



Review

# Deciphering the Mysterious Relationship between the Cross-Pathogenetic Mechanisms of Neurodegenerative and Oncological Diseases

Yulia Aleksandrova <sup>1</sup> and Margarita Neganova <sup>1,2,\*</sup>

<sup>1</sup> Institute of Physiologically Active Compounds at Federal Research Center of Problems of Chemical Physics and Medicinal Chemistry, Russian Academy of Sciences, 142432 Chernogolovka, Russia; aleksandrova@ipac.ac.ru

<sup>2</sup> Arbusov Institute of Organic and Physical Chemistry, FRC Kazan Scientific Center, Russian Academy of Sciences, 420088 Kazan, Russia

\* Correspondence: neganovam@ipac.ac.ru; Tel.: +7-(496)-524-26-06

**Abstract:** The relationship between oncological pathologies and neurodegenerative disorders is extremely complex and is a topic of concern among a growing number of researchers around the world. In recent years, convincing scientific evidence has accumulated that indicates the contribution of a number of etiological factors and pathophysiological processes to the pathogenesis of these two fundamentally different diseases, thus demonstrating an intriguing relationship between oncology and neurodegeneration. In this review, we establish the general links between three intersecting aspects of oncological pathologies and neurodegenerative disorders, i.e., oxidative stress, epigenetic dysregulation, and metabolic dysfunction, examining each process in detail to establish an unusual epidemiological relationship. We also focus on reviewing the current trends in the research and the clinical application of the most promising chemical structures and therapeutic platforms that have a modulating effect on the above processes. Thus, our comprehensive analysis of the set of molecular determinants that have obvious cross-functional pathways in the pathogenesis of oncological and neurodegenerative diseases can help in the creation of advanced diagnostic tools and in the development of innovative pharmacological strategies.

**Keywords:** neurodegenerative diseases; Alzheimer's disease; cancer; molecular mechanisms of pathogenesis; oxidative stress; epigenetics; metabolism; drugs

**Citation:** Aleksandrova, Y.; Neganova, M. Deciphering the Mysterious Relationship between the Cross-Pathogenetic Mechanisms of Neurodegenerative and Oncological Diseases. *Int. J. Mol. Sci.* **2023**, *24*, 14766. <https://doi.org/10.3390/ijms241914766>

Academic Editor: Cristoforo Comi

Received: 10 August 2023

Revised: 22 September 2023

Accepted: 28 September 2023

Published: 29 September 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Studies conducted over the past three decades show that the treatment of oncological diseases and neurodegenerative disorders remains very limited [1,2]. This is primarily due to the numerous adverse effects of the existing drugs as a result of their systemic action due to the lack of knowledge about the etiopathogenesis of these disorders. As a result, in recent years, there has been a fundamental revision of the currently prevailing traditional therapeutic paradigms with the aim of developing the most promising pharmacological approaches.

Despite the fact that malignant neoplasms and neurodegenerative disorders are considered to be two completely different groups of diseases, and their epidemiological association is extremely complex, in both cases, the mechanisms of cellular regulation are disturbed. Moreover, if the occurrence and progression of cancers are associated with uncontrolled cell proliferation [3], then dementia, on the contrary, is characterized by brain atrophy as a result of extensive neuronal death [4].

The growing interest in the problem of determining the relationship between oncological and neurodegenerative diseases in order to find effective therapeutic agents has

allowed us to identify a number of common pathological cascades. These are associated with the dysregulation of signaling pathways and changes in the expressions of genes and proteins [5]. Such changes mainly occur in antagonistic directions. For example, long before the first symptoms of Alzheimer's disease appear in non-neuronal brain cells and on the periphery, there is an overexpression and aberrant activity of the incorrectly folded protein p53 [6], while its functional insufficiency is associated with tumor transformation and the progression of a wide range of malignant neoplasms [7]. Meanwhile, the reduced level of the apoptosis regulator Bcl-2 observed in neurodegenerative diseases [8,9] reflects the vulnerability of neuronal cells to death [10]. This phenomenon is the opposite to the overexpression of the anti-apoptotic protein in oncological diseases [11,12], leading to the increased survival of tumor cells and the development of their chemo- and radioresistance [13–15].

However, special attention should be paid to the processes involved in the etiopathogenesis of the diseases, which have significant pathophysiological correlations with each other. The list of such processes is dominated by oxidative stress and mitochondrial dysfunction [16–21], alterations in the bioenergetic cell metabolism [22,23], and epigenetic dysregulation [24,25]. To confirm this, an inverse correlation between cancers and neurodegeneration may be considered [26] when the prevalence of malignant neoplasms is significantly reduced in people with various forms of neurodegenerative disorders receiving maintenance therapy (approximately 70%) [27]. Meanwhile, among patients with a history of successfully cured cancer, there is a reduction in the risk of developing Alzheimer's disease (approximately 50%) [27], Parkinson's disease [28], and other dementia syndromes.

In this regard, having analyzed many experimental and review works conducted by leading researchers worldwide, we attempted to establish complex biological links between the pathogenesis of malignant neoplasms and neurodegenerative disorders. Since the data on the general mechanisms of the development of these diseases suggest the introduction of new therapeutic strategies, we also provided a detailed definition of potential targets for the action of promising therapeutic agents. All this can lead to a better understanding of their etiopathological mechanisms, as well as contribute to the discovery of promising new areas in the development of effective pharmacological strategies.

## **2. Oxidative Stress and Mitochondrial Dysfunction in Oncological and Neurodegenerative Diseases**

### *2.1. Concept of Oxidative Stress and Sources of Free Radicals*

For almost four decades, in the research community in experimental and clinical medicine, there has been a genuine interest in the role of reactive oxygen species (ROS) in the treatment of various diseases. Among the most important are both radical molecules (hydroxyl radicals ( $\text{OH}^\bullet$ ), superoxide anion radicals ( $\text{O}_2^{\bullet-}$ ), and peroxy radicals ( $\text{ROO}^\bullet$ ,  $\text{RCOO}^\bullet$ )), and non-radical molecules (hydrogen peroxide ( $\text{H}_2\text{O}_2$ ), singlet oxygen ( $^1\text{O}_2$ ), nitric oxide (NO), and peroxynitrite ( $\text{ONOO}^-$ )) [29].

The initial integration of reactive oxygen species into biomedical concepts was associated exclusively with their toxic effects and irreversible functional changes in various disorders. Since then, knowledge of the cell redox balance has developed rapidly. Nowadays, the dual role of reactive oxygen species in the functioning of live systems is well known. Acting as secondary messengers, reactive oxygen species at the basal level (in low or moderate concentrations) regulate the flow of numerous intracellular signaling cascades [30–33]. In turn, their overproduction and cumulative production in biological systems as a result of endogenous or exogenous effects are toxic. It makes ROS important mediators of nonspecific damage to cellular structures (known as oxidative stress), including proteins, lipids, carbohydrates, and nucleic acids [34]. In other words, the concept of oxidative stress includes disturbances in the homeostasis of free radicals that occur as a

result of the overproduction of reactive oxygen species and a deficiency of antioxidant enzymes that can detoxify reactive intermediates and repair damage [35].

The formation of reactive oxygen species is a consequence of oxidative phosphorylation, and mitochondrial ROS play a crucial role in a number of redox signaling processes and in the formation and progression of various pathological conditions. It is believed that mitochondria are the main contributors to the intracellular production of reactive oxygen species [36].

Since the delicate balance between the levels of free radicals is a key aspect in the normal functioning of the body, the processes related to oxidative stress are considered to be optimal therapeutic targets for the action of potential drug agents.

## 2.2. Free Radical Theory of the Occurrence of a Pathological Condition in a Cell

Currently, there are many theories and hypotheses that attempt to explain the driving forces behind the onset and progression of malignant neoplasms and neurodegenerative disorders. One of the most popular and fundamental postulates is the free radical theory, which interprets these pathological processes at the molecular level.

The free radical theory was first suggested in 1956 by Denham Harman to explain the attack of cellular components by free radicals produced by mitochondria as byproducts during normal metabolism [37]. It is now known that these organelles are one of the key sources of reactive oxygen species formed as a result of electron leakage in the electron transport chain [38].

Owing to ongoing discoveries in this area, 16 years later, Harman made an important statement, arguing that the lifespans of mammals depend on the rate of oxygen utilization and proposing a mitochondrial-free radical theory of aging [37]. Thus, the main postulate of Harman's theory states that "the prolonged presence of these unstable and reactive molecules in the system can lead to a direct or indirect damage to cellular components and connective tissues" [37]. Since then, Harman's theory has led to the idea that the removal of such molecules will reduce cell damage and, as a result, can slow down the pathological process. Much effort has been made to verify the validity of this position, and, to date, it has been confirmed in a large number of works demonstrating the abilities of antioxidant therapy, leading to an increase in the lifespan of model organisms [39].

It is well known that the course of various types of neurodegenerative disorders is accompanied by progressive modifications or the deterioration of brain metabolism. One of the main causes of this phenomenon, oxidative stress, affects a number of metabolic pathways [40], and evidence of the involvement of free radicals in cognitive impairment has been obtained from patients suffering from neurodegeneration. In Alzheimer's disease, redox-mediated damage to various biomolecules has often been reported [41–52]. Thus, in the search for the mechanisms underlying this disorder, in addition to the dominant hypothesis of the amyloid cascade, an alternative explanation for the pathogenesis of this disease was proposed, which consisted of the relationship between mitochondrial dysfunction and the hyperproduction of reactive oxygen species. These signs are commonly exhibited in patients with this neuropathology [41]. High levels of biomarkers of oxidative stress in blood plasma and urine have been found in both animal models [42–47] and patients with Alzheimer's disease [48–51], which correlates with the aggregation and deposits of  $\beta$ -amyloid. In addition, a proteomic study has shown that the higher risk of developing neurodegenerative disorders in patients with Down syndrome can be explained by their higher susceptibility to damage caused by oxidative stress [52].

Oxidative damage to lipids plays a key role in the hyperproduction of reactive oxygen species in the brain [53]. Their high contents in this organ (36–40% in gray matter, 49–66% in white matter, and 78–81% in the myelin sheath) are well known, as is the need for significant oxygen consumption by nerve cells [54]. During this process, lipids are exposed to ROS; as a result, through the mechanism of a chain reaction of free radicals, corresponding products are formed [55]. Interestingly, histological studies show the co-localization

of lipid peroxidation products and  $\beta$ -amyloid plaques in the brains of patients with Alzheimer's disease [56,57]. Thus, the evaluation of the malondialdehyde content showed significantly higher levels of this marker in the hippocampus, occipital and temporal cortices, and serum and plasma [58–60] in people with Alzheimer's disease compared to a healthy group of people. Similar results were obtained in the analysis of levels of 4-hydroxy-2-trans-nonenal (4-HNE), isoprostanes and neurostans, acroleins, oxidized low-density lipoproteins, phospholipids, and hydroperoxides [60–63].

Because mitochondria are the main sources of reactive oxygen species in the central nervous system, these organelles take a leading position in the pathological hierarchy of neurodegenerative diseases [64]. It has been proven that almost all aspects of mitochondrial function are disrupted in Alzheimer's disease. The current hypothesis of the mitochondrial cascade, explaining the onset and progression of AD, states that the dysfunction of these organelles affects the expression and processing of the  $\beta$ -amyloid precursor protein (APP), contributing to the oligomerization of pathological forms of the peptide [65,66]. Later, it was discovered that  $\beta$ -amyloid is itself a source of oxidative stress, and the incubation of neurons with oligomers of peptide  $A\beta_{1-42}$  leads to lipid peroxidation, as evidenced by high levels of the marker of this process, 4-HNE [67]. Interestingly, 4-HNE levels are proportional to the degree of neuronal damage [68]. In addition, it has been shown that oxidative stress caused by amyloid  $\beta$  (mainly as a result of the formation of metalamyloid complexes of protein with redox-active metals, i.e., copper, zinc, and iron) leads to the disruption of mitochondrial functioning [69] and contributes to the depolarization of the organelle membrane and the development of the phenomenon of excitotoxicity. At the same time, the alleviation of mitochondrial dysfunction leads to the attenuation of the pathological signs of Alzheimer's disease. This allows us to consider targeting damaged mitochondria as a key strategy for reducing oxidative stress in models of this disease.

It is obvious that, initially, the free radical theory was associated with aging and degenerative processes occurring in the brains of patients with various types of dementia. With the development of interdisciplinary cooperation between the pharmaceutical and biochemical fields of science with the aim of better understanding the specificity of redox processes, the levels of reactive oxygen species began to attract the attention of researchers in the field of cancer.

Traditionally, the increase in free radical levels has been considered a promising therapeutic strategy for inducing oxidative stress, damaging cellular components, killing tumor cells by inducing apoptosis [70], and triggering autophagy [71]. However, the use of such prooxidant therapies can lead to a number of adverse effects in relation to a healthy microenvironment. For example, the stimulation of oxidative stress via the well-known cytostatic alkylating effect of cisplatin occurs not only in tumor cells but also in cells of normal origin, which is the cause of a common adverse effect of the drug: ototoxicity [72]. Similar to cisplatin, for doxorubicin, the conventional anthracycline antibiotic that leads to the direct generation of hydrogen peroxide and, as a result, the subsequent depolarization of the mitochondrial membrane and caspase-dependent cell death, non-selective localization was found in the mitochondria of untransformed cells; this causes its high cardiotoxicity and imposes serious limitations on its clinical use [73].

Such examples allowed for a revision of the existing concept; therefore, further studies became the basis for the assumption that reactive oxygen species are the driving factor of oncogenesis [74]. Thus, it was found that the elevated levels of ROS shown in various types of tumor cells are, on the contrary, oncogenic due to the damaging effects of DNA, proteins, and lipids, which contribute to genetic instability and the emergence and progression of oncogenesis [32]. This is shown by a significant increase in the above-mentioned markers of oxidative stress in the biological material of patients with malignant neoplasms. In particular, an increase in malondialdehyde is found in carcinoma of the breast [75,76], lung [77], prostate [78–80] and bladder [81,82], oral cavity, and oropharynx

[83,84], the levels of which correlate with the clinical stages of cancer, reaching their maximum in stages III and IV [85], which indicates the direct role of oxidative stress in the progression of diseases. Damage in DNA malignancies as a result of free radicals is confirmed by an increase in excreted 8-hydroxy-2-deoxyguanosine (8-OHdG), which plays an important role in carcinogenesis for the transformation of healthy cells to tumor cells [86]. Studies show that high levels of 8-OHdG in urine, plasma, and serum are prognostic factors in carcinoma of the esophagus [86,87], ovaries [88,89], and large intestine [90–92]. Such ectopic accumulations of reactive oxygen species and the development of permanent oxidative stress lead to dedifferentiation [93] and abnormal cell growth [94,95], metastasis [96], the emergence of resistance to apoptosis (in particular, due to the increased glucose metabolism and adaptation to hypoxic conditions) [97], angiogenesis [98], and the generation of oncogenic mutations [99].

Thus, despite all of the inconsistencies detailed above, to date, both an inhibition of free radical reactions and a stimulation of the production of reactive oxygen species for the specific destruction of transformed cells are considered to be promising strategies for antitumor therapy. However, the use of the latter as a promising therapeutic tool should have a strict selective focus on tumor cells.

### 2.3. Dysfunction of the Cell Antioxidant Defense System

Because the generation of reactive oxygen species is an inevitable phenomenon, not only in pathological conditions but also in normal conditions, living organisms have evolutionarily formed the intrinsic antioxidant defense system, aimed at the strict regulation of redox homeostasis and striving to eliminate excessive amounts of reactive oxygen species without the inhibition of their useful role [100,101].

The cellular system of endogenous antioxidant defense consists of a number of components, the protective mechanisms of which function in two main directions: (1) the elimination of free radicals and reactive forms with the enzymes superoxide dismutase, catalase, glutathione peroxidase, and glutathione reductase [102] and (2) the elimination of free radicals by the electron donor glutathione (GSH) [103].

The first line of the intrinsic antioxidant system, which plays a key role in the protective mechanisms of the cell, consists directly of the above-mentioned enzymes, which act by binding superoxide and other peroxides. The enzymes convert them into stable compounds and thus have an important biological defense role against attacks by ROS [101].

The enzyme superoxide dismutase, which catalyzes the dismutation reaction of the superoxide anion radical into hydrogen peroxide, is localized in the cytosol and mitochondria [104]. Furthermore, the resulting hydrogen peroxide is converted into water and oxygen as a result of the enzymatic activity of catalase [105]. Despite the fact that this antioxidant is abundantly localized in various compartments of cells and is considered the main antioxidant [106], catalase is not found in mitochondria. Another enzyme, glutathione peroxidase, is needed for the degradation of hydrogen peroxide molecules in these organelles, where its main activity is implemented by converting  $\text{H}_2\text{O}_2$  into two  $\text{H}_2\text{O}$  molecules. It is known that glutathione peroxidase is the first enzyme activated under high ROS levels [107]. In addition to  $\text{H}_2\text{O}_2$ , glutathione peroxidase also converts peroxides and hydroxyl radicals to non-toxic compounds by sequentially oxidizing reduced glutathione to glutathione disulfide, which is reduced to GSH under the action of glutathione reductase [108]. One of the most important functions of glutathione reductase is to maintain the ratio of reduced and oxidized forms of glutathione [109].

Glutathione is one of the main non-enzymatic antioxidants that is involved in many cellular functions by acting as a free radical scavenger [110]. It is synthesized in the cytosol and then distributed to almost all cell compartments, including the mitochondrial matrix, where it reacts with ROS and prevents apoptosis [111]. Under physiological conditions, thiol-reduced glutathione (GSH) is the basic form, which is present at much higher concentrations than its disulfide-oxidized form (GSSG); meanwhile, under oxidative stress conditions, the ratio shifts towards GSSG [112].

Various age-related and metabolic diseases are closely associated with abnormal levels of endogenous antioxidants [19,113]. Thus, in the biological materials of patients with cervical carcinoma, significantly lower activity levels of superoxide dismutase [114], catalase [115], glutathione peroxidase, and glutathione reductase [116] are found compared with a healthy control group. A similar pattern has been shown in breast [117] and lung [118,119] carcinoma, progressive bladder carcinoma [120], and lymphocytic leukemia [121,122]. It should be noted that the observed changes in the activity of enzymes are significantly worsened as disease progresses, which indicates their role in the pathogenesis of malignant neoplasms. As for GSH, the levels of this antioxidant play a diametrically opposite role in the development of cancer diseases [123]. Because glutathione, in addition to antioxidant protection, plays a role in many metabolic processes, increasing the sensitivity of the GSH system in response to redox changes in cancer does not have a positive effect. On the contrary, it protects transformed cells from death in a stressful microenvironment, contributing to proliferation, metastatic activity, and the acquisition of resistance to the action of therapeutic agents [124–126]. In particular, direct correlations between high levels of the reduced form of glutathione and unfavorable prognostic signs were found in carcinoma of the ovaries [127], breast [128], colon [129], and lung [130], along with leukemia [131] and a number of other types of malignancies.

The attenuation of the antioxidant defense system of the cell is also directly related to the pathogenesis of neurodegenerative disorders [132]. Thus, the measurement of a wide range of enzymatic and non-enzymatic antioxidants in samples obtained from patients with Alzheimer-type dementia demonstrated a decrease in their activity, with a positive correlation as the clinical prognosis worsened [133,134].

#### 2.4. ROS-Mediated Signaling Pathways in Oncology and Neurodegeneration

In recent years, the understanding of the role of oxidative stress has expanded significantly, and nowadays it is often considered to be an imbalance that plays a direct role in the regulation of gene expression and related signaling pathways. As important physiological modulators of intracellular signaling pathways, reactive oxygen species are involved in the progression of malignancies and neurodegenerative disorders through the regulation of mitogen-activated protein kinase (MAPK) and phosphoinositide 3-kinase (PI3K)/Akt, the activation of nuclear factor of activated B cells ( $\kappa$ B (NF- $\kappa$ B)), and mutations in transcription factors.

1. Mitogen-activated protein kinase (MAPK) are serine–threonine kinase that mediate both the extra- and intracellular signaling that regulates all aspects of cellular functions, including proliferation, differentiation, survival, death, and transformation [135]. MAPK pathways are represented by the three-level kinase cascade, which, when active, phosphorylates various substrate proteins [136]. One of the key signals in the response to which the pathological MARK response occurs is oxidative stress [137], which causes point mutations [18].

To date, the important role of MAPK signaling pathways in the emergence and progression of many diseases has been convincingly proven [138,139], with the focus in recent years on the RAS/RAF/MEK/ERK pathway [140], modifications of which have been proven in almost 50% of cases of human cancers [141]. Thus, the transmission of signals along this pathway and the phosphorylation of the corresponding proteins contribute to the migration and survival of tumor cells [142,143], the degradation of extracellular matrix proteins, and subsequent tumor invasion [144]. Thus, the RAS/RAF/MEK/ERK signaling pathway is considered an important therapeutic target for the development of anti-neoplastic drugs. The significance of other MAPK pathways in the development of malignancies is ambiguous, but, in the case of neurodegenerative diseases, reactive oxygen species (mainly hydroxyl radicals, superoxide anion radicals, and hydrogen peroxide) are typical activators of the JNK and p38 pathways that mediate the downstream negative regulation of  $\beta$ - and  $\gamma$ -secretase activity and the phosphorylation of amyloid precursor protein and tau protein, leading to neuronal death.

2. PI3K/AKT (the signaling pathway of phosphatidylinositol-3-kinase/protein kinase B) is the most important coordinator of intracellular signaling in response to extracellular stimulants, which are free radicals. This signaling pathway plays a central role in the perception of metabolic changes in the environment [145]. Its role is evolutionarily related to the regulation and support of cell growth, proliferation, and survival. Hyperactivation of the PI3K/AKT signaling cascades is one of the most common disorders in cancer [146–148], while neurodegenerative disorders are characterized by impaired signaling in this pathway [149,150].

Studies have shown that, in cancers, the PI3K/Akt signaling pathway affects the cell cycle by phosphorylating cyclin-dependent kinase inhibitors and preventing translocation to the nucleus of the tumor suppressor gene p27, thereby attenuating its inhibitory effect on the cell cycle and directly promoting tumor cell proliferation [151]. The activation of this signaling pathway also determines the resistance of transformed cells to apoptotic death by inhibiting the pro-apoptotic factors Bad and procaspase-9 [152], as well as resistance to antitumor therapy in various types of malignant neoplasms, including carcinomas of the prostate [153,154], lung [155,156], breast [146,157,158], esophagus [159], glioma [160], etc.

In the context of the role of PI3K/AKT in the progression of neurodegenerative diseases, it should be noted that, in the brain, this signaling pathway performs a wide range of functions, including complex processes such as dendrite and axon elongation [161,162]. This gives PI3K/AKT a unique role in the maintenance of synaptic plasticity and has a significant impact on the processes of memory formation [163–165]. PI3K-Akt disorder, found in the brain in neurodegenerative diseases, provokes mitochondrial dysfunction, which leads to a surge in the generation of reactive oxygen species and the further development of pathological mechanisms [166,167]. In particular, it has been shown that inactivation of the PI3K/AKT pathway correlates with an increase in the level of hyperphosphorylated tau protein [168] and A $\beta$ <sub>40-42</sub> plaques in the brain [169], mainly due to the increased activity of glycogen synthase-3 $\beta$  (GSK-3 $\beta$ ) [168]. To date, it has been documented that antioxidant therapy targeting this signaling pathway is a promising strategy for the treatment of Alzheimer's disease [166,168,170].

3. Activated B cell nuclear factor (NF- $\kappa$ B) comprises a family of redox-sensitive transcription factors that regulate the expression of various genes [171] and are involved in inflammatory processes [172,173]. NF- $\kappa$ B is a well-known oxidative stress “sensor” that detects H<sub>2</sub>O<sub>2</sub> at low levels. Under normal conditions, this factor is in a “rest” condition due to its association with the inhibitor of  $\kappa$ B proteins (I $\kappa$ B) [174] while inducing stimuli, including free radicals, to trigger the activation of the I $\kappa$ B kinase complex. Thus, in an in vitro model of neurotoxicity, it was found that the treatment of a human neuroblastoma cell culture with H<sub>2</sub>O<sub>2</sub> promoted the translocation of NF- $\kappa$ B into the nucleus and the subsequent transcription of pro-inflammatory cytokines and chemokines [175].

In recent years, NF- $\kappa$ B has been increasingly recognized as a key factor in all stages of tumor initiation and progression, both as an independent unit [171,176] and in cross-interactions with a variety of other signaling molecules [177]. Free-radical-induced oncogenic mutations leading to NF- $\kappa$ B activation have been identified in various types of malignancies and are associated with a negative prognosis. For example, in the cells of squamous cell carcinoma of the oral cavity, a decrease in the activity of the antioxidant superoxide dismutase and the increased production of ROS correlate with high activity levels of NF- $\kappa$ B [178]. It has been shown that, under the conditions of tumor development, NF- $\kappa$ B has a selective effect in transformed cells: it triggers the activation of survival genes and genes in the tumor microenvironment, contributing to inflammation. It is also of interest that, in neurodegenerative diseases, NF- $\kappa$ B exerts negative effects, not only by inducing neuroinflammation [179,180] but also by stimulating amyloidogenic cascades due to the presence of sites of synthesis in the promoter region of genes involved in amyloidogenesis [181]. All of this evidence suggests that there is a close correlation between this transcription factor and the pathogenesis of cancer and Alzheimer's disease.

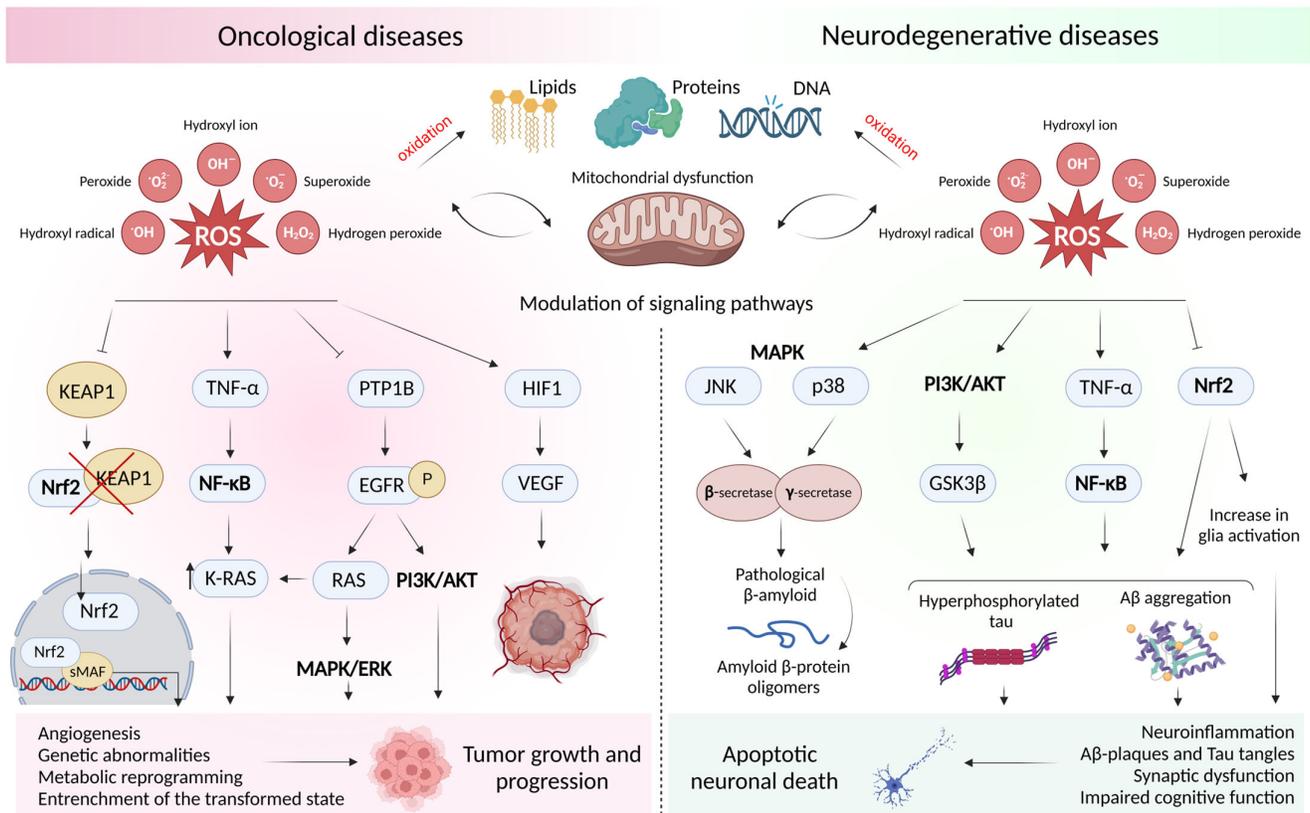
4. Nuclear factor erythroid 2-related factor 2 (Nrf2) is a transcription factor that is considered one of the main coordinators of the cellular antioxidant response. Traditionally, the activation of Nrf2 has been associated with oncological diseases that increase the production of antioxidant proteins and maintain redox balance in tumor cells [182]. However, recent studies have shown numerous functions of Nrf2 that lie beyond its original purpose, opening up the possibility of targeting this factor in the treatment of other diseases, including Alzheimer's disease [183].

Under normal conditions, basal levels of Nrf2 are maintained at a low level, but when pathological conditions occur, it is activated, accompanied by a rapid suppression of reactive oxygen species and the restoration of oxidative damage through the expression of target genes [184]. This positive feature of this protein in the treatment of various diseases has been discussed in detail since the last century in a number of works, and it had no competition until 2006. Researchers have begun to actively highlight the role of Nrf2 activation, mainly in transformed cells, in the progression of malignant neoplasms, contributing to tumor metastasis [185,186] and the acquisition of resistance to therapy [187] by maintaining reprogrammed cell metabolism under hypoxia [188]. This phenomenon has been described as the "dark side" of Nrf2 [189], which has allowed researchers to make significant progress in understanding the role of the transcription factor in the pathogenesis of cancers.

