

**Table S1.** Bacterial strains, plasmids and primers used in this study.

Name	Description	Origin
<b>Strain</b>		
<i>Pectobacterium atrosepticum</i> SCRI1043 ( <i>Pba</i> )	Wild type	[1]
<i>Pectobacterium atrosepticum</i> SCRI1043 $\Delta$ <i>expI</i>	Mutant strain of SCRI1043 containing the kanamycin cassette in the chromosome; <i>Km<sup>R</sup></i>	This study
<i>Pectobacterium atrosepticum</i> SCRI1043 $\Delta$ <i>luxS</i>	Mutant strain of SCRI1043 containing the kanamycin cassette in the chromosome; <i>Km<sup>R</sup></i>	This study
<i>Escherichia coli</i> cc118	Host for suicidal vector pKNG101; $\Delta$ ( <i>ara</i> , <i>leu</i> ) <i>araD</i> $\Delta$ <i>lacX</i> 74 <i>galE</i> <i>galK</i> <i>PhoA20</i> <i>thi-1</i> <i>rpsE</i> <i>rpoB</i> <i>argE</i> ( <i>am</i> ) <i>recA1</i> , <i>Sm<sup>R</sup></i>	[2]
<i>Escherichia coli</i> HH26/pNJ5000	Mobilizing strain for conjugative transfer of the suicide vector pKNG101 into <i>Pba</i> cells; <i>tra+</i> ; <i>Tet<sup>R</sup></i>	[3]
<i>Escherichia coli</i> NovaBlue	<i>endA</i> <i>1hsdR17</i> ( <i>rK12-mK12+</i> ) <i>supE44</i> <i>thi-1</i> <i>recA1</i> <i>gyrA96</i> <i>relA1</i> <i>lacF'</i> [ <i>proA+</i> <i>B+</i> <i>lacIqZ</i> $\Delta$ M15:: <i>Tn10</i> ( <i>Tet<sup>R</sup></i> )]	Novagen
<i>Pantoea ananatis</i> LMG 20103	Wild type	[4]
<i>Dickeya solani</i> DSM28711	Wild type	[5]
<i>Xanthomonas vesicatoria</i> DSM 22252	Wild type	[6]
<i>Escherichia coli</i> K-12 MG1655	Wild type	NCBI:txid511145
<i>Pseudomonas syringae</i> DC3000	Wild type	[7]
<i>Bacillus subtilis</i> DSM10	Wild type	[8]
<i>E. coli</i> JLD271	Host for the bioluminescence reporter vector pAL103 or pAL104 as a negative control; <i>WM54 sdiA271::cam</i>	[9]
<i>Vibrio harveyi</i> BB170	Strain was used as a sensor AI-2; <i>luxN::Tn5</i>	[10]
<i>Vibrio harveyi</i> BB152	Strain was used as a positive control; <i>luxI::Tn5</i>	[10]
<b>Plasmids</b>		
pGEM-T Easy	Linearized vector for cloning; <i>f1 ori Amp<sup>R</sup> lacZ</i>	Promega
pGEM: <i>expI</i>	<i>f1 ori Amp<sup>R</sup> lacZ expI</i>	This study
pGEM: <i>luxS</i>	<i>f1 ori Amp<sup>R</sup> lacZ luxS</i>	This study
pGEM: $\Delta$ <i>expI</i> ;Km <sup>R</sup>	<i>f1 ori Amp<sup>R</sup> lacZ Km<sup>R</sup></i>	This study
pGEM: $\Delta$ <i>luxS</i> ;Km <sup>R</sup>	<i>f1 ori Amp<sup>R</sup> lacZ Km<sup>R</sup></i>	This study
pKD4	Matrix for PCR amplification of kanamycin resistance cassette; <i>oriR<math>\gamma</math> rgnB bla Km<sup>R</sup></i>	[11]
pKNG101	Suicide mobilized vector for inactivation of target genes; <i>pir-ori R6K mobRK2 sacB Sm<sup>R</sup></i>	[12]
pKNG101: $\Delta$ <i>expI</i> ;Km <sup>R</sup>	Suicide plasmid carrying mutant locus $\Delta$ <i>expI</i> ;Km <sup>R</sup> ; <i>Km<sup>R</sup> Sm<sup>R</sup> sacB</i>	This study
pKNG101: $\Delta$ <i>luxS</i> ;Km <sup>R</sup>	Suicide plasmid carrying mutant locus $\Delta$ <i>luxS</i> ;Km <sup>R</sup> ; <i>Km<sup>R</sup> Sm<sup>R</sup> sacB</i>	This study
pAL103	The bioluminescence reporter vector; <i>Tet<sup>R</sup></i> ; <i>luxR+luxI::luxCDABE</i> ; <i>Tet<sup>R</sup> p15A origin</i>	[9]
pAL104	The negative control for vector pAL103; <i>Tet<sup>R</sup></i> ; <i>luxI::luxCDABE</i> ; <i>Tet<sup>R</sup> p15A origin</i>	[9]

Primers	
Primer name	Primer sequence 5'–3'
Primers for mutagenesis <i>ΔexpI</i>	
<b>upexpIF</b>	CGGCCGTCAGGAGATGTCTGAG
<b>dnexpIR</b>	ACAATTAGACTTAGGTACAGGTGTGG
<b>dnexpIKmF</b>	GCAGCTCCAGCCTACACGGAATTAGCGTAGTTGAACAAGG
<b>upexpIKmR</b>	CCCATGTCAGCCGTGTCAGCAAGAGTTCACACACG
<b>KmexpIF</b>	CGGCTGACATGGGGTGTAGGCTGGAGCTGCTTC
<b>KmexpIR</b>	GTGTAGGCTGGAGCTGCACGGCTGACATGGGAATTAG
<b>CheckexpIF</b>	CCTGCTGACCAATGCTGAACTG
<b>CheckexpIR</b>	ACATTCCCTGATGTTAGCCAATC
Primers for mutagenesis <i>ΔluxS</i>	
<b>upluxSF</b>	CCTGCCTTATGCGACCTCTCTAC
<b>dnluxSR</b>	CGCCTACACACTCAGCCTCG
<b>dnluxSKmF</b>	CCATGTCAGCCGTTAAGACATATCTAGTTTATTGG
<b>upluxSKmR</b>	CTCCAGCCTACACAATCAGTAACGGCATAATGCCTCC
<b>KmluxSF</b>	CATTATGCCGTTACTGATTGTGTAGGCTGGAGCTGCTTC
<b>KmluxSR</b>	CAATAAACTAGATATGTCTTAACGGCTGACATGGGAATTAGC
<b>CheckluxSF</b>	GCAGAAAGCGGGAAAAAGGC
<b>CheckluxSR</b>	CGCTGCTGACGGAGAACTGTTG

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