



Editorial

Myelinodegeneration vs. Neurodegeneration in MS Progressive Forms

Serge Nataf ^{1,2,3}

¹ Bank of Tissues and Cells, Hospices Civils de Lyon, Hôpital Edouard Herriot, Place d'Arsonval, F-69003 Lyon, France; serge.nataf@inserm.fr

² Stem-Cell and Brain Research Institute, 18 Avenue de Doyen Lépine, F-69500 Bron, France

³ Lyon-Est School of Medicine, University Claude Bernard Lyon 1, 43 Bd du 11 Novembre 1918, F-69100 Villeurbanne, France

In MS patients with a progressive form of the disease, the slow deterioration of neurological functions is thought to result from a combination of neuronal cell death, axonal damages and synaptic dysfunctions. Axonal alterations were first reported in the historical neuropathological observations made by Charcot in the 19th century [1]. However, approximately 25 years ago, the notion of inflammation-associated neurodegeneration was put on the front line of MS pathophysiology. Indeed, in the late 1990s, axonal alterations up to the stage of axonal transection were demonstrated in active MS lesions [2,3]. Axonal loss was then found to occur throughout the normal-appearing white matter (NAWM) in patients suffering from a primary progressive or a secondary progressive form of MS [4,5]. Such diffuse axonal alterations were proposed to essentially result from the sum of retrograde and anterograde degenerative processes initiated either at sites of axonal transection or from dying neurons [6,7]. Several works also provided evidence that axonal degeneration in MS may stem from a failure of mitochondrial energy metabolism [8–10]. On this basis, MS progression was proposed to arise from a diffuse axonopathy targeting predominantly long axons, i.e., axons with a high energy demand [11]. Although diffuse parenchymal inflammation was shown to correlate with diffuse axonal loss [4,5], the neurodegeneration-promoting impact of meningeal inflammation drew increasing interest in the last decade. In particular, neuropathological studies demonstrated that meningeal inflammation correlates with either the extent of neuronal cell loss in MS brains [12,13] or the level of axonal loss in MS spinal cords [14]. Importantly, a causative link between meningeal inflammation and MS-associated neurodegeneration was recently demonstrated in vivo in a rat model of experimentally induced meningeal inflammation [15]. More specifically, the over-expression of lymphotoxin-alpha in meningeal cells was shown to cause the death of cortical neurons and to induce the formation of meningeal tertiary lymphoid structures (TLS) [15]. In this regard, it should be noticed that TLS are essentially observed in MS patients with an accelerated and aggressive course of the disease [16]. Thus, one may argue that mechanisms involved in MS “aggressiveness” may not relate to those involved in MS “progressiveness”. Furthermore, although supported by a substantial amount of data, the neuroinflammation/neurodegeneration hypothesis fails to integrate the existence of diffuse myelin alterations in the central nervous system (CNS) of MS patients. Notably, we and others demonstrated large areas of partial demyelination in periplaque regions, which are associated with low levels of inflammation [17–19]. Interestingly, irrespective of localization (brain vs. spinal cord) and plaque activity (chronic-active vs. silent) an important molecular feature shared by periplaques is the down-regulation of *NDRG1*, an oligodendrocyte gene that plays a crucial role in myelin maintenance [20,21]. This observation is reminiscent of previous work that demonstrated the epigenetic silencing of *NDRG1* in “pathology-free” MS brain areas [22]. Additionally, in contrast with the expected inflammatory profile, periplaques exhibit a TGF-beta molecular signature reflecting an anti-inflammatory rather than a pro-inflammatory response [20,21,23]. Other



Citation: Nataf, S.