As mentioned above, a growing body of evidence suggests that altered Nrf2 expression is significantly associated with neurodegenerative diseases, including Alzheimer's disease [190]. However, in this case, a decrease in the expression of Nrf2 and protein-controlled genes is associated with an increased risk of the development and early onset of Alzheimer's disease [191]. Thus, in animal models with this cognitive disorder, it was found that, under the conditions of redox imbalance in the brain, the Nrf2-mediated antioxidant response is suppressed, correlating with higher levels of the insoluble form of hyperphosphorylated tau protein [192] and  $\beta$ -amyloid [193]. In addition, in the work conducted by Branca et al., it was shown that Nrf2 deficiency significantly aggravates the cognitive dysfunctions of transgenic APP/PS 1 animals in a study of various types of memory, including spatial, working, and associative, which was associated, in particular, with an increase in the level of  $A\beta$  [194]. In turn, the induction of Nrf2 expression can promote the excretion of the precursor protein  $\beta$ -amyloid and tau protein by influencing the downstream genes involved in the processes of autophagy and macroautophagy [195].

All of these data indicate that the impaired expression of Nrf2 as a result of redox imbalance can be considered an important therapeutic target in the search for promising therapeutic agents for the treatment of both malignant neoplasms and Alzheimer's disease.

Thus, oxidative stress is certainly a significant common denominator that is shared by cancer and neurodegenerative disorders (Figure 1). Although this process is not the only factor in their etiopathogenesis, it creates interesting opportunities for the development of new strategies for the treatment of these socially significant diseases.



**Figure 1.** Schematic illustration of the negative role of mitochondrial dysfunction and reactive oxygen species in the development and progression of oncological and neurodegenerative diseases, with an emphasis on the cross-effects of signaling pathways associated with oxidative stress. This figure was created by the authors using BioRender.com (<https://www.biorender.com/> (14 June, 2023)).

### 2.5. Potential Neuroprotective and Antitumor Therapeutic Candidates Targeting ROS

Guided by the fact that most of the known therapeutic agents are directly or indirectly derived from natural resources (more than 70% of the currently existing anti-tumor agents are natural products or their derivatives [196]), we focused on substances of natural origin with high pharmacological potential.

A prominent representative of a natural substance with good potential in the treatment of a wide range of diseases is quercetin (3, 3', 4', 5, 7-pentahydroxyflavone), which is a polyphenolic compound.

Among the anti-tumor effects of quercetin is its ability to induce cell cycle arrest in G2/M and G1 phases through the regulation of the PI3K/Akt [197] and MAPK signaling pathways [198], inhibit proliferation, and trigger cascades of apoptotic tumor cell death [199], as well as reverse resistance to the action of cytostatic agents [200]. These abilities are primarily due to the bioflavonoid's ability to modulate levels of reactive oxygen species [201]. In particular, in the work of Lu et al. [197], quercetin demonstrated the ability to effectively inhibit the PI3K/Akt signaling pathway in docetaxel-resistant prostate cancer cells LNCaP/R and PC-3/R and, in so doing, to restore the sensitivity of transformed cells to the action of cytostatic agents. This effect also correlated with reduced tumor growth in mice with a xenograft model [197].

This effect was also found in the results of investigations into the chemosensitizing ability of quercetin in ovarian adenocarcinoma [202], where flavonoids, regulating the PI3 signaling pathway K/Akt/mTOR and inhibiting Nrf 2 expression, reversed the resistance of the SKOV-3/CDDP cell line to the action of cisplatin. The sequential treatment of PC3

and DU145 prostate carcinoma cells with vitamin C and quercetin led to a significant decrease in Nrf2 expression and in the activity of glutathione enzymes, which were accompanied by the lower production of endogenous ROS and remarkable levels of cell death [203]. Similar results were obtained in [204] in a study of the therapeutic potential of such a combination against the human breast cancer cells MDA-MB 231. Another mechanism of anti-tumor action of quercetin is its anti-inflammatory function [205], and Lin et al. [206] demonstrated quercetin's ability to restore the number of leukocytes and reduce the expression of markers of oxidative stress in mice with a model of colorectal carcinoma, which was accompanied by a decrease in the number and size of colon tumors.

The neuroprotective effects of quercetin, associated with the implementation of its antioxidant properties, have also been widely studied [207]. In particular, in the work of Rishitha et al. [208], due to the inhibition of lipid peroxidation and the modulation of glutathione levels, solid lipid nanoparticles of quercetin effectively inhibited Danio rerio cognitive dysfunction with pentylenetetrazole-induced neurodegeneration. The neuroprotective action of quercetin has also been confirmed in a lipopolysaccharide-induced (LPS) model, where the chronic administration of flavonoids to 18-month-old mice with LPS-induced dementia led to significant improvements in memory performance [209]. While evaluating the cognitive function of rats with a model of Alzheimer's disease stimulated by toxic forms of A $\beta$ 1-42, Li et al. [210] also identified quercetin's positive effects in the Morris water maze test. This was directly correlated with Nrf 2 activation, altered levels of oxidative stress markers, decreased MDA, and the increased expressions of superoxide dismutase, catalase, and glutathione, which ultimately inhibited neuronal damage. Moreover, the three-month administration of the quercetin glycoside quercetrin to transgenic 5xFAD mice led to the leveling of cognitive disorders mediated by the previously mentioned anti-inflammatory function [211]. As such, by modulating transcription factor NF- $\kappa$ B, which is overactivated in neurodegeneration [212], quercitrin interfered with the excessive secretion of inflammatory cytokines, blocking microglial proliferation.

Among natural compounds, sesquiterpene lactones also play an important role in the development of therapeutic agents due to their wide range of biological activities and promising pharmacological profiles [213–218]. One of the outstanding discoveries of traditional Chinese medicine is artemisinin, which exhibits excellent anti-malarial activity and is isolated from *Artemisia annua* L. [219,220]. Almost 50 years have passed since its introduction into clinical practice as a priority therapy for tropical malaria, while the range of pharmacological properties exhibited by artemisinin is being continuously supplemented [221].

In many studies, artemisinin and its derivatives have been found to exert anti-tumor effects by inducing oxidative stress. In particular, ROS-dependent cytotoxicity has been shown for water-soluble artemisinin–artesunate in cell models of colorectal carcinoma [222], ovarian cancer [223], non-small-cell lung cancer [224], etc. Interestingly, the anti-tumor effects of artesunate are not limited to an increase in the levels of free radicals, and their toxic effects are strictly selective to cells of tumorous origin [225]. As an inhibitor of NF- $\kappa$ B, artesunate reverses the resistance of metastatic cells of castration-resistant prostate cancer to androgen receptor antagonists, leading to their ubiquitin-mediated degradation [226].

Due to its pronounced antioxidant properties in cells of non-tumorous origin and its ability to penetrate the blood–brain barrier without serious adverse effects, artemisinin has been actively considered by researchers for use as a promising neuroprotective agent. Thus, in a study of the protective properties of artemisinin in Parkinson's disease induced by the MPP<sup>+</sup> cell model, it was found that sesquiterpene lactone may inhibit the apoptotic death of SH-SY5Y cells by reducing oxidative damage due to the increase in the activity of the endogenous antioxidant enzymes superoxide dismutase and glutathione and the suppression of levels of malondialdehyde [227]. In an in vivo model of this neurodegenerative disorder, artemisinin reduced damage to dopaminergic neurons in mice treated

with the neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine [228]. The neuroprotective properties of artemisinin have also been shown in the treatment of the HT-22 neuronal cell line with glutamate-induced neurotoxicity, which manifested as the blocking of the production of reactive oxygen species and the activation of the Akt signaling pathway, and, as a result, increased cell survival [229]. In a study conducted by Okorji et al. [230], the authors described the possible therapeutic neuroprotective effect of the fat-soluble artemisinin derivative artemether. Artemether led to the activation of Nrf2 and its binding to elements of the antioxidant response in LPS-stimulated microglia BV2, which directly correlated with a decrease in the levels of inflammatory mediators (prostaglandin E, microsomal prostaglandin E-synthase 1, cyclooxygenase 2, TNF- $\alpha$ , and interleukin 6), as well as in the content of A $\beta$  and the activity of  $\beta$ -secretase 1. Zhao et al. [231] also found that, due to the reduction in oxidative stress and its anti-inflammatory effects, artemisinin has the ability to improve the cognitive functions of triple-transgenic 3xTg cells in the APPSwe, TauP301L, and PS1M146V genes, precisely reproducing the disorders associated with Alzheimer's disease.

Other representatives of the group of sesquiterpene lactones also exhibit potent therapeutic properties due to the modulation of the redox balance: costunolide, first obtained from the roots of *Saussurea lappa*, C.B. Clarke, and parthenolide, extracted from *Tanacetum parthenium* L. The decreased viability of human bladder cancer cells under the action of costunolide is associated with the hyperproduction of reactive oxygen species and the disruption of the transmembrane potential of mitochondria, leading to the overexpression of apoptotic proteins, the suppression of tumor suppressors and, ultimately, the triggering of an apoptotic cascade of cell death [232]. Similar effects of costunolide were demonstrated by Hua et al. [233] in a study of the anti-tumor properties of costunolide in a model of human esophageal squamous cell carcinoma, as well as breast ductal adenocarcinoma [234]. In turn, study of the therapeutic potential of parthenolide has shown that it may induce the death of cervical cancer cells by blocking the PI3K/Akt signaling pathway and inducing intensive ROS formation, leading to the dissipation of mitochondrial membrane potential [235]. The possibilities of the administration of parthenolide in the treatment of triple-negative breast cancer have been demonstrated in a study [236]. Thus, the generation of ROS caused by this compound in the MDA-MB231 cell line led to the depletion of the reduced form of glutathione and the shutdown of transcription factor NF- $\kappa$ B, with subsequent cell death. Moreover, recent work carried out by the research team led by Jorge studied the antitumor activity of parthenolide in lymphoid neoplasms: multiple myeloma, diffuse large-B-cell lymphoma, T- and B-cell acute lymphoblastic leukemia, and Burkitt's lymphoma [237]. Interestingly, the mechanisms of the antitumor action of parthenolide differed depending on the cell line, but, in all cases, sesquiterpene lactone promoted cell death along the apoptosis pathway due to a significant increase in ROS and a decrease in GSH activity.

Similar to artemisinin, costunolide and parthenolide are also actively being considered as potential neuroprotective agents. Treatment with costunolide of the PC12 cell line obtained from the pheochromocytoma of rat adrenal medulla prevented damage to cells with neurotoxin H<sub>2</sub>O<sub>2</sub> [238]. Due to a decrease in the level of intracellular reactive oxygen species, costunolide reduced the expression of caspase 3, which is involved in the apoptosis process. Similar neuroprotective properties were found for parthenolide. In a model of transgenic APP/PS1 mice, parthenolide significantly improved memory performance in the Morris water maze test, perhaps due to its antioxidant properties and ability to reduce neuroinflammation by blocking the AKT/MAPK/NF- $\kappa$ B signaling pathway [239]. Moreover, in the work of Arslan et al. [240], the ability of costunolide and parthenolide to inhibit the activity of the enzyme monoamine oxidase B was considered a possible mechanism of neuroprotective activity in a cellular model of Parkinson's disease.

Table 1 presents the key features of the chemical compounds described above, allowing them to be considered as medicinal agents capable of influencing processes associated with oxidative stress.

**Table 1.** Therapeutic potential of natural products as modulators of oxidative stress in the treatment of oncological and neurodegenerative diseases.

Therapeutic Agent	Molecular Mechanisms of Action	Prognostic Significance	Disease
Quercetin	Inhibition of the PI3K/Akt signaling pathway	Restoration of sensitivity of transformed cells to docetaxel action [197]	PC
	Reduction in the expression of oxidative stress markers and restoration of the leukocyte count	Reduction in inflammation, reduction in tumor size [206]	CC
	Regulation of the PI3K/Akt/mTOR signaling pathway and inhibition of Nrf2 expression	Reversing drug resistance to cisplatin [202]	EOC
Quercetin + Vitamin C	Decreased Nrf2 expression and activity of glutathione enzymes	Stimulation of tumor cell death [203,204]	BC
Artesunate	ROS-dependent cell cycle arrest due to changes in cyclin D3, E2F-1, and p21 expression	Antiproliferative effect [223]	EOC
	Induction of ROS-dependent apoptosis by reducing the VDAC and increasing the cleavage of caspase 3		NSCLC
	Cell cycle arrest due to an increase in p16, p21, p-IRE1a, and LC3B and a decrease in Ki67 and cyclin D1		CC
	Inhibition of NF- $\kappa$ B, ubiquitin-mediated degradation of castration-resistant prostate cancer cells		PC
Costunolide	Triggering of apoptosis as a result of ROS hyperproduction and $\varphi_m$ violation	Transformed cell death [232–234]	BCa
			ESCC
			BC
Parthenolide	PI3K/Akt pathway blocking, ROS hyperproduction, GSH depletion, NF- $\kappa$ B shutdown	Antiproliferative action [235–237]	CCa
			BC
			LN
Quercetin	Inhibition of LPO, increase in GSH expression	Danio rerio cognitive dysfunction restoration in PTZ-induced neurodegeneration [208]	AD
	Reducing the COX-2 level	Improvement of memory parameters in mice with LPS-induced neurodegeneration [209]	
	Activation of Nrf2, decrease in the MDA level, increased expression of SOD, CAT, and GSH	Preventing neuronal damage, leveling cognitive impairment in rats with Alzheimer's disease model stimulated by toxic A $\beta_{1-42}$ forms [210]	
Artemisinin	Decrease in the MDA level, increased SOD and GSH expression	SH-SY5Y cell death inhibition in MPP <sup>+</sup> -induced neurotoxicity [227], reduction in damage to dopaminergic neurons with MPTP-induced toxicity [228]	PD
	Blocking of ROS production as a result of Akt signaling pathway activation	Increased HT-22 survival with glutamate-induced neurotoxicity [229]	
	Activating the ERK/CREB pathway	Inhibition of SH-SY5Y cell death in A $\beta_{1-42}$ toxicity, 3xTg transgenic mice cognitive function improvement [231]	
Artemeter	Activation of the Nrf2 signaling pathway, decrease in the level of inflammatory mediators, A $\beta$ levels, and activity of $\beta$ -secretase 1	Inhibition of neuroinflammation in LPS-stimulated BV2 microglia [230]	AD
Costunolide	Decrease in the intracellular ROS level and caspase 3 expression	Preventing damage to the PC12 cell line by H <sub>2</sub> O <sub>2</sub> [238]	
Parthenolide	Blocking of the AKT/MAPK/NF- $\kappa$ B signaling pathway, neuroinflammation reduction	Improvement of memory indicators in the APP/PS1 transgenic mice line [239]	
	Inhibition of MAO B activity	Cell death decrease in MPP <sup>+</sup> -induced toxicity [240]	PD

Oncological diseases

Neurodegenerative diseases

Abbreviations: PC, pancreatic cancer; CC, colorectal carcinoma; EOC, epithelial ovarian carcinoma; BC, breast cancer; NSCLC, non-small-cell lung cancer; PC, prostate cancer; BCa, bladder cancer; ESCC, esophageal squamous cell carcinoma; CCa, cervical cancer; LN, lymphoid neoplasms; AD, Alzheimer's disease; PD, Parkinson's disease; PTZ, pentylentetrazole; LPS, lipopolysaccharide; VDAC, voltage-dependent anion-selective channel 1; SOD, superoxide dismutase; CAT, catalase; GSH, reduced form of glutathione; MAO B, monoamine oxidase B; MPP<sup>+</sup>, 1-methyl-4-phenylpyridinium; MPTP, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine.

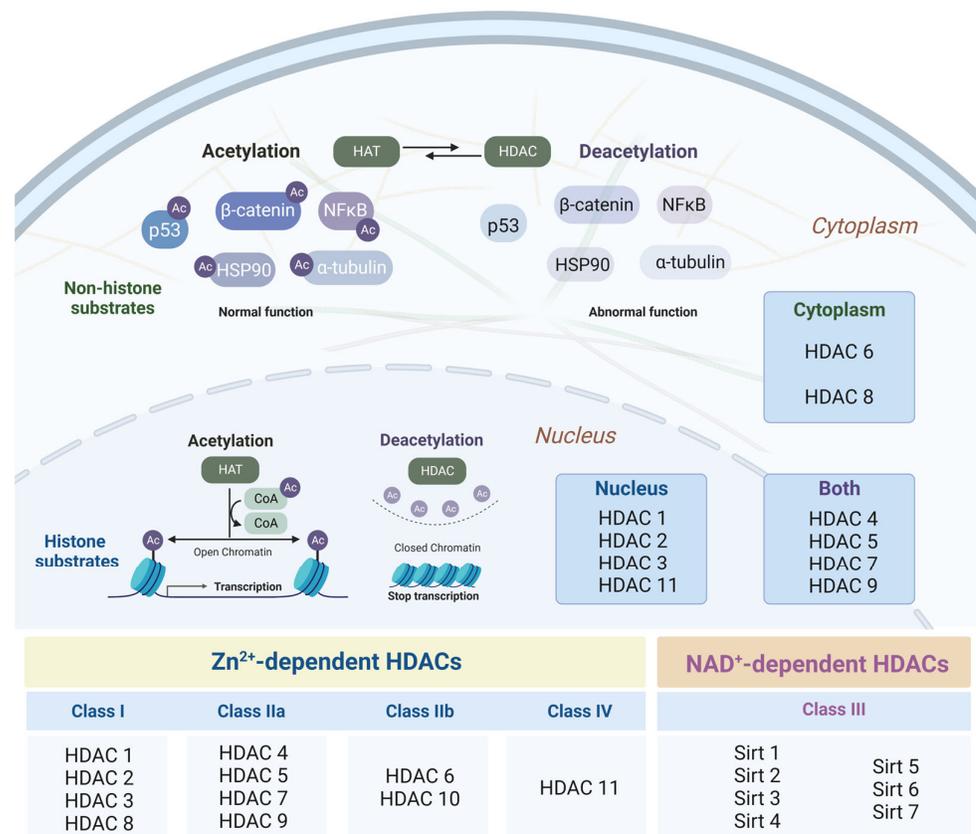
### 3. General Aspects of the Epigenetic Regulation of Neurodegenerative Diseases and Cancer Pathogenesis

#### 3.1. Histone Deacetylases as Major Epigenetic Regulators: Structure and Function

It is well known that the structure of chromatin is made up of DNA and histones. In total, 146 pairs of DNA bases are tightly wrapped around an octamer of histone proteins, including two copies of H2A, H2B, H3, and H4, with H1 being a linker histone protein [241,242]. The long N-terminal regions of histone proteins undergo many post-translational modifications, including acetylation, methylation, phosphorylation, sumoylation, ubiquitination, etc. [243–245].

Histone acetylation and deacetylation are the most important epigenetic processes that influence chromatin status and gene expression. Representing tightly regulated dynamic processes, they are controlled by a fluctuating balance between the reversible activity of enzymes of two families: histone acetyltransferases (HATs) and histone deacetylases (HDACs) (Figure 2) [246–248].

The acetylation process consists of adding an acetyl group to the N-terminal lysine of substrates, which leads to a decrease in the positive charge of histone proteins to neutral and prevents them from binding to negatively charged DNA [246,249]. This formation of the loose structure of chromatin, euchromatin, is associated with transcription activation. In turn, deacetylation, the reverse process of acetylation, involves the removal of the acetyl group from lysine residues in the tails of proteins, which enhances the interaction between positively charged histones and negatively charged DNA [250]. This strong binding of DNA to histone proteins contributes to the formation of a permissive state of chromatin, which interferes with gene transcription. Interestingly, as enzymes that limit this process, histone deacetylases also have the ability to regulate other post-translational modifications, such as methylation, ubiquitination, and sumoylation [251]. For example, acetylation has been shown to inhibit proteasome-mediated protein degradation, dependent on ubiquitination [252]. All of this has led to a genuine interest in the role of HDACs as critical regulators of the normal and pathological functioning of the body. Despite the fact that the first documented studies of the enzymatic activity of histone deacetylases date back to relatively recent works published in the early 1970s, since then, approximately 15,000 articles have been published on this topic (according to a systematic literature search for original articles in the PubMed database); as a result, important discoveries have already been conducted in the field of HDAC investigations.



**Figure 2.** Histone acetylation and deacetylation processes as fundamental mechanisms of epigenetic regulation that controls gene expression. Acetylation is mediated by the activity of histone acetyltransferase enzymes (HATs), which catalyze the attachment of acetyl groups to the lysine residues of histone proteins, which contributes to the relaxation of chromatin conformation and the triggering of transcription. Histone deacetylases, on the contrary, remove acetyl groups from histones, which leads to a thickening of the chromatin structure and the suppression of gene transcription. Histone deacetylases also implement their functions through modifications of non-histone substrates. This figure was created by the authors using BioRender.com <https://www.biorender.com/> (20 June, 2023).

The range of biological functions of HDACs includes cell proliferation [253] and differentiation [254], inflammatory responses [255,256], DNA damage [257], and apoptosis [256]. In addition to their role in transcriptional repression, HDACs also act as modulators of non-histone post-translational modifications of proteins of various natures, including transcription factors and signaling mediators [258]; therefore, histone deacetylases potentially play a role in almost every aspect of the body's functioning. As a result, HDACs have been closely studied by researchers in therapeutic experimental paradigms [253]. Among the many roles that HDACs play in human diseases, oncological diseases are the most frequently discussed.

### 3.2. Changes in the Intensity of Histone Acetylation during Oncogenesis

The aberrant activity of histone deacetylases is often associated with tumor progression; as a result, enzymes of this class have been recognized for more than 30 years as key targets for the action of therapeutic agents against various types of malignant neoplasms [258–260].

Numerous reports state that the overexpression of HDACs is observed in both solid tumors and hematologic malignancies, a finding that correlates with multiple clinical and pathological parameters and low patient survival.

According to preclinical and clinical studies, it is class I HDAC that contributes to the development of malignant neoplasms.

Thus, HDAC1 is an important epigenetic factor in lung carcinoma [261], and there is a close correlation between its expression and the degree of histological differentiation, as well as the subtype [261]. HDAC1 expression was found to be higher in squamous cell carcinoma than in lung adenocarcinoma [262]. The level of HDAC1 expression can also serve as a good diagnostic and prognostic marker of malignant neoplasms of the gastrointestinal tract [261], especially in colorectal carcinoma [263,264]. When analyzing the relationship between HDAC1 expression and clinical features, it was found that the activity of this isoform of histone deacetylase was higher (1) in patients with stages III–IV than in patients with stages I–II of gastric cancer [265–267] and liver cancer [268]; (2) in patients with poorly differentiated liver cancer [269] and adenocarcinoma of the large intestine than in patients with medium- and high-grade cancers [270]; and (3) in groups of persons with positive lymph node metastasis in gastric [265,271,272] and hepatic [268] carcinomas and distant metastases in colorectal carcinoma [263]. A recent study of the possible mechanism of action of histone deacetylase 1, conducted by Yu et al. [273], demonstrated that HDAC1 is involved in the stimulation of the proliferation of malignant gastric cells by enhancing the expression of long non-coding RNAs that regulate the activity of both oncogenes and tumor suppressors [274–277]. Moreover, HDAC 1 has been found to play an integral role in tumor cell evasion from the immune response due to the  $\gamma$ -interferon-induced expression of homologue B7 1 (B7-H1), which plays a fundamental role in the initiation and progression of gastric carcinoma [278]. At the same time, in the study conducted by Jiang et al. [279], the analysis of primary tumor samples obtained from patients with gastric cancer showed that the overexpression of HDAC1 is accompanied by a high value of maximum standardized uptake and a poor prognosis. This is due to the direct effect of the enzyme on the activity of the factor induced by hypoxia 1-alpha (HIF-1 $\alpha$ ), leading to a shift in the metabolic state of tumor cells towards glycolysis.

Epigenetic modifications associated with the overexpression of HDAC1 also play a causal role in the progression of breast cancer (BC). Guo et al. [280] have shown that high levels of the expression of histone deacetylase 1 correlate with clinical and pathological signs and a negative prognosis in patients with breast cancer, with a direct relationship between HDAC 1 levels and the histone binding protein RBBP4 responsible for tumor cell invasion and migration. HDAC1 levels in the cells of this malignancy are higher than those in normal cells; this contributes to their proliferation and migration by regulating the transcriptional and promotional activity of interleukin 8 [281], which plays a significant role in numerous oncogenic pathways [282]. At the same time, in a recent review by Sukocheva et al. [283], the defining role of epigenetic regulation in breast cancer resistance by estrogenic receptor modulators was noted. Thus, the abnormal expression profile of HDAC1 acts as an enhancer for the development of multidrug resistance (MDR) in breast cancer cells, leading to the blocking of estrogen receptors along with the induction of tumor activator genes. A similar effect may also be related to the ability of HDAC1 to enhance the expression of P-glycoprotein (P-gp), a membrane protein that is a critical transporter of drug efflux [284], by recruiting transcription coactivator P300/CBP-associated factor (PCAF) and nuclear transcription factor Y (NF-Y) subunit  $\alpha$  to the P-gp promoter region [285]. The recent studies by Duan et al. [286,287] were the first to investigate the role of HDAC1 in the regulation of another protein with similar P-gp functions: placental breast cancer resistance protein (BCRP), which is overexpressed in tumor cells. It was found that histone deacetylase 1 activity has a positive correlation with BCRP expression in placental breast cancer cells.

Increased HDAC1 activity has also been found in prostate cancer [288,289]. In particular, in a study conducted by Halkidou et al. [290], an immunohistochemical analysis of HDAC1 expression was performed in samples obtained from malignant prostate lesions in humans and mice with a CWR 22 xenograft model. The significant activation of this

enzyme produced an aggressive, strongly proliferating phenotype and the metastatic potential of cells mediated by the suppression of the activity of the tumor suppressor p53 and inhibitors of the cyclin-dependent kinases p 21 and p27. A pronounced prognostic effect of HDAC1 on prostate cancer was shown in a large-scale analysis of samples in a study conducted by Burdelski et al. [288], where nuclear accumulation HDAC 1 was closely correlated with tumor aggressivity and poor prognosis, as was also confirmed in earlier studies [291,292]. Considering one of the possible mechanisms of oncogenic action of histone deacetylase 1, Shankar et al. [293] identified the HDAC1-mediated repression of Maspin, a tumor-suppressor gene that regulates cell invasion, angiogenesis, and apoptosis, processes important for both tumor growth and metastasis [294,295].

Histone deacetylase 1 is a predictor of an unfavorable tumor phenotype in gynecological cancers: ovarian carcinomas [296] and cervical carcinomas [297]. A global study of the relationship between the immunohistochemical expression of HDAC1 and the clinical and pathological data of patients with ovarian cancer revealed the maximum increase in nuclear expression in the samples obtained in patients with mucinous carcinoma, 80% with clear cell carcinoma, more than 70% with serous carcinoma, and 53% with endometrioid carcinoma [298], indicating a clear correlation of HDAC 1 levels with the prognosis. A study on chemoresistant A2780-AD ovarian cancer cells of the molecular mechanisms of epigenetic modifications associated with HDAC1 overexpression showed the mediated HDAC1 suppression of the G-protein 10 signaling regulator (RGS10), which performs a key function in inflammation and cell survival [299]. A number of studies have shown that the deficiency of this protein is associated with the development of resistance to therapy in transformed cells by increasing the production of TNF- $\alpha$  and cyclooxygenase 2 (COX-2), which mediates the production of prostaglandin E2 (PGE) 2 [300,301]. Two years later, the same team of authors, led by Cacan [302], found that an increase in the activity of HDAC 1 in the A2780-AD cell line also accompanied a decrease in the level of acetylated histone 3 (H3) in the protective region of the FAS antigen, as a result, blocking its receptor function, which consists of the induction of programmed apoptotic cell death. In the work by Liu et al. [303], HDAC1 activation was also shown to be an important event in the development of drug resistance, with the results indicating that the possible mechanism of this enzyme's action was of particular interest. The team of authors found that the abnormal expression of HDAC1 in cisplatin-resistant ovarian cancer cells stimulates cell proliferation and chemoresistance by regulating the c-Myc-miR-34a pathway, which stimulates the aberrant expression of the transcription factor c-Myc and suppresses the activity of the powerful miR-34, a tumor suppressor, two known regulators of multidrug resistance [304–307].

As for cervical cancer, Liu et al. [297] found that the aberrant activity of HDAC1 in C-33A cells significantly increases the expression of octamer-binding embryonic transcription factor 4 (Oct4), a prognostic biomarker of various types of malignancies that plays a critical role in maintaining the pluripotency and self-renewal of embryonic stem cells [308–311]. The contribution of histone deacetylase 1 to the maintenance of stem properties in transformed cells was also shown in a recent study by Yokoi et al. [312], where an abnormal HDAC 1 expression profile in the ME180 and CaSki cell lines led to the activation of the Oct4, Nanog, and SOX2 genes that exhibit oncogenic function.

Changes in the activity of other isoforms of class I HDACs may also have clinical value as therapeutic targets. However, nowadays, the understanding of their role in the pathogenesis of cancer is at fairly early stages compared to the understanding of HDAC1.

However, it has been thoroughly proven that HDAC2 is overexpressed in tissue samples obtained from patients with lung cancer, with a negative correlation between HDAC2 levels and prognosis [313]. Such a function of histone deacetylase 2 in the processes of the migration and invasion of lung cancer cells can be implemented by regulating the expression of protein complexes, such as eukaryotic initiation factors 5 and 6 (eIF5 and eIF6), which play a decisive role in the occurrence and progression of tumors [314–317]. The ability of HDAC 2 to control the metastasis of transformed cells may also be mediated by

other mechanisms. In [318], an increase in HDAC 2 levels in non-small-cell lung cancer (NSCLC) cells led to an increase in the expression of fibronectin, a protein that is an extracellular driver of malignancies [319–321]. Wang et al. have shown that HDAC2 activates the expression of c-Myc and cyclin D1, which promote the proliferation, migration, and invasion of NSCLC cells [322]. The expression levels of histone deacetylase 2 are also impaired in colorectal cancer [323]. The analysis of specimens obtained from patients with colorectal carcinoma showed a significant increase in HDAC2 and abnormal levels of H3K56 acetylation in tissues [263]. Clinical and pathological data presented in previous work [324] showed that a higher expression of HDAC 2 correlates with poor overall survival and is associated with liver metastasis. Histone deacetylase 2 levels are also associated with tumor aggressiveness in gastric cancer [325]. Thus, there is a statistically significant increase in HDAC2 expression as the disease progresses, along with positive metastasis to the lymph nodes. HDAC2 may also be a potential biomarker for the prognosis of tumor progression in squamous cell carcinoma of the oral cavity [326], esophageal squamous cell carcinoma [327], gallbladder carcinoma [328], and breast carcinoma [329].

