

## A novel lipase from *Streptomyces exfoliatus* DSMZ 41693 for biotechnological applications

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**Table S1.** Strains, plasmids, and oligonucleotides used in this study.

Strain	Relevant description or sequence	Reference
<i>Streptomyces exfoliatus</i> DSMZ 41693	Producer of lipases	[26]
<i>Escherichia coli</i> DH5 $\alpha$	Host for recombinant plasmids F <sup>-</sup> $\phi$ 80 <i>lacZ</i> $\Delta$ M15 $\Delta$ ( <i>lacZ</i> YA- <i>argF</i> )U169 <i>recA1</i> <i>endA1 hsdR17</i> (r <sub>K</sub> <sup>-</sup> , m <sub>K</sub> <sup>+</sup> ) <i>phoA supE44</i> $\lambda$ <sup>-</sup> <i>thi-1</i> <i>gyrA96 relA1</i>	[101]
<i>Rhodococcus sp.</i> T104 KACC 21099	Host for gene expression (Kn <sup>s</sup> )	[102]
<i>Rhodococcus sp.</i> T104 KACC 21099 pENV19SeLipA	<i>Rhodococcus sp.</i> T104 KACC 21099 recombinant strain harboring the plasmid pENV19SeLipA	This study
<i>Rhodococcus sp.</i> T104 KACC 21099 pENV19SeLipB	<i>Rhodococcus sp.</i> T104 KACC 21099 recombinant strain harboring the plasmid pENV19SeLipB	This study
<i>Rhodococcus sp.</i> T104 KACC 21099 pENV19SeLipC	<i>Rhodococcus sp.</i> T104 KACC 21099 recombinant strain harboring the plasmid pENV19SeLipC	This study
<i>Rhodococcus sp.</i> T104 KACC 21099 pENV19SeLipD	<i>Rhodococcus sp.</i> T104 KACC 21099 recombinant strain harboring the plasmid pENV19SeLipD	This study
<b>Plasmids</b>		
pENV19	Shuttle vector for <i>E. coli</i> - <i>Rhodococcus</i> with the constitutive <i>perme</i> * promoter (5.1 Kb, Kn <sup>R</sup> pAL5000ori <i>perme</i> * ColE1ori)	[35,37,38]
pENV19SeLipA	pENV19 containing <i>lipA</i> gene from <i>S. exfoliatus</i> DSMZ 41693	This study
pENV19SeLipB	pENV19 containing <i>lipB</i> gene from <i>S. exfoliatus</i> DSMZ 41693	This study
pENV19SeLipC	pENV19 containing <i>lipC</i> gene from <i>S. exfoliatus</i> DSMZ 41693	This study
pENV19SeLipD	pENV19 containing <i>lipPs</i> gene from <i>S. exfoliatus</i> DSMZ 41693	This study
<b>Primers</b>		
Sequence (5'→3') <sup>#</sup>		
SeLipAFWsp	GCTCTAGAGAGGACGACTCCGTCATGAAACGACAC	This study
SeLipAREV	<b>GGAATTC</b> <u>TCAGGAACCGAGCGGGCAGGTG</u>	This study
SeLipBFWsp	GCTCTAGAGGAGGTTCCATGAGACTGTCC	This study
SeLipBREV	<b>GGAATTC</b> <u>CTACGCCGCCCGTTG</u>	This study
SeLipCFWsp	GCTCTAGACAGTGTGGCTGGAACGACTGGTCCTGCAAGCCC	This study
SeLipCREV	<b>GGAATTC</b> <u>CCTAGCCGATCACCGAGAGGCAGGTGGTCGGGGTTCG</u>	This study
SeLipDFW	GCTCTAGAAGGAGTTCCTATGTCAGGGAG	This study
SeLipDREV	<b>GGAATTC</b> <u>CTCAGGCCGTGGACCGGGAATCTG</u>	This study

<sup>#</sup>The restriction sites *Xba*I and *Eco*RI are shown in bold, ribosomal binding site sequence is shown in italics, and the start and the stop codons are shown underlined.

