



Amyloid Precursor Protein and Alzheimer's Disease

Kseniia S. Orobets and Andrey L. Karamyshev *D

Department of Cell Biology and Biochemistry, Texas Tech University Health Sciences Center, Lubbock, TX 79430, USA; korobets@ttuhsc.edu

* Correspondence: andrey.karamyshev@ttuhsc.edu; Tel.: +1-806-743-4102

Abstract: Alzheimer's disease (AD) is one of the most common neurodegenerative disorders associated with age or inherited mutations. It is characterized by severe dementia in the late stages that affect memory, cognitive functions, and daily life overall. AD progression is linked to the accumulation of cytotoxic amyloid beta ($A\beta$) and hyperphosphorylated tau protein combined with other pathological features such as synaptic loss, defective energy metabolism, imbalances in protein, and metal homeostasis. Several treatment options for AD are under investigation, including antibodybased therapy and stem cell transplantation. Amyloid precursor protein (APP) is a membrane protein considered to play a main role in AD pathology. It is known that APP in physiological conditions follows a non-amyloidogenic pathway; however, it can proceed to an amyloidogenic scenario, which leads to the generation of extracellular deleterious $A\beta$ plaques. Not all steps of APP biogenesis are clear so far, and these questions should be addressed in future studies. AD is a complex chronic disease with many factors that contribute to disease progression.

Keywords: neurodegenerative disease; Alzheimer's disease; amyloid precursor protein (APP); amyloid beta; protein biogenesis; protein transport; membrane proteins; SRP-dependent targeting

1. Introduction

Alzheimer's disease (AD) is a severe neurological disorder and the most common type of dementia across the world. According to Alzheimer's Association, AD contributes to 60-80% of all dementia cases worldwide. As estimated, in 2023, there will be 6.7 million people who are 65 years old or older living in the United States with Alzheimer's disease [1]. It is predicted that a dramatic elevation of AD pathology will occur in the future, with 13.85 million Americans affected by Alzheimer's dementia and 152 million affected around the world by the year 2050 [2,3]. In 2019, the World Health Organization (WHO) reported USD 1.3 trillion as the dementia cost around the world, including care expenses from family members and friends who do not fall into the category of professional caregivers and medical personnel. A huge number of current AD patients, their dramatic increase with an aging population in the near future, and the devastating economic costs put pressure on governments to address these issues through new policies for medical care providers to find efficient ways to treat patients and for the scientific community and pharmacologists to discover the mechanism of this disorder, developing markers for its early detection and finding new potential effective treatment and the disease cure. However, despite the fact that intensive studies and significant funding for Alzheimer's disease research have been undertaken, no breakthrough discovery has been made regarding the mechanism, and many promising therapies have failed; currently, only a few pharmacological treatments have received approval or are under consideration by the FDA, providing only mild improvement in patients [4]. Thus, the significance of the study related to AD is obvious.

Alzheimer's disease is represented in two forms—familial (inherited) and sporadic. Familial AD is the autosomal-dominant form of the disease and is characterized by relatively early onset under the age of 65, contributing to around 1% of all cases [1]. The sporadic form usually develops after 65 years and, therefore, is referred to as late-onset



Citation: Orobets, K.S.; Karamyshev, A.L. Amyloid Precursor Protein and Alzheimer's Disease. *Int. J. Mol. Sci.* 2023, 24, 14794. https://doi.org/ 10.3390/ijms241914794

Academic Editors: Hari Shanker Sharma and Claudia Ricci

Received: 30 July 2023 Revised: 20 September 2023 Accepted: 26 September 2023 Published: 30 September 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Alzheimer's disease (LOAD). It is the most common form of AD. Familial and sporadic cases are triggered by mutations in different genes (discussed in detail below) or by alternative mRNA splicing [5]. There are several studied cases with a very early onset of AD reported, suggesting the increasing number of affected people, even in the younger generation [6–9]. With the age of disease decreasing and the general population getting older, the development of functional treatment, preventive medicine, and effective diagnostics stay in the focus of attention and is the most wanted.

The clinical picture of Alzheimer's disease is identical for inherited and familial cases. It comprises memory loss, decreasing thinking skills and solving problems, and the inability to cope with daily tasks. This functional decline is accompanied by changes in personality and behavior and withdrawal from social life and work. Finally, in the late stages, patients are fully dependent on caregivers or special facilities. Alzheimer's disease affects not only patients diagnosed with this disorder but also their families, with the patients being a large burden in many ways. The progression of this disease usually takes years and starts much earlier than the first symptoms can be detected. Biological changes, such as the presence of specific biomarkers in patient samples or the accumulation of pathological hallmarks, can help to diagnose the disease at the so-called preclinical or pre-symptomatic stage [1,10]. Mild cognitive impairment represents the next stage of the disease progression, characterized by mild symptoms without much interference with daily tasks. The final stage is Alzheimer's dementia, which can also be in mild, moderate, or severe, causing minor to drastic interference with everyday life.

Alzheimer's disease has a strong association with genetics and cellular mechanisms, yet it is a chronic and complex disorder where additional risk factors contribute to disease onset and progression. Genetics and age are the strongest and nonmodifiable risk factors. Health factors (heart and blood vessel conditions, hypertension, and diabetes) and behavior factors (diet, physical activity, level of education, and cognitive engagement) are mixed together in an intricate interplay where the variables depend on each other.

2. The Genetics of Alzheimer's Disease

The molecular basis of Alzheimer's disease has been studied for decades. Among the hallmarks of neurodegenerative disorders, the most recent data define not only the aberrant aggregation of proteins but also the dysfunction of neuronal networks, defective energy metabolism, abnormalities in the cytoskeleton, and alterations to protein and metal homeostasis, as well as declining memory, language, and thinking abilities [11].

The most known and studied molecular marker of AD is the accumulation of extracellular plaques built up by amyloid β protein (A β) and intracellular neurofibrillary tangles (NFTs) formed by hyperphosphorylated tau-protein in brain neurons. To date, several hypotheses of Alzheimer's disease onset are being discussed in the field. The major hypothesis implicates the defective cleavage of amyloid precursor protein (APP) and the consequent amyloid beta plaque formation as a predominant basis for Alzheimer's disease, giving rise to a downstream cascade that leads to tau-pathology [12]. However, nowadays, there is a tendency to show the interplay between these two factors [13,14].

Although aggregated A β and tau are the major characteristics of AD on the microscopic level, the molecular pathology of the disease is not limited to only these two proteins. There is a plethora of genes associated with a higher risk for Alzheimer's disease. Genomewide association studies (GWASs) help to identify novel mutations in those genes related to the sporadic form of the disease. This topic has been in the research field for years, widening the list of potentially pathogenic mutations and confirming the genetic complexity of Alzheimer's disease [15–17]. There has been progress in genetic screens that are linked to other genes, including *APOE*, *TREM2*, *SORL1*, and *ABCA7*, with the disease [18,19]. Recent studies identified 75 loci for AD (42 of them were new, and 33 were previously found) [20]. Some of these newly identified genes may regulate APP recycling in the endosomal system and modulate APP metabolism by influencing lipid metabolism and inflammation [21–23].

APOE4, a variant of the APOE gene, is associated with a high risk of the development of the sporadic form of AD, but the reason for such effect is still not clear. Apolipoprotein E, which is encoded by this gene, regulates lipoprotein uptake and interferes with lipid transport and lipid metabolism in the brain. Defects in APOE4 lead to the common pathological characteristics of AD, such as mitochondrial dysfunction, changes in synaptic plasticity, and neuroinflammation [24]. Transcriptomic analyses of APOE4 neurons, astrocytes, and microglia-like cells (derived from induced pluripotent stem cells—iPSC) revealed many differentially expressed genes. Notably, in APOE4 neurons, the production of A β is increased, as well as the number of endosomes, where major A β generation takes place [25]. With elevated neuronal A β production, A β uptake by astrocytes is compromised, leading to an increase in extracellular amyloid deposition. These events are accompanied by the activation of an inflammatory response in microglia-like cells and the upregulation of proinflammatory genes [26,27]. The removal of the APOE4 allele in a mouse model leads to a decrease in another AD hallmark, hyperphosphorylated tau and tau-associated neurodegeneration in microglia. It indicates that APOE4 affects tau pathology [26,28,29]. Changes in cholesterol metabolism were also observed. Another study using transcriptomic analysis demonstrated alterations in lipid metabolism in APOE4 astrocytes and microglia, resulting in increased cholesterol synthesis in combination with high cholesterol accumulation in lysosomes, suggesting defects in cholesterol turnover in these cell types; however, this was only in humans [30]. Eventually, an oversupply of cholesterol by astrocytes promotes amyloidogenic APP processing in neurons due to the increased formation of lipid rafts, which APP is associated with [31].

Familial forms of AD are connected to the defective proteins involved in the generation of A β and are caused by mutations in *PSEN1*, *PSEN2*, or *APP* genes. For the *APP* gene, 25 mutations were described as pathogenic [32]. For *PSEN1* and *PSEN2*, there are around 200 different pathogenic mutations that have been identified as contributing to disease development [33,34]. The *PSEN1* and *PSEN2* genes encode the proteins presenilin 1 and presenilin 2, respectively. Both these proteins modulate the activity of γ -secretase, a membrane-associated complex responsible for the cleavage of different proteins, including APP. It was established that mutations in *PSENs* affect γ -secretase activity through the destabilization of the enzyme-substrate complex. In APP processing, the production of longer A β peptides is what stimulates amyloid generation and shifts the balance towards A β accumulation [35]. Additionally, it was suggested that *PSENs* mutations trigger pathological alterations in mitochondrial metabolism, which is one of the cellular hallmarks of AD [36]. Mutations in *APP* contribute to AD by increasing the production of the most toxic A β 42 peptides or through stimulating A β aggregation but not through the alterations of APP function [34].

3. Early Biogenesis of Amyloid Precursor Protein

Despite extensive research into APP biology, especially its processing, the early steps of APP biogenesis are still unknown. Generally, newly synthesized proteins are marked with specific targeting signals for the final protein destination. Depending on these signals, the proteins are transported to different organelles such as the endoplasmic reticulum (ER), Golgi apparatus, plasma membrane, nucleus, mitochondrion, endosomes, lysosomes, or peroxisomes. APP is located in the plasma membrane and other membrane organelles. Thus, it must undergo certain trafficking to reach these subcellular locations. Here, we discuss the general protein trafficking pathway in eukaryotic organisms and analyze its relevance to APP biogenesis.

In general, secretory and membrane proteins follow a specific path during their biogenesis. For proper folding and transport, they are targeted to the endoplasmic reticulum with the assistance of the signal recognition particle (SRP), which is a major route of protein transport in eukaryotes [37,38]. SRP is a ribonucleoprotein complex that is able to bind signal peptides and ribosomes, and is able to transport its cargo to the SRP receptor (SR) in the ER membrane. Mutations in the SRP subunits are associated with multiple human

diseases [39]. SRP recognizes a specific part of a polypeptide emerging from the ribosome exit tunnel during translation. This cleavable N-terminal region of secretory proteins is known as a signal peptide; its properties and features were originally described in Dr. G von Heijne's works [40-42]. It was shown that signal peptides do not have amino acid sequence homology; instead, they have common physico-chemical properties, including a stretch of hydrophobic amino acids in the central part. The importance of this signal peptide's parameters was highlighted in several studies [43–46]. The defective signal peptide of preprolactin (PPL) does not allow for normal interaction between SRP and the nascent chain of PPL, triggering a specific mechanism of mRNA degradation, named regulation of aberrant protein production (RAPP) [44]. RAPP is one of the protein quality control mechanisms in eukaryotes, and it is activated when SRP cannot bind the nascent chain and the targeting of secretory and membrane proteins is compromised [47,48]. So far, RAPP has been associated with the degradation of the mRNAs of several different secretory proteins in addition to preprolactin. Thus, it was shown that disease-associated mutations in multiple secretory proteins, including granulin (the protein associated with neurodegenerative disease frontotemporal lobar degeneration or FTLD), activate the RAPP pathway [49,50]. It was also suggested that SRP is involved in alpha-synuclein biogenesis, and RAPP may play a role in Parkinson's disease [51]. Finally, a deep RNA-seq analysis revealed the connection between the loss of SRP interaction with a signal peptide and various metabolic, immune, and age-related disorders, as well as cancer [52]. It was established that RAPP is a general pathway that controls the quality of SRP-dependent secretory and membrane proteins in the ribosome [52]. However, despite the in-depth studies on the interaction between SRP and ribosome-associated polypeptides and the control of their quality during translation, the fundamental questions of which proteins are SRP-dependent and which proteins are SRP-independent have not been answered yet.

Similar to many secretory and membrane proteins, APP has an N-terminal signal peptide, which is remarkably hydrophobic. The APP signal peptide consists of 17 amino acid residues, and five of them are leucines, which makes it a potential candidate for being an SRP substrate. The APP signal sequence marks this protein for ER targeting, but it was linked to SRP only indirectly [53] and was briefly discussed as a client for cotranslational targeting [54]. There are few studies in which the early stages of APP biogenesis are the focus of the interest. APP was identified as a client of the SEC61 translocon, one of the main entry gates to the ER [55]. The SEC61 translocon is a protein complex in the ER membrane, to which SRP cotranslationally brings its cargo [56,57]. SEC61 is one of the major entry points to the ER, and it can be engaged with other trafficking partners [58,59]. Thus, it is still to be determined how APP is targeted to the ER and what partners are involved in its transport; this can shed light on early APP biogenesis and its possible effect on Alzheimer's disease onset.

4. Amyloid Precursor Protein Processing

Amyloid precursor protein is a type I membrane protein. It is encoded by the *APP* gene located on chromosome 21 in humans [60–62]. APP is widely expressed in the body, with higher expression in the neuronal tissues in the brain. Differential processing and alternative splicing generate different isoforms of APP in a tissue-dependent manner [63]. The three major variants are APP695, APP751, and APP770, and all of them are capable of producing amyloid beta [64]. Isoforms APP751 and APP770 are mostly present in non-neuronal tissues, while APP695 is predominantly found in neurons and is considered the most toxic. APP functions are diverse and are associated with the neurogenesis and differentiation of neuronal cells, synaptic mechanisms, cell cycle and adhesion, and calcium metabolism [65–67]. APP-deficient mice exhibit a shortened lifespan, cognitive and learning impairment, and altered metal homeostasis in the brain regions typically affected by the disease [68–70].

APP processing is a multistep mechanism that involves several cleavage events to release different products. APP biogenesis can be divided into distinct general steps, as shown in Figure 1.

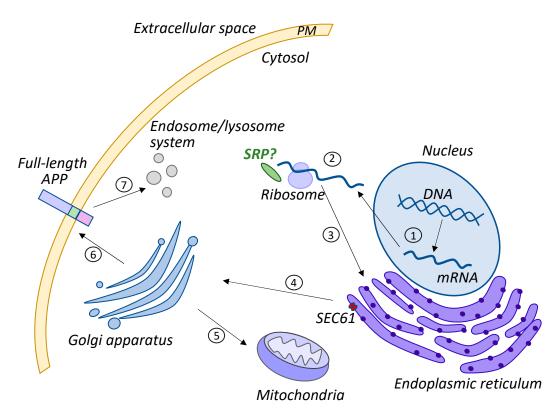


Figure 1. Intracellular APP trafficking. (1) *APP* transcription in the nucleus and mRNA export to the cytoplasm. (2) APP mRNA translation on a ribosome with the assistance of SRP or unestablished targeting factors (no experimental evidence of SRP involvement yet, thus, it is indicated by a question mark). (3) The transport of nascent APP to ER for further biogenesis. (4) The transition to Golgi for post-translational modifications. (5) The transport of full-length APP to mitochondria and the insertion into the mitochondria membrane. (6) The transport of full-length APP to the plasma membrane (PM). (7) The internalization of full-length APP into the endosomal system for further cleavage.

The vast majority of the studies focus on the late stages of APP processing when the full-length APP is inserted into the plasma membrane or other intracellular membrane organelles and undergoes cleavage events. The cleavage of membrane-inserted APP can follow two pathways—amyloidogenic or non-amyloidogenic (the most common one) (Figure 2). Three secretases play a central role in the late processing of APP: α -, β -, and γ -secretase. The non-amyloidogenic pathway starts with α -secretase releasing the N-terminal extracellular soluble APP domain (sAPP α) and the membrane-attached C83 fragment. α -secretase cuts the middle of the A β region; thus, this cleavage prevents the further formation of A β . Extracellular sAPP α can mitigate amyloid beta production via the inhibition of β -secretase (BACE1), which is the enzyme responsible for one of the steps in the amyloidogenic pathway of APP processing. Thus, sAPP α stimulates the non-amyloidogenic pathway [71,72]. The reintroduction of sAPP α into APP-depleted mouse models leads to the restoration of a normal phenotype, indicating the pivotal role of the sAPP α fragment in development [73].

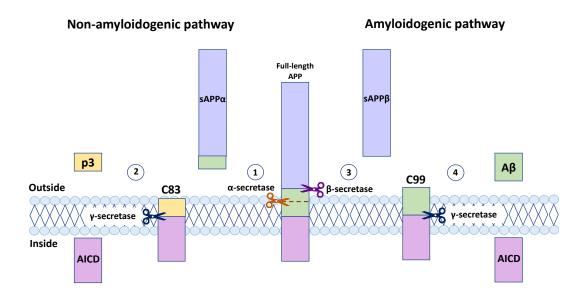


Figure 2. Amyloid precursor protein processing pathways. Full-length APP is inserted into the plasma membrane or intracellular membrane organelles, where it can proceed to the nonamyloidogenic or amyloidogenic pathway. (1) The non-amyloidogenic pathway starts with cleavage by α -secretase, which cuts full-length APP at the A β mid-region. This cleavage produces sAPP α and the membrane-bound C83 fragment. (2) The C83 fragment is cleaved further by γ -secretase to release the p3 molecule extracellularly and AICD (amyloid precursor protein intracellular domain) intracellularly. (3) The amyloidogenic pathway starts with β -secretase cleavage, which occurs on the membrane. It produces extracellular sAPP β and the C99 fragment associated with the membrane. (4) γ -secretase cuts the C99 fragment, and A β is released outside of the cell, whereas AICD stays inside.

The first cleavage in the amyloidogenic pathway is performed by β -secretase, also known as β -site APP-cleaving enzyme-1 (BACE1). This cleavage produces the extracellular soluble APP β (sAPP β) fragment and membrane-bound C99 domain. The importance of BACE1 for the production of aberrant amyloid beta was demonstrated in several studies. Remarkably, the experiments with mouse models for Alzheimer's disease revealed the complete absence of amyloid beta when BACE1 was silenced [74–77]. β -secretase is a membrane-associated enzyme with complex trafficking and diverse cellular routs. It is synthesized in the ER in a proenzyme form, which acquires its full activity after several post-translational modifications in the Golgi, including palmitoylation, glycosylation, acetylation, and phosphorylation, which have been shown to be essential for this enzyme to trigger amyloidogenic events [78,79]. After β -secretase insertion into the plasma membrane lipid rafts, it can be extracellularly released; therefore, APP processing by this enzyme rarely occurs on the plasma membrane. Then, this extracellular BACE1 is endocytosed to appear in the endosomes for functioning or to proceed to the lysosomes for degradation. Inside of the cell, BACE1 is mostly located on the membrane of the trans-Golgi network (TGN) and endosomes, where APP processing takes place [80]. Interestingly, initially, the APP from the plasma membrane is internalized through a clathrin-mediated mechanism, whereas BACE1 uses another clathrin-independent mechanism [81]. The optimal pH for this enzyme is 5.5; therefore, the predominant location of the possible APP processing and generation of A β is endosomes and lysosomes. Eventually, APP and β -secretase colocalize in Rab5-positive endosomes, where APP is cleaved by a fully active enzyme [82,83].

The first cleavage in both pathways results in the formation of the C83 and C99 membrane-bound domains in non-amyloidogenic and amyloidogenic scenarios, respectively. The physiological role of the C83 and C99 fragments is still uncharacterized. Both fragments are substrates for the γ -secretase enzyme complex. The cleavage of C83 or C99 by γ -secretase results in the production of amyloid precursor protein intracellular domain

(AICD) in both scenarios, amyloidogenic and non-amyloidogenic. AICD is known as a transcription factor containing the motif YENPTY, facilitating binding to other proteins [84–87]. AICD has been shown to be one of the regulators of APP processing, promoting intracellular APP trafficking. The APP intracellular domain can be phosphorylated at S655, stimulating the non-amyloidogenic pathway due to directing APP from endosomes with active BACE1 to TGN [88–90]. Phosphorylation at the position T668 may promote the amyloidogenic pathway [91], and likely, it interferes with APP intracellular processing [90,92]. Membrane-associated γ -secretase is a complex consisting of at least four transmembrane enzymes—presenilin (PS1 or PS2), presenilin-enhancer 2 (PEN-2), nicastrin (NCT), and anteriorpharynx-defective-1 (APH-1) [93,94]. As mentioned earlier, mutations in PSEN1 or *PSEN2* genes contribute to the development of familial AD. γ -secretase is not exclusively associated with APP processing. It is implicated in the Notch-pathway and tumorigenesis. There are more than 50 proteins, including E-Cadherin, CD44, and IGF1R (insulin-like growth factor receptor) among γ -secretase's substrates [95,96]. The location of γ -secretase is not limited to the plasma membrane; it is also located in mitochondria and lysosome membranes, as well as in early and late endosomes [97,98]. The ubiquitous localization of this enzyme complex supports the idea of the highly intricate processing of APP with many subcellular loci available for the potential generation of AB. Noticeably, the non-amyloidogenic pathway is predominantly associated with the plasma membrane [99,100], whereas amyloidogenic is connected to the endosomal system [82,83,101]. When C83 is cleaved by γ secretase, another product, p3, is released into extracellular space in the non-amyloidogenic pathway. To date, the biological role of this molecule has not been established.