As a key component involved in DNA replication and repair and maintenance of the chromatin structure, histone deacetylase 3 plays a number of important roles in regulating cell progression, differentiation, and other processes associated with the progression of malignant neoplasms [330–332]. Abnormalities in HDAC expression are often found in the most common types of oncological diseases, in particular, in various histological forms of non-small-cell lung cancer [333–336], prostate cancer [337], gastrointestinal malignancies [332,338–343], glioma [344], etc. Beyer et al. [345] found that the overexpression of HDAC3 mediates the growth of transformed cells in human acute myeloid leukemia by modulating the leukemia-related transcription factors  $\beta$ -catenin, the Wilms tumor suppressor gene (WT1), and the myelocytomatosis oncogene (MYC). The aberrant activity of HDAC3 in colon adenocarcinoma cells, SW480, significantly reduces H4-K12 histone acetylation and modulates gene expression in the intracellular signaling pathway Wnt [346], the hyperactivation of which positively controls a number of key cascades regulating stem properties, metastasis, and immune control in neoplastic cells [347–349]. Moreover, it has been found that HDAC3 promotes not only the proliferation and invasion of transformed cells but also the acquisition of drug resistance by suppressing the transcription of tumor suppressor genes p 53, p27, and Bax [350] and activating the PI3K-Akt-mTOR pathway [351].

Another histone deacetylase of class I, the deregulation and overexpression of which are involved in various aspects of the progression of malignant neoplasms, is HDAC8. Despite the fact that this isoform was identified relatively recently as the last representative of class I HDACs, advanced methods allowed researchers to clearly identify the structure and function of the enzyme and determine its significance in oncological diseases [352–354]. By acting on both histone and non-histone substrates, overexpressed HDAC8 implements its oncogenic functions through the regulation of a number of signaling cascades in various types of malignant neoplasms, in particular, in hepatocellular carcinoma [355], gastric adenocarcinoma [356], acute lymphoblastic leukemia [357], squamous cell carcinoma of the oral cavity [358], neuroblastoma [359], etc. [360–362]. Thus, the study of the molecular mechanisms of the HDAC8-induced proliferation of tumor cells showed that, by reducing the expression of cytokine signaling suppressors 1 and 3 (SOCS1 and SOCS3), histone deacetylase 8 contributes to the constitutive activation of the Janus kinase signaling pathway 2/signal transducers and activators of transcription signaling (JAK2/STAT) [363], which play an important role in the oncogenesis of myeloproliferative neoplasms and leukemia [364]. A recent study by Zhang et al. [365] showed that the deacetylated conserved residue of K62, the key enzyme pyruvate kinase glycolysis M2 (PKM2), facilitates the transport of PKM2 into the nucleus, enhancing its enzymatic activity; it also binds  $\beta$ -catenin, promoting the transcription of the gene encoding cyclin D (CCND1), which stimulates the progression of numerous malignancies [366–368] and cell cycle development. HDAC 8 is also involved in the metastasis of many cancers

[354,369,370]. Similar to the first isoform of histone deacetylase, HDAC 8 also promotes the migration of prostate carcinoma cells by inhibiting the expression of the tumor suppressor maspin [293], and the functional redundancy of HDAC8 in human cervical cancer cells (HeLa) promotes metastasis through the excessive deacetylation of tubulin [371]. Neoplastic cell migration in triple-negative breast cancer is significantly enhanced by the HDAC8-induced modulation of mesenchymal markers such as matrix metalloproteinases 2 and 9 (MMP-2 and MMP-9), N-cadherin, fibronectin, and vimentin [372].

### 3.3. Role of Histone Deacetylases in the Pathogenesis of Neurodegenerative Disorders

The role of histone deacetylases in the regulation of brain functions, neurological status, and pathogenesis of a wide range of neurodegenerative conditions has been investigated using a huge number of experimental models [373]. It was found that, by reducing the acetylation of histone proteins, as well as non-histone substrates, individual isoforms of histone deacetylases cause the suppression of the transcription of the regulatory genes involved in neuroplasticity, learning, and memory processes, leading to the development and progression of central nervous system disorders.

For 15 years, researchers have focused on the relationship between the sixth isoform of histone deacetylase and Alzheimer's disease. Because of this, nowadays, HDAC6 has become a widespread therapeutic target for the treatment of this neuropathology [374]. Histone deacetylase 6 is a unique enzyme since it implements its functions through both epigenetic and non-epigenetic mechanisms, regulating a variety of signaling pathways associated with neurodegenerative disorders [375]. In an analysis of the levels of different isoforms of histone deacetylases in the frontal lobe of the brains of patients with mild, moderate, and severe Alzheimer's disease, a significant increase in HDAC6 was found, which negatively correlated with overall cognitive status as the disease progressed [376]. Meanwhile, in the work of Bai et al., positron emission tomography imaging of the brain of transgenic 5xFAD mice, modeling dementia of the Alzheimer type, showed a significantly higher radioactivity of the probe [<sup>18</sup>F]PB118 for HDAC6 imaging in the cortex and hippocampus, the regions most susceptible to the disorder [377].

Due to the predominantly cytoplasmic localization inherent in histone deacetylase 6, which differentiates this enzyme from other HDACs, the spectrum of specific non-histone substrates and proteins deacetylated by HDAC6 includes tau protein,  $\alpha$ -tubulin, ubiquitin, heat shock protein 90, etc. [378,379].

It is known that the level of acetylation of  $\alpha$ -tubulin, the main substrate of HDAC6 [378], plays a key role in the formation of stable microtubules, which are important in various biological processes, including learning and memory [380,381]. The overexpression of HDAC 6 leads to intensive deacetylation of  $\alpha$ -tubulin, leading to the destabilization of microtubules and, as a result, the pathological death of neuronal cells that contribute to neurodegeneration [382]. Moreover, HDAC6 is a critical regulator of the ubiquitin-proteasome system [383,384], processes responsible for the degradation of tau protein [385,386]. In the brain organelles of transgenic mice overexpressing HDAC6, there are high levels of aggregated forms of phosphorylated tau protein [387]. Additionally, in the work of Balmik et al. [388,389], the excessive interaction of the ubiquitin-binding domain of histone deacetylase 6 with tau led to conformational changes in the protein, as well as a decrease in the trend of disaggregation of already formed aggregates. The correlation of high tau protein concentrations with abnormal HDAC6 levels was proven via the quantification of neurons in the cerebral cortices of mice exhibiting high expression of this enzyme, while neurons knocked out by this protein showed a decrease in the formation of neurofibrillary tangles by approximately 90%, which was accompanied by an increase in viability [390]. A similar clearance of hyperphosphorylated tau protein under the conditions of blocking HDAC6 activity was also shown by Cook [391] and Sreenivasamurthy [392].

The overexpression of HDAC6 also influences the processes associated with autophagy. There are several ways to regulate the HDAC6 process, the most important of which

is deacetylation of the autophagy related transcription factors TFEB and FOXO1, which leads to blocking their translocation to the nucleus and the inhibition of transcription associated with gene autophagy. As a result, the processes of the fusion of autophagosomes with lysosomes are disrupted, and there is no degradation of misfolded proteins (tau protein, A $\beta$ , and others) and mitochondria with impaired functions, which ultimately leads to the death of the neuronal cell [393–395]. In recent work by Liu et al., it was found that the aberrant activity of HDAC6 is a critical regulator of neuroinflammatory responses, leading to the overexpression of inflammatory cytokines such as interleukins 6 and 1  $\beta$  (IL-6, IL1 $\beta$ ), as well as TNF- $\alpha$ .

In addition, the aberrant activity of HDAC6 has been demonstrated in other neurodegenerative diseases such as Parkinson's disease [396] and Huntington's disease [397].

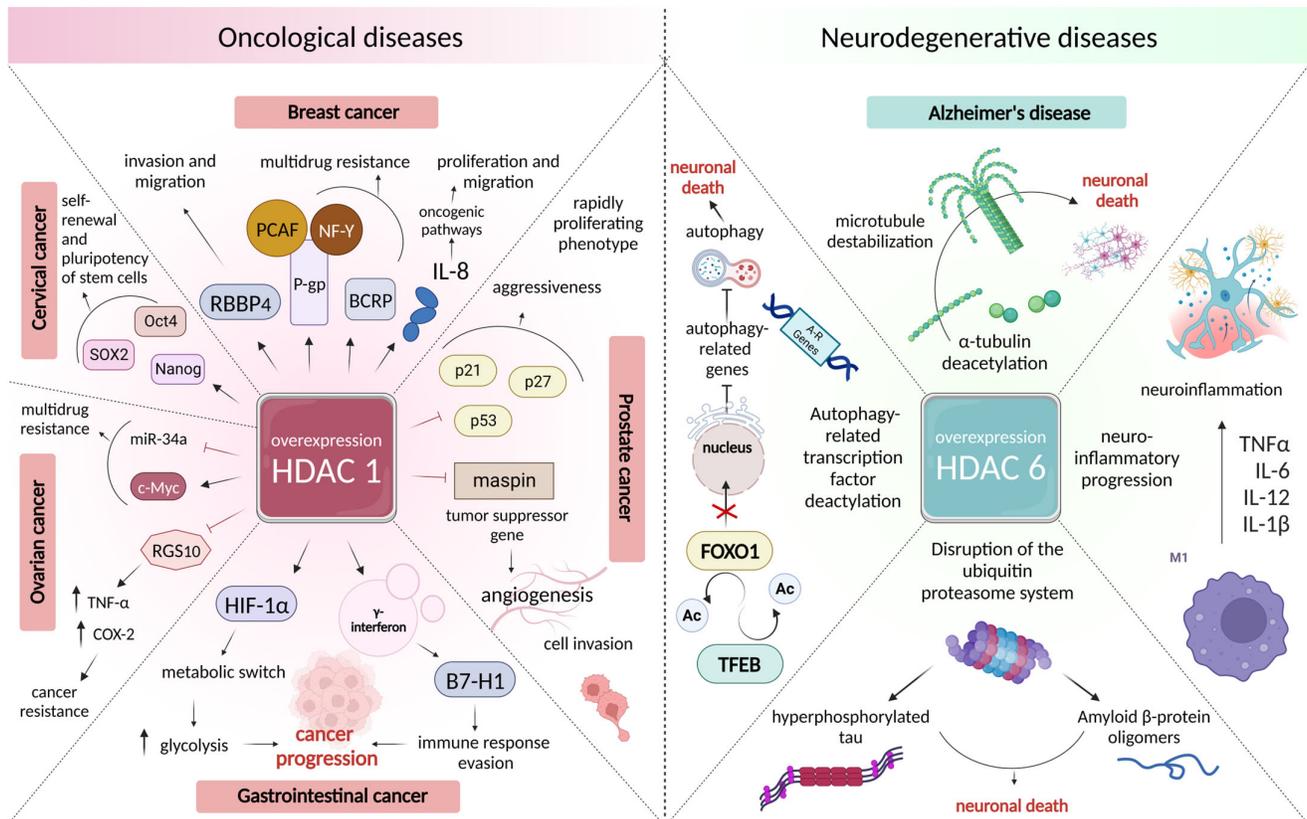
Researchers in the field of Alzheimer's disease have also focused on other isoforms of histone deacetylases, which are also involved in pathogenesis, though to different degrees. There is a clear correlation between the overexpression of histone deacetylase 2, belonging to class I, and impaired brain function. In particular, in the work of Guan et al. [398], the analysis of promoter occupancy showed that histone deacetylase 2 may negatively regulate the activity of the genes involved in synaptic plasticity and memory formation, while the overexpression of HDAC2 in the brains of genetically modified mice contributed to a significant disruption of these functions. Such a repressor function of HDAC2 was also shown in p25 transgenic animals modeling Alzheimer's disease [399], in which an increase in the level of this histone deacetylase isoform correlated with cognitive deficits due to the higher recruitment of HDAC2 on the promoters of key genes associated with learning, memory, and neuroplasticity when compared with wild-type animals. Genes affected by HDAC2 include the brain-derived neurotrophic factor and the early growth response protein 1. It is interesting that, despite the 85% structural homology of HDAC2 to HDAC1, it is the second isoform of histone deacetylase that is involved in cognitive processes [398]. However, even though histone deacetylase 2 would seem to represent an appropriate target for the treatment of neurodegenerative disorders, caution is needed in the use of modulators of its activity. This is because the catalytic function of HDAC2 is a critical requirement of brain neurogenesis in adults [400].

Among representatives of Class I, nuclear-localized HDAC3 is also significantly increased in the hippocampus of APP<sup>swe</sup>/PS1<sup>dE9</sup> transgenic animals, which correlates with high levels of A $\beta$ , the activation of microglia, and a decrease in the density of dendritic spines in the brain. This causes the negative regulation of histone deacetylase 3 and spatial memory in the line of mice [401]. In addition, Bardai and d'Mello [402] found that HDAC3 is a protein with prominent neurotoxic activity since the overexpression of this enzyme leads to the selective death of neuronal brain cells without influencing the survival of cell lines of other origins. With regard to HDAC4, Fitzsimons et al. [403] proved its role in the memory processes of the model genetic organism of *Drosophila* (fruit fly), but it was shown that, depending on the localization, this isoform is not only a repressor of long-term memory, but also modulates its normal formation, which, as was the case for HDAC2, complicates the use of targeted therapy.

For other isoforms of histone deacetylases, specific correlations with Alzheimer's disease have not been proven to date.

Thus, nowadays, we have a good understanding of 18 HDAC proteins in humans, which function as transcriptional repressors and corepressors, leading to genuine interest in enzymes of this class. Despite the fact that information about the structural and functional features of histone deacetylases is increasing exponentially, there are still many questions regarding how particular isoforms of HDACs regulate signaling cascades and gene activity, which largely remain unexplored. Nevertheless, there is indisputable evidence that enzymes of this class are involved in the pathogenesis of a wide range of diseases, mainly malignant neoplasms and neurodegenerative disorders, which lays the theoretical foundation for the clinical use of HDACs as targets for the action of promising pharmacological agents.

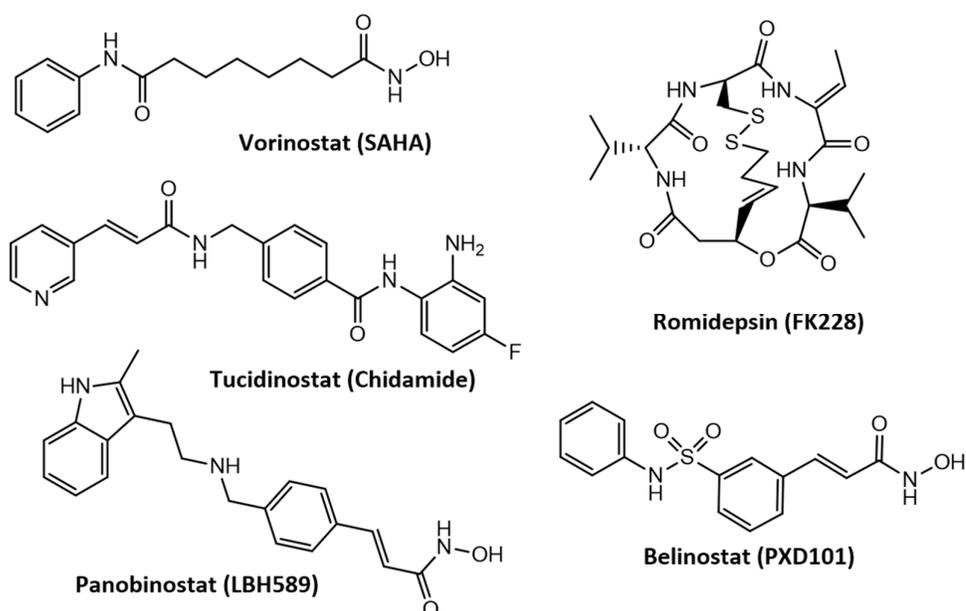
In Figure 3, we combine the most important information about the mechanisms of action of the most common histone deacetylases, HDAC 1 and HDAC6, as key epigenetic regulators of the pathogenesis of onco- and neurodegenerative diseases.



**Figure 3.** Schematic representation of the pathological functions of HDAC1 in various types of malignant neoplasms and HDAC6 in Alzheimer’s neurodegeneration. HDAC1 overexpression promotes oncogenesis by enhancing the proliferation of tumor cells and suppressing apoptosis, as well as metastasis and the development of drug resistance. In turn, aberrant HDAC6 levels are involved in the pathogenesis of neurodegenerative diseases, leading to the destabilization of microtubules and participating in the processes of neuroinflammation, as well as causing disorders in the ubiquitin–proteasomal system and autophagy processes, preventing the degradation of improperly folded proteins. This figure was created by the authors using BioRender.com (<https://www.biorender.com/> (17 June, 2023)).

### 3.4. Advances in the Development of Histone Deacetylase Inhibitors in the Treatment of Cancer and Neurodegenerative Diseases

Currently, the Food and Drug Administration (FDA) has approved the use of five histone deacetylase inhibitors: vorinostat (suberoylanilide hydroxamic acid, SAHA), romidepsin (FK228), tucidinostat (hidamide), belinostat (PXD101), and panobinostat (LBH 589) (Figure 4).



**Figure 4.** FDA-approved histone deacetylase inhibitors.

Based on the formulas presented in Figure 4, it is easy to see that HDAC-inhibitory ability has been found for representatives of different chemical structures. According to convention, all of the histone deacetylase inhibitors that currently exist can be divided into four large groups: (1) hydroxamic acids, (2) benzamides, (3) cyclic peptides, and (4) aliphatic fatty acids. Meanwhile, of all the HDAC inhibitors, it is hydroxamic acid-based compounds that represent the largest and most clinically successful class due to the fact that they exhibit the most promising profile of pharmacological activity [404–406].

Nowadays, there are many studies on the therapeutic potential of hydroxamic derivatives, and it is not possible to cover all such works in this paper. Moreover, many review manuscripts, including our recently published large-scale review, have already been devoted to this problem [407]. In this regard, in this section, we decided to select an original strategy for considering the pharmacological prospects of one of the already approved representatives of HDAC inhibitors: vorinostat. Our review will provide convincing evidence of the prospects of the hydroxamic acid class as unique therapeutic agents and will not only summarize the well-known antitumor properties for them, but also the relatively recently discovered neuroprotective effect.

Vorinostat was the first histone deacetylase inhibitor approved by the FDA in 2006 as a monotherapy for the treatment of patients with refractory or recurrent cutaneous T-cell lymphoma [408,409]. To date, a large number of preclinical and clinical trials of vorinostat in the treatment of other hematological and solid tumors have been conducted. However, studies of the antitumor properties of vorinostat monotherapy for solid tumors showed insufficiently satisfactory results [410,411]. This prompted the teams of authors to attempt to study the efficacy of vorinostat in combination with other active compounds that are already used in the treatment of various malignant neoplasms. In particular, the combined use of vorinostat and radioligand  $^{131}\text{I}$ -metaiodobenzylguanidine has demonstrated a high true-response rate in patients with relapsed or refractory neuroblastoma [412]. The inclusion of vorinostat and isotretinoin in the intensive chemotherapy regimen of medulloblastoma allowed researchers to achieve an improvement in the rates of five-year progression-free survival and overall survival in younger children [413]. The administration of SAHA in combination with autophagy targeting hydroxychloroquine in patients with metastatic colorectal cancer showed a high response of antitumor immunity in

a randomized phase II trial [414]. This method enhanced the expression of lysosomal protease of cathepsin D, the p62 protein, and, as a result, the inhibition of autophagy and the subsequent apoptosis of transformed cells [415]. The enhancement of the antiangiogenic properties of pazopanib under the action of SAHA was also found in phase I studies of metastatic solid tumors with TP53 mutations [416]. Here, patients treated with vorinostat + pazopanib demonstrated a significantly longer average time of overall survival and progression-free life, which may be associated with the triggering of mutant p53 degradation mechanisms and the suppression of vascular endothelial growth factor (VEGF)-mediated overexpression of HIF-1 $\alpha$ . The potential of the strategy of the combined administration of antitumor agents with epigenetic modulators was also confirmed by the results obtained when combining vorinostat with chemoradiotherapy for the treatment of squamous cell carcinoma of the head and neck, where the selected administration regimen demonstrated high efficacy (96.2% of patients showed a positive response) and safety [417], which has also been shown for other cancer types [418,419].

Based on the results obtained in recent years, there is strong evidence of the therapeutic value of vorinostat in diseases affecting the brain [417]. This is because, as a pan-selective inhibitor, vorinostat modulates the activity not only of HDACs of class I, which perform a predominantly oncogenic function, but also of HDAC6, an isoform that plays a leading role in the functioning of neurons. Over the past 10 years, researchers have accumulated an impressive pool of data on the potential of reprofiling the drug agent as a drug for neurodegenerative disorders.

In the work conducted by Chen et al. [420], vorinostat demonstrated a protective effect under the modeling conditions of neurotoxicity induced by lipopolysaccharides and the N-methyl-4-phenylpyridinium cation (MPP<sup>+</sup>), increasing the viability of dopaminergic neurons by inhibiting deacetylation histones and mediating the release of neurotrophic factors from astroglia. It has also been reported that vorinostat exerts neuroprotective effects by stimulating the expression of glycoprotein clustering in human astrocyte cells, which play a known role in modulating A $\beta$  aggregation in Alzheimer's disease [421]. Kilgore et al. [422] showed that intravenous injections of vorinostat into transgenic APP<sup>swe</sup>/PS1<sup>dE9</sup> mice led to the restoration of contextual memory in the animals, which directly correlated with the inhibition of HDACs of class I and HDAC6. Further in vitro studies of the neuroprotective properties of vorinostat also showed that a decrease in the activity of histone deacetylase as a result of SAHA action was accompanied by improvements in synaptic function. However, in vivo behavioral experiments on a model of transgenic Tg2576 mice failed to confirm the protective effect of the compound, which is associated with its limited availability to the brain and effective peripheral distribution [423]. In this regard, the teams of authors focused on another known property of vorinostat: its adjuvant ability. Thus, in the work of Sarathlal et al. [424], the administration of vorinostat as a chemosensitizing agent, in combination with the hypoglycemic agent rosiglitazone, resulted in the significantly higher gene expression of neurotrophic factors and attenuated biochemical, cellular, and behavioral abnormalities in a mouse model of streptozotocin-induced Alzheimer's disease. Interestingly, the authors also managed to improve the profile of the therapeutic efficacy and bioavailability of this system by improving the forms of agent administration using a poloxamer-stabilized system of polymer nanocarriers. One year later, the same group of authors also managed to confirm the potential of vorinostat as a means of enhancing the activity of anti-insulin-resistant drugs [425]. The combined administration of vorinostat and rapamycin resulted in the alleviation of cognitive dysfunction in rats with advanced insulin resistance (IR) undergoing the intracerebroventricular injection of A $\beta$ <sub>1-42</sub>. The subsequent analysis of biomarkers associated with neurodegeneration exacerbated by IR showed a significant decrease in amyloid precursor protein (APP) due to the increased expression of Beclin 1, glial-cell-line-derived neurotrophic factors (GDNF), brain-derived neurotrophic factors (BDNF), neuronal growth factors (NGF), and the neuronal markers MAP 2 (microtubule-associated protein 2) and LAMP 2 (lysosome-bound membrane protein 2). The ability to effectively alleviate cognitive deficit

symptoms in a mouse model of Alzheimer's disease has also been shown for the combination of vorinostat with tadalafil. This combination is aimed at another target, phosphodiesterase type 5, which is a critical component of the cyclic guanosine monophosphate/protein kinase G (cGMP/PKG) signaling pathway that regulates nerve cell apoptosis [426]. It is notable that the synergistic effect of these drug agents also manifested as a decrease in amyloid and tau pathologies and as an obstacle to the death of hippocampal neurons.

The strategy of using vorinostat as a chemosensitizing therapeutic agent has a significant advantage: the use of extremely low doses of the drug during treatment. It allows for the prevention of the adverse effects it has on a healthy microenvironment due to the absence of the selectivity of action on a particular isoform, ensuring optimal safety values in chronic use.

Table 2 summarizes the above materials, focusing on the most significant points in the development of medicinal agents for the treatment of oncological and neurodegenerative diseases.

**Table 2.** Therapeutic potential of vorinostat as an adjuvant agent in the treatment of oncological and neurodegenerative diseases.

Therapeutic Agent in Combination	Molecular Mechanisms of Action	Prognostic Significance	Disease	
<sup>131</sup> I-methaiodbenzylguanidine	Increase in human NET protein expression	Increased radioligand absorption and frequency of true response [412,427,428]	NB	Oncological diseases
Isotretinoin	Modulation of APF2 levels	Five-year progression-free and overall survival improvement [413,429,430]	MB	
Hydroxychloroquine	Autophagy inhibition due to increased cathepsin D and p62 levels	Strengthening of antitumor immunity [414,415,431]	CC	
Pazopanib	Degradation of mutant p53, increased VEGF expression, decreased HIF-1 $\alpha$ levels	Increase in the average duration of overall survival and life without progression of the disease [416,432]	EOC	
			BC	
			CC	
			GC	
			HNSCC	
Chemoradiotherapy	Increased apoptosis rate due to increased Bax and p21 expression	Improved overall survival [417–419]	NSCLC	
			PC	
			GB	
Rosiglitazone	Increased expression of neurotrophic factor genes	Reduced biochemical, cellular, and behavioral disorders in the STZ mouse model of Alzheimer's disease [424]	AD	Neurodegenerative diseases
Rapamycin	Decreased APP due to increased expression of Beclin-1, neurotrophic factors GDNF, BDNF, NGF, and neuronal markers MAP2 and LAMP2	Relief of cognitive dysfunction in rats with an insulin resistance and intracerebroventricular injection A $\beta$ <sub>1-42</sub> [425]		
Tadalafil	Restoration of long-term potentiation, A $\beta$ , and tau pathology relief through the Akt/GSK3 $\beta$ pathway	Restoration of cognitive functions in APP/PS1 transgenic mice [426]		

Abbreviations: NB, neuroblastoma; MB, medulloblastoma; CC, colorectal carcinoma; EOC, epithelial ovarian carcinoma; BC, breast cancer; GC, gastric carcinoma; NSCLC, non-small-cell lung cancer; HNSCC, head and neck squamous cell carcinoma; PC, pancreatic cancer; GB, glioblastoma; AD, Alzheimer's disease; NET, norepinephrine transporter; APF2, angiotensin converting enzyme; VEGF, vascular endothelial growth factor; HIF-1 $\alpha$ , hypoxia-induced factor 1- $\alpha$ ; APP, amyloid precursor protein; GDNF, glial neurotrophic factor; BDNF, brain neurotrophic factor; NGF, nerve growth factor; MAP2, microtubule-associated protein 2; LAMP2, lysosome-associated membrane protein 2; A  $\beta$ <sub>1-42</sub>, pathological form of  $\beta$ -amyloid peptide 1-42.

Thus, to date, the treatment of cancer and neurodegenerative disorders (mainly Alzheimer's disease) represents most indications for the class of drugs targeting HDACs. In particular, based on our assessment of the current status of studies involving vorinostat, the large-scale potential of epigenetic therapy was confirmed by representatives of the class of hydroxamic acids, which have already achieved promising clinical progress.

### 4. Alterations in the Bioenergetic Metabolism of Cells during Oncogenesis and Neurodegeneration

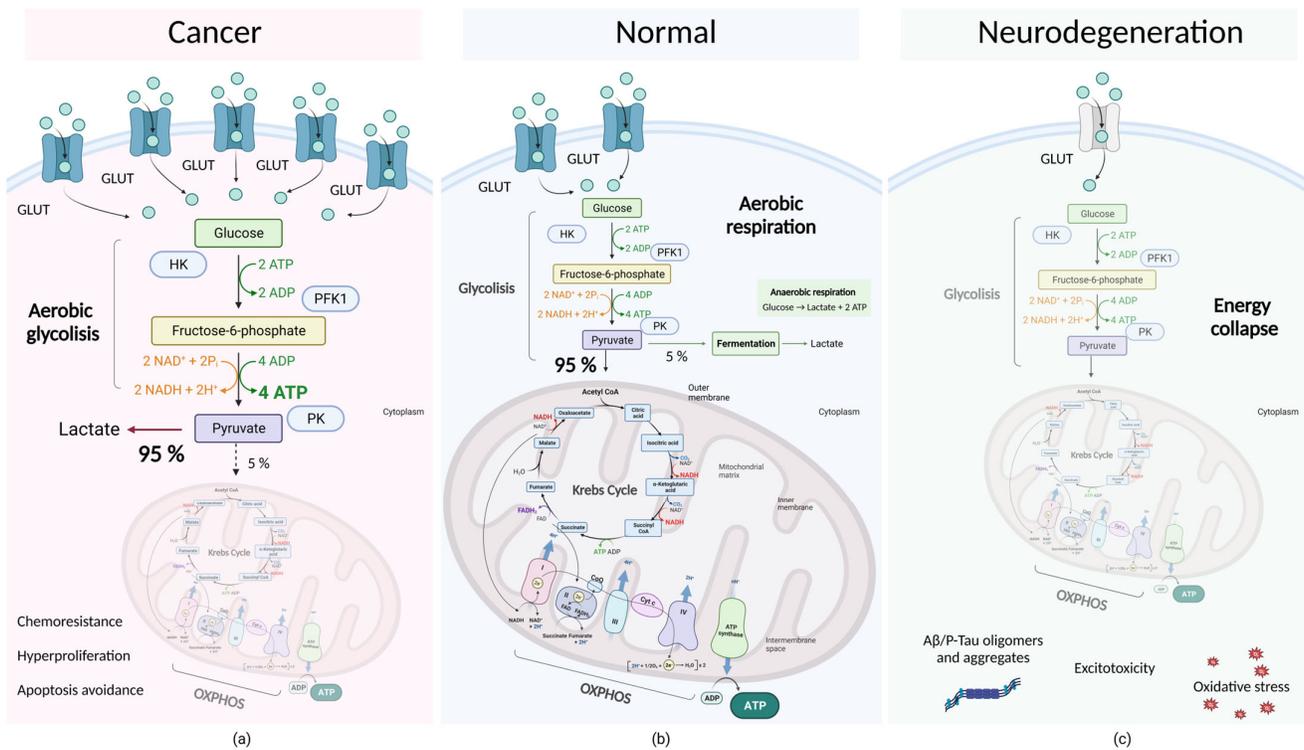
#### 4.1. Determination of the Main Metabolic Processes of the Cell and Energy Metabolism

Cellular metabolism involves many interrelated pathways, the main purpose of which is to provide the cell with the energy necessary for its functioning [433]. In general, metabolism can be divided into a number of chemical reactions, which include both synthesis (anabolism, assimilation, plastic metabolism) and the cleavage of complex macromolecules (catabolism, dissimulation, energy metabolism). These processes are closely interrelated with each other. Thus, during anabolism, energy and products formed in dissimulation reactions are used. In turn, the enzymes formed as a result of assimilation reactions are necessary for catabolism.