Myelinodegeneration vs.
Neurodegeneration in MS
Progressive Forms. *Int. J. Mol. Sci.*
2023, 24, 1596. <https://doi.org/10.3390/ijms24021596>

Received: 9 December 2022

Accepted: 14 December 2022

Published: 13 January 2023



Copyright: © 2023 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

works firmly demonstrated that, beyond periplaques, oligodendrocytes exhibit profound molecular and functional alterations throughout the CNS. This has been demonstrated ex vivo by epigenetics analyses, as mentioned above [22] but also, more recently, via single-cell RNA-seq analyses [24]. Similarly, the in vivo measures of myelin water fraction (MWF) by magnetic resonance imaging (MRI) clearly illustrate the occurrence of diffuse myelin alterations in the brain of MS patients [25,26]. Moreover, in MS patients, such decreased levels of MWF correlate with cognitive decline [27], disability scores [28], and extent of axonal loss [29]. This later point is of particular interest, as it raises the possibility that a process of myelin degeneration may precede or at least accompany neurodegeneration, as previously proposed [30,31]. Indeed, the functional coupling between oligodendrocytes and axons is now acknowledged, and oligodendrocytes were shown to support axonal energy metabolism via specific glycolytic functions [32,33]. Supporting this view, it was previously shown that mice KO for the myelin genes Mag, Plp or Cnp develop a neurodegenerative process characterized by a diffuse axonopathy [34–37]. Finally, in mice, genetically determined alterations of myelinating oligodendrocyte are sufficient to provoke or amplify CNS neuroinflammation [38,39].

Future studies should aim to further characterize myelin degeneration in the CNS of MS patients and determine how diffuse myelin alterations, neuroinflammation and neurodegeneration develop concurrently during the course of MS progressive forms.

Funding: This research received no external funding.

Conflicts of Interest: The author declares no conflict of interest.

References

1. Kornek, B.; Lassmann, H. Axonal pathology in multiple sclerosis. A historical note. *Brain Pathol.* **1999**, *9*, 651–656. [[CrossRef](#)] [[PubMed](#)]
2. Trapp, B.D.; Peterson, J.; Ransohoff, R.M.; Rudick, R.; Mörk, S.; Bö, L. Axonal Transection in the Lesions of Multiple Sclerosis. *N. Engl. J. Med.* **1998**, *338*, 278–285. [[CrossRef](#)] [[PubMed](#)]
3. Ferguson, B.; Matyszak, M.K.; Esiri, M.M.; Perry, V.H. Axonal damage in acute multiple sclerosis lesions. *Brain* **1997**, *120*, 393–399. [[CrossRef](#)]
4. Frischer, J.M.; Bramow, S.; Dal-Bianco, A.; Lucchinetti, C.F.; Rauschka, H.; Schmidbauer, M.; Laursen, H.; Sorensen, P.S.; Lassmann, H. The relation between inflammation and neurodegeneration in multiple sclerosis brains. *Brain* **2009**, *132*, 1175–1189. [[CrossRef](#)] [[PubMed](#)]