**Table S2.** PCR conditions for amplification of lipase genes from *Streptomyces exfoliatus* DSMZ 41693.

Step	Temperature (°C)	Time
1. Denaturalization	95	3 minutes
2. Denaturalization	95	30 seconds
3. Annealing	83	30 seconds
4. Extension	72	55 seconds
5. Repeat 17 cycles steps 2-4 decreasing 1 °C annealing temperature by cycle		
6. Denaturalization	95	30 seconds
7. Annealing	58-72	30 seconds
8. Extension	72	55 seconds
9. Repeat 20 cycles steps 6-8 increasing 1 s annealing time by cycle		
10. Extension	72	10 minutes

**Table S3.** pNP-ester hydrolytic specific activity in the culture broth of the recombinant strains.

pNP-esters (number of carbon atoms in acyl chain)	Specific activity (IU/mg protein)			
	SeLipA	SeLipB	SeLipC	SeLipD
pNP-acetate (C2)	n.d.	n.d.	n.d.	n.d.
pNP-butyrate (C4)	0.9	n.d.	0.9	n.d.
pNP-valerate (C5)	2.5	n.d.	2.5	n.d.
pNP-hexanoate (C6)	3.1	n.d.	6.5	n.d.
pNP-octanoate (C8)	4.8	n.d.	13.3	n.d.
pNP-decanoate (C10)	8.0	n.d.	17.2	n.d.
pNP-laurate (C12)	7.1	n.d.	12.8	n.d.
pNP-myristate (C14)	4.8	n.d.	9.5	n.d.
pNP-palmitate (C16)	5.3	n.d.	12	n.d.
pNP-stearate (C18)	4.6	n.d.	7.8	n.d.

n.d.: not detected

**Table S4.** Purification of SeLipC produced by *Rhodococcus* pENV19SeLipC by hydrophobic interaction chromatography (HIC).

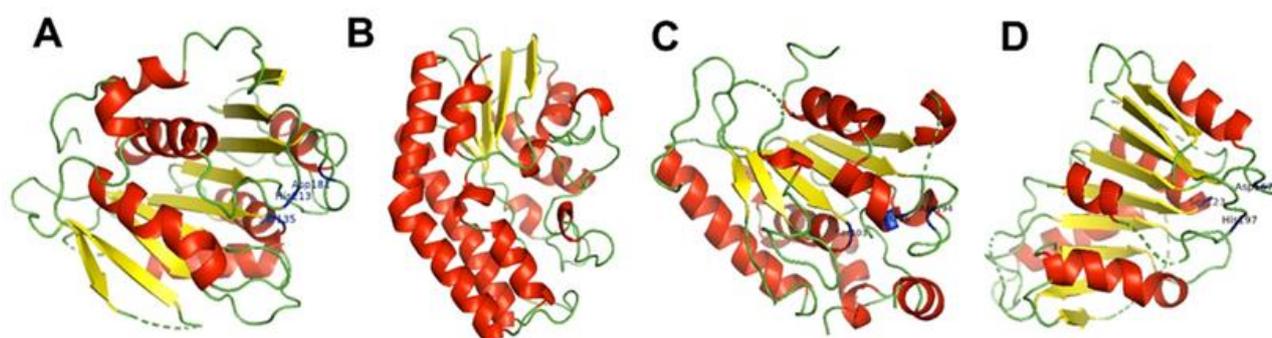
Step	V (mL)	Activity (IU)	Protein concentration (mg/mL)	Total protein (mg)	Specific activity (IU/mg)	Purification factor (fold)	Yield (%)
Broth	700	1356	0.163	114.1	12	1	100
Broth (NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	740	1202	0.151	111.7	11	0.9	89
HIC	92	245.7	0.023	2.12	116.6	3.6	17.2
Dialysis	140	240.8	0.007	1.04	232.4	7.2	16.9

**Table S5.** Effect of different reducing agents on SeLipC activity.

Chemical reagent	Concentration	Residual activity (%)
No addition	-	100.0 ± 7.2
Dithiothreitol (DTT)	1 mM	94.9 ± 6.0
	10 mM	101.6 ± 5.7
2-Mercaptoethanol	1 mM	99.8 ± 1.9
	10 mM	99.4 ± 2.9

**Table S6.** Effect of chemical modification with PMSF on SeLipC.

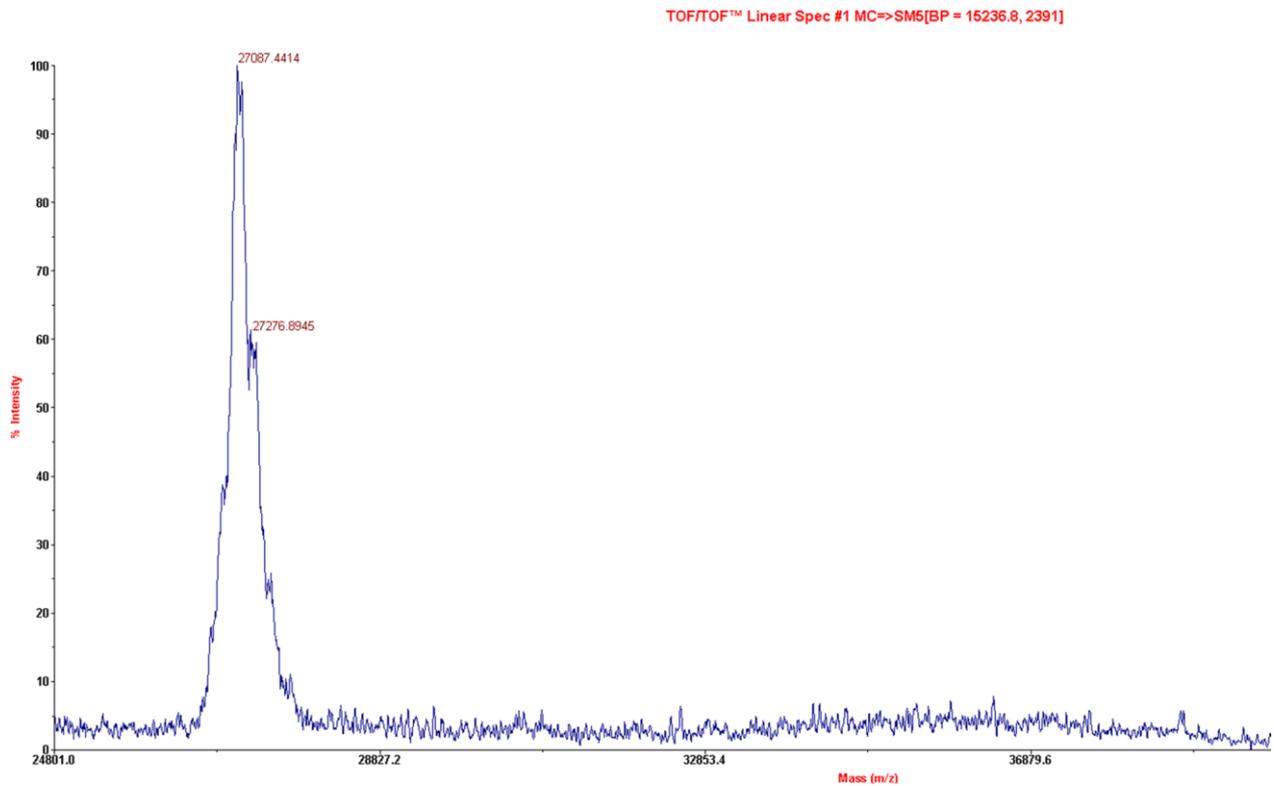
Chemical reagent	Concentration	Residual activity (%)
<i>In absence of substrate (active site not protected)</i>		
No addition	-	100 ± 2.6
PMSF	1 mM	84.7 ± 3.5
	10 mM	51.2 ± 5.5
<i>In presence of substrate (active site protected)</i>		
No addition	-	100 ± 0.5
PMSF	1 mM	100.5 ± 1.2
	10 mM	90.0 ± 1.3

**Figure S1.** Predicted 3D structures of hypothetical lipases from *Streptomyces exfoliatus* DSMZ 41693. A. SeLipA; B. SeLipB; C. SeLipC; D. SeLipD. Models were obtained by RoseTTAFold program [29] and their structures were visualized by PyMOL program (Schrodinger, LLC, version 2.5).