As the energy that is necessary for maintaining cell functions, the active transport of ions and substrates, the biosynthesis of complex macromolecules, etc., is formed as a result of catabolism reactions (dissimulation, energy metabolism), and this process has been studied by biochemistry specialists for more than 150 years [434]. Such investigations are the key to understanding both normal physiological functions and their role in the pathological conditions of the body.

Under normal conditions, the production of energy by living organisms occurs through the subsequent oxidation of organic compounds during the three main stages of cellular respiration: (1) glycolysis, (2) the tricarboxylic acid cycle, and (3) mitochondrial electron transport (Figure 5).

Figure 5 clearly illustrates disorders in the cellular metabolism in oncological and neurodegenerative diseases, a detailed consideration of which is presented below in the text of the manuscript.



**Figure 5.** Metabolic reactions occurring in oncological (a) and neurodegenerative (c) diseases, as well as under the conditions of normal cell functioning (b). (a) Schematic representation of the biochemical process occurring in tumor cells, which consists of reprogramming the metabolism and the means of obtaining energy from oxidative phosphorylation to glycolysis. (b) Illustration of the

stages of cellular respiration occurring under physiological conditions. (c) Mitochondrial dysfunction and related metabolic disorders that occur in neurodegenerative diseases and lead to an imbalance between energy production and consumption, as well as the hyperproduction of reactive oxygen species. This figure was created by the authors using BioRender.com (<https://www.biorender.com/> (25 June, 2023)).

#### *4.2. Molecular Subtleties of Tumor Cell Metabolism: Dysregulation of Aerobic Glycolysis and the Warburg Effect*

It is well known that tumor cells have a reprogrammed metabolism that promotes their growth, metastasis, and survival [435]. This proliferative metabolic phenotype [436] is characterized by an increased glycolytic function, in which, due to the intensive increase in glucose and its reduction of pyruvate to lactate, transformed cells not only satisfy their high energy requirements but also obtain intermediates that are critical for the synthesis of macromolecules. This phenomenon, often referred to as the “Warburg effect” [437], has been known for about 100 years and is now the object of studies by a large number of scientific groups. Targeting enhanced glycolysis in tumor cells is considered to be a promising strategy for improving the efficacy of the treatment of malignant neoplasms [438].

Until recently, the predominance of the glycolytic pathway in neoplastic cells was a mystery to the scientific community, since, unlike mitochondrial-dependent oxidative phosphorylation (OXPHOS), this pathway is a less efficient way to obtain energy [439], while cells consume a large amount of energy to maintain the tumor phenotype [440–442]. However, the results produced in recent years have shed light on the long-standing debate about the significance of this unique phenomenon in tumor cells. Pfeiffer et al. postulated that glycolysis, as a method of producing ATP at a high rate, gives neoplastic cells a selective advantage in the face of competition for common energy sources, thereby giving evolutionary significance to this process [443]. In the study conducted by Locasale et al., the rate of ATP production during glycolysis was recorded as being 100 times higher than that with OXPHOS [444]. In addition, the prevalence of glycolysis may be due to the hypoxic microenvironment of tumors, when mitochondrial defects are observed [445], and oxidative phosphorylation becomes inactive, and obtaining energy as a result of high-rate glucose conversion becomes the only possible way of doing so. For example, mitochondrial dysfunction is observed in malignant neoplasms of the large intestine [446], mammary duct [447], and gastric carcinomas [448].

However, since the Warburg effect is observed even in the presence of fully functional mitochondria, and mitochondrial dysfunction is not necessary for oncogenesis, it has been found that neoplastic cells can acquire a hybrid phenotype using both intensive glycolysis and a standard rate of oxidative phosphorylation (glycolysis/OXPHOS) [449]. It is believed that, due to this phenomenon, tumor cells acquire the property of metabolic plasticity, which plays a particularly important role in metastasis and the development of resistance to therapy. Such properties can be achieved through the glycolysis-dependent activation of the PI3K/AKT signaling pathway, which allows tumor cells to acquire stem properties [450].

In addition to providing cellular energy, metabolic intermediates of glycolysis also play a key role in the biosynthesis of macromolecules, thereby providing a selective advantage to cancer cells under the conditions of a limited nutrient supply. In particular, it has been shown that, by enhancing the hexosamine biosynthesis pathway and the pentose phosphate pathway, aberrant glycolysis can contribute to an increase in the turnover of the nucleotides necessary for the effective repair of DNA damage caused by the action of therapeutic agents [451,452]. In turn, such DNA repairs induce the activation of prooncogenic signaling pathways, triggering the mechanisms of resistance of tumor cells to apoptosis [450,453]. In addition, NADPH and ribose-5-phosphate, formed as a result of the pentose phosphate pathway, are essential for the biosynthesis of lipids and nucleic acids in the tumor, and NADPH also allows neoplastic cells to maintain elevated levels of GSH, which exerts the positive effects for tumor progression described above [454,455].

As shown in Figure 5, which depicts the main biochemical reactions of glycolysis under the action of a number of enzymes, the initial consumption of glucose depends on the specific transporters of GLUT [456], which ensure its transfer from the systemic circulation to the cell [457]. Among the 14 proteins of this family belonging to the major facilitator superfamily of membrane transporters, four isoforms of GLUTs 1–4 play a specific role in the development of glucose, exclusively in glucose homeostasis in cells and in the body as a whole [458,459]. At the same time, it is the overexpression of the representative of GLUT-1 that is observed in many types of malignant neoplasms. In particular, the prognostic value of GLUT-1 expression is indicated in adenocarcinomas of the stomach [460,461], pancreas [462], and prostate [463,464]; carcinomas of the lungs [465] and endometrium [466,467]; and osteosarcoma [468], meningioma [469], and malignant tumors of the oral epithelial tissue [470,471]. In addition, high levels of GLUT-1 expression are found in patients with poorly differentiated breast tumors [472] and are associated with a negative prognosis [473–475].

It seems that the blocking of GLUT can be considered a direct approach to glycolysis inhibition as a result of terminating the flow of the substrate into tumor cells, and, as a result, completely disrupting this pathway. However, due to the ubiquitous expression of these transport proteins, this strategy has a critical limitation: the difficulty of developing selective GLUT blockers in neoplastic cells. For this reason, the study of the glycolytic function continues to expand, and, to date, it has been found that the intensity of glycolysis in tumor cells closely correlates with the activity of key glycolytic enzymes, hexokinase, 6-phosphofructo-2-kinase, and pyruvate kinase (Figure 5) [476], which catalyze the irreversible stages of this process; targeting them is thus a promising strategy for the repair of impaired tumor metabolism.

Hexokinase is an enzyme that catalyzes the first stage of glycolysis by converting glucose to glucose-6-phosphate. The limiting role of this enzyme arises due to its kinetic properties, due to which it has an excellent affinity for glucose, which allows the glycolysis process to be triggered even when glucose levels are low, thus playing a key role in the energy metabolism of tumor cells [477]. Among the four existing isoforms of hexokinase in mammals, the contribution of the second isoenzyme (HK 2) to the regulation of the protective functions of neoplastic cells is now well known [478]. In particular, the ectopic expression of hexokinase 2 was found to attenuate apoptosis in a glucose-dependent manner and triggers the Akt-mediated survival of rat fibroblast cell cultures [479]. In addition, a wide range of oncogenic effects of HK2 are shown in hepatocellular carcinoma [480], ovarian malignancy [481,482], non-small-cell lung cancer [483], and prostate [484] and gastric carcinomas [485] and correlates with the recurrence and poor prognosis of breast cancer [486,487].

It has also been shown that, along with an increase in glucose consumption and lactate production, the expression of hexokinase 2, which is found in the prostate carcinoma cell line and has undergone sumoylation, reduces mitochondrial respiration, contributing to the metabolic reprogramming of cells and the triggering of anti-apoptotic mechanisms [488]. Moreover, the oncogenic functions of hexokinase 2 can also be implemented via an alternative non-metabolic pathway. In particular, Wang et al. found a direct correlation between the overexpression of HK 2 and CD133, a key marker of stem cell self-renewal in small-cell lung cancer, due to the glycolytic activity of the enzyme [489]. Similar properties of HK2 have been shown in a model of acute myeloid leukemia (AML). For example, Thomas et al. showed that the interaction of HK2 with nuclear proteins of leukemic stem cells regulates the openness of chromatin and its availability to the action of therapeutic agents, thereby contributing to the development of the mechanism of chemoresistance [490].

6-phosphofructo-2-kinase is an enzyme that catalyzes another irreversible stage of glycolysis: the conversion of fructose-6-phosphate to fructose-1,6-bisphosphate [491]. As with most other glycolytic enzymes, the activity and expression of 6-phosphofructo-2-kinase are impaired in various cancers and are strongly correlated with aggressiveness and

poor prognoses. It was found that the PFKFB3 gene, which encodes the third isoform of 6-phosphofructo-2-kinase and is a key effector protein of transforming growth factor  $\beta$ 1 (TGF $\beta$ 1), stimulates glycolysis in the pancreatic carcinoma cell line Panc1, thereby mediating the epithelial–mesenchymal transition necessary for the acquisition of the invasive ability of cells [492]. A similar situation, along with strengthening of the epithelial–mesenchymal transition mediated by the activation of NF- $\kappa$ B pathway signaling, is shown in gastric carcinoma, where high levels of expression of the PFKFB3 gene are positively correlated with tumor size, differentiation, invasion, metastasis, and poor patient survival [493]. In the work conducted by Moon et al., it was shown that the constitutive activation of 6-phosphofructo-2-kinase, which is found in the cells of L-lymphatic nodular carcinoma of the prostate, leads to a high rate of glycolysis and, as a result, plays an important role in stimulating androgen-induced lipogenesis [494], a process necessary for the proliferation of neoplastic cells [495]. The role of 6-phosphofructo-2-kinase as an activator of the glycolytic rearrangement of neoplastic cells is also shown in cancers of the central nervous system. Thus, the levels of the PFKFB3 gene are significantly higher in high-grade gliomas (G3 and G4 degrees of differentiation) than in non-pathological brain tissues or gliomas with G1 and G2 degrees of differentiation [496].

Pyruvate kinase is an enzyme that catalyzes the third physiologically irreversible glycolysis reaction, namely, the conversion of phosphoenolpyruvate to form one molecule of pyruvate and generate one ATP molecule. Among the various isoforms, the M2 isoform (PKM2) has attracted the most attention as a target for potential anti-neoplastic therapeutic agents due to its increased expression in transformed cells [497]. Since pyruvate kinase M2 is widely expressed during embryogenesis and regeneration, it is clear that its enzymatic activity abilities are extremely important in actively proliferating cells, such as tumor cells [498]. As a prognostic and diagnostic marker of malignant neoplasms, PKM2 plays a critical role in maintaining the metabolic program of neoplastic cells in non-muscle-invasive and highly differentiated muscle-invasive bladder carcinomas [499], colorectal cancer [500], ovarian carcinoma [501], etc. In addition to its involvement in the development of solid tumors, in an evaluation of the blood plasma of patients with acute myeloid leukemia, acute lymphoblastic leukemia (ALL) showed significantly higher levels of PKM 2, the values of which were negatively correlated with the prognosis of survival [502]. Moreover, Wang et al. found that the oncogenic functions of PKM2 are implemented not only because of the ability of the enzyme to enhance glycolysis but also to mediate the activation of autophagy by increasing the phosphorylation of the key protein of the early process-initiation Beclin-1 [503], contributing to cell survival in AML.

Since the exposure to the glycolysis process in general and directly to glycolytic enzymes can increase the efficacy of existing treatment strategies by sensitizing tumor cells to therapy, the selective targeting of specific enzymes or isoforms of enzymes is a promising area for the development of new anticancer drugs.

#### *4.3. Correction of Anomalies in Oxidative Phosphorylation in Mitochondria as a Promising Therapeutic Approach in the Development of Neuroprotective Drugs*

Maintaining mitochondrial integrity and function is of the highest priority for neuronal cells, as neurons are both key consumers of ATP and provide its highest yield [504]. There is strong evidence demonstrating that, as a result of oxidative phosphorylation disorders, the cerebral hypometabolism precedes clinical manifestations of neurodegenerative conditions [505]. This provides a rationale for corrections aimed at the metabolism and mitochondrial function constituting a potential strategy for modifying the diseases by blocking their progression.

The analysis of brain specimens from patients with various forms of affective and neurodegenerative pathologies shows disorders of mitochondrial functions as dysfunctions of electron transport and oxidative phosphorylation. In particular, the NADH dehydrogenase and cytochrome C oxidase complexes are particularly impaired in bipolar disorder [506,507], schizophrenia [508,509], Parkinson's disease [510,511], and Alzheimer's

disease [512,513]. A large-scale study by Lunnon et al. [514] showed that a significant decrease in the activity of complex I was observed in temporal cortex specimens obtained from patients with Alzheimer's disease; this was accompanied by the lower expression of the NDUFA1/4/7-9, NDUFB2/3/6, and NDUF3/4/5 subunits in the blood. Meanwhile, the activity of cytochrome C oxidase was lower in the frontal, motor, occipital, parietal, and temporal lobes of the cerebral cortex, as well as in the hippocampal regions, compared to clinically healthy controls. It is clear that these lobes of the brain are potentially involved in cognitive decline in Alzheimer's disease [515–518], suggesting their critical contribution to neurodegeneration. Perluigi et al. [519] reported a close correlation between the level of mitochondrial dysfunction and neuronal death in the hippocampal and polysensory regions of the neocortex, as well as in the entorhinal cortex.

The role of the other three complexes—succinate dehydrogenase, cytochrome reductase, and ATP synthase—in the formation of the pathological phenotype of these diseases should not be underestimated, but, to date, they have been studied to a much lesser extent than complexes I and IV. Nevertheless, interesting results were obtained in parallel evaluations of the activity of NADH + cytochrome with reductase complexes in the work of Francis et al. [520], a study conducted in transgenic TgCRND8 mice at 45 weeks of age. The authors were able to identify bioenergetic insufficiency in the brains of animals modeling Alzheimer's disease, comprising a decrease in the activity of complexes I and III and an impairment of ATP levels. A proteomic study of specimens of frozen brain tissue from patients with this neurodegenerative disorder showed a pattern of low activity of complexes I and III and ATP synthase in the early stages of Alzheimer's disease, while dysfunction of the cytochrome C oxidase complex was observed in later stages [521]. In turn, a study by Emmerzaal et al. [522] showed an age-related decrease in mitochondrial complex II activity in APP<sup>swe</sup>/PS1 $\Delta$ E9 mice aged 9 to 14 months. Interestingly, the cognitive dysfunction observed in animals of this line was observed exactly in the age interval of 8–10 months [523], which suggests the critical role of this mitochondrial abnormality in cognitive decline in Alzheimer's disease. Another study showed a parallel decrease in the activity of the succinate dehydrogenase complex in the brains of APP/PS1 mice [524], confirming the significant contribution of this complex to the energy function of mitochondria.

There are numerous disputes about whether anomalies in the functioning of the mitochondrial electron transport chain occur at the earliest stages of disease development, even before the appearance of any histopathological abnormalities, or whether they depend on the direct influence of previous pathological signs, mainly toxic deposits of  $\beta$ -amyloid peptide ( $A\beta$ ). The way in which the interaction between mitochondria and  $A\beta$  influences the pathogenesis of Alzheimer's disease is still largely unknown. There is a significant amount of evidence that  $A\beta$ , accumulating in the mitochondrial matrix, plays an important role in mitochondrial collapse and the neuronal damage mediated by the dysfunction of these organelles in Alzheimer's disease [525–528]. By inhibiting the enzymatic activity of complexes III and IV, toxic forms of  $\beta$ -amyloid peptide significantly inhibit mitochondrial respiration, thereby reducing the production of ATP [529]. In addition,  $A\beta$  has been shown to be colocalized with the  $\alpha$  subunit of ATP synthase in the cortex and hippocampal regions of the brains of mice modeling Alzheimer's disease, leading to a decrease in extracellular ATP levels due to damage to the electron transport chain and, as a result, increased long-term depression in synapses [530]. Meanwhile, the opposite situation has been well proven, when mitochondrial dysfunction provokes  $A\beta$  toxicity [531,532]. In particular, in the studies of Emmerzaal [522] and Djordjevic [533], decreases in the activity of complex II in APP/PS1 animal models and protein levels in all five subunits of electron transport chain complexes in 3xTg mice were observed even before the appearance of  $\beta$ -amyloid plaques, detected later in life. In addition, the critical role of impaired NADH dehydrogenase activity in the pathological  $A\beta$  mechanisms has been proven in a cell model of SH-SY5Y that overexpresses the Swedish mutation of the human amyloid pre-

cursor protein, where rotenone-induced dysfunction of mitochondrial complex I increased  $\beta$ -amyloid toxicity [534], allowing us to consider mitochondrial dysfunction as the starting point of the amyloid cascade.

The literature is also rich in examples showing the synergy of another biomarker of Alzheimer's disease, phosphorylated tau protein, and defects in electron transport chain complexes in the formation of disease pathogenesis [535]. In an in vivo model of genetically modified animals overexpressing the mutant human tau protein P301L, the accumulation of hyperphosphorylated tau led to a decrease in the activity of the NADH-dehydrogenase complex and impaired ATP production [536]. Later, Rhein et al. demonstrated the tau-dependent dysregulation of complex I in transgenic pR5 mice at the age of eight months [537] with abnormally phosphorylated protein [538], thus confirming the close relationship between the two main pathological features of Alzheimer's disease. In turn, Yamada et al. [539] showed that the injection of the mitochondrial complex I inhibitor annonacin in mice expressing the R406W-tau mutation led to an increase in the number of neurons with hyperphosphorylated tau protein in the frontal and parietal lobes, as well as in the hippocampal region of the brain.

However, regardless of which process has the function of a trigger, nowadays, there are extremely powerful arguments proving that an impairment of oxidative phosphorylation causes a general bioenergetic crisis in neurons, leading to cell death under neurodegenerative conditions. Moreover, since the normal function of neuronal cells requires well-coordinated mechanisms, which, in turn, require an appropriate supply of energy substrates, focusing on the pathological process as a therapeutic target constitutes a sensible direction in the development of potential neuroprotective drugs.

#### 4.4. Determination of the Main Metabolic Processes of the Cell and Energy Metabolism

Targeting the glycolysis process is an appealing prospect for therapeutic interventions in the treatment of malignant neoplasms. To date, several approaches to modulating metabolism in tumor cells have been outlined (Table 3).

**Table 3.** Therapeutic potential of promising modulators of glycolytic function.

Therapeutic Agent	Key Target	Molecular Mechanisms of Action	Prognostic Significance	Disease
WZB117	GLUT1	Cell cycle arrest, necrotic cell death	Tumor size reduction in a xenograft mouse model [540]	NSCLC
			Decreased cell viability [541]	NB
		Self-renewal of stem cell obstruction	Tumor initiation inhibition in a xenograft mouse model [542]	PC
		Blocking the STAT3/PKM2 pathway	Overcoming resistance to apatinib [543]	M
		AMPK activation, blocking the mTOR pathway, increased Bax and Bcl-2 translocation in mitochondria	Increased sensitivity to Adriamycin and radiation [544]	BC
GRg3	GLUT1, GLUT4	IL-6/STAT3/p-STAT3 pathway inhibition, MDSC suppression, CAF and collagen fibers decrease, cell death	Overcoming resistance to paclitaxel in an in vivo xenograft model [545]	BC
			Decrease in the level of AKR1C1	Induction of cell death, overcoming resistance to bortezomib [546]
Methyl jasmonate	HK2	Opening of the mPTP due to dissociation of the HK2/VDAC1 complex, triggering apoptotic cell death	Increased cell sensitivity to 5-fluorouracil, Adriamycin, and sorafenib in a xenograft mouse model [547]	HCC
3PO	PFKFB3	Decreased survivin expression, c-IAP1 and c-IAP2, NF- $\kappa$ B activation	Cell death induction [548]	EOC
Shikonin	PKM2	Decrease in Bcl-2 expression, apoptotic cell death	Increasing the therapeutic effect of cisplatin [549,550]	BC
		Exosome secretion inhibition		NSCLC

Oncological diseases

DNA damage, decreased in BRCA1	Overcoming resistance to Olaparib [501]	EOC
PKM2/STAT3 pathway inhibition	Reduced tumor growth in an in vivo xenograft model [551]	ESCC

Abbreviations: NSCLC, non-small-cell lung cancer; NB, neuroblastoma; PC, pancreatic cancer; M, melanoma; BC, breast cancer; GIST, gastrointestinal stromal tumor; MM, multiple myeloma; HCC, hepatocellular carcinoma; EOC, epithelial ovarian carcinoma; BC, bladder cancer; ESCC, esophageal squamous cell carcinoma; AMPK, 5'AMP-activated protein kinase; MDSCs, myeloid-derived suppressor cells; CAF, cancer-associated fibroblasts.

One of the most obvious strategies is the use of agents that block the glucose transporter GLUT, thereby preventing glucose from entering the cell and disrupting the glycolytic pathway.

Just over 10 years ago, Liu et al. identified a small molecule inhibitor, GLUT1 WZB 117 [540], of which studies of the antitumor activity continue to this day. During this period, research teams were able to detect a wide range of antineoplastic properties in WZB117 in various models of malignant neoplasms. In particular, in their first work [540], the authors reported that, by blocking GLUT1, this compound deprives transformed cells of glucose as an energy source and reduces the levels of intracellular ATP and glycolytic enzymes, which are accompanied by cell cycle arrest and the necrotic death of neoplastic cells. All of this mediated a reduction in tumor size in NU/J nude mice with a xenograft model of non-small-cell lung cancer by more than 70%. Similar properties of WZB117 were shown in an evaluation of the effect of the compound on the regulation of the metabolism and the viability of the neuroblastoma cell line [541]. Thus, due to the effect on GLUT1, the treatment of SH-SY5Y with WZB117 reduced the contents of glycolytic metabolites and ATP, inducing cell cycle arrest and the initiation of cell death up to necrosis. Notably, as a specific inhibitor of GLUT1, WZB117 also inhibits the self-renewal of cancer cells [542], one of the typical properties of which is intense glycolysis [552].

Years later, the range of positive properties of WZB117 was expanded, as evidenced by a number of studies on its chemosensitizing properties [544,553–556], as the involvement of GLUT1 in the development of drug resistance is well known [557,558]. In particular, the use of WZB117 allowed colon cancer cells to overcome resistance to the action of 5-fluorouracil [555], Adriamycin [556], and radiation [544] in the case of the human breast cancer cells MCF-7/ADR and MDA-MB-231/MCF-7, imatinib in gastrointestinal stromal tumors [559], and apatinib against the human melanoma cells A375 and SK-MEL-28 [543].

Ginsenoside GRg 3, derived from *Panax ginseng*, has the ability to significantly reduce the proteins GLUT 1 (~50%) and GLUT4 (~40%), which are overexpressed in tumor cells HGC-27 (derived from the metastatic lymph nodes of gastric carcinoma) and AGS (atypical glandular cells of gastric cancer) [560]. Clearly, this may be one of the possible mechanisms discovered earlier for GRg3 antitumor properties. Interestingly, in a recently published study by Zhu et al. [545], GRg 3 was used as an active ingredient to develop unique liposomes loaded with the known cytostatic paclitaxel (Rg3-PTX-LP). Thus, by targeting GRg3 to GLUT1 [561], the authors were able to reverse the resistance of cells of tumorous origin MCF 7/T to paclitaxel and achieve more than 90% tumor inhibition.

In the work of Chen et al. [562], owing to comprehensive in vitro and in vivo analyses of the antitumor properties of SMI 277—a new modulator of GLUT1—the authors were able to detect the high inhibitory ability of the compound in relation to the process of glucose uptake, which was confirmed by a decrease in lactate production. This mechanism of action of SMI277 effectively inhibited proliferation and triggered the apoptotic death of the transformed cells, which, in in vivo experiments in a mouse model, was accompanied by a decrease in tumor size by more than 50%.

Based on the above findings, it is easy to see that studies of the antitumor properties of the current GLUT inhibitors are in the initial stages. To date, the FDA has not approved a single GLUT modulator, and its development and implementation in clinical practice

remain serious problems due to the widespread expression of these proteins and the impossibility of targeting only those localized in tumor cells. This has led to the use of alternative approaches in modulating the metabolism of malignant neoplasms; among these, the effect on the key glycolytic enzymes mentioned in the previous subsection is promising.

As one of the promising inhibitors of hexokinase 2, lonidamine was recently assessed, passing the third phase of clinical trials, but it was never introduced into clinical practice due to its high systemic toxicity [563]. However, the literature is rich in experimental data obtained in the study of the antitumor properties of lonidamine derivatives. The introduction of the lonidamine motif to the structure of organometallic ruthenium led to a significant increase in cytotoxic properties in relation to the MCF 7 cell line, mediated by the activation of apoptotic caspases [564]. Additionally, the combination of cisplatin, tegafur, and lonidamine in one molecule demonstrated excellent cytotoxic properties on the triple-negative breast cancer cell line due to DNA damage and the disruption of the metabolic state of cells [565].

The role of jasmonates in the inhibition of hexokinase 2 and, as a result, their exertion of antitumor effects, have been well described in the literature [566]. In particular, the derivative methyl jasmonate represents a promising strategy for the treatment of multiple myeloma [546]. Thus, treatment with a compound of cells obtained from patients with this disease led to the impaired activity of HK2, a decrease in ATP production, and subsequent death due to the dysfunction of intracellular oncogenic signaling pathways. These properties of methyl jasmonate have prompted researchers to consider the possibility of using it as an adjuvant therapy. In the work of Klippel et al. [546], the combination of methyl jasmonate with bortezomib, which is a known proteasome inhibitor [567], demonstrated synergistic action in cell models of multiple myeloma MM.1S and INA6, which may indicate the possibility of using this agent in the treatment of hematological diseases. The chemosensitizing properties of methyl jasmonate have also been shown when combined with 5-fluorouracil, Adriamycin, and sorafenib in the treatment of hepatocellular carcinoma [547]. Of particular interest are the results obtained in [546], where the authors found that the inhibitory effect of methyl jasmonate on the glycolysis process is selective for cells of tumorous origin, which explains the absence of severe damage to the liver, lungs, and kidneys, and is an absolute advantage for future clinical use.

There is a relatively small pool of data on the inhibitors of two other key glycolysis enzymes, 6-phosphofructo-2-kinase and pyruvate kinase. However, in the work of Jiang et al. [548], (2 E)-3-(3-Pyridinyl)-1-(4-pyridinyl)-2-propen-1-one (3 PO), as a modulator of PFKFB3 activity, led to a decrease in lactate production and in the expression of the anti-apoptotic proteins survivin, c-IAP1, and c-IAP 2, as well as the activation of the NF- $\kappa$ B-mediated signaling pathway, leading to the death of A2780CP ovarian cancer cells. The chemotherapeutic potential of 3PO was also confirmed in [568], where the inhibition of PFKFB3 resulted in the effective suppression of the proliferation of a number of cell lines of tumorous origin, which was accompanied by a decrease in the intracellular concentrations of fructose-2,6-bisphosphate, lactate, and ATP. Despite the positive properties of 3PO described above, the mechanism of PFKFB3's inhibitory action is questionable, as recent studies have found that this agent does not interact with PFKFB3, and the effects it shows may not be due to the action of this enzyme at all [569,570]. This set the direction for new work in the field, prompting researchers to design selective modulators of 6-phosphofructo-2-kinase; AZ67 [571] was developed as a result, the uniqueness of which is explained by the fact that, due to its targeted binding to PFKFB3, AZ67 in critically low doses implements an antiangiogenic effect without affecting glycolysis.

Approximately 100 publications were found in the PubMed search engine when processing the query "shikonin-pkm2". Thus, it is possible to comprehensively analyze the promising results obtained by teams of authors on the inhibition of pyruvate kinase by this representative of the M2 isoform and its associated therapeutic effects. Among the most prominent is the work of Wang et al. [549], in which, due to the addition of shikonin

to the standard treatment regimen for bladder cancer, the overactivation of PKM 2 was convincingly proven. The authors were able to enhance the therapeutic effect of cisplatin in cytostatic-resistant T24 cells. This ability of shikonin to lead to cell death due to necrosis or apoptosis was accompanied by disturbances in the regulation of proteins of the Bcl-2 family. A similar effect has also been shown in a model of non-small-cell lung cancer, where the increased sensitivity to cisplatin by shikonin may be due to the inhibition of exosome secretion [550]—direct participants in the formation of the resistant phenotype of tumor cells [572]. The inhibition of pyruvate kinase activity by shikonin in an ovarian cancer model enhanced the antitumor properties of Olaparib [501], reaffirming the adjuvant potential of a naturally occurring product. In turn, the evaluation of the effects of shikonin in esophageal squamous cell carcinoma in an *in vivo* xenotransplant model showed the ability of the agent to inhibit cell proliferation by blocking PKM 2 and, as a result, aerobic glycolysis by regulating the PKM 2/STAT3 signaling pathway [551]. A pool of data was discovered for shikonin, describing the current state of research on its anti-tumor properties due to its PKM2-inhibitory ability; these data allow us to anticipate the rapid introduction of the agent into preclinical and clinical trials in order to confirm its pharmacological prospects and bring to the market a new drug with an exceptional therapeutic profile.

