about 315 km straight line distance in a mountainous area) from the original one. Our sample contained a *cytb* sequence of the neotype (AY786154). We can state that, so far, the *F. candida* clade is limited to the available specimens from Kyrgyzstan and was not found elsewhere. The type specimen of *F. picea* is old and highly damaged [13]. It was collected in the vicinity of Helsingfors (currently Helsinki, Finland) with no precise information on its whereabouts. Both locations available in our sample are several dozen km from Helsinki. Given the fact that no other group of the complex was found in Finland, we can suggest that these accessions represent the «real» *F. picea*.

Whether *F. picea* and *F. candida* are reciprocally monophyletic remains an open question. On our *cytb* tree *F. picea* was not supported as a clade (Figure 2), but more data are needed to verify that.

Based on the data on the type specimen origin, we cannot assign the *Formica* sp. clade to either *F. picea* or *F. candida* and have to conclude that it represents a new taxon that is still to be described. From the maps, we can see that all three ant taxa have distinct distributions: *Formica* sp. is found in Asian Russia, Siberia, and East Asia; *F. candida*, in Central Asia; *F. picea*, in Europe, with a single finding in Tibet. As suggested by multiple authors, morphological differences between these taxa are elusive [11,13,18]. There might be some differences in ecological preferences of these species: the European *F. picea* is found in swamps or in the alpine belt of mountains. The Asian *Formica* sp. is considered to be eurytopic in Siberia, while in the Russian Far East, it is found mostly in floodplains ([11] and our observations). However, it remains to be proved that these ecological differences coincide with the distribution boundaries of these species.

5. Conclusions

By trying to resolve the relationships within the *«F. candida—F. picea»* complex, in order to separate these two species, we found a new third member of this complex. It is important to note that genetic distances between *F. candida* and *F. picea* are significantly

Diversity 2022, 14, 613 8 of 9

smaller than between them and the newly found taxon. The latter seems to not be the only cryptic taxon in this group: we obtained preliminary evidence that *F. gagatoides* might also represent as many as three cryptic taxa.

Author Contributions: Conceptualization, Z.A.Z. and D.I.B.; methodology, S.V.S. and T.V.P.; investigation, S.V.S. and T.V.P.; data curation, S.V.S.; writing—original draft preparation, D.I.B.; writing—review and editing, Z.A.Z. and S.V.S.; visualization, D.I.B. and S.V.S.; supervision, D.I.B.; project administration, D.I.B.; funding acquisition, D.I.B. All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported by the State Budget Projects no. 1021060307698-5 (Faunogenesis and adaptive strategies of poikilothermic animals in extreme environments of the North) and no. FWNR-2022-0022.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The obtained sequences were deposited in GenBank under accession numbers ON228270-ON228284 (*cytb*) and ON220872-ON220887 (*cox1*).

Acknowledgments: We are grateful to N.A. Bulakhova, E.B. Fedoseeva, N.G. Gagarinova, T.V. Popkova, Yu. N. Sundukov, S.V. Chesnokova, Z.M. Yusupov and J. Sorvari for their help with specimen collection. Sanger sequencing was performed in SB RAS Genomics Core Facility (ICBFM SB RAS, Novosibirsk, Russia).

Conflicts of Interest: The authors declare no conflict of interest.