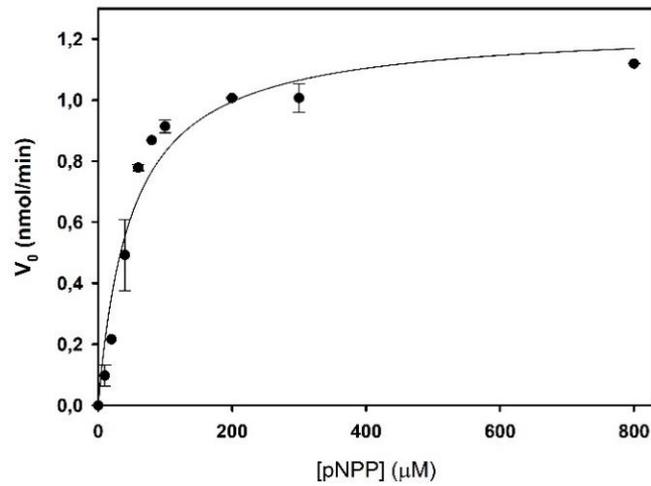
gi|639917083 Mass: 31194 Score: 659 Expect: 1.3e-066 Matches: 21  
lipase [*Streptomyces exfoliatus*]

Query	Start	End	Observed	Mr(expt)	Mr(calc)	ppm	M	Score	Peptide
59	84	109	2896.5251	2895.5178	2895.4418	26.2	0		R.GYCVFSLDYGQLPNVPPFFHGLGPIAK.S
5	110	120	1266.5919	1265.5846	1265.5888	-3.28	0		K.SAEQLDAYVDR.V
6	110	120	1266.5919	1265.5846	1265.5888	-3.28	0	66	K.SAEQLDAYVDR.V
72	110	143	3544.8044	3543.7971	3543.6722	35.2	1		K.SAEQLDAYVDRVLAATGAPEADLVGHSQGGMMMPR.W + 2 Oxidation (M)
39	121	143	2297.1086	2296.1013	2296.0940	3.19	0		R.VLAATGAPEADLVGHSQGGMMMPR.W + 2 Oxidation (M)
56	121	147	2887.5613	2886.5540	2886.4157	47.9	1		R.VLAATGAPEADLVGHSQGGMMMPRWYK.F + 2 Oxidation (M)
48	148	175	2836.6228	2835.6155	2835.5495	23.3	1		K.FLGGAEKVNTLVGIAPDNHGTLLGLTK.L
32	155	175	2134.1792	2133.1719	2133.1794	-3.50	0		K.VNTLVGIAPDNHGTLLGLTK.L
33	155	175	2134.1792	2133.1719	2133.1794	-3.50	0	69	K.VNTLVGIAPDNHGTLLGLTK.L
67	176	206	3163.7524	3162.7451	3162.6489	30.4	0		K.LLPYFPGAEDLISSATPGLADQIAGSAFITK.L
68	176	206	3163.7524	3162.7451	3162.6489	30.4	0	112	K.LLPYFPGAEDLISSATPGLADQIAGSAFITK.L
3	207	218	1213.6143	1212.6070	1212.6099	-2.35	0		K.LNEGGDTPGV.R.Y
4	207	218	1213.6143	1212.6070	1212.6099	-2.35	0	42	K.LNEGGDTPGV.R.Y
27	219	234	1939.9414	1938.9341	1938.9476	-6.94	0		R.YHVIASQYDEVVTPYR.S
28	219	234	1939.9414	1938.9341	1938.9476	-6.94	0	143	R.YHVIASQYDEVVTPYR.S
29	219	234	1939.9414	1938.9341	1938.9476	-6.94	0	152	R.YHVIASQYDEVVTPYR.S
22	235	251	1859.9799	1858.9726	1858.9789	-3.37	0		R.SQFLSGPNVTNLIQDK.C
12	252	267	1755.8586	1754.8513	1754.8621	-6.16	0		K.CALDSEHVAIGTVDR.V
70	252	281	3276.6643	3275.6570	3275.6357	6.51	1		K.CALDSEHVAIGTVDRVTFHEVANALDPAR.A
9	268	281	1539.7820	1538.7747	1538.7841	-6.12	0		R.VTFHEVANALDPAR.A
10	268	281	1539.7820	1538.7747	1538.7841	-6.12	0	70	R.VTFHEVANALDPAR.A