As mentioned earlier, disorders of energy metabolism are a typical feature of a wide range of diseases, especially neurodegenerative disorders. The central target for the action of therapeutic agents with a neuroprotective type of activity is mitochondria, the dysfunction of which is involved in various pathological cascades in NDD. Meanwhile, at first glance, the concept of strengthening mitochondrial bioenergy by activating the work of electron transport chain complexes seems obvious. In particular, for the metabolite of soy isoflavone daidzein S-equol, the ability to potentiate the functions of brain mitochondria was found to arise due to the positive regulation of the activity of the IV complex [573], which directly correlates with the protective effect of the compound in the modeling of A $\beta$ -induced neurotoxicity [574].

However, an equally interesting approach in the development of neuroprotective drugs is the production of drugs with an inverse function, the so-called “soft” inhibitors of mitochondrial respiration. The positive effects exhibited by such agents may be due to two phenomena: (1) the decrease in the production of mitochondrial ROS and (2) the development of adaptive mechanisms acquired by organelles in response to a kind of positive mobilizing stress. One of the most prominent examples of such compounds is a drug with antihyperglycemic activity: metformin. As a reversible inhibitor of the activity of the NADH dehydrogenase complex, metformin leads to the induction of chaperone-mediated autophagy in transgenic APP/PS1 mice, which mediates a decrease in the number of A $\beta$  deposits in the brain and prevents the development of a pathological behavioral phenotype [575]. Moreover, a number of clinical trials have already been conducted that aim to evaluate the effect of metformin on the cognitive functions of patients. In particular, in a meta-analysis performed by Zhang et al. [576], it was found that this drug significantly improved the neurobehavioral profile of patients with type 2 diabetes.

Thus, based on the above findings, it can be concluded that a number of studies have indeed demonstrated the efficacy of the therapeutic approach that consists of modulating metabolic shifts in malignant neoplasms and neurodegenerative disorders, thereby confirming its scientific justification. However, the preclinical and clinical evaluations of metabolic modulators for the treatment of these diseases remain in a relatively embryonic condition, which only increases the interest in continuing research in order to explore the potential of therapeutic options. Already, we can be sure that the development of highly specific drug agents, aimed at the metabolism of transformed cells in cancer diseases and neurons in neurodegenerative disorders, will create a completely new range of tools for use against these diseases.

## 5. Conclusions

The relationship between oncological diseases and neurodegenerative disorders is extremely complex. However, in recent years, convincing scientific data have been accumulated indicating the contribution of a number of etiological factors and pathophysiological processes to the pathogenesis of these two radically different diseases, thereby demonstrating an intriguing relationship between oncology and neurodegeneration.

Based on our detailed review of critical molecular mechanisms that simultaneously contribute to the development of both types of diseases, several important points can be identified.

The conceptual origin of the term “oxidative stress” can be traced back to the second half of the twentieth century. It was then that the researchers wondered about the fleeting nature of free radicals and the molecular mess caused by their action. In this regard, for more than 50 years, the concept of oxidative stress has largely focused only on the biology of free radicals, considered trigger factors of a cascade of pathological reactions associated mainly with the aging process. In recent years, the idea of oxidative stress has evolved significantly. It has been proven that the increase in its intensity leads to fatal consequences in a wide range of chronic and acute diseases, especially, paradoxically, two diametrically different pathologies: oncological and neurodegenerative. Moreover, over the past 20 years, the relevant research has focused on the most promising paradigms based on the components of the cell’s own intrinsic antioxidant defense system and intracellular signaling pathways regulating the production of free radicals. Because free radicals are key regulatory factors in the molecular mechanisms of the pathogenesis of malignant neoplasms and neurodegenerative disorders, targeting various chains of oxidative stress can help to determine a rational therapeutic strategy for these diseases. The development of effective therapeutic strategies aimed at modulating oxidative stress in various types of diseases is an important link in the creation of treatment protocols. The most promising profile of this activity type has been demonstrated for products of natural origin. Due to direct or indirect regulation of intracellular levels of reactive oxygen species, they are able to have therapeutic effects. In this regard, the modification of compounds of this class, which are valuable sources of biologically active molecules, is certainly considered as a priority direction in the creation of drugs.

Epigenetic modifications are another molecular determinant that has obvious cross-functional pathways in the pathogenesis of oncological and neurodegenerative diseases. Remarkably, the curiosity shown in the middle of the last century about the enzymatic activity that catalyzes the removal of acetyl groups from histone proteins of calf thymus extract turned into the scientific community’s pronounced interest in the field of epigenetics, playing a critical role in understanding the pathogenetic mechanisms of various diseases. As a result, there is already convincing evidence that a number of expression patterns of various HDAC isoforms are altered in people with oncological diseases; this plays a direct role in the manifestation of numerous pathological changes. As a result of such epigenetic modifications, neoplastic cells manage to maintain a highly proliferating tumor phenotype. At the same time, there is unequivocal evidence that HDACs regulate signaling cascades, gene activity, and the expression levels of other molecules involved in the pathogenesis of neurodegenerative diseases. It also demonstrates positive epidemiological correlations between oncology and neurodegeneration. Because there is a clear relationship between HDACs and the pathological phenotype in these human diseases, therapeutic agents that are effective against these enzymes are considered to be innovative treatment options both for monotherapy and in combination with already known drugs for the treatment of oncological and neurodegenerative disorders. Current HDAC inhibitors have rather low specificity, which limits their therapeutic effectiveness due to significant side effects. In this regard, one of the priority directions in the creation of HDACi as therapeutic agents for neurodegenerative and oncological diseases treatment is the development of selective agents targeting specific isoforms of the enzyme. This feature could

lead to the active targeting of specific pathological signs and increase the chances of candidates for success in the clinic.

Disorders in cellular metabolism are also of fundamental importance in future research in the search for additional biomarkers and the development of new therapeutic strategies for the treatment of these diseases. Approximately 30 Nobel Prizes have been awarded for work related to the studies of the mechanisms of metabolic reactions, the understanding of which has, over hundreds of years (since the first mention in the work of Ibn al-Nafis—XIII century AD), expanded from individual enzymes and metabolites to complex pathways that have a close relationship and dependence on each other. Such discoveries allowed us to define metabolism in detail, which helped us to understand metabolic changes in various human diseases. To date, the range of pathological conditions associated with disorders in the glycolysis and oxidative phosphorylation processes has been described in many review and experimental papers. Thus, the bioenergetic hypothesis of the pathogenesis of malignant neoplasms covers a variety of altered metabolic pathways that provide tumor cells with the energy necessary for intensive proliferation and resistance to apoptosis. This is achieved due to the property of metabolic heterogeneity, when neoplastic cells, producing ATP, are equally oriented to aerobic glycolysis and oxidative phosphorylation. In turn, in neurodegenerative diseases, mitochondrial respiration is a vulnerability, the dysfunction of which describes successive events leading to a decrease in cognitive function ranging from metabolic deficiency to neuronal death. In this regard, the multifaceted modulation of metabolic pathways is considered a modern therapeutic solution for the treatment of these diseases. A number of experimental and pre-clinical studies have convincingly proven the role of cellular metabolism modulators in the treatment of cancer and neurodegenerative disorders. Based on our analysis of existing data, a critical conclusion can be drawn: real clinical success of such drugs can only be achieved by developing agents with a very high degree of molecular target specificity; which is mainly due to the ubiquity of metabolic enzymes and transporters.

Thus, increasing the awareness of researchers based on our comprehensive analysis of the possible relationship between oncological diseases and neurodegenerative disorders, as well as a review of the current trends in research and in the clinical application of the most promising therapeutic platforms of various chemical structures, can contribute to the development of both advanced diagnostic tools and innovative methods of treating these diseases.

**Author Contributions:** Conceptualization, M.N. and Y.A.; investigation, Y.A.; supervision, M.N.; data curation, Y.A.; visualization, M.N.; writing—original draft preparation, Y.A.; writing—review and editing, M.N.; funding acquisition, M.N. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the Ministry of Science and Higher Education of the Russian Federation at the FRC Kazan Scientific Center (grant No. 075-15-2022-1128).

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Acknowledgments:** We would like to sincerely apologize to those colleagues whose work could not be quoted due to lack of space in the text of the manuscript. However, we wholeheartedly thank everyone for their invaluable contributions to this topic. We also thank BioRender.com, which made it possible to create unique drawings. This research was supported by the Centre for Collective Use of IPAC RAS (IPAC research topic FFSN-2021-0013).

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Zugazagoitia, J.; Guedes, C.; Ponce, S.; Ferrer, I.; Molina-Pinelo, S.; Paz-Ares, L. Current Challenges in Cancer Treatment. *Clin. Ther.* **2016**, *38*, 1551–1566.
2. Passeri, E.; Elkhoury, K.; Morsink, M.; Broersen, K.; Linder, M.; Tamayol, A.; Malaplate, C.; Yen, F.T.; Arab-Tehrany, E. Alzheimer's Disease: Treatment Strategies and Their Limitations. *Int. J. Mol. Sci.* **2022**, *23*, 13954.
3. Schiliro, C.; Firestein, B.L. Mechanisms of Metabolic Reprogramming in Cancer Cells Supporting Enhanced Growth and Proliferation. *Cells* **2021**, *10*, 1056.
4. Katsnelson, A.; De Strooper, B.; Zoghbi, H.Y. Neurodegeneration: From cellular concepts to clinical applications. *Sci. Transl. Med.* **2016**, *8*, 364ps18.
5. Lanni, C.; Masi, M.; Racchi, M.; Govoni, S. Cancer and Alzheimer's disease inverse relationship: an age-associated diverging derailment of shared pathways. *Mol. Psychiatry* **2021**, *26*, 280–295.
6. French, P.W. Unfolded p53 in non-neuronal cells supports bacterial etiology of Alzheimer's disease. *Neural Regen. Res.* **2022**, *17*, 2619–2622.
7. Lacroix, M.; Riscal, R.; Arena, G.; Linares, L.K.; Le Cam, L. Metabolic functions of the tumor suppressor p53: Implications in normal physiology, metabolic disorders, and cancer. *Mol. Metab.* **2020**, *33*, 2–22.
8. Liu, J.; Liu, W.; Yang, H. Balancing Apoptosis and Autophagy for Parkinson's Disease Therapy: Targeting BCL-2. *ACS Chem. Neurosci.* **2019**, *10*, 792–802.
9. Callens, M.; Kraskovskaya, N.; Derevtsova, K.; Annaert, W.; Bultynck, G.; Bezprozvanny, I.; Vervliet, T. The role of Bcl-2 proteins in modulating neuronal Ca<sup>2+</sup> signaling in health and in Alzheimer's disease. *Biochim. Biophys. Acta Mol. Cell Res.* **2021**, *1868*, 118997.
10. Pemberton, J.M.; Pogmore, J.P.; Andrews, D.W. Neuronal cell life, death, and axonal degeneration as regulated by the BCL-2 family proteins. *Cell Death Differ.* **2021**, *28*, 108–122.
11. Fairlie, W.D.; Lee, E.F. Co-Operativity between MYC and BCL-2 Pro-Survival Proteins in Cancer. *Int. J. Mol. Sci.* **2021**, *22*, 2841.
12. Opferman, J.T. Attacking cancer's Achilles heel: antagonism of anti-apoptotic BCL-2 family members. *FEBS J.* **2016**, *283*, 2661–2675.
13. Jin, J.; Xiong, Y.; Cen, B. Bcl-2 and Bcl-xL mediate resistance to receptor tyrosine kinase-targeted therapy in lung and gastric cancer. *Anticancer Drugs* **2017**, *28*, 1141–1149.
14. Thomalla, D.; Beckmann, L.; Grimm, C.; Oliverio, M.; Meder, L.; Herling, C.D.; Nieper, P.; Feldmann, T.; Merkel, O.; Lorsy, E.; et al. Deregulation and epigenetic modification of BCL2-family genes cause resistance to venetoclax in hematologic malignancies. *Blood* **2022**, *140*, 2113–2126.
15. Kaloni, D.; Diepstraten, S.T.; Strasser, A.; Kelly, G.L. BCL-2 protein family: attractive targets for cancer therapy. *Apoptosis* **2023**, *28*, 20–38.
16. Jelic, M.D.; Mandic, A.D.; Maricic, S.M.; Srdjenovic, B.U. Oxidative stress and its role in cancer. *J. Cancer Res. Ther.* **2021**, *17*, 22–28.
17. Klaunig, J.E. Oxidative Stress and Cancer. *Curr. Pharm. Des.* **2018**, *24*, 4771–4778.
18. Sosa, V.; Moline, T.; Somoza, R.; Paciucci, R.; Kondoh, H.; ME, L.L. Oxidative stress and cancer: an overview. *Ageing Res. Rev.* **2013**, *12*, 376–390.
19. Santos, R.; Ruiz de Almodovar, C.; Bulteau, A.L.; Gomes, C.M. Neurodegeneration, neurogenesis, and oxidative stress. *Oxid. Med. Cell Longev.* **2013**, *2013*, 730581.
20. Xu, S.C.; Chen, Y.B.; Lin, H.; Pi, H.F.; Zhang, N.X.; Zhao, C.C.; Shuai, L.; Zhong, M.; Yu, Z.P.; Zhou, Z.; et al. Damage to mtDNA in liver injury of patients with extrahepatic cholestasis: the protective effects of mitochondrial transcription factor A. *Free Radic. Biol. Med.* **2012**, *52*, 1543–1551.
21. Lin, J.C.; Wang, X.Z.; Shen, T.; Zhang, J.Y. iTRAQ-based quantitative analysis reveals the mechanism underlying the changes in physiological activity in a glutamate racemase mutant strain of *Streptococcus mutans* UA159. *Mol. Biol. Rep.* **2020**, *47*, 3719–3733.
22. Pavlova, N.N.; Thompson, C.B. The Emerging Hallmarks of Cancer Metabolism. *Cell Metab.* **2016**, *23*, 27–47.
23. Yin, F.; Sancheti, H.; Patil, I.; Cadenas, E. Energy metabolism and inflammation in brain aging and Alzheimer's disease. *Free Radic. Biol. Med.* **2016**, *100*, 108–122.
24. Goldsamt, A.; Damayanti, N.P.; De Nigris, F.; Pili, R. Epigenetic Dysregulation in Advanced Kidney Cancer: Opportunities for Therapeutic Interventions. *Cancer J.* **2020**, *26*, 399–406.
25. Maity, S.; Farrell, K.; Navabpour, S.; Narayanan, S.N.; Jarome, T.J. Epigenetic Mechanisms in Memory and Cognitive Decline Associated with Aging and Alzheimer's Disease. *Int. J. Mol. Sci.* **2021**, *22*, 12280.
26. Zhang, Q.; Guo, S.; Zhang, X.; Tang, S.; Shao, W.; Han, X.; Wang, L.; Du, Y. Inverse relationship between cancer and Alzheimer's disease: a systemic review meta-analysis. *Neurol. Sci.* **2015**, *36*, 1987–1994.
27. Driver, J.A.; Beiser, A.; Au, R.; Kreger, B.E.; Splansky, G.L.; Kurth, T.; Kiel, D.P.; Lu, K.P.; Seshadri, S.; Wolf, P.A. Inverse association between cancer and Alzheimer's disease: results from the Framingham Heart Study. *BMJ* **2012**, *344*, e1442.
28. Gao, X.; Ning, Y. Cancer and Parkinson's disease: the odd couple. *Drugs Today* **2011**, *47*, 215–222.
29. Brieger, K.; Schiavone, S.; Miller, F.J., Jr.; Krause, K.H. Reactive oxygen species: from health to disease. *Swiss Med. Wkly.* **2012**, *142*, w13659.

30. Zhao, Y.; Hu, X.; Liu, Y.; Dong, S.; Wen, Z.; He, W.; Zhang, S.; Huang, Q.; Shi, M. ROS signaling under metabolic stress: cross-talk between AMPK and AKT pathway. *Mol. Cancer* **2017**, *16*, 79.
31. Bouchez, C.; Devin, A. Mitochondrial Biogenesis and Mitochondrial Reactive Oxygen Species (ROS): A Complex Relationship Regulated by the cAMP/PKA Signaling Pathway. *Cells* **2019**, *8*, 287.
32. Moloney, J.N.; Cotter, T.G. ROS signalling in the biology of cancer. *Semin. Cell Dev. Biol.* **2018**, *80*, 50–64.
33. Zhang, J.; Wang, X.; Vikash, V.; Ye, Q.; Wu, D.; Liu, Y.; Dong, W. ROS and ROS-Mediated Cellular Signaling. *Oxid. Med. Cell Longev.* **2016**, *2016*, 4350965.
34. Neganova, M.; Liu, J.; Aleksandrova, Y.; Klochkov, S.; Fan, R. Therapeutic Influence on Important Targets Associated with Chronic Inflammation and Oxidative Stress in Cancer Treatment. *Cancers* **2021**, *13*, 6062.
35. Forman, H.J.; Zhang, H. Targeting oxidative stress in disease: promise and limitations of antioxidant therapy. *Nat. Rev. Drug Discov.* **2021**, *20*, 689–709.
36. Dan Dunn, J.; Alvarez, L.A.; Zhang, X.; Soldati, T. Reactive oxygen species and mitochondria: A nexus of cellular homeostasis. *Redox Biol.* **2015**, *6*, 472–485.
37. Harman, D. Aging: a theory based on free radical and radiation chemistry. *J. Gerontol.* **1956**, *11*, 298–300.
38. Shadel, G.S.; Horvath, T.L. Mitochondrial ROS signaling in organismal homeostasis. *Cell* **2015**, *163*, 560–569.
39. Lu, W.; Shi, Y.; Wang, R.; Su, D.; Tang, M.; Liu, Y.; Li, Z. Antioxidant Activity and Healthy Benefits of Natural Pigments in Fruits: A Review. *Int. J. Mol. Sci.* **2021**, *22*, 4945.
40. Kang, S.W.; Lee, S.; Lee, E.K. ROS and energy metabolism in cancer cells: alliance for fast growth. *Arch. Pharm. Res.* **2015**, *38*, 338–345.
41. Grimm, A.; Eckert, A. Brain aging and neurodegeneration: from a mitochondrial point of view. *J. Neurochem.* **2017**, *143*, 418–431.
42. Kim, J.; Kim, J.; Huang, Z.; Goo, N.; Bae, H.J.; Jeong, Y.; Park, H.J.; Cai, M.; Cho, K.; Jung, S.Y.; et al. Theracurmin Ameliorates Cognitive Dysfunctions in 5XFAD Mice by Improving Synaptic Function and Mitigating Oxidative Stress. *Biomol. Ther.* **2019**, *27*, 327–335.
43. Peng, A.; Gao, Y.; Zhuang, X.; Lin, Y.; He, W.; Wang, Y.; Chen, W.; Chen, T.; Huang, X.; Yang, R.; et al. Bazhu Decoction, a Traditional Chinese Medical Formula, Ameliorates Cognitive Deficits in the 5xFAD Mouse Model of Alzheimer's Disease. *Front. Pharmacol.* **2019**, *10*, 1391.
44. Shin, S.W.; Kim, D.H.; Jeon, W.K.; Han, J.S. 4-Hydroxynonenal Immunoreactivity Is Increased in the Frontal Cortex of 5XFAD Transgenic Mice. *Biomedicines* **2020**, *8*, 326.
45. Kim, J.H.; Lim, D.K.; Suh, Y.H.; Chang, K.A. Long-Term Treatment of Cuban Policosanol Attenuates Abnormal Oxidative Stress and Inflammatory Response via Amyloid Plaques Reduction in 5xFAD Mice. *Antioxidants* **2021**, *10*, 1321.
46. Park, M.W.; Cha, H.W.; Kim, J.; Kim, J.H.; Yang, H.; Yoon, S.; Boonpraman, N.; Yi, S.S.; Yoo, I.D.; Moon, J.S. NOX4 promotes ferroptosis of astrocytes by oxidative stress-induced lipid peroxidation via the impairment of mitochondrial metabolism in Alzheimer's diseases. *Redox Biol.* **2021**, *41*, 101947.
47. Yang, Y.; Chen, W.; Wang, X.; Ge, W. Impact of mitochondrial aldehyde dehydrogenase 2 on cognitive impairment in the AD model mouse. *Acta Biochim. Biophys. Sin.* **2021**, *53*, 837–847.
48. Foroumandi, E.; Javan, R.; Moayed, L.; Fahimi, H.; Kheirabadi, F.; Neamatshahi, M.; Shogofteh, F.; Zarghi, A. The effects of fenugreek seed extract supplementation in patients with Alzheimer's disease: A randomized, double-blind, placebo-controlled trial. *Phytother. Res.* **2023**, *37*, 285–294.
49. Khalil, A.; Berrougui, H.; Pawelec, G.; Fulop, T. Impairment of the ABCA1 and SR-BI-mediated cholesterol efflux pathways and HDL anti-inflammatory activity in Alzheimer's disease. *Mech. Ageing Dev.* **2012**, *133*, 20–29.
50. Ton, A.M.M.; Campagnaro, B.P.; Alves, G.A.; Aires, R.; Coco, L.Z.; Arpini, C.M.; Guerra, E.O.T.; Campos-Toimil, M.; Meyrelles, S.S.; Pereira, T.M.C.; et al. Oxidative Stress and Dementia in Alzheimer's Patients: Effects of Synbiotic Supplementation. *Oxid. Med. Cell Longev.* **2020**, *2020*, 2638703.
51. Pena-Bautista, C.; Tirlle, T.; Lopez-Nogueroles, M.; Vento, M.; Baquero, M.; Chafer-Pericas, C. Oxidative Damage of DNA as Early Marker of Alzheimer's Disease. *Int. J. Mol. Sci.* **2019**, *20*, 6136.
52. Hasina, Z.; Wang, N.; Wang, C.C. Developmental Neuropathology and Neurodegeneration of Down Syndrome: Current Knowledge in Humans. *Front. Cell Dev. Biol.* **2022**, *10*, 877711.
53. Butterfield, D.A. Brain lipid peroxidation and alzheimer disease: Synergy between the Butterfield and Mattson laboratories. *Ageing Res. Rev.* **2020**, *64*, 101049.
54. Leu, T.; Schutzhold, V.; Fandrey, J.; Ferenz, K.B. When the Brain Yearns for Oxygen. *Neurosignals* **2019**, *27*, 50–61.
55. Gaschler, M.M.; Stockwell, B.R. Lipid peroxidation in cell death. *Biochem. Biophys. Res. Commun.* **2017**, *482*, 419–425.
56. Guan, L.; Mao, Z.; Yang, S.; Wu, G.; Chen, Y.; Yin, L.; Qi, Y.; Han, L.; Xu, L. Dioscin alleviates Alzheimer's disease through regulating RAGE/NOX4 mediated oxidative stress and inflammation. *Biomed. Pharmacother.* **2022**, *152*, 113248.
57. Meng, M.; Zhang, L.; Ai, D.; Wu, H.; Peng, W. beta-Asarone Ameliorates beta-Amyloid-Induced Neurotoxicity in PC12 Cells by Activating P13K/Akt/Nrf2 Signaling Pathway. *Front. Pharmacol.* **2021**, *12*, 659955.
58. Song, T.; Song, X.; Zhu, C.; Patrick, R.; Skurla, M.; Santangelo, I.; Green, M.; Harper, D.; Ren, B.; Forester, B.P.; et al. Mitochondrial dysfunction, oxidative stress, neuroinflammation, and metabolic alterations in the progression of Alzheimer's disease: A meta-analysis of in vivo magnetic resonance spectroscopy studies. *Ageing Res. Rev.* **2021**, *72*, 101503.

59. Schrag, M.; Mueller, C.; Zabel, M.; Crofton, A.; Kirsch, W.M.; Ghribi, O.; Squitti, R.; Perry, G. Oxidative stress in blood in Alzheimer's disease and mild cognitive impairment: a meta-analysis. *Neurobiol. Dis.* **2013**, *59*, 100–110.
60. Zabel, M.; Nackenoff, A.; Kirsch, W.M.; Harrison, F.E.; Perry, G.; Schrag, M. Markers of oxidative damage to lipids, nucleic acids and proteins and antioxidant enzymes activities in Alzheimer's disease brain: A meta-analysis in human pathological specimens. *Free Radic. Biol. Med.* **2018**, *115*, 351–360.
61. Trares, K.; Chen, L.J.; Schottker, B. Association of F(2)-isoprostane levels with Alzheimer's disease in observational studies: A systematic review and meta-analysis. *Ageing Res. Rev.* **2022**, *74*, 101552.
62. Pena-Bautista, C.; Alvarez, L.; Baquero, M.; Ferrer, I.; Garcia, L.; Hervas-Marin, D.; Chafer-Pericas, C. Plasma isoprostanoids assessment as Alzheimer's disease progression biomarkers. *J. Neurochem.* **2021**, *157*, 2187–2194.
63. Miyazawa, T.; Nakagawa, K.; Takekoshi, H.; Higuchi, O.; Kato, S.; Kondo, M.; Kimura, F.; Miyazawa, T. Ingestion of Chlorella reduced the oxidation of erythrocyte membrane lipids in senior Japanese subjects. *J. Oleo Sci.* **2013**, *62*, 873–881.
64. Streck, E.L.; Czapski, G.A.; Goncalves da Silva, C. Neurodegeneration, mitochondrial dysfunction, and oxidative stress. *Oxid. Med. Cell Longev.* **2013**, *2013*, 826046.
65. Echtay, K.S.; Esteves, T.C.; Pakay, J.L.; Jekabsons, M.B.; Lambert, A.J.; Portero-Otin, M.; Pamplona, R.; Vidal-Puig, A.J.; Wang, S.; Roebuck, S.J.; et al. A signalling role for 4-hydroxy-2-nonenal in regulation of mitochondrial uncoupling. *EMBO J.* **2003**, *22*, 4103–4110.
66. Takeda, A.; Smith, M.A.; Avila, J.; Nunomura, A.; Siedlak, S.L.; Zhu, X.; Perry, G.; Sayre, L.M. In Alzheimer's disease, heme oxygenase is coincident with Alz50, an epitope of tau induced by 4-hydroxy-2-nonenal modification. *J. Neurochem.* **2000**, *75*, 1234–1241.
67. Takagane, K.; Nojima, J.; Mitsuhashi, H.; Suo, S.; Yanagihara, D.; Takaiwa, F.; Urano, Y.; Noguchi, N.; Ishiura, S. Aβ induces oxidative stress in senescence-accelerated (SAMP8) mice. *Biosci. Biotechnol. Biochem.* **2015**, *79*, 912–918.
68. Akude, E.; Zherebitskaya, E.; Roy Chowdhury, S.K.; Girling, K.; Fernyhough, P. 4-Hydroxy-2-nonenal induces mitochondrial dysfunction and aberrant axonal outgrowth in adult sensory neurons that mimics features of diabetic neuropathy. *Neurotox. Res.* **2010**, *17*, 28–38.
69. Jiang, D.; Men, L.; Wang, J.; Zhang, Y.; Chickenyen, S.; Wang, Y.; Zhou, F. Redox reactions of copper complexes formed with different beta-amyloid peptides and their neuropathological [correction of neuropathological] relevance. *Biochemistry* **2007**, *46*, 9270–9282.
70. Firczuk, M.; Bajor, M.; Graczyk-Jarzynka, A.; Fidyk, K.; Goral, A.; Zagozdzon, R. Harnessing altered oxidative metabolism in cancer by augmented prooxidant therapy. *Cancer Lett.* **2020**, *471*, 1–11.
71. Li, L.; Tan, J.; Miao, Y.; Lei, P.; Zhang, Q. ROS and Autophagy: Interactions and Molecular Regulatory Mechanisms. *Cell Mol. Neurobiol.* **2015**, *35*, 615–621.
72. Santos, N.; Ferreira, R.S.; Santos, A.C.D. Overview of cisplatin-induced neurotoxicity and ototoxicity, and the protective agents. *Food Chem. Toxicol.* **2020**, *136*, 111079.
73. Christidi, E.; Brunham, L.R. Regulated cell death pathways in doxorubicin-induced cardiotoxicity. *Cell Death Dis.* **2021**, *12*, 339.
74. Wang, Y.; Qi, H.; Liu, Y.; Duan, C.; Liu, X.; Xia, T.; Chen, D.; Piao, H.L.; Liu, H.X. The double-edged roles of ROS in cancer prevention and therapy. *Theranostics* **2021**, *11*, 4839–4857.
75. Sadati Zarrini, A.; Moslemi, D.; Parsian, H.; Vessal, M.; Mosapour, A.; Shirkhani Kelagari, Z. The status of antioxidants, malondialdehyde and some trace elements in serum of patients with breast cancer. *Casp. J. Intern. Med.* **2016**, *7*, 31–36.
76. do Val Carneiro, J.L.; Nixdorf, S.L.; Mantovani, M.S.; da Silva do Amaral Herrera, A.C.; Aoki, M.N.; Amarante, M.K.; Fabris, B.A.; Pelegrinelli Fungaro, M.H.; Ehara Watanabe, M.A. Plasma malondialdehyde levels and CXCR4 expression in peripheral blood cells of breast cancer patients. *J. Cancer Res. Clin. Oncol.* **2009**, *135*, 997–1004.
77. Gonenc, A.; Ozkan, Y.; Torun, M.; Simsek, B. Plasma malondialdehyde (MDA) levels in breast and lung cancer patients. *J. Clin. Pharm. Ther.* **2001**, *26*, 141–144.
78. Arif, M.; Rashid, A.; Majeed, A.; Qaiser, F.; Razak, S. Evaluation of correlation between expression of P53 and Malondialdehyde levels in prostate cancer patients. *J. Pak. Med. Assoc.* **2018**, *68*, 1373–1377.
79. Dillioglugil, M.O.; Mekik, H.; Muezzinoglu, B.; Ozkan, T.A.; Demir, C.G.; Dillioglugil, O. Blood and tissue nitric oxide and malondialdehyde are prognostic indicators of localized prostate cancer. *Int. Urol. Nephrol.* **2012**, *44*, 1691–1696.
80. Drozd-Afelt, J.M.; Koim-Puchowska, B.B.; Kaminski, P. Analysis of oxidative stress indicators in Polish patients with prostate cancer. *Environ. Sci. Pollut. Res. Int.* **2022**, *29*, 4632–4640.
81. Lepara, Z.; Lepara, O.; Fajkic, A.; Rebic, D.; Alic, J.; Spahovic, H. Serum malondialdehyde (MDA) level as a potential biomarker of cancer progression for patients with bladder cancer. *Rom. J. Intern. Med.* **2020**, *58*, 146–152.
82. Gecit, I.; Eryilmaz, R.; Kavak, S.; Meral, I.; Demir, H.; Pirincci, N.; Gunes, M.; Taken, K. The Prolidase Activity, Oxidative Stress, and Nitric Oxide Levels of Bladder Tissues with or Without Tumor in Patients with Bladder Cancer. *J. Membr. Biol.* **2017**, *250*, 455–459.
83. Firdausa, A.Y.; Ahimsa, S.S.; Ahmada, R.A.; Sukmawati, N.F.; Ernawati, D.S.; Parmadiati, A.E.; Soebadi, B.; Radithia, D.; Winias, S.; Mahdani, F.Y.; et al. Malondialdehyde Level and Tissue Apoptosis Count as an Early-Detection Marker of Oral Potentially Malignant Disorders. *Eur. J. Dent.* **2023**, *17*, 155–160.
84. Marakala, V.; Malathi, M.; Shivashankara, A.R. Lipid peroxidation and antioxidant vitamin status in oral cavity and oropharyngeal cancer patients. *Asian Pac. J. Cancer Prev.* **2012**, *13*, 5763–5765.

