



Review

Promising Strategy of mPTP Modulation in Cancer Therapy: An Emerging Progress and Future Insight

Mohammad Waseem ¹ and Bi-Dar Wang ^{1,2,*}

¹ Department of Pharmaceutical Sciences, School of Pharmacy and Health Professions, University of Maryland Eastern Shore, Princess Anne, MD 21853, USA; mwaseem@umes.edu

² Hormone Related Cancers Program, University of Maryland Greenebaum Comprehensive Cancer Center, Baltimore, MD 21201, USA

* Correspondence: bwang@umes.edu; Tel.: +1-410-621-2240; Fax: +1-410-651-8394

Abstract: Cancer has been progressively a major global health concern. With this developing global concern, cancer determent is one of the most significant public health challenges of this era. To date, the scientific community undoubtedly highlights mitochondrial dysfunction as a hallmark of cancer cells. Permeabilization of the mitochondrial membranes has been implicated as the most considerable footprint in apoptosis-mediated cancer cell death. Under the condition of mitochondrial calcium overload, exclusively mediated by oxidative stress, an opening of a nonspecific channel with a well-defined diameter in mitochondrial membrane allows free exchange between the mitochondrial matrix and the extra mitochondrial cytosol of solutes and proteins up to 1.5 kDa. Such a channel/nonspecific pore is recognized as the mitochondrial permeability transition pore (mPTP). mPTP has been established for regulating apoptosis-mediated cancer cell death. It has been evident that mPTP is critically linked with the glycolytic enzyme hexokinase II to defend cellular death and reduce cytochrome c release. However, elevated mitochondrial Ca^{2+} loading, oxidative stress, and mitochondrial depolarization are critical factors leading to mPTP opening/activation. Although the exact mechanism underlying mPTP-mediated cell death remains elusive, mPTP-mediated apoptosis machinery has been considered as an important clamp and plays a critical role in the pathogenesis of several types of cancers. In this review, we focus on structure and regulation of the mPTP complex-mediated apoptosis mechanisms and follow with a comprehensive discussion addressing the development of novel mPTP-targeting drugs/molecules in cancer treatment.

Keywords: mPTP; therapeutic targets; mPTP-mediated apoptosis; MOMP; oxidative stress



Citation: Waseem, M.; Wang, B.-D. Promising Strategy of mPTP Modulation in Cancer Therapy: An Emerging Progress and Future Insight. *Int. J. Mol. Sci.* **2023**, *24*, 5564. <https://doi.org/10.3390/ijms24065564>

Academic Editors: Elena Levantini and Daniela Sanchez Bassères

Received: 7 February 2023

Revised: 4 March 2023

Accepted: 7 March 2023

Published: 14 March 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Mitochondria are considered as intracellular organelles crucial for energy generation and are also involved in several kinds of cellular damage and/or death including necrosis, apoptosis, and other impaired mechanisms via mitochondrial outer membrane permeabilization (MOMP) [1]. Mitochondria have been revealed to mediate the intrinsic apoptotic pathway provoked by the cytotoxic potency of different kinds of chemotherapeutic molecules [2]. These mitochondria-mediated intrinsic machinery could be triggered by different stimuli, such as high contents of cytoplasmic Ca^{2+} , overproduction of reactive oxygen species (ROS), and other environmental factors such as UV damage [3–5]. Such stimuli activate apoptotic cascade, increase apoptotic protein expression, promote release of cytochrome c (Cyt C), and consequently promote/complete the eradication of the cells [6].

Despite the fact the events underlying MOMP remain elusive, several protein complexes on mitochondrial membranes have been functionally linked with MOMP. In addition, the mitochondrial permeability transition (mPT) is believed to be critically involved in the mitochondria-mediated apoptosis [7,8]. mPT has been known as an alteration in the permeability of the inner mitochondrial membrane (IMM). During the past four decades,

it was accepted that calcium overloaded/energized mitochondria could lead to substantial swelling episodes [9,10]. It has been suggested that swelling is due to a nonspecific permeabilization of the IMM via activation of Ca-sensitive phospholipases and thereby causes the production of fatty acids and lysophospholipids in the membrane. Nevertheless, pioneer studies by Haworth and Hunter [11] and later by Crompton and his coworkers [12] corroborated the evidence on increased permeability and showed the opening of a non-specific channel with a well-defined diameter in the mitochondrial membrane that allows free exchange between the mitochondrial matrix and the extra mitochondrial cytosol of solutes and proteins smaller than 1.5 kDa. Later, this channel/pore was recognized as the mitochondrial permeability transition pore (mPTP) (Figure 1).

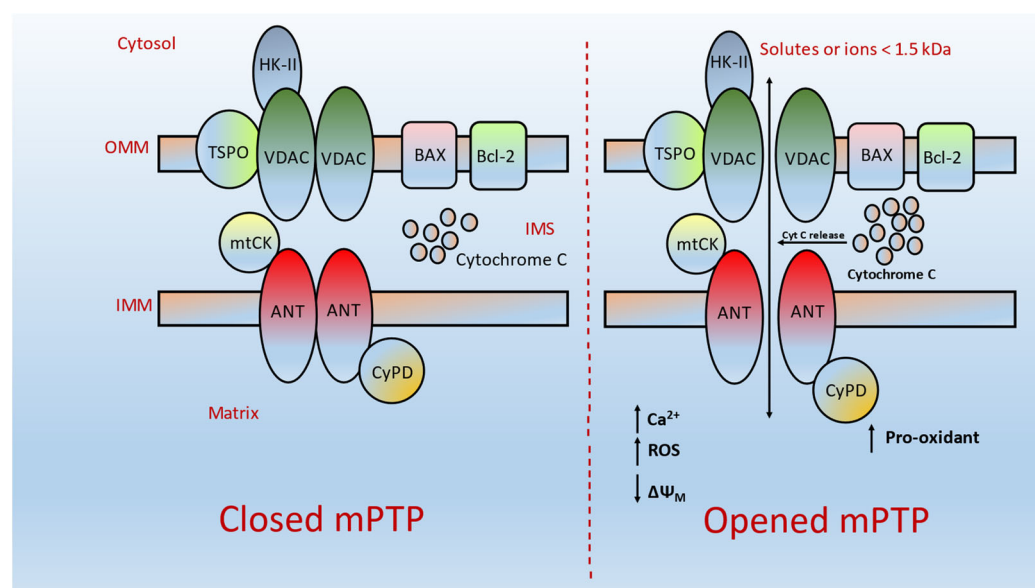


Figure 1. Schematic representation of the junction of mPTP at both opened and closed states. OMM, outer mitochondrial membrane; IMM, inner mitochondrial membrane; VDAC, voltage-dependent anion channel (mitochondrial porin); HK-II, hexokinase II; TSPO, translocator protein; ANT, adenine nucleotide transporter; CypD, cyclophilin D; Bcl-2, antiapoptotic protein; BAX, proapoptotic protein. Numerous stimuli such as pro-oxidant, enhanced intracellular Ca^{2+} levels, and overproduction of ROS are able to allow the free exchange of solutes and proteins smaller than 1.5 kDa between the mitochondrial matrix and the extra mitochondrial cytosol, ultimately causing mPTP opening.

mPTP is a multiprotein channel complex found at the sites of mitochondrial membranes and is considered as a crucial regulator for the maintenance of the mitochondrial respiratory chain [13]. mPTP is predominantly composed of key proteins such as the voltage-dependent anion channel (VDAC) at the mitochondrial membrane (OMM), adenine nucleotide translocator 1 (ANT-1) at the inner mitochondrial membrane (IMM), and cyclophilin-D (Cyp-D) at the mitochondrial matrix platform [8]. Under normal physiological conditions, the mitochondrial inner membrane is impermeable, and only limited metabolites and/or ions can pass across the membranes. In healthy cells, mPTP normally remains closed and is triggered or opened upon external stimuli. It has been suggested that stimuli could induce the upregulation of both ANT-1 and Cyp-D, leading to mPTP opening and ROS production, ATP depletion and Cyt C release from the opened pore [14] (Figure 1). Notably, mPTP-mediated apoptosis machinery has been considered as an important clamp and plays a critical role in the pathogenesis of several kinds of cancers. It has been implicated that increased influxes of mitochondrial Ca^{2+} are generally predisposed to cellular death that could lead to intensifying mPTP-mediated cell invasion and proliferation in multiple cancer episodes. These events occur via a posttranslational scenario and/or the inhibitory action of several kinds of promalignant factors.

2. Cancer: Global Burden

Cancer has been recognized as a disorder arisen from genetic or epigenetic remodeling in somatic cells [15]. It is believed that cancer cells develop a degree of autonomy from alarming signals or stimuli, leading to unregulated growth and proliferation. If proliferation continues and transmits to the surrounding cells, it potentially leads to a more destructive/deadly episode. Indeed, almost 90% of cancer-associated mortality was observed owing to tumor spread, so-called metastasis [16–18].

Cancer is the second leading cause of mortality at the global level. Overall, the prevalence of cancer has exponentially increased; 1.9 million new cancer cases and 609,360 cancer deaths were projected to occur in 2022 in the United States [16]. According to global demographic investigations, about 0.42 billion new cases of cancer have been anticipated by 2025 every year, representing an expanding incidence of cancer in upcoming years. On this planet, new cancer was recorded as about 1,918,030 cases in 2022; overall prevalence in men was documented as about 983,160 and women about 934,870 cases. Globally, about 9.5 million casual cases or deaths were predicted in cancer. By 2022, for males the most prevalent and death cases in cancers are recorded as lung (1,435,943), prostate (1,414,259), and colorectal (1,065,960) cancer. For females, the most prevalent and death cases are recorded as breast (2,261,419), colorectal (865,630), and lung cancer, contributing 44.5% of the total number of new cases diagnosed in 2022.

3. Possible Functional Implications of mPTP in Cancer

The mPTP junction has emerged as a promising target for novel chemotherapeutics [16,19]. In this review, we focus on structure and regulation of the mPTP complex in apoptosis-mediated cancer cell death, followed by the development of novel drugs/compounds targeting mPTP for cancer therapy. The closure of mPTP establishes no chance of cellular collapse, which then constantly pushes to execute cellular blebbing or shrinkage due to the stimulation of caspases via the apoptosis mechanism. The greater capacity to enhance cellular death triggers such conditions and is an anchor for targeting malignant cell progression [17], and most chemotherapeutics function by directly or indirectly activating the apoptotic cascade in neoplastic cells. Accordingly, a deeper knowledge of mPTP structure and its regulation in cancer provides an appealing way to develop anticancer strategies. mPTP opening is triggered by both the Ca^{2+} overload in the mitochondrial matrix and the overproduction of ROS-mediated oxidative stress. It has been implicated that the opening of mPTP results in the increase of mitochondrial permeability that facilitates free fluxes of solutes (including water, molecules, and ions) into the mitochondrial matrix. In later stages, the opening of mPTP induces mitochondrial swelling, OMM rupture, and impairment in ETC, causing a massive release of ROS. Furthermore, the enhanced mitochondrial permeability also leads to loss of mitochondrial membrane potential ($\Delta\Psi_M$), thereby causing a decrease of the cellular mitochondrial ATP and an increase of the intracellular Ca^{2+} levels. Consequently, the opening of mPTP induces the activation of endogenous apoptotic machinery, promoting chemotherapeutic-induced cancer cell death (Figure 2).

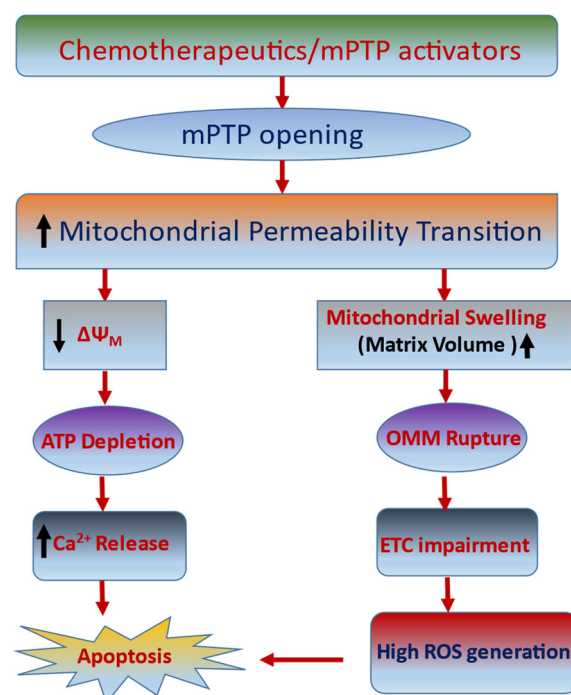


Figure 2. Plausible role of chemotherapeutics or modulators targeting mPTP in cancer. In cancer therapy, mPTP remains open, which causes the mitochondrial membrane potential to deteriorate followed by disruption of the OMM. This mitochondrial impairment results in the depletion of ATP generation, defective ETC, and enhanced Ca^{2+} flux, which leads to a massive generation of ROS and ultimately causes apoptosis.

4. Mitochondrial Participation in Oxidative Stress and Cancer Development

It has been implicated that in healthy replicative eukaryotic cells, mitochondria have shown to be a crucial regulator for prominent cellular processes including proliferation, Ca^{2+} homeostasis, metabolic adaptation, and death. In addition, mitochondria are also the hub for extensive reactions, such as fatty acid oxidation (FAO), the tricarboxylic acid (TCA) cycle, and oxidative phosphorylation (OXPHOS). The primary steps are gluconeogenesis, ketogenesis, and heme biosynthesis [20,21]. In accordance with mitochondrial functionality, it is not surprising that mitochondrial impairment participates in a series of diseases, including cancer [22,23]. There have been more considerable mechanisms wherein mitochondria are involved in the progression of the malignant phenotype over the metabolic reprogramming of cancer cells. Primarily, it is ubiquitously investigated that most cancer diseases are critically linked with DNA mutations affecting mitochondrial functions, predominantly due to the modulation of subunits of the ETC. Interestingly, highlights of cancers including liver and prostate have been alarmingly linked with mutations in mitochondrial Complex I, and brain related cancers also harbor mutations in Complex II at mitochondrial sites [24–26].

Additionally, ROS-induced oxidative stress is a more prominent stimulus/signal for development and progression of cancer. It is evident that ROS is primarily produced in mitochondria for delivering superoxide as a by-product of oxidative phosphorylation. Mitochondrial ROS was shown with a massive production via the TCA cycle or in the ETC [24]. Unlike inducible reactions, ROS could also act as toxic species for cellular macromolecules. An enhanced-level ROS is frequently observed in cancer cells owing to the raised metabolic rates and modified antioxidant potential [27]. Multiple molecular events/changes leading to enhanced cell proliferation and anti-apoptosis are induced in mitochondria during tumorigenesis [28]. One such change is the hindrance of telomere erosion by an integral telomerase expression that establishes the maintenance of telomere length. It has been shown that telomerase reverses the translocation of transcriptase

from nucleus to mitochondria under oxidative stress, restoring mitochondrial functionality and declining oxidative stress and therefore defending mitochondrial DNA and nuclear DNA from oxidative damage [29]. Many investigators have documented increased ROS production that then triggers protumorigenic signaling events, raising cellular proliferation and survival and stimulating DNA damage and genetic alterations [29–32]. Nevertheless, an increased level of ROS has been pointed out as enhancing antitumorigenic machinery by activating oxidative stress-mediated tumor cell death. Tumor cells evolve a cascade wherein an adjustment of high ROS was replenished by altered increased levels of antioxidant molecules to detoxify for maintaining protumorigenic signaling and resistance to apoptosis. Therefore, manipulation of ROS levels has been of great interest for potential cancer therapy. Taken together, mitochondrial dysfunction plays a critical role in cancer development/progression, and the molecular mechanism underlying mitochondria-mediated cancer clinical outcomes remains elusive.

5. Mitochondria as Model Organelle for Regulating Apoptosis Machinery

Each second, billions of cells encounter a well-controlled form of cell death called apoptosis. This cellular event plays a key role in maintaining our health and preventing us from cancer [33]. Cancer has been the result of accumulated gene mutations that result in uncontrolled cell proliferation. It has been well reported that apoptosis activity is frequently suppressed during tumorigenesis through different mechanisms. Likewise, matrix detachment of cells has an imperative effect for metastasis, which is then able to induce a critical stage of apoptosis called anoikis [34]. Such kinds of cell death events could play a key role in cancer therapeutic interventions. In fact, several anticancer therapies were developed based on provoking cell death/apoptosis via targeting/disturbing various mechanisms in cancer cells. In contrast, the anti-apoptosis phenotype observed in cancers play a critical role in promoting tumorigenesis and blocking therapeutic responses. However, apoptotic inhibition promotes cancer, and indeed, tumor cells are frequently more sensitive to cell death than is normal tissue.

In recent years, mitochondrion has been accepted as an excellent model for studying apoptotic cell death, and the molecular mechanism underlying mitochondrial-induced apoptosis is extensively investigated. The mitochondria-mediated apoptotic pathway has been characterized by caspase activation machinery via MOMP and ensuing a release of Cyt C from mitochondria to cytoplasm, thereby causing caspases activation that ultimately leads to cellular shrinkage and blebbing of the plasma membrane [35]. On the contrary, MOMP is controlled by the Bcl-2 family of apoptotic proteins. Such events play a key role in not only normal development and maintenance of tissue homeostasis but also human diseases including cancer [36,37]. An increase in mitochondrial membrane permeability is one of the considerable episodes in apoptotic death. However, the exact mechanism is yet to be clarified. It has also been confirmed that mitochondria-mediated apoptosis is a programmed event that requires some specified protein molecules to either directly or indirectly target cell death in cancer [38–40]. For the past two decades, development of therapeutic drugs and conventional chemotherapies has shown a considerable induction of apoptosis against different kinds of cancers including prostate cancer [41–43].