References

- Borowiec, M.L.; Moreau, C.S.; Rabeling, C. Ants: Phylogeny and Classification. In Encyclopedia of Social Insects; Springer International Publishing: Cham, Switzerland, 2020; pp. 1–18.
- Valenzuela-González, J.E.; Pérez-Toledo, G.R.; García-Martínez, M.A. Adelomyrmex dorae sp. nov. Garcia-Martinez (Hymenoptera: Formicidae): A New Species Supported by Parsimony Analysis of Morphological Characters. Trans. Am. Entomol. Soc. 2017, 143, 713–727. [CrossRef]
- 3. Goropashnaya, A. Phylogeographic Structure and Genetic Variation in *Formica* Ants. Ph.D. Thesis, Uppsala University, Uppsala, Sweden, 20 December 2003.
- 4. Schär, S.; Talavera, G.; Espadaler, X.; Rana, J.D.; Andersen, A.; Cover, S.P.; Vila, R. Do Holarctic ant species exist? Trans-Beringian dispersal and homoplasy in the Formicidae. *J. Biogeogr.* **2018**, *45*, 1917–1928. [CrossRef]
- 5. Goropashnaya, A.V.; Fedorov, V.B.; Seifert, B.; Pamilo, P. Phylogenetic relationships of Palaearctic *Formica* species (Hymenoptera, Formicidae) based on mitochondrial cytochrome b sequences. *PLoS ONE* **2012**, *7*, e41697. [CrossRef] [PubMed]
- 6. Roslin, T.; Somervuo, P.; Pentinsaari, M.; Hebert, P.D.N.; Agda, J.; Ahlroth, P.; Anttonen, P.; Aspi, J.; Blagoev, G.; Blanco, S.; et al. A molecular-based identification resource for the arthropods of Finland. *Mol. Ecol. Resour.* **2022**, 22, 803–822. [CrossRef]
- 7. Chen, Y.; Zhou, S. Phylogenetic relationships based on DNA barcoding among 16 species of the ant genus *Formica* (Hymenoptera: Formicidae) from China. *J. Insect Sci.* **2017**, *17*, 117. [CrossRef]
- 8. Park, S.; Noh, P.; Kang, J.-Y. Endosymbionts and phage WO infections in Korean ant species (Hymenoptera: Formicidae). *Proc. Natl. Inst. Ecol. Repub. Korea* **2020**, *1*, 52–57. [CrossRef]
- 9. Chen, Z.; Zhou, S.Y.; Ye, D.; Chen, Y.; Lu, C. Molecular phylogeny of the ant subfamily Formicinae (Hymenoptera, Formicidae) from China Based on mitochondrial genes. *Sociobiology* **2013**, *60*, 135–144. [CrossRef]
- 10. Shekhovtsov, S.V.; Berman, D.I.; Bulakhova, N.A.; Makarova, O.L.; Peltek, S.E. Phylogeography of earthworms from high latitudes of Eurasia. *Acta Zool. Acad. Sci. Hung.* **2018**, *64*, 369–382. [CrossRef]
- 11. Dlussky, G.M. Ants of the Genus Formica (Hymenoptera: Formicidae g. Formica); Nauka: Moscow, Russia, 1967; p. 236.
- 12. Bolton, B. A New General Catalogue of the Ants of the World; Harvard University Press: Cambridge, UK, 1995; p. 502.
- 13. Seifert, B. The "Black Bog Ant" *Formica picea* Nylander, 1846—A species different from *Formica candida* Smith, 1878 (Hymenoptera: Formicidae). *Myrmecolog. Nachr.* **2004**, *6*, 29–38.
- 14. Zakharov, A.A.; Dlussky, G.M.; Goryunov, D.N.; Gilyov, A.V.; Zryanin, V.A.; Fedoseeva, E.B.; Gorokhovskaya, E.A.; Radchenko, A.G. *Monitoring of the Formica Ants*; KMK Scientific Press: Moscow, Russia, 2013; p. 99.
- 15. Ward, P.S.; Brady, S.G.; Fisher, B.L.; Schultz, T.R. The evolution of myrmicine ants: Phylogeny and biogeography of a hyperdiverse ant clade (Hymenoptera: Formicidae). *Syst. Entomol.* **2015**, *40*, 61–81. [CrossRef]
- 16. Antonov, I.A.; Bukin, Y.S. Molecular phylogenetic analysis of the ant genus *Formica* L. (Hymenoptera: Formicidae) from Palearctic region. *Russ. J. Genet.* **2016**, 52, 810–820. [CrossRef]

Diversity 2022, 14, 613 9 of 9

17. Berman, D.I.; Alfimov, A.V.; Zhigulskaya, Z.A.; Leirikh, A.N. Overwintering and Cold-Hardiness of Ants in the Northeast of Asia; Pensoft Publishers: Sofia-Moscow, Russia, 2010; p. 294.