85. Pande, D.; Negi, R.; Khanna, S.; Khanna, R.; Khanna, H.D. Vascular endothelial growth factor levels in relation to oxidative damage and antioxidant status in patients with breast cancer. *J. Breast Cancer* **2011**, *14*, 181–184.
86. Valavanidis, A.; Vlachogianni, T.; Fiotakis, C. 8-hydroxy-2'-deoxyguanosine (8-OHdG): A critical biomarker of oxidative stress and carcinogenesis. *J. Environ. Sci. Health C Environ. Carcinog. Ecotoxicol. Rev.* **2009**, *27*, 120–139.
87. Yamamoto, T.; Hosokawa, K.; Tamura, T.; Kanno, H.; Urabe, M.; Honjo, H. Urinary 8-hydroxy-2'-deoxyguanosine (8-OHdG) levels in women with or without gynecologic cancer. *J. Obstet. Gynaecol. Res.* **1996**, *22*, 359–363.
88. Pylvas, M.; Puistola, U.; Laatio, L.; Kauppila, S.; Karihtala, P. Elevated serum 8-OHdG is associated with poor prognosis in epithelial ovarian cancer. *Anticancer. Res.* **2011**, *31*, 1411–1415.
89. Xu, X.; Wang, Y.; Guo, W.; Zhou, Y.; Lv, C.; Chen, X.; Liu, K. The significance of the alteration of 8-OHdG in serous ovarian carcinoma. *J. Ovarian Res.* **2013**, *6*, 74.
90. Plachetka, A.; Adamek, B.; Strzelczyk, J.K.; Krakowczyk, L.; Migula, P.; Nowak, P.; Wiczkowski, A. 8-hydroxy-2'-deoxyguanosine in colorectal adenocarcinoma—is it a result of oxidative stress? *Med. Sci. Monit.* **2013**, *19*, 690–695.
91. Sato, T.; Takeda, H.; Otake, S.; Yokozawa, J.; Nishise, S.; Fujishima, S.; Orii, T.; Fukui, T.; Takano, J.; Sasaki, Y.; et al. Increased plasma levels of 8-hydroxydeoxyguanosine are associated with development of colorectal tumors. *J. Clin. Biochem. Nutr.* **2010**, *47*, 59–63.
92. Rozalski, R.; Gackowski, D.; Siomek-Gorecka, A.; Starczak, M.; Modrzejewska, M.; Banaszkiwicz, Z.; Olinski, R. Urinary 5-hydroxymethyluracil and 8-oxo-7,8-dihydroguanine as potential biomarkers in patients with colorectal cancer. *Biomarkers* **2015**, *20*, 287–291.
93. Sedlic, F.; Seiwerth, F.; Sepac, A.; Sikiric, S.; Cindric, M.; Milavic, M.; Batelja Vuletic, L.; Jakopovic, M.; Seiwerth, S. Mitochondrial ROS Induce Partial Dedifferentiation of Human Mesothelioma via Upregulation of NANOG. *Antioxidants* **2020**, *9*, 606.
94. Migliario, M.; Pittarella, P.; Fanuli, M.; Rizzi, M.; Reno, F. Laser-induced osteoblast proliferation is mediated by ROS production. *Lasers Med. Sci.* **2014**, *29*, 1463–1467.
95. Diebold, L.; Chandel, N.S. Mitochondrial ROS regulation of proliferating cells. *Free Radic. Biol. Med.* **2016**, *100*, 86–93.
96. Chang, C.H.; Pauklin, S. ROS and TGFbeta: from pancreatic tumour growth to metastasis. *J. Exp. Clin. Cancer Res.* **2021**, *40*, 152.
97. Cui, Q.; Wang, J.Q.; Assaraf, Y.G.; Ren, L.; Gupta, P.; Wei, L.; Ashby, C.R., Jr.; Yang, D.H.; Chen, Z.S. Modulating ROS to overcome multidrug resistance in cancer. *Drug Resist. Updat.* **2018**, *41*, 1–25.
98. Fukai, T.; Ushio-Fukai, M. Cross-Talk between NADPH Oxidase and Mitochondria: Role in ROS Signaling and Angiogenesis. *Cells* **2020**, *9*, 1849.
99. Perillo, B.; Di Donato, M.; Pezone, A.; Di Zazzo, E.; Giovannelli, P.; Galasso, G.; Castoria, G.; Migliaccio, A. ROS in cancer therapy: the bright side of the moon. *Exp. Mol. Med.* **2020**, *52*, 192–203.
100. Pisoschi, A.M.; Pop, A. The role of antioxidants in the chemistry of oxidative stress: A review. *Eur. J. Med. Chem.* **2015**, *97*, 55–74.
101. Demirci-Cekic, S.; Ozkan, G.; Avanc, A.N.; Uzunboy, S.; Capanoglu, E.; Apak, R. Biomarkers of Oxidative Stress and Antioxidant Defense. *J. Pharm. Biomed. Anal.* **2022**, *209*, 114477.
102. Grazioli, V.; Schiavo, R.; Casari, E.; Marzatico, F.; Rodriguez y Baena, R.; Gaetani, P. Antioxidant enzymatic activities and lipid peroxidation in cultured human chondrocytes from vertebral plate cartilage. *FEBS Lett.* **1998**, *431*, 149–153.
103. Forman, H.J.; Zhang, H.; Rinna, A. Glutathione: overview of its protective roles, measurement, and biosynthesis. *Mol. Aspects Med.* **2009**, *30*, 1–12.
104. Wang, Y.; Branicky, R.; Noe, A.; Hekimi, S. Superoxide dismutases: Dual roles in controlling ROS damage and regulating ROS signaling. *J. Cell Biol.* **2018**, *217*, 1915–1928.
105. Goyal, M.M.; Basak, A. Human catalase: looking for complete identity. *Protein Cell* **2010**, *1*, 888–897.
106. Glorieux, C.; Calderon, P.B. Catalase, a remarkable enzyme: targeting the oldest antioxidant enzyme to find a new cancer treatment approach. *Biol. Chem.* **2017**, *398*, 1095–1108.
107. Handy, D.E.; Loscalzo, J. The role of glutathione peroxidase-1 in health and disease. *Free Radic. Biol. Med.* **2022**, *188*, 146–161.
108. Brigelius-Flohe, R.; Flohe, L. Regulatory Phenomena in the Glutathione Peroxidase Superfamily. *Antioxid. Redox Signal* **2020**, *33*, 498–516.
109. Couto, N.; Wood, J.; Barber, J. The role of glutathione reductase and related enzymes on cellular redox homeostasis network. *Free Radic. Biol. Med.* **2016**, *95*, 27–42.
110. Averill-Bates, D.A. The antioxidant glutathione. *Vitam. Horm.* **2023**, *121*, 109–141.
111. Alanazi, A.M.; Mostafa, G.A.; Al-Badr, A.A. Glutathione. *Profiles Drug Subst. Excip. Relat. Methodol.* **2015**, *40*, 43–158.
112. Owen, J.B.; Butterfield, D.A. Measurement of oxidized/reduced glutathione ratio. *Methods Mol. Biol.* **2010**, *648*, 269–277.
113. Poljsak, B.; Milisav, I. The Role of Antioxidants in Cancer, Friends or Foes? *Curr. Pharm. Des.* **2018**, *24*, 5234–5244.
114. Zahra, K.; Patel, S.; Dey, T.; Pandey, U.; Mishra, S.P. A study of oxidative stress in cervical cancer- an institutional study. *Biochem. Biophys. Rep.* **2021**, *25*, 100881.
115. Manju, V.; Balasubramanian, V.; Nalini, N. Oxidative stress and tumor markers in cervical cancer patients. *J. Biochem. Mol. Biol. Biophys.* **2002**, *6*, 387–390.
116. Manju, V.; Kalaiyani Sailaja, J.; Nalini, N. Circulating lipid peroxidation and antioxidant status in cervical cancer patients: a case-control study. *Clin. Biochem.* **2002**, *35*, 621–625.
117. Bel'skaya, L.V.; Sarf, E.A.; Solomatin, D.V.; Kosenok, V.K. Metabolic Features of Saliva in Breast Cancer Patients. *Metabolites* **2022**, *12*, 166.

118. Cobanoglu, U.; Demir, H.; Duran, M.; Sehitogullari, A.; Mergan, D.; Demir, C. Erythrocyte catalase and carbonic anhydrase activities in lung cancer. *Asian Pac. J. Cancer Prev.* **2010**, *11*, 1377–1382.
119. Skorska, K.B.; Placzkowska, S.; Prescha, A.; Porebska, I.; Kosacka, M.; Pawelczyk, K.; Zablocka-Slowinska, K. Serum Total SOD Activity and SOD1/2 Concentrations in Predicting All-Cause Mortality in Lung Cancer Patients. *Pharmaceuticals* **2021**, *14*.
120. Pirincci, N.; Gecit, I.; Gunes, M.; Yuksel, M.B.; Kaba, M.; Tanik, S.; Demir, H.; Aslan, M. Serum adenosine deaminase, catalase and carbonic anhydrase activities in patients with bladder cancer. *Clinics* **2012**, *67*, 1443–1446.
121. Cavallini, C.; Chignola, R.; Dando, I.; Perbellini, O.; Mimiola, E.; Lovato, O.; Laudanna, C.; Pizzolo, G.; Donadelli, M.; Scupoli, M.T. Low catalase expression confers redox hypersensitivity and identifies an indolent clinical behavior in CLL. *Blood* **2018**, *131*, 1942–1954.
122. Oltra, A.M.; Carbonell, F.; Tormos, C.; Iradi, A.; Saez, G.T. Antioxidant enzyme activities and the production of MDA and 8-oxo-dG in chronic lymphocytic leukemia. *Free Radic. Biol. Med.* **2001**, *30*, 1286–1292.
123. Bansal, A.; Simon, M.C. Glutathione metabolism in cancer progression and treatment resistance. *J. Cell Biol.* **2018**, *217*, 2291–2298.
124. Liu, W.; Zhou, Y.; Duan, W.; Song, J.; Wei, S.; Xia, S.; Wang, Y.; Du, X.; Li, E.; Ren, C.; et al. Glutathione peroxidase 4-dependent glutathione high-consumption drives acquired platinum chemoresistance in lung cancer-derived brain metastasis. *Clin. Transl. Med.* **2021**, *11*, e517.
125. Estrela, J.M.; Ortega, A.; Mena, S.; Sirerol, J.A.; Obrador, E. Glutathione in metastases: From mechanisms to clinical applications. *Crit. Rev. Clin. Lab. Sci.* **2016**, *53*, 253–267.
126. Hatem, E.; El Banna, N.; Huang, M.E. Multifaceted Roles of Glutathione and Glutathione-Based Systems in Carcinogenesis and Anticancer Drug Resistance. *Antioxid. Redox Signal* **2017**, *27*, 1217–1234.
127. Nunes, S.C.; Serpa, J. Glutathione in Ovarian Cancer: A Double-Edged Sword. *Int. J. Mol. Sci.* **2018**, *19*, 1882.
128. Miran, T.; Vogg, A.T.J.; Drude, N.; Mottaghy, F.M.; Morgenroth, A. Modulation of glutathione promotes apoptosis in triple-negative breast cancer cells. *FASEB J.* **2018**, *32*, 2803–2813.
129. Skrzydlewska, E.; Sulkowski, S.; Koda, M.; Zalewski, B.; Kanczuga-Koda, L.; Sulkowska, M. Lipid peroxidation and antioxidant status in colorectal cancer. *World J. Gastroenterol.* **2005**, *11*, 403–406.
130. Kontakiotis, T.; Katsoulis, K.; Hagizisi, O.; Kougioulis, M.; Gerou, S.; Papakosta, D. Bronchoalveolar lavage fluid alteration in antioxidant and inflammatory status in lung cancer patients. *Eur. J. Intern. Med.* **2011**, *22*, 522–526.
131. Pei, S.; Minhajuddin, M.; Callahan, K.P.; Balys, M.; Ashton, J.M.; Neering, S.J.; Lagadinou, E.D.; Corbett, C.; Ye, H.; Liesveld, J.L.; et al. Targeting aberrant glutathione metabolism to eradicate human acute myelogenous leukemia cells. *J. Biol. Chem.* **2013**, *288*, 33542–33558.
132. Aoyama, K. Glutathione in the Brain. *Int. J. Mol. Sci.* **2021**, *22*, 5010.
133. Nandi, A.; Yan, L.J.; Jana, C.K.; Das, N. Role of Catalase in Oxidative Stress- and Age-Associated Degenerative Diseases. *Oxid. Med. Cell Longev.* **2019**, *2019*, 9613090.
134. Haddad, M.; Herve, V.; Ben Khedher, M.R.; Rabanel, J.M.; Ramassamy, C. Glutathione: An Old and Small Molecule with Great Functions and New Applications in the Brain and in Alzheimer's Disease. *Antioxid. Redox Signal* **2021**, *35*, 270–292.
135. Ronkina, N.; Gaestel, M. MAPK-Activated Protein Kinases: Servant or Partner? *Annu. Rev. Biochem.* **2022**, *91*, 505–540.
136. Zhang, Y.; Zhao, M.; Gao, H.; Yu, G.; Zhao, Y.; Yao, F.; Yang, W. MAPK signalling-induced phosphorylation and subcellular translocation of PDHE1 $\alpha$  promotes tumour immune evasion. *Nat. Metab.* **2022**, *4*, 374–388.
137. Behl, T.; Upadhyay, T.; Singh, S.; Chigurupati, S.; Alsubayiel, A.M.; Mani, V.; Vargas-De-La-Cruz, C.; Uivarosan, D.; Bustea, C.; Sava, C.; et al. Polyphenols Targeting MAPK Mediated Oxidative Stress and Inflammation in Rheumatoid Arthritis. *Molecules* **2021**, *26*.
138. Kim, E.K.; Choi, E.J. Pathological roles of MAPK signaling pathways in human diseases. *Biochim. Biophys. Acta* **2010**, *1802*, 396–405.
139. Kim, E.K.; Choi, E.J. Compromised MAPK signaling in human diseases: an update. *Arch. Toxicol.* **2015**, *89*, 867–882.
140. Degirmenci, U.; Wang, M.; Hu, J. Targeting Aberrant RAS/RAF/MEK/ERK Signaling for Cancer Therapy. *Cells* **2020**, *9*, 198.
141. Santarpia, L.; Lippman, S.M.; El-Naggar, A.K. Targeting the MAPK-RAS-RAF signaling pathway in cancer therapy. *Expert. Opin. Ther. Targets* **2012**, *16*, 103–119.
142. Huang, C.; Jacobson, K.; Schaller, M.D. MAP kinases and cell migration. *J. Cell Sci.* **2004**, *117 Pt 20*, 4619–4628.
143. Balmanno, K.; Cook, S.J. Tumour cell survival signalling by the ERK1/2 pathway. *Cell Death Differ.* **2009**, *16*, 368–377.
144. Chakraborti, S.; Mandal, M.; Das, S.; Mandal, A.; Chakraborti, T. Regulation of matrix metalloproteinases: an overview. *Mol. Cell Biochem.* **2003**, *253*, 269–285.
145. Xie, Y.; Shi, X.; Sheng, K.; Han, G.; Li, W.; Zhao, Q.; Jiang, B.; Feng, J.; Li, J.; Gu, Y. PI3K/Akt signaling transduction pathway, erythropoiesis and glycolysis in hypoxia (Review). *Mol. Med. Rep.* **2019**, *19*, 783–791.
146. Miricescu, D.; Totan, A.; Stanescu, S., II; Badoiu, S.C.; Stefani, C.; Greabu, M. PI3K/AKT/mTOR Signaling Pathway in Breast Cancer: From Molecular Landscape to Clinical Aspects. *Int. J. Mol. Sci.* **2020**, *22*, 173.
147. Noorolyai, S.; Shajari, N.; Baghbani, E.; Sadreddini, S.; Baradaran, B. The relation between PI3K/AKT signalling pathway and cancer. *Gene* **2019**, *698*, 120–128.
148. Chen, H.; Zhou, L.; Wu, X.; Li, R.; Wen, J.; Sha, J.; Wen, X. The PI3K/AKT pathway in the pathogenesis of prostate cancer. *Front. Biosci. (Landmark Ed)* **2016**, *21*, 1084–1091.

149. Long, H.Z.; Cheng, Y.; Zhou, Z.W.; Luo, H.Y.; Wen, D.D.; Gao, L.C. PI3K/AKT Signal Pathway: A Target of Natural Products in the Prevention and Treatment of Alzheimer's Disease and Parkinson's Disease. *Front. Pharmacol.* **2021**, *12*, 648636.
150. Razani, E.; Pourbagheri-Sigaroodi, A.; Safaroghli-Azar, A.; Zoghi, A.; Shanaki-Bavarsad, M.; Bashash, D. The PI3K/Akt signaling axis in Alzheimer's disease: a valuable target to stimulate or suppress? *Cell Stress. Chaperones* **2021**, *26*, 871–887.
151. Shin, I.; Yakes, F.M.; Rojo, F.; Shin, N.Y.; Bakin, A.V.; Baselga, J.; Arteaga, C.L. PKB/Akt mediates cell-cycle progression by phosphorylation of p27(Kip1) at threonine 157 and modulation of its cellular localization. *Nat. Med.* **2002**, *8*, 1145–1152.
152. Chen, X.; Thakkar, H.; Tyan, F.; Gim, S.; Robinson, H.; Lee, C.; Pandey, S.K.; Nwokorie, C.; Onwudiwe, N.; Srivastava, R.K. Constitutively active Akt is an important regulator of TRAIL sensitivity in prostate cancer. *Oncogene* **2001**, *20*, 6073–6083.
153. Shorning, B.Y.; Dass, M.S.; Smalley, M.J.; Pearson, H.B. The PI3K-AKT-mTOR Pathway and Prostate Cancer: At the Crossroads of AR, MAPK, and WNT Signaling. *Int. J. Mol. Sci.* **2020**, *21*, 4507.
154. Pungsrinont, T.; Kallenbach, J.; Baniahmad, A. Role of PI3K-AKT-mTOR Pathway as a Pro-Survival Signaling and Resistance-Mediating Mechanism to Therapy of Prostate Cancer. *Int. J. Mol. Sci.* **2021**, *22*, 11088.
155. Jin, Y.; Chen, Y.; Tang, H.; Hu, X.; Hubert, S.M.; Li, Q.; Su, D.; Xu, H.; Fan, Y.; Yu, X.; et al. Activation of PI3K/AKT Pathway Is a Potential Mechanism of Treatment Resistance in Small Cell Lung Cancer. *Clin. Cancer Res.* **2022**, *28*, 526–539.
156. Li, X.; Li, C.; Guo, C.; Zhao, Q.; Cao, J.; Huang, H.Y.; Yue, M.; Xue, Y.; Jin, Y.; Hu, L.; et al. PI3K/Akt/mTOR signaling orchestrates the phenotypic transition and chemo-resistance of small cell lung cancer. *J. Genet. Genom.* **2021**, *48*, 640–651.
157. Guerrero-Zotano, A.; Mayer, I.A.; Arteaga, C.L. PI3K/AKT/mTOR: role in breast cancer progression, drug resistance, and treatment. *Cancer Metastasis Rev.* **2016**, *35*, 515–524.
158. Dong, C.; Wu, J.; Chen, Y.; Nie, J.; Chen, C. Activation of PI3K/AKT/mTOR Pathway Causes Drug Resistance in Breast Cancer. *Front. Pharmacol.* **2021**, *12*, 628690.
159. An, J.; Yang, L.; Pan, Y.; He, Y.; Xie, H.; Tao, Y.; Li, W.; Yan, Y.; Chen, S.; Liu, Y.; et al. SPAG5 Activates PI3K/AKT Pathway and Promotes the Tumor Progression and Chemo-Resistance in Gastric Cancer. *DNA Cell Biol.* **2022**, *41*, 893–902.
160. Sun, S.; Guo, C.; Gao, T.; Ma, D.; Su, X.; Pang, Q.; Zhang, R. Hypoxia Enhances Glioma Resistance to Sulfasalazine-Induced Ferroptosis by Upregulating SLC7A11 via PI3K/AKT/HIF-1alpha Axis. *Oxid. Med. Cell Longev.* **2022**, *2022*, 7862430.
161. Xiang, M.; Liu, T.; Tian, C.; Ma, K.; Gou, J.; Huang, R.; Li, S.; Li, Q.; Xu, C.; Li, L.; et al. Kinsenoside attenuates liver fibroinflammation by suppressing dendritic cells via the PI3K-AKT-FoxO1 pathway. *Pharmacol. Res.* **2022**, *177*, 106092.
162. Wang, B.N.; Wu, C.B.; Chen, Z.M.; Zheng, P.P.; Liu, Y.Q.; Xiong, J.; Xu, J.Y.; Li, P.F.; Mamun, A.A.; Ye, L.B.; et al. DL-3-n-butylphthalide ameliorates diabetes-associated cognitive decline by enhancing PI3K/Akt signaling and suppressing oxidative stress. *Acta Pharmacol. Sin.* **2021**, *42*, 347–360.
163. Fabbri, S.B.; Girardi, B.A.; de Lorena Wendel, A.; Coelho Ilha Valin, C.; Pillat, M.M.; Viero, F.T.; Mello, C.F.; Rubin, M.A. Spermidine-induced improvement of memory consolidation involves PI3K/Akt signaling pathway. *Brain Res. Bull.* **2020**, *164*, 208–213.
164. Knox, D.; Della Valle, R.; Mohammadmirzaei, N.; Shultz, B.; Biddle, M.; Farkash, A.; Chamness, M.; Moulton, E. PI3K-Akt Signaling in the Basolateral Amygdala Facilitates Traumatic Stress Enhancements in Fear Memory. *Int. J. Neuropsychopharmacol.* **2021**, *24*, 229–238.
165. Li, H.; Xue, X.; Li, L.; Li, Y.; Wang, Y.; Huang, T.; Wang, Y.; Meng, H.; Pan, B.; Niu, Q. Aluminum-Induced Synaptic Plasticity Impairment via PI3K-Akt-mTOR Signaling Pathway. *Neurotox. Res.* **2020**, *37*, 996–1008.
166. Ali, T.; Kim, T.; Rehman, S.U.; Khan, M.S.; Amin, F.U.; Khan, M.; Ikram, M.; Kim, M.O. Natural Dietary Supplementation of Anthocyanins via PI3K/Akt/Nrf2/HO-1 Pathways Mitigate Oxidative Stress, Neurodegeneration, and Memory Impairment in a Mouse Model of Alzheimer's Disease. *Mol. Neurobiol.* **2018**, *55*, 6076–6093.
167. Wang, C.; Hao, J.; Liu, X.; Li, C.; Yuan, X.; Lee, R.J.; Bai, T.; Wang, D. Isoforsythiaside Attenuates Alzheimer's Disease via Regulating Mitochondrial Function Through the PI3K/AKT Pathway. *Int. J. Mol. Sci.* **2020**, *21*.
168. Yang, W.; Liu, Y.; Xu, Q.Q.; Xian, Y.F.; Lin, Z.X. Sulforaphene Ameliorates Neuroinflammation and Hyperphosphorylated Tau Protein via Regulating the PI3K/Akt/GSK-3beta Pathway in Experimental Models of Alzheimer's Disease. *Oxid. Med. Cell Longev.* **2020**, *2020*, 4754195.
169. Kumar, M.; Bansal, N. Implications of Phosphoinositide 3-Kinase-Akt (PI3K-Akt) Pathway in the Pathogenesis of Alzheimer's Disease. *Mol. Neurobiol.* **2022**, *59*, 354–385.
170. Salem, M.A.; Budzynska, B.; Kowalczyk, J.; El Sayed, N.S.; Mansour, S.M. Tadalafil and bergapten mitigate streptozotocin-induced sporadic Alzheimer's disease in mice via modulating neuroinflammation, PI3K/Akt, Wnt/beta-catenin, AMPK/mTOR signaling pathways. *Toxicol. Appl. Pharmacol.* **2021**, *429*, 115697.
171. Mitchell, S.; Vargas, J.; Hoffmann, A. Signaling via the NFkappaB system. *Wiley Interdiscip. Rev. Syst. Biol. Med.* **2016**, *8*, 227–241.
172. Barnabei, L.; Laplantine, E.; Mbongo, W.; Rieux-Laucat, F.; Weil, R. NF-kappaB: At the Borders of Autoimmunity and Inflammation. *Front. Immunol.* **2021**, *12*, 716469.
173. Mitchell, J.P.; Carmody, R.J. NF-kappaB and the Transcriptional Control of Inflammation. *Int. Rev. Cell Mol. Biol.* **2018**, *335*, 41–84.
174. Mulero, M.C.; Huxford, T.; Ghosh, G. NF-kappaB, IkappaB, and IKK: Integral Components of Immune System Signaling. *Adv. Exp. Med. Biol.* **2019**, *1172*, 207–226.
175. Su, S.Y.; Cheng, C.Y.; Tsai, T.H.; Hsiang, C.Y.; Ho, T.Y.; Hsieh, C.L. Paeonol attenuates H(2)O(2)-induced NF-kappaB-associated amyloid precursor protein expression. *Am. J. Chin. Med.* **2010**, *38*, 1171–1192.

176. Yu, H.; Lin, L.; Zhang, Z.; Zhang, H.; Hu, H. Targeting NF-kappaB pathway for the therapy of diseases: mechanism and clinical study. *Signal Transduct. Target. Ther.* **2020**, *5*, 209.
177. Fan, Y.; Mao, R.; Yang, J. NF-kappaB and STAT3 signaling pathways collaboratively link inflammation to cancer. *Protein Cell* **2013**, *4*, 176–185.
178. Wang, Y.; Huang, X.; Cang, H.; Gao, F.; Yamamoto, T.; Osaki, T.; Yi, J. The endogenous reactive oxygen species promote NF-kappaB activation by targeting on activation of NF-kappaB-inducing kinase in oral squamous carcinoma cells. *Free Radic. Res.* **2007**, *41*, 963–971.
179. Singh, S.S.; Rai, S.N.; Birla, H.; Zahra, W.; Rathore, A.S.; Singh, S.P. NF-kappaB-Mediated Neuroinflammation in Parkinson's Disease and Potential Therapeutic Effect of Polyphenols. *Neurotox. Res.* **2020**, *37*, 491–507.
180. Shabab, T.; Khanabdali, R.; Moghadamtousi, S.Z.; Kadir, H.A.; Mohan, G. Neuroinflammation pathways: a general review. *Int. J. Neurosci.* **2017**, *127*, 624–633.
181. Du, Y.; Chen, X.; Wei, X.; Bales, K.R.; Berg, D.T.; Paul, S.M.; Farlow, M.R.; Maloney, B.; Ge, Y.W.; Lahiri, D.K. NF-(kappa)B mediates amyloid beta peptide-stimulated activity of the human apolipoprotein E gene promoter in human astroglial cells. *Brain Res. Mol. Brain Res.* **2005**, *136*, 177–188.
182. Chatterjee, P.; Yadav, M.; Chauhan, N.; Huang, Y.; Luo, Y. Cancer Cell Metabolism Featuring Nrf2. *Curr. Drug Discov. Technol.* **2020**, *17*, 263–271.
183. Osama, A.; Zhang, J.; Yao, J.; Yao, X.; Fang, J. Nrf2: a dark horse in Alzheimer's disease treatment. *Ageing Res. Rev.* **2020**, *64*, 101206.
184. Kasai, S.; Shimizu, S.; Tatara, Y.; Mimura, J.; Itoh, K. Regulation of Nrf2 by Mitochondrial Reactive Oxygen Species in Physiology and Pathology. *Biomolecules* **2020**, *10*, 320.
185. Jeong, W.S.; Jun, M.; Kong, A.N. Nrf2: a potential molecular target for cancer chemoprevention by natural compounds. *Antioxid. Redox Signal* **2006**, *8*, 99–106.
186. Rojo de la Vega, M.; Chapman, E.; Zhang, D.D. NRF2 and the Hallmarks of Cancer. *Cancer Cell* **2018**, *34*, 21–43.
187. Sivinski, J.; Zhang, D.D.; Chapman, E. Targeting NRF2 to treat cancer. *Semin. Cancer Biol.* **2021**, *76*, 61–73.
188. He, F.; Antonucci, L.; Karin, M. NRF2 as a regulator of cell metabolism and inflammation in cancer. *Carcinogenesis* **2020**, *41*, 405–416.
189. Choi, B.H.; Kim, J.M.; Kwak, M.K. The multifaceted role of NRF2 in cancer progression and cancer stem cells maintenance. *Arch. Pharm. Res.* **2021**, *44*, 263–280.
190. George, M.; Tharakan, M.; Culbertson, J.; Reddy, A.P.; Reddy, P.H. Role of Nrf2 in aging, Alzheimer's and other neurodegenerative diseases. *Ageing Res. Rev.* **2022**, *82*, 101756.
191. Davies, D.A.; Adlimoghaddam, A.; Albensi, B.C. Role of Nrf2 in Synaptic Plasticity and Memory in Alzheimer's Disease. *Cells* **2021**, *10*.
192. Riordan, R.; Rong, W.; Yu, Z.; Ross, G.; Valerio, J.; Dimas-Munoz, J.; Heredia, V.; Magnusson, K.; Galvan, V.; Perez, V.I. Effect of Nrf2 loss on senescence and cognition of tau-based P301S mice. *Geroscience* **2023**, *45*, 1451–1469.
193. Simoni, E.; Serafini, M.M.; Caporaso, R.; Marchetti, C.; Racchi, M.; Minarini, A.; Bartolini, M.; Lanni, C.; Rosini, M. Targeting the Nrf2/Amyloid-Beta Liaison in Alzheimer's Disease: A Rational Approach. *ACS Chem. Neurosci.* **2017**, *8*, 1618–1627.
194. Branca, C.; Ferreira, E.; Nguyen, T.V.; Doyle, K.; Caccamo, A.; Oddo, S. Genetic reduction of Nrf2 exacerbates cognitive deficits in a mouse model of Alzheimer's disease. *Hum. Mol. Genet.* **2017**, *26*, 4823–4835.
195. Pajares, M.; Rojo, A.I.; Arias, E.; Diaz-Carretero, A.; Cuervo, A.M.; Cuadrado, A. Transcription factor NFE2L2/NRF2 modulates chaperone-mediated autophagy through the regulation of LAMP2A. *Autophagy* **2018**, *14*, 1310–1322.
196. Deng, L.J.; Qi, M.; Li, N.; Lei, Y.H.; Zhang, D.M.; Chen, J.X. Natural products and their derivatives: Promising modulators of tumor immunotherapy. *J. Leukoc. Biol.* **2020**, *108*, 493–508.
197. Lu, X.; Yang, F.; Chen, D.; Zhao, Q.; Chen, D.; Ping, H.; Xing, N. Quercetin reverses docetaxel resistance in prostate cancer via androgen receptor and PI3K/Akt signaling pathways. *Int. J. Biol. Sci.* **2020**, *16*, 1121–1134.
198. Jia, X.B.; Zhang, Q.; Xu, L.; Yao, W.J.; Wei, L. Lotus leaf flavonoids induce apoptosis of human lung cancer A549 cells through the ROS/p38 MAPK pathway. *Biol. Res.* **2021**, *54*, 7.
199. Reyes-Farias, M.; Carrasco-Pozo, C. The Anti-Cancer Effect of Quercetin: Molecular Implications in Cancer Metabolism. *Int. J. Mol. Sci.* **2019**, *20*, 3177.
200. Ghafouri-Fard, S.; Shabestari, F.A.; Vaezi, S.; Abak, A.; Shoorei, H.; Karimi, A.; Taheri, M.; Basiri, A. Emerging impact of quercetin in the treatment of prostate cancer. *Biomed. Pharmacother.* **2021**, *138*, 111548.
201. Biswas, P.; Dey, D.; Biswas, P.K.; Rahaman, T.I.; Saha, S.; Parvez, A.; Khan, D.A.; Lily, N.J.; Saha, K.; Sohel, M.; et al. A Comprehensive Analysis and Anti-Cancer Activities of Quercetin in ROS-Mediated Cancer and Cancer Stem Cells. *Int. J. Mol. Sci.* **2022**, *23*, 11746.
202. Hasan, A.A.S.; Kalinina, E.V.; Tatarskiy, V.V.; Volodina, Y.L.; Petrova, .S.; Novichkova, M.D.; Zhdanov, D.D.; Shtil, A.A. Suppression of the Antioxidant System and PI3K/Akt/mTOR Signaling Pathway in Cisplatin-Resistant Cancer Cells by Quercetin. *Bull. Exp. Biol. Med.* **2022**, *173*, 760–764.
203. Abbasi, A.; Mostafavi-Pour, Z.; Amiri, A.; Keshavarzi, F.; Nejabat, N.; Ramezani, F.; Sardarian, A.; Zal, F. Chemoprevention of Prostate Cancer Cells by Vitamin C plus Quercetin: role of Nrf2 in Inducing Oxidative Stress. *Nutr. Cancer* **2021**, *73*, 2003–2013.
204. Mostafavi-Pour, Z.; Ramezani, F.; Keshavarzi, F.; Samadi, N. The role of quercetin and vitamin C in Nrf2-dependent oxidative stress production in breast cancer cells. *Oncol. Lett.* **2017**, *13*, 1965–1973.