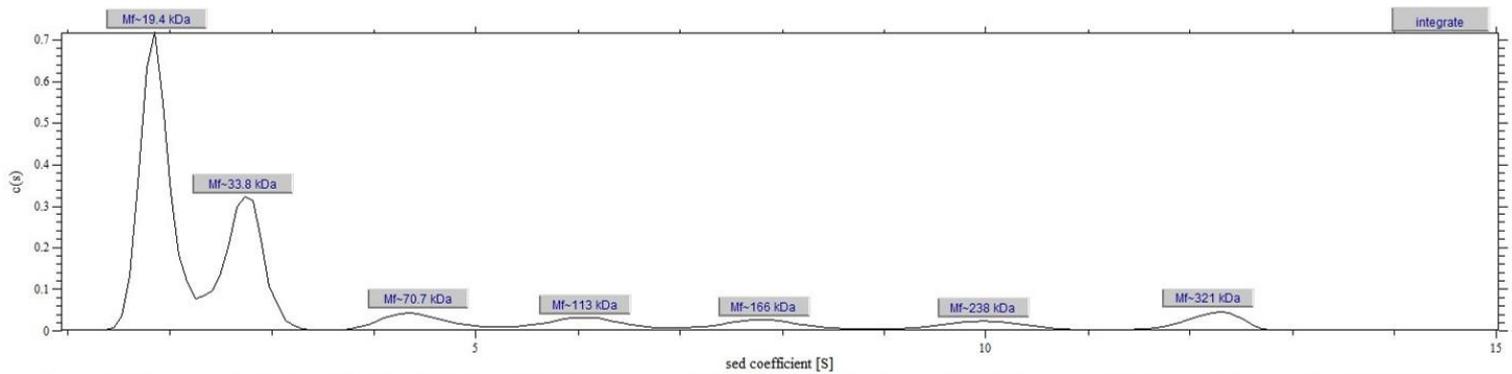
**Figure S2.** Peptide mass fingerprinting of SeLipC obtained by MALDI-TOF analysis. The protein was identified as a lipase from *Streptomyces exfoliatus* (gi|639917083) after peptide matching process using MASCOT Software (Score 659).



**Figure S3.** Determination of the molecular mass of SeLipC by MALDI-TOF. Mass spectrum recorded with 0.127 mg/mL of pure lipase.



**Figure S4.** Michaelis-Menten hyperbola plot for the determination of kinetic parameters of SeLipC using pNPP as substrate. Conditions: 200  $\mu\text{L}$  assay volume, 0.05  $\mu\text{g/mL}$  of enzyme (1.81 nM), assay time 10 min, 37°C, 50 mM Tris HCl pH 8.0, 0.01% gum arabic and 0.02% sodium deoxycholate.  $K_M = 49.6 \pm 11.2 \mu\text{M}$  and  $V_{max} = 1.24 \pm 0.08 \times 10^{-3} \mu\text{moles/min}$  ( $k_{cat} = 57 \text{ s}^{-1}$  and  $k_{cat} / K_M = 1.15 \times 10^6 \text{ s}^{-1} \cdot \text{M}^{-1}$ ).



**Figure S5.** Analytical ultracentrifugation analysis of SeLipC:  $c(s)$  distribution as a function of sedimentation coefficient ( $S$ ). Experiment was performed with 0.320 mg/mL of pure lipase, and the presence of two main species with sedimentation coefficient ( $S$ ) values of 1.9 and 2.7 were observed. These values are compatible with the presence of monomers and dimers with spheroid oblate-shape in both forms.