6. mPTP: A Gate for Apoptotic-Mediated Cancer Cell Death

It has been investigated that targeting these specialized mPTP components could lead to initiating cell death, and therefore mPTP is considered a key player in cancer. To develop novel therapeutic strategies targeting cancer cells, researchers have been exploring the possibility of modulating mPTP activity to inhibit tumor initiation and progression. In recent years, many scientific investigations on mPTP have been performed in cell lines derived from prostate [43], colon [44,45], cervical [46], osteosarcomas [47], pancreatic [48], and other kinds of cancers. In these cancer cell models, different drug and/or molecules were considered to evaluate mitochondrial integrity by examining mitochondrial membrane potential followed by the opening of mPTP due to enhanced mitochondrial oxygen con-

sumption. It has been noted that reduced mitochondrial membrane potential could release Cyt C from mitochondria and activate a series of caspase events to establish the mechanistic model of apoptosis in cancer cells. mPTP has been considered as a molecular mechanism for increasing mitochondrial inner membrane permeability, allowing solutes, ions, or proteins up to 1.5 kDa to pass through [48,49]. mPTP consists of a voltage-dependent anion channel (VDAC) and the adenosine nucleotide transporter (ANT) located on the OMM and IMM, respectively. These two proteins were considered major components of the pore. These molecules and/or proteins are enveloped by a sequence of regulators, such as kinases including hexokinase II (HKII), glycogen synthase kinase 3 β (GSK3 β), and mitochondrial creatine kinase (mtCK); translocator protein (TSPO, formerly known as benzodiazepene receptor); cyclophilin D (CypD, a mitochondrial matrix protein found for peptidylprolyl cis-trans-isomerase activity); and members of the Bcl-2 family [49–52]. Precisely, the two proapoptotic candidates BAX and Bcl-2 were considered as homologous antagonists triggering the mPTP opening, as evident from the BAX- and BAK-knockout rodent models [53]. Previous studies have suggested that inhibiting BAX and BAX causes impairment of mitochondrial Ca²⁺ uptake, triggering mPTP opening by increasing the permeability of the OMM [54].

7. Targeting Mitochondria as Cancer Therapeutic Strategy

It has been well studied that a complex crosstalk occurs between mitochondria and ion channels, proteins and/or enzymes for ATP production, ROS deterioration, and proper supply of glucose at the metabolic platform [55,56]. These key functions are most prominent for cellular survival and growth. In cancer therapeutic intervention, targeting such kinds of channels, enzymes, and proteins linked with mitochondria could lead to a destructions of cancer cells [57]. In the context of ion channels/pores, there are specialized channels/pores that stay on the mitochondrial outer membrane and are responsible for the entrance of almost all the nutrients or ions and expel several apoptosis mediators. Such resident ion channels and/or pores on the mitochondrial outer membrane are considered as translocases that have the ability to transport protein, and the mitochondrial apoptosis-induced channel allows for the release of Cyt C and VDAC, able to transport metabolites. All these specialized channels play a key role in the formation of mPTP. This mPTP induction could further lead to apoptosis [40]. Overproduction of mitochondrial ROS and cytoplasmic Ca²⁺ stimulates mPTP opening, facilitating mitochondrial-mediated apoptosis. In addition, external stimuli-induced mPTP opening could lead to the translocation of Cyt C from the mitochondrial matrix into cytosol followed by strong binding with Apaf-1 and caspases to form a structure called apoptosome, thereby causing caspase-dependent apoptotic cell death pathways in cancer cells. Taken together, mPTP is considered to be a promising therapeutic target. Drugs/compounds targeting mPTP and inducing pore opening could lead to activation of the mPTP-mediated apoptosis cascade in mitochondria, representing a novel therapeutic strategy for cancer treatment. In the following sections, we discuss several therapeutic strategies by targeting mPTP components.

8. Targeting mPTP Components as Novel Cancer Interventions

8.1. Targeting VDAC and HK-II

VDAC is a critical component of mPTP and has been investigated as a frequently targeted hub for cancer therapy. VDAC has also been recognized as mitochondrial porins. Such porins have been implicated as facilitating the transport of key ions including Cl[−], K⁺, Na⁺, and Ca²⁺; the delivery of ATP/ADP and NAD⁺/NADH; the movement of fatty acids and cholesterol; and the transfer of ROS between the inner mitochondrial membrane and cytosol [49]. In mammalian mitochondrial studies, three isoforms of VDAC have been determined: VDAC1, VDAC2, and VDAC3 [58]. Of these three isoforms, VDAC1 has been identified as being present in a more abundant form in mammalian mitochondria and has been shown to play a crucial role in mitochondrial apoptosis [59]. VDAC1 communicates with other apoptosis switching proteins, including members of the Bcl-2, antiapoptotic

proteins such as Bcl-xL and apoptotic-repressing glycolytic enzymes such as HK, could activate cell survival [60]. Once the attachment and/or binding of apoptosis-mediated proteins to VDAC1 is hindered, this leads to an increase in the mitochondrial permeability transition (mPT) (Figure 3). This increase of mPT would further activate proapoptotic proteins (BAX and BAK) and thereby lead to moving across the mitochondrial outer membrane and entering into the cytosol wherein cellular apoptosis is executed [61]. Several compounds are recognized as proapoptotic molecules (cytotoxic agents) that are able to act (target/interfere) on either VDAC1 or HKII. This could inhibit their possible interactions or promote their detachment, inducing cell apoptosis [62,63]. Prezma et al. have synthesized VDAC1-based peptides and considered them as Antp-LP4 and N-terminal-Antp that could specifically destroy peripheral blood mononuclear cells pulled from patients with chronic lymphocytic leukemia. Interestingly, this did not show any impaired effects to normal cells obtained from healthy donors [64]. A similar research group has also formulated one new cell-penetrating peptide synthesized to improve target specificity and cell stability. This peptide was named R-Tf-D-LP4 and consists of a transferrin receptor that has a property to enhance the targeting of a peptide to cancer cells [65]. A study by Yuan et al. has implicated that the endostatin stimulates the opening of mPTP via targeting VDAC1, resulting in infiltration of cytochrome c from mitochondria into the cytosol and ultimately inducing cell apoptosis [66].

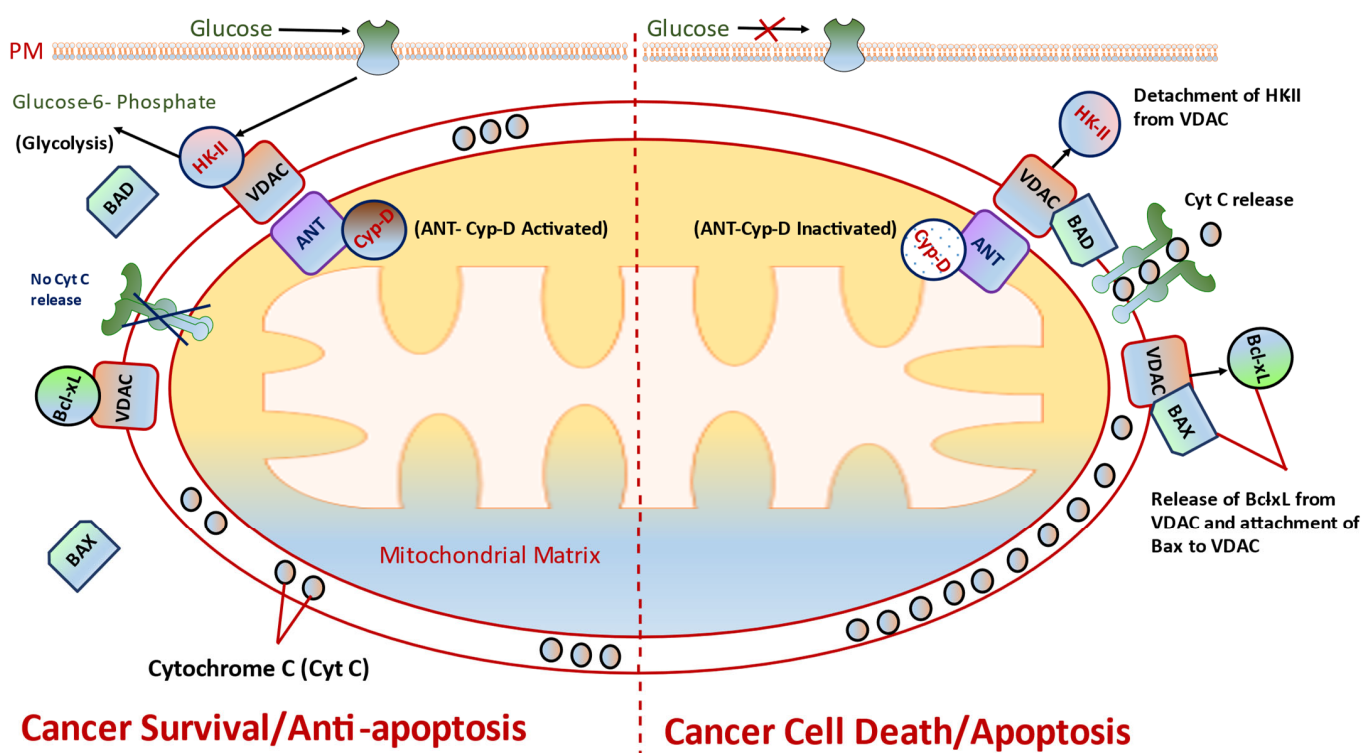


Figure 3. Mitochondrial-bound HKII is a well-considered player for providing glucose supplementation to cancer cells, thereby preventing tumor apoptosis. Targeting HKII or disrupting HKII–VDAC interaction will then initiate cell apoptosis. On left: cancer survival or antiapoptotic state persists in cancer cells within the critical conditions existing in tumor microenvironment. For instance, cytochrome c release, detachment of HKII from VDAC/ANT/CyD, and inactivation of BAX or BAD block the cell apoptotic cascade, and therefore no activation of the mPTP is allowed. On right: cancer cell death/apoptosis. HK-II is capable of detaching from VDAC, which blocks glucose supplementation and inhibits/decreases Bcl-2 proteins that in turn facilitates the switch-on of both apoptotic proteins BAX and BAD and alters both ANT and CyD, ultimately triggering mPTP opening and leading to the release of cytochrome c from the IMM to the cytosol.

Cancer cells exhibit active aerobic glycolysis for their energy production in the form of ATP [67]. Most notably, hexokinase (HK) is a well-recognized enzyme for catalyzing the primary step of glycolysis, wherein glucose conversion to glucose-6-phosphate (G6P) was observed (Figure 3). There are four well-defined isoforms of hexokinase, including HKI, HKII, HKIII, and HKIV. Among all four isoforms, HKII has been characterized and known for high overexpression in cancer cells [68,69]. This kinase has the capability of binding with VDAC1 on the mitochondrial outer membrane, which leads to the interaction of VDAC1 with another mPTP component ANT on the mitochondrial inner membrane. This possible interaction facilitates the coupling of aerobic glycolysis that favors a strong relationship with OXPHOS in metabolic events. Such episodes could also show a rapid rate of glycolysis by prodding HKII to exchange ADP for ATP from the mitochondria [7,70]. Therefore, inhibiting HKII could lead to delaying or ceasing aerobic glycolysis, thereby causing cancer cell death. Therefore, HKII inhibition would not only hinder glycolysis but also repress the antiapoptosis and HKII–VDAC interaction in cancer (Figure 3).

8.2. Targeting CyP-D, a Key Regulator of HKII Binding to VDAC

Previous studies have indicated that CyP-D is overexpressed in many human cancers. Overexpression of CyP-D leads to repression of the apoptosis cascade [71]. Accumulating studies have demonstrated that the antiapoptotic regulation of CyP-D might be critically linked with the modeling of HKII-bound mitochondria [72,73]. This has been implicated as an inactivation of CyP-D with either cyclosporine A or siRNA knockdown of *PPID* (gene encoding CyP-D) to move out HKII from mitochondria. CyP-D is a well-defined mitochondrial matrix protein and is persistent for its alteration of HKII binding to VDAC. As discussed above, ANT at the IMM junction could play a role as mediator for such events [74]. Nevertheless, Chiara et al. have reported an opposite functional role of CyP-D as a proapoptotic protein [75]. Researchers also demonstrated that HKII detachment-provoked apoptosis might be linked with a deterioration of the interaction of CyP-D with ANT. Furthermore, inactivation of CyP-D could also promote an opening of the mPTP (Figure 3). Further studies are needed to clarify the possible role of CyP-D in apoptotic cascade. Disruption of HKII and other proapoptotic proteins to interact with VDAC is required for mPTP opening and is subsequently linked to the mPTP-mediated cell apoptosis. Moreover, the reduced form of CyP-D is involved in mPTP closing, while the oxidized form promotes mPTP opening [75,76]. Earlier reports have suggested an interaction between ATP synthase and the mPTP. The molecules that are considered to regulate mPTP opening/closing also have a binding affinity to ATP synthase and/or CypD. The CypD has more capacity to either directly or indirectly bind to ATP synthase and Bcl-2 family member Bcl-xL and could interact with the β -subunit of ATP synthase, leading to an enhanced efficiency of ATP production in the IMM. In addition, CypD is more capable of binding with ANT. This kind of interaction has reflected a more interesting fact. Therefore, ANT and ATP synthase in synthasomes also consist of mtCK, which binds to ANT [77]. One of the studies conducted by Bernardi et al. showed that mPTP forms at the mitochondrial membranes enclosing the dimers of ATP synthase [9].

8.3. Targets Bcl-2 Family Proteins

Bcl-2 is well recognized as a family member of antiapoptotic B-cell lymphoma 2, which is implicated in tumor episodes including initiation, progression, apoptosis, and/or programmed cellular death and response to chemotherapy [77,78]. Based on its molecular architecture and function, Bcl-2 has been indicated as both antiapoptotic and proapoptotic proteins. At the antiapoptotic platform, four Bcl-2 homology domains (BH1, BH2, BH3, and BH4) includes Bcl-2 and Bcl-2-related protein A1 (Bfl-1/A1), myeloid cell leukemia 1 (Mcl-1), and BCLB/Boo protein [78,79]. Likewise, the proapoptotic platform consists of two subfamilies: the multidomain proapoptotic ‘effectors’ (such as BAK, BAX and BOK) and those members that are known as ‘BH3-only proteins’ (BAD, BID, BIK, BIM, BMF, HRK, PUMA, and NOXA), as they possess only the short BH3 domain [80]. Such key

proteins have great effects on mitochondrial permeabilization. It has been suggested that the activation of BAK and BAX could lead to oligomerization of BAK/BAX that further forms a pore, therefore enhancing mitochondrial permeability and ultimately leading to a sequence of events activating apoptosis cascade [81] (Figure 3).

9. Signaling Pathways Mediating mPTP Opening/Closing in Cancers

9.1. ERK Signaling

In recent years, a cluster of well-recognized oncogenes and/or tumor suppressor genes have shown the ability to alter Ca^{2+} homeostasis responsible for the regulation of mPT induction [13]. Convincingly, such factors are more capable of localizing on the mitochondrial membrane, possibly enhancing both selectivity and efficiency of their regulatory effect in AKT and two tumor suppressors, PML and PTEN. At this juncture, genes such as AKT, PML, and PTEN appear to exist in a complex that interacts with IP₃, and this cascade has demonstrated a particular intention, as AKT and PTEN are both members of the PI3K pathway, which is an extensively investigated signaling mechanism and is considered as a key player for cancer therapies [82]. In the context of mitochondrial events, AKT has shown a great effect on mPTP activation via another mechanism that is indirectly associated to Ca^{2+} signaling [82–85].

In general, AKT has the propensity to induce the phosphorylation of GSK3 β that leads to its inactivation. Notably, GSK3 β kinase is a significant contributor for mPTP modulation. Its inactivation by AKT could enhance the association between HKII and VDAC, leading to an inhibition of mPTP opening and an increase in cancer cell survival [84]. Modulation and alteration in GSK3 β activity have been investigated in different kinds of cancers [85–87]. Moreover, GSK3 β has compelling effects, as it has the strength to act as a convergence factor, which allows survival-signaling molecules to interact with the mPTP. In addition, ERK/GSK3/CyP-D signaling has been implicated as a transduction axis and is considered the critical component for modulation of the mPTP opening/closing in tumor cells. In recent years, ERK has been delineated to be an integral activator of mitochondria in different kinds of cancers including prostate [88–90], osteosarcoma [91], endometrial [92], and Gastric cancer [93]. Several published reports have implicated that ERK activation inhibits the release of apoptogenic proteins, and on the contrary, the interference of ERK could also induce a profound reduction in cellular ATP, leading to loss of mitochondrial membrane potential and inducing cell apoptosis [90]. Additionally, ERK activation suppresses the GSK3-dependent phosphorylation of CyP-D (a mPTP regulator), altering mitochondrial permeability and blocking the opening of mPTP at the mitochondrial junction in cancer cells.

9.2. mTOR Signaling

mTOR, an extensively negative regulator of cellular autophagy, and plays crucial functions at the downstream of AKT signaling. mTOR is frequently upregulated in cancers, and mTOR overexpression could also block the mPTP opening to the same extent as activating AKT and inactivating GSK3 β . In contrast, inhibition of mTOR interferes with the binding of HKII to VDAC, thereby activating cell apoptosis [94]. Therefore, kinase events are more necessarily activated in cancers and could retain HKII bound to the mitochondrial sites by blocking GSK3, hence guarding the closure of mPTP. Such a cascade may also affect the antiapoptotic phenotype of cancer cells and the progress of therapeutic resistance. In contrast, molecular strategy mediating GSK3 activation may enhance the sensitivity of mPTP opening and increase the probability of tumor regression [13].

10. Other Targeting Candidates for mPTP Opening

Cyt C has been considered “a point of no return” causing apoptotic cellular death [6]. mPTP is the only gate-regulating translocation of Cyt C from the mitochondrial inner membrane space to cytosol. Most importantly, Cyt C release is regarded as a critical step of mitochondria-mediated apoptotic cascade. Accordingly, modulation of mPTP

function/opening is more focused on the release of Cyt C from mitochondria and could lead to an opportunity for competent and selective anticancer therapy.

11. Pharmacological Switches of mPTP Opening/Closing in Cancer Therapeutics

11.1. Modulator/Antagonist of Bcl-2 Family Proteins

A novel range of anticancer molecules has developed with the progress of BH3-mimetic agents. Such new agents and/or compounds have been designed in such a way to selectively regulate or kill cancer cells by targeting the cellular machinery involved in their survival [95]. These developed agents can induce apoptosis by mimicking the action of natural negative switches of Bcl-2 and other related proteins [96]. There is an investigation wherein the concept was utilized with selective inhibitors targeting the different antiapoptotic Bcl-2 family members expressed at the mitochondrial level to determine if they influence mitochondrial matrix Ca^{2+} retention capacity and mPTP sensitivity. Bcl-2, Bcl-xL, and Mcl-1 have been reported to play a key role for mPTP opening. In addition, Mcl-1 is mainly located at the mitochondrial outer membrane, the inner membrane, and the matrix that may strengthen Mcl-1 to execute mPTP sensitivity as compared to the other members localized on to the outer membrane. It has been suggested that the inhibition of each antiapoptotic Bcl-2 family member could lead to decreased mitochondrial calcium retention capacity due to the sensitization of mPTP opening.

