



Review

Gut Microbiota and Alzheimer's Disease: How to Study and Apply Their Relationship

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Abstract: Gut microbiota (GM), the microorganisms in the gastrointestinal tract, contribute to the regulation of brain homeostasis through bidirectional communication between the gut and the brain. GM disturbance has been discovered to be related to various neurological disorders, including Alzheimer's disease (AD). Recently, the microbiota-gut-brain axis (MGBA) has emerged as an enticing subject not only to understand AD pathology but also to provide novel therapeutic strategies for AD. In this review, the general concept of the MGBA and its impacts on the development and progression of AD are described. Then, diverse experimental approaches for studying the roles of GM in AD pathogenesis are presented. Finally, the MGBA-based therapeutic strategies for AD are discussed. This review provides concise guidance for those who wish to obtain a conceptual and methodological understanding of the GM and AD relationship with an emphasis on its practical application.

Keywords: gut microbiota; Alzheimer's disease; microbiota-gut-brain axis; experimental models; therapeutic strategy



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1. Relationship between Gut Microbiota and Brain Function

Gut microbiota (GM) refers to the microorganisms living in the gastrointestinal (GI) tract [1]. Primarily, they come from all three taxa of microorganisms, including Bacteria, Archaea, and Eukarya, with Bacteria being the most predominant. Among them, the most popular gut bacterial species consist of *Proteobacteria*, *Firmicutes*, *Actinobacteria*, and *Bacteroidetes* [2]. From the evolutionary perspective, the GM has fostered a mutually beneficial relationship with the host in several ways [3]. For example, GM plays a critical role in metabolism by producing enzymes and metabolic intermediates not synthesizable by the host. In addition, the GM resident on the enteric mucosal epithelium serves as an immuno-neutral zone to defend against the entry of pathogenic microorganisms into the host. Beyond its direct effects on host metabolism and immunophysiology, it seems to indirectly impact the communication between the GI tract and central nervous system (CNS) in both health and disease states. In particular, GM can influence the neural network controlling gut function through the intrinsic and extrinsic nervous systems, such as the autonomic nervous system, enteric nervous system, and neuroendocrine and immune systems [4]. As a consequence, GM has been implicated in the regulation of neural development, neurotransmission, and the maintenance of brain homeostasis.

Considering the critical roles of GM in the normal neurophysiology of the host, some researchers suggested its direct involvement in the development of various brain diseases under pathogenic conditions. The current predominant theory postulates that the mechanism of GM-mediated neuropathogenesis involves GM-induced enhanced neuroinflammation and imbalanced inhibitory/excitatory neurotransmission. In this theory, GM dysbiosis, which is defined as a pathogenic imbalance in the gut microbial community, can upregulate local and systemic inflammations by secreting several bacterial endotoxins, typically lipopolysaccharides (LPSs). The translocation of these bacterially secreted LPSs from

the gut to the bloodstream can lead to the so-called “leaky-gut” state, which is characterized by increased intestinal permeability, followed by infiltration of monocytes and secretion of neuroinflammatory cytokines and chemokines [5–7]. In addition, GM dysbiosis can also upregulate the microbial production of a wide range of neurotransmitters and neuromodulators, including dopamine, norepinephrine, serotonin, γ -aminobutyric acid (GABA), and short-chain fatty acids (SCFAs), as well as neurotoxic metabolites, such as D-lactic acid and ammonia [6,8,9]. Such GM-induced neuroinflammation and GM-derived neurochemicals can adversely affect host neural functions and inter-bacterial communication processes in a pathogenic manner, ultimately leading to various brain disorders [8].

To emphasize the critical roles of GM in the normal and pathological regulation of a variety of gut and brain functions, a new term, “microbiota-gut-brain axis” (MGBA), was defined, to describe the communication and influence between GM and brain. MGBA has been implicated in various CNS disorders such as depression, anxiety, autism, and Alzheimer’s disease (AD) [9–13]. Accumulating evidence suggests the direct involvement of GM in the regulation of social behaviors, stress resistance, and cognitive functions [5,14–16]. In particular, a number of studies suggested the pathogenic roles of GM in the development of Alzheimer’s disease, Parkinson’s disease (PD), and Huntington’s disease (HD) [17,18]. In regards to PD, alternation of GM was confirmed in PD patients by two separate studies [19,20]. Interestingly, the presence of α -synuclein-aggregates in the enteric nervous system before their appearance in the brain suggests their gut-to-to-brain spread theory [21]. In line with this, α -synuclein-mediated motor deficits were aggravated by GM in the mouse model [22]. GM dybiosis was also confirmed in the mouse models of HD [23–25]. Based on these observations, already-known pathogenic mechanisms for various brain diseases have been re-evaluated in the context of this newly-found relationship between GM and the brain.

2. Gut Microbiota and Alzheimer’s Disease

AD is one of the most common neurodegenerative diseases, characterized by a decline in cognitive function and neuronal loss. Neuritic plaques and neurofibrillary tangles (NFTs) are considered to be pathological hallmarks of AD [26]. Recently, numerous studies have described the notable impact of GM on the pathogenesis of AD [27–32]. In particular, GM dysbiosis was shown to have negative effects on brain function and host behavior through MGBA dysregulation, suggesting its potential role in the development of AD [33,34]. The GM is thought to be involved in AD through alterations of at least five different pathogenic processes. These include amyloid-beta ($A\beta$) deposit, increased tau (a microtubule-associated protein) phosphorylation, neuroinflammation, metabolic dysfunction, and oxidative stress (Figure 1). Individuals with GM dysbiosis due to intestinal diseases were also shown to be at high risk for developing AD [35,36]. Additionally, the degree of alteration in the fecal microbiome has been correlated with the severity of AD [37–39]. Approximately 85% of patients with dementia were reported to have alteration in GM compositions as compared to populations of healthy individuals [40] and decreases in GM richness and diversity were observed in patients with AD [30]. Moreover, the levels of the specific bacterial species were found to be correlated with the cerebrospinal fluid (CSF) biomarkers of AD pathology. These data indicate that the alterations in GM can be linked to neuropathological changes in AD [30].

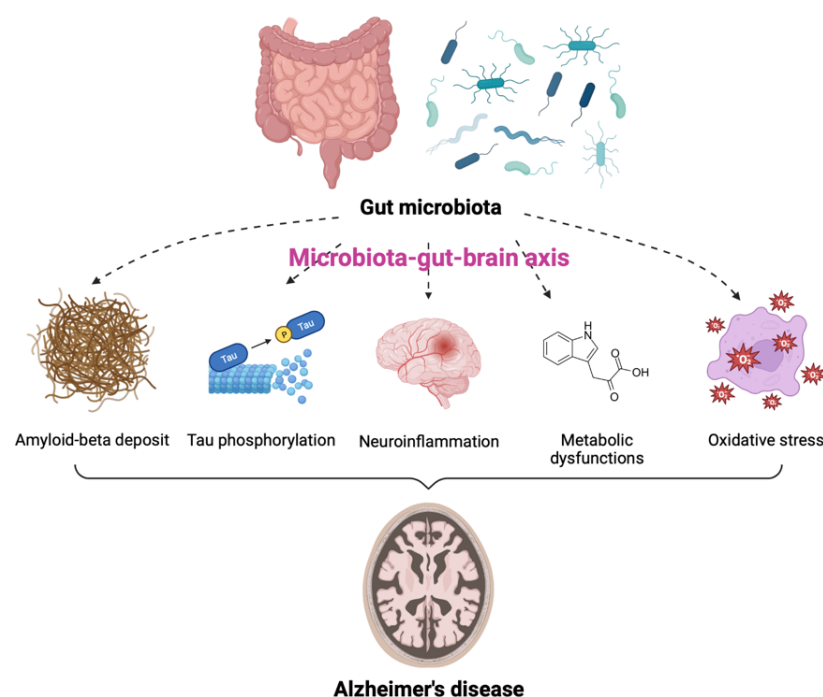


Figure 1. Contribution of gut microbiota (GM) to the pathogenesis of Alzheimer's disease (AD). GM is involved in the onset and progression of AD through alterations of at least five pathogenic processes, including amyloid-beta deposit, increased tau phosphorylation, neuroinflammation, metabolic dysfunctions, and oxidative stress via the microbiota-gut-brain axis.

Several GM species are associated with the production of amyloid fibers [41]. These amyloid fibers can cross the intestinal epithelium and blood-brain barrier (BBB) and induce several effects on the deposition of A β in the brain, resulting in AD pathogenesis [28,42]. Bacterial amyloids produced by GM promoted the misfolding of A β fibrils and oligomers [43]. In addition, due to the similarity in structure and immunogenicity, amyloids secreted by GM can bind to Toll-like receptor 2 (TLR2) on microglia and activate inflammatory responses [27]. These factors may enter the brain and directly affect AD pathology. Moreover, IL-17-expressing T helper cells migrate to the CNS via the gut-associated lymphoid tissue and interact with microglia, contributing to neurodegeneration in AD [27]. Furthermore, GM dysbiosis reduces the clearance of A β by affecting the gut mucosal barrier and energy homeostasis [44]. Collectively, GM increases the production of A β fibers, accelerates their aggregation and misfolding, and reduces A β clearance, all of which may contribute to the development and progression of AD.

Hyperphosphorylated tau and disintegrated microtubules are characteristics of NFTs, another pathological hallmark of AD [45]. A recent study found elevated levels of a GM-derived metabolite, trimethylamine N-oxide (TMAO), in the CSF of patients with AD dementia [46]. TMAO was involved in tau pathology, suggesting the influence of GM in AD pathogenesis through tau dysregulation [46]. Other researchers revealed GM-induced hyperphosphorylation of tau through the activation of the glycogen synthase kinase 3 beta (GSK-3 β) pathway, resulting in cognitive impairment [47]. Moreover, another study found alleviation of A β deposition, tau pathology, and memory impairment following fecal microbiota transplantation (FMT) from healthy wild-type mice into transgenic (Tg) mice with AD-like pathology, including amyloid and NFTs [48]. Although the comprehensive tau-mediated AD pathogenesis by GM remains elusive, these studies indicate that the modulation of GM may be a potential strategy to alleviate tauopathy for AD treatment.

Neuroinflammation is one of the plausible mechanisms to explain AD pathogenesis. This physiological response to stimuli maintains homeostasis, but excessive inflammatory responses cause harmful effects on the CNS. Changes in GM composition can trigger

peripheral immune responses by activating immune components and regulating the levels of pro-inflammatory cytokines in the brain [49–51]. Recent evidence suggests that GM dysbiosis is associated with the development of AD through neuroinflammation [52–54]. The increase in pro-inflammatory GM is accompanied by enhanced systemic inflammation and neuroinflammatory processes. Since GM dysbiosis can lead to defects in the activities of microglia, its activation can contribute to the progression of AD. Several gut microorganisms were reported to produce nitric oxide (NO) and activate microglia, thereby exacerbating the AD condition [55].

Metabolic dysfunction is also another hallmark of AD. As mentioned above, GM can produce bioactive metabolites, such as SCFAs and neurotransmitters, which can modulate the immune system and influence brain activities [56,57]. These metabolites can cross the BBB to affect cognition directly or indirectly through immune, neuroendocrine, or vagal mechanisms [58,59]. A meta-analysis of observational studies reported that increased levels of GABA were associated with a lower risk of AD [59]. There is a significant difference in the level of GM metabolites between AD patients and the control group [60]. Several metabolites upregulated by GM in patients with AD, including indole-3-pyruvic acid, an endogenous metabolite of tryptophan, and SCFAs, were correlated with cognitive impairment [60]. Interestingly, PD patients showed the opposite phenomenon, which is the reduction of SCFAs [61]. In addition, serotonin concentrations in the serum and urine of AD patients were significantly lower than in controls [62]. Most GM-associated metabolic pathways have been predicted based on sequencing analysis of the microbiome [63]. Therefore, validation is essential to prove the relationship between certain GM and AD pathogenesis.

Oxidative stress is another factor responsible for AD pathology. GM dysbiosis can involve in AD development by influencing the levels of oxidative stress in the CNS. For example, NO conversion from nitrate and nitrite by *Lactobacillus*, *E. coli*, and *Bifidobacterium* increases the permeability of the BBB and contributes to neurotoxicity in AD [64,65]. Increased intestinal permeability and following GM dysbiosis were also confirmed in the mouse model of HD [66]. Pathogenic enteric bacteria, such as *Salmonella* and *E. coli*, can induce hydrogen sulfide production in the gut, resulting in decreased mitochondrial oxygen consumption and overexpression of pro-inflammatory cytokines [67]. Hydrogen is a highly diffusible bioactive gas produced mainly by anaerobic cocci, which belong to the *Enterobacteriaceae* family [68]. GM dysbiosis may lead to reduced hydrogen production and limit gas supply to neurons in the CNS. Taken together, alterations in the GM favor oxidative stress, resulting in the pathogenesis of AD.

3. How to Study the Relationship between GM and AD

Overall, similar to other research, there are three different types of experimental approaches used to explore the relationship between GM and AD. These include in vitro, in vivo, and human studies (Figure 2). In this section, the concepts, strengths, and weaknesses of these three methods to study the role of the MGBA in the pathogenesis of AD are presented.

3.1. In Vitro Study

Although conceptually possible, cell-based studies have several limitations in studying the interconnection between GM and AD. The study of the MGBA has almost exclusively relied on the genomic or metagenomics analysis of samples collected from animal or human models. Up to now, there is a lack in in vitro models to simulate the communication between GM complex and intestinal epithelium or host-microbiome interactions [69,70]. To discover the disease status or intervention related to GM thoroughly, there is a need for longitudinal studies in animals and humans, which are difficult, expensive, and time-consuming. Therefore, this review will focus on in vitro gut fermentation models and their application for longitudinal studies to supplement in vivo microbiome studies.