Amyloid beta is one of the final products in the amyloidogenic pathway of APP processing. It is a small peptide consisting of 37–43 amino acids, where the A β 42 isoform is known to be the most deleterious. A β peptides can form extracellular soluble oligomers and plaques and insoluble fibrils, which are the main hallmark of Alzheimer's disease. This accumulation gives rise to pathological events, such as neuroinflammation, cytotoxic effects, and neuronal death. The aggregation of A β and its dynamics in laboratory conditions in vitro has been carefully investigated through various methods, including cryo-electron microscopy, atomic force microscopy, nuclear magnetic resonance, electron paramagnetic resonance, and X-ray [102–106]. A β peptides can build up in a different fashion to form diverse structures of β -sheets, depending on the arrangement of monomers and the orientation of β -strands and β -sheets [107]. An intriguing phenomenon of aggregated A β peptides was observed in several studies. Amyloids consisting of 2-12 monomers are considered to possess the highest level of toxicity, whereas longer forms can interact with their shorter counterparts to "isolate" them, reducing the harmful effects. Therefore, the aggregation of A β , despite being a main pathological signature of the disease, can actually help cells to survive via the mitigation of cytotoxic effects [108,109]. Another deleterious effect of A β accumulation is the disruption of the plasma membrane followed by changes in calcium (Ca^{2+}) flux. The pore-forming hypothesis is still controversial; however, growing evidence indicates that, indeed, $A\beta$ oligomers are inserted into the plasma membrane where they form Ca²⁺-permeable pores and disrupt calcium homeostasis, which also leads to neuronal damage and cell death [110,111].

5. Amyloid Precursor Protein and Mitochondria

It has been known and investigated for years that mitochondria are connected to amyloid precursor protein and Alzheimer's disease. This field can be divided into three major research questions: (1) how is APP transported to mitochondria, and where is it localized? (2) What is the role of APP in mitochondria, and how is mitochondrial metabolism affected during the disease? (3) Can mitochondria be a therapeutic target for AD treatment?

As of today, mitochondrial dysfunction is one of the pathological hallmarks of Alzheimer's disease; nevertheless, this condition is present in the majority of neurodegenerative disorders [112,113]. In addition to the plasma membrane, APP is transported to mitochondria due

APP was found in the mitochondria of AD patient brain samples, a recent study showed APP presence in both healthy and pathological brains [116,117]. Mitochondrial dysfunction was attributed to Alzheimer's disease, causing defects in metabolism, protein maturation in mitochondria, energy production, oxygen consumption, and mitochondrial calcium homeostasis [118–120]. Multiple attempts were made to assess the changes in mitochondrial metabolism with the overexpression of wild-type APP or its mutated forms. The results vary from one cell line to another as well as between study groups; therefore, there is no consensus about APP effects in mitochondria. A comprehensive review describes, in detail, the results of recent studies on how APP effects mitochondria in in vitro and in vivo models [121]. Mitochondria metabolism seems an appealing target for potential AD therapy. Cell replacement therapy with MSCs or MSC-conditioned media has the potential for a reduction in oxidative stress and the restoration of normal mitochondrial function in a mouse model [122]. Exploiting nanoparticles for targeting mitochondria was also investigated. MSCs-derived extracellular vesicles (EVs) with tyrosine phosphatase-2 (SHP2) deliver SHP2 to the brain, where it induces mitophagy and helps with the clearance of aberrant proteins [123,124].

6. Alzheimer's Disease Is a Complex Disorder

Alzheimer's disease pathology is triggered by genetic factors, such as inherited mutations in familial cases or sporadic mutations with a connection to age. However, there are other factors affecting the disease progression and severity of the symptoms. AD contributing factors are spread over all parts of APP biogenesis (Figure 3).

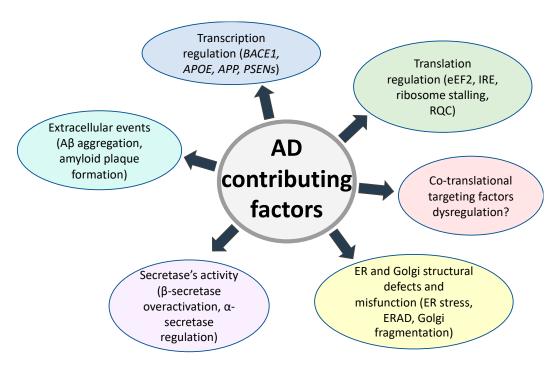


Figure 3. Alzheimer's disease contributing factors.

Versatile evidence indicates that Alzheimer's disease emerges from an imbalance between amyloid plaque accumulation and its degradation. The regulation of amyloid β production starts on the level of transcription. Autophagy is the main cellular mechanism for the clearance of aggregates and aberrant proteins. PPARA/PPAR α (peroxisome proliferator-activated receptor alpha) regulates the gene expression of autophagy, with lipid and glucose metabolism genes serving as some of the central regulators for mitochondrial function [125]. The pharmacological targeting of transcription factor PPARA activates autophagy in human microglial and glioma cells expressing APP, which leads to the partial removal of amyloid plaques and causes a shift towards A β clearance through transcriptional regulation [126]. A shift towards amyloid β production can be triggered by the regulation of genes directly involved in APP processing. For instance, *BACE1*, the gene that encodes β -secretase, has several transcription factor-binding sites that allow for the regulation of this gene by multiple transcription factors, for example, peroxisome proliferator-activated receptor gamma (PPAR γ), NF- κ B, specificity protein 1 (SP1). PPAR γ is a nuclear transcription factor that reduces the activity of the *BACE1* promotor when overexpressed. In AD patient, samples of a lower level of PPAR γ was detected, suggesting the overactivation of *BACE1* [127]. NF- κ B regulates *BACE1* expression differently under different conditions, such as lowering BACE1 expression in physiological conditions but promoting A β generation in pathology [128]. SP1 is one of the first identified regulators for *BACE1*, working as an activator for β -secretase expression and also interacting with NF- κ B [129,130]. The transcription of *APOE* can be upregulated by cyclic AMP (cAMP), retinoic acid (RA), PPAR γ , and A β itself. In the case of *APOE* transcriptional regulation via A β , it can be considered as a neuroprotective mechanism since ApoE helps prevent against cytotoxicity [131].

APP transcription can be activated in different cell types by heat-shock factor 1 (HSF-1), NF- κ B, and Rac1 [128]. *PSEN1* transcriptional regulation has been studied more than *PSEN2* and can be controlled by diverse transcription elements (Ets, ZNF237, cAMP-responsive element-binding protein, and p300) and chromatin modifications [132–134].

The activity of α -secretase is also subject to regulation, leading to changes in the balance of amyloid production. The protease furin effectively promotes the non-amyloidogenic pathway and the production of sAPP α , which, in turn, further stimulates this pathway. When furin is inhibited, the level of sAPP α is decreased (when the APP level is not changed), suggesting amyloidogenic pathway activation [135]. Remarkably, a recent study revealed an interplay between iron overload in neurodegeneration and the downregulation of furin, leading to elevated production of amyloids [136].

Defects in other stages of APP processing may also play a significant role in A β accumulation and aggregation. It was shown that alterations to APP mRNA translation can happen in the initiation and elongation steps. Elongation factor eEF2, when phosphorylated, slows down protein synthesis and leads to ribosome stalling. In AD mouse models, the phosphorylation of eEF2 is enhanced, suggesting the involvement of this elongation factor in AD pathology development [137]. Another mechanism that was shown to alter APP mRNA translation is iron-dependent. It was demonstrated that patients with neurodegenerative disorders, such as Alzheimer's or Parkinson's disease, have an elevated level of iron in the brain [138]. Iron toxicity is connected to the generation of reactive oxygen species in the brain following oxidative stress and neuronal death [139]. The iron-responsive element (IRE) of APP mRNA was identified and shown to regulate APP protein expression. When iron is chelated in a neuroblastoma cell line, APP protein synthesis is drastically decreased, demonstrating the involvement of iron in the regulation of APP translation [140]. If ribosomes with an APP nascent chain are stuck in the ER during translation, this activates ribosome-associated quality control (RQC), triggering a down-stream cascade of reactions. Abnormal RQC causes endolysosomal misfunction, which leads to the formation of an amyloid plaque core intracellularly [55]. Yet, as mentioned before, the APP targeting factors are still unestablished, and this can be another aspect of APP processing that might contribute to AD development.

The prevalent part of APP maturation takes place in the ER and Golgi, where posttranslational modifications occur; therefore, these subcellular locations are important for physiologically normal APP biogenesis. A plethora of studies have focused on ER stress and its connection to Alzheimer's disease. The unfolded protein response (UPR) is a protein quality control mechanism associated with ER. During mild ER stress, this mechanism is highly functional and beneficial for cells since it works for balancing ER and protein homeostasis [141]. But under severe stress conditions in AD, the UPR turns maladaptive and stimulates apoptosis, increasing neuronal death [142]. Another connection between APP biogenesis and ER was made through ER degradation-enhancing α -mannosidase-like protein 1 (EDEM1), a targeting factor for misfolded ER proteins. It targets aberrant proteins for degradation in the ER-associated protein degradation (ERAD) pathway [143]. In cell cultures expressing APP, EDEM1 promotes APP targeting from the ER to the cytoplasm, where ERAD takes place. It stimulates the proteosome degradation of APP inside the cell and leads to a consequent decrease in A β 40-42 production [144]. Golgi fragmentation was reported in AD pathology cases; however, the primary reason and consequence in the APP-Golgi relationship is still poorly understood. It was suggested that despite this, Golgi defects appear as a consequences of AD pathology in the early stages, and these defects also enhance amyloid formation and stimulate the amyloidogenic pathway [145]. The fragmentation of the Golgi apparatus is due to the phosphorylation of Golgi structural proteins (GRASP65), which happens through the A β -associated activation of cyclin-dependent kinase-5 (cdk5) [146].

7. Alzheimer's Disease Diagnostic and Treatment

Nowadays, technical progress has allowed for the diagnosis of Alzheimer's disease, even in the preclinical stage. An inadequate level of Aβ and hyperphosphorylated tau protein can be detected in cerebrospinal fluid (CSF). Positron emission tomography (PET) allows for the detection of accumulated amyloids and tau tangles in the brain. Recently, a new diagnostic technique was introduced. Blood-based biomarkers (BBMs) for Alzheimer's disease screening have several advantages, such as the simplicity of the test and its performance (blood tests can be carried out in any medical facility, whereas CSF analysis or PET can only be conducted in specialized clinics), lower cost, and additional biomarkers for neurodegeneration (neurofilament light chain (NfL) and glial fibrillary acidic protein (GFAP)) can be detected [147–149]. Several other methods, such as computed tomography (CT) or magnetic resonance imaging (MRI), may be used to refute Alzheimer's disease pathology or to support other possible diagnoses.

There are many potential options for the treatment of Alzheimer's disease, which are broadly being explored to this day. One of the most promising fields is antibody-based therapy to target $A\beta$. Several developed immunotherapy compounds have already entered clinical trials; however, many of them failed. By the end of 2022, there were four antibodybased therapeutic agents undergoing the final clinical phase: aducanumab, lecanemab, gantenerumab, and donanemab. These are monoclonal IgG1 antibodies with an affinity to aggregate amyloid beta forms [150]. In spite of high hopes for finding a curative medicine, it is too early to say if some of the suggested options may revolutionize Alzheimer's disease treatment.

Another treatment option that has become widely explored is stem cell therapy. Mesenchymal stem cells (MSCs), neural stem cells (NSCs), and embryonic stem cells (ESCs) are used for transplantation into AD mouse models to evaluate their potential curative effect on neurodegenerative pathology. Among the common effects between these different MSC lines, there has been an increase in synaptic plasticity, mitigation of inflammatory response, improved short-term memory and learning abilities, and cognitive improvement [151–156].

Since the sporadic form of Alzheimer's disease has a strong association with aging, anti-aging therapy is considered another approach for AD treatment, which has been actively investigated. Several existing anti-aging drugs are under investigation with nanoparticle-based delivery agents in animal models [157,158]. Nanoparticle-based treatment delivery is believed to be effective because nanoparticles can penetrate the blood–brain barrier (BBB) efficiently and perform targeted delivery with a lower chance of crossing peripheral circulation.

8. Conclusions

Alzheimer's disease is a complex chronic disorder where genetic defects are enhanced by age, other health conditions, and environmental factors. The investigation of the genetic features of AD using modern technological approaches has allowed for a broader picture of the diagnostics of the disease. Detailed studies on the molecular biology of APP and all the related products, including secretases, have helped determine the relationship between them and how they affect the amyloidogenic process. Despite decades of research on Alzheimer's disease and APP, we are still far from a complete understanding of its biological basis. Many questions about APP biogenesis, especially the early steps, interacting partners, APP's role in mitochondria, and the potential therapeutic targets, must be addressed in future studies.

Author Contributions: A.L.K. contributed to the conceptualization; K.S.O. wrote the manuscript, designed and prepared figures; and all authors discussed and edited the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the Center of Excellence for Translational Neuroscience and Therapeutics (CTNT) and the TTUHSC Office of Research (award number CTNT-OR 2022-04 AKMM).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Alzheimer's Association Report. 2023 Alzheimer's disease facts and figures. *Alzheimers Dement.* 2023, *19*, 1598–1695. [CrossRef] [PubMed]
- Rajan, K.B.; Weuve, J.; Barnes, L.L.; McAninch, E.A.; Wilson, R.S.; Evans, D.A. Population estimate of people with clinical Alzheimer's disease and mild cognitive impairment in the United States (2020–2060). *Alzheimers Dement*. 2021, 17, 1966–1975. [CrossRef]
- Passeri, E.; Elkhoury, K.; Morsink, M.; Broersen, K.; Linder, M.; Tamayol, A.; Malaplate, C.; Yen, F.T.; Arab-Tehrany, E. Alzheimer's Disease: Treatment Strategies and Their Limitations. *Int. J. Mol. Sci.* 2022, 23, 13954. [CrossRef] [PubMed]
- 4. US Food and Drug Administration. *FDA Grants Accelerated Approval for Alzheimer's Disease Treatment;* US Food and Drug Administration: Rockville, MD, USA, 2023.
- 5. Course, M.M.; Gudsnuk, K.; Keene, C.D.; Bird, T.D.; Jayadev, S.; Valdmanis, P.N. Aberrant splicing of PSEN2, but not PSEN1, in individuals with sporadic Alzheimer's disease. *Brain* 2023, 146, 507–518. [CrossRef]
- Jia, J.; Zhang, Y.; Shi, Y.; Yin, X.; Wang, S.; Li, Y.; Zhao, T.; Liu, W.; Zhou, A.; Jia, L. A 19-Year-Old Adolescent with Probable Alzheimer's Disease. J. Alzheimers Dis. 2023, 91, 915–922. [CrossRef] [PubMed]
- Csaban, D.; Illes, A.; Renata, T.B.; Balicza, P.; Pentelenyi, K.; Molnar, V.; Gezsi, A.; Grosz, Z.; Gal, A.; Kovacs, T.; et al. Genetic landscape of early-onset dementia in Hungary. *Neurol. Sci.* 2022, 43, 5289–5300. [CrossRef] [PubMed]
- Barthélemy, N.R.; Li, Y.; Joseph-Mathurin, N.; Gordon, B.A.; Hassenstab, J.; Benzinger, T.L.S.; Buckles, V.; Fagan, A.M.; Perrin, R.J.; Goate, A.M.; et al. A soluble phosphorylated tau signature links tau, amyloid and the evolution of stages of dominantly inherited Alzheimer's disease. *Nat. Med.* 2020, 26, 398–407. [CrossRef]
- Gordon, B.A.; Blazey, T.M.; Su, Y.; Hari-Raj, A.; Dincer, A.; Flores, S.; Christensen, J.; McDade, E.; Wang, G.; Xiong, C.; et al. Spatial patterns of neuroimaging biomarker change in individuals from families with autosomal dominant Alzheimer's disease: A longitudinal study. *Lancet Neurol.* 2018, 17, 241–250. [CrossRef]
- Porsteinsson, A.P.; Isaacson, R.S.; Knox, S.; Sabbagh, M.N.; Rubino, I. Diagnosis of Early Alzheimer's Disease: Clinical Practice in 2021. J. Prev. Alzheimers Dis. 2021, 8, 371–386. [CrossRef]
- 11. Wilson, D.M.; Cookson, M.R.; Van Den Bosch, L.; Zetterberg, H.; Holtzman, D.M.; Dewachter, I. Hallmarks of neurodegenerative diseases. *Cell* **2023**, *186*, 693–714. [CrossRef]
- 12. Hardy, J.A.; Higgins, G.A. Alzheimer's disease: The amyloid cascade hypothesis. Science 1992, 256, 184–185. [CrossRef]
- Edwards, F.A. A Unifying Hypothesis for Alzheimer's Disease: From Plaques to Neurodegeneration. *Trends Neurosci.* 2019, 42, 310–322. [CrossRef]
- Busche, M.A.; Hyman, B.T. Synergy between amyloid-β and tau in Alzheimer's disease. *Nat. Neurosci.* 2020, 23, 1183–1193. [CrossRef]
- 15. Bertram, L.; McQueen, M.B.; Mullin, K.; Blacker, D.; Tanzi, R.E. Systematic meta-analyses of Alzheimer disease genetic association studies: The AlzGene database. *Nat. Genet.* 2007, *39*, 17–23. [CrossRef]
- Hollingworth, P.; Harold, D.; Sims, R.; Gerrish, A.; Lambert, J.C.; Carrasquillo, M.M.; Abraham, R.; Hamshere, M.L.; Pahwa, J.S.; Moskvina, V.; et al. Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. *Nat. Genet.* 2011, 43, 429–435. [CrossRef] [PubMed]

- Lambert, J.C.; Ibrahim-Verbaas, C.A.; Harold, D.; Naj, A.C.; Sims, R.; Bellenguez, C.; DeStafano, A.L.; Bis, J.C.; Beecham, G.W.; Grenier-Boley, B.; et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat. Genet.* 2013, 45, 1452–1458. [CrossRef] [PubMed]
- 18. Selkoe, D.J. SnapShot: Pathobiology of Alzheimer's disease. Cell 2013, 154, 468–468.e1. [CrossRef] [PubMed]
- 19. Waring, S.C.; Rosenberg, R.N. Genome-wide association studies in Alzheimer disease. Arch. Neurol. 2008, 65, 329–334. [CrossRef]
- Bellenguez, C.; Küçükali, F.; Jansen, I.E.; Kleineidam, L.; Moreno-Grau, S.; Amin, N.; Naj, A.C.; Campos-Martin, R.; Grenier-Boley, B.; Andrade, V.; et al. New insights into the genetic etiology of Alzheimer's disease and related dementias. *Nat. Genet.* 2022, 54, 412–436. [CrossRef]
- 21. Li, R.Y.; Qin, Q.; Yang, H.C.; Wang, Y.Y.; Mi, Y.X.; Yin, Y.S.; Wang, M.; Yu, C.J.; Tang, Y. TREM2 in the pathogenesis of AD: A lipid metabolism regulator and potential metabolic therapeutic target. *Mol. Neurodegener.* **2022**, *17*, 40. [CrossRef] [PubMed]
- Mishra, S.; Knupp, A.; Szabo, M.P.; Williams, C.A.; Kinoshita, C.; Hailey, D.W.; Wang, Y.; Andersen, O.M.; Young, J.E. The Alzheimer's gene SORL1 is a regulator of endosomal traffic and recycling in human neurons. *Cell. Mol. Life Sci.* 2022, 79, 162. [CrossRef] [PubMed]
- 23. Dib, S.; Pahnke, J.; Gosselet, F. Role of ABCA7 in Human Health and in Alzheimer's Disease. *Int. J. Mol. Sci.* 2021, 22, 4603. [CrossRef]
- Zhao, N.; Liu, C.C.; Qiao, W.; Bu, G. Apolipoprotein E, Receptors, and Modulation of Alzheimer's Disease. *Biol. Psychiatry* 2018, 83, 347–357. [CrossRef] [PubMed]
- Cataldo, A.M.; Peterhoff, C.M.; Troncoso, J.C.; Gomez-Isla, T.; Hyman, B.T.; Nixon, R.A. Endocytic pathway abnormalities precede amyloid beta deposition in sporadic Alzheimer's disease and Down syndrome: Differential effects of APOE genotype and presenilin mutations. *Am. J. Pathol.* 2000, 157, 277–286. [CrossRef] [PubMed]
- Lin, Y.T.; Seo, J.; Gao, F.; Feldman, H.M.; Wen, H.L.; Penney, J.; Cam, H.P.; Gjoneska, E.; Raja, W.K.; Cheng, J.; et al. APOE4 Causes Widespread Molecular and Cellular Alterations Associated with Alzheimer's Disease Phenotypes in Human iPSC-Derived Brain Cell Types. *Neuron* 2018, *98*, 1141–1154.e7. [CrossRef]
- de Leeuw, S.M.; Kirschner, A.W.T.; Lindner, K.; Rust, R.; Budny, V.; Wolski, W.E.; Gavin, A.C.; Nitsch, R.M.; Tackenberg, C. APOE2, E3, and E4 differentially modulate cellular homeostasis, cholesterol metabolism, and inflammatory response in isogenic iPSC-derived astrocytes. *Stem Cell Rep.* 2022, *17*, 110–126. [CrossRef] [PubMed]
- Wang, C.; Xiong, M.; Gratuze, M.; Bao, X.; Shi, Y.; Andhey, P.S.; Manis, M.; Schroeder, C.; Yin, Z.; Madore, C.; et al. Selective removal of astrocytic APOE4 strongly protects against tau-mediated neurodegeneration and decreases synaptic phagocytosis by microglia. *Neuron* 2021, 109, 1657–1674.e7. [CrossRef]
- 29. Zhao, J.; Fu, Y.; Yamazaki, Y.; Ren, Y.; Davis, M.D.; Liu, C.C.; Lu, W.; Wang, X.; Chen, K.; Cherukuri, Y.; et al. APOE4 exacerbates synapse loss and neurodegeneration in Alzheimer's disease patient iPSC-derived cerebral organoids. *Nat. Commun.* 2020, *11*, 5540. [CrossRef]
- 30. Tcw, J.; Qian, L.; Pipalia, N.H.; Chao, M.J.; Liang, S.A.; Shi, Y.; Jain, B.R.; Bertelsen, S.E.; Kapoor, M.; Marcora, E.; et al. Cholesterol and matrisome pathways dysregulated in astrocytes and microglia. *Cell* **2022**, *185*, 2213–2233.e25. [CrossRef]
- Lee, S.I.; Jeong, W.; Lim, H.; Cho, S.; Lee, H.; Jang, Y.; Cho, J.; Bae, S.; Lin, Y.T.; Tsai, L.H.; et al. APOE4-carrying human astrocytes oversupply cholesterol to promote neuronal lipid raft expansion and Aβ generation. *Stem Cell Rep.* 2021, 16, 2128–2137. [CrossRef]
- 32. Li, N.M.; Liu, K.F.; Qiu, Y.J.; Zhang, H.H.; Nakanishi, H.; Qing, H. Mutations of beta-amyloid precursor protein alter the consequence of Alzheimer's disease pathogenesis. *Neural Regen. Res.* **2019**, *14*, 658–665. [PubMed]
- Selkoe, D.J. Presenilin, Notch, and the genesis and treatment of Alzheimer's disease. Proc. Natl. Acad. Sci. USA 2001, 98, 11039–11041. [CrossRef]
- 34. Weggen, S.; Beher, D. Molecular consequences of amyloid precursor protein and presenilin mutations causing autosomaldominant Alzheimer's disease. *Alzheimers Res. Ther.* **2012**, *4*, 9. [CrossRef]
- Szaruga, M.; Munteanu, B.; Lismont, S.; Veugelen, S.; Horré, K.; Mercken, M.; Saido, T.C.; Ryan, N.S.; De Vos, T.; Savvides, S.N.; et al. Alzheimer's-Causing Mutations Shift Aβ Length by Destabilizing γ-Secretase-Aβn Interactions. *Cell* 2017, 170, 443–456.e14. [CrossRef]
- Sarasija, S.; Laboy, J.T.; Ashkavand, Z.; Bonner, J.; Tang, Y.; Norman, K.R. Presenilin mutations deregulate mitochondrial Ca. *Elife* 2018, 7, e33052. [CrossRef]
- Walter, P.; Blobel, G. Translocation of proteins across the endoplasmic reticulum III. Signal recognition protein (SRP) causes signal sequence-dependent and site-specific arrest of chain elongation that is released by microsomal membranes. *J. Cell Biol.* 1981, 91 Pt 1, 557–561. [CrossRef] [PubMed]
- Kellogg, M.K.; Miller, S.C.; Tikhonova, E.B.; Karamyshev, A.L. SRPassing Co-translational Targeting: The Role of the Signal Recognition Particle in Protein Targeting and mRNA Protection. *Int. J. Mol. Sci.* 2021, 22, 6284. [CrossRef]
- Kellogg, M.K.; Tikhonova, E.B.; Karamyshev, A.L. Signal Recognition Particle in Human Diseases. Front. Genet. 2022, 13, 898083. [CrossRef]
- 40. von Heijne, G. Signal sequences. The limits of variation. J. Mol. Biol. 1985, 184, 99–105. [CrossRef]
- 41. von Heijne, G. Analysis of the distribution of charged residues in the N-terminal region of signal sequences: Implications for protein export in prokaryotic and eukaryotic cells. *EMBO J.* **1984**, *3*, 2315–2318. [CrossRef] [PubMed]
- 42. von Heijne, G. Protein targeting signals. Curr. Opin. Cell Biol. 1990, 2, 604–608. [CrossRef] [PubMed]

- Nilsson, I.; Lara, P.; Hessa, T.; Johnson, A.E.; von Heijne, G.; Karamyshev, A.L. The code for directing proteins for translocation across ER membrane: SRP cotranslationally recognizes specific features of a signal sequence. J. Mol. Biol. 2015, 427 Pt A, 1191–1201. [CrossRef]
- Karamyshev, A.L.; Patrick, A.E.; Karamysheva, Z.N.; Griesemer, D.S.; Hudson, H.; Tjon-Kon-Sang, S.; Nilsson, I.; Otto, H.; Liu, Q.; Rospert, S.; et al. Inefficient SRP interaction with a nascent chain triggers a mRNA quality control pathway. *Cell* 2014, 156, 146–157. [CrossRef] [PubMed]
- 45. Karamyshev, A.L.; Tikhonova, E.B.; Karamysheva, Z.N. Translational Control of Secretory Proteins in Health and Disease. *Int. J. Mol. Sci.* 2020, *21*, 2538. [CrossRef] [PubMed]
- Tikhonova, E.B.; Karamysheva, Z.N.; von Heijne, G.; Karamyshev, A.L. Silencing of Aberrant Secretory Protein Expression by Disease-Associated Mutations. J. Mol. Biol. 2019, 431, 2567–2580. [CrossRef]
- 47. Karamyshev, A.L.; Karamysheva, Z.N. Lost in Translation: Ribosome-Associated mRNA and Protein Quality Controls. *Front. Genet.* **2018**, *9*, 431. [CrossRef] [PubMed]
- Karamysheva, Z.N.; Karamyshev, A.L. Aberrant protein targeting activates quality control on the ribosome. *Front. Cell Dev. Biol.* 2023, 11, 1198184. [CrossRef]
- Pinarbasi, E.S.; Karamyshev, A.L.; Tikhonova, E.B.; Wu, I.H.; Hudson, H.; Thomas, P.J. Pathogenic Signal Sequence Mutations in Progranulin Disrupt SRP Interactions Required for mRNA Stability. *Cell Rep.* 2018, 23, 2844–2851. [CrossRef]
- Karamysheva, Z.N.; Tikhonova, E.B.; Karamyshev, A.L. Granulin in Frontotemporal Lobar Degeneration: Molecular Mechanisms of the Disease. Front. Neurosci. 2019, 13, 395. [CrossRef]
- 51. Hernandez, S.M.; Tikhonova, E.B.; Baca, K.R.; Zhao, F.; Zhu, X.; Karamyshev, A.L. Unexpected Implication of SRP and AGO2 in Parkinson's Disease: Involvement in Alpha-Synuclein Biogenesis. *Cells* **2021**, *10*, 2792. [CrossRef]
- Tikhonova, E.B.; Gutierrez Guarnizo, S.A.; Kellogg, M.K.; Karamyshev, A.; Dozmorov, I.M.; Karamysheva, Z.N.; Karamyshev, A.L. Defective Human SRP Induces Protein Quality Control and Triggers Stress Response. J. Mol. Biol. 2022, 434, 167832. [CrossRef] [PubMed]
- 53. Gadhave, K. The signal peptide of the amyloid precursor protein forms amyloid-like aggregates and enhances Ab42 aggregation. *Cell Rep. Phys. Sci.* **2021**, *2*, 100599. [CrossRef]
- 54. Selkoe, D.J. Alzheimer's disease: Genes, proteins, and therapy. Physiol. Rev. 2001, 81, 741–766. [CrossRef]
- 55. Rimal, S.; Li, Y.; Vartak, R.; Geng, J.; Tantray, I.; Li, S.; Huh, S.; Vogel, H.; Glabe, C.; Grinberg, L.T.; et al. Inefficient quality control of ribosome stalling during APP synthesis generates CAT-tailed species that precipitate hallmarks of Alzheimer's disease. *Acta Neuropathol. Commun.* **2021**, *9*, 169. [CrossRef] [PubMed]
- 56. Simon, S.M.; Blobel, G. A protein-conducting channel in the endoplasmic reticulum. Cell 1991, 65, 371–380. [CrossRef]
- 57. Rapoport, T.A. Protein transport across the endoplasmic reticulum membrane. FEBS J. 2008, 275, 4471–4478. [CrossRef]
- 58. Ast, T.; Cohen, G.; Schuldiner, M. A network of cytosolic factors targets SRP-independent proteins to the endoplasmic reticulum. *Cell* **2013**, *152*, 1134–1145. [CrossRef]
- Liaci, A.M.; Förster, F. Take Me Home, Protein Roads: Structural Insights into Signal Peptide Interactions during ER Translocation. Int. J. Mol. Sci. 2021, 22, 11871. [CrossRef]
- 60. Goldgaber, D.; Lerman, M.I.; McBride, O.W.; Saffiotti, U.; Gajdusek, D.C. Characterization and chromosomal localization of a cDNA encoding brain amyloid of Alzheimer's disease. *Science* **1987**, 235, 877–880. [CrossRef]
- Yoshikai, S.; Sasaki, H.; Doh-ura, K.; Furuya, H.; Sakaki, Y. Genomic organization of the human amyloid beta-protein precursor gene. *Gene* 1990, 87, 257–263. [CrossRef]
- 62. Lamb, B.T.; Sisodia, S.S.; Lawler, A.M.; Slunt, H.H.; Kitt, C.A.; Kearns, W.G.; Pearson, P.L.; Price, D.L.; Gearhart, J.D. Introduction and expression of the 400 kilobase amyloid precursor protein gene in transgenic mice [corrected]. *Nat. Genet.* **1993**, *5*, 22–30. [CrossRef]
- 63. Delvaux, E.; Bentley, K.; Stubbs, V.; Sabbagh, M.; Coleman, P.D. Differential processing of amyloid precursor protein in brain and in peripheral blood leukocytes. *Neurobiol. Aging* **2013**, *34*, 1680–1686. [CrossRef]
- 64. Kang, J.; Müller-Hill, B. Differential splicing of Alzheimer's disease amyloid A4 precursor RNA in rat tissues: PreA4(695) mRNA is predominantly produced in rat and human brain. *Biochem. Biophys. Res. Commun.* 1990, 166, 1192–1200. [CrossRef]
- 65. Korte, M.; Herrmann, U.; Zhang, X.; Draguhn, A. The role of APP and APLP for synaptic transmission, plasticity, and network function: Lessons from genetic mouse models. *Exp. Brain Res.* **2012**, *217*, 435–440. [CrossRef] [PubMed]
- 66. Soba, P.; Eggert, S.; Wagner, K.; Zentgraf, H.; Siehl, K.; Kreger, S.; Löwer, A.; Langer, A.; Merdes, G.; Paro, R.; et al. Homo- and heterodimerization of APP family members promotes intercellular adhesion. *EMBO J.* **2005**, *24*, 3624–3634. [CrossRef] [PubMed]
- 67. Yamazaki, T.; Koo, E.H.; Selkoe, D.J. Cell surface amyloid beta-protein precursor colocalizes with beta 1 integrins at substrate contact sites in neural cells. *J. Neurosci.* **1997**, *17*, 1004–1010. [CrossRef] [PubMed]
- Senechal, Y.; Kelly, P.H.; Dev, K.K. Amyloid precursor protein knockout mice show age-dependent deficits in passive avoidance learning. *Behav. Brain Res.* 2008, 186, 126–132. [CrossRef]
- White, A.R.; Reyes, R.; Mercer, J.F.; Camakaris, J.; Zheng, H.; Bush, A.I.; Multhaup, G.; Beyreuther, K.; Masters, C.L.; Cappai, R. Copper levels are increased in the cerebral cortex and liver of APP and APLP2 knockout mice. *Brain Res.* 1999, 842, 439–444. [CrossRef]
- Zhang, X.; Zhong, W.; Brankačk, J.; Weyer, S.W.; Müller, U.C.; Tort, A.B.; Draguhn, A. Impaired theta-gamma coupling in APP-deficient mice. *Sci. Rep.* 2016, 6, 21948. [CrossRef]

- 71. Young-Pearse, T.L.; Chen, A.C.; Chang, R.; Marquez, C.; Selkoe, D.J. Secreted APP regulates the function of full-length APP in neurite outgrowth through interaction with integrin beta1. *Neural Dev.* **2008**, *3*, 15. [CrossRef]
- 72. Obregon, D.; Hou, H.; Deng, J.; Giunta, B.; Tian, J.; Darlington, D.; Shahaduzzaman, M.; Zhu, Y.; Mori, T.; Mattson, M.P.; et al. Soluble amyloid precursor protein-α modulates β-secretase activity and amyloid-β generation. *Nat. Commun.* 2012, *3*, 777. [CrossRef]
- 73. Ring, S.; Weyer, S.W.; Kilian, S.B.; Waldron, E.; Pietrzik, C.U.; Filippov, M.A.; Herms, J.; Buchholz, C.; Eckman, C.B.; Korte, M.; et al. The secreted beta-amyloid precursor protein ectodomain APPs alpha is sufficient to rescue the anatomical, behavioral, and electrophysiological abnormalities of APP-deficient mice. *J. Neurosci.* **2007**, *27*, 7817–7826. [CrossRef]
- 74. Farzan, M.; Schnitzler, C.E.; Vasilieva, N.; Leung, D.; Choe, H. BACE2, a beta -secretase homolog, cleaves at the beta site and within the amyloid-beta region of the amyloid-beta precursor protein. *Proc. Natl. Acad. Sci. USA* 2000, 97, 9712–9717. [CrossRef] [PubMed]
- 75. Laird, F.M.; Cai, H.; Savonenko, A.V.; Farah, M.H.; He, K.; Melnikova, T.; Wen, H.; Chiang, H.C.; Xu, G.; Koliatsos, V.E.; et al. BACE1, a major determinant of selective vulnerability of the brain to amyloid-beta amyloidogenesis, is essential for cognitive, emotional, and synaptic functions. *J. Neurosci.* 2005, 25, 11693–11709. [CrossRef] [PubMed]
- 76. Ohno, M.; Chang, L.; Tseng, W.; Oakley, H.; Citron, M.; Klein, W.L.; Vassar, R.; Disterhoft, J.F. Temporal memory deficits in Alzheimer's mouse models: Rescue by genetic deletion of BACE1. *Eur. J. Neurosci.* **2006**, *23*, 251–260. [CrossRef] [PubMed]
- 77. Ohno, M.; Sametsky, E.A.; Younkin, L.H.; Oakley, H.; Younkin, S.G.; Citron, M.; Vassar, R.; Disterhoft, J.F. BACE1 deficiency rescues memory deficits and cholinergic dysfunction in a mouse model of Alzheimer's disease. *Neuron* 2004, 41, 27–33. [CrossRef] [PubMed]
- 78. Araki, W. Post-translational regulation of the β-secretase BACE1. Brain Res. Bull. 2016, 126 Pt 2, 170–177. [CrossRef] [PubMed]
- Andrew, R.J.; Fernandez, C.G.; Stanley, M.; Jiang, H.; Nguyen, P.; Rice, R.C.; Buggia-Prévot, V.; De Rossi, P.; Vetrivel, K.S.; Lamb, R.; et al. Lack of BACE1 S-palmitoylation reduces amyloid burden and mitigates memory deficits in transgenic mouse models of Alzheimer's disease. *Proc. Natl. Acad. Sci. USA* 2017, *114*, E9665–E9674. [CrossRef] [PubMed]
- Huse, J.T.; Liu, K.; Pijak, D.S.; Carlin, D.; Lee, V.M.; Doms, R.W. Beta-secretase processing in the trans-Golgi network preferentially generates truncated amyloid species that accumulate in Alzheimer's disease brain. *J. Biol. Chem.* 2002, 277, 16278–16284. [CrossRef]
- Schneider, A.; Rajendran, L.; Honsho, M.; Gralle, M.; Donnert, G.; Wouters, F.; Hell, S.W.; Simons, M. Flotillin-dependent clustering of the amyloid precursor protein regulates its endocytosis and amyloidogenic processing in neurons. *J. Neurosci.* 2008, 28, 2874–2882. [CrossRef]
- Sannerud, R.; Declerck, I.; Peric, A.; Raemaekers, T.; Menendez, G.; Zhou, L.; Veerle, B.; Coen, K.; Munck, S.; De Strooper, B.; et al. ADP ribosylation factor 6 (ARF6) controls amyloid precursor protein (APP) processing by mediating the endosomal sorting of BACE1. *Proc. Natl. Acad. Sci. USA* 2011, 108, E559–E568. [CrossRef] [PubMed]
- Sakurai, T.; Kaneko, K.; Okuno, M.; Wada, K.; Kashiyama, T.; Shimizu, H.; Akagi, T.; Hashikawa, T.; Nukina, N. Membrane microdomain switching: A regulatory mechanism of amyloid precursor protein processing. J. Cell Biol. 2008, 183, 339–352. [CrossRef]
- Bukhari, H.; Glotzbach, A.; Kolbe, K.; Leonhardt, G.; Loosse, C.; Müller, T. Small things matter: Implications of APP intracellular domain AICD nuclear signaling in the progression and pathogenesis of Alzheimer's disease. *Prog. Neurobiol.* 2017, 156, 189–213. [CrossRef] [PubMed]
- 85. Greenwood, E.K.; Angelova, D.M.; Büchner, H.M.I.; Brown, D.R. The AICD fragment of APP initiates a FoxO3a mediated response via FANCD2. *Mol. Cell. Neurosci.* 2022, 122, 103760. [CrossRef] [PubMed]
- Zhou, D.; Noviello, C.; D'Ambrosio, C.; Scaloni, A.; D'Adamio, L. Growth factor receptor-bound protein 2 interaction with the tyrosine-phosphorylated tail of amyloid beta precursor protein is mediated by its Src homology 2 domain. *J. Biol. Chem.* 2004, 279, 25374–25380. [CrossRef]
- 87. Chen, W.J.; Goldstein, J.L.; Brown, M.S. NPXY, a sequence often found in cytoplasmic tails, is required for coated pit-mediated internalization of the low density lipoprotein receptor. *J. Biol. Chem.* **1990**, *265*, 3116–3123. [CrossRef]
- Vieira, S.I.; Rebelo, S.; Domingues, S.C.; da Cruz e Silva, E.F.; da Cruz e Silva, O.A. S655 phosphorylation enhances APP secretory traffic. *Mol. Cell. Biochem.* 2009, 328, 145–154. [CrossRef]
- Vieira, S.I.; Rebelo, S.; Esselmann, H.; Wiltfang, J.; Lah, J.; Lane, R.; Small, S.A.; Gandy, S.; da Cruz e Silva, E.F.; da Cruz e Silva, O.A. Retrieval of the Alzheimer's amyloid precursor protein from the endosome to the TGN is S655 phosphorylation state-dependent and retromer-mediated. *Mol. Neurodegener.* 2010, *5*, 40. [CrossRef]
- Jiang, S.; Li, Y.; Zhang, X.; Bu, G.; Xu, H.; Zhang, Y.W. Trafficking regulation of proteins in Alzheimer's disease. *Mol. Neurodegener.* 2014, 9, 6. [CrossRef]
- 91. Lee, M.S.; Kao, S.C.; Lemere, C.A.; Xia, W.; Tseng, H.C.; Zhou, Y.; Neve, R.; Ahlijanian, M.K.; Tsai, L.H. APP processing is regulated by cytoplasmic phosphorylation. *J. Cell Biol.* **2003**, *163*, 83–95. [CrossRef]
- 92. Suzuki, T.; Nakaya, T. Regulation of amyloid beta-protein precursor by phosphorylation and protein interactions. *J. Biol. Chem.* **2008**, *283*, 29633–29637. [CrossRef]
- 93. De Strooper, B. Aph-1, Pen-2, and Nicastrin with Presenilin generate an active gamma-Secretase complex. *Neuron* **2003**, *38*, 9–12. [CrossRef]

- Krishnaswamy, S.; Verdile, G.; Groth, D.; Kanyenda, L.; Martins, R.N. The structure and function of Alzheimer's gamma secretase enzyme complex. *Crit. Rev. Clin. Lab. Sci.* 2009, 46, 282–301. [CrossRef]
- 95. Haapasalo, A.; Kovacs, D.M. The many substrates of presenilin/γ-secretase. J. Alzheimers Dis. 2011, 25, 3–28. [CrossRef] [PubMed]
- 96. Beel, A.J.; Sanders, C.R. Substrate specificity of gamma-secretase and other intramembrane proteases. *Cell. Mol. Life Sci.* 2008, 65, 1311–1334. [CrossRef] [PubMed]
- Kanatsu, K.; Tomita, T. Membrane trafficking and proteolytic activity of γ-secretase in Alzheimer's disease. *Biol. Chem.* 2016, 397, 827–835. [CrossRef]
- Hansson, C.A.; Frykman, S.; Farmery, M.R.; Tjernberg, L.O.; Nilsberth, C.; Pursglove, S.E.; Ito, A.; Winblad, B.; Cowburn, R.F.; Thyberg, J.; et al. Nicastrin, presenilin, APH-1, and PEN-2 form active gamma-secretase complexes in mitochondria. *J. Biol. Chem.* 2004, 279, 51654–51660. [CrossRef] [PubMed]
- 99. Sisodia, S.S. Beta-amyloid precursor protein cleavage by a membrane-bound protease. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 6075–6079. [CrossRef]
- Carey, R.M.; Balcz, B.A.; Lopez-Coviella, I.; Slack, B.E. Inhibition of dynamin-dependent endocytosis increases shedding of the amyloid precursor protein ectodomain and reduces generation of amyloid beta protein. BMC Cell Biol. 2005, 6, 30. [CrossRef]
- Goodger, Z.V.; Rajendran, L.; Trutzel, A.; Kohli, B.M.; Nitsch, R.M.; Konietzko, U. Nuclear signaling by the APP intracellular domain occurs predominantly through the amyloidogenic processing pathway. J. Cell Sci. 2009, 122 Pt 20, 3703–3714. [CrossRef]
- 102. Campioni, S.; Mannini, B.; Zampagni, M.; Pensalfini, A.; Parrini, C.; Evangelisti, E.; Relini, A.; Stefani, M.; Dobson, C.M.; Cecchi, C.; et al. A causative link between the structure of aberrant protein oligomers and their toxicity. *Nat. Chem. Biol.* 2010, *6*, 140–147. [CrossRef]
- 103. Fitzpatrick, A.W.; Debelouchina, G.T.; Bayro, M.J.; Clare, D.K.; Caporini, M.A.; Bajaj, V.S.; Jaroniec, C.P.; Wang, L.; Ladizhansky, V.; Müller, S.A.; et al. Atomic structure and hierarchical assembly of a cross-β amyloid fibril. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 5468–5473. [CrossRef]
- 104. Gu, L.; Liu, C.; Guo, Z. Structural insights into Aβ42 oligomers using site-directed spin labeling. *J. Biol. Chem.* **2013**, 288, 18673–18683. [CrossRef]
- 105. Sawaya, M.R.; Sambashivan, S.; Nelson, R.; Ivanova, M.I.; Sievers, S.A.; Apostol, M.I.; Thompson, M.J.; Balbirnie, M.; Wiltzius, J.J.; McFarlane, H.T.; et al. Atomic structures of amyloid cross-beta spines reveal varied steric zippers. *Nature* 2007, 447, 453–457. [CrossRef] [PubMed]
- 106. Scheidt, H.A.; Morgado, I.; Rothemund, S.; Huster, D. Dynamics of amyloid β fibrils revealed by solid-state NMR. *J. Biol. Chem.* 2012, 287, 2017–2021. [CrossRef] [PubMed]
- 107. Eisenberg, D.; Jucker, M. The amyloid state of proteins in human diseases. Cell 2012, 148, 1188–1203. [CrossRef]
- 108. Chen, J.; Armstrong, A.H.; Koehler, A.N.; Hecht, M.H. Small molecule microarrays enable the discovery of compounds that bind the Alzheimer's Aβ peptide and reduce its cytotoxicity. *J. Am. Chem. Soc.* **2010**, *132*, 17015–17022. [CrossRef]
- 109. Zhang, C.; Liu, Y.; Gilthorpe, J.; van der Maarel, J.R. MRP14 (S100A9) protein interacts with Alzheimer beta-amyloid peptide and induces its fibrillization. *PLoS ONE* 2012, *7*, e32953. [CrossRef]
- 110. Camandola, S.; Mattson, M.P. Aberrant subcellular neuronal calcium regulation in aging and Alzheimer's disease. *Biochim. Biophys. Acta* 2011, 1813, 965–973. [CrossRef]
- Demuro, A.; Smith, M.; Parker, I. Single-channel Ca(2+) imaging implicates Aβ1-42 amyloid pores in Alzheimer's disease pathology. J. Cell Biol. 2011, 195, 515–524. [CrossRef] [PubMed]
- 112. Johri, A.; Beal, M.F. Mitochondrial dysfunction in neurodegenerative diseases. J. Pharmacol. Exp. Ther. 2012, 342, 619–630. [CrossRef] [PubMed]
- 113. Moon, H.E.; Paek, S.H. Mitochondrial Dysfunction in Parkinson's Disease. Exp. Neurobiol. 2015, 24, 103–116. [CrossRef] [PubMed]
- 114. Anandatheerthavarada, H.K.; Biswas, G.; Mullick, J.; Sepuri, N.B.; Otvos, L.; Pain, D.; Avadhani, N.G. Dual targeting of cytochrome P4502B1 to endoplasmic reticulum and mitochondria involves a novel signal activation by cyclic AMP-dependent phosphorylation at ser128. *EMBO J.* **1999**, *18*, 5494–5504. [CrossRef] [PubMed]
- Robin, M.A.; Anandatheerthavarada, H.K.; Biswas, G.; Sepuri, N.B.; Gordon, D.M.; Pain, D.; Avadhani, N.G. Bimodal targeting of microsomal CYP2E1 to mitochondria through activation of an N-terminal chimeric signal by cAMP-mediated phosphorylation. *J. Biol. Chem.* 2002, 277, 40583–40593. [CrossRef]
- Devi, L.; Prabhu, B.M.; Galati, D.F.; Avadhani, N.G.; Anandatheerthavarada, H.K. Accumulation of amyloid precursor protein in the mitochondrial import channels of human Alzheimer's disease brain is associated with mitochondrial dysfunction. *J. Neurosci.* 2006, 26, 9057–9068. [CrossRef]
- 117. Vaillant-Beuchot, L.; Mary, A.; Pardossi-Piquard, R.; Bourgeois, A.; Lauritzen, I.; Eysert, F.; Kinoshita, P.F.; Cazareth, J.; Badot, C.; Fragaki, K.; et al. Accumulation of amyloid precursor protein C-terminal fragments triggers mitochondrial structure, function, and mitophagy defects in Alzheimer's disease models and human brains. *Acta Neuropathol.* 2021, 141, 39–65. [CrossRef]
- 118. Mossmann, D.; Vögtle, F.N.; Taskin, A.A.; Teixeira, P.F.; Ring, J.; Burkhart, J.M.; Burger, N.; Pinho, C.M.; Tadic, J.; Loreth, D.; et al. Amyloid-β peptide induces mitochondrial dysfunction by inhibition of preprotein maturation. *Cell Metab.* 2014, 20, 662–669. [CrossRef]
- 119. Sorrentino, V.; Romani, M.; Mouchiroud, L.; Beck, J.S.; Zhang, H.; D'Amico, D.; Moullan, N.; Potenza, F.; Schmid, A.W.; Rietsch, S.; et al. Enhancing mitochondrial proteostasis reduces amyloid-β proteotoxicity. *Nature* 2017, 552, 187–193. [CrossRef]

- Calvo-Rodriguez, M.; Bacskai, B.J. Mitochondria and Calcium in Alzheimer's Disease: From Cell Signaling to Neuronal Cell Death. *Trends Neurosci.* 2021, 44, 136–151. [CrossRef]
- 121. Strope, T.A.; Wilkins, H.M. Amyloid precursor protein and mitochondria. Curr. Opin. Neurobiol. 2023, 78, 102651. [CrossRef]
- 122. Lykhmus, O.; Koval, L.; Voytenko, L.; Uspenska, K.; Komisarenko, S.; Deryabina, O.; Shuvalova, N.; Kordium, V.; Ustymenko, A.; Kyryk, V.; et al. Intravenously Injected Mesenchymal Stem Cells Penetrate the Brain and Treat Inflammation-Induced Brain Damage and Memory Impairment in Mice. *Front. Pharmacol.* 2019, 10, 355. [CrossRef]
- 123. Xu, F.; Wu, Y.; Yang, Q.; Cheng, Y.; Xu, J.; Zhang, Y.; Dai, H.; Wang, B.; Ma, Q.; Chen, Y.; et al. Engineered Extracellular Vesicles with SHP2 High Expression Promote Mitophagy for Alzheimer's Disease Treatment. *Adv. Mater.* 2022, 34, e2207107. [CrossRef] [PubMed]
- 124. Yin, T.; Liu, Y.; Ji, W.; Zhuang, J.; Chen, X.; Gong, B.; Chu, J.; Liang, W.; Gao, J.; Yin, Y. Engineered mesenchymal stem cell-derived extracellular vesicles: A state-of-the-art multifunctional weapon against Alzheimer's disease. *Theranostics* 2023, 13, 1264–1285. [CrossRef]
- Vamecq, J.; Latruffe, N. Medical significance of peroxisome proliferator-activated receptors. *Lancet* 1999, 354, 141–148. [CrossRef]
 [PubMed]
- 126. Luo, R.; Su, L.Y.; Li, G.; Yang, J.; Liu, Q.; Yang, L.X.; Zhang, D.F.; Zhou, H.; Xu, M.; Fan, Y.; et al. Activation of PPARA-mediated autophagy reduces Alzheimer disease-like pathology and cognitive decline in a murine model. *Autophagy* 2020, 16, 52–69. [CrossRef] [PubMed]
- 127. Sastre, M.; Dewachter, I.; Rossner, S.; Bogdanovic, N.; Rosen, E.; Borghgraef, P.; Evert, B.O.; Dumitrescu-Ozimek, L.; Thal, D.R.; Landreth, G.; et al. Nonsteroidal anti-inflammatory drugs repress beta-secretase gene promoter activity by the activation of PPARgamma. *Proc. Natl. Acad. Sci. USA* 2006, 103, 443–448. [CrossRef] [PubMed]
- Chen, X.F.; Zhang, Y.W.; Xu, H.; Bu, G. Transcriptional regulation and its misregulation in Alzheimer's disease. *Mol. Brain* 2013, *6*, 44. [CrossRef] [PubMed]
- 129. Christensen, M.A.; Zhou, W.; Qing, H.; Lehman, A.; Philipsen, S.; Song, W. Transcriptional regulation of BACE1, the beta-amyloid precursor protein beta-secretase, by Sp1. *Mol. Cell. Biol.* **2004**, *24*, 865–874. [CrossRef]
- 130. Hirano, F.; Tanaka, H.; Hirano, Y.; Hiramoto, M.; Handa, H.; Makino, I.; Scheidereit, C. Functional interference of Sp1 and NF-kappaB through the same DNA binding site. *Mol. Cell. Biol.* **1998**, *18*, 1266–1274. [CrossRef] [PubMed]
- 131. Rossello, X.S.; Igbavboa, U.; Weisman, G.A.; Sun, G.Y.; Wood, W.G. AP-2β regulates amyloid beta-protein stimulation of apolipoprotein E transcription in astrocytes. *Brain Res.* **2012**, *1444*, 87–95. [CrossRef]
- Das, H.K. Transcriptional regulation of the presenilin-1 gene: Implication in Alzheimer's disease. Front. Biosci. 2008, 13, 822–832.
 [CrossRef]
- 133. Pastorcic, M.; Das, H.K. Regulation of transcription of the human presenilin-1 gene by ets transcription factors and the p53 protooncogene. *J. Biol. Chem.* **2000**, 275, 34938–34945. [CrossRef]
- 134. Pastorcic, M.; Das, H.K. Analysis of transcriptional modulation of the presenilin 1 gene promoter by ZNF237, a candidate binding partner of the Ets transcription factor ERM. *Brain Res.* 2007, *1128*, 21–32. [CrossRef]
- Hwang, E.M.; Kim, S.K.; Sohn, J.H.; Lee, J.Y.; Kim, Y.; Kim, Y.S.; Mook-Jung, I. Furin is an endogenous regulator of alpha-secretase associated APP processing. *Biochem. Biophys. Res. Commun.* 2006, 349, 654–659. [CrossRef]
- Zhang, Y.; Bai, X.; Yao, S.; Cui, Y.; You, L.H.; Yu, P.; Chang, Y.Z.; Gao, G. Hippocampal Iron Accumulation Impairs Synapses and Memory via Suppressing Furin Expression and Downregulating BDNF Maturation. *Mol. Neurobiol.* 2022, 59, 5574–5590. [CrossRef]
- 137. Beckelman, B.C.; Yang, W.; Kasica, N.P.; Zimmermann, H.R.; Zhou, X.; Keene, C.D.; Ryazanov, A.G.; Ma, T. Genetic reduction of eEF2 kinase alleviates pathophysiology in Alzheimer's disease model mice. *J. Clin. Investig.* **2019**, *129*, 820–833. [CrossRef]
- Zhang, N.; Yu, X.; Xie, J.; Xu, H. New Insights into the Role of Ferritin in Iron Homeostasis and Neurodegenerative Diseases. *Mol. Neurobiol.* 2021, 58, 2812–2823. [CrossRef]
- Altamura, S.; Muckenthaler, M.U. Iron toxicity in diseases of aging: Alzheimer's disease, Parkinson's disease and atherosclerosis. J. Alzheimers Dis. 2009, 16, 879–895. [CrossRef]
- Rogers, J.T.; Randall, J.D.; Cahill, C.M.; Eder, P.S.; Huang, X.; Gunshin, H.; Leiter, L.; McPhee, J.; Sarang, S.S.; Utsuki, T.; et al. An iron-responsive element type II in the 5'-untranslated region of the Alzheimer's amyloid precursor protein transcript. *J. Biol. Chem.* 2002, 277, 45518–45528. [CrossRef]
- 141. Hetz, C. The unfolded protein response: Controlling cell fate decisions under ER stress and beyond. *Nat. Rev. Mol. Cell Biol.* **2012**, 13, 89–102. [CrossRef]
- 142. Cozachenco, D.; Ribeiro, F.C.; Ferreira, S.T. Defective proteostasis in Alzheimer's disease. *Ageing Res. Rev.* 2023, *85*, 101862. [CrossRef]
- 143. Hosokawa, N.; Wada, I.; Hasegawa, K.; Yorihuzi, T.; Tremblay, L.O.; Herscovics, A.; Nagata, K. A novel ER alpha-mannosidase-like protein accelerates ER-associated degradation. *EMBO Rep.* 2001, 2, 415–422. [CrossRef]
- 144. Nowakowska-Gołacka, J.; Czapiewska, J.; Sominka, H.; Sowa-Rogozińska, N.; Słomińska-Wojewódzka, M. EDEM1 Regulates Amyloid Precursor Protein (APP) Metabolism and Amyloid-β Production. *Int. J. Mol. Sci.* **2021**, 23, 117. [CrossRef]
- 145. Joshi, G.; Wang, Y. Golgi defects enhance APP amyloidogenic processing in Alzheimer's disease. *Bioessays* 2015, 37, 240–247. [CrossRef]

- 146. Joshi, G.; Chi, Y.; Huang, Z.; Wang, Y. Aβ-induced Golgi fragmentation in Alzheimer's disease enhances Aβ production. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, E1230–E1239. [CrossRef]
- 147. Teunissen, C.E.; Verberk, I.M.W.; Thijssen, E.H.; Vermunt, L.; Hansson, O.; Zetterberg, H.; van der Flier, W.M.; Mielke, M.M.; Del Campo, M. Blood-based biomarkers for Alzheimer's disease: Towards clinical implementation. *Lancet Neurol.* 2022, 21, 66–77. [CrossRef]
- 148. Hansson, O. Biomarkers for neurodegenerative diseases. Nat. Med. 2021, 27, 954–963. [CrossRef]
- 149. Hansson, O.; Edelmayer, R.M.; Boxer, A.L.; Carrillo, M.C.; Mielke, M.M.; Rabinovici, G.D.; Salloway, S.; Sperling, R.; Zetterberg, H.; Teunissen, C.E. The Alzheimer's Association appropriate use recommendations for blood biomarkers in Alzheimer's disease. *Alzheimers Dement.* 2022, 18, 2669–2686. [CrossRef] [PubMed]
- 150. Söderberg, L.; Johannesson, M.; Nygren, P.; Laudon, H.; Eriksson, F.; Osswald, G.; Möller, C.; Lannfelt, L. Lecanemab, Aducanumab, and Gantenerumab—Binding Profiles to Different Forms of Amyloid-Beta Might Explain Efficacy and Side Effects in Clinical Trials for Alzheimer's Disease. *Neurotherapeutics* **2023**, *20*, 195–206. [CrossRef] [PubMed]
- 151. Jia, Y.; Cao, N.; Zhai, J.; Zeng, Q.; Zheng, P.; Su, R.; Liao, T.; Liu, J.; Pei, H.; Fan, Z.; et al. HGF Mediates Clinical-Grade Human Umbilical Cord-Derived Mesenchymal Stem Cells Improved Functional Recovery in a Senescence-Accelerated Mouse Model of Alzheimer's Disease. Adv. Sci. 2020, 7, 1903809. [CrossRef] [PubMed]
- Neves, A.F.; Camargo, C.; Premer, C.; Hare, J.M.; Baumel, B.S.; Pinto, M. Intravenous administration of mesenchymal stem cells reduces Tau phosphorylation and inflammation in the 3xTg-AD mouse model of Alzheimer's disease. *Exp. Neurol.* 2021, 341, 113706. [CrossRef]
- 153. Yang, H.; Yue, C.; Xie, Z.; Hu, H.; Wei, L.; Wang, P.; Zhao, C.; Bi, J. Intravenous Administration of Human Umbilical Cord Mesenchymal Stem Cells Improves Cognitive Impairments and Reduces Amyloid-Beta Deposition in an AβPP/PS1 Transgenic Mouse Model. *Neurochem. Res.* 2013, 38, 2474–2482. [CrossRef] [PubMed]
- 154. Lim, J.Y.; In Park, S.; Park, S.A.; Jeon, J.H.; Jung, H.Y.; Yon, J.M.; Jeun, S.S.; Lim, H.K.; Kim, S.W. Potential application of human neural crest-derived nasal turbinate stem cells for the treatment of neuropathology and impaired cognition in models of Alzheimer's disease. *Stem Cell Res. Ther.* 2021, 12, 402. [CrossRef] [PubMed]
- 155. Lim, J.Y.; Lee, J.E.; Park, S.A.; Park, S.I.; Yon, J.M.; Park, J.A.; Jeun, S.S.; Kim, S.J.; Lee, H.J.; Kim, S.W.; et al. Protective Effect of Human-Neural-Crest-Derived Nasal Turbinate Stem Cells against Amyloid-β Neurotoxicity through Inhibition of Osteopontin in a Human Cerebral Organoid Model of Alzheimer's Disease. *Cells* 2022, *11*, 1029. [CrossRef]
- 156. Zhang, H.A.; Yuan, C.X.; Liu, K.F.; Yang, Q.F.; Zhao, J.; Li, H.; Yang, Q.H.; Song, D.; Quan, Z.Z.; Qing, H. Neural stem cell transplantation alleviates functional cognitive deficits in a mouse model of tauopathy. *Neural Regen. Res.* 2022, 17, 152–162. [PubMed]
- 157. Chu, J.J.; Ji, W.B.; Zhuang, J.H.; Gong, B.F.; Chen, X.H.; Cheng, W.B.; Liang, W.D.; Li, G.R.; Gao, J.; Yin, Y. Nanoparticles-based anti-aging treatment of Alzheimer's disease. *Drug Deliv.* 2022, 29, 2100–2116. [CrossRef]
- 158. Zhong, G.; Long, H.; Zhou, T.; Liu, Y.; Zhao, J.; Han, J.; Yang, X.; Yu, Y.; Chen, F.; Shi, S. Blood-brain barrier Permeable nanoparticles for Alzheimer's disease treatment by selective mitophagy of microglia. *Biomaterials* **2022**, *288*, 121690. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.