ABT-737 is the foremost developed agent designed by Abbott Laboratories (North Chicago, IL, USA) and has been considered the prototype of BH3 mimetics, as it was the 'first line' compound established to mimic the function of BH3-only proteins [97]. It has been studied that the obstinacy of several kinds of cancer cells to ABT-737 may be due to ABT-737's inability to target another prosurvival protein, Mcl-1. The predicted influence of targeting such protein mechanisms of apoptosis resistance was investigated in several kinds of cancers [98,99]. Notably, ABT-737 could act as an apoptotic executioner synergistically with chemotherapeutic molecules to overcome the chemoresistance and in particular kill cancer cells without causing severe toxicity [100,101]. Furthermore, ABT-737 has also been reported to sensitize radiation and chemotherapeutic agents including etoposide [102], doxorubicin [103], and cisplatin [104,105] in different cancer cell lines.

ABT-263, an orally bioavailable derivative of ABT-263, is a potent Bcl-2 selective inhibitor [106] (Figure 4). ABT-263 has been shown as an in vitro anticancer activity against a panel of cancer cell lines, either as single agents or in combination with chemotherapeutics [107,108]. In addition, in vivo experiments showed that this Bcl-2 inhibitor could also induce rapid and complete tumor suppression in multiple xenograft models established using small-cell lung cancer [109,110] and hematologic cell lines [111].

To overcome the off-target effect and toxicity linked with Bcl-2 inhibition, Venetoclax (ABT-199), developed by Abbvie, demonstrated a great robustness of anticancer activity against several kinds of hematological malignancies [112,113]. This US FDA-approved drug was employed for the therapeutic intervention of chronic lymphocytic leukemia and small lymphocytic lymphoma as a single therapy and in combination with azacitidine [114,115], decitabine, or low-dose cytarabine [116,117].

In addition, gossypol, a natural phenolic compound found in cotton plants, has been depicted as inhibiting Bcl-2, Bcl-xL, Bcl-W, and Mcl-1 in various cancers, including prostate cancer, breast cancer, leukemia, and glioblastoma [118–122]. Likewise, Gx15-070 (obatoclax), established as a small molecule indole bipyrrrole agent, could show significant inhibition of Bcl-2, Bcl-xL, Bcl-W, and Mcl-1 [123]. This inhibitor was developed and investigated to efficiently disrupt the interaction between BAK and Mcl-1 in multiple myeloma [123,124] (Figure 4).

Furthermore, the organic compound HA14-1 was put forward in as to pointedly inhibit Bcl-2 [124,125]. However, HA14-1 is reported to enhance the sensitivity of cancer cells to chemotherapy and radiotherapy [126–128]. In the context of molecular signature, oblimersen, an 18-mer phosphorothioate Bcl-2 mRNA antisense oligonucleotides, has the capacity to anneal the initiation codon region of Bcl-2 mRNA, thereby inhibiting Bcl-2

biosynthesis [129]. This mRNA was also studied to directly bind to VDAC, decreasing the channel conductance of VDAC [130]. AZD5991, a selective BH3-mimetic, was recently investigated in clinical trials and showed significant inhibition of Mcl-1 in murine lymphoma models and resulted in a prolonged survival of Mcl-expressing PDX mice [131]. BH3 mimetic antagonists of other antiapoptotic proteins have also been developed as A-1155463 and A-1331852 for Bcl-xL or S63845, AMG 176, for Mcl-1 [131–133] in cancer (Figure 4). The overall discussed inhibitors of Bcl-2 have been shown to precisely induce apoptosis in malignant cells and have been greatly demonstrated as single agents and in combination with other drugs in several malignancies, including acute leukemia, lymphomas, and solid tumors (Table 1).

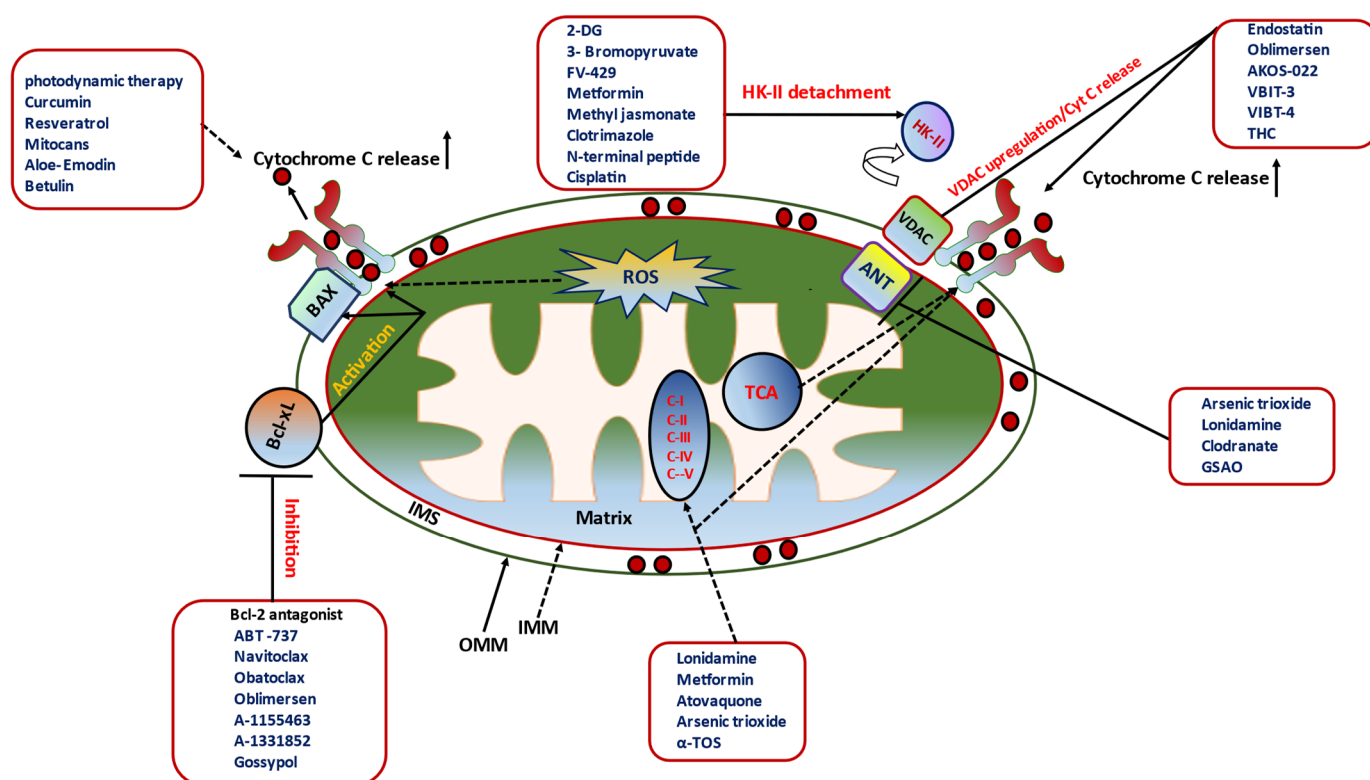


Figure 4. Pharmacologically targeting mPTP-mediated cancer cell death signaling in mitochondrion using chemotherapeutic agents and natural compounds.

11.2. Pharmacological Disruptors of HKII–VDAC Interaction

In the context of glucose analogues employed to target HKII, some key modulators such as 2-deoxy-D-glucose (2-DG) and 3-Bromopyruvate (3-BPA) were reported in previous studies [70,134]. 2-DG administration has been shown a competence with glucose for its entrance into the cell, and once it is penetrated into the cell, it is phosphorylated by HKII to form 2-deoxyglucosephosphate (2-DG-P), which further blocks the glycolysis [70]. 2-DG has been under clinical trials (Phase I/II) to test its efficacies in advanced and hormone refractory prostate cancer (Table 1). It has been shown that a cellular gain of 2-DG-P could also inhibit of HKII [135,136]. The HKII attachment to mitochondria is a crucial step for the initiation of apoptotic cascade. Combined formulations of metformin and 2-DG collapsed ATP in a synergistic way and was observed with a considerable combined therapy in pancreatic cancer cells. In addition to its antimetabolic action, some preclinical investigations have reported that 2-DG possesses an additional anticancer efficacy comprising an antiangiogenic role [137–139] for blocking cancer metastasis. 2-DG also showed an improved efficacy in autophagy inhibition [140–142]. Nevertheless, in vitro and in vivo preclinical investigations suggested possible synergistic therapeutics in combination with conventional chemotherapy, including cisplatin [143,144] and doxorubicin [145,146]. Con-

vincing data have also been displayed in combination with radiotherapy for the treatment of glioblastoma [147,148]. By combining with mitochondria-targeted carboxy-proxyl (Mito-CP), 2-DG has also demonstrated significant tumor degeneration, advocating that a dual focus of mitochondrial bioenergetics metabolism and glycolytic inhibition may represent a promising chemotherapeutic paradigm [149].

3-BPA is also highlighted as an alkylating agent that tends to covalently modulate sulphhydryl groups of HKII. Such kinds of chemical alteration could lead to deterioration of HKII from mitochondria, which provokes apoptosis and further induces cellular destruction [150]. In addition, FV-429 has been recognized as a synthetic flavone that has a potential action to trigger apoptosis in cancer cells. Specifically, FV-429 disrupts the interaction between HKII and VDAC, thereby inhibiting glycolysis-inducing mitochondrial-mediated cell apoptosis [85] (Table 1). An antidiabetic drug, Metformin, could also suppress different kinds of cancers [151–153]. Previous reports suggested that metformin could inhibit HKII in lung carcinoma cells, leading to minimizing the glucose uptake and phosphorylation of glucose [154].

Cancer cells are characterized by enhanced glycolytic activity and decreased mitochondrial oxidation, regardless of oxygen availability (i.e., Warburg effect). The overstimulation of glycolysis causes lactate overproduction, triggering a condition of metabolic acidosis in tumor microenvironment [155]. Dichloroacetate (DCA), an orally available small molecule, prevents the metabolic acidosis condition in tumor microenvironment [155,156]. Specifically, DCA inhibits pyruvate dehydrogenase kinase (PDK), selectively targeting cancer cells and switching their metabolism from glycolysis to oxidative phosphorylation. The enhanced pyruvate flux into mitochondria further triggers mPTP activation, increasing efflux of Cyt C and other apoptotic-inducing factors and overproduction of ROS, subsequently reducing cancer cell viability [156].

Bao et al. have observed that the steroid derivatives extracted from *Ganoderma sinense*, such as (22E, 24R)-6 β -methoxyergosta-7,9, 22-triene-3 β and 5 α -diol, demonstrated a high HKII binding affinity [157] (Figure 4). This was the foremost nutraceutical that is reported to show HKII inhibitory action for cancer therapy [157,158]. Another natural compound, methyl jasmonate, also exhibits an excellent binding affinity with HKII [157]. The high affinity of methyl jasmonate with HKII causes dissociation of HKII from VDAC1, causing mPTP opening, which further promotes cytochrome c release and caspase-mediated cell apoptosis [159,160]. In addition, clotrimazole displays its capacity to induce HKII detachment from mitochondria in different cancer cells and efficiently triggered a dose dependent cell apoptosis [74,161]. Nevertheless, the toxic profile of clotrimazole on mitochondria emerged as an option for interfering mitochondrial respiration by promptly binding to a molecular target other than HKII [162] (Figure 4) (Table 1). In addition, several compounds have already been investigated for their capacity to either directly interact or alter the activity of VDAC [163,164]. Erastin binds to VDAC and disrupts the HKII–VDAC interaction, triggering the opening of mPTP and subsequently inducing cell apoptosis [163,164]. In addition, furanonaphthoquinones and oblimersen could be able to interact with VDAC1 and prevent or inhibit the VDAC1 oligomerization and thereby cause mitochondrial dysfunction and apoptosis, and thus are considered as novel anticancer agents that facilitate the regulation of VDAC.

Shulga et al. have highlighted the importance of HKII–VDAC interaction as one the most prominent components regulating the apoptotic signaling in cells [165]. This research group has synthesized an N-terminal peptide of HKII (15 amino acid residues) that targets VDAC. The therapeutic setup of N-terminal peptide of HKII resulted in dissociation of HKII from the mitochondria to the cytosol. It was also suggested that the dissociation of HKII from the mitochondria may sensitize the mitochondria to the cisplatin-triggered cellular impairment, causing an increased cytotoxicity [165].

11.3. Pharmacological Inhibition of ANT

ANT is considered an excellent target of arsenic trioxide within the mPTP hub. Arsenic trioxide is well proven as a potent agent against acute promyelocytic leukemia [166]. In the context of proapoptotic execution, one of the key mechanisms was established with an apoptotic effect via the opening of the mPTP [167]. Arsenic trioxide was also considered as an agent disrupting mitochondrial membrane potential, and the proapoptotic effect triggered by arsenic trioxide could also be due to its ability to inhibit Bcl-2 [168]. Likewise, Lonidamine (LND) was developed as an indazole carboxylate and has been investigated to target ANT to induce mitochondria-mediated apoptotic cell death [169] (Figure 4). Instead of its cytostatic potential in recurrent glioblastoma multiforme and further improvement of response rate in epirubicin therapy for advanced breast cancer patients, LND was determined to be as toxic to nontumor tissues and was shown to cause hepatotoxicity [170]. Nevertheless, LND has also been considered in clinical trials with the supplementation of chemotherapeutics in several cancers [171,172]. For instance, LND has been reported with limited efficacy tested in lung cancers in both Phase II and III trials [173,174]. In addition, clodronate (a nitrogen-free bisphosphonate) exhibits a competitive inhibition in ANT, causing the impairment of the mitochondrial membrane potential and leading to apoptotic cell death [169]. However, the underlying mechanisms are still unclear. It was also observed that the administration of clodronate to postoperative adjuvant therapy has shown considerable improvement in overall survival among breast cancer patients [175]. Furthermore, GSAO (4-(N-(S-glutathionylacetyl)amino) phenylarsenoxide) is a well-recognized cross-linker to cysteine residues of ANT and showed an inhibition in ATP/ADP exchange via ANT, causing depolarization in the mitochondrial membrane and inducing apoptosis [176] (Figure 4). Interestingly, GSAO is established to be a more promising anticancer drug due to its inhibitory activity on tumor angiogenesis by targeting mitochondria in proliferating endothelial cells [176] (Table 1).

11.4. Pharmacological Triggers of Cyt C Release in Cancer Cell Death

It has been implicated that apoptotic sensors are actively able to trigger Cyt C release that could be employed as a powerful alarm to induce apoptotic cancer cell death [177,178]. Several reports have hypothesized that photodynamic therapy (PDT) is more capable of causing cancer cellular damage by altering mitochondrial membrane potential, thereby stimulating the release of Cyt C and activating other caspase-dependent apoptotic cellular death [179,180]. There is an important recognition for distinct photosensitizers applied in PDT, which was considerably approved for clinical utilization and/or is still under clinical trials [181,182]. In the context of nutraceutical therapy, several compounds have been developed to induce apoptosis-mediated cancer cell death and has displayed promising data of particular interest. Likewise, resveratrol [183,184], curcumin [185], aloemodin [186], and betulin [187] were investigated for trigger apoptosis in several kinds of cancer cell lines via enhancing the release of Cyt C from mPTP to cytosol (Table 1). Resveratrol has been clinically tested under Phase I trials with colon cancer patients. There are some promising clinical trials for using the FDA-approved curcumin for chronic myeloid leukemia, multiple myeloma, and prostate, colorectal, and pancreatic cancer patients [188]. Other compounds, such as mitocans, have been used to target mitochondria and induce apoptosis in breast, colorectal, small-cell lung, and prostate cancer [189,190].

11.5. Pharmacological Modulation of ETC in Cancer Cell Death

ETC complexes act as an entrance gate of electrons in the ETC and are capable of regulating the mitochondrial ATP synthesis by pumping protons toward the intermembrane space. It has been evident that such complexes participate in tumor formation and trigger metastasis by elevating ROS contents. Interestingly, it has been established that complex modulators could alter mPTP architecture. ETC complexes directly interact with mPTP and regulate the mPTP closing/opening status. In addition, cancer cells were placed in a cross talk between the complex junction and mPTP to achieve cellular death and/or

regulate the complex activity. ETC has also been considered as a major culprit in apoptosis in cancer cells. Indeed, metformin has been clinically tested in different phases and trials for cancer patients [191]. So far, extensive data has been reflected toward Phase III clinical trial wherein the potential of metformin was tested for the therapeutic manifestation of breast cancer, low-grade malignant cancers, and other benign cancers [192]. Metformin was shown as a key modulator for the Complex I platform in mitochondria-mediated apoptosis. In contrast to Complex I, no well-developed Complex II inhibitor was discussed at the biomedical hub (Figure 4). Nevertheless, LND has been placed in a research pharmacological rack and shown to hinder Complex II in isolated mitochondria and in DB-1 melanoma cells [193–195]. Atovaquone is another FDA-approved molecule considered to target pneumocystis, pneumonia, and malaria [196]. This drug has been known as a ubiquinone analogue that shows an inhibition of Complex III in cancer cell lines, reducing the mitochondrial oxygen consumption rate and diminishing the hypoxia induced in cancer with considerable doses [197–201]. In addition, arsenic trioxide has been well documented as a Complex IV inhibitor and is a FDA-approved compound for targeting acute promyelocytic leukemia [166]. One of the crucial mechanisms of the arsenic trioxide-mediated apoptosis is through the opening of the mPTP [166,202] (Figure 4). For the past few years, arsenic trioxide-mediated apoptosis has been well demonstrated as mPTP and other mitochondrial-mediated apoptotic cell death in several kinds of cancers [203–206]. At the preclinical stage, this mechanism was shown as a great inhibitor for mitochondria-mediated cellular respiration and exhibited as minimizing hypoxia in lung carcinoma (Table 1).