5. Kutzelnigg, A.; Lucchinetti, C.F.; Stadelmann, C.; Brück, W.; Rauschka, H.; Bergmann, M.; Schmidbauer, M.; Parisi, J.E.; Lassmann, H. Cortical demyelination and diffuse white matter injury in multiple sclerosis. *Brain* **2005**, *128*, 2705–2712. [[CrossRef](#)] [[PubMed](#)]
6. Balk, L.J.; Steenwijk, M.D.; Tewarie, P.; Daams, M.; Killestein, J.; Wattjes, M.P.; Vrenken, H.; Barkhof, F.; Polman, C.H.; Uitdehaag, B.M.J.; et al. Bidirectional trans-synaptic axonal degeneration in the visual pathway in multiple sclerosis. *J. Neurol. Neurosurg. Psychiatry* **2015**, *86*, 419–424. [[CrossRef](#)] [[PubMed](#)]
7. Gabilondo, I.; Martínez-Lapiscina, E.H.; Martínez-Heras, E.; Fraga-Pumar, E.; Llufriu, S.; Ortiz, S.; Bullich, S.; Sepulveda, M.; Falcon, C.; Berenguer, J.; et al. Trans-synaptic axonal degeneration in the visual pathway in multiple sclerosis. *Ann. Neurol.* **2014**, *75*, 98–107. [[CrossRef](#)]
8. Lassmann, H.; Van Horssen, J.; Mahad, D. Progressive multiple sclerosis: Pathology and pathogenesis. *Nat. Rev. Neurol.* **2012**, *8*, 647–656. [[CrossRef](#)]
9. Licht-Mayer, S.; Campbell, G.R.; Canizares, M.; Mehta, A.R.; Gane, A.B.; McGill, K.; Ghosh, A.; Fullerton, A.; Menezes, N.; Dean, J.; et al. Enhanced axonal response of mitochondria to demyelination offers neuroprotection: Implications for multiple sclerosis. *Acta Neuropathol.* **2020**, *140*, 143–167. [[CrossRef](#)]
10. Campbell, G.R.; Worrall, J.T.; Mahad, D.J. The central role of mitochondria in axonal degeneration in multiple sclerosis. *Mult. Scler. J.* **2014**, *20*, 1806–1813. [[CrossRef](#)]
11. Giovannoni, G.; Cutter, G.; Pia-Sormani, M.; Belachew, S.; Hyde, R.; Koendgen, H.; Knappertz, V.; Tomic, D.; Leppert, D.; Herndon, R.; et al. Is multiple sclerosis a length-dependent central axonopathy? The case for therapeutic lag and the asynchronous progressive MS hypotheses. *Mult. Scler. Relat. Disord.* **2017**, *12*, 70–78. [[CrossRef](#)]
12. Magliozzi, R.; Fadda, G.; Brown, R.A.; Bar-Or, A.; Howell, O.W.; Hametner, S.; Marastoni, D.; Poli, A.; Nicholas, R.; Calabrese, M.; et al. “Ependymal-in” Gradient of Thalamic Damage in Progressive Multiple Sclerosis. *Ann. Neurol.* **2022**, *92*, 670–685. [[CrossRef](#)]
13. Magliozzi, R.; Howell, O.W.; Reeves, C.; Roncaroli, F.; Nicholas, R.; Serafini, B.; Aloisi, F.; Reynolds, R. A Gradient of neuronal loss and meningeal inflammation in multiple sclerosis. *Ann. Neurol.* **2010**, *68*, 477–493. [[CrossRef](#)]
14. Androdias, G.; Reynolds, R.; Chanal, M.; Ritleng, C.; Confavreux, C.; Nataf, S. Meningeal T cells associate with diffuse axonal loss in multiple sclerosis spinal cords. *Ann. Neurol.* **2010**, *68*, 465–476. [[CrossRef](#)]

15. James Bates, R.E.; Browne, E.; Schalks, R.; Jacobs, H.; Tan, L.; Parekh, P.; Magliozi, R.; Calabrese, M.; Mazarakis, N.D.; Reynolds, R. Lymphotxin-alpha expression in the meninges causes lymphoid tissue formation and neurodegeneration. *Brain* **2022**, *12*. [[CrossRef](#)]
16. Howell, O.W.; Reeves, C.A.; Nicholas, R.; Carassiti, D.; Radotra, B.; Gentleman, S.M.; Serafini, B.; Aloisi, F.; Roncaroli, F.; Magliozi, R.; et al. Meningeal inflammation is widespread and linked to cortical pathology in multiple sclerosis. *Brain* **2011**, *134*, 2755–2771. [[CrossRef](#)]
17. Möller, J.R.; Yanagisawa, K.; Brady, R.O.; Tourtellotte, W.W.; Quarles, R.H. Myelin-associated glycoprotein in multiple sclerosis lesions: A quantitative and qualitative analysis. *Ann. Neurol.* **1987**, *22*, 469–474. [[CrossRef](#)]
18. Johnson, D.; Sato, S.; Quarles, R.H.; Inuzuka, T.; Brady, R.O.; Tourtellotte, W.W. Quantitation of the Myelin-Associated Glycoprotein in Human Nervous Tissue from Controls and Multiple Sclerosis Patients. *J. Neurochem.* **1986**, *46*, 1086–1093. [[CrossRef](#)]
19. Lieury, A.; Chanal, M.; Androdias, G.; Reynolds, R.; Cavagna, S.; Giraudon, P.; Confavreux, C.; Nataf, S. Tissue remodeling in periplaque regions of multiple sclerosis spinal cord lesions. *Glia* **2014**, *62*, 1645–1658. [[CrossRef](#)]
20. Nataf, S.; Guillen, M.; Pays, L. Irrespective of plaque activity, multiple sclerosis brain periplaques exhibit alterations of myelin genes and a TGF-beta signature. *Int. J. Mol. Sci.* **2022**, *23*, 14993. [[CrossRef](#)]
21. Nataf, S.; Barritault, M.; Pays, L. A unique TGFB1-driven genomic program links astrocytosis, low-grade inflammation and partial demyelination in spinal cord periplaques from progressive multiple sclerosis patients. *Int. J. Mol. Sci.* **2017**, *18*, 2097. [[CrossRef](#)] [[PubMed](#)]
22. Huynh, J.L.; Garg, P.; Thin, T.H.; Yoo, S.; Dutta, R.; Trapp, B.D.; Haroutunian, V.; Zhu, J.; Donovan, M.J.; Sharp, A.J.; et al. Epigenome-wide differences in pathology-free regions of multiple sclerosis-affected brains. *Nat. Neurosci.* **2014**, *17*, 121–130. [[CrossRef](#)] [[PubMed](#)]
23. Nataf, S.; Guillen, M.; Pays, L. TGFB1-mediated gliosis in multiple sclerosis spinal cords is favored by the regionalized expression of HOXA5 and the age-dependent decline in androgen receptor ligands. *Int. J. Mol. Sci.* **2019**, *20*, 5934. [[CrossRef](#)] [[PubMed](#)]
24. Jäkel, S.; Agirre, E.; Mendanha Falcão, A.; van Bruggen, D.; Lee, K.W.; Knuesel, I.; Malhotra, D.; Ffrench-Constant, C.; Williams, A.; Castelo-Branco, G. Altered human oligodendrocyte heterogeneity in multiple sclerosis. *Nature* **2019**, *566*, 543–547. [[CrossRef](#)]
25. Johnson, P.; Vavasour, I.M.; Stojkova, B.J.; Abel, S.; Lee, L.E.; Laule, C.; Tam, R.; Li, D.K.B.; Ackermans, N.; Schabas, A.J.; et al. Myelin heterogeneity for assessing normal appearing white matter myelin damage in multiple sclerosis. *J. Neuroimaging*, **2022**; *Online ahead of print*. [[CrossRef](#)]
26. Laule, C.; Vavasour, I.M.; Moore, G.R.W.; Oger, J.; Li, D.K.B.; Paty, D.W.; MacKay, A.L. Water content and myelin water fraction in multiple sclerosis: A T 2 relaxation study. *J. Neurol.* **2004**, *251*, 284–293. [[CrossRef](#)]
27. Abel, S.; Vavasour, I.; Lee, L.E.; Johnson, P.; Ackermans, N.; Chan, J.; Dvorak, A.; Schabas, A.; Wiggermann, V.; Tam, R.; et al. Myelin Damage in Normal Appearing White Matter Contributes to Impaired Cognitive Processing Speed in Multiple Sclerosis. *J. Neuroimaging* **2020**, *30*, 205–211. [[CrossRef](#)]
28. Kolind, S.; Matthews, L.; Johansen-Berg, H.; Leite, M.I.; Williams, S.C.R.; Deoni, S.; Palace, J. Myelin water imaging reflects clinical variability in multiple sclerosis. *Neuroimage* **2012**, *60*, 263–270. [[CrossRef](#)]
29. Yik, J.T.; Becquart, P.; Gill, J.; Petkau, J.; Traboulsee, A.; Carruthers, R.; Kolind, S.H.; Devonshire, V.; Sayao, A.L.; Schabas, A.; et al. Serum neurofilament light chain correlates with myelin and axonal magnetic resonance imaging markers in multiple sclerosis. *Mult. Scler. Relat. Disord.* **2022**, *57*, 103366. [[CrossRef](#)]
30. Laule, C.; Vavasour, I.M.; Leung, E.; Li, D.K.B.; Kozlowski, P.; Traboulsee, A.L.; Oger, J.; MacKay, A.L.; Wayne Moore, G.R. Pathological basis of diffusely abnormal white matter: Insights from magnetic resonance imaging and histology. *Mult. Scler. J.* **2011**, *17*, 144–150. [[CrossRef](#)]
31. Laule, C.; Pavlova, V.; Leung, E.; Zhao, G.; Mackay, A.L.; Kozlowski, P.; Traboulsee, A.L.; Li, D.K.B.; Moore, G.R.W. Diffusely abnormal white matter in multiple sclerosis: Further histologic studies provide evidence for a primary lipid abnormality with neurodegeneration. *J. Neuropathol. Exp. Neurol.* **2013**, *72*, 42–52. [[CrossRef](#)]
32. Saab, A.S.; Tzvetanova, I.D.; Nave, K.A. The role of myelin and oligodendrocytes in axonal energy metabolism. *Curr. Opin. Neurobiol.* **2013**, *23*, 1065–1072. [[CrossRef](#)]
33. Fünfschilling, U.; Supplie, L.M.; Mahad, D.; Boretius, S.; Saab, A.S.; Edgar, J.; Brinkmann, B.G.; Kassmann, C.M.; Tzvetanova, I.D.; Möbius, W.; et al. Glycolytic oligodendrocytes maintain myelin and long-term axonal integrity. *Nature* **2012**, *485*, 517–521. [[CrossRef](#)]
34. Lappe-Siefke, C.; Goebels, S.; Gravel, M.; Nicksch, E.; Lee, J.; Braun, P.E.; Griffiths, I.R.; Navel, K.A. Disruption of Cnp1 uncouples oligodendroglial functions in axonal support and myelination. *Nat. Genet.* **2003**, *33*, 366–374. [[CrossRef](#)]
35. Griffiths, I.; Klugmann, M.; Anderson, T.; Yool, D.; Thomson, C.; Schwab, M.H.; Schneider, A.; Zimmermann, F.; McCulloch, M.; Nadon, N.; et al. Axonal swellings and degeneration in mice lacking the major proteolipid of myelin. *Science* **1998**, *280*, 1610–1613. [[CrossRef](#)]
36. Quarles, R.H. Myelin-associated glycoprotein (MAG): Past, present and beyond. *J. Neurochem.* **2007**, *100*, 1431–1448. [[CrossRef](#)]
37. Pan, B.; Fromholt, S.E.; Hess, E.J.; Crawford, T.O.; Griffin, J.W.; Sheikh, K.A.; Schnaar, R.L. Myelin-associated glycoprotein and complementary axonal ligands, gangliosides, mediate axon stability in the CNS and PNS: Neuropathology and behavioral deficits in single- and double-null mice. *Exp. Neurol.* **2005**, *195*, 208–217. [[CrossRef](#)]

38. Madsen, P.M.; Desu, H.L.; Vaccari, J.P.d.R.; Florimon, Y.; Ellman, D.G.; Keane, R.W.; Clausen, B.H.; Lambertsen, K.L.; Brambilla, R. Oligodendrocytes modulate the immune-inflammatory response in EAE via TNFR2 signaling. *Brain. Behav. Immun.* **2020**, *84*, 132–146. [[CrossRef](#)]
39. Qiu, S.; Palavicini, J.P.; Wang, J.; Gonzalez, N.S.; He, S.; Dustin, E.; Zou, C.; Ding, L.; Bhattacharjee, A.; Van Skike, C.E.; et al. Adult-onset CNS myelin sulfatide deficiency is sufficient to cause Alzheimer’s disease-like neuroinflammation and cognitive impairment. *Mol. Neurodegener.* **2021**, *16*, 64. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.