



Article

Human Colonoid–Myofibroblast Coculture for Study of Apical Na⁺/H⁺ Exchangers of the Lower Cryptal Neck Region

Azam Salari ¹, Kunyan Zhou ^{1,2}, Katerina Nikolovska ¹, Ursula Seidler ^{1,*} and Mahdi Amiri ^{1,*}

¹ Department of Gastroenterology, Hepatology and Endocrinology, Hannover Medical School, 30625 Hannover, Germany

² Department of Thyroid Surgery, The First Affiliated Hospital, School of Medicine, Zhejiang University, Hangzhou 310027, China

* Correspondence: seidler.ursula@mh-hannover.de (U.S.); amiri.mahdi@mh-hannover.de (M.A.); Tel.: +49-511-532-9427; Fax: +49-511-532-8428 (U.S.)

Supplementary Materials

Table S1. Sequence of primers used for RT-qPCR experiments.

Gene	Forward Primer	Reverse Primer
ACTB	CGAGGACTTGATTGCACATTGTT	TGGGGTGGCTTAGGATGG
AE2	TGCCAAAGGTTCCACACA	CAACTCATTCACTCCACAAAC
ALPI	CGCTTAACCAGTGCAACAC	GTCACCACTCCTACTGACTTC
CFTR	CTACCACTGGTGCATACTCTAATC	ACGTGTTGAGGGTGCACATAG
CLDN1	CTGGGAGGTGCCCTACTTTG	ACACGTAGTCTTCCCGCTG
CLDN2	GATCCTACGGGACTCTACTCA	CAGGGAGAACAGGGAAAGAAATAA
CLDN3	ACCGAGAAGAAGTACACGG	TAGACGTAGTCCTGCGGTC
CLDN4	TTCTACAAATCCGCTGGTGGC	GCGGAGTAAGGCTGTCTGT
CLDN7	GGGGGAGACGACAAAGTGAA	CATACAGGAGCAAGCTACCA
CLDN8	TGCCCAAAAACGTGAGCTTG	TGTGCGATGGGAAGGTATCG
GAPDH	TGCACCACTGCTTAGC	GGCATGGACTGTGGTATGAG
GREM1	GCAAGCCAAGAAATTCACTAC	TGCAACGACACTGCTTCA
KI67	GACCTCAAACGGCTCTAAC	GCTGCCAGATAGACTCAGAAAG
LGR5	CCTGCTGACTTTGAGGAAGACC	CCAGCCATCAAGCAGGTGTTCA
LYZ	GGCTTGCCTCCTTCTGTTA	GTAGCCATCCATTCCAATCT
MUC2	AGTTTGGGAGCACTCGAG	TCTTCCACGCAGTGGTAAC
NBCe1	CCGGCTTGTGGTCACTAT	CAAGTGATACCCCTGCTCCTTC
NBCn1	CTGCTATTCCGTGTTGCTTG	GTGATAGCCAGCTCCTTCTT
NHE1	GCTGGTGGCAGACCCCTACGA	ATAGGCCAGTGGCTGAGCCGA
NHE2	TGTCTACCGTGGCAAGAAC	AACGCAAAACAGATGGCACC
NHE3	ACCGTGCCTACACCATGAAGATG	ATGCGGTAGCGGTTCAGAACCC
NHE8	CATGTGTGTTGCATTCTTGGC	AGCACTATGCACCAGATGACA
NKCC1	AAAGGAACATTCAAGCACAGC	CTAGACACAGCACCTTTCGTG
OCLN	TCGACCAATGCTCTCAGC	CTCCTGGAGGAGAGGTCCAT
PDGFRA	CTTGTCAGTGTGCTCTTGG	GTTCACCTCTGTGGCTATT
RSPO3	GAAAGAGGAGAAAGGAAGGGAAG	GGCTGCCATGTATTCCATAAA
SI	CGCTACACCTTATTACCCCTCC	CCAGCTGTTCGTATCCTCATAAA
SLC26A3	CCAGCGTCTATTCCCTCAAAT	TCCCAGCAAATCCTCTGAATAC
SLC26A6	AGAAACTGCTCAAGAACAGGA	CCATCTTATCTCCTGAGCTCACC
THY1 (CD90)	TGACCCGTGAGACAAAGAAC	GCTAGTGAAGGCGGATAAGTAG
WNT2B	CCTTGGAGTGGTAGCCATAAG	AACGCTGACTGTAGGTATG
WNT5a	GTGATGCAGATAGGCAGCCG	GCCATAGTCGATGTTGTCGC
ZO-1	CCTGAGTTGACACTGGAGTT	GCTGAAGGACTCACAGGAATAG

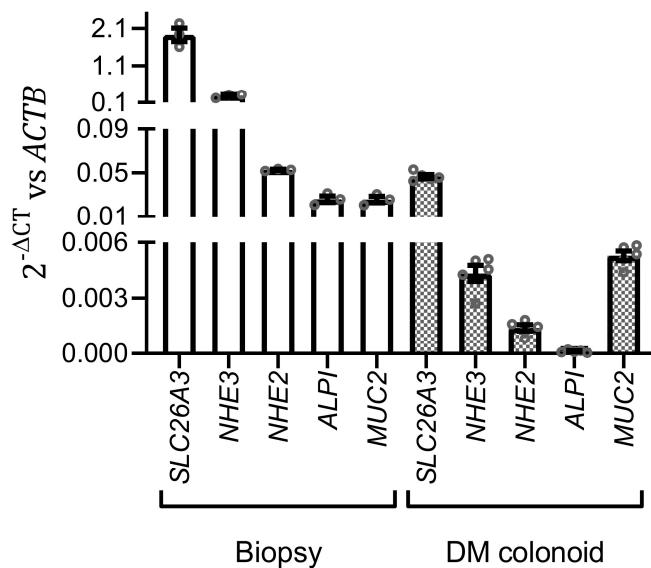


Figure S1. *SLC26A3* mRNA is highly expressed in human colon. RT-qPCR analysis of human transverse colon biopsy material from which the colonoids were derived shows that among the selected panel of genes, *SLC26A3* has substantially higher expression levels. Similarly in differentiated colonoid monolayers, *SLC26A3* expression is drastically higher than other differentiation marker genes including *NHE3*, *MUC2* or *ALPI*.

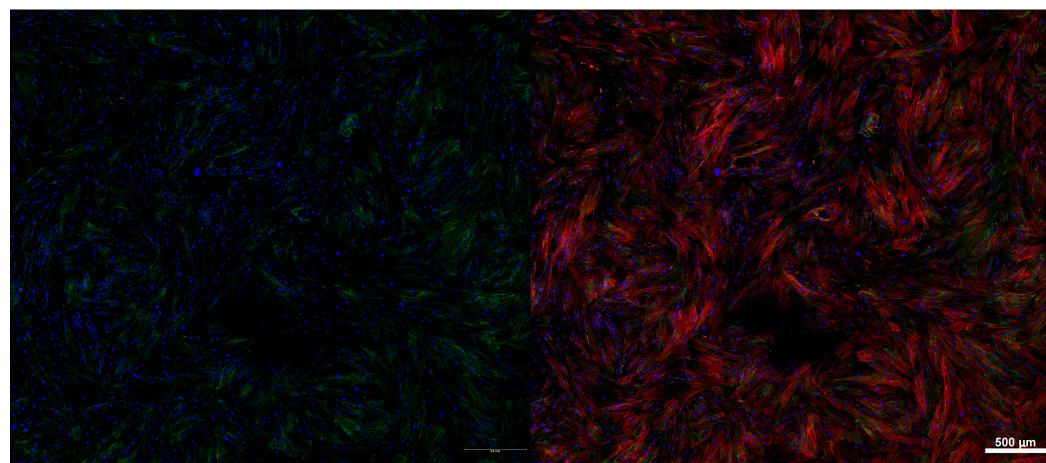


Figure S2. Majority of the cells in the myofibroblast culture used in CM-CE cocultures are CD90-positive. This immunofluorescent image shows intestinal myofibroblasts from the lamina propria of human transverse colon stained for nuclei (blue), CD90 (green) and F-actin (red). The image is generated by stitching multiple tile scan acquisitions to cover a broad area of the culture. The left and right panels are identical, except that in the left panel the F-actin signal is excluded for a better visualization of the CD90 signal. The majority of the myofibroblasts are positively stained for CD90.

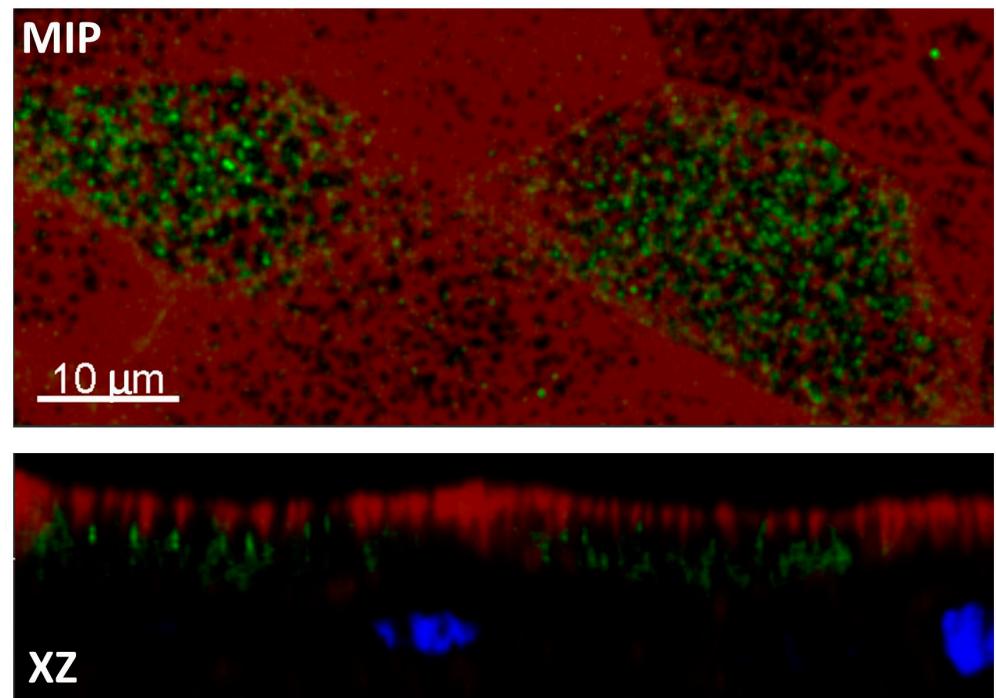


Figure S3. immunohistochemical staining of NHE8 in human CM-CE monolayers. Maximum intensity projection (MIP) and XZ cross section of CM-CE monolayer. NHE8 is detected as organellar structure mainly distributed between nucleus and subapical region. Green: NHE8, Red: F-actin, Blue: nuclei (excluded in MIP).