11.6. Nutraceuticals (Natural Compounds): Excellent Armor for Apoptotic Cancer Cell Death

Nutraceuticals have been known to minimize the level of antiapoptotic signals in cancer cells. Resveratrol is a polyphenolic agent extracted from grapes and wine and has been shown to be able to inhibit mitochondrial ATP synthesis and thereby trigger MOMP [169]. One of the synthetic resveratrol analogues, HS-1793, was screened to disrupt the mitochondrial membrane potential, subsequently promoting release of Cyt C and triggering the opening of mPTP in murine breast cancer cells [207]. Additionally, this analogue was also shown to enhance Bcl-2-mediated antiapoptosis in leukemia cells [208]. Resveratrol has been considered in a few clinical trials in cancer patients [209] (Table 1). In recent years, 4-amido-2,4-pentadieneoate (APD)-class peptide was prepared from a bacterial extract (*Nocardiopsis* sp.) from marine mollusks. Natural contents of such an APD-class have been established as hypoxia-triggered cytotoxins, targeting mitochondria [94]. On the other hand, cernumidine (50) is a well renowned guanidinic alkaloid that exhibits anticancer efficacy via the downregulation of Bcl-2/BAX ratio, leading to loss of mitochondrial membrane permeability with synergistically treated cisplatin in T24 cells [210]. Lycorine (51) was extracted from the Amaryllidaceae family and could cause the mPTP opening and mitochondrial Ca^{2+} and Cyt C release and ultimately induce apoptosis in HepG2 cells [211]. Likewise, amorfrutin C (73) is another nutraceutical armor that belongs to the amorfrutin benzoic acid class of compounds found in *Glycyrrhiza foetida*. The therapeutic regime using amorfrutin C shows considerable disruption of mitochondrial integrity, which results in an irreversible opening of mPTP in HT-29 cells [44]. There is increasing recognition of other nutraceuticals such as allyl isothiocyanate [212], α -conidendrin [213], dehydrobruceine B [214], frugoside [215], methyl caffeate [216], tetrahydrocurcumin [217], phloretin [218], and sesamol [219] for their anticancer capacities. These nutraceuticals demonstrate anticancer capacities through disruption of mitochondria through ROS production, Cyt C release, loss of mitochondrial membrane potential, and modulation of Bcl-2 family members, ultimately leading to mPTP opening and inducing cell apoptosis. In recent decades, curcumin has been extensively employed as a major constituent of turmeric powder from the plant *Curcuma longa*. In the context of its anticancer potential, curcumin is established as an excellent modulator of Bcl-2 family proteins and cellular ROS generation. Correspondingly, betulinic acid, a natural pentacyclic triterpenoid, has been known to stimulate mitochondria-mediated apoptosis in cancer cells [169]. This com-

pound not only enhances MOMP but also inhibits Bcl-2 family proteins in cancer [169,220] (Table 1). Alkaloids, such as phyto-compounds, have been well investigated for their use in inhibiting proliferation and metastasis in cancer cells. Berberine (an alkaloid), derived from the Berberidaceae plant family, exhibits potency either directly on mitochondria or selectively with a possible interaction of ANT at the mPTP juncture [221]. Several other cellular machineries underlying apoptotic induction were triggered by berberine through its modulation on ROS generation and reduction in mitochondrial membrane potential, leading to mPTP opening, the irreversible release of Cyt C, and alteration in the Bcl-2/BAX ratio [169]. In addition, pre- and postsupplementation of vitamins has also played a crucial role in cancer cell impairment. For example, α -Tocopheryl succinate (α -TOS) has been known as a vitamin E analogue competitively bound with ubiquinone to specific Q sites at ETC Complex II. The α -TOS/ETC Complex II binding subsequently leads to the detachment/destabilization of ubiquinone from Complex II, disruption of electron flux, and subsequently overproduction of ROS [222]. It has been known that ROS is more capable of catalyzing the construction of disulfide bridges between the monomers of BAX in the cytosolic compartment, leading to a conformational alteration for dimerization or forming channels by translocated BAX in the outer mitochondrial membrane [223] (Figure 4). Likewise, one hypothetical scheme was also proved with α -TOS-induced apoptosis that could involve the Noxa-Bak axis [224]. Polyphenols, isolated from herbal species, have been demonstrated with promising anticancer activities. For instance, honokiol is a well-established polyphenolic molecule extracted from the genus *Magnolia* [225]. In recent years, both antiangiogenic and antitumor activities of honokiol have been investigated in preclinical studies. It has been suggested that honokiol can induce CyP-D and promote mPTP opening. Notably, the honokiol-induced CyP-D expression further impaired Bcl-2 and Bcl-xL balance, triggering apoptosis in the resistant leukemia cells [226].

Table 1. Overview of the clinical trials of drugs targeting mPTP-mediated apoptotic cascade in cancers.

Compounds/Inhibitors	Targets	Preclinical/Clinical Status	Clinical Trial.gov Identifier/Reference
Bcl-2 Modulators/Inhibitors			
ABT737	Bcl-2 family	Preclinical	[97,100,101]
ABT199	Bcl-2 family	Phase II	NCT01889186 [112,113]
Gossypol	Bcl-2 family	Preclinical	[118,121]
Obatoclax	Bcl-2 family & Mcl-1	Preclinical	[122]
AZD5991	Mcl-1	Phase I	NCT03218683
AMG176	Mcl-1	Phase I	NCT02675452
S63845	Mcl-1	Phase I	NCT02992483
HKII -VDAC-Mediated Metabolic Inhibitors			
3-Bromopyruvate	HKII-VDAC interaction	Preclinical	[70]
2-DG	HKII-DAC interaction	Preclinical Phase I/III	NCT00096707, [135,136]
FV-429	HKII-AkT	Preclinical	[85]
Methyl Jasonate	HKII-VDAC interaction	Preclinical	[159,160]
Clotrimazole	HKII-VDAC interaction	Preclinical	[74,161]
Erastin	HKII-VDAC interaction	Preclinical	[164]
ANT Inhibitor			
Arsenite	ANT proteoliposomes	Preclinical	[168]
Lonidamine	ANT	Preclinical	[171–174]
Clodranate	ANT	Preclinical	[169,175]
GSAO	ADP/ATP exchange via ANT	Preclinical	[176]
Cytochrome C Sensitizer			
Photodynamic therapy	Cyt C	Phase III	NCT02064673, [180,181]
Resveratrol	Cyt C	Preclinical	[183,184]
Curcumin	Cyt C	Phase II	NCT02944578/NCT02782949
Aloe-Emodin	Cyt C	Preclinical	[186]
Betulin	Cyt C	Preclinical	[187]

Table 1. Cont.

Compounds/Inhibitors	Targets	Preclinical/Clinical Status	Clinical Trial.gov Identifier/Reference
ETC Modulators			
Metformin	Complex I	Phase III	NCT01101438, [192]
Lonidamine	Complex II	Phase II/III	NCT00237536/NCT00435448
Atovaquone	Complex III	Phase I/Preclinical	NCT02628080, [195–200]
Arsenic Trioxide	Complex IV	Preclinical	[166,203–206]
Nutraceuticals on mPTP Modulation			
Resveratrol (analogue, HS-1793)	Cyt C/Bcl-2	Preclinical	[207–209]
Cernumidine	Bcl-2/BAX ratio	Preclinical	[210]
Lycorine	mitochondrial Ca^{2+}	Preclinical	[211]
Amorfrutin C	ETC	Preclinical	[44]
Isothiocyanate	Cyt C	Preclinical	[212]
α -conidendrin	Bcl-2	Preclinical	[213]
Dehydrobruceine B	Cyt C	Preclinical	[214]
Frugoside	Cyt C	Preclinical	[215]
Methyl caffeate	Bcl-2	Preclinical	[216]
Phloretin	ETC	Preclinical	[218]
Sesamol	ETC	Preclinical	[219]
Betulinic acid	Bcl-2	Preclinical	[220]
Berberine	ANT	Preclinical	[221]
α -TOS	Complex II	Preclinical	[222,223]
Honokiol	CypD	Preclinical	[164,225]

12. Preclinical and Clinical Models for mPTP-Mediated Cancer Cell Death

In recent years, several studies were published to discuss VDAC-binding compounds and/or agents [164] and their functions on regulating mPTP status in drug-treated colorectal cancer cells. In addition, mitochondrial immunoprecipitation (Mito-IP) assay demonstrated that ANT-1 and Cyp-D formed a complex in erastin-treated HT29 colorectal cells and is considered as a window of opportunity for induction of mPTP opening; the level of cytosolic Cyt C was also increased in HT-29 cells, inducing opening of the mPTP junction [164]. Several pharmacological mPTP blockers, including sanglifehrin A [227,228], were applied to modulate mPTP in cancer cells. Zhang et al. (2018) have suggested that hirsutine, a nutraceutical, could potentially provoke mitochondrial impairment and induce caspase-dependent apoptotic cascade, enhancing the opening of mPTP, in human lung cancer cells. In addition, hirsutine promotes the dephosphorylation of GSK3 β the depletion of ATP and facilitates mPTP-mediated cell apoptosis in cancer cells [229]. In 2021, Zhang et al. observed that Cr(VI) exposure could cause Ca^{2+} elevation and activate the Ca^{2+} -associated signaling mechanism, which is also a main culprit for an extensive opening of mPTP, leading to activation of apoptotic cascade in liver cancer (Hep3B) cells [230]. It has been well noted that in the resting state, CypD stays in the mitochondrial matrix to regulate mPTP closure. Nevertheless, CypD also connects with ANT at the inner mitochondrial membrane to open the mPTP if masking is under crucial conditions [231]. One report has demonstrated that CsA could effectively modulate apoptosis, advocating the cytotoxic potential of esculetin critically linked with CypD, and it was highlighted as a key player in esculetin-induced cell death. This observation was also verified by silencing CypD in human gastric cancer (SGC-7901, MGC-803, BGC-823) cells [232]. One interesting study showed that ABT-737 and curcumin significantly suppress melanoma (WM-115 and B16) cells through a massive induction of cell apoptosis. Furthermore, combining curcumin with ABT-737 synergistically provoked mPTP opening in melanoma cells [233]. Recently, Xue et al. (2021) found that accumulation of Ca^{2+} in mitochondria would lead to mitochondrial ROS accumulation, $\Delta\Psi_m$ depolarization, mPTP opening, and ultimately cellular death and/or apoptosis established in breast cancer cell lines MDA-MB-231, MCF-7, and SKBR-3 [234,235].

Moreover, the isolated proteins and reconstituted liposomes established on the VDAC at the OMM and the ANT at the IMM were extensively studied [236,237]. Interestingly, knocking down VDAC2 using siRNA showed enhanced sensitivity to altering mPT in VDAC1^{-/-} and VDAC3^{-/-} mice [236]. In addition, HKII functions as a key component in the mPTP complex and is involved in HKII-mediated tumorigenesis in multiple cancer types [238,239]. Sun and Zhang suggested that miR-143 targets HKII, which suppresses oral squamous cell carcinoma (OSCC) cell growth, invasion, and glucose metabolism in both in vitro and in vivo systems [238–240].

13. Concluding Remarks

In conclusion, this review aims to focus on mitochondrial function in regulating cellular survival/death switching and how mitochondrial dysfunctions execute the translocation of cytochrome c for initiating apoptotic cell death in cancer episodes. Accumulating evidence highlights mPTP as the major gate for the cytochrome c release, and thereby causing apoptotic cancer cell death. We also discussed disruption of MOMP as a more promising possibility that could activate/stimulate upstream proapoptotic signaling cascade that is intermittently deregulated and more resistant to chemotherapeutic agents in cancers. As described in this review, modulating mPTP opening, via small molecules/compounds and natural products, has proven to be a promising target-specific therapeutic regimen in treating cancers. Understanding mPTP structure and functional mechanisms will provide molecular insight and facilitate the development of novel approaches toward improving the quantity of precision medicine in cancers. Further investigation is also summoned to explore the detailed mechanism of MOMP via the mPTP hub in cancers.

We have clarified in this review that permeability of the mitochondrial membranes is a decisive location wherein cancer cellular survival or death could be determined. Moreover, a deeper understanding of the molecular mechanisms underlying mPTP-mediated apoptotic cancer cell death would pave a new path for developing novel therapeutic strategies against cancers.

Funding: This article was supported by NIH/NIGMS grant 5SC1GM127256 (to B.D.W.).

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Bock, F.J.; Tait, S.W. Mitochondria as multifaceted regulators of cell death. *Nat. Rev. Mol. Cell Biol.* **2020**, *21*, 85–100. [\[CrossRef\]](#) [\[PubMed\]](#)
2. Wenner, C.E. Targeting mitochondria as a therapeutic target in cancer. *J. Cell Physiol.* **2012**, *227*, 450–456. [\[CrossRef\]](#) [\[PubMed\]](#)
3. Kim, B.; Song, Y.S. Mitochondrial dynamics altered by oxidative stress in cancer. *Free. Radic. Res.* **2016**, *50*, 1065–1070. [\[CrossRef\]](#) [\[PubMed\]](#)
4. Chakraborti, T.; Das, S.; Mondal, M.; Roychoudhury, S.; Chakraborti, S. Oxidant, Mitochondria and Calcium: An Overview. *Cell. Signal.* **1999**, *11*, 77–85. [\[CrossRef\]](#)
5. Vakifahmetoglu-Norberg, H.; Ouchida, A.T.; Norberg, E. The role of mitochondria in metabolism and cell death. *Biochem. Biophys. Res. Commun.* **2017**, *482*, 426–431. [\[CrossRef\]](#)
6. Gogvadze, V.; Orrenius, S.; Zhivotovsky, B. Multiple pathways of cytochrome c release from mitochondria in apoptosis. *Biochim. Biophys. Acta Bioenerg.* **2006**, *1757*, 639–647. [\[CrossRef\]](#)
7. Mathupala, S.P.; Ko, Y.H.; Pedersen, P.L. Hexokinase II: Cancer's double-edged sword acting as both facilitator and gatekeeper of malignancy when bound to mitochondria. *Oncogene* **2006**, *25*, 4777–4786. [\[CrossRef\]](#)
8. Halestrap, A.P. What is the mitochondrial permeability transition pore? *J. Mol. Cell Cardiol.* **2009**, *46*, 821–831. [\[CrossRef\]](#)
9. Bernardi, P.; Rasola, A. Calcium and Cell Death: The Mitochondrial Connection. In *Calcium Signalling and Disease*; Springer: Berlin/Heidelberg, Germany, 2007; Volume 45, pp. 481–506. [\[CrossRef\]](#)
10. Wong, R.; Steenbergen, C.; Murphy, E. Mitochondrial Permeability Transition Pore and Calcium Handling. In *Mitochondrial Bioenergetics*; Springer: Berlin/Heidelberg, Germany, 2012; Volume 810, pp. 235–242. [\[CrossRef\]](#)
11. Haworth, R.A.; Hunter, D.R. Control of the mitochondrial permeability transition pore by high-affinity ADP binding at the ADP/ATP translocase in permeabilized mitochondria. *J. Bioenerg. Biomembr.* **2000**, *32*, 91–96. [\[CrossRef\]](#)
12. Crompton, M. Mitochondria and aging: A role for the permeability transition? *Aging Cell.* **2004**, *3*, 3–6. [\[CrossRef\]](#)
13. Bonora, M.; Giorgi, C.; Pinton, P. Molecular mechanisms and consequences of mitochondrial permeability transition. *Nat. Rev. Mol. Cell Biol.* **2022**, *23*, 266–285. [\[CrossRef\]](#)
14. Rao, V.K.; Carlson, E.A.; Yan, S.S. Mitochondrial permeability transition pore is a potential drug target for neurodegeneration. *Biochim. Biophys. Acta Mol. Basis Dis.* **2014**, *1842*, 1267–1272. [\[CrossRef\]](#)