205. Li, Y.; Yao, J.; Han, C.; Yang, J.; Chaudhry, M.T.; Wang, S.; Liu, H.; Yin, Y. Quercetin, Inflammation and Immunity. *Nutrients* **2016**, *8*, 167.
206. Lin, R.; Piao, M.; Song, Y.; Liu, C. Quercetin Suppresses AOM/DSS-Induced Colon Carcinogenesis through Its Anti-Inflammation Effects in Mice. *J. Immunol. Res.* **2020**, *2020*, 9242601.
207. Xu, D.; Hu, M.J.; Wang, Y.Q.; Cui, Y.L. Antioxidant Activities of Quercetin and Its Complexes for Medicinal Application. *Molecules* **2019**, *24*, 1123.
208. Rishitha, N.; Muthuraman, A. Therapeutic evaluation of solid lipid nanoparticle of quercetin in pentylenetetrazole induced cognitive impairment of zebrafish. *Life Sci.* **2018**, *199*, 80–87.
209. Patil, C.S.; Singh, V.P.; Satyanarayan, P.S.; Jain, N.K.; Singh, A.; Kulkarni, S.K. Protective effect of flavonoids against aging- and lipopolysaccharide-induced cognitive impairment in mice. *Pharmacology* **2003**, *69*, 59–67.
210. Li, Y.; Tian, Q.; Li, Z.; Dang, M.; Lin, Y.; Hou, X. Activation of Nrf2 signaling by sitagliptin and quercetin combination against beta-amyloid induced Alzheimer's disease in rats. *Drug Dev. Res.* **2019**, *80*, 837–845.
211. Wang, L.; Sun, J.; Miao, Z.; Jiang, X.; Zheng, Y.; Yang, G. Quercitrin improved cognitive impairment through inhibiting inflammation induced by microglia in Alzheimer's disease mice. *Neuroreport* **2022**, *33*, 327–335.
212. Chen, T.; Zhang, X.; Zhu, G.; Liu, H.; Chen, J.; Wang, Y.; He, X. Quercetin inhibits TNF-alpha induced HUVECs apoptosis and inflammation via downregulating NF-kB and AP-1 signaling pathway in vitro. *Medicine* **2020**, *99*, e22241.
213. Paco, A.; Bras, T.; Santos, J.O.; Sampaio, P.; Gomes, A.C.; Duarte, M.F. Anti-Inflammatory and Immunoregulatory Action of Sesquiterpene Lactones. *Molecules* **2022**, *27*, 1142.
214. Wang, F.; Zhong, H.; Fang, S.; Zheng, Y.; Li, C.; Peng, G.; Shen, X. Potential Anti-inflammatory Sesquiterpene Lactones from *Eupatorium lindleyanum*. *Planta Med.* **2018**, *84*, 123–128.
215. Neganova, M.E.; Klochkov, S.G.; Pukhov, S.A.; Afanasieva, S.V.; Aleksandrova, Y.R.; Yandulova, E.Y.; Avila-Rodriguez, M.F.; Mikhaleva, L.M.; Nikolenko, V.N.; Somasundaram, S.G.; et al. Synthesis and Cytotoxic Activity of Azine Derivatives of 6-Hydroxyxanthanodiene. *Curr. Cancer Drug Targets* **2020**, *20*, 666–674.
216. Neganova, M.; Semakov, A.; Aleksandrova, Y.; Yandulova, E.; Pukhov, S.; Anikina, L.; Klochkov, S. N-Alkylation of Anthracycline Antibiotics by Natural Sesquiterpene Lactones as a Way to Obtain Antitumor Agents with Reduced Side Effects. *Biomedicines* **2021**, *9*, 547.
217. Neganova, M.E.; Smirnova, E.V.; Sharova, E.V.; Artyushin, O.I.; Aleksandrova, Y.R.; Yandulova, E.Y.; Nikolaeva, N.S.; Brel, V.K. Design of Conjugates Based on Sesquiterpene Lactones with Polyalkoxybenzenes by “Click” Chemistry to Create Potential Anticancer Agents. *Molecules* **2022**, *27*, 8411.
218. Neganova, M.; Liu, J.; Aleksandrova, Y.; Vasilieva, N.; Semakov, A.; Yandulova, E.; Sukocheva, O.; Balakin, K.; Klochkov, S.; Fan, R. Development of Neuroprotective Agents for the Treatment of Alzheimer's Disease Using Conjugates of Serotonin with Sesquiterpene Lactones. *Curr. Med. Chem.* **2022**, *30*, 529–551.
219. Talman, A.M.; Clain, J.; Duval, R.; Menard, R.; Arley, F. Artemisinin Bioactivity and Resistance in Malaria Parasites. *Trends Parasitol.* **2019**, *35*, 953–963.
220. Ma, N.; Zhang, Z.; Liao, F.; Jiang, T.; Tu, Y. The birth of artemisinin. *Pharmacol. Ther.* **2020**, *216*, 107658.
221. Chen, G.Q.; Benthani, F.A.; Wu, J.; Liang, D.; Bian, Z.X.; Jiang, X. Artemisinin compounds sensitize cancer cells to ferroptosis by regulating iron homeostasis. *Cell Death Differ.* **2020**, *27*, 242–254.
222. Huang, Z.; Gan, S.; Zhuang, X.; Chen, Y.; Lu, L.; Wang, Y.; Qi, X.; Feng, Q.; Huang, Q.; Du, B.; et al. Artesunate Inhibits the Cell Growth in Colorectal Cancer by Promoting ROS-Dependent Cell Senescence and Autophagy. *Cells* **2022**, *11*, 2472.
223. Greenshields, A.L.; Shepherd, T.G.; Hoskin, D.W. Contribution of reactive oxygen species to ovarian cancer cell growth arrest and killing by the anti-malarial drug artesunate. *Mol. Carcinog.* **2017**, *56*, 75–93.
224. Zhang, Q.; Yi, H.; Yao, H.; Lu, L.; He, G.; Wu, M.; Zheng, C.; Li, Y.; Chen, S.; Li, L.; et al. Artemisinin Derivatives Inhibit Non-small Cell Lung Cancer Cells Through Induction of ROS-dependent Apoptosis/Ferroptosis. *J. Cancer* **2021**, *12*, 4075–4085.
225. Roh, J.L.; Kim, E.H.; Jang, H.; Shin, D. Nrf2 inhibition reverses the resistance of cisplatin-resistant head and neck cancer cells to artesunate-induced ferroptosis. *Redox Biol.* **2017**, *11*, 254–262.
226. Nunes, J.J.; Pandey, S.K.; Yadav, A.; Goel, S.; Ateeq, B. Targeting NF-kappa B Signaling by Artesunate Restores Sensitivity of Castrate-Resistant Prostate Cancer Cells to Antiandrogens. *Neoplasia* **2017**, *19*, 333–345.
227. Yan, J.; Ma, H.; Lai, X.; Wu, J.; Liu, A.; Huang, J.; Sun, W.; Shen, M.; Zhang, Y. Artemisinin attenuated oxidative stress and apoptosis by inhibiting autophagy in MPP(+)-treated SH-SY5Y cells. *J. Biol. Res.* **2021**, *28*, 6.
228. Lv, J.; Zhu, J.; Wang, P.; Liu, T.; Yuan, J.; Yin, H.; Lan, Y.; Sun, Q.; Zhang, Z.; Ding, G.; et al. Artemisinin exerts a protective effect in the MPTP mouse model of Parkinson's disease by inhibiting microglial activation via the TLR4/Myd88/NF-KB pathway. *CNS Neurosci. Ther.* **2023**, *29*, 1012–1023.
229. Lin, S.P.; Li, W.; Winters, A.; Liu, R.; Yang, S.H. Artemisinin Prevents Glutamate-Induced Neuronal Cell Death Via Akt Pathway Activation. *Front. Cell Neurosci.* **2018**, *12*, 108.
230. Okorji, U.P.; Velagapudi, R.; El-Bakoush, A.; Fiebich, B.L.; Olajide, O.A. Antimalarial Drug Artemether Inhibits Neuroinflammation in BV2 Microglia Through Nrf2-Dependent Mechanisms. *Mol. Neurobiol.* **2016**, *53*, 6426–6443.
231. Zhao, X.; Li, S.; Gaur, U.; Zheng, W. Artemisinin Improved Neuronal Functions in Alzheimer's Disease Animal Model 3xtg Mice and Neuronal Cells via Stimulating the ERK/CREB Signaling Pathway. *Aging Dis.* **2020**, *11*, 801–819.
232. Rasul, A.; Bao, R.; Malhi, M.; Zhao, B.; Tsuji, I.; Li, J.; Li, X. Induction of apoptosis by costunolide in bladder cancer cells is mediated through ROS generation and mitochondrial dysfunction. *Molecules* **2013**, *18*, 1418–1433.

233. Hua, P.; Sun, M.; Zhang, G.; Zhang, Y.; Song, G.; Liu, Z.; Li, X.; Zhang, X.; Li, B. Costunolide Induces Apoptosis through Generation of ROS and Activation of P53 in Human Esophageal Cancer Eca-109 Cells. *J. Biochem. Mol. Toxicol.* **2016**, *30*, 462–469.
234. Choi, Y.J.; Choi, Y.K.; Ko, S.G.; Cheon, C.; Kim, T.Y. Investigation of Molecular Mechanisms Involved in Sensitivity to the Anti-Cancer Activity of Costunolide in Breast Cancer Cells. *Int. J. Mol. Sci.* **2023**, *24*, 4009.
235. Jeyamohan, S.; Moorthy, R.K.; Kannan, M.K.; Arockiam, A.J. Parthenolide induces apoptosis and autophagy through the suppression of PI3K/Akt signaling pathway in cervical cancer. *Biotechnol. Lett.* **2016**, *38*, 1251–1260.
236. D’Anneo, A.; Carlisi, D.; Lauricella, M.; Puleio, R.; Martinez, R.; Di Bella, S.; Di Marco, P.; Emanuele, S.; Di Fiore, R.; Guercio, A.; et al. Parthenolide generates reactive oxygen species and autophagy in MDA-MB231 cells. A soluble parthenolide analogue inhibits tumour growth and metastasis in a xenograft model of breast cancer. *Cell Death Dis.* **2013**, *4*, e891.
237. Jorge, J.; Neves, J.; Alves, R.; Geraldes, C.; Goncalves, A.C.; Sarmiento-Ribeiro, A.B. Parthenolide Induces ROS-Mediated Apoptosis in Lymphoid Malignancies. *Int. J. Mol. Sci.* **2023**, *24*, 9167.
238. Cheong, C.U.; Yeh, C.S.; Hsieh, Y.W.; Lee, Y.R.; Lin, M.Y.; Chen, C.Y.; Lee, C.H. Protective Effects of Costunolide against Hydrogen Peroxide-Induced Injury in PC12 Cells. *Molecules* **2016**, *21*, 898.
239. Fan, M.; Wang, C.; Zhao, X.; Jiang, Y.; Wang, C. Parthenolide alleviates microglia-mediated neuroinflammation via MAPK/TRIM31/NLRP3 signaling to ameliorate cognitive disorder. *Int. Immunopharmacol.* **2023**, *120*, 110287.
240. Arslan, M.E.; Turkez, H.; Sevim, Y.; Selvitopi, H.; Kadi, A.; Oner, S.; Mardinoglu, A. Costunolide and Parthenolide Ameliorate MPP+ Induced Apoptosis in the Cellular Parkinson’s Disease Model. *Cells* **2023**, *12*, 992.
241. Reyes, A.A.; Marcum, R.D.; He, Y. Structure and Function of Chromatin Remodelers. *J. Mol. Biol.* **2021**, *433*, 166929.
242. Hauer, M.H.; Gasser, S.M. Chromatin and nucleosome dynamics in DNA damage and repair. *Genes. Dev.* **2017**, *31*, 2204–2221.
243. Zhang, Y.; Sun, Z.; Jia, J.; Du, T.; Zhang, N.; Tang, Y.; Fang, Y.; Fang, D. Overview of Histone Modification. *Adv. Exp. Med. Biol.* **2021**, *1283*, 1–16.
244. Strahl, B.D.; Allis, C.D. The language of covalent histone modifications. *Nature* **2000**, *403*, 41–45.
245. Neganova, M.E.; Klochkov, S.G.; Aleksandrova, Y.R.; Aliev, G. Histone modifications in epigenetic regulation of cancer: Perspectives and achieved progress. *Semin. Cancer Biol.* **2022**, *83*, 452–471.
246. Shvedunova, M.; Akhtar, A. Modulation of cellular processes by histone and non-histone protein acetylation. *Nat. Rev. Mol. Cell Biol.* **2022**, *23*, 329–349.
247. Shen, Y.; Wei, W.; Zhou, D.X. Histone Acetylation Enzymes Coordinate Metabolism and Gene Expression. *Trends Plant Sci.* **2015**, *20*, 614–621.
248. Peserico, A.; Simone, C. Physical and functional HAT/HDAC interplay regulates protein acetylation balance. *J. Biomed. Biotechnol.* **2011**, *2011*, 371832.
249. Chen, Y.; Zhou, Y.; Yin, H. Recent advances in biosensor for histone acetyltransferase detection. *Biosens. Bioelectron.* **2021**, *175*, 112880.
250. Khangura, R.K.; Bali, A.; Jaggi, A.S.; Singh, N. Histone acetylation and histone deacetylation in neuropathic pain: An unresolved puzzle? *Eur. J. Pharmacol.* **2017**, *795*, 36–42.
251. Seto, E.; Yoshida, M. Erasers of histone acetylation: the histone deacetylase enzymes. *Cold Spring Harb. Perspect. Biol.* **2014**, *6*, a018713.
252. Caron, C.; Boyault, C.; Khochbin, S. Regulatory cross-talk between lysine acetylation and ubiquitination: role in the control of protein stability. *Bioessays* **2005**, *27*, 408–415.
253. De Souza, C.; Chatterji, B.P. HDAC Inhibitors as Novel Anti-Cancer Therapeutics. *Recent. Pat. Anticancer. Drug Discov.* **2015**, *10*, 145–162.
254. Sweet, M.J.; Shakespear, M.R.; Kamal, N.A.; Fairlie, D.P. HDAC inhibitors: modulating leukocyte differentiation, survival, proliferation and inflammation. *Immunol. Cell Biol.* **2012**, *90*, 14–22.
255. Glauben, R.; Sonnenberg, E.; Zeitz, M.; Siegmund, B. HDAC inhibitors in models of inflammation-related tumorigenesis. *Cancer Lett.* **2009**, *280*, 154–159.
256. Leus, N.G.; Zwinderman, M.R.; Dekker, F.J. Histone deacetylase 3 (HDAC 3) as emerging drug target in NF-kappaB-mediated inflammation. *Curr. Opin. Chem. Biol.* **2016**, *33*, 160–168.
257. Robert, C.; Rassool, F.V. HDAC inhibitors: roles of DNA damage and repair. *Adv. Cancer Res.* **2012**, *116*, 87–129.
258. Zhao, C.; Dong, H.; Xu, Q.; Zhang, Y. Histone deacetylase (HDAC) inhibitors in cancer: a patent review (2017-present). *Expert. Opin. Ther. Pat.* **2020**, *30*, 263–274.
259. Ramaiah, M.J.; Tangutur, A.D.; Manyam, R.R. Epigenetic modulation and understanding of HDAC inhibitors in cancer therapy. *Life Sci.* **2021**, *277*, 119504.
260. Li, Y.; Seto, E. HDACs and HDAC Inhibitors in Cancer Development and Therapy. *Cold Spring Harb. Perspect. Med.* **2016**, *6*, a026831.
261. Cao, L.L.; Song, X.; Pei, L.; Liu, L.; Wang, H.; Jia, M. Histone deacetylase HDAC1 expression correlates with the progression and prognosis of lung cancer: A meta-analysis. *Medicine* **2017**, *96*, e7663.
262. Park, J.H.; Hong, Y.S.; Choi, P.J.; Kim, N.Y.; Lee, K.E.; Roh, M.S. The Overexpression of Histone Deacetylase 1 and Its Relationship with p16INK4a Gene Hypermethylation in Pulmonary Squamous Cell Carcinoma and Adenocarcinoma. *Korean J. Pathol.* **2009**, *43*, 107–112.

263. Benard, A.; Goossens-Beumer, I.J.; van Hoesel, A.Q.; Horati, H.; de Graaf, W.; Putter, H.; Zeestraten, E.C.; Liefers, G.J.; van de Velde, C.J.; Kuppen, P.J. Nuclear expression of histone deacetylases and their histone modifications predicts clinical outcome in colorectal cancer. *Histopathology* **2015**, *66*, 270–282.
264. Higashijima, J.; Kurita, N.; Miyatani, T.; Yoshikawa, K.; Morimoto, S.; Nishioka, M.; Iwata, T.; Shimada, M. Expression of histone deacetylase 1 and metastasis-associated protein 1 as prognostic factors in colon cancer. *Oncol. Rep.* **2011**, *26*, 343–348.
265. Weichert, W.; Roske, A.; Gekeler, V.; Beckers, T.; Ebert, M.P.; Pross, M.; Dietel, M.; Denkert, C.; Rocken, C. Association of patterns of class I histone deacetylase expression with patient prognosis in gastric cancer: a retrospective analysis. *Lancet Oncol.* **2008**, *9*, 139–148.
266. Mutze, K.; Langer, R.; Becker, K.; Ott, K.; Novotny, A.; Luber, B.; Hapfelmeier, A.; Gottlicher, M.; Hofler, H.; Keller, G. Histone deacetylase (HDAC) 1 and 2 expression and chemotherapy in gastric cancer. *Ann. Surg. Oncol.* **2010**, *17*, 3336–3343.
267. Wisniewski, F.; Calcagno, D.Q.; Leal, M.F.; Chen, E.S.; Gigek, C.O.; Santos, L.C.; Pontes, T.B.; Rasmussen, L.T.; Payao, S.L.; Assumpcao, P.P.; et al. Differential expression of histone deacetylase and acetyltransferase genes in gastric cancer and their modulation by trichostatin A. *Tumour Biol.* **2014**, *35*, 6373–6381.
268. Morine, Y.; Shimada, M.; Iwahashi, S.; Utsunomiya, T.; Imura, S.; Ikemoto, T.; Mori, H.; Hanaoka, J.; Miyake, H. Role of histone deacetylase expression in intrahepatic cholangiocarcinoma. *Surgery* **2012**, *151*, 412–419.
269. Rikimaru, T.; Taketomi, A.; Yamashita, Y.; Shirabe, K.; Hamatsu, T.; Shimada, M.; Maehara, Y. Clinical significance of histone deacetylase 1 expression in patients with hepatocellular carcinoma. *Oncology* **2007**, *72*, 69–74.
270. Weichert, W.; Roske, A.; Niesporek, S.; Noske, A.; Buckendahl, A.C.; Dietel, M.; Gekeler, V.; Boehm, M.; Beckers, T.; Denkert, C. Class I histone deacetylase expression has independent prognostic impact in human colorectal cancer: specific role of class I histone deacetylases in vitro and in vivo. *Clin. Cancer Res.* **2008**, *14*, 1669–1677.
271. Sudo, T.; Mimori, K.; Nishida, N.; Kogo, R.; Iwaya, T.; Tanaka, F.; Shibata, K.; Fujita, H.; Shirouzu, K.; Mori, M. Histone deacetylase 1 expression in gastric cancer. *Oncol. Rep.* **2011**, *26*, 777–782.
272. Jiang, Z.; Sun, X.; Zhang, Q.; Ji, X.; Yu, Q.; Huang, T.; Chen, D.; Chen, H.; Mei, X.; Wang, L.; et al. Identification of candidate biomarkers that involved in the epigenetic transcriptional regulation for detection gastric cancer by iTRAQ based quantitative proteomic analysis. *Clin. Chim. Acta* **2017**, *471*, 29–37.
273. Yu, Z.; Zeng, J.; Liu, H.; Wang, T.; Yu, Z.; Chen, J. Role of HDAC1 in the progression of gastric cancer and the correlation with lncRNAs. *Oncol. Lett.* **2019**, *17*, 3296–3304.
274. Peng, W.X.; Koirala, P.; Mo, Y.Y. LncRNA-mediated regulation of cell signaling in cancer. *Oncogene* **2017**, *36*, 5661–5667.
275. Bhan, A.; Soleimani, M.; Mandal, S.S. Long Noncoding RNA and Cancer: A New Paradigm. *Cancer Res.* **2017**, *77*, 3965–3981.
276. McCabe, E.M.; Rasmussen, T.P. lncRNA involvement in cancer stem cell function and epithelial-mesenchymal transitions. *Semin. Cancer Biol.* **2021**, *75*, 38–48.
277. Yan, H.; Bu, P. Non-coding RNA in cancer. *Essays Biochem.* **2021**, *65*, 625–639.
278. Deng, R.; Zhang, P.; Liu, W.; Zeng, X.; Ma, X.; Shi, L.; Wang, T.; Yin, Y.; Chang, W.; Zhang, P.; et al. HDAC is indispensable for IFN-gamma-induced B7-H1 expression in gastric cancer. *Clin. Epigenetics* **2018**, *10*, 153.
279. Jiang, Z.; Yang, H.; Zhang, X.; Wang, Z.; Rong, R.; Wang, X. Histone deacetylase-1 as a prognostic factor and mediator of gastric cancer progression by enhancing glycolysis. *Hum. Pathol.* **2019**, *85*, 194–201.
280. Guo, Q.; Cheng, K.; Wang, X.; Li, X.; Yu, Y.; Hua, Y.; Yang, Z. Expression of HDAC1 and RBBP4 correlate with clinicopathologic characteristics and prognosis in breast cancer. *Int. J. Clin. Exp. Pathol.* **2020**, *13*, 563–572.
281. Tang, Z.; Ding, S.; Huang, H.; Luo, P.; Qing, B.; Zhang, S.; Tang, R. HDAC1 triggers the proliferation and migration of breast cancer cells via upregulation of interleukin-8. *Biol. Chem.* **2017**, *398*, 1347–1356.
282. Alfaro, C.; Sanmamed, M.F.; Rodriguez-Ruiz, M.E.; Teijeira, A.; Onate, C.; Gonzalez, A.; Ponz, M.; Schalper, K.A.; Perez-Gracia, J.L.; Melero, I. Interleukin-8 in cancer pathogenesis, treatment and follow-up. *Cancer Treat. Rev.* **2017**, *60*, 24–31.
283. Sukocheva, O.A.; Lukina, E.; Friedemann, M.; Menschikowski, M.; Hagelgans, A.; Aliev, G. The crucial role of epigenetic regulation in breast cancer anti-estrogen resistance: Current findings and future perspectives. *Semin. Cancer Biol.* **2022**, *82*, 35–59.
284. Zhang, H.; Xu, H.; Ashby, C.R., Jr.; Assaraf, Y.G.; Chen, Z.S.; Liu, H.M. Chemical molecular-based approach to overcome multidrug resistance in cancer by targeting P-glycoprotein (P-gp). *Med. Res. Rev.* **2021**, *41*, 525–555.
285. Xu, Y.; Jiang, Z.; Yin, P.; Li, Q.; Liu, J. Role for Class I histone deacetylases in multidrug resistance. *Exp. Cell Res.* **2012**, *318*, 177–186.
286. Duan, H.; Zhou, K.; Zhang, Y.; Yue, P.; Wang, T.; Li, Y.; Qiu, D.; Hua, Y.; Wang, C. HDAC1 was involved in placental breast cancer resistance protein regulation in vitro: A preliminary study. *J. Cell Mol. Med.* **2019**, *23*, 5818–5821.
287. Wang, C.; Ma, D.; Hua, Y.; Duan, H. Modulation of Placental Breast Cancer Resistance Protein by HDAC1 in Mice: Implications for Optimization of Pharmacotherapy During Pregnancy. *Reprod. Sci.* **2021**, *28*, 3540–3546.
288. Burdelski, C.; Ruge, O.M.; Melling, N.; Koop, C.; Simon, R.; Steurer, S.; Sauter, G.; Kluth, M.; Hube-Magg, C.; Minner, S.; et al. HDAC1 overexpression independently predicts biochemical recurrence and is associated with rapid tumor cell proliferation and genomic instability in prostate cancer. *Exp. Mol. Pathol.* **2015**, *98*, 419–426.
289. Kim, N.H.; Kim, S.N.; Kim, Y.K. Involvement of HDAC1 in E-cadherin expression in prostate cancer cells; its implication for cell motility and invasion. *Biochem. Biophys. Res. Commun.* **2011**, *404*, 915–921.
290. Halkidou, K.; Gaughan, L.; Cook, S.; Leung, H.Y.; Neal, D.E.; Robson, C.N. Upregulation and nuclear recruitment of HDAC1 in hormone refractory prostate cancer. *Prostate* **2004**, *59*, 177–189.