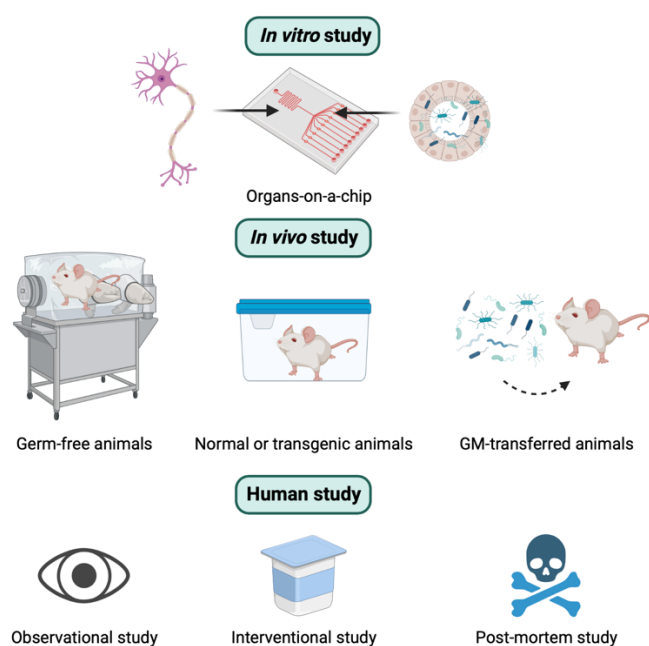


Figure 2. Three different types of experimental approaches to studying the role of the microbiota-gut-brain axis in the pathogenesis of Alzheimer’s disease. See the text below for the details.

To study the role of GM in the pathogenesis of AD, technology has proved a powerful tool to support in combination with cellular biology. Organ-on-a-chip (OOC) technology is a typical example of this combination [71–73]. This design successfully integrates the physiological features [70] in a model with bacterial molecular flux, microbial co-cultivation at the anoxic–oxic interface, a mucus bilayer with physiological thickness, or physical deformations for peristalsis-like motion [74,75]. However, most of today’s *in vitro* gut models are designed without the presence of tissue-resident immune cells [76]. The inclusion of the typical GM and human immune cells offers a promising gap to develop a reliable and predictive *in vitro* model to elucidate the relationship between GM and AD.

In the *in vitro* three-dimensional (3D) models used in AD studies by Sommer, brain cells were loaded in a suitable hydrogel matrix and cultured in a novel microfluidic device under perfusion with flow rates similar to the interstitial flow in brain tissue [70]. An innovative microfluidic device constitutes the building block of a multi-organ platform, and a hydrogel-based 3D model of brain cells can be housed in the microfluidic device to facilitate the expression of APP and accumulate amyloid, which is related to AD pathology. Based on this feature, the device can host suspended 2D and 3D cell-based models cultured alone or interconnected with other OOC units, to represent biological barriers, such as the BBB. Human neuroglioma H4-SW cells were chosen to test this model for their simplicity and capability to produce high levels of toxic A β . In particular, neural progenitor cells were cultured in a microfluidic chip under continuous flow conditions and a gradient of oligomeric assemblies of A β [77]. The main drawback of this study is the static conditions, although it provided evidence about the amyloid hypothesis in AD. Therefore, it is important to improve the dynamic conditions in *in vitro* studies for the simulation of AD pathogenesis.

Microfluidic OOCs *in vitro* modeling is a successful production of combining cellular biology and technology. This model provides an extraordinary opportunity to study the MGBA mechanisms in the development of neurodegenerative disorders, including AD. However, there is room left to improve more reliable and physiologically relevant OOCs. Currently, the most challenging aspects are building a dynamic environment and co-culture of multiple cell populations in multi-stage OOCs [78].

3.2. In Vivo Study

In vivo studies play a major role and provide a powerful tool for studying the underlying pathogenic mechanisms of GM-mediated AD. Various animal models, such as mice, rats, pigs, and zebrafish, have been used for GM research [79–82]. Furthermore, inbred and knockout techniques are regularly used for some animals like mice or rats [83,84]. There are different types of animal models depending on the origin of GM. They can be germ-free (GF) models (animals without microorganisms living in them), conventionally raised animals (animals colonized with their native GM), gnotobiotic animals (animals colonized with specific microbe(s), or human GM-transferred animals (animals colonized with human GM) [85]. These models are illustrated in Figure 2. The major advantage of animal models is the availability of pharmacological interventions that are not approved for humans and the access to organs and tissues after scarification [84]. The advantages and disadvantages of in vivo animal studies are summarized in Table 1.

Table 1. The advantages and disadvantages of animal models in studying the effects of gut microbiota on Alzheimer’s disease [86].

Advantages	Disadvantages
<ul style="list-style-type: none"> • Allowance of experiments that are not permissible in humans due to ethical reasons • Use of numerous genetically modified and knockout mouse models • Less burden on maintenance cost, high reproduction, and short life span • Homogenous genetic background and enhanced reproducibility of experiments • Control of sources of variations, minimizing unwanted environmental noises to GM 	<ul style="list-style-type: none"> • Fundamentally different physiology from humans in animal models • Alteration of GM composition in different animal models • No human-specific crosstalk between the GM and the host in animal models • No inherent genetic variations in animal models • Inability to recapitulate “real” human-GM relationship in animal models

Among different types of animal models, the GF model is the most popular one to study GM-host interactions. Almost human microbiota can be colonized in GF mice, while it is difficult to transfer predominant taxa in the gut of the human to ordinary mouse gut. In several cases, after colonization, the growth of microbiota may be modified and loss the characteristics of human donors [87,88]. Similarly, GM composition is likely to be altered by the disease states of GM dysbiosis after the colonization of GF mice, which may no longer reflect the composition in the disease state [89]. Several factors including diet, housing conditions, and sample collection procedures are also known or suspected to alter GM in animals [90]. Furthermore, origins or commercial suppliers of animal models are other factors to affect the variety of GM composition [83,91–93]. Despite the huge advantages and wide application of in vivo research, there are still other disadvantages of animal studies. For example, the reduction of animal experiments is necessary for ethical reasons. In addition, animal models are economically more burdensome than most in vitro models. On top of this, they need appropriate and special animal housing facilities, including GF facilities [84]. Moreover, longitudinal studies with animals are expensive and challenging since numerous animals need to be sacrificed at each time point [94,95]. Ultimately, there is a significant gap in the GM and immune systems between humans and animals, making it difficult to translate the results of in vivo research to human clinical trials [96].

The use of GF animals has been instrumental in understanding microbe-host relationships [97]. The first GF model in rodents was successfully generated in the 20th century [98]. Similar methods are now typically used to produce many generations of GF animals. Cesarean section is preferred to avoid inoculation of pups by microbiota [99,100]. Regular examinations of cages and feces are carried out to confirm the absence of bacteria [101]. Subsequent GF animals can be bred in an isolator, and GF pups can be born virginally. Alternatively, an embryonic transfer can be performed in GF animals into a GF host mother at the two-cell stage [101]. GF animals have shown completely different developmental

and physiological processes when compared with animals hosting commensal bacteria. GF animals are lighter and live longer. They have reduced levels of most GI luminal amino acids when compared with specific pathogen-free mice [100,102,103]. The lack of commensal microbes renders GF animals have an abnormal impact on immune systems, hormone signaling, metabolism, and neurotransmission [104–106]. Interestingly, phenotypes of GF animals are different across species, sex, research group, and even strain. This indicates the importance of microbiota and host genetics in determining several animal phenotypes [107,108].

Despite its many strengths, GF mice have several limitations. They include physiology, neurodevelopment, and immunity that are aberrant from human counterparts. All these factors limit translatability to clinical applications [86]. Nonetheless, GF mice have been used as the first of choice to investigate the involvement of microbiota in a given process [109,110]. Moreover, the results of GF studies begin to be applied to other non-rodent species such as pigs to maximize their translational value [109]. Alternatively, the transfer of mice with specific, known strains of bacteria (gnotobiotic animals) has also been utilized to investigate the specific microbiota-host relationship [111]. Among such methods, the altered Schaedler flora (ASF) mouse line has been the most widely used [112,113]. Eight bacterial colonization used in ASF mice simplified the study of microbiota involvement in brain diseases. A study with ASF mice was also able to produce more clinically relevant data than GF studies. When performing the experiments using GF mice, several host developmental defects including an underdeveloped immune system, slower intestinal epithelial turnover, differing nutritional requirements, and less body fat were frequently found. ASF mice turned out to be able to reduce these limitations [114]. Therefore, the ASF model is regarded as an attractive alternative to studying the effects of GM on stress-related brain disorders [112].

In the below paragraphs, illustrations and descriptions are mentioned about the utilization of animal models. To elucidate the impact of GM manipulation on AD pathology, a study used the 5xFAD model mice treated with antibiotics or probiotics [115]. While antibiotics treatment significantly reduced viable commensals, probiotics treatment transiently increased *Lactobacillaceae*. An analysis of A β deposition in the hippocampus confirmed the finding of ameliorated pathology. This study provides evidence that antibiotics might elicit a beneficial effect on AD pathology by the subsequent decrease in the A β influx. Another study also used the 5xFAD Tg mice model to understand the role of commensal gut bacteria on the progression of cognitive decline in AD [116]. The oral administration of *Bifidobacterium* brought alterations in the GI tract related to AD pathogenesis. These alterations included changes in GM composition, reduced fecal and blood LPS levels in feces and blood, suppressed nuclear factor kappa B (NF- κ B) activation, and tumor necrosis factor- α (TNF- α) expression in 5xFAD Tg and aged mice. These results suggest that gut dysbiosis and excessive endotoxin production can lead to endotoxemia and systemic inflammation, and CNS disorders. Moreover, the administration of *Bifidobacterium* was able to suppress the GI inflammation, resulting in the attenuation of cognitive decline in AD and aged mice through the regulation of neuroinflammation by the MGBA.

Various AD mouse models have been used to investigate the correlation between MGBA and AD. The D-galactose (D-Gal)/AlCl₃-induced AD mouse model was used to examine the effect of the water extract of *Gastrodia elata* rhizoma (WERG) on MGBA in AD treatment. WERG treatment enriched the gut probiotics and decreased the levels of phosphorylated tau, therefore, the cognitive impairment of D-Gal/AlCl₃-induced mice was improved in the WERG-treated group [117]. The regulation of GM by gastrodin (Gas) from *G. elata* for neuroprotection in AD was also determined using a D-Gal-induced AD model [118]. In that study, Gas was found to mitigate the memory dysfunction of AD mice. Interestingly, the antibiotic cocktail partially reversed the neuroprotective effect of Gas by changing the GM composition. In conclusion, Gas could improve the memory function of AD mice by partly targeting the MGBA and mitigating neuronal inflammation [118].

In addition to AD mice models, ordinary mice models are also used in the study of GM and AD. In research assessing the effect of GM depletion on anxiety- and depression-related behaviors, C57BL/6 mice were treated with an antibiotic cocktail for a long period of time from weaning to adulthood [119]. Results showed that there is a significant decrease in anxiety-like behaviors in the healthy antibiotic-treated group. Antibiotic treatment from early adolescence prevented the development of anxiety- and depression-related behaviors in AD-induced mice. In another study investigating the effect of *Lactobacillus* dominance by Korean red ginseng on the improvement of AD, Tg mice (Tg2576) were used for the experimental model of AD [120]. It was found that Korean red ginseng improved the cognitive behavior of mice and decreased the A β 42/A β 40, indicating reduced A β accumulation. In particular, the diversity of GM was altered, showing the increased population of *Lactobacillus* species.

Interventions, such as FMT, were introduced to in vivo studies to prove the effect of GM on the pathogenesis and treatment of AD. Research with an APPswe/PS1dE9 Tg mouse model was used to evaluate the efficacy of FMT for AD treatment [121]. FMT treatment improved cognitive deficits and reduced the brain deposition of A β in APPswe/PS1dE9 Tg mice. FMT treatment reversed the changes in the GM and SCFAs. In another study, FMT was proved sufficient to induce behavioral phenotypes in GF mice [122]. Kundu et al. used C57BL/6 mice to study FMT for AD treatment [122]. In their study, FMT transferred from 5xFAD mice to normal C57BL/6 mice decreased hippocampal neurogenesis and brain-derived neurotrophic factor expression, resulting in memory decline. This result can clarify the role of 5xFAD mouse-derived microbiota in AD development.

3.3. Human Study

GM can be studied in humans with a wide variety of individuals, including healthy volunteers, patients with a disease, patients with ileostomies, and individuals with sudden death [84]. The major advantage of human studies of GM is biological significance. Several antibiotics, including cefepime [123], amoxicillin [124], rifampicin [125], D-cycloserine [126], and doxycycline [127], reduced A β pathology and improved cognition in clinical studies. Furthermore, live and post-mortem studies using plasma [128] and brain samples [129–131] further indicated the greater LPS abundance in patients with AD compared with control individuals. However, it seems to be very hard to draw solid mechanistic conclusions based on these results because we cannot distinguish the direct or indirect effects of pharmacological interventions on the structure or functionality of the GM [95]. On top of this, additional challenges need to be overcome in human microbiome studies. For example, stringent ethical requirements for human study need to be satisfied beforehand. This may include modification of the research protocols and the prohibition of the usage of uncharacterized compounds [84]. In general, the time-series measurement methods used in human studies are costly and time-consuming. Extensive clinical data need to be provided to distinguish whether GM variability comes from pharmacological intervention or not. Moreover, due to higher percentages of withdrawal in long-term studies, researchers must consider patient numbers, which should be great enough to draw statistically meaningful conclusions [84]. Cohort variability due to host heterogeneity is another challenging factor to control in human studies [132,133]. Interindividual baseline variation of the GM with different responses to the same treatment is an additional confounding aspect to correlate the changes in the GM to the experimental intervention [134]. For example, the fecal microbiota transfer to different individuals exhibited different responses to the ciprofloxacin treatment [134,135]. Even the repeated administration of ciprofloxacin in the same patient also demonstrated different responses due to the composition change of the GM before and after drug treatment [134]. Several improvements to current research methodologies have been proposed to address these challenges associated with GM human studies. Stratification of study participants based on baseline GM may be helpful for better identification of GM alteration after drug administration. Diet has emerged as one of the most important factors responsible for normal variation in GM, as the relative

composition of GM is strongly influenced by nutrients [136]. However, the data from properly controlled dietary intervention studies are very hard to interpret. Walker et al. studied the fecal microbiota of 14 overweight men who consumed a precisely controlled diet for 10 weeks [137]. While there is a dramatic increase in certain bacteria after starting the diet, they found that fecal microbiota clustered more closely by individuals than by diet. Patient compliance and the accurate follow-up of food consumption by study participants are other important factors to determine the quality of these types of studies. The source of the samples needs to be diversified in data analysis since microbial communities found in feces are not representative of the whole GI tract [94,138,139]. If researchers are interested in the interaction between bacteria and the gut mucosa, the collection of intestinal biopsies may be more desirable.

Several studies have reported that GM composition is related to AD pathology, but the observed differences are inconsistent across studies. An observational study in the Netherlands investigated the associations between GM composition and AD biomarkers using machine learning models [140]. Results showed that GM composition was associated with amyloid and p-tau status, which were the two characteristics of AD. Another study, which was conducted in China, examined the structural and functional dysbiosis of GM in AD patients [38]. The data demonstrated a remarkable reduction in bacterial diversity and alterations in the taxonomic composition of the fecal microbiota of patients with AD. The study established the structural and functional dysbiosis of fecal microbiota in AD patients. The results further suggest the potential for the use of gut bacteria for early diagnosis of AD and personalized treatment for patients with AD.

To thoroughly understand the relationship between MGBA and AD, a study of the microbial-derived metabolite is another approach. TMAO, which is generated by choline metabolism, is a known risk factor for AD [46]. TMAO was found to be higher in the CSF of individuals with AD dementia than in the control group. In addition, elevated TMAO in the CSF was associated with p-tau and p-tau/A β 42, as well as neuronal degeneration. These findings provide additional insight into the involvement of GM in AD [46].

Several interventional human studies were conducted regarding the role of GM in AD pathogenesis and treatment, including case reports and clinical trials. A case report about cognitive function improvement after FMT in patients with AD dementia in 2021 provided further knowledge about this subject [141]. In that study, the cognitive function showed an improvement after FMT intervention based on the test score. The intervention also changed the GM composition in the recipient's feces. This finding suggests a relationship between GM and cognitive function in AD. Furthermore, it also suggests a novel therapeutic option, FMT, for patients with dementia [141]. Another case showed a rapid improvement in AD symptoms following FMT [142]. The patient reported improvements in mental acuity. These findings supported the central role of GM in neurological dysfunctions, such as AD [142].

A randomized multicenter trial was conducted to examine the effects of probiotics on cognition and mood in the elderly [143]. The results showed that the probiotics group had greater improvements in the mental flexibility test and stress score than the placebo group. Probiotics change the composition of GM, promote mental flexibility, and reduce stress in healthy older adults. These results support the hypothesis that probiotics provide health-promoting properties as a part of a healthy diet in older adults [143]. Another explorative intervention study aimed to examine the effect of probiotic supplementation in patients with AD dementia [144]. The results showed that supplementing patients with a wide variety of probiotics affected not only tryptophan metabolism in serum, but also GM composition [144].

Although there is an increase in advanced human studies of MGBA and AD, there is still a gap in the application of knowledge about MGBA in AD therapies. Further human research, especially interventional studies, should be conducted in the future to build a concrete conclusion about the role of GM in AD pathogenesis and provide helpful evidence for GM-related treatments in AD.

3.4. Relevant Techniques

Elucidation of GM composition is necessary to study the relationship between GM and AD [145]. The 16S rRNA gene sequencing has been the main technique in GM analysis for decades. This technique is applied in both in vivo studies and human studies [119,141,146,147]. Fecal samples are usually collected to investigate the GM profile, total DNA is then isolated, measured, and 16S rRNA gene sequencing is performed [148].

To determine the diversity and richness of GM, alpha diversity is usually represented by indexes such as Shannon, Chao1, and Simpson. Meanwhile, beta diversity is usually represented by Principal Coordinate Analysis (PCoA) for the differences in composition between groups [118]. While alpha diversity is a measure of microbiome diversity applicable to a single sample, beta diversity is a measure of the similarity or dissimilarity of two communities. Furthermore, LEfSe analysis is used to identify the specific individual bacterial taxa which cause the differences between groups. LEfSe analysis refers to the LDA (Linear discriminant analysis) Effect Size analysis. LEfSe is used to discover high-dimensional biomarkers and reveal genomic characteristics, including genes, metabolism, and classification [118,146].

Recently, high-throughput sequencing of the full 16S gene has been applied widely to investigate the relationship between GM and AD [149–151]. Low-throughput sequencing methods used the polymorphisms within the gene to distinguish strains or subspecies. Meanwhile, the complete full-length 16S gene is sequenced in a high-throughput method. The full 16S gene sequencing provides better taxonomic resolution, and real and significant advantages over sequencing commonly targeted variable regions [152].

4. GM-Directed Therapeutic Options to Ameliorate the Progression of AD

Due to the importance of maintenance of a healthy microbiota, several modulators of GM have been proposed such as microecological regulators, including prebiotics and probiotics, dietary intervention, and FMT. In this section, as depicted in Figure 3, the therapeutic options based on GM for the treatment of AD are discussed.

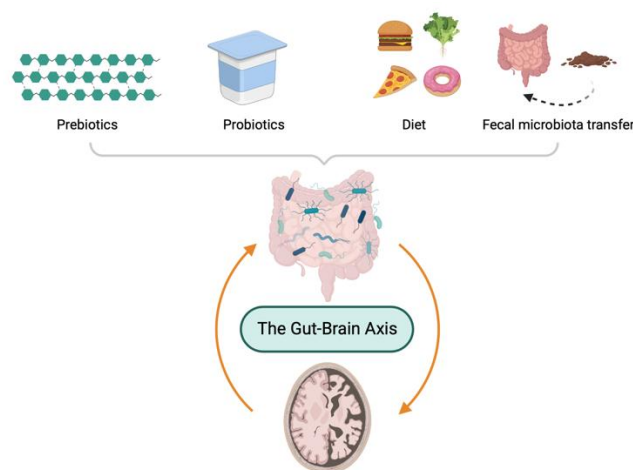


Figure 3. Gut microbiota-directed therapeutic options to ameliorate pathology of Alzheimer's disease.

4.1. Prebiotics

Prebiotics are defined as non-digestible short-chain carbohydrates that possess the ability to change the composition and metabolism of GM beneficially [153]. They can act as specific fermentation substrates for SCFA-producing probiotic genera, thus affecting both GI and extra-intestinal functions [154]. Recent studies in animal models and humans have shown possible effects on psychiatric symptoms [145]. Some promising results have been reported regarding the use of prebiotics for the prevention or treatment of AD [145,153]. The administration of yeast beta-glucan to mouse models of AD is effective in restoring the balance between pro- and anti-inflammatory GM species [146]. Lactulose, which was

the first commercially available prebiotic, was found to improve short-term memory and learning retrieval in AD mice [147]. In addition, mannan oligosaccharide treatment for 8 weeks induced the growth of *Lactobacillus* species and decreased *Helicobacter* abundance, resulting in reduced LPS leakage and BBB dysfunctions in 5xFAD Tg mice [63]. Interestingly, GM reconstitution by this prebiotic was also accompanied by decreased A β accumulation, restoration of redox homeostasis, and increased butyrate levels [63]. Similar results were seen in rodent models of AD treated with *Marinda officinalis*-derived oligosaccharides, with the effect of improved memory and learning ability [36,148]. Although the mechanism of action remains to be elucidated, prebiotic administration may be beneficial in the treatment of AD [148,149]. Moreover, a combination of probiotics and prebiotics (or synbiotics) seems to be more effective in increasing neurogenesis and reducing neuroinflammation as compared to prebiotics alone [149].

Daily administration of fructan, a well-known prebiotic, reduced the risk of AD development, as data from a large longitudinal study in older adults [150]. However, other authors suggest that more evidence for the use of prebiotics in clinical practice is still needed for concluding the normalization of several factors such as age, gender, ethnicity, and diet [151]. In conclusion, prebiotics may be helpful as a preventive or therapeutic therapy for AD, and there is a need for more human trials to concrete the importance of prebiotic treatment.

4.2. Probiotics

Probiotics are living microorganisms that are beneficial to the host with an adequate amount [152]. Probiotics have recently gained attention in brain function because they improve GM by positively influencing the MGBA. They are also known as psychobiotics because they attempt to restore the imbalances in the MGBA. They can release neuroactive substances and directly affect the human brain [155–157]. Many of these responses arise from the regulation of intracellular signaling pathways, such as mitogen-activated protein kinases (MAPKs) and NF- κ B [158]. Several studies have shown that probiotic supplementation can restore the GM, improve the integrity of the gut barrier and BBB, and reduce neuroinflammation, as well as cognitive decline [159]. Administration of probiotics increased *Actinobacteria* and *Bacteroides* species in the GM composition of AD animal model, significantly affecting long-term memory, inflammation, and neural plasticity [160]. Recently, a study on Tg AD mice demonstrated that the administration of a probiotic formulation significantly reduced oxidative stress by inducing sirtuin-1-dependent mechanisms [161]. In addition, the probiotics from a mixture of *Lactobacillus* and *Bifidobacterium* modified specific neurotransmitters, such as GABA and glutamate [162]. In another work, short-term administration of *Bifidobacterium breve* strain A1 suppressed immune response and neural inflammation in A β -injected mice [163]. Furthermore, a mixture of *Lactobacillus acidophilus*, *Lactobacillus fermentum*, *Bifidobacterium lactis*, and *Bifidobacterium longum* improved learning disability and oxidative stress of rats intra-hippocampally injected with A β 1–42 [164]. By using in vitro Caco-2 cell monolayer, di Vito et al. confirmed the modulation of tight and adherent junction and prevention of LPS-induced inflammatory damage by administration of commercially available probiotic formulation [165].

In a recent randomized trial, 60 patients with AD were divided into two groups and administered milk (control group) or probiotics (probiotic group). After 12 weeks of daily administration of probiotics, a significant improvement in the mini-mental state exam score was reported in the treated group, as compared to the control group [166]. Similarly, data from another meta-analysis reported a significant amelioration in cognition and a consistent reduction in post-intervention levels of malondialdehyde and high-sensitivity C-reactive protein in subjects receiving probiotics. These results indicate that probiotics, even when taken alone or in a combination supplement, have shown great potential in the reduction of AD progression. However, the appropriate strains, doses, time of treatment, routes of administration, and safe use of probiotics for AD need to be studied in the future.

4.3. Diet

Interactions between diet, GM, and host are important factors influencing health. Diet is one of the major factors involved in shaping GM composition [167,168]. Based on recent evidence, the Mediterranean diet (MD) and the ketogenic diet (KD) are likely to be the most promising dietary therapies for AD.

MD is a way of eating based on the traditional cuisine of countries bordering the Mediterranean Sea. This diet is characterized by a high intake of fruits, vegetables, cereals, and legumes; and a low intake of meat, high-fat dairy, and sweets. It is considered to be an anti-inflammatory diet and prevents the occurrence of several chronic diseases [169,170]. MD is also associated with a lower risk of AD [171]. Two large randomized controlled trials have demonstrated a positive correlation between “MD plus olive oil” or “MD plus nuts” with cognitive performance [172,173]. Recently, another clinical study linked MD with improved cognition [174]. Furthermore, a narrative systematic review and meta-analysis demonstrated a protective and likely therapeutic role of MD in AD and confirmed its ability to prevent cognitive impairment [175]. In general, a dietary pattern rich in fruits, vegetables, and legumes and low in saturated fats and sweets seems to provide protective effects [176]. Similarly, results highlighted the benefits of MD as a protective factor against AD [177]. Possible neuroprotective mechanisms common to these diets include the presence of antioxidants and anti-inflammatory compounds that help reduce inflammation and oxidative stress in the brain, and high levels of fiber, vitamin C, β -carotene, and folate. As a result, it improves brain integrity and increases the amount of brain tissue [178]. It has also been reported that saturated and trans fatty acid deficiency may reduce BBB dysfunction and amyloid aggregation [179,180].

KD is a term for a low-carbohydrate and adequate protein diet [181]. Recent studies have demonstrated a role for KD in the compositional remodeling of GM, thereby promoting its protective effects in various CNS disorders, including AD [182,183]. When sugar is in short supply, ketone bodies, which are used as alternative energy substrates for glucose in many organs, including the brain, are produced to break down and oxidize fat [184]. In mice models, ketone bodies have been demonstrated to influence neurotransmission, reduce neuroinflammation and oxidative stress, as well as reduce amyloid accumulation, and improve learning and memory abilities [185,186]. In humans, KD may benefit people with mild cognitive impairment or AD [183,187]. Similar to the mechanism (ketone body production), the medium-chain triglyceride diet/supplementation and modified Atkins diet are effective not only for symptoms such as fatigue and daytime sleepiness in Parkinson’s disease but also for cognitive decline in AD, epileptic seizures, and mood swings in depression [187,188]. Additionally, the modified Atkins diet, which does not restrict protein intake as the KD diet, allows much more flexibility than the classic KD. Overall, dietary patterns that lead to ketone production appear to represent a promising treatment for AD, although they reveal human protective mechanisms and adverse effects such as inflexibility and variability in dietary plans. To do so, more research is needed. It can easily lead to school dropouts and a lack of plant foods rich in vitamins and other antioxidant compounds [182].

In summary, dietary interventions are generally safer and more beneficial than drug therapy because they are inexpensive, easy to administer, and reduce the burden on caregivers of AD patients.

4.4. Fecal Microbiota Transplantation

FMT is a procedure that transfers the healthy donor’s GM to the recipient for therapeutic purposes. It is considered a safe procedure with minor and transient side effects [189]. It has been shown to be effective in the treatment of recurrent *Clostridium difficile* infections [190]. To date, most of the studies have been conducted in animal models, with promising results but a concrete conclusion has not been drawn yet.

Transplantation of feces from AD model donor mice into healthy mice resulted in impaired neurogenesis, increased memory impairment, increased circulating inflammatory

cytokines, and A β plaque deposition [191,192]. Similarly, GF mice that received feces from APP/PS1-Tg mice that developed brain A β deposition showed increased plaque formation [49,193]. Furthermore, FMT from AD patients to GF mice accelerated cognitive decline and the reduction of microbiota-derived metabolites important for nervous system function [193]. Another study confirmed improved cognition, decreased amyloid accumulation and tau expression, improved synaptic plasticity, and increased SCFA-producing gut bacteria [194]. FMT effectively restored the microbiota composition in the APP/PS1 Tg mouse model of AD, improving the conditions of microglia and A β deposition [194].

In terms of human studies, two case studies are showing promising results [141,142]. Hazan demonstrated an improvement in AD symptoms (including cognitive function, memory, and mood) in a man aged 82 after FMT from the recipient's wife [142]. In a second case study, a woman aged 90 with AD and severe *C. difficile* infection who underwent FMT from a healthy young donor showed improvements in cognitive function, GM composition, and SCFA production [141]. Interestingly, FMT also improved GM dysbiosis and cognitive deficits in the mouse model of HD [195].

These studies demonstrate that FMT can rapidly and effectively restore GM dysbiosis and brain dysfunction in patients, suggesting that restoration of GM homeostasis by FMT may have beneficial effects on AD treatment. However, several limitations remain for its wide application, such as standardization of the procedures, timepoint, and treatment period, as well as inclusion criteria of donor and recipient [196–198]. Therefore, more human trials will be conducted in the future to provide evidence for the efficacy of FMT and optimize the intervention.

5. Conclusions

The MGBA is an enticing target to understand the pathogenesis of AD, as well as to develop new therapeutic options to prevent and treat this disease. The MGBA can influence the development and progression of AD through various pathways, from A β deposition and tau phosphorylation to neuroinflammation, metabolic dysfunction, and oxidative stress. Numerous methodologies using in vitro cell models, animal models, and humans have been developed to gain insight into the relationship between GM and AD. Understanding of normal and pathogenic roles of the MGBA in host neurophysiology is critical for the development of mechanism-based prophylactic and/or therapeutic strategies for AD.

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References

1. Mayer, E.A.; Knight, R.; Mazmanian, S.K.; Cryan, J.F.; Tillisch, K. Gut Microbes and the Brain: Paradigm Shift in Neuroscience. *J. Neurosci.* **2014**, *34*, 15490–15496. [[CrossRef](#)] [[PubMed](#)]
2. Hugon, P.; Dufour, J.-C.; Colson, P.; Fournier, P.-E.; Sallah, K.; Raoult, D. A comprehensive repertoire of prokaryotic species identified in human beings. *Lancet Infect. Dis.* **2015**, *15*, 1211–1219. [[CrossRef](#)] [[PubMed](#)]

3. Cho, I.; Blaser, M.J. The human microbiome: At the interface of health and disease. *Nat. Rev. Genet.* **2012**, *13*, 260–270. [[CrossRef](#)] [[PubMed](#)]
4. Carabotti, M.; Scirocco, A.; Maselli, M.A.; Severi, C. The gut-brain axis: Interactions between enteric microbiota, central and enteric nervous systems. *Ann. Gastroenterol.* **2015**, *28*, 203–209.
5. Foster, J.A.; McVey Neufeld, K.-A. Gut-brain axis: How the microbiome influences anxiety and depression. *Trends Neurosci.* **2013**, *36*, 305–312. [[CrossRef](#)] [[PubMed](#)]
6. Galland, L. The Gut Microbiome and the Brain. *J. Med. Food* **2014**, *17*, 1261–1272. [[CrossRef](#)]
7. Stolfi, C.; Maresca, C.; Monteleone, G.; Laudisi, F. Implication of Intestinal Barrier Dysfunction in Gut Dysbiosis and Diseases. *Biomedicines* **2022**, *10*, 289. [[CrossRef](#)]
8. Lyte, M. Host-microbiota neuroendocrine interactions influencing brain and behavior. *Gut Microbes* **2014**, *5*, 381–389. [[CrossRef](#)]
9. Mayer, E.A.; Tillisch, K.; Gupta, A. Gut/brain axis and the microbiota. *J. Clin. Investig.* **2015**, *125*, 926–938. [[CrossRef](#)]
10. Borre, Y.E.; O’Keeffe, G.W.; Clarke, G.; Stanton, C.; Dinan, T.G.; Cryan, J.F. Microbiota and neurodevelopmental windows: Implications for brain disorders. *Trends Mol. Med.* **2014**, *20*, 509–518. [[CrossRef](#)]
11. Hsiao, E.Y.; McBride, S.W.; Hsien, S.; Sharon, G.; Hyde, E.R.; McCue, T.; Codelli, J.A.; Chow, J.; Reisman, S.E.; Petrosino, J.F.; et al. Microbiota Modulate Behavioral and Physiological Abnormalities Associated with Neurodevelopmental Disorders. *Cell* **2013**, *155*, 1451–1463. [[CrossRef](#)]
12. Cryan, J.F.; O’Riordan, K.J.; Sandhu, K.; Peterson, V.; Dinan, T.G. The gut microbiome in neurological disorders. *Lancet Neurol.* **2020**, *19*, 179–194. [[CrossRef](#)] [[PubMed](#)]
13. Tremlett, H.; Bauer, K.C.; Appel-Cresswell, S.; Finlay, B.B.; Waubant, E. The gut microbiome in human neurological disease: A review. *Ann. Neurol.* **2017**, *81*, 369–382. [[CrossRef](#)] [[PubMed](#)]
14. Heijtz, R.D.; Wang, S.; Anuar, F.; Qian, Y.; Björkholm, B.; Samuelsson, A.; Hibberd, M.L.; Forssberg, H.; Pettersson, S. Normal gut microbiota modulates brain development and behavior. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 3047–3052. [[CrossRef](#)] [[PubMed](#)]
15. Needham, B.D.; Kaddurah-Daouk, R.; Mazmanian, S.K. Gut microbial molecules in behavioural and neurodegenerative conditions. *Nat. Rev. Neurosci.* **2020**, *21*, 717–731. [[CrossRef](#)]
16. Gubert, C.; Kong, G.; Renoir, T.; Hannan, A.J. Exercise, diet and stress as modulators of gut microbiota: Implications for neurodegenerative diseases. *Neurobiol. Dis.* **2020**, *134*, 104621. [[CrossRef](#)]
17. Conte, C.; Sichert, M.; Traina, G. Gut–Brain Axis: Focus on Neurodegeneration and Mast Cells. *Appl. Sci.* **2020**, *10*, 1828. [[CrossRef](#)]
18. Quigley, E.M.M. Microbiota-Brain-Gut Axis and Neurodegenerative Diseases. *Curr. Neurol. Neurosci. Rep.* **2017**, *17*, 94. [[CrossRef](#)]
19. Scheperjans, F.; Aho, V.; Pereira, P.A.B.; Koskinen, K.; Paulin, L.; Pekkonen, E.; Haapaniemi, E.; Kaakkola, S.; Eerola-Rautio, J.; Pohja, M.; et al. Gut microbiota are related to Parkinson’s disease and clinical phenotype. *Mov. Disord.* **2015**, *30*, 350–358. [[CrossRef](#)]
20. Hill-Burns, E.M.; Debelius, J.W.; Morton, J.T.; Wissemann, W.T.; Lewis, M.R.; Wallen, Z.D.; Peddada, S.D.; Factor, S.A.; Molho, E.; Zabetian, C.P.; et al. Parkinson’s disease and Parkinson’s disease medications have distinct signatures of the gut microbiome. *Mov. Disord.* **2017**, *32*, 739–749. [[CrossRef](#)]
21. Felice, V.D.; Quigley, E.M.; Sullivan, A.M.; O’Keeffe, G.W.; O’Mahony, S.M. Microbiota-gut-brain signalling in Parkinson’s disease: Implications for non-motor symptoms. *Park. Relat. Disord.* **2016**, *27*, 1–8. [[CrossRef](#)] [[PubMed](#)]
22. Sampson, T.R.; Debelius, J.W.; Thron, T.; Janssen, S.; Shastri, G.G.; Ilhan, Z.E.; Challis, C.; Schretter, C.E.; Rocha, S.; Gradinaru, V.; et al. Gut Microbiota Regulate Motor Deficits and Neuroinflammation in a Model of Parkinson’s Disease. *Cell* **2016**, *167*, 1469–1480.e12. [[CrossRef](#)] [[PubMed](#)]
23. Kong, G.; Cao, K.-A.L.; Judd, L.M.; Li, S.; Renoir, T.; Hannan, A.J. Microbiome profiling reveals gut dysbiosis in a transgenic mouse model of Huntington’s disease. *Neurobiol. Dis.* **2020**, *135*, 104268. [[CrossRef](#)] [[PubMed](#)]
24. Wasser, C.I.; Mercieca, E.-C.; Kong, G.; Hannan, A.J.; McKeown, S.J.; Glikmann-Johnston, Y.; Stout, J.C. Gut dysbiosis in Huntington’s disease: Associations among gut microbiota, cognitive performance and clinical outcomes. *Brain Commun.* **2020**, *2*, fcaa110. [[CrossRef](#)]
25. Kong, G.; Ellul, S.; Narayana, V.K.; Kanojia, K.; Ha, H.T.T.; Li, S.; Renoir, T.; Cao, K.-A.L.; Hannan, A.J. An integrated metagenomics and metabolomics approach implicates the microbiota-gut-brain axis in the pathogenesis of Huntington’s disease. *Neurobiol. Dis.* **2021**, *148*, 105199. [[CrossRef](#)]
26. De-Paula, V.J.; Radanovic, M.; Diniz, B.S.; Forlenza, O.V. Alzheimer’s Disease. *Subcell. Biochem.* **2012**, *65*, 329–352. [[CrossRef](#)]
27. Janeiro, M.H.; Ramírez, M.J.; Solas, M. Dysbiosis and Alzheimer’s Disease: Cause or Treatment Opportunity? *Cell. Mol. Neurobiol.* **2022**, *42*, 377–387. [[CrossRef](#)]
28. Pistollato, F.; Sumalla Cano, S.; Elio, I.; Masias Vergara, M.; Giampieri, F.; Battino, M. Role of gut microbiota and nutrients in amyloid formation and pathogenesis of Alzheimer disease. *Nutr. Rev.* **2016**, *74*, 624–634. [[CrossRef](#)]
29. Shabbir, U.; Arshad, M.S.; Sameen, A.; Oh, D.-H. Crosstalk between Gut and Brain in Alzheimer’s Disease: The Role of Gut Microbiota Modulation Strategies. *Nutrients* **2021**, *13*, 690. [[CrossRef](#)]
30. Vogt, N.M.; Kerby, R.L.; Dill-McFarland, K.A.; Harding, S.J.; Merluzzi, A.P.; Johnson, S.C.; Carlsson, C.M.; Asthana, S.; Zetterberg, H.; Blennow, K.; et al. Gut microbiome alterations in Alzheimer’s disease. *Sci. Rep.* **2017**, *7*, 13537. [[CrossRef](#)]
31. Bostanciklioğlu, M. The role of gut microbiota in pathogenesis of Alzheimer’s disease. *J. Appl. Microbiol.* **2019**, *127*, 954–967. [[CrossRef](#)] [[PubMed](#)]