15. You, J.S.; Jonas, P.A. Cancer genetics and epigenetics: Two sides of the same coin? *Cancer Cell* **2012**, *22*, 9–20. [[CrossRef](#)]
16. Siegel, R.L.; Miller, K.D.; Fuchs, H.E.; Jemal, A. Cancer statistics. *CA Cancer J. Clin.* **2022**, *72*, 7–33. [[CrossRef](#)]
17. Morita, M.; Gravel, S.-P.; Hulea, L.; Larsson, O.; Pollak, M.; St-Pierre, J.; Topisirovic, I. mTOR coordinates protein synthesis, mitochondrial activity and proliferation. *Cell Cycle* **2015**, *14*, 473–480. [[CrossRef](#)]
18. Chen, X.; Zhao, Y.; Luo, W.; Chen, S.; Lin, F.; Zhang, X.; Fan, S.; Shen, X.; Wang, Y.; Liang, G. Celastrol induces ROS-mediated apoptosis via directly targeting peroxiredoxin-2 in gastric cancer cells. *Theranostics* **2020**, *10*, 10290–10308. [[CrossRef](#)]
19. Leanza, L.; Venturini, E.; Kadow, S.; Carpinteiro, A.; Gulbins, E.; Becker, K.A. Targeting a mitochondrial potassium channel to fight cancer. *Cell Calcium* **2014**, *58*, 131–138. [[CrossRef](#)]
20. Vaupel, P.; Multhoff, G. Revisiting the Warburg effect: Historical dogma versus current understanding. *J. Physiol.* **2021**, *599*, 1745–1757. [[CrossRef](#)]
21. Nowinski, S.M.; Solmonson, A.; Rusin, S.F.; Maschek, J.A.; Bensard, C.L.; Fogarty, S.; Jeong, M.-Y.; Lettlova, S.; A Berg, J.; Morgan, J.T.; et al. Author response: Mitochondrial fatty acid synthesis coordinates oxidative metabolism in mammalian mitochondria. *Elife* **2020**, *9*, e58041. [[CrossRef](#)]
22. Luo, Y.; Ma, J.; Lu, W. The Significance of Mitochondrial Dysfunction in Cancer. *Int. J. Mol. Sci.* **2020**, *21*, 5598. [[CrossRef](#)]
23. Hsu, C.-C.; Tseng, L.-M.; Lee, H.-C. Role of mitochondrial dysfunction in cancer progression. *ExBiol. Med.* **2016**, *241*, 1281–1295. [[CrossRef](#)] [[PubMed](#)]
24. Yang, Y.; Karakhanova, S.; Hartwig, W.; D’Haese, J.G.; Philippov, P.P.; Werner, J.; Bazhin, A.V. Mitochondria and Mitochondrial ROS in Cancer: Novel Targets for Anticancer Therapy. *J. Cell. Physiol.* **2016**, *231*, 2570–2581. [[CrossRef](#)] [[PubMed](#)]
25. Nomoto, S.; Yamashita, K.; Koshikawa, K.; Nakao, A.; Sidransky, D. Mitochondrial D-loop mutations as clonal markers in multicentric hepatocellular carcinoma and plasma. *Clin. Cancer Res.* **2002**, *8*, 481–487. [[PubMed](#)]
26. Keith, C.G.; Arnold, R.S.; Petros, J.A. Mitochondrial DNA mutations in prostate cancer bone metastases. *J. Nat. Sci.* **2015**, *1*, e147.
27. Mazat, J.-P.; Devin, A.; Ransac, S. Modelling mitochondrial ROS production by the respiratory chain. *Cell. Mol. Life Sci.* **2020**, *77*, 455–465. [[CrossRef](#)]
28. Guo, L.; Yang, Y.; Sheng, Y.; Wang, J.; Ruan, S.; Han, C. Mechanism of piperine in affecting apoptosis and proliferation of gastric cancer cells via ROS-mitochondria-associated signalling pathway. *J. Cell Mol. Med.* **2021**, *25*, 9513–9522. [[CrossRef](#)]
29. Inghapal, C.; Pal, D.; Czapiewski, R.; Porika, M.; Nelson, G.; Saretzki, G.C. Mitochondrial telomerase protects cancer cells from nuclear DNA damage and apoptosis. *PLoS ONE* **2013**, *8*, e52989.
30. Moloney, J.N.; Cotter, T.G. ROS signalling in the biology of cancer. *Semin. Cell Dev. Biol.* **2018**, *80*, 50–64. [[CrossRef](#)]
31. Wang, X.; Ni, H.; Xu, W.; Wu, B.; Xie, T.; Zhang, C.; Cheng, J.; Li, Z.; Tao, L.; Zhang, Y. Difenconazole induces oxidative DNA damage and mitochondria mediated apoptosis in SH-SY5Y cells. *Chemosphere* **2021**, *283*, 131160. [[CrossRef](#)]
32. Ruiz-Ramos, R.; Lopez-Carrillo, L.; Rios-Perez, A.D.; De Vizcaya-Ruiz, A.; Cebrian, M.E. Sodium arsenite induces ROS generation, DNA oxidative damage, HO-1 and c-Myc proteins, NF-kappaB activation and cell proliferation in human breast cancer MCF-7 cells. *Mutat. Res.* **2009**, *674*, 109–115. [[CrossRef](#)]
33. Strickertsson, J.A.B.; Desler, C.; Martin-Bertelsen, T.; Machado, A.M.D.; Wadström, T.; Winther, O.; Rasmussen, L.J.; Friis-Hansen, L. Enterococcus faecalis Infection Causes Inflammation, Intracellular Oxphos-Independent ROS Production, and DNA Damage in Human Gastric Cancer Cells. *PLoS ONE* **2013**, *8*, e63147. [[CrossRef](#)]
34. Schwartz, G.K.; Shah, M.A. Targeting the cell cycle: A new approach to cancer therapy. *J. Clin. Oncol.* **2005**, *23*, 9408–9421. [[CrossRef](#)]
35. Paoli, P.; Giannoni, E.; Chiarugi, P. Anoikis molecular pathways and its role in cancer progression. *Biochim. Biophys. Acta Mol. Cell Res.* **2013**, *1833*, 3481–3498. [[CrossRef](#)]
36. Waseem, M.; Sahu, U.; Salman, M.; Choudhury, A.; Kar, S.; Tabassum, H.; Parvez, S. Melatonin pre-treatment mitigates SHSY-5Y cells against oxaliplatin induced mitochondrial stress and apoptotic cell death. *PLoS ONE* **2017**, *12*, e0180953. [[CrossRef](#)]
37. Lopez, J.; Tait, S.W. Mitochondrial apoptosis: Killing cancer using the enemy within. *Br. J. Cancer* **2015**, *112*, 957–962. [[CrossRef](#)]
38. Llambi, F.; Green, D.R. Apoptosis and oncogenesis: Give and take in the BCL-2 family. *Curr. Opin. Genet. Dev.* **2011**, *21*, 12–20. [[CrossRef](#)]
39. Cidado, J.; Boiko, S.; Proia, T.; Ferguson, D.; Criscione, S.W.; Martin, M.S.; Pop-Damkov, P.; Su, N.; Franklin, V.N.R.; Chilamakuri, C.S.R.; et al. AZD4573 Is a Highly Selective CDK9 Inhibitor That Suppresses MCL-1 and Induces Apoptosis in Hematologic Cancer Cells. *Clin. Cancer Res.* **2020**, *26*, 922–934. [[CrossRef](#)]
40. Ji, B.L.; Xia, L.P.; Zhou, F.X.; Mao, G.Z.; Xu, L.X. Aconitine induces cell apoptosis in human pancreatic cancer via NF-κB signaling pathway. *Eur. Rev. Med. Pharmacol. Sci.* **2016**, *20*, 4955–4964.
41. Vasan, K.; Werner, M.; Chandel, N.S. Mitochondrial Metabolism as a Target for Cancer Therapy. *Cell Metab.* **2020**, *300*, 341–352.
42. Scorrano, L.; Oakes, S.A.; Opferman, J.T.; Cheng, E.H.; Sorcinelli, M.D.; Pozzan, T.; Korsmeyer, S.J. BAX and BAK Regulation of Endoplasmic Reticulum Ca²⁺: A Control Point for Apoptosis. *Science* **2003**, *300*, 135–139. [[CrossRef](#)]
43. Zhang, L.Y.; Wu, Y.L.; Gao, X.H.; Guo, F. Mitochondrial protein cyclophilin-D-mediated programmed necrosis attributes to berberine-induced cytotoxicity in cultured prostate cancer cells. *Biochem. Biophys. Res. Commun.* **2014**, *450*, 697–703. [[CrossRef](#)]
44. Weidner, C.; Rousseau, M.; Micikas, R.J.; Fischer, C.; Plauth, A.; Wowro, S.J.; Siems, K.; Hetterling, G.; Kliem, M.; Schroeder, F.C.; et al. Amorfrutin C Induces Apoptosis and Inhibits Proliferation in Colon Cancer Cells through Targeting Mitochondria. *J. Nat. Prod.* **2016**, *79*, 2–12. [[CrossRef](#)] [[PubMed](#)]
45. Ji, Y.B.; Ji, C.F.; Zhang, H. Laminarin induces apoptosis of human colon cancer LOVO cells through a mitochondrial pathway. *Molecules* **2012**, *17*, 9947–9960. [[CrossRef](#)] [[PubMed](#)]

46. Wang, F. Role of mitochondria and mitochondrial cytochrome c in tubeimoside I-mediated apoptosis of human cervical carcinoma HeLa cell line. *Cancer Chemother. Pharmacol.* **2006**, *57*, 389–399. [[CrossRef](#)] [[PubMed](#)]
47. Chen, Y.; Li, M.; Li, Z.; Gao, P.; Zhou, X.; Zhang, J. Bufalin induces apoptosis in the U-2OS human osteosarcoma cell line via triggering the mitochondrial pathway. *Mol. Med. Rep.* **2016**, *13*, 817–822. [[CrossRef](#)]
48. Yao, L.C. Panax notoginseng Saponins Promote Cell Death and Chemosensitivity in Pancreatic Cancer through the Apoptosis and Autophagy Pathways. *Anticancer Agents Med. Chem.* **2021**, *21*, 1680–1688. [[CrossRef](#)]
49. Kinnally, K.W.; Peixoto, P.M.; Ryu, S.-Y.; Dejean, L.M. Is mPTP the gatekeeper for necrosis, apoptosis, or both? *Biochim. Biophys. Acta* **2011**, *1813*, 616–622. [[CrossRef](#)]
50. Shoshan-Barmatz, V.; Mizrahi, D. VDAC1: From structure to cancer therapy. *Front. Oncol.* **2012**, *2*, 164. [[CrossRef](#)]
51. Bonora, M.; Pinton, P. The Mitochondrial Permeability Transition Pore and Cancer: Molecular Mechanisms Involved in Cell Death. *Front. Oncol.* **2014**, *4*, 302. [[CrossRef](#)]
52. Baines, C.P.; Kaiser, R.A.; Purcell, N.H.; Blair, N.S.; Osinska, H.; Hambleton, M.A.; Brunskill, E.W.; Sayen, M.R.; Gottlieb, R.A.; Dorn, G.W., II; et al. Loss of cyclophilin D reveals a critical role for mitochondrial permeability transition in cell death. *Nature* **2005**, *434*, 658–662. [[CrossRef](#)]
53. Kroemer, G.; Galluzzi, L.; Brenner, C. Mitochondrial Membrane Permeabilization in Cell Death. *Physiol. Rev.* **2007**, *87*, 99–163. [[CrossRef](#)]
54. Karch, J.; Kwong, J.Q.; Burr, A.R.; Sargent, M.A.; Elrod, J.W.; Peixoto, P.M.; Martinez-Caballero, S.; Osinska, H.; Cheng, E.H.-Y.; Robbins, J.; et al. Bax and Bak function as the outer membrane component of the mitochondrial permeability pore in regulating necrotic cell death in mice. *Elife* **2013**, *2*, e00772. [[CrossRef](#)]
55. Zunino, S.J.; Storms, D.H. Resveratrol-induced apoptosis is enhanced in acute lymphoblastic leukemia cells by modulation of the mitochondrial permeability transition pore. *Cancer Lett.* **2006**, *240*, 123–134. [[CrossRef](#)]
56. Lemesko, V.V. VDAC electronics: 1. VDAC-hexo(gluc)okinase generator of the mitochondrial outer membrane potential. *Biochim. Biophys. Acta* **2014**, *1838*, 1362–1371. [[CrossRef](#)]
57. Chang, W.M. Mitochondrial calcium-mediated reactive oxygen species are essential for the rapid induction of the grp78 gene in 9L rat brain tumour cells. *Cell Signal.* **2003**, *15*, 57–64. [[CrossRef](#)]
58. Szabo, I.; Zoratti, M.; Biasutto, L. Targeting mitochondrial ion channels for cancer therapy. *Redox Biol.* **2021**, *42*, 101846. [[CrossRef](#)]
59. Messina, A.; Reina, S.; Guarino, F.; De Pinto, V. VDAC isoforms in mammals. *Biochim. Biophys. Acta Biomembr.* **2012**, *1818*, 1466–1476. [[CrossRef](#)]
60. De Stefani, D.; Bononi, A.; Romagnoli, A.; Messina, A.; De Pinto, V.; Pinton, P.; Rizzuto, R. VDAC1 selectively transfers apoptotic Ca²⁺ signals to mitochondria. *Cell Death Differ.* **2012**, *19*, 267–273. [[CrossRef](#)]
61. Vander Heiden, M.G. Bcl-xL promotes the open configuration of the voltage-dependent anion channel and metabolite passage through the outer mitochondrial membrane. *J. Biol. Chem.* **2001**, *276*, 19414–19419. [[CrossRef](#)]
62. Tsujimoto, Y.; Shimizu, S. Role of the mitochondrial membrane permeability transition in cell death. *Apoptosis* **2007**, *12*, 835–840. [[CrossRef](#)]
63. Shoshan-Barmatz, V.; Zakar, M.; Rosenthal, K.; Abu-Hamad, S. Key regions of VDAC1 functioning in apoptosis induction and regulation by hexokinase. *Biochim. Biophys. Acta Bioenerg.* **2009**, *1787*, 421–430. [[CrossRef](#)] [[PubMed](#)]
64. Prezma, T.; Shteinfer, A.; Admoni, L.; Raviv, Z.; Sela, I.; Levi, I.; Shoshan-Barmatz, V. VDAC1-based peptides: Novel pro-apoptotic agents and potential therapeutics for B-cell chronic lymphocytic leukemia. *Cell Death Dis.* **2013**, *4*, e809. [[CrossRef](#)] [[PubMed](#)]
65. Shteinfer-Kuzmine, A.; Amsalem, Z.; Arif, T.; Zooravlov, A.; Shoshan-Barmatz, V. Selective induction of cancer cell death by VDAC 1-based peptides and their potential use in cancer therapy. *Mol. Oncol.* **2018**, *12*, 1077–1103. [[CrossRef](#)] [[PubMed](#)]
66. Yuan, S.; Fu, Y.; Wang, X.; Shi, H.; Huang, Y.; Song, X.; Li, L.; Song, N.; Luo, Y. Voltage-dependent anion channel 1 is involved in endostatin-induced endothelial cell apoptosis. *FASEB J.* **2008**, *22*, 2809–2820. [[CrossRef](#)] [[PubMed](#)]
67. Shoshan-Barmatz, V.; Shteinfer-Kuzmine, A.; Verma, A. VDAC1 at the Intersection of Cell Metabolism, Apoptosis, and Diseases. *Biomolecules* **2020**, *10*, 1485. [[CrossRef](#)]
68. Abbaszadeh, Z.; Çeşmeli, S.; Avci, B. Crucial players in glycolysis: Cancer progress. *Gene* **2020**, *726*, 144158. [[CrossRef](#)]
69. Patra, K.C.; Wang, Q.; Bhaskar, P.T.; Miller, L.; Wang, Z.; Wheaton, W.; Chandel, N.; Laakso, M.; Muller, W.J.; Allen, E.L.; et al. Hexokinase 2 Is Required for Tumor Initiation and Maintenance and Its Systemic Deletion Is Therapeutic in Mouse Models of Cancer. *Cancer Cell* **2013**, *24*, 213–228. [[CrossRef](#)]
70. Krasnov, G.S.; Dmitriev, A.A.; Lakunina, V.A.; A Kirpiy, A.; Kudryavtseva, A.V. Targeting VDAC-bound hexokinase II: A promising approach for concomitant anti-cancer therapy. *Expert Opin. Ther. Targets* **2013**, *17*, 1221–1233. [[CrossRef](#)]
71. Lin, D.T.; Lechleiter, J.D. Mitochondrial targeted cyclophilin D protects cells from cell death by peptidyl prolyl isomerization. *J. Biol. Chem.* **2002**, *277*, 31134–31141. [[CrossRef](#)]
72. Schubert, A.; Grimm, S. Cyclophilin D, a component of the permeability transition-pore, is an apoptosis repressor. *Cancer Res.* **2004**, *64*, 85–93. [[CrossRef](#)]
73. Machida, K.; Ohta, Y.; Osada, H. Suppression of Apoptosis by Cyclophilin D via Stabilization of Hexokinase II Mitochondrial Binding in Cancer Cells. *J. Biol. Chem.* **2006**, *281*, 14314–14320. [[CrossRef](#)]
74. Chiara, F.; Castellaro, D.; Marin, O.; Petronilli, V.; Brusilow, W.S.; Juhaszova, M.; Sollott, S.J.; Forte, M.; Bernardi, P.; Rasola, A. Hexokinase II Detachment from Mitochondria Triggers Apoptosis through the Permeability Transition Pore Independent of Voltage-Dependent Anion Channels. *PLoS ONE* **2008**, *3*, e1852. [[CrossRef](#)]
75. Choudhary, D.; Goykar, H.; Karanwad, T.; Kannaujia, S.; Gadekar, V.; Misra, M. An understanding of mitochondria and its role in targeting nanocarriers for diagnosis and treatment of cancer. *Asian J. Pharm. Sci.* **2021**, *16*, 397–418. [[CrossRef](#)]

76. Folda, A.; Citta, A.; Scalcon, V.; Cali, T.; Zonta, F.; Scutari, G.; Bindoli, A.; Rigobello, M.P. Mitochondrial Thioredoxin System as a Modulator of Cyclophilin D Redox State. *Sci. Rep.* **2016**, *6*, 23071. [\[CrossRef\]](#)
77. Chevrollier, A.; Loiseau, D.; Reynier, P.; Stepien, G. Adenine nucleotide translocase 2 is a key mitochondrial protein in cancer metabolism. *Biochim. Biophys. Acta Bioenerg.* **2011**, *1807*, 562–567. [\[CrossRef\]](#)
78. Suh, D.H.; Kim, M.-K.; Kim, H.S.; Chung, H.H.; Song, Y.S. Mitochondrial permeability transition pore as a selective target for anti-cancer therapy. *Front. Oncol.* **2013**, *3*, 41. [\[CrossRef\]](#)
79. Gogvadze, V.; Orrenius, S.; Zhivotovsky, B. Mitochondria as targets for cancer chemotherapy. *Semin. Cancer Biol.* **2009**, *19*, 57–66. [\[CrossRef\]](#)
80. Liu, Z.; Wild, C.; Ding, Y.; Ye, N.; Chen, H.; Wold, E.A.; Zhou, J. BH4 domain of Bcl-2 as a novel target for cancer therapy. *Drug Discov. Today* **2016**, *21*, 989–996. [\[CrossRef\]](#)
81. Roufayel, R.; Younes, K.; Al-Sabi, A.; Murshid, N. BH3-Only Proteins Noxa and Puma Are Key Regulators of Induced Apoptosis. *Life* **2022**, *12*, 256. [\[CrossRef\]](#)
82. Danese, A.; Patergnani, S.; Bonora, M.; Wieckowski, M.R.; Previati, M.; Giorgi, C.; Pinton, P. Calcium regulates cell death in cancer: Roles of the mitochondria and mitochondria-associated membranes (MAMs). *Biochim. Biophys. Acta Bioenerg.* **2017**, *1858*, 615–627. [\[CrossRef\]](#)
83. Ciardiello, F.; Tortora, G. Inhibition of bcl-2 as cancer therapy. *Ann Oncol.* **2002**, *13*, 501–502. [\[CrossRef\]](#)
84. Papa, A.; Pandolfi, P.P. The PTEN–PI3K Axis in Cancer. *Biomolecules* **2019**, *9*, 153. [\[CrossRef\]](#) [\[PubMed\]](#)
85. Zhou, Y.; Lu, N.; Qiao, C.; Ni, T.; Li, Z.; Yu, B.; Guo, Q.; Wei, L. FV-429 induces apoptosis and inhibits glycolysis by inhibiting Akt-mediated phosphorylation of hexokinase II in MDA-MB-231 cells. *Mol. Carcinog.* **2016**, *55*, 1317–1328. [\[CrossRef\]](#) [\[PubMed\]](#)
86. Ding, L.; Billadeau, D.D. Glycogen synthase kinase-3 β : A novel therapeutic target for pancreatic cancer. *Expert Opin. Ther. Targets* **2020**, *24*, 417–426. [\[CrossRef\]](#) [\[PubMed\]](#)
87. Yu, J.; Liu, D.; Sun, X.; Yang, K.; Yao, J.; Cheng, C.; Wang, C.; Zheng, J. CDX2 inhibits the proliferation and tumor formation of colon cancer cells by suppressing Wnt/ β -catenin signaling via transactivation of GSK-3 β and Axin2 expression. *Cell Death Dis.* **2019**, *10*, 26. [\[CrossRef\]](#)
88. Liu, F.; Wu, X.; Jiang, X.; Qian, Y.; Gao, J. Prolonged inhibition of class I PI3K promotes liver cancer stem cell expansion by augmenting SGK3/GSK-3 β / β -catenin signalling. *J. Exp. Clin. Cancer Res.* **2018**, *37*, 122. [\[CrossRef\]](#)
89. Rasola, A.; Sciacovelli, M.; Chiara, F.; Pantic, B.; Brusilow, W.S.; Bernardi, P. Activation of mitochondrial ERK protects cancer cells from death through inhibition of the permeability transition. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 726–731. [\[CrossRef\]](#)
90. Monick, M.M.; Powers, L.S.; Barrett, C.W.; Hinde, S.; Ashare, A.; Groskreutz, D.J.; Nyunoya, T.; Coleman, M.; Spitz, D.R.; Hunninghake, G.W. Constitutive ERK MAPK Activity Regulates Macrophage ATP Production and Mitochondrial Integrity. *J. Immunol.* **2008**, *180*, 7485–7496. [\[CrossRef\]](#)
91. Fan, J.; Zhou, W.; Ding, Q.; Zhang, J.; Wu, X.; Tang, P.; Zhou, H.; Wan, B.; Yin, G. ERK inhibition sensitizes CZ415-induced anti-osteosarcoma activity in vitro and in vivo. *Oncotarget* **2017**, *8*, 82027–82036.
92. An, J.; Li, L.; Zhang, X. Curcucione C induces apoptosis in endometrial cancer cells via mitochondria-dependent apoptotic and ERK pathway. *Biotechnol. Lett.* **2021**, *43*, 329–338. [\[CrossRef\]](#)
93. Yang, Z.; Zhao, Y.; Yao, Y.; Li, J.; Wang, W.; Wu, X. Equol Induces Mitochondria-Dependent Apoptosis in Human Gastric Cancer Cells via the Sustained Activation of ERK1/2 Pathway. *Mol. Cells* **2016**, *39*, 742–749. [\[CrossRef\]](#)
94. Pastorino, J.G.; Hoek, J.B. Regulation of hexokinase binding to VDAC. *J. Bioenerg. Biomembr.* **2008**, *40*, 171–182. [\[CrossRef\]](#)
95. Ahmad, A.; Sakr, W.A.; Rahman, K.W. Novel targets for detection of cancer and their modulation by chemopreventive natural compounds. *Front. Biosci.* **2012**, *4*, 410–425. [\[CrossRef\]](#)
96. Nemec, K.N.; Khaled, A.R. Therapeutic modulation of apoptosis: Targeting the BCL-2 family at the interface of the mitochondrial membrane. *Yonsei Med. J.* **2008**, *49*, 689–697. [\[CrossRef\]](#)
97. Mark, F.V.D.; Wei, A.H.; Mason, K.D.; Vandenberg, C.J.; Chen, L.; Czabotar, P.E.; Willis, S.N.; Scott, C.L.; Day, C.L.; Cory, S.; et al. The BH3 mimetic ABT-737 targets selective Bcl-2 proteins and efficiently induces apoptosis via Bak/Bax if Mcl-1 is neutralized. *Cancer Cell* **2006**, *10*, 389–399.
98. Bray, K.; Chen, H.-Y.; Karp, C.M.; May, M.; Ganesan, S.; Karantza-Wadsworth, V.; DiPaola, R.S.; White, E. Bcl-2 Modulation to Activate Apoptosis in Prostate Cancer. *Mol. Cancer Res.* **2009**, *7*, 1487–1496. [\[CrossRef\]](#)
99. Lu, Q.; Zhang, Y.; Wang, J.-H. Bag3 promotes resistance to apoptosis through Bcl-2 family members in non-small cell lung cancer. *Oncol. Rep.* **2012**, *27*, 109–113. [\[CrossRef\]](#)
100. Zinn, R.L.; E Gardner, E.; Dobromilskaya, I.; Murphy, S.; Marchionni, L.; Hann, C.L.; Rudin, C.M. Combination treatment with ABT-737 and chloroquine in preclinical models of small cell lung cancer. *Mol. Cancer* **2013**, *12*, 16. [\[CrossRef\]](#)
101. Kutuk, O.; Letai, A. Alteration of the mitochondrial apoptotic pathway is key to acquired paclitaxel resistance and can be reversed by ABT-737. *Cancer Res.* **2008**, *68*, 7985–7994. [\[CrossRef\]](#)
102. Zall, H.; Weber, A.; Besch, R.; Zantl, N.; Häcker, G. Chemotherapeutic drugs sensitize human renal cell carcinoma cells to ABT-737 by a mechanism involving the Noxa-dependent inactivation of Mcl-1 or A1. *Mol. Cancer* **2010**, *9*, 164. [\[CrossRef\]](#)
103. Du, Y.; Wu, J.; Luo, L. Secreted Heat Shock Protein 90 α Attenuated the Effect of Anticancer Drugs in Small-Cell Lung Cancer Cells Through AKT/GSK3 β / β -Catenin Signaling. *Cancer Control.* **2018**, *25*, 1073274818804489. [\[CrossRef\]](#) [\[PubMed\]](#)
104. Kim, E.Y.; Jung, J.Y.; Kim, A.; Chang, Y.S.; Kim, S.K. ABT-737 Synergizes with Cisplatin Bypassing Aberration of Apoptotic Pathway in Non-small Cell Lung Cancer. *Neoplasia* **2017**, *19*, 354–363. [\[CrossRef\]](#) [\[PubMed\]](#)

105. Chen, Z.-J.; Zhang, B.; Pan, S.-H.; Zhao, H.-M.; Zhang, Y.; Feng, W.-H.; Li, Y.-Y.; Cao, X.-C. Bcl-2 inhibitor ABT-737 enhances the cisplatin-induced apoptosis in breast cancer T47D cells. *Zhonghua Zhong Liu Za Zhi* **2011**, *33*, 891–895. [\[PubMed\]](#)
106. Lee, E.Y.; Gong, E.-Y.; Shin, J.-S.; Moon, J.-H.; Shim, H.J.; Kim, S.-M.; Lee, S.; Jeong, J.; Gong, J.H.; Kim, M.J.; et al. Human breast cancer cells display different sensitivities to ABT-263 based on the level of survivin. *Toxicol. In Vitro* **2018**, *46*, 229–236. [\[CrossRef\]](#) [\[PubMed\]](#)
107. Stamelos, V.A. Navitoclax augments the activity of carboplatin and paclitaxel combinations in ovarian cancer cells. *Gynecol Oncol.* **2013**, *128*, 377–382. [\[CrossRef\]](#)
108. Shao, H.; Jing, K.; Mahmoud, E.; Huang, H.; Fang, X.; Yu, C. Apigenin Sensitizes Colon Cancer Cells to Antitumor Activity of ABT-263. *Mol. Cancer Ther.* **2013**, *12*, 2640–2650. [\[CrossRef\]](#)
109. Wang, H.; Hong, B.; Li, X.; Deng, K.; Li, H.; Lui, V.W.Y.; Lin, W. JQ1 synergizes with the Bcl-2 inhibitor ABT-263 against MYCN-amplified small cell lung cancer. *Oncotarget* **2017**, *8*, 86312–86324. [\[CrossRef\]](#)
110. Gardner, E.E.; Connis, N.; Poirier, J.T.; Cope, L.; Dobromilskaya, I.; Gallia, G.L.; Rudin, C.M.; Hann, C.L. Rapamycin rescues ABT-737 efficacy in small cell lung cancer. *Cancer Res.* **2014**, *74*, 2846–2856. [\[CrossRef\]](#)
111. Ackler, S.; Mitten, M.J.; Foster, K.; Oleksijew, A.; Refici, M.; Tahir, S.K.; Xiao, Y.; Tse, C.; Frost, D.J.; Fesik, S.W.; et al. The Bcl-2 inhibitor ABT-263 enhances the response of multiple chemotherapeutic regimens in hematologic tumors in vivo. *Cancer Chemother. Pharmacol.* **2010**, *66*, 869–880. [\[CrossRef\]](#)
112. Scheffold, A.; Jebaraj, B.M.C.; Stilgenbauer, S. Venetoclax: Targeting BCL2 in Hematological Cancers. In *Small Molecules in Hematology. Recent Results in Cancer Research*; Martens, U., Ed.; Springer: Cham, Switzerland, 2018; Volume 212.
113. Jakubowska, M.A.; Kerkhofs, M.; Martines, C.; Efremov, D.; Gerasimenko, J.; Gerasimenko, O.; Petersen, O.; Bultynck, G.; Vervliet, T.; Ferdek, P. ABT-199 (Venetoclax), a BH3-mimetic Bcl-2 inhibitor, does not cause Ca²⁺ -signalling dysregulation or toxicity in pancreatic acinar cells. *Br. J. Pharmacol.* **2019**, *176*, 4402–4415. [\[CrossRef\]](#)
114. Drozd-Sokolowska, J.; Mądry, K.; Siewiorek, K.; Feliksbrat-Bratosiewicz, M.; Stokłosa, T.; Gierej, B.; Stefaniak, A.; Paszkowska-Kowalewska, M.; Sokołowski, J.; Sankowski, B.; et al. The Clinical Tumor Lysis Syndrome in a Patient with Mixed Phenotype Acute Leukemia Under-going Induction with Venetoclax and Azacitidine: A Case Report. *Chemotherapy* **2022**, *67*, 173–177. [\[CrossRef\]](#)
115. Waggoner, M.; Katsetos, P.-C.J.; Thomas, M.E.; Galinsky, B.I.; Fox, P.-C.H. Practical Management of the Venetoclax-Treated Patient in Chronic Lymphocytic Leukemia and Acute Myeloid Leukemia. *J. Adv. Pr. Oncol.* **2022**, *13*, 400–415. [\[CrossRef\]](#)
116. Dinardo, C.D.; Pratz, K.; Pullarkat, V.; Jonas, B.A.; Arellano, M.; Becker, P.S.; Frankfurt, O.; Konopleva, M.; Wei, A.H.; Kantarjian, H.M.; et al. Venetoclax combined with decitabine or azacitidine in treatment-naïve, elderly patients with acute myeloid leukemia. *Blood* **2019**, *133*, 7–17. [\[CrossRef\]](#)
117. Wei, A.H.; Strickland, S.A., Jr.; Hou, J.Z.; Fiedler, W.; Lin, T.L.; Walter, R.B.; Enjeti, A.; Tiong, S.; Savona, M.; Lee, S.; et al. Venetoclax Combined with Low-Dose Cytarabine for Previously Untreated Patients with Acute Myeloid Leukemia: Results from a Phase Ib/II Study. *J. Clin. Oncol.* **2019**, *37*, 1277–1284. [\[CrossRef\]](#)
118. Meng, Y.; Tang, W.; Dai, Y.; Wu, X.; Liu, M.; Ji, Q.; Ji, M.; Pienta, K.; Lawrence, T.; Xu, L. Natural BH3 mimetic (-)-gossypol chemosensitizes human prostate cancer via Bcl-xL inhibition accompanied by increase of Puma and Noxa. *Mol. Cancer Ther.* **2008**, *7*, 2192–2202. [\[CrossRef\]](#)
119. Oliver, C.L.; Miranda, M.B.; Shangary, S.; Land, S.; Wang, S.; E Johnson, D. (-)-Gossypol acts directly on the mitochondria to overcome Bcl-2- and Bcl-X(L)-mediated apoptosis resistance. *Mol. Cancer Ther.* **2005**, *4*, 23–31. [\[CrossRef\]](#)
120. Ye, W.; Chang, H.-L.; Wang, L.-S.; Huang, Y.-W.; Shu, S.; Sugimoto, Y.; Dowd, M.K.; Wan, P.J.; Lin, Y.C. Induction of apoptosis by (-)-gossypol-enriched cottonseed oil in human breast cancer cells. *Int. J. Mol. Med.* **2010**, *26*, 113–119.
121. Warnsmann, V.; Meyer, N.; Hamann, A.; Kögel, D.; Osiewacz, H.D. A novel role of the mitochondrial permeability transition pore in (-)-gossypol-induced mitochondrial dysfunction. *Mech. Ageing Dev.* **2018**, *170*, 45–58. [\[CrossRef\]](#)
122. Azmi, A.S.; Mohammad, R.M. Non-peptidic small molecule inhibitors against Bcl-2 for cancer therapy. *J. Cell Physiol.* **2009**, *218*, 13–21. [\[CrossRef\]](#)
123. O'Brien, S.M.; Claxton, D.F.; Crump, M.; Faderl, S.; Kipps, T.; Keating, M.J.; Viallet, J.; Cheson, B.D. Phase I study of obatoclax mesylate (GX15-070), a small molecule pan-Bcl-2 family antagonist, in patients with advanced chronic lymphocytic leukemia. *Blood* **2009**, *113*, g299–g305. [\[CrossRef\]](#)
124. Nguyen, M.; Cencic, R.; Ertel, F.; Bernier, C.; Pelletier, J.; Roulston, A.; Silvius, J.R.; Shore, G.C. Obatoclax is a direct and potent antagonist of membrane-restricted Mcl-1 and is synthetic lethal with treatment that induces Bim. *BMC Cancer* **2015**, *15*, 568. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Manero, F.; Gautier, F.; Gallenne, T.; Cauquil, N.; Gréé, D.; Cartron, P.-F.; Geneste, O.; Gréé, R.; Vallette, F.M.; Juin, P. The Small Organic Compound HA14-1 Prevents Bcl-2 Interaction with Bax to Sensitize Malignant Glioma Cells to Induction of Cell Death. *Cancer Res.* **2006**, *66*, 2757–2764. [\[CrossRef\]](#) [\[PubMed\]](#)
126. Wang, J.-L.; Liu, D.; Zhang, Z.-J.; Shan, S.; Han, X.; Srinivasula, S.M.; Croce, C.M.; Alnemri, E.S.; Huang, Z. Structure-based discovery of an organic compound that binds Bcl-2 protein and induces apoptosis of tumor cells. *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 7124–7129. [\[CrossRef\]](#) [\[PubMed\]](#)
127. Arisan, E.D.; Kutuk, O.; Tezil, T.; Bodur, C.; Telci, D.; Basaga, H. Small inhibitor of Bcl-2, HA14-1, selectively enhanced the apoptotic effect of cisplatin by modulating Bcl-2 family members in MDA-MB-231 breast cancer cells. *Breast Cancer Res. Treat.* **2010**, *119*, 271–281. [\[CrossRef\]](#) [\[PubMed\]](#)

128. Hamada, N.; Kataoka, K.; Sora, S.; Hara, T.; Omura-Minamisawa, M.; Funayama, T.; Sakashita, T.; Nakano, T.; Kobayashi, Y. The small-molecule Bcl-2 inhibitor HA14-1 sensitizes cervical cancer cells, but not normal fibroblasts, to heavy-ion radiation. *Radiother. Oncol.* **2008**, *89*, 227–230. [\[CrossRef\]](#)
129. Klasa, R.J.; Gillum, A.M.; Klem, R.E.; Frankel, S.R. Oblimersen Bcl-2 antisense: Facilitating apoptosis in anticancer treatment. *Antisense Nucleic Acid Drug Dev.* **2002**, *12*, 193–213. [\[CrossRef\]](#)
130. Moreira, J.N. Bcl-2-targeted antisense therapy (Oblimersen sodium): Towards clinical reality. *Rev. Recent Clin. Trials* **2006**, *1*, 217–235. [\[CrossRef\]](#)
131. Tan, W.; Loke, Y.-H.; Stein, C.; Miller, P.; Colombini, M. Phosphorothioate Oligonucleotides Block the VDAC Channel. *Biophys. J.* **2007**, *93*, 1184–1191. [\[CrossRef\]](#)
132. Liu, T.; Lam, V.; Thieme, E.; Sun, D.; Wang, X.; Xu, F.; Wang, L.; Danilova, O.V.; Xia, Z.; Tyner, J.W.; et al. Pharmacologic Targeting of Mcl-1 Induces Mitochondrial Dysfunction and Apoptosis in B-Cell Lymphoma Cells in a TP53- and BAX-Dependent Manner. *Clin. Cancer Res.* **2021**, *27*, 4910–4922. [\[CrossRef\]](#)
133. De Ridder, I.; Kerkhofs, M.; Veettil, S.P.; Dehaen, W.; Bultynck, G. Cancer cell death strategies by targeting Bcl-2's BH4 domain. *Biochim. Biophys. Acta Mol. Cell Res.* **2021**, *1868*, 118983. [\[CrossRef\]](#)
134. Ho, N.; Morrison, J.; Silva, A.; Coomber, B.L. The effect of 3-bromopyruvate on human colorectal cancer cells is dependent on glucose concentration but not hexokinase II expression. *Biosci. Rep.* **2016**, *36*, e00299. [\[CrossRef\]](#)
135. Pajak, B.; Siwiak, E.; Sołtyka, M.; Pribe, A.; Zieliński, R.; Fokt, I.; Ziemniak, M.; Jaśkiewicz, A.; Borowski, R.; Domoradzki, T.; et al. 2-Deoxy-d-Glucose and Its Analogs: From Diagnostic to Therapeutic Agents. *Int. J. Mol. Sci.* **2019**, *21*, 234. [\[CrossRef\]](#)
136. Roberts, D.J.; Tan-Sah, V.P.; Ding, E.; Smith, J.M.; Miyamoto, S. Hexokinase-II positively regulates glucose starvation-induced autophagy through TORC1 inhibition. *Mol. Cell* **2014**, *53*, 521–533. [\[CrossRef\]](#)
137. Wu, Y.; Gao, W.-N.; Xue, Y.-N.; Zhang, L.-C.; Zhang, J.-J.; Lu, S.-Y.; Yan, X.-Y.; Yu, H.-M.; Su, J.; Sun, L.-K. SIRT3 aggravates metformin-induced energy stress and apoptosis in ovarian cancer cells. *Exp. Cell Res.* **2018**, *367*, 137–149. [\[CrossRef\]](#)
138. Cheng, G.; Zielonka, J.; McAllister, D.L.; Tsai, S.; Dwinell, M.B.; Kalyanaraman, B. Profiling and targeting of cellular bioenergetics: Inhibition of pancreatic cancer cell proliferation. *Br. J. Cancer* **2014**, *111*, 85–93. [\[CrossRef\]](#)
139. Bizjak, M.; Malavašič, P.; Dolinar, K.; Pohar, J.; Pirkmajer, S.; Pavlin, M. Combined treatment with Metformin and 2-deoxy glucose induces detachment of viable MDA-MB-231 breast cancer cells in vitro. *Sci. Rep.* **2017**, *7*, 1761. [\[CrossRef\]](#)
140. Xi, H.; Kurtoglu, M.; Liu, H.; Wangpaichitr, M.; You, M.; Liu, X.; Savaraj, N.; Lampidis, T.J. 2-Deoxy-d-glucose activates autophagy via endoplasmic reticulum stress rather than ATP depletion. *Cancer Chemother. Pharmacol.* **2011**, *67*, 899–910. [\[CrossRef\]](#)
141. DiPaola, R.S.; Dvorzhinski, D.; Thalasila, A.; Garikapaty, V.; Doram, D.; May, M.; Bray, K.; Mathew, R.; Beaudoin, B.; Karp, C.; et al. Therapeutic starvation and autophagy in prostate cancer: A new paradigm for targeting metabolism in cancer therapy. *Prostate* **2008**, *68*, 1743–1752. [\[CrossRef\]](#)
142. Wu, H.; Zhu, H.; Liu, D.X.; Niu, T.-K.; Ren, X.; Patel, R.; Hait, W.N.; Yang, J.-M. Silencing of Elongation Factor-2 Kinase Potentiates the Effect of 2-Deoxy-d-Glucose against Human Glioma Cells through Blunting of Autophagy. *Cancer Res.* **2009**, *69*, 2453–2460. [\[CrossRef\]](#)
143. Zhang, L.; Su, J.; Xie, Q.; Zeng, L.; Wang, Y.; Yi, D.; Yu, Y.; Liu, S.; Li, S.; Xu, Y. 2-Deoxy-d-Glucose Sensitizes Human Ovarian Cancer Cells to Cisplatin by Increasing ER Stress and Decreasing ATP Stores in Acidic Vesicles. *J. Biochem. Mol. Toxicol.* **2015**, *29*, 572–578. [\[CrossRef\]](#)
144. Jalota, A.; Kumar, M.; Das, B.C.; Yadav, A.K.; Chosdol, K.; Sinha, S. Synergistic increase in efficacy of a combination of 2-deoxy-d-glucose and cisplatin in normoxia and hypoxia: Switch from autophagy to apoptosis. *Tumor. Biol.* **2016**, *37*, 12347–12358. [\[CrossRef\]](#) [\[PubMed\]](#)
145. Mustafa, E.H.; Mahmoud, H.T.; Al-Hudhud, M.Y.; Abdalla, M.Y.; Ahmad, I.M.; Yasin, S.R.; Elkarmi, A.Z.; Tahtamouni, L.H. 2-deoxy-D-Glucose Synergizes with Doxorubicin or L-Buthionine Sulfoximine to Reduce Adhesion and Migration of Breast Cancer Cells. *Asian Pac. J. Cancer Prev.* **2015**, *16*, 3213–3222. [\[CrossRef\]](#) [\[PubMed\]](#)
146. Wang, S.-Y.; Wei, Y.-H.; Shieh, D.-B.; Lin, L.-L.; Cheng, S.-P.; Wang, P.-W.; Chuang, J.-H. 2-Deoxy-d-Glucose Can Complement Doxorubicin and Sorafenib to Suppress the Growth of Papillary Thyroid Carcinoma Cells. *PLoS ONE* **2015**, *10*, e0130959. [\[CrossRef\]](#) [\[PubMed\]](#)
147. Singh, D.; Banerji, A.K.; Tripathi, R.P.; Gupta, J.P.; Mathew, T.L.; Ravindranath, T.; Jain, V. Optimizing cancer radiotherapy with 2-deoxy-d-glucose dose escalation studies in patients with glioblastoma multiforme. *Strahlenther. Onkol.* **2005**, *181*, 507–514. [\[CrossRef\]](#)
148. Venkataramana, N.K.; Venkatesh, P.; Dwarakanath, B.; Vani, S. Protective effect on normal brain tissue during a combin\ational therapy of 2-deoxy-d-glucose and hypofractionated irradiation in malignant gliomas. *Asian J. Neurosurg.* **2013**, *8*, 9–14.
149. Cheng, G.; Zielonka, J.; Dranka, B.P.; McAllister, D.; Mackinnon, A.C., Jr.; Joseph, J.; Kalyanaraman, B. Mitochondria-Targeted Drugs Synergize with 2-Deoxyglucose to Trigger Breast Cancer Cell Death. *Cancer Res.* **2012**, *72*, 2634–2644. [\[CrossRef\]](#)
150. Kim, W.; Yoon, J.-H.; Jeong, J.-M.; Cheon, G.-J.; Lee, T.-S.; Yang, J.-I.; Park, S.-C.; Lee, H.-S. Apoptosis-inducing antitumor efficacy of hexokinase II inhibitor in hepatocellular carcinoma. *Mol. Cancer Ther.* **2007**, *6*, 2554–2562. [\[CrossRef\]](#)
151. DeWaal, D.; Nogueira, V.; Terry, A.R.; Patra, K.C.; Jeon, S.-M.; Guzman, G.; Au, J.; Long, C.P.; Antoniewicz, M.R.; Hay, N. Hexokinase-2 depletion inhibits glycolysis and induces oxidative phosphorylation in hepatocellular carcinoma and sensitizes to metformin. *Nat. Commun.* **2018**, *9*, 446. [\[CrossRef\]](#)
152. Wang, T.; Zhang, J.; Hu, M.; Zhang, Y.; Cui, P.; Li, X.; Li, J.; Vestin, E.; Brännström, M.; Shao, L.; et al. Differential Expression Patterns of Glycolytic Enzymes and Mitochondria-Dependent Apoptosis in PCOS Pa-tients with Endometrial Hyperplasia, an Early Hallmark of Endometrial Cancer, In Vivo and the Impact of Metformin In Vitro. *Int. J. Biol. Sci.* **2019**, *15*, 714–725. [\[CrossRef\]](#)

153. Han, C.Y.; Patten, D.A.; Lee, S.G.; Parks, R.J.; Chan, D.W.; Harper, M.-E.; Tsang, B.K. p53 Promotes chemoresponsiveness by regulating hexokinase II gene transcription and metabolic reprogramming in epithelial ovarian cancer. *Mol. Carcinog.* **2019**, *58*, 2161–2174. [\[CrossRef\]](#)
154. Kang, Y.-T.; Hsu, W.-C.; Wu, C.-H.; Hsin, I.-L.; Wu, P.-R.; Yeh, K.-T.; Ko, J.-L. Metformin alleviates nickel-induced autophagy and apoptosis via inhibition of hexokinase-2, activating lipocalin-2, in human bronchial epithelial cells. *Oncotarget* **2017**, *8*, 105536–105552. [\[CrossRef\]](#)
155. Michelakis, E.D.; Webster, L.; Mackey, J.R. Dichloroacetate (DCA) as a potential metabolic-targeting therapy for cancer. *Br. J. Cancer* **2008**, *99*, 989–994. [\[CrossRef\]](#)
156. Tataranni, T.; Piccoli, C. Dichloroacetate (DCA) and Cancer: An Overview towards Clinical Applications. *Oxid. Med. Cell Longev.* **2019**, *2019*, 8201079. [\[CrossRef\]](#)
157. Bao, F.; Yang, K.; Wu, C.; Gao, S.; Wang, P.; Chen, L.; Li, H. New natural inhibitors of hexokinase 2 (HK2): Steroids from *Ganoderma sinense*. *Fitoterapia* **2018**, *125*, 123–129. [\[CrossRef\]](#)
158. Goldin, N.; Arzoin, L.; Heyfets, A.; Israelson, A.; Zaslavsky, Z.; Bravman, T.; Bronner, V.; Notcovich, A.; Shoshan-Barmatz, V.; Flescher, E. Methyl jasmonate binds to and detaches mitochondria-bound hexokinase. *Oncogene* **2008**, *27*, 4636–4643. [\[CrossRef\]](#)
159. Uludağ, D.; Bay, S.; Sucu, B.O.; İpek, Ö.S.; Mohr, T.; Güzel, M.; Karakaş, N. Potential of Novel Methyl Jasmonate Analogs as Anticancer Agents to Metabolically Target HK-2 Activity in Glioblastoma Cells. *Front. Pharmacol.* **2022**, *13*, 828400.
160. Pastorino, J.G.; Shulga, N.; Hoek, J.B. Mitochondrial binding of hexokinase II inhibits Bax-induced cytochrome c release and apoptosis. *J. Biol. Chem.* **2002**, *277*, 7610–7618. [\[CrossRef\]](#)
161. Neary, C.L. Akt inhibition promotes hexokinase 2 redistribution and glucose uptake in cancer cells. *J. Cell Physiol.* **2013**, *228*, 1943–1948. [\[CrossRef\]](#)
162. Reina, S.; De Pinto, V. Anti-Cancer Compounds Targeted to VDAC: Potential and Perspectives. *Curr. Med. Chem.* **2017**, *24*, 4447–4469. [\[CrossRef\]](#)
163. Ben-Hail, D.; Begas-Shvartz, R.; Shalev, M.; Shteinfein-Kuzmine, A.; Gruzman, A.; Reina, S.; De Pinto, V.; Shoshan-Barmatz, V. Novel Compounds Targeting the Mitochondrial Protein VDAC1 Inhibit Apoptosis and Protect against Mitochondrial Dysfunction. *J. Biol. Chem.* **2016**, *291*, 24986–25003. [\[CrossRef\]](#)
164. Huo, H.; Zhou, Z.; Qin, J.; Liu, W.; Wang, B.; Gu, Y. Erastin Disrupts Mitochondrial Permeability Transition Pore (mPTP) and Induces Apoptotic Death of Colorectal Cancer Cells. *PLoS ONE* **2016**, *11*, e0154605. [\[CrossRef\]](#) [\[PubMed\]](#)
165. Shulga, N.; Wilson-Smith, R.; Pastorino, J.G. Hexokinase II detachment from the mitochondria potentiates cisplatin induced cytotoxicity through a caspase-2 dependent mechanism. *Cell Cycle* **2009**, *8*, 3355–3364. [\[CrossRef\]](#) [\[PubMed\]](#)
166. Powell, B.L.; Moser, B.; Stock, W.; Gallagher, R.E.; Willman, C.L.; Stone, R.M.; Rowe, J.M.; Coutre, S.; Feusner, J.H.; Gregory, J.; et al. Arsenic trioxide improves event-free and overall survival for adults with acute promyelocytic leukemia: North American Leukemia Intergroup Study C9710. *Blood* **2010**, *116*, 3751–3757. [\[CrossRef\]](#) [\[PubMed\]](#)
167. Waseem, M.; Tabassum, H.; Parvez, S. Melatonin modulates permeability transition pore and 5-hydroxydecanoate induced KATP channel inhibition in isolated brain mitochondria. *Mitochondrion* **2016**, *31*, 1–8. [\[CrossRef\]](#)
168. Wang, Z.F.; Guo, X. Arsenite-induced apoptosis is prevented by selenite in A375 cell line. *Biol. Trace Elem. Res.* **2011**, *140*, 7–17. [\[CrossRef\]](#)
169. Fulda, S.; Galluzzi, L.; Kroemer, G. Targeting mitochondria for cancer therapy. *Nat. Rev. Drug Discov.* **2010**, *9*, 447–464. [\[CrossRef\]](#)
170. Barbosa, I.A. Mitochondrial remodeling in cancer metabolism and survival: Potential for new therapies. *Biochim. Biophys. Acta* **2012**, *1826*, 238–254. [\[CrossRef\]](#)
171. Mansi, J.L.; De Graeff, A.; Newell, D.R.; Glaholm, J.; Button, D.; Leach, M.; Payne, G.; Smith, I.E. A phase II clinical and pharmacokinetic study of Lonidamine in patients with advanced breast cancer. *Br. J. Cancer* **1991**, *64*, 593–597. [\[CrossRef\]](#)
172. De Lena, M.; Lorusso, V.; Latorre, A.; Fanizza, G.; Gargano, G.; Caporusso, L.; Guida, M.; Catino, A.; Crucitta, E.; Sambiasi, D.; et al. Paclitaxel, cisplatin and lonidamine in advanced ovarian cancer. A phase II study. *Eur. J. Cancer* **2001**, *37*, 364–368. [\[CrossRef\]](#)
173. Cervantes-Madrid, D.; Romero, Y.; Dueñas-González, A. Reviving Lonidamine and 6-Diazo-5-oxo-L-norleucine to Be Used in Combination for Metabolic Cancer Therapy. *BioMed Res. Int.* **2015**, *2015*, 690492. [\[CrossRef\]](#)
174. Cheng, G.; Zhang, Q.; Pan, J.; Lee, Y.; Ouari, O.; Hardy, M.; Zielonka, M.; Myers, C.R.; Zielonka, J.; Weh, K.; et al. Targeting lonidamine to mitochondria mitigates lung tumorigenesis and brain metastasis. *Nat. Commun.* **2019**, *10*, 2205. [\[CrossRef\]](#)
175. Diel, I.J.; Jaschke, A.; Solomayer, E.F.; Gollan, C.; Bastert, G.; Sohn, C.; Schuetz, F. Adjuvant oral clodronate improves the overall survival of primary breast cancer patients with micrometastases to the bone marrow: A long-term follow-up. *Ann. Oncol.* **2008**, *19*, 2007–2011. [\[CrossRef\]](#)
176. Don, A.S. A peptide trivalent arsenical inhibits tumor angiogenesis by perturbing mitochondrial function in angiogenic endothelial cells. *Cancer Cell.* **2003**, *3*, 497–509. [\[CrossRef\]](#)
177. Alotaibi, A.A.; Bepari, A.; Assiri, R.A.; Niazi, S.K.; Nayaka, S.; Rudrappa, M.; Nagaraja, S.K.; Bhat, M.P. Saussurea lappa Exhibits Anti-Oncogenic Effect in Hepatocellular Carcinoma, HepG2 Cancer Cell Line by Bcl-2 Mediated Apoptotic Pathway and Mitochondrial Cytochrome C Release. *Curr. Issues Mol. Biol.* **2021**, *43*, 1114–1132. [\[CrossRef\]](#)
178. Shin, M.; Lee, B.-M.; Kim, O.; Tran, H.N.K.; Lee, S.; Hwangbo, C.; Min, B.-S.; Lee, J.-H. Triterpenoids from *Ziziphus jujuba* induce apoptotic cell death in human cancer cells through mitochondrial reactive oxygen species production. *Food Funct.* **2018**, *9*, 3895–3905. [\[CrossRef\]](#)
179. Qi, S.; Guo, L.; Yan, S.; Lee, R.J.; Yu, S.; Chen, S. Hypocrellin A-based photodynamic action induces apoptosis in A549 cells through ROS-mediated mitochondrial signaling pathway. *Acta Pharm. Sin. B* **2019**, *9*, 279–293. [\[CrossRef\]](#)

180. Mahalingam, S.M.; Ordaz, J.D.; Low, P.S. Targeting of a Photosensitizer to the Mitochondrion Enhances the Potency of Photodynamic Therapy. *ACS Omega* **2018**, *3*, 6066–6074. [\[CrossRef\]](#)
181. Panzarini, E.; Tenuzzo, B.; Dini, L. Photodynamic therapy-induced apoptosis of HeLa cells. *Ann. N. Y. Acad. Sci.* **2009**, *1171*, 617–626.
182. Baskaran, R.; Lee, J.; Yang, S.-G. Clinical development of photodynamic agents and therapeutic applications. *Biomater. Res.* **2018**, *22*, 25. [\[CrossRef\]](#)
183. Sareen, D.; Darjatmoko, S.R.; Albert, D.M.; Polans, A.S. Mitochondria, Calcium, and Calpain are Key Mediators of Resveratrol-Induced Apoptosis in Breast Cancer. *Mol. Pharmacol.* **2007**, *72*, 1466–1475. [\[CrossRef\]](#)
184. Ma, X. Resveratrol-induced mitochondrial dysfunction and apoptosis are associated with Ca²⁺ and mCICR-mediated MPT activation in HepG2 cells. *Mol. Cell Biochem.* **2007**, *302*, 99–109. [\[CrossRef\]](#)
185. Li, Y.; Zhang, S.; Geng, J.X.; Hu, X.Y. Curcumin inhibits human non-small cell lung cancer A549 cell proliferation through regulation of Bcl-2/Bax and cytochrome C. *Asian Pac. J. Cancer Prev.* **2013**, *14*, 4599–4602. [\[CrossRef\]](#)
186. Chen, S.-H.; Lin, K.-Y.; Chang, C.-C.; Fang, C.-L.; Lin, C.-P. Aloe-emodin-induced apoptosis in human gastric carcinoma cells. *Food Chem. Toxicol.* **2007**, *45*, 2296–2303. [\[CrossRef\]](#) [\[PubMed\]](#)
187. Li, Y.; He, K.; Huang, Y.; Zheng, D.; Gao, C.; Cui, L.; Jin, Y.-H. Betulin induces mitochondrial cytochrome c release associated apoptosis in human cancer cells. *Mol. Carcinog.* **2010**, *49*, 630–640. [\[CrossRef\]](#) [\[PubMed\]](#)
188. Arslan, A.K.; Uzunhisarçikli, E.; Yerer, M.B.; Bishayee, A. The golden spice curcumin in cancer: A perspective on finalized clinical trials during the last 10 years. *J. Cancer Res. Ther.* **2022**, *18*, 19–26.
189. Ralph, S.J.; Neuzil, J. Mitochondria as targets for cancer therapy. *Mol. Nutr. Food Res.* **2009**, *53*, 9–28. [\[CrossRef\]](#)
190. Neuzil, J.; Dong, L.-F.; Rohlena, J.; Truksa, J.; Ralph, S.J. Classification of mitocans, anti-cancer drugs acting on mitochondria. *Mitochondrion* **2013**, *13*, 199–208. [\[CrossRef\]](#)
191. Saraei, P.; Asadi, I.; Kakar, M.A.; Moradi-Kor, N. The beneficial effects of metformin on cancer prevention and therapy: A comprehensive review of recent advances. *Cancer Manag. Res.* **2019**, *11*, 3295–3313. [\[CrossRef\]](#)
192. Coyle, C.; Cafferty, F.; Vale, C.; Langley, R. Metformin as an adjuvant treatment for cancer: A systematic review and meta-analysis. *Ann. Oncol.* **2016**, *27*, 2184–2195. [\[CrossRef\]](#)
193. Martinez-Outschoorn, U.E.; Peiris-Pagés, M.; Pestell, R.G.; Sotgia, F.; Lisanti, M.P. Cancer metabolism: A therapeutic perspective. *Nat. Rev. Clin. Oncol.* **2017**, *14*, 11–31. [\[CrossRef\]](#)
194. Guo, L.; Shestov, A.A.; Worth, A.J.; Nath, K.; Nelson, D.S.; Leeper, D.B.; Glickson, J.D.; Blair, I.A. Inhibition of Mitochondrial Complex II by the Anticancer Agent Lonidamine. *J. Biol. Chem.* **2016**, *291*, 42–57. [\[CrossRef\]](#)
195. Nixon, G.L.; Moss, D.M.; Shone, A.E.; Lalloo, D.G.; Fisher, N.; O'Neill, P.M.; Ward, S.A.; Biagini, G.A. Antimalarial pharmacology and therapeutics of atovaquone. *J. Antimicrob. Chemother.* **2013**, *68*, 977–985. [\[CrossRef\]](#)
196. Ashton, T.M.; Fokas, E.; Kunz-Schughart, L.A.; Folkes, L.K.; Anbalagan, S.; Huether, M.; Kelly, C.J.; Pirovano, G.; Buffa, F.M.; Hammond, E.M.; et al. The anti-malarial atovaquone increases radiosensitivity by alleviating tumour hypoxia. *Nat. Commun.* **2016**, *7*, 12308. [\[CrossRef\]](#)
197. Birth, D.; Kao, W.-C.; Hunte, C. Structural analysis of atovaquone-inhibited cytochrome bc1 complex reveals the molecular basis of antimalarial drug action. *Nat. Commun.* **2014**, *5*, 4029. [\[CrossRef\]](#)
198. Dixon, R.; Pozniak, A.L.; Watt, H.M.; Rolan, P.; Posner, J. Single-dose and steady-state pharmacokinetics of a novel microfluidized suspension of atovaquone in human immunodeficiency virus-seropositive patients. *Antimicrob. Agents Chemother.* **1996**, *40*, 556–560. [\[CrossRef\]](#)
199. Fiorillo, M.; Lamb, R.; Tanowitz, H.B.; Mutti, L.; Krstic-Demonacos, M.; Cappello, A.R.; Martinez-Outschoorn, U.E.; Sotgia, F.; Lisanti, M.P. Repurposing atovaquone: Targeting mitochondrial complex III and OXPHOS to eradicate cancer stem cells. *Oncotarget* **2016**, *7*, 34084–34099. [\[CrossRef\]](#)
200. Falloon, J.; Sargent, S.; Piscitelli, S.C.; Bechtel, C.; Lafon, S.W.; Sadler, B.; Walker, R.E.; Kovacs, J.A.; Polis, M.A.; Davey, R.T.; et al. Atovaquone Suspension in HIV-Infected Volunteers: Pharmacokinetics, Pharmacodynamics, and TMP-SMX Interaction Study. *Pharmacother. J. Hum. Pharmacol. Drug Ther.* **1999**, *19*, 1050–1056. [\[CrossRef\]](#)
201. Anderson, N.M. The emerging role and targetability of the TCA cycle in cancer metabolism. *Protein Cell.* **2018**, *9*, 216–237. [\[CrossRef\]](#)
202. Larochette, N.; Decaudinab, D.; Jacotota, E.; Brennerac, C.; Marzo, I.; Susin, S.A.; Zamzamia, N.; Xied, Z.; Reedd, J.; Kroemer, G. Arsenite Induces Apoptosis via a Direct Effect on the Mitochondrial Permeability Transition Pore. *ExCell Res.* **1999**, *249*, 413–421. [\[CrossRef\]](#)
203. Tian, C.; Gao, P.; Zheng, Y.; Yue, W.; Wang, X.; Jin, H.; Chen, Q. Redox status of thioredoxin-1 (TRX1) determines the sensitivity of human liver carcinoma cells (HepG2) to arsenic trioxide-induced cell death. *Cell Res.* **2008**, *18*, 458–471. [\[CrossRef\]](#)
204. Stevens, J.J.; Graham, B.; Dugo, E.; Sumner, B.B.; Ndebele, K.; Tchounwou, P.B. Arsenic Trioxide Induces Apoptosis via Specific Signaling Pathways in HT-29 Colon Cancer Cells. *J. Cancer Sci. Ther.* **2017**, *9*, 298–306. [\[CrossRef\]](#)
205. Tsai, C.-W.; Yang, M.-D.; Hsia, T.-C.; Chang, W.-S.; Hsu, C.-M.; Hsieh, Y.-H.; Chung, J.-G.; Bau, D.-T. Dithiothreitol enhanced arsenic-trioxide-induced cell apoptosis in cultured oral cancer cells via mitochondrial dysfunction and endoplasmic reticulum stress. *Environ. Toxicol.* **2017**, *32*, 17–27. [\[CrossRef\]](#) [\[PubMed\]](#)
206. Shen, Z.Y.; Shen, J.; Cai, W.J.; Hong, C.; Zheng, M.H. The alteration of mitochondria is an early event of arsenic trioxide induced apoptosis in esophageal carcinoma cells. *Int. J. Mol. Med.* **2000**, *5*, 155–158. [\[CrossRef\]](#) [\[PubMed\]](#)
207. Kim, H.-J.; Yang, K.-M.; Park, Y.-S.; Choi, Y.-J.; Yun, J.-H.; Son, C.-H.; Suh, H.-S.; Jeong, M.-H.; Jo, W.-S. The novel resveratrol analogue HS-1793 induces apoptosis via the mitochondrial pathway in murine breast cancer cells. *Int. J. Oncol.* **2012**, *41*, 1628–1634. [\[CrossRef\]](#) [\[PubMed\]](#)

208. Jeong, S.H. A novel resveratrol derivative, HS1793, overcomes the resistance conferred by Bcl-2 in human leukemic U937 cells. *Biochem. Pharmacol.* **2009**, *77*, 1337–1347. [[CrossRef](#)]
209. Nguyen, A.V.; Martinez, M.; Stamos, M.J.; Moyer, M.P.; Planutis, K.; Hope, C.; Holcombe, R.F. Results of a phase I pilot clinical trial examining the effect of plant-derived resveratrol and grape powder on Wnt pathway target gene expression in colonic mucosa and colon cancer. *Cancer Manag. Res.* **2009**, *1*, 25–37.
210. Miranda, M.A.; Mondal, A.; Sachdeva, M.; Cabral, H.; Neto, Y.A.A.H.; Khan, I.; Groppo, M.; McChesney, J.D.; Bastos, J.K. Chemosensitizing Effect of Cernumidine Extracted from *Solanum cernuum* on Bladder Cancer Cells in Vitro. *Chem. Biodivers.* **2019**, *16*, e1900334. [[CrossRef](#)]
211. Liu, W.Y. Lycorine Induces Mitochondria-Dependent Apoptosis in Hepatoblastoma HepG2 Cells Through ROCK1 Activation. *Front. Pharmacol.* **2019**, *10*, 651. [[CrossRef](#)]
212. Bo, P.; Lien, J.-C.; Chen, Y.-Y.; Yu, F.-S.; Lu, H.-F.; Yu, C.-S.; Chou, Y.-C.; Yu, C.-C.; Chung, J.-G. Allyl Isothiocyanate Induces Cell Toxicity by Multiple Pathways in Human Breast Cancer Cells. *Am. J. Chin. Med.* **2016**, *44*, 415–437. [[CrossRef](#)]
213. Hafezi, K.; Hemmati, A.A.; Abbaszadeh, H.; Valizadeh, A.; Makvandi, M. Anticancer activity and molecular mechanisms of α -conidendrin, a polyphenolic compound present in *Taxus yunnanensis*, on human breast cancer cell lines. *Phytother. Res.* **2020**, *34*, 1397–1408. [[CrossRef](#)]
214. Zhao, L.; Wen, Q.; Yang, G.; Huang, Z.; Shen, T.; Li, H.; Ren, D. Apoptosis induction of dehydrobruceine B on two kinds of human lung cancer cell lines through mitochondri-al-dependent pathway. *Phytomedicine* **2016**, *23*, 114–122. [[CrossRef](#)]
215. Song, L.-S.; Jeong, Y.J.; Kim, J.E.; Shin, J.; Jang, S.-W. Frugoside Induces Mitochondria-Mediated Apoptotic Cell Death through Inhibition of Sulfiredoxin Expression in Melanoma Cells. *Cancers* **2019**, *11*, 854. [[CrossRef](#)]
216. Balachandran, C.; Emi, N.; Arun, Y.; Yamamoto, Y.; Ahilan, B.; Sangeetha, B.; Duraipandiyan, V.; Inaguma, Y.; Okamoto, A.; Ignacimuthu, S.; et al. In vitro anticancer activity of methyl caffeate isolated from *Solanum torvum* Swartz. *Fruit Chem. Interactions* **2015**, *242*, 81–90. [[CrossRef](#)]
217. Han, X.; Deng, S.; Wang, N.; Liu, Y.; Yang, X. Inhibitory effects and molecular mechanisms of tetrahydrocurcumin against human breast cancer MCF-7 cells. *Food Nutr. Res.* **2016**, *60*, 30616. [[CrossRef](#)]
218. Duan, H.; Wang, R.; Yan, X.; Liu, H.; Zhang, Y.; Mu, D.; Han, J.; Li, X. Phloretin induces apoptosis of human esophageal cancer via a mitochondria-dependent pathway. *Oncol. Lett.* **2017**, *14*, 6763–6768. [[CrossRef](#)]
219. Liu, Z.; Ren, B.; Wang, Y.; Zou, C.; Qiao, Q.; Diao, Z.; Mi, Y.; Zhu, D.; Liu, X. Sesamol Induces Human Hepatocellular Carcinoma Cells Apoptosis by Impairing Mitochondrial Function and Suppressing Autophagy. *Sci. Rep.* **2017**, *7*, srep45728. [[CrossRef](#)]
220. Selzer, E.; Thallinger, C.; Hoeller, C.; Oberkleiner, P.; Wacheck, V.; Pehamberger, H.; Jansen, B. Betulinic acid-induced Mcl-1 expression in human melanoma—mode of action and functional significance. *Mol. Med.* **2002**, *8*, 877–884. [[CrossRef](#)]
221. Pereira, C.V.; Machado, N.G.; Oliveira, P.J. Mechanisms of berberine (natural yellow 18)-induced mitochondrial dysfunction: Interaction with the ad-enine nucleotide translocator. *Toxicol. Sci.* **2008**, *105*, 408–417. [[CrossRef](#)]
222. Dong, L.F.; Low, P.; Dyason, J.C.; Wang, X.-F.; Prochazka, L.; Witting, P.K.; Freeman, R.; Swettenham, E.; Valis, K.; Liu, J.; et al. Alpha-tocopheryl succinate induces apoptosis by targeting ubiquinone-binding sites in mitochondrial respiratory complex II. *Oncogene* **2008**, *27*, 4324–4335. [[CrossRef](#)]
223. Neuzil, J.; Wang, X.-F.; Dong, L.; Low, P.; Ralph, S. Molecular mechanism of ‘mitocan’-induced apoptosis in cancer cells epitomizes the multiple roles of reactive oxygen species and Bcl-2 family proteins. *FEBS Lett.* **2006**, *580*, 5125–5129. [[CrossRef](#)]
224. Prochazka, L.; Dong, L.-F.; Valis, K.; Freeman, R.; Ralph, S.J.; Turánek, J.; Neuzil, J. α -Tocopheryl succinate causes mitochondrial permeabilization by preferential formation of Bak channels. *Apoptosis* **2010**, *15*, 782–794. [[CrossRef](#)]
225. Fried, L.E.; Arbiser, J.L. Honokiol, a multifunctional antiangiogenic and antitumor agent. *Antioxid. Redox Signal.* **2009**, *11*, 1139–1148. [[CrossRef](#)]
226. Li, L.; Han, W.; Gu, Y.; Qiu, S.; Lu, Q.; Jin, J.; Luo, J.; Hu, X. Honokiol Induces a Necrotic Cell Death through the Mitochondrial Permeability Transition Pore. *Cancer Res.* **2007**, *67*, 4894–4903. [[CrossRef](#)] [[PubMed](#)]
227. Mazure, N.M. VDAC in cancer. *Biochim. Biophys. Acta Bioenerg.* **2017**, *1858*, 665–673. [[CrossRef](#)] [[PubMed](#)]
228. Yang, L.; Zheng, L.-Y.; Tian, Y.; Zhang, Z.-Q.; Dong, W.-L.; Wang, X.-F.; Zhang, X.-Y.; Cao, C. C6 ceramide dramatically enhances docetaxel-induced growth inhibition and apoptosis in cultured breast cancer cells: A mechanism study. *ExCell Res.* **2015**, *332*, 47–59. [[CrossRef](#)] [[PubMed](#)]
229. Zhang, R.; Li, G.; Zhang, Q.; Tang, Q.; Huang, J.; Hu, C.; Liu, Y.; Wang, Q.; Liu, W.; Gao, N.; et al. Hirsutine induces mPTP-dependent apoptosis through ROCK1/PTEN/PI3K/GSK3 β pathway in human lung cancer cells. *Cell Death Dis.* **2018**, *9*, 598. [[CrossRef](#)]
230. Zhang, X.; Wang, Y.; Chen, M.; Zeng, M. Hexavalent chromium-induced apoptosis in Hep3B cells is accompanied by calcium overload, mitochondrial damage, and AIF translocation. *Ecotoxicol. Environ. Saf.* **2021**, *208*, 111391. [[CrossRef](#)]
231. Porter, G.A., Jr.; Beutner, G. Cyclophilin D, Somehow a Master Regulator of Mitochondrial Function. *Biomolecules* **2018**, *8*, 176. [[CrossRef](#)]
232. Pan, H.; Wang, B.-H.; Lv, W.; Jiang, Y.; He, L. Esculetin induces apoptosis in human gastric cancer cells through a cyclophilin D-mediated mitochondrial permeability transition pore associated with ROS. *Chem. Interactions* **2015**, *242*, 51–60. [[CrossRef](#)]
233. Yu, T.; Chen, C.; Sun, Y.; Sun, H.; Li, T.-H.; Meng, J.; Shi, X. ABT-737 sensitizes curcumin-induced anti-melanoma cell activity through facilitating mPTP death pathway. *Biochem. Biophys. Res. Commun.* **2015**, *464*, 286–291. [[CrossRef](#)]
234. Xue, P.; Chen, Q.; Ren, X.; Liu, D.; Yang, X. A novel protoapigenone analog RY10-4 induces apoptosis of breast cancer cells by exacerbating mitochondrial Ca²⁺ influx through mitochondrial calcium uniporter. *Toxicol. Appl. Pharmacol.* **2021**, *433*, 115776. [[CrossRef](#)]

235. Patenaude, A.; Deschesnes, R.G.; Rousseau, J.L.; Petitclerc, E.; Lacroix, J.; Côté, M.-F.C.; Gaudreault, R. New Soft Alkylating Agents with Enhanced Cytotoxicity against Cancer Cells Resistant to Chemotherapeutics and Hypoxia. *Cancer Res.* **2007**, *67*, 2306–2316. [[CrossRef](#)]
236. Gainutdinov, T.; Molkentin, J.D.; Siemen, D.; Ziemer, M.; Debska-Vielhaber, G.; Vielhaber, S.; Gizatullina, Z.; Orynbayeva, Z.; Gellerich, F.N. Knockout of cyclophilin D in *Ppif*^{−/−} mice increases stability of brain mitochondria against Ca²⁺ stress. *Arch. Biochem. Biophys.* **2015**, *579*, 40–46. [[CrossRef](#)]
237. Chin, H.S.; Li, M.X.; Tan, I.K.L.; Ninnis, R.L.; Reljic, B.; Scicluna, K.; Dagley, L.F.; Sandow, J.J.; Kelly, G.L.; Samson, A.L.; et al. VDAC2 enables BAX to mediate apoptosis and limit tumor development. *Nat. Commun.* **2018**, *9*, 4976. [[CrossRef](#)]
238. Shanguan, X.; He, J.; Ma, Z.; Zhang, W.; Ji, Y.; Shen, K.; Yue, Z.; Li, W.; Xin, Z.; Zheng, Q.; et al. SUMOylation controls the binding of hexokinase 2 to mitochondria and protects against prostate cancer tumorigenesis. *Nat. Commun.* **2021**, *12*, 1812. [[CrossRef](#)]
239. Wang, N.N.; Zhang, P.-Z.; Zhang, J.; Wang, H.-N.; Li, L.; Ren, F.; Dai, P.-F.; Li, H.; Lv, X.-F. Penfluridol triggers mitochondrial-mediated apoptosis and suppresses glycolysis in colorectal cancer cells through down-regulating hexokinase-2. *Anat. Rec.* **2021**, *304*, 520–530. [[CrossRef](#)]
240. Sun, X.; Zhang, L. MicroRNA-143 suppresses oral squamous cell carcinoma cell growth, invasion and glucose metabolism through targeting hexokinase 2. *Biosci. Rep.* **2017**, *37*, BSR20160404. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.