- 18. Radchenko, A.G. Ants (Hymenoptera, Formicidae) of Ukraine; Institute of Zoology of Ukraine NAS: Kyiv, Ukraine, 2016; p. 495.
- 19. Folmer, O.; Hoeh, W.R.; Black, M.B.; Vrijenhoek, R.C. Conserved primers for PCR amplification of mitochondrial DNA from different invertebrate phyla. *Mol. Mar. Biol. Biotechnol.* **1994**, *3*, 294–299.
- 20. Shekhovtsov, S.V.; Shipova, A.A.; Poluboyarova, T.V.; Vasiliev, G.V.; Golovanova, E.V.; Geraskina, A.P.; Bulakhova, N.A.; Szederjesi, T.; Peltek, S.E. Species delimitation of the *Eisenia nordenskioldi* complex (Oligochaeta, Lumbricidae) using transcriptomic data. *Front. Genet.* **2020**, *11*, 1508. [CrossRef]
- 21. Stamatakis, A. RAxML version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **2014**, 30, 1312–1313. [CrossRef]
- 22. Nylander, J.A.A. MrModeltest Ver. 2; Evolutionary Biology Centre, Uppsala University: Uppsala, Sweden, 2004.
- 23. Ronquist, F.; Teslenko, M.; Van Der Mark, P.; Ayres, D.L.; Darling, A.; Höhna, S.; Larget, B.; Liu, L.; Suchard, M.A.; Huelsenbeck, J.P. MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst. Biol.* **2012**, *61*, 539–542. [CrossRef]
- 24. Seifert, B. Cryptic species in ants (Hymenoptera: Formicidae) revisited: We need a change in the alpha-taxonomic approach. *Myrmecol. News* **2009**, *12*, 149–166.
- 25. Bernasconi, C.; Cherix, D.; Seifert, B.; Pamilo, P. Molecular taxonomy of the *Formica rufa* group (red wood ants) (Hymenoptera: Formicidae): A new cryptic species in the Swiss Alps. *Myrmecol. News* **2011**, *14*, 37–47.
- 26. Seifert, B. Inconvenient hyperdiversity—The traditional concept of "*Pheidole pallidula*" includes four cryptic species (Hymenoptera: Formicidae). *SOIL Org.* **2016**, *88*, 1–17.
- 27. Fournier, D.; Tindo, M.; Kenne, M.; Mbenoun Masse, P.S.; Van Bossche, V.; De Coninck, E.; Aron, S. Genetic structure, nestmate recognition and behaviour of two cryptic species of the invasive big-headed ant *Pheidole megacephala*. *PLoS ONE* **2012**, 7, e31480. [CrossRef]
- 28. Seifert, B.; Yazdi, A.B.; Schultz, R. *Myrmica martini* sp. n.—A cryptic species of the *Myrmica scabrinodis* species complex (Hymenoptera: Formicidae) revealed by geometric morphometrics and nest-centroid clustering. *Myrmecol. News* **2014**, *19*, 171–183.
- 29. Ebsen, J.R.; Boomsma, J.J.; Nash, D.R. Phylogeography and cryptic speciation in the *Myrmica scabrinodis* Nylander, 1846 species complex (Hymenoptera: Formicidae), and their conservation implications. *Insect Conserv. Divers.* **2019**, 12, 467–480. [CrossRef]
- 30. Seifert, B. A taxonomic revision of the Palaearctic members of the *Formica rufa* group (Hymenoptera: Formicidae)–The famous mound-building red wood ants. *Myrmecol. News* **2021**, *31*, 133–179. [CrossRef]
- 31. Pamilo, P.; Kulmuni, J. Genetic identification of *Formica rufa* group species and their putative hybrids in northern Europe. *Myrmecol. News* **2022**, 32. [CrossRef]
- 32. Sanllorente, O.; Lorite, P.; Ruano, F.; Palomeque, T.; Tinaut, A. Phylogenetic relationships between the slave-making ants *Rossomyrmex* and their *Proformica* hosts in relation to other genera of the ant tribe Formicini (Hymenoptera: Formicidae). *J. Zool. Syst. Evol. Res.* **2018**, *56*, 48–60. [CrossRef]
- Smith, F. Scientific Results of the Second Yarkand Mission; Based upon the Collections and Notes of the Late Ferdinand Stoliczka, Ph.D.; Superintendent of Government Printing (Government of India): Calcutta, India, 1878; p. 22.