32. Jiang, C.; Li, G.; Huang, P.; Liu, Z.; Zhao, B. The Gut Microbiota and Alzheimer's Disease. *J. Alzheimer's Dis.* **2017**, *58*, 1–15. [[CrossRef](#)] [[PubMed](#)]
33. Guo, M.; Peng, J.; Huang, X.; Xiao, L.; Huang, F.; Zuo, Z. Gut Microbiome Features of Chinese Patients Newly Diagnosed with Alzheimer's Disease or Mild Cognitive Impairment. *J. Alzheimer's Dis.* **2021**, *80*, 299–310. [[CrossRef](#)] [[PubMed](#)]
34. Haran, J.P.; Bhattarai, S.K.; Foley, S.E.; Dutta, P.; Ward, D.V.; Bucci, V.; McCormick, B.A. Alzheimer's Disease Microbiome Is Associated with Dysregulation of the Anti-Inflammatory P-Glycoprotein Pathway. *Mbio* **2019**, *10*, e00632-19. [[CrossRef](#)] [[PubMed](#)]
35. Caini, S.; Bagnoli, S.; Palli, D.; Saieva, C.; Ceroti, M.; Bendinelli, B.; Assedi, M.; Masala, G. Total and cancer mortality in a cohort of ulcerative colitis and Crohn's disease patients: The Florence inflammatory bowel disease study, 1978–2010. *Dig. Liver Dis.* **2016**, *48*, 1162–1167. [[CrossRef](#)] [[PubMed](#)]
36. Chen, C.-H.; Lin, C.-L.; Kao, C.-H. Irritable Bowel Syndrome Is Associated with an Increased Risk of Dementia: A Nationwide Population-Based Study. *PLoS ONE* **2016**, *11*, e0144589. [[CrossRef](#)]
37. Ling, Z.; Zhu, M.; Liu, X.; Shao, L.; Cheng, Y.; Yan, X.; Jiang, R.; Wu, S. Fecal Fungal Dysbiosis in Chinese Patients with Alzheimer's Disease. *Front. Cell Dev. Biol.* **2020**, *8*, 631460. [[CrossRef](#)]
38. Ling, Z.; Zhu, M.; Yan, X.; Cheng, Y.; Shao, L.; Liu, X.; Jiang, R.; Wu, S. Structural and Functional Dysbiosis of Fecal Microbiota in Chinese Patients with Alzheimer's Disease. *Front. Cell Dev. Biol.* **2020**, *8*, 634069. [[CrossRef](#)]
39. Liu, P.; Wu, L.; Peng, G.; Han, Y.; Tang, R.; Ge, J.; Zhang, L.; Jia, L.; Yue, S.; Zhou, K.; et al. Altered microbiomes distinguish Alzheimer's disease from amnesic mild cognitive impairment and health in a Chinese cohort. *Brain Behav. Immun.* **2019**, *80*, 633–643. [[CrossRef](#)]
40. Morris, G.; Berk, M.; Carvalho, A.; Caso, J.R.; Sanz, Y.; Walder, K.; Maes, M. The Role of the Microbial Metabolites Including Tryptophan Catabolites and Short Chain Fatty Acids in the Pathophysiology of Immune-Inflammatory and Neuroimmune Disease. *Mol. Neurobiol.* **2017**, *54*, 4432–4451. [[CrossRef](#)]
41. Li, B.; He, Y.; Ma, J.; Huang, P.; Du, J.; Cao, L.; Wang, Y.; Xiao, Q.; Tang, H.; Chen, S. Mild cognitive impairment has similar alterations as Alzheimer's disease in gut microbiota. *Alzheimer's Dement.* **2019**, *15*, 1357–1366. [[CrossRef](#)] [[PubMed](#)]
42. Paranjapye, N.; Daggett, V. De Novo Designed α -Sheet Peptides Inhibit Functional Amyloid Formation of Streptococcus mutans Biofilms. *J. Mol. Biol.* **2018**, *430*, 3764–3773. [[CrossRef](#)] [[PubMed](#)]
43. Zhan, X.; Stamova, B.; Sharp, F.R. Lipopolysaccharide Associates with Amyloid Plaques, Neurons and Oligodendrocytes in Alzheimer's Disease Brain: A Review. *Front. Aging Neurosci.* **2018**, *10*, 42. [[CrossRef](#)] [[PubMed](#)]
44. Hrnčir, T.; Hrnčírova, L.; Kverka, M.; Tlaskalová-Hogenová, H. The role of gut microbiota in intestinal and liver diseases. *Lab. Anim.* **2019**, *53*, 271–280. [[CrossRef](#)]
45. Takashima, A. Tau aggregation is a therapeutic target for Alzheimer's disease. *Curr. Alzheimer Res.* **2010**, *7*, 665–669. [[CrossRef](#)]
46. Vogt, N.M.; Romano, K.A.; Darst, B.F.; Engelman, C.D.; Johnson, S.C.; Carlsson, C.M.; Asthana, S.; Blennow, K.; Zetterberg, H.; Bendlin, B.B.; et al. The gut microbiota-derived metabolite trimethylamine N-oxide is elevated in Alzheimer's disease. *Alzheimer's Res. Ther.* **2018**, *10*, 124. [[CrossRef](#)]
47. Wei, S.; Peng, W.; Mai, Y.; Li, K.; Wei, W.; Hu, L.; Zhu, S.; Zhou, H.; Jie, W.; Wei, Z.; et al. Outer membrane vesicles enhance tau phosphorylation and contribute to cognitive impairment. *J. Cell. Physiol.* **2020**, *235*, 4843–4855. [[CrossRef](#)]
48. Kim, M.-S.; Kim, Y.; Choi, H.; Kim, W.; Park, S.; Lee, D.; Kim, D.K.; Kim, H.J.; Choi, H.; Hyun, D.-W.; et al. Transfer of a healthy microbiota reduces amyloid and tau pathology in an Alzheimer's disease animal model. *Gut* **2020**, *69*, 283–294. [[CrossRef](#)]
49. Harach, T.; Marungruang, N.; Duthilleul, N.; Cheatham, V.; Mc Coy, K.D.; Frisoni, G.B.; Neher, J.J.; Fåk, F.; Jucker, M.; Lasser, T.; et al. Reduction of A β amyloid pathology in APPPS1 transgenic mice in the absence of gut microbiota. *Sci. Rep.* **2017**, *7*, 41802. [[CrossRef](#)]
50. Park, A.-M.; Omura, S.; Fujita, M.; Sato, F.; Tsunoda, I. *Helicobacter pylori* and gut microbiota in multiple sclerosis versus Alzheimer's disease: 10 pitfalls of microbiome studies. *Clin. Exp. Neuroimmunol.* **2017**, *8*, 215–232. [[CrossRef](#)]
51. Fung, T.C.; Olson, C.A.; Hsiao, E.Y. Interactions between the microbiota, immune and nervous systems in health and disease. *Nat. Neurosci.* **2017**, *20*, 145–155. [[CrossRef](#)] [[PubMed](#)]
52. Qureshi, I.A.; Mehler, M.F. Towards a 'systems'-level understanding of the nervous system and its disorders. *Trends Neurosci.* **2013**, *36*, 674–684. [[CrossRef](#)] [[PubMed](#)]
53. Sun, J.; Xu, J.; Yang, B.; Chen, K.; Kong, Y.; Fang, N.; Gong, T.; Wang, F.; Ling, Z.; Liu, J. Effect of *Clostridium butyricum* against Microglia-Mediated Neuroinflammation in Alzheimer's Disease via Regulating Gut Microbiota and Metabolites Butyrate. *Mol. Nutr. Food Res.* **2020**, *64*, e1900636. [[CrossRef](#)] [[PubMed](#)]
54. Pellegrini, C.; Antonioli, L.; Colucci, R.; Blandizzi, C.; Fornai, M. Interplay among gut microbiota, intestinal mucosal barrier and enteric neuro-immune system: A common path to neurodegenerative diseases? *Acta Neuropathol.* **2018**, *136*, 345–361. [[CrossRef](#)] [[PubMed](#)]
55. Sun, Y.-X.; Jiang, X.-J.; Lu, B.; Gao, Q.; Chen, Y.-F.; Wu, D.-B.; Zeng, W.-Y.; Yang, L.; Li, H.-H.; Yu, B. Roles of Gut Microbiota in Pathogenesis of Alzheimer's Disease and Therapeutic Effects of Chinese Medicine. *Chin. J. Integr. Med.* **2022**, *28*, 1048–1056. [[CrossRef](#)]
56. Rutsch, A.; Kantsjö, J.B.; Ronchi, F. The Gut-Brain Axis: How Microbiota and Host Inflammasome Influence Brain Physiology and Pathology. *Front. Immunol.* **2020**, *11*, 604179. [[CrossRef](#)]
57. McCarville, J.L.; Chen, G.Y.; Cuevas, V.D.; Troha, K.; Ayres, J.S. Microbiota Metabolites in Health and Disease. *Annu. Rev. Immunol.* **2020**, *38*, 147–170. [[CrossRef](#)]

58. Fülling, C.; Dinan, T.G.; Cryan, J.F. Gut Microbe to Brain Signaling: What Happens in Vagus . . . *Neuron* **2019**, *101*, 998–1002. [\[CrossRef\]](#)
59. Manyevitch, R.; Protas, M.; Scarpiello, S.; DeLiso, M.; Bass, B.; Nanajian, A.; Chang, M.; Thompson, S.M.; Khoury, N.; Gonnella, R.; et al. Evaluation of Metabolic and Synaptic Dysfunction Hypotheses of Alzheimer’s Disease (AD): A Meta-Analysis of CSF Markers. *Curr. Alzheimer Res.* **2018**, *15*, 164–181. [\[CrossRef\]](#)
60. Wu, L.; Han, Y.; Zheng, Z.; Peng, G.; Liu, P.; Yue, S.; Zhu, S.; Chen, J.; Lv, H.; Shao, L.; et al. Altered Gut Microbial Metabolites in Amnesic Mild Cognitive Impairment and Alzheimer’s Disease: Signals in Host–Microbe Interplay. *Nutrients* **2021**, *13*, 228. [\[CrossRef\]](#)
61. Unger, M.M.; Spiegel, J.; Dillmann, K.-U.; Grundmann, D.; Philippeit, H.; Bürmann, J.; Faßbender, K.; Schwiertz, A.; Schäfer, K.H. Short chain fatty acids and gut microbiota differ between patients with Parkinson’s disease and age-matched controls. *Parkinsonism Relat. Disord.* **2016**, *32*, 66–72. [\[CrossRef\]](#)
62. Whiley, L.; on behalf of AddNeuroMed consortium; Chappell, K.E.; D’Hondt, E.; Lewis, M.R.; Jiménez, B.; Snowden, S.G.; Soininen, H.; Kłoszewska, I.; Mecocci, P.; et al. Metabolic phenotyping reveals a reduction in the bioavailability of serotonin and kynurenine pathway metabolites in both the urine and serum of individuals living with Alzheimer’s disease. *Alzheimer’s Res. Ther.* **2021**, *13*, 20. [\[CrossRef\]](#)
63. Liu, Q.; Xi, Y.; Wang, Q.; Liu, J.; Li, P.; Meng, X.; Liu, K.; Chen, W.; Liu, X.; Liu, Z. Mannan oligosaccharide attenuates cognitive and behavioral disorders in the 5xFAD Alzheimer’s disease mouse model via regulating the gut microbiota-brain axis. *Brain Behav. Immun.* **2021**, *95*, 330–343. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Parker, A.; Fonseca, S.; Carding, S.R. Gut microbes and metabolites as modulators of blood-brain barrier integrity and brain health. *Gut Microbes* **2020**, *11*, 135–157. [\[CrossRef\]](#) [\[PubMed\]](#)
65. Tiso, M.; Schechter, A.N. Nitrate Reduction to Nitrite, Nitric Oxide and Ammonia by Gut Bacteria under Physiological Conditions. *PLoS ONE* **2015**, *10*, e0119712. [\[CrossRef\]](#)
66. Stan, T.L.; Soylu-Kucharz, R.; Burleigh, S.; Prykhodko, O.; Cao, L.; Franke, N.; Sjögren, M.; Haikal, C.; Hållénus, F.; Björkqvist, M. Increased intestinal permeability and gut dysbiosis in the R6/2 mouse model of Huntington’s disease. *Sci. Rep.* **2020**, *10*, 18270. [\[CrossRef\]](#)
67. Beaumont, M.; Andriamihaja, M.; Lan, A.; Khodorova, N.; Audebert, M.; Blouin, J.-M.; Grauso, M.; Lancha, L.; Benetti, P.-H.; Benamouzig, R.; et al. Detrimental effects for colonocytes of an increased exposure to luminal hydrogen sulfide: The adaptive response. *Free. Radic. Biol. Med.* **2016**, *93*, 155–164. [\[CrossRef\]](#) [\[PubMed\]](#)
68. Ohta, S. Molecular hydrogen as a preventive and therapeutic medical gas: Initiation, development and potential of hydrogen medicine. *Pharmacol. Ther.* **2014**, *144*, 1–11. [\[CrossRef\]](#)
69. Eain, M.M.G.; Baginska, J.; Greenhalgh, K.; Fritz, J.V.; Zenhausern, F.; Wilmes, P. Engineering Solutions for Representative Models of the Gastrointestinal Human-Microbe Interface. *Engineering* **2017**, *3*, 60–65. [\[CrossRef\]](#)
70. Sommer, M.O. Advancing gut microbiome research using cultivation. *Curr. Opin. Microbiol.* **2015**, *27*, 127–132. [\[CrossRef\]](#)
71. Ingber, D.E. Reverse Engineering Human Pathophysiology with Organs-on-Chips. *Cell* **2016**, *164*, 1105–1109. [\[CrossRef\]](#) [\[PubMed\]](#)
72. Wu, Q.; Liu, J.; Wang, X.; Feng, L.; Wu, J.; Zhu, X.; Wen, W.; Gong, X. Organ-on-a-chip: Recent breakthroughs and future prospects. *Biomed. Eng. Online* **2020**, *19*, 1–19. [\[CrossRef\]](#) [\[PubMed\]](#)
73. Yesil-Celiktas, O.; Hassan, S.; Miri, A.K.; Maharjan, S.; Al-Kharboosh, R.; Quiñones-Hinojosa, A.; Zhang, Y.S. Mimicking Human Pathophysiology in Organ-on-Chip Devices. *Adv. Biosyst.* **2018**, *2*, 1800109. [\[CrossRef\]](#)
74. Ceppa, F.A.; Izzo, L.; Sardelli, L.; Raimondi, I.; Tunesi, M.; Albani, D.; Giordano, C. Human Gut-Microbiota Interaction in Neurodegenerative Disorders and Current Engineered Tools for Its Modeling. *Front. Cell. Infect. Microbiol.* **2020**, *10*, 297. [\[CrossRef\]](#) [\[PubMed\]](#)
75. Kim, R.; Attayek, P.J.; Wang, Y.; Furtado, K.L.; Tamayo, R.; E Sims, C.; Allbritton, N.L. An *in vitro* intestinal platform with a self-sustaining oxygen gradient to study the human gut/microbiome interface. *Biofabrication* **2019**, *12*, 015006. [\[CrossRef\]](#) [\[PubMed\]](#)
76. Ambrosini, Y.M.; Shin, W.; Min, S.; Kim, H.J. Microphysiological Engineering of Immune Responses in Intestinal Inflammation. *Immune Netw.* **2020**, *20*, e13. [\[CrossRef\]](#)
77. Choi, Y.J.; Chae, S.; Kim, J.H.; Barald, K.F.; Park, J.Y.; Lee, S.-H. Neurotoxic amyloid beta oligomeric assemblies recreated in microfluidic platform with interstitial level of slow flow. *Sci. Rep.* **2013**, *3*, srep01921. [\[CrossRef\]](#)
78. Raimondi, M.T.; Albani, D.; Giordano, C. An Organ-On-A-Chip Engineered Platform to Study the Microbiota–Gut–Brain Axis in Neurodegeneration. *Trends Mol. Med.* **2019**, *25*, 737–740. [\[CrossRef\]](#)
79. Bry, L.; Falk, P.G.; Midtvedt, T.; Gordon, J.I. A Model of Host-Microbial Interactions in an Open Mammalian Ecosystem. *Science* **1996**, *273*, 1380–1383. [\[CrossRef\]](#)
80. Gerard, P.; Beguet, F.; Lepercq, P.; Rigottier-Gois, L.; Rochet, V.; Andrieux, C.; Juste, C. Gnotobiotic rats harboring human intestinal microbiota as a model for studying cholesterol-to-coprostanol conversion. *FEMS Microbiol. Ecol.* **2004**, *47*, 337–343. [\[CrossRef\]](#)
81. Meurens, F.; Berri, M.; Siggers, R.H.; Willing, B.P.; Salmon, H.; Van Kessel, A.G.; Gerdts, V. Commensal Bacteria and Expression of Two Major Intestinal Chemokines, TECK/CCL25 and MEC/CCL28, and Their Receptors. *PLoS ONE* **2007**, *2*, e677. [\[CrossRef\]](#) [\[PubMed\]](#)
82. Rawls, J.F.; Samuel, B.S.; Gordon, J.I. Gnotobiotic zebrafish reveal evolutionarily conserved responses to the gut microbiota. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 4596–4601. [\[CrossRef\]](#) [\[PubMed\]](#)

83. Hufeldt, M.R.; Nielsen, D.S.; Vogensen, F.K.; Midtvedt, T.; Hansen, A.K. Variation in the gut microbiota of laboratory mice is related to both genetic and environmental factors. *Comp. Med.* **2010**, *60*, 336–347. [[PubMed](#)]
84. Macfarlane, G.T.; Macfarlane, S. Models for intestinal fermentation: Association between food components, delivery systems, bioavailability and functional interactions in the gut. *Curr. Opin. Biotechnol.* **2007**, *18*, 156–162. [[CrossRef](#)] [[PubMed](#)]
85. Sekirov, I.; Russell, S.L.; Antunes, L.C.M.; Finlay, B.B.; Galla, S.; Chakraborty, S.; Cheng, X.; Yeo, J.; Mell, B.; Zhang, H.; et al. Gut Microbiota in Health and Disease. *Physiol. Rev.* **2010**, *90*, 859–904. [[CrossRef](#)]
86. Nguyen, T.L.A.; Vieira-Silva, S.; Liston, A.; Raes, J. How informative is the mouse for human gut microbiota research? *Dis. Model. Mech.* **2015**, *8*, 1–16. [[CrossRef](#)]
87. Chung, H.; Pamp, S.J.; Hill, J.A.; Surana, N.K.; Edelman, S.M.; Troy, E.B.; Reading, N.C.; Villablanca, E.J.; Wang, S.; Mora, J.R.; et al. Gut Immune Maturation Depends on Colonization with a Host-Specific Microbiota. *Cell* **2012**, *149*, 1578–1593. [[CrossRef](#)]
88. Turnbaugh, P.J.; Ridaura, V.K.; Faith, J.J.; Rey, F.E.; Knight, R.; Gordon, J.I. The Effect of Diet on the Human Gut Microbiome: A Metagenomic Analysis in Humanized Gnotobiotic Mice. *Sci. Transl. Med.* **2009**, *1*, 6ra14. [[CrossRef](#)]
89. Arrieta, M.-C.; Walter, J.; Finlay, B.B. Human Microbiota-Associated Mice: A Model with Challenges. *Cell Host Microbe* **2016**, *19*, 575–578. [[CrossRef](#)]
90. Ericsson, A.C.; Franklin, C.L. Manipulating the Gut Microbiota: Methods and Challenges: Figure. *ILAR J.* **2015**, *56*, 205–217. [[CrossRef](#)]
91. Denning, T.L.; Norris, B.A.; Medina-Contreras, O.; Manicassamy, S.; Geem, D.; Madan, R.; Karp, C.L.; Pulendran, B. Functional Specializations of Intestinal Dendritic Cell and Macrophage Subsets That Control Th17 and Regulatory T Cell Responses Are Dependent on the T Cell/APC Ratio, Source of Mouse Strain, and Regional Localization. *J. Immunol.* **2011**, *187*, 733–747. [[CrossRef](#)] [[PubMed](#)]
92. Ericsson, A.C.; Davis, J.W.; Spollen, W.; Bivens, N.; Givan, S.; Hagan, C.E.; McIntosh, M.; Franklin, C.L. Effects of Vendor and Genetic Background on the Composition of the Fecal Microbiota of Inbred Mice. *PLoS ONE* **2015**, *10*, e0116704. [[CrossRef](#)] [[PubMed](#)]
93. Ivanov, I.I.; de Llanos Frutos, R.; Manel, N.; Yoshinaga, K.; Rifkin, D.B.; Sartor, R.B.; Finlay, B.B.; Littman, D.R. Specific Microbiota Direct the Differentiation of IL-17-Producing T-Helper Cells in the Mucosa of the Small Intestine. *Cell Host Microbe* **2008**, *4*, 337–349. [[CrossRef](#)] [[PubMed](#)]
94. Macfarlane, G.; Macfarlane, S.; Gibson, G. Validation of a Three-Stage Compound Continuous Culture System for Investigating the Effect of Retention Time on the Ecology and Metabolism of Bacteria in the Human Colon. *Microb. Ecol.* **1998**, *35*, 180–187. [[CrossRef](#)] [[PubMed](#)]
95. Van den Abbeele, P.; Grootaert, C.; Marzorati, M.; Possemiers, S.; Verstraete, W.; Gérard, P.; Rabot, S.; Bruneau, A.; El Aidy, S.; Derrien, M.; et al. Microbial Community Development in a Dynamic Gut Model Is Reproducible, Colon Region Specific, and Selective for Bacteroidetes and Clostridium Cluster IX. *Appl. Environ. Microbiol.* **2010**, *76*, 5237–5246. [[CrossRef](#)]
96. Shultz, L.D.; Brehm, M.A.; Garcia-Martinez, J.V.; Greiner, D.L. Humanized mice for immune system investigation: Progress, promise and challenges. *Nat. Rev. Immunol.* **2012**, *12*, 786–798. [[CrossRef](#)]
97. Weger, B.D.; Gobet, C.; Yeung, J.; Martin, E.; Jimenez, S.; Betrisey, B.; Foata, F.; Berger, B.; Balvay, A.; Foussier, A.; et al. The Mouse Microbiome Is Required for Sex-Specific Diurnal Rhythms of Gene Expression and Metabolism. *Cell Metab.* **2019**, *29*, 362–382.e8. [[CrossRef](#)]
98. Gustafsson, B. Germ-free rearing of rats. *Acta Anat.* **1946**, *2*, 376–391. [[CrossRef](#)]
99. Al-Asmakh, M.; Zadjali, F. Use of Germ-Free Animal Models in Microbiota-Related Research. *J. Microbiol. Biotechnol.* **2015**, *25*, 1583–1588. [[CrossRef](#)]
100. Gordon, H.A.; Bruckner-Kardoss, E.; Wostmann, B.S. Aging in Germ-free Mice: Life Tables and Lesions Observed at Natural Death. *J. Gerontol.* **1966**, *21*, 380–387. [[CrossRef](#)]
101. Bibiloni, R. Rodent models to study the relationships between mammals and their bacterial inhabitants. *Gut Microbes* **2012**, *3*, 536–543. [[CrossRef](#)] [[PubMed](#)]
102. Aluwihare, A.P.R. An ultrastructural study of the effect of neomycin on the colon in the human subject and in the conventional and the germ-free mouse. *Gut* **1971**, *12*, 341–349. [[CrossRef](#)] [[PubMed](#)]
103. Yamamoto, Y.; Nakanishi, Y.; Murakami, S.; Aw, W.; Tsukimi, T.; Nozu, R.; Ueno, M.; Hioki, K.; Nakahigashi, K.; Hirayama, A.; et al. A Metabolomic-Based Evaluation of the Role of Commensal Microbiota throughout the Gastrointestinal Tract in Mice. *Microorganisms* **2018**, *6*, 101. [[CrossRef](#)] [[PubMed](#)]
104. Kawase, T.; Nagasawa, M.; Ikeda, H.; Yasuo, S.; Koga, Y.; Furuse, M. Gut microbiota of mice putatively modifies amino acid metabolism in the host brain. *Br. J. Nutr.* **2017**, *117*, 775–783. [[CrossRef](#)] [[PubMed](#)]
105. Pan, W.-H.; Sommer, F.; Falk-Paulsen, M.; Ulas, T.; Best, P.; Fazio, A.; Kachroo, P.; Luzius, A.; Jentzsch, M.; Rehman, A.; et al. Exposure to the gut microbiota drives distinct methylome and transcriptome changes in intestinal epithelial cells during postnatal development. *Genome Med.* **2018**, *10*, 27. [[CrossRef](#)]
106. Sudo, N.; Chida, Y.; Aiba, Y.; Sonoda, J.; Oyama, N.; Yu, X.-N.; Kubo, C.; Koga, Y. Postnatal microbial colonization programs the hypothalamic-pituitary-adrenal system for stress response in mice. *J. Physiol.* **2004**, *558*, 263–275. [[CrossRef](#)]
107. Arentsen, T.; Khalid, R.; Qian, Y.; Heijtz, R.D. Sex-dependent alterations in motor and anxiety-like behavior of aged bacterial peptidoglycan sensing molecule 2 knockout mice. *Brain Behav. Immun.* **2018**, *67*, 345–354. [[CrossRef](#)]

108. Nishino, R.; Mikami, K.; Takahashi, H.; Tomonaga, S.; Furuse, M.; Hiramoto, T.; Aiba, Y.; Koga, Y.; Sudo, N. Commensal microbiota modulate murine behaviors in a strictly contamination-free environment confirmed by culture-based methods. *Neurogastroenterol. Motil.* **2013**, *25*, 521–e371. [\[CrossRef\]](#)
109. Charbonneau, M.R.; O'Donnell, D.; Blanton, L.V.; Totten, S.M.; Davis, J.C.C.; Barratt, M.J.; Cheng, J.; Guruge, J.; Talcott, M.; Bain, J.R.; et al. Sialylated Milk Oligosaccharides Promote Microbiota-Dependent Growth in Models of Infant Undernutrition. *Cell* **2016**, *164*, 859–871. [\[CrossRef\]](#)
110. Luczynski, P.; McVey Neufeld, K.-A.; Oriach, C.S.; Clarke, G.; Dinan, T.G.; Cryan, J.F. Growing up in a Bubble: Using Germ-Free Animals to Assess the Influence of the Gut Microbiota on Brain and Behavior. *Int. J. Neuropsychopharmacol.* **2016**, *19*, pyw020. [\[CrossRef\]](#)
111. Gordon, H.A.; Pesti, L. The gnotobiotic animal as a tool in the study of host microbial relationships. *Bacteriol. Rev.* **1971**, *35*, 390–429. [\[CrossRef\]](#) [\[PubMed\]](#)
112. Lyte, J.; Proctor, A.; Phillips, G.; Lyte, M.; Wannemuehler, M. Altered Schaedler flora mice: A defined microbiota animal model to study the microbiota-gut-brain axis. *Behav. Brain Res.* **2019**, *356*, 221–226. [\[CrossRef\]](#) [\[PubMed\]](#)
113. Brand, M.W.; Wannemuehler, M.J.; Phillips, G.J.; Proctor, A.; Overstreet, A.-M.; Jergens, A.E.; Orcutt, R.P.; Fox, J.G. The Altered Schaedler Flora: Continued Applications of a Defined Murine Microbial Community. *ILAR J.* **2015**, *56*, 169–178. [\[CrossRef\]](#)
114. Cryan, J.F.; O'Riordan, K.J.; Cowan, C.S.M.; Sandhu, K.V.; Bastiaansen, T.F.S.; Boehme, M.; Codagnone, M.G.; Cussotto, S.; Fülling, C.; Golubeva, A.V.; et al. The Microbiota-Gut-Brain Axis. *Physiol. Rev.* **2019**, *99*, 1877–2013. [\[CrossRef\]](#)
115. Guilherme, M.; Nguyen, V.; Reinhardt, C.; Endres, K. Impact of Gut Microbiome Manipulation in 5xFAD Mice on Alzheimer's Disease-Like Pathology. *Microorganisms* **2021**, *9*, 815. [\[CrossRef\]](#) [\[PubMed\]](#)
116. Lee, H.-J.; Lee, K.-E.; Kim, J.-K.; Kim, D.-H. Suppression of gut dysbiosis by *Bifidobacterium longum* alleviates cognitive decline in 5XFAD transgenic and aged mice. *Sci. Rep.* **2019**, *9*, 11814. [\[CrossRef\]](#)
117. Zhao, W.; Wang, J.; Latta, M.; Wang, C.; Liu, Y.; Ma, W.; Zhou, Z.; Hu, S.; Chen, P.; Liu, Y. Rhizoma Gastrodiae Water Extract Modulates the Gut Microbiota and Pathological Changes of P-TauThr231 to Protect Against Cognitive Impairment in Mice. *Front. Pharmacol.* **2022**, *13*, 903659. [\[CrossRef\]](#)
118. Fasina, O.B.; Wang, J.; Mo, J.; Osada, H.; Ohno, H.; Pan, W.; Xiang, L.; Qi, J. Gastrodin From *Gastrodia elata* Enhances Cognitive Function and Neuroprotection of AD Mice via the Regulation of Gut Microbiota Composition and Inhibition of Neuron Inflammation. *Front. Pharmacol.* **2022**, *13*, 814271. [\[CrossRef\]](#) [\[PubMed\]](#)
119. Mosafari, B.; Jand, Y.; Salari, A.-A. Gut microbiota depletion from early adolescence alters anxiety and depression-related behaviours in male mice with Alzheimer-like disease. *Sci. Rep.* **2021**, *11*, 22941. [\[CrossRef\]](#)
120. Lee, M.; Lee, S.-H.; Kim, M.-S.; Ahn, K.-S.; Kim, M. Effect of *Lactobacillus* dominance modified by Korean Red Ginseng on the improvement of Alzheimer's disease in mice. *J. Ginseng Res.* **2020**, *46*, 464–472. [\[CrossRef\]](#)
121. Sun, J.; Xu, J.; Ling, Y.; Wang, F.; Gong, T.; Yang, C.; Ye, S.; Ye, K.; Wei, D.; Song, Z.; et al. Fecal microbiota transplantation alleviated Alzheimer's disease-like pathogenesis in APP/PS1 transgenic mice. *Transl. Psychiatry* **2019**, *9*, 189. [\[CrossRef\]](#) [\[PubMed\]](#)
122. Kundu, P.; Stagaman, K.; Kasschau, K.; Holden, S.; Shulzhenko, N.; Sharpton, T.J.; Raber, J. Fecal Implants From AppNL-G-F and AppNL-G-F/E4 Donor Mice Sufficient to Induce Behavioral Phenotypes in Germ-Free Mice. *Front. Behav. Neurosci.* **2022**, *16*, 791128. [\[CrossRef\]](#) [\[PubMed\]](#)
123. Payne, L.E.; Gagnon, D.J.; Riker, R.R.; Seder, D.B.; Glisic, E.K.; Morris, J.G.; Fraser, G.L. Cefepime-induced neurotoxicity: A systematic review. *Crit. Care* **2017**, *21*, 276. [\[CrossRef\]](#) [\[PubMed\]](#)
124. Kountouras, J.; Boziki, M.; Gavalas, E.; Zavos, C.; Grigoriadis, N.; Deretzi, G.; Tzilves, D.; Katsinelos, P.; Tsolaki, M.; Chatzopoulos, D.; et al. Eradication of *Helicobacter pylori* may be beneficial in the management of Alzheimer's disease. *J. Neurol.* **2009**, *256*, 758–767. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Yulug, B.; Hanoglu, L.; Ozansoy, M.; Isik, D.; Kilic, U.; Kilic, E.; Schabitz, W.R. Therapeutic role of rifampicin in Alzheimer's disease. *Psychiatry Clin. Neurosci.* **2018**, *72*, 152–159. [\[CrossRef\]](#)
126. Tsai, G.E.; Falk, W.E.; Gunther, J.; Coyle, J.T. Improved Cognition in Alzheimer's Disease With Short-Term D-Cycloserine Treatment. *Am. J. Psychiatry* **1999**, *156*, 467–469. [\[CrossRef\]](#)
127. Loeb, M.B.; Molloy, D.W.; Smieja, M.; Standish, T.; Goldsmith, C.H.; Mahony, J.; Smith, S.; Borrie, M.; Decoteau, E.; Davidson, W.; et al. A Randomized, Controlled Trial of Doxycycline and Rifampin for Patients with Alzheimer's Disease. *J. Am. Geriatr. Soc.* **2004**, *52*, 381–387. [\[CrossRef\]](#)
128. Zhang, R.; Miller, R.G.; Gascon, R.; Champion, S.; Katz, J.; Lancero, M.; Narvaez, A.; Honrada, R.; Ruvalcaba, D.; McGrath, M.S. Circulating endotoxin and systemic immune activation in sporadic amyotrophic lateral sclerosis (sALS). *J. Neuroimmunol.* **2009**, *206*, 121–124. [\[CrossRef\]](#)
129. Zhao, Y.; Cong, L.; Jaber, V.; Lukiw, W.J. Microbiome-Derived Lipopolysaccharide Enriched in the Perinuclear Region of Alzheimer's Disease Brain. *Front. Immunol.* **2017**, *8*, 1064. [\[CrossRef\]](#)
130. Zhao, Y.; Cong, L.; Lukiw, W.J. Lipopolysaccharide (LPS) Accumulates in Neocortical Neurons of Alzheimer's Disease (AD) Brain and Impairs Transcription in Human Neuronal-Glial Primary Co-cultures. *Front. Aging Neurosci.* **2017**, *9*, 407. [\[CrossRef\]](#)
131. Zhao, Y.; Jaber, V.; Lukiw, W.J. Secretory Products of the Human GI Tract Microbiome and Their Potential Impact on Alzheimer's Disease (AD): Detection of Lipopolysaccharide (LPS) in AD Hippocampus. *Front. Cell. Infect. Microbiol.* **2017**, *7*, 318. [\[CrossRef\]](#) [\[PubMed\]](#)

132. Dave, M.; Higgins, P.D.; Middha, S.; Rioux, K.P. The human gut microbiome: Current knowledge, challenges, and future directions. *Transl. Res.* **2012**, *160*, 246–257. [[CrossRef](#)] [[PubMed](#)]
133. Lozupone, C.A.; Stombaugh, J.I.; Gordon, J.I.; Jansson, J.K.; Knight, R. Diversity, stability and resilience of the human gut microbiota. *Nature* **2012**, *489*, 220–230. [[CrossRef](#)] [[PubMed](#)]
134. Dethlefsen, L.; Relman, D.A. Incomplete recovery and individualized responses of the human distal gut microbiota to repeated antibiotic perturbation. *Proc. Natl. Acad. Sci. USA* **2011**, *108* (Suppl. S1), 4554–4561. [[CrossRef](#)] [[PubMed](#)]
135. Dethlefsen, L.; Huse, S.; Sogin, M.L.; Relman, D.A. The Pervasive Effects of an Antibiotic on the Human Gut Microbiota, as Revealed by Deep 16S rRNA Sequencing. *PLoS Biol.* **2008**, *6*, e280. [[CrossRef](#)]
136. Spor, A.; Koren, O.; Ley, R. Unravelling the effects of the environment and host genotype on the gut microbiome. *Nat. Rev. Microbiol.* **2011**, *9*, 279–290. [[CrossRef](#)]
137. Walker, A.; Ince, J.; Duncan, S.H.; Webster, L.M.; Holtrop, G.; Ze, X.; Brown, D.; Stares, M.D.; Scott, P.; Bergerat, A.; et al. Dominant and diet-responsive groups of bacteria within the human colonic microbiota. *ISME J.* **2010**, *5*, 220–230. [[CrossRef](#)] [[PubMed](#)]
138. Macfarlane, G.; Gibson, G.; Cummings, J. Comparison of fermentation reactions in different regions of the human colon. *J. Appl. Bacteriol.* **1992**, *72*, 57–64. [[CrossRef](#)]
139. Salzman, N.H.; Bevins, C.L. Negative Interactions with the Microbiota: IBD. *Adv. Exp. Med. Biol.* **2008**, *635*, 67–78. [[CrossRef](#)]
140. Verhaar, B.J.H.; Hendriksen, H.M.A.; de Leeuw, F.A.; Doorduijn, A.S.; van Leeuwenstijn, M.; Teunissen, C.E.; Barkhof, F.; Scheltens, P.; Kraaij, R.; van Duijn, C.M.; et al. Gut Microbiota Composition Is Related to AD Pathology. *Front. Immunol.* **2021**, *12*, 794519. [[CrossRef](#)]
141. Park, S.-H.; Lee, J.H.; Shin, J.; Kim, J.-S.; Cha, B.; Lee, S.; Kwon, K.S.; Shin, Y.W.; Choi, S.H. Cognitive function improvement after fecal microbiota transplantation in Alzheimer’s dementia patient: A case report. *Curr. Med Res. Opin.* **2021**, *37*, 1739–1744. [[CrossRef](#)] [[PubMed](#)]
142. Hazan, S. Rapid improvement in Alzheimer’s disease symptoms following fecal microbiota transplantation: A case report. *J. Int. Med Res.* **2020**, *48*, 0300060520925930. [[CrossRef](#)] [[PubMed](#)]
143. Kim, C.-S.; Cha, L.; Sim, M.; Jung, S.; Chun, W.Y.; Baik, H.W.; Shin, D.-M. Probiotic Supplementation Improves Cognitive Function and Mood with Changes in Gut Microbiota in Community-Dwelling Older Adults: A Randomized, Double-Blind, Placebo-Controlled, Multicenter Trial. *J. Gerontol. Ser. A* **2021**, *76*, 32–40. [[CrossRef](#)] [[PubMed](#)]
144. Leblhuber, F.; Steiner, K.; Schuetz, B.; Fuchs, D.; Gostner, J.M. Probiotic Supplementation in Patients with Alzheimer’s Dementia—An Explorative Intervention Study. *Curr. Alzheimer Res.* **2018**, *15*, 1106–1113. [[CrossRef](#)]
145. Rajanala, K.; Kumar, N.; Chamallamudi, M.R. Modulation of Gut-Brain Axis by Probiotics: A Promising Anti-depressant Approach. *Curr. Neuropharmacol.* **2021**, *19*, 990–1006. [[CrossRef](#)]
146. Xu, M.; Mo, X.; Huang, H.; Chen, X.; Liu, H.; Peng, Z.; Chen, L.; Rong, S.; Yang, W.; Xu, S.; et al. Yeast β -glucan alleviates cognitive deficit by regulating gut microbiota and metabolites in A β 1–42-induced AD-like mice. *Int. J. Biol. Macromol.* **2020**, *161*, 258–270. [[CrossRef](#)]
147. Lee, Y.-S.; Lai, D.-M.; Huang, H.-J.; Lee-Chen, G.-J.; Chang, C.-H.; Hsieh-Li, H.M.; Lee, G.-C. Prebiotic Lactulose Ameliorates the Cognitive Deficit in Alzheimer’s Disease Mouse Model through Macroautophagy and Chaperone-Mediated Autophagy Pathways. *J. Agric. Food Chem.* **2021**, *69*, 2422–2437. [[CrossRef](#)]
148. Xin, Y.; Diling, C.; Jian, Y.; Ting, L.; Guoyan, H.; Hualun, L.; Xiaocui, T.; Guoxiao, L.; Ou, S.; Chaoqun, Z.; et al. Effects of Oligosaccharides From *Morinda officinalis* on Gut Microbiota and Metabolome of APP/PS1 Transgenic Mice. *Front. Neurol.* **2018**, *9*, 412. [[CrossRef](#)]
149. Deng, S.; Chen, C.; Lin, H.; Cheng, I.H. The beneficial effect of synbiotics consumption on Alzheimer’s disease mouse model via reducing local and systemic inflammation. *IUBMB Life* **2021**, *74*, 748–753. [[CrossRef](#)]
150. Gu, Y.; Nishikawa, M.; Brickman, A.M.; Manly, J.J.; Schupf, N.; Mayeux, R.P. Association of Dietary Prebiotic Consumption with Reduced Risk of Alzheimer’s Disease in a Multiethnic Population. *Curr. Alzheimer Res.* **2021**, *18*, 984–992. [[CrossRef](#)]
151. Barbosa, R.S.D.; Vieira-Coelho, M.A. Probiotics and prebiotics: Focus on psychiatric disorders—A systematic review. *Nutr. Rev.* **2020**, *78*, 437–450. [[CrossRef](#)] [[PubMed](#)]
152. Lilly, D.M.; Stillwell, R.H. Probiotics: Growth-Promoting Factors Produced by Microorganisms. *Science* **1965**, *147*, 747–748. [[CrossRef](#)] [[PubMed](#)]
153. Davani-Davari, D.; Negahdaripour, M.; Karimzadeh, I.; Seifan, M.; Mohkam, M.; Masoumi, S.; Berenjian, A.; Ghasemi, Y. Prebiotics: Definition, Types, Sources, Mechanisms, and Clinical Applications. *Foods* **2019**, *8*, 92. [[CrossRef](#)] [[PubMed](#)]
154. Den Besten, G.; van Eunen, K.; Groen, A.K.; Venema, K.; Reijngoud, D.-J.; Bakker, B.M. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *J. Lipid Res.* **2013**, *54*, 2325–2340. [[CrossRef](#)] [[PubMed](#)]
155. Collado, M.C.; Gueimonde, M.; Sanz, Y.; Salminen, S. Adhesion Properties and Competitive Pathogen Exclusion Ability of Bifidobacteria with Acquired Acid Resistance. *J. Food Prot.* **2006**, *69*, 1675–1679. [[CrossRef](#)]
156. Collado, M.C.; Hernández, M.; Sanz, Y. Production of Bacteriocin-Like Inhibitory Compounds by Human Fecal Bifidobacterium Strains. *J. Food Prot.* **2005**, *68*, 1034–1040. [[CrossRef](#)]
157. Martinez, F.A.C.; Balciunas, E.M.; Converti, A.; Cotter, P.D.; de Souza Oliveira, R.P. Bacteriocin production by *Bifidobacterium* spp. A review. *Biotechnol. Adv.* **2013**, *31*, 482–488. [[CrossRef](#)]
158. Bajaj, B.K.; Claes, I.J.; Lebeer, S. Functional mechanisms of probiotics. *J. Microbiol. Biotechnol. Food Sci.* **2021**, *2021*, 321–327.

159. Sichetti, M.; De Marco, S.; Pagiotti, R.; Traina, G.; Pietrella, D. Anti-inflammatory effect of multistrain probiotic formulation (*L. rhamnosus*, *B. lactis*, and *B. longum*). *Nutrition* **2018**, *53*, 95–102. [\[CrossRef\]](#)
160. Distrutti, E.; O'Reilly, J.-A.; McDonald, C.; Cipriani, S.; Renga, B.; Lynch, M.A.; Fiorucci, S. Modulation of Intestinal Microbiota by the Probiotic VSL#3 Resets Brain Gene Expression and Ameliorates the Age-Related Deficit in LTP. *PLoS ONE* **2014**, *9*, e106503. [\[CrossRef\]](#)
161. Bonfili, L.; Cecarini, V.; Cuccioloni, M.; Angeletti, M.; Berardi, S.; Scarpona, S.; Rossi, G.; Eleuteri, A.M. SLAB51 Probiotic Formulation Activates SIRT1 Pathway Promoting Antioxidant and Neuroprotective Effects in an AD Mouse Model. *Mol. Neurobiol.* **2018**, *55*, 7987–8000. [\[CrossRef\]](#)
162. O'Hagan, C.; Li, J.V.; Marchesi, J.R.; Plummer, S.; Garaiova, I.; Good, M.A. Long-term multi-species *Lactobacillus* and *Bifidobacterium* dietary supplement enhances memory and changes regional brain metabolites in middle-aged rats. *Neurobiol. Learn. Mem.* **2017**, *144*, 36–47. [\[CrossRef\]](#) [\[PubMed\]](#)
163. Kobayashi, Y.; Sugahara, H.; Shimada, K.; Mitsuyama, E.; Kuhara, T.; Yasuoka, A.; Kondo, T.; Abe, K.; Xiao, J.-Z. Therapeutic potential of *Bifidobacterium* breve strain A1 for preventing cognitive impairment in Alzheimer's disease. *Sci. Rep.* **2017**, *7*, 13510. [\[CrossRef\]](#) [\[PubMed\]](#)
164. Azm, S.A.N.; Djazayeri, A.; Safa, M.; Azami, K.; Ahmadvand, B.; Sabbaghziarani, F.; Sharifzadeh, M.; Vafa, M. Lactobacilli and bifidobacteria ameliorate memory and learning deficits and oxidative stress in β -amyloid (1–42) injected rats. *Appl. Physiol. Nutr. Metab.* **2018**, *43*, 718–726. [\[CrossRef\]](#)
165. di Vito, R.; Conte, C.; Traina, G. A Multi-Strain Probiotic Formulation Improves Intestinal Barrier Function by the Modulation of Tight and Adherent Junction Proteins. *Cells* **2022**, *11*, 2617. [\[CrossRef\]](#) [\[PubMed\]](#)
166. Akbari, E.; Asemi, Z.; Daneshvar Kakhaki, R.; Bahmani, F.; Kouchaki, E.; Tamtaji, O.R.; Hamidi, G.A.; Salami, M. Effect of probiotic supplementation on cognitive function and metabolic status in Alzheimer's disease: A randomized, double-blind and controlled trial. *Front. Aging Neurosci.* **2016**, *8*, 256. [\[CrossRef\]](#) [\[PubMed\]](#)
167. Klingbeil, E.A.; de La Serre, C.B. Microbiota modulation by eating patterns and diet composition: Impact on food intake. *Am. J. Physiol. Integr. Comp. Physiol.* **2018**, *315*, R1254–R1260. [\[CrossRef\]](#)
168. Merra, G.; Noce, A.; Marrone, G.; Cintoni, M.; Tarsitano, M.G.; Capacci, A.; De Lorenzo, A. Influence of Mediterranean Diet on Human Gut Microbiota. *Nutrients* **2021**, *13*, 7. [\[CrossRef\]](#) [\[PubMed\]](#)
169. Fung, T.T.; Rexrode, K.M.; Mantzoros, C.S.; Manson, J.E.; Willett, W.C.; Hu, F.B. Mediterranean Diet and Incidence of and Mortality From Coronary Heart Disease and Stroke in Women. *Circulation* **2009**, *119*, 1093–1100. [\[CrossRef\]](#)
170. Lopez-Garcia, E.; Rodriguez-Artalejo, F.; Li, T.Y.; Fung, T.T.; Li, S.; Willett, W.C.; Rimm, E.B.; Hu, F.B. The Mediterranean-style dietary pattern and mortality among men and women with cardiovascular disease. *Am. J. Clin. Nutr.* **2014**, *99*, 172–180. [\[CrossRef\]](#)
171. Sofi, F.; Macchi, C.; Abbate, R.; Gensini, G.F.; Casini, A. Mediterranean diet and health. *Biofactors* **2013**, *39*, 335–342. [\[CrossRef\]](#)
172. Martínez-Lapiscina, E.H.; Clavero, P.; Toledo, E.; Estruch, R.; Salas-Salvadó, J.; San Julián, B.; Sanchez-Tainta, A.; Ros, E.; Valls-Pedret, C.; Martínez-Gonzalez, M.Á. Mediterranean diet improves cognition: The PREDIMED-NAVARRA randomised trial. *J. Neurol. Neurosurg. Psychiatry* **2013**, *84*, 1318–1325. [\[CrossRef\]](#)
173. Valls-Pedret, C.; Sala-Vila, A.; Serra-Mir, M.; Corella, D.; De La Torre, R.; Martínez-González, M.Á.; Martínez-Lapiscina, E.H.; Fitó, M.; Pérez-Heras, A.; Salas-Salvadó, J.; et al. Mediterranean Diet and Age-Related Cognitive Decline: A Randomized Clinical Trial. *JAMA Intern. Med.* **2015**, *175*, 1094–1103. [\[CrossRef\]](#)
174. Marseglia, A.; Xu, W.; Fratiglioni, L.; Fabbri, C.; Berendsen, A.A.M.; Bialecka-Debek, A.; Jennings, A.; Gillings, R.; Meunier, N.; Caumon, E.; et al. Effect of the NU-AGE Diet on Cognitive Functioning in Older Adults: A Randomized Controlled Trial. *Front. Physiol.* **2018**, *9*, 349. [\[CrossRef\]](#) [\[PubMed\]](#)
175. Bartochowski, Z.; Conway, J.; Wallach, Y.; Chakkampambal, B.; Alakkassery, S.; Grossberg, G.T. Dietary Interventions to Prevent or Delay Alzheimer's Disease: What the Evidence Shows. *Curr. Nutr. Rep.* **2020**, *9*, 210–225. [\[CrossRef\]](#) [\[PubMed\]](#)
176. Liu, Y.H.; Gao, X.; Na, M.; Kris-Etherton, P.; Mitchell, D.C.; Jensen, G.L. Dietary pattern, diet quality, and dementia: A systematic review and meta-analysis of prospective cohort studies. *J. Alzheimers Dis.* **2020**, *78*, 151–158. [\[CrossRef\]](#) [\[PubMed\]](#)
177. Barbaresko, J.; Lellmann, A.W.; Schmidt, A.; Lehmann, A.; Amini, A.M.; Egert, S.; Schlesinger, S.; Nöthlings, U. Dietary Factors and Neurodegenerative Disorders: An Umbrella Review of Meta-Analyses of Prospective Studies. *Adv. Nutr. Int. Rev. J.* **2020**, *11*, 1161–1173. [\[CrossRef\]](#) [\[PubMed\]](#)
178. Prinelli, F.; Fratiglioni, L.; Kalpouzos, G.; Musicco, M.; Adorni, F.; Johansson, I.; Marseglia, A.; Xu, W. Specific nutrient patterns are associated with higher structural brain integrity in dementia-free older adults. *Neuroimage* **2019**, *199*, 281–288. [\[CrossRef\]](#) [\[PubMed\]](#)
179. Kao, Y.-C.; Ho, P.-C.; Tu, Y.-K.; Jou, I.-M.; Tsai, K.-J. Lipids and Alzheimer's Disease. *Int. J. Mol. Sci.* **2020**, *21*, 1505. [\[CrossRef\]](#) [\[PubMed\]](#)
180. Rainey-Smith, S.R.; Gu, Y.; Gardener, S.L.; Doecke, J.D.; Villemagne, V.L.; Brown, B.M.; Taddei, K.; Laws, S.M.; Sohrabi, H.R.; Weinborn, M.; et al. Mediterranean diet adherence and rate of cerebral A β -amyloid accumulation: Data from the Australian Imaging, Biomarkers and Lifestyle Study of Ageing. *Transl. Psychiatry* **2018**, *8*, 238. [\[CrossRef\]](#) [\[PubMed\]](#)
181. Ułamek-Kozioł, M.; Czuczwar, S.J.; Januszewski, S.; Pluta, R. Ketogenic Diet and Epilepsy. *Nutrients* **2019**, *11*, 2510. [\[CrossRef\]](#) [\[PubMed\]](#)
182. Bostock, E.C.S.; Kirkby, K.C.; Taylor, B.V.M. The Current Status of the Ketogenic Diet in Psychiatry. *Front. Psychiatry* **2017**, *8*, 43. [\[CrossRef\]](#) [\[PubMed\]](#)

183. Rusek, M.; Pluta, R.; Ułamek-Kozioł, M.; Czuczwar, S.J. Ketogenic Diet in Alzheimer's Disease. *Int. J. Mol. Sci.* **2019**, *20*, 3892. [[CrossRef](#)] [[PubMed](#)]
184. Masino, S.A.; Rho, J.M. Mechanisms of ketogenic diet action. *Epilepsia* **2010**, *51*, 85. [[CrossRef](#)]
185. Kashiwaya, Y.; Bergman, C.; Lee, J.-H.; Wan, R.; King, M.T.; Mughal, M.R.; Okun, E.; Clarke, K.; Mattson, M.P.; Veech, R.L. A ketone ester diet exhibits anxiolytic and cognition-sparing properties, and lessens amyloid and tau pathologies in a mouse model of Alzheimer's disease. *Neurobiol. Aging* **2013**, *34*, 1530–1539. [[CrossRef](#)]
186. Rojas-Morales, P.; Pedraza-Chaverri, J.; Tapia, E. Ketone bodies, stress response, and redox homeostasis. *Redox Biol.* **2020**, *29*, 101395. [[CrossRef](#)]
187. Taylor, M.K.; Sullivan, D.K.; Mahnken, J.D.; Burns, J.; Swerdlow, R.H. Feasibility and efficacy data from a ketogenic diet intervention in Alzheimer's disease. *Alzheimer's Dementia Transl. Res. Clin. Interv.* **2018**, *4*, 28–36. [[CrossRef](#)]
188. Włodarek, D. The possibility of use of the ketogenic diet and medium chain triglycerides supplementation in the support therapy of Alzheimer disease. *Curr. Opin. Clin. Nutr. Metab. Care* **2021**, *24*, 385–391. [[CrossRef](#)]
189. Gupta, S.; Allen-Vercoe, E.; Petrof, E.O. Fecal microbiota transplantation: In perspective. *Ther. Adv. Gastroenterol.* **2016**, *9*, 229–239. [[CrossRef](#)]
190. Surawicz, C.M.; Brandt, L.J.; Binion, D.G.; Ananthakrishnan, A.N.; Curry, S.R.; Gilligan, P.H.; McFarland, L.V.; Mellow, M.; Zuckerbraun, B.S. Guidelines for diagnosis, treatment, and prevention of *Clostridium difficile* infections. *Am. J. Gastroenterol.* **2013**, *108*, 478–498. [[CrossRef](#)]
191. Kim, N.; Jeon, S.H.; Ju, I.G.; Gee, M.S.; Do, J.; Oh, M.S.; Lee, J.K. Transplantation of gut microbiota derived from Alzheimer's disease mouse model impairs memory function and neurogenesis in C57BL/6 mice. *Brain Behav. Immun.* **2021**, *98*, 357–365. [[CrossRef](#)]
192. Wang, M.; Cao, J.; Gong, C.; Amakye, W.K.; Yao, M.; Ren, J. Exploring the microbiota-Alzheimer's disease linkage using short-term antibiotic treatment followed by fecal microbiota transplantation. *Brain Behav. Immun.* **2021**, *96*, 227–238. [[CrossRef](#)] [[PubMed](#)]
193. Fujii, Y.; Nguyen, T.T.T.; Fujimura, Y.; Kameya, N.; Nakamura, S.; Arakawa, K.; Morita, H. Fecal metabolite of a gnotobiotic mouse transplanted with gut microbiota from a patient with Alzheimer's disease. *Biosci. Biotechnol. Biochem.* **2019**, *83*, 2144–2152. [[CrossRef](#)] [[PubMed](#)]
194. Dodiya, H.B.; Kuntz, T.; Shaik, S.M.; Baufeld, C.; Leibowitz, J.; Zhang, X.; Gottel, N.; Zhang, X.; Butovsky, O.; Gilbert, J.A.; et al. Sex-specific effects of microbiome perturbations on cerebral A β amyloidosis and microglia phenotypes. *J. Exp. Med.* **2019**, *216*, 1542–1560. [[CrossRef](#)]
195. Gubert, C.; Choo, J.M.; Love, C.J.; Kodikara, S.; Masson, B.A.; Liew, J.J.M.; Wang, Y.; Kong, G.; Narayana, V.K.; Renoir, T.; et al. Faecal microbiota transplant ameliorates gut dysbiosis and cognitive deficits in Huntington's disease mice. *Brain Commun.* **2022**, *4*, fcac205. [[CrossRef](#)]
196. Choi, H.H.; Cho, Y.-S. Fecal Microbiota Transplantation: Current Applications, Effectiveness, and Future Perspectives. *Clin. Endosc.* **2016**, *49*, 257–265. [[CrossRef](#)] [[PubMed](#)]
197. Schmulson, M.; Bashashati, M. Fecal microbiota transfer for bowel disorders: Efficacy or hype? *Curr. Opin. Pharmacol.* **2018**, *43*, 72–80. [[CrossRef](#)] [[PubMed](#)]
198. Tan, P.; Li, X.; Shen, J.; Feng, Q. Fecal Microbiota Transplantation for the Treatment of Inflammatory Bowel Disease: An Update. *Front. Pharmacol.* **2020**, *11*, 574533. [[CrossRef](#)]

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