291. Nakagawa, M.; Oda, Y.; Eguchi, T.; Aishima, S.; Yao, T.; Hosoi, F.; Basaki, Y.; Ono, M.; Kuwano, M.; Tanaka, M.; et al. Expression profile of class I histone deacetylases in human cancer tissues. *Oncol. Rep.* **2007**, *18*, 769–774.
292. Kitamura, H.; Torigoe, T.; Asanuma, H.; Hisasue, S.I.; Suzuki, K.; Tsukamoto, T.; Satoh, M.; Sato, N. Cytosolic overexpression of p62 sequestosome 1 in neoplastic prostate tissue. *Histopathology* **2006**, *48*, 157–161.
293. Shankar, E.; Pandey, M.; Verma, S.; Abbas, A.; Candamo, M.; Kanwal, R.; Shukla, S.; MacLennan, G.T.; Gupta, S. Role of class I histone deacetylases in the regulation of maspin expression in prostate cancer. *Mol. Carcinog.* **2020**, *59*, 955–966.
294. Bodenstine, T.M.; Seftor, R.E.; Khalkhali-Ellis, Z.; Seftor, E.A.; Pemberton, P.A.; Hendrix, M.J. Maspin: molecular mechanisms and therapeutic implications. *Cancer Metastasis Rev.* **2012**, *31*, 529–551.
295. Dzinic, S.H.; Bernardo, M.M.; Oliveira, D.S.; Wahba, M.; Sakr, W.; Sheng, S. Tumor suppressor maspin as a modulator of host immune response to cancer. *Bosn. J. Basic. Med. Sci.* **2015**, *15*, 1–6.
296. Jin, K.L.; Pak, J.H.; Park, J.Y.; Choi, W.H.; Lee, J.Y.; Kim, J.H.; Nam, J.H. Expression profile of histone deacetylases 1, 2 and 3 in ovarian cancer tissues. *J. Gynecol. Oncol.* **2008**, *19*, 185–190.
297. Liu, D.; Zhou, P.; Zhang, L.; Gong, W.; Huang, G.; Zheng, Y.; He, F. HDAC1/DNMT3A-containing complex is associated with suppression of Oct4 in cervical cancer cells. *Biochemistry (Moscow)* **2012**, *77*, 934–940.
298. Yano, M.; Yasuda, M.; Sakaki, M.; Nagata, K.; Fujino, T.; Arai, E.; Hasebe, T.; Miyazawa, M.; Miyazawa, M.; Ogane, N.; et al. Association of histone deacetylase expression with histology and prognosis of ovarian cancer. *Oncol. Lett.* **2018**, *15*, 3524–3531.
299. Cacan, E.; Ali, M.W.; Boyd, N.H.; Hooks, S.B.; Greer, S.F. Inhibition of HDAC1 and DNMT1 modulate RGS10 expression and decrease ovarian cancer chemoresistance. *PLoS ONE* **2014**, *9*, e87455.
300. Hooks, S.B.; Murph, M.M. Cellular deficiency in the RGS10 protein facilitates chemoresistant ovarian cancer. *Future Med. Chem.* **2015**, *7*, 1483–1489.
301. Cruceriu, D.; Baldasici, O.; Balacescu, O.; Berindan-Neagoe, I. The dual role of tumor necrosis factor-alpha (TNF-alpha) in breast cancer: molecular insights and therapeutic approaches. *Cell Oncol.* **2020**, *43*, 1–18.
302. Cacan, E. Histone Deacetylase-1-mediated Suppression of FAS in Chemoresistant Ovarian Cancer Cells. *Anticancer. Res.* **2016**, *36*, 2819–2826.
303. Liu, X.; Yu, Y.; Zhang, J.; Lu, C.; Wang, L.; Liu, P.; Song, H. HDAC1 Silencing in Ovarian Cancer Enhances the Chemotherapy Response. *Cell Physiol. Biochem.* **2018**, *48*, 1505–1518.
304. Zhu, Y.; Piao, C.; Zhang, Z.; Jiang, Y.; Kong, C. The potential role of c-MYC and polyamine metabolism in multiple drug resistance in bladder cancer investigated by metabolomics. *Genomics* **2022**, *114*, 125–137.
305. Krysan, K.; Kusko, R.; Grogan, T.; O'Hearn, J.; Reckamp, K.L.; Walser, T.C.; Garon, E.B.; Lenburg, M.E.; Sharma, S.; Spira, A.E.; et al. PGE2-driven expression of c-Myc and oncomiR-17-92 contributes to apoptosis resistance in NSCLC. *Mol. Cancer Res.* **2014**, *12*, 765–774.
306. Yang, Y.; Yuan, H.; Zhao, L.; Guo, S.; Hu, S.; Tian, M.; Nie, Y.; Yu, J.; Zhou, C.; Niu, J.; et al. Targeting the miR-34a/LRP-PRC/MDR1 axis collapse the chemoresistance in P53 inactive colorectal cancer. *Cell Death Differ.* **2022**, *29*, 2177–2189.
307. Li, W.J.; Wang, Y.; Liu, R.; Kasinski, A.L.; Shen, H.; Slack, F.J.; Tang, D.G. MicroRNA-34a: Potent Tumor Suppressor, Cancer Stem Cell Inhibitor, and Potential Anticancer Therapeutic. *Front. Cell Dev. Biol.* **2021**, *9*, 640587.
308. Wei, Y.; Liu, H.; Wang, C.; Zhang, W.; Wen, X.; Long, H.; Xu, Z.; Guo, H.; Liu, Y.; Wei, D.; et al. Clinicopathological and prognostic significance of octamer-binding transcription factor 4 in patients with gastric cancer: a systematic review and meta-analysis. *Biomark. Med.* **2019**, *13*, 219–234.
309. Gao, Z.Y.; Liu, X.B.; Yang, F.M.; Liu, L.; Zhao, J.Z.; Gao, B.; Li, S.B. Octamer binding transcription factor-4 expression is associated with cervical cancer malignancy and histological differentiation: a systematic review and meta-analysis. *Biosci. Rep.* **2019**, *39*, BSR20182328.
310. Zhao, X.; Lu, H.; Sun, Y.; Liu, L.; Wang, H. Prognostic value of octamer binding transcription factor 4 for patients with solid tumors: A meta-analysis. *Medicine* **2020**, *99*, e22804.
311. Upadhyay, V.A.; Shah, K.A.; Makwana, D.P.; Raval, A.P.; Shah, F.D.; Rawal, R.M. Putative stemness markers octamer-binding transcription factor 4, sex-determining region Y-box 2, and NANOG in non-small cell lung carcinoma: A clinicopathological association. *J. Cancer Res. Ther.* **2020**, *16*, 804–810.
312. Yokoi, E.; Mabuchi, S.; Shimura, K.; Komura, N.; Kozasa, K.; Kuroda, H.; Takahashi, R.; Sasano, T.; Kawano, M.; Matsumoto, Y.; et al. Lurbinectedin (PM01183), a selective inhibitor of active transcription, effectively eliminates both cancer cells and cancer stem cells in preclinical models of uterine cervical cancer. *Investig. New Drugs* **2019**, *37*, 818–827.
313. Cai, S.X.; Chen, W.S.; Zeng, W.; Cheng, X.F.; Lin, M.B.; Wang, J.S. Roles of HDAC2, eIF5, and eIF6 in Lung Cancer Tumorigenesis. *Curr. Med. Sci.* **2021**, *41*, 764–769.
314. Wang, F.W.; Guan, X.Y.; Xie, D. Roles of eukaryotic initiation factor 5A2 in human cancer. *Int. J. Biol. Sci.* **2013**, *9*, 1013–1020.
315. Zhang, H.; Zhang, K.; Ning, L.; Chen, D.; Hao, F.; Li, P. Clinical significance of eukaryotic translation initiation factor 5A2 in papillary thyroid cancer. *Bioengineered* **2020**, *11*, 1325–1333.
316. Gantenbein, N.; Bernhart, E.; Anders, I.; Golob-Schwarzl, N.; Krassnig, S.; Wodlej, C.; Brcic, L.; Lindenmann, J.; Fink-Neuboeck, N.; Gollowitsch, F.; et al. Influence of eukaryotic translation initiation factor 6 on non-small cell lung cancer development and progression. *Eur. J. Cancer* **2018**, *101*, 165–180.
317. Golob-Schwarzl, N.; Wodlej, C.; Kleinegger, F.; Gogg-Kamerer, M.; Birkl-Toeglhofer, A.M.; Petzold, J.; Aigelsreiter, A.; Thalhammer, M.; Park, Y.N.; Haybaeck, J. Eukaryotic translation initiation factor 6 overexpression plays a major role in the translational control of gallbladder cancer. *J. Cancer Res. Clin. Oncol.* **2019**, *145*, 2699–2711.

318. Li, L.; Mei, D.T.; Zeng, Y. HDAC2 promotes the migration and invasion of non-small cell lung cancer cells via upregulation of fibronectin. *Biomed. Pharmacother.* **2016**, *84*, 284–290.
319. Kumra, H.; Reinhardt, D.P. Fibronectin-targeted drug delivery in cancer. *Adv. Drug Deliv. Rev.* **2016**, *97*, 101–110.
320. Rick, J.W.; Chandra, A.; Dalle Ore, C.; Nguyen, A.T.; Yagnik, G.; Aghi, M.K. Fibronectin in malignancy: Cancer-specific alterations, protumoral effects, and therapeutic implications. *Semin. Oncol.* **2019**, *46*, 284–290.
321. Lin, T.C.; Yang, C.H.; Cheng, L.H.; Chang, W.T.; Lin, Y.R.; Cheng, H.C. Fibronectin in Cancer: Friend or Foe. *Cells* **2019**, *9*, 27.
322. Wang, B.; Shen, X.Y.; Pan, L.Y.; Li, Z.; Chen, C.J.; Yao, Y.S.; Tang, D.F.; Gao, W. The HDAC2-MTA3 interaction induces non-small cell lung cancer cell migration and invasion by targeting c-Myc and cyclin D1. In *Molecular Carcinogenesis*; Wiley: Hoboken, NJ, USA, 2023.
323. Conte, M.; Di Mauro, A.; Capasso, L.; Montella, L.; De Simone, M.; Nebbioso, A.; Altucci, L. Targeting HDAC2-Mediated Immune Regulation to Overcome Therapeutic Resistance in Mutant Colorectal Cancer. *Cancers* **2023**, *15*, 1960.
324. Qi, Z.P.; Yalikong, A.; Zhang, J.W.; Cai, S.L.; Li, B.; Di, S.; Lv, Z.T.; Xu, E.P.; Zhong, Y.S.; Zhou, P.H. HDAC2 promotes the EMT of colorectal cancer cells and via the modular scaffold function of ENSG00000274093.1. *J. Cell Mol. Med.* **2021**, *25*, 1190–1197.
325. Song, J.; Noh, J.H.; Lee, J.H.; Eun, J.W.; Ahn, Y.M.; Kim, S.Y.; Lee, S.H.; Park, W.S.; Yoo, N.J.; Lee, J.Y.; et al. Increased expression of histone deacetylase 2 is found in human gastric cancer. *APMIS* **2005**, *113*, 264–268.
326. Krishna, A.; Singh, V.; Singh, S.; Kumar, S.; Kumar, V.; Mehrotra, D.; Singh, U.S.; Mahdi, A.A. Upregulated histone deacetylase 2 gene correlates with the progression of oral squamous cell carcinoma. *Cancer Biomark.* **2020**, *29*, 543–552.
327. Li, S.; Wang, F.; Qu, Y.; Chen, X.; Gao, M.; Yang, J.; Zhang, D.; Zhang, N.; Li, W.; Liu, H. HDAC2 regulates cell proliferation, cell cycle progression and cell apoptosis in esophageal squamous cell carcinoma EC9706 cells. *Oncol. Lett.* **2017**, *13*, 403–409.
328. Du, X.; Zhao, H.; Zang, L.; Song, N.; Yang, T.; Dong, R.; Yin, J.; Wang, C.; Lu, J. Overexpression of histone deacetylase 2 predicts unfavorable prognosis in human gallbladder carcinoma. *Pathol. Oncol. Res.* **2013**, *19*, 397–403.
329. Shan, W.; Jiang, Y.; Yu, H.; Huang, Q.; Liu, L.; Guo, X.; Li, L.; Mi, Q.; Zhang, K.; Yang, Z. HDAC2 overexpression correlates with aggressive clinicopathological features and DNA-damage response pathway of breast cancer. *Am. J. Cancer Res.* **2017**, *7*, 1213–1226.
330. Ma, L.; Qi, L.; Li, S.; Yin, Q.; Liu, J.; Wang, J.; She, C.; Li, P.; Liu, Q.; Wang, X.; et al. Aberrant HDAC3 expression correlates with brain metastasis in breast cancer patients. *Thorac. Cancer* **2020**, *11*, 2493–2505.
331. Hu, G.; He, N.; Cai, C.; Cai, F.; Fan, P.; Zheng, Z.; Jin, X. HDAC3 modulates cancer immunity via increasing PD-L1 expression in pancreatic cancer. *Pancreatol.* **2019**, *19*, 383–389.
332. Edderkaoui, M.; Xu, S.; Chheda, C.; Morvaridi, S.; Hu, R.W.; Grippo, P.J.; Mascarinas, E.; Principe, D.R.; Knudsen, B.; Xue, J.; et al. HDAC3 mediates smoking-induced pancreatic cancer. *Oncotarget* **2016**, *7*, 7747–7760.
333. Eichner, L.J.; Curtis, S.D.; Brun, S.N.; McGuire, C.K.; Gushterova, I.; Baumgart, J.T.; Trefts, E.; Ross, D.S.; Rymoff, T.J.; Shaw, R.J. HDAC3 is critical in tumor development and therapeutic resistance in Kras-mutant non-small cell lung cancer. *Sci. Adv.* **2023**, *9*, eadd3243.
334. Minamiya, Y.; Ono, T.; Saito, H.; Takahashi, N.; Ito, M.; Motoyama, S.; Ogawa, J. Strong expression of HDAC3 correlates with a poor prognosis in patients with adenocarcinoma of the lung. *Tumour Biol.* **2010**, *31*, 533–539.
335. Miao, L.J.; Huang, F.X.; Sun, Z.T.; Zhang, R.X.; Huang, S.F.; Wang, J. Stat3 inhibits Beclin 1 expression through recruitment of HDAC3 in non-small cell lung cancer cells. *Tumour Biol.* **2014**, *35*, 7097–7103.
336. Chen, C.Q.; Chen, C.S.; Chen, J.J.; Zhou, L.P.; Xu, H.L.; Jin, W.W.; Wu, J.B.; Gao, S.M. Histone deacetylases inhibitor trichostatin A increases the expression of Dleu2/miR-15a/16-1 via HDAC3 in non-small cell lung cancer. *Mol. Cell Biochem.* **2013**, *383*, 137–148.
337. McLeod, A.B.; Stice, J.P.; Wardell, S.E.; Alley, H.M.; Chang, C.Y.; McDonnell, D.P. Validation of histone deacetylase 3 as a therapeutic target in castration-resistant prostate cancer. *Prostate* **2018**, *78*, 266–277.
338. Zheng, Y.; Wu, C.; Yang, J.; Zhao, Y.; Jia, H.; Xue, M.; Xu, D.; Yang, F.; Fu, D.; Wang, C.; et al. Insulin-like growth factor 1-induced enolase 2 deacetylation by HDAC3 promotes metastasis of pancreatic cancer. *Signal Transduct. Target. Ther.* **2020**, *5*, 53.
339. Wu, L.M.; Yang, Z.; Zhou, L.; Zhang, F.; Xie, H.Y.; Feng, X.W.; Wu, J.; Zheng, S.S. Identification of histone deacetylase 3 as a biomarker for tumor recurrence following liver transplantation in HBV-associated hepatocellular carcinoma. *PLoS One* **2010**, *5*, e14460.
340. Liu, C.; Liu, L.; Shan, J.; Shen, J.; Xu, Y.; Zhang, Q.; Yang, Z.; Wu, L.; Xia, F.; Bie, P.; et al. Histone deacetylase 3 participates in self-renewal of liver cancer stem cells through histone modification. *Cancer Lett.* **2013**, *339*, 60–69.
341. Xu, G.; Zhu, H.; Zhang, M.; Xu, J. Histone deacetylase 3 is associated with gastric cancer cell growth via the miR-454-mediated targeting of CHD5. *Int. J. Mol. Med.* **2018**, *41*, 155–163.
342. Nemat, M.; Ajami, N.; Estari, M.A.; Rezapour, S.; Gavani, R.R.; Hashemzadeh, S.; Kafil, H.S.; Sakhinia, E. Deregulated expression of HDAC3 in colorectal cancer and its clinical significance. *Adv. Clin. Exp. Med.* **2018**, *27*, 305–311.
343. Jiao, F.; Hu, H.; Han, T.; Zhuo, M.; Yuan, C.; Yang, H.; Wang, L.; Wang, L. Aberrant expression of nuclear HDAC3 and cytoplasmic CDH1 predict a poor prognosis for patients with pancreatic cancer. *Oncotarget* **2016**, *7*, 16505–16516.
344. Zhong, S.; Fan, Y.; Wu, B.; Wang, Y.; Jiang, S.; Ge, J.; Hua, C.; Zhao, G.; Chen, Y.; Xu, H. HDAC3 Expression Correlates with the Prognosis and Grade of Patients with Glioma: A Diversification Analysis Based on Transcriptome and Clinical Evidence. *World Neurosurg.* **2018**, *119*, e145–e158.

345. Beyer, M.; Romanski, A.; Mustafa, A.M.; Pons, M.; Buchler, I.; Vogel, A.; Pautz, A.; Sellmer, A.; Schneider, G.; Bug, G.; et al. HDAC3 Activity is Essential for Human Leukemic Cell Growth and the Expression of beta-catenin, MYC, and WT1. *Cancers* **2019**, *11*, 1436.
346. Godman, C.A.; Joshi, R.; Tierney, B.R.; Greenspan, E.; Rasmussen, T.P.; Wang, H.W.; Shin, D.G.; Rosenberg, D.W.; Giardina, C. HDAC3 impacts multiple oncogenic pathways in colon cancer cells with effects on Wnt and vitamin D signaling. *Cancer Biol. Ther.* **2008**, *7*, 1570–1580.
347. Zhan, T.; Rindtorff, N.; Boutros, M. Wnt signaling in cancer. *Oncogene* **2017**, *36*, 1461–1473.
348. Zhou, Y.; Xu, J.; Luo, H.; Meng, X.; Chen, M.; Zhu, D. Wnt signaling pathway in cancer immunotherapy. *Cancer Lett.* **2022**, *525*, 84–96.
349. Parsons, M.J.; Tammela, T.; Dow, L.E. WNT as a Driver and Dependency in Cancer. *Cancer Discov.* **2021**, *11*, 2413–2429.
350. Jiao, F.; Hu, H.; Yuan, C.; Jin, Z.; Guo, Z.; Wang, L.; Wang, L. Histone deacetylase 3 promotes pancreatic cancer cell proliferation, invasion and increases drug-resistance through histone modification of P27, P53 and Bax. *Int. J. Oncol.* **2014**, *45*, 1523–1530.
351. Ren, H.; Tang, L. HDAC3-mediated lncRNA-LOC101928316 contributes to cisplatin resistance in gastric cancer via activating the PI3K-Akt-mTOR pathway. *Neoplasma* **2021**, *68*, 1043–1051.
352. Kim, J.Y.; Cho, H.; Yoo, J.; Kim, G.W.; Jeon, Y.H.; Lee, S.W.; Kwon, S.H. Pathological Role of HDAC8: Cancer and Beyond. *Cells* **2022**, *11*, 3161.
353. An, P.; Chen, F.; Li, Z.; Ling, Y.; Peng, Y.; Zhang, H.; Li, J.; Chen, Z.; Wang, H. HDAC8 promotes the dissemination of breast cancer cells via AKT/GSK-3beta/Snail signals. *Oncogene* **2020**, *39*, 4956–4969.
354. Chen, J.; Cao, L.; Ma, J.; Yue, C.; Zhu, D.; An, R.; Wang, X.; Guo, Y.; Gu, B. HDAC8 Promotes Liver Metastasis of Colorectal Cancer via Inhibition of IRF1 and Upregulation of SUCNR1. *Oxid. Med. Cell Longev.* **2022**, *2022*, 2815187.
355. Wu, J.; Du, C.; Lv, Z.; Ding, C.; Cheng, J.; Xie, H.; Zhou, L.; Zheng, S. The up-regulation of histone deacetylase 8 promotes proliferation and inhibits apoptosis in hepatocellular carcinoma. *Dig. Dis. Sci.* **2013**, *58*, 3545–3553.
356. Song, S.; Wang, Y.; Xu, P.; Yang, R.; Ma, Z.; Liang, S.; Zhang, G. The inhibition of histone deacetylase 8 suppresses proliferation and inhibits apoptosis in gastric adenocarcinoma. *Int. J. Oncol.* **2015**, *47*, 1819–1828.
357. Moreno, D.A.; Scrideli, C.A.; Cortez, M.A.; de Paula Queiroz, R.; Valera, E.T.; da Silva Silveira, V.; Yunes, J.A.; Brandalise, S.R.; Tone, L.G. Differential expression of HDAC3, HDAC7 and HDAC9 is associated with prognosis and survival in childhood acute lymphoblastic leukaemia. *Br. J. Haematol.* **2010**, *150*, 665–673.
358. Ahn, M.Y.; Yoon, J.H. Histone deacetylase 8 as a novel therapeutic target in oral squamous cell carcinoma. *Oncol. Rep.* **2017**, *37*, 540–546.
359. Oehme, I.; Deubzer, H.E.; Wegener, D.; Pickert, D.; Linke, J.P.; Hero, B.; Kopp-Schneider, A.; Westermann, F.; Ulrich, S.M.; von Deimling, A.; et al. Histone deacetylase 8 in neuroblastoma tumorigenesis. *Clin. Cancer Res.* **2009**, *15*, 91–99.
360. Wilmott, J.S.; Colebatch, A.J.; Kakavand, H.; Shang, P.; Carlino, M.S.; Thompson, J.F.; Long, G.V.; Scolyer, R.A.; Hersey, P. Expression of the class 1 histone deacetylases HDAC8 and 3 are associated with improved survival of patients with metastatic melanoma. *Mod. Pathol.* **2015**, *28*, 884–894.
361. Mormino, A.; Cocozza, G.; Fontemaggi, G.; Valente, S.; Esposito, V.; Santoro, A.; Bernardini, G.; Santoni, A.; Fazi, F.; Mai, A.; et al. Histone-deacetylase 8 drives the immune response and the growth of glioma. *Glia* **2021**, *69*, 2682–2698.
362. Lopez, G.; Bill, K.L.; Bid, H.K.; Braggio, D.; Constantino, D.; Prudner, B.; Zewdu, A.; Batte, K.; Lev, D.; Pollock, R.E. HDAC8, A Potential Therapeutic Target for the Treatment of Malignant Peripheral Nerve Sheath Tumors (MPNST). *PLoS ONE* **2015**, *10*, e0133302.
363. Gao, S.M.; Chen, C.Q.; Wang, L.Y.; Hong, L.L.; Wu, J.B.; Dong, P.H.; Yu, F.J. Histone deacetylases inhibitor sodium butyrate inhibits JAK2/STAT signaling through upregulation of SOCS1 and SOCS3 mediated by HDAC8 inhibition in myeloproliferative neoplasms. *Exp. Hematol.* **2013**, *41*, 261–270 e4.
364. Chen, S.S.; Wu, W.Z.; Zhang, Y.P.; Huang, W.J. Gene polymorphisms of SOCS1 and SOCS2 and acute lymphoblastic leukemia. *Eur. Rev. Med. Pharmacol. Sci.* **2020**, *24*, 5564–5572.
365. Zhang, R.; Shen, M.; Wu, C.; Chen, Y.; Lu, J.; Li, J.; Zhao, L.; Meng, H.; Zhou, X.; Huang, G.; et al. HDAC8-dependent deacetylation of PKM2 directs nuclear localization and glycolysis to promote proliferation in hepatocellular carcinoma. *Cell Death Dis.* **2020**, *11*, 1036.
366. Nie, M.; Wang, Y.; Yu, Z.; Li, X.; Deng, Y.; Wang, Y.; Yang, D.; Li, Q.; Zeng, X.; Ju, J.; et al. AURKB promotes gastric cancer progression via activation of CCND1 expression. *Aging (Albany NY)* **2020**, *12*, 1304–1321.
367. Zhu, D.; Huang, J.; Liu, N.; Li, W.; Yan, L. PSMC2/CCND1 axis promotes development of ovarian cancer through regulating cell growth, apoptosis and migration. *Cell Death Dis.* **2021**, *12*, 730.
368. Valla, M.; Klaestad, E.; Ytterhus, B.; Bofin, A.M. CCND1 Amplification in Breast Cancer -associations With Proliferation, Histopathological Grade, Molecular Subtype and Prognosis. *J. Mammary Gland. Biol. Neoplasia* **2022**, *27*, 67–77.
369. Tang, X.; Li, G.; Su, F.; Cai, Y.; Shi, L.; Meng, Y.; Liu, Z.; Sun, J.; Wang, M.; Qian, M.; et al. HDAC8 cooperates with SMAD3/4 complex to suppress SIRT7 and promote cell survival and migration. *Nucleic Acids Res.* **2020**, *48*, 2912–2923.
370. Kim, J.Y.; Cho, H.; Yoo, J.; Kim, G.W.; Jeon, Y.H.; Lee, S.W.; Kwon, S.H. HDAC8 Deacetylates HIF-1alpha and Enhances Its Protein Stability to Promote Tumor Growth and Migration in Melanoma. *Cancers* **2023**, *15*, 1123.
371. Vanaja, G.R.; Ramulu, H.G.; Kalle, A.M. Overexpressed HDAC8 in cervical cancer cells shows functional redundancy of tubulin deacetylation with HDAC6. *Cell Commun. Signal* **2018**, *16*, 20.

372. Wu, S.; Luo, Z.; Yu, P.J.; Xie, H.; He, Y.W. Suberoylanilide hydroxamic acid (SAHA) promotes the epithelial mesenchymal transition of triple negative breast cancer cells via HDAC8/FOXA1 signals. *Biol. Chem.* **2016**, *397*, 75–83.
373. Kumar, V.; Kundu, S.; Singh, A.; Singh, S. Understanding the Role of Histone Deacetylase and their Inhibitors in Neurodegenerative Disorders: Current Targets and Future Perspective. *Curr. Neuropharmacol.* **2022**, *20*, 158–178.
374. Zhang, L.; Sheng, S.; Qin, C. The role of HDAC6 in Alzheimer's disease. *J. Alzheimers Dis.* **2013**, *33*, 283–295.
375. Simoes-Pires, C.; Zwick, V.; Nurisso, A.; Schenker, E.; Carrupt, P.A.; Cuendet, M. HDAC6 as a target for neurodegenerative diseases: what makes it different from the other HDACs? *Mol. Neurodegener.* **2013**, *8*, 7.
376. Mahady, L.; Nadeem, M.; Malek-Ahmadi, M.; Chen, K.; Perez, S.E.; Mufson, E.J. Frontal Cortex Epigenetic Dysregulation During the Progression of Alzheimer's Disease. *J. Alzheimers Dis.* **2018**, *62*, 115–131.
377. Bai, P.; Mondal, P.; Bagdasarian, F.A.; Rani, N.; Liu, Y.; Gomm, A.; Tocci, D.R.; Choi, S.H.; Wey, H.Y.; Tanzi, R.E.; et al. Development of a potential PET probe for HDAC6 imaging in Alzheimer's disease. *Acta Pharm. Sin. B* **2022**, *12*, 3891–3904.
378. Li, L.; Yang, X.J. Tubulin acetylation: responsible enzymes, biological functions and human diseases. *Cell Mol. Life Sci.* **2015**, *72*, 4237–4255.
379. Pulya, S.; Amin, S.A.; Adhikari, N.; Biswas, S.; Jha, T.; Ghosh, B. HDAC6 as privileged target in drug discovery: A perspective. *Pharmacol. Res.* **2021**, *163*, 105274.
380. Dent, E.W. Of microtubules and memory: implications for microtubule dynamics in dendrites and spines. *Mol. Biol. Cell* **2017**, *28*, 1–8.
381. Pena-Ortega, F.; Robles-Gomez, A.A.; Xolalpa-Cueva, L. Microtubules as Regulators of Neural Network Shape and Function: Focus on Excitability, Plasticity and Memory. *Cells* **2022**, *11*, 923.
382. Boiarska, Z.; Passarella, D. Microtubule-targeting agents and neurodegeneration. *Drug Discov. Today* **2021**, *26*, 604–615.
383. Lee, S.; Kwon, Y.; Kim, S.; Jo, M.; Jeon, Y.M.; Cheon, M.; Lee, S.; Kim, S.R.; Kim, K.; Kim, H.J. The Role of HDAC6 in TDP-43-Induced Neurotoxicity and UPS Impairment. *Front. Cell Dev. Biol.* **2020**, *8*, 581942.
384. Pandey, U.B.; Nie, Z.; Batlevi, Y.; McCray, B.A.; Ritson, G.P.; Nedelsky, N.B.; Schwartz, S.L.; DiProspero, N.A.; Knight, M.A.; Schuldiner, O.; et al. HDAC6 rescues neurodegeneration and provides an essential link between autophagy and the UPS. *Nature* **2007**, *447*, 859–863.
385. Hamano, T.; Enomoto, S.; Shirafuji, N.; Ikawa, M.; Yamamura, O.; Yen, S.H.; Nakamoto, Y. Autophagy and Tau Protein. *Int. J. Mol. Sci.* **2021**, *22*.
386. Xu, Y.; Propson, N.E.; Du, S.; Xiong, W.; Zheng, H. Autophagy deficiency modulates microglial lipid homeostasis and aggravates tau pathology and spreading. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2023418118.
387. Choi, H.; Kim, H.J.; Yang, J.; Chae, S.; Lee, W.; Chung, S.; Kim, J.; Choi, H.; Song, H.; Lee, C.K.; et al. Acetylation changes tau interactome to degrade tau in Alzheimer's disease animal and organoid models. *Aging Cell* **2020**, *19*, e13081.
388. Balmik, A.A.; Chidambaram, H.; Dangi, A.; Marelli, U.K.; Chinnathambi, S. HDAC6 ZnF UBP as the Modifier of Tau Structure and Function. *Biochemistry* **2020**, *59*, 4546–4562.
389. Qureshi, T.; Chinnathambi, S. Histone deacetylase-6 modulates Tau function in Alzheimer's disease. *Biochim. Biophys. Acta Mol. Cell Res.* **2022**, *1869*, 119275.
390. Tseng, J.H.; Xie, L.; Song, S.; Xie, Y.; Allen, L.; Ajit, D.; Hong, J.S.; Chen, X.; Meeker, R.B.; Cohen, T.J. The Deacetylase HDAC6 Mediates Endogenous Neuritic Tau Pathology. *Cell Rep.* **2017**, *20*, 2169–2183.
391. Cook, C.; Gendron, T.F.; Scheffel, K.; Carlomagno, Y.; Dunmore, J.; DeTure, M.; Petrucelli, L. Loss of HDAC6, a novel CHIP substrate, alleviates abnormal tau accumulation. *Hum. Mol. Genet.* **2012**, *21*, 2936–2945.
392. Sreenivasmurthy, S.G.; Iyaswamy, A.; Krishnamoorthi, S.; Reddi, R.N.; Kammala, A.K.; Vasudevan, K.; Senapati, S.; Zhu, Z.; Su, C.F.; Liu, J.; et al. Bromo-protopine, a novel protopine derivative, alleviates tau pathology by activating chaperone-mediated autophagy for Alzheimer's disease therapy. *Front. Mol. Biosci.* **2022**, *9*, 1030534.
393. Zhang, L.; Zhang, Z.; Li, C.; Zhu, T.; Gao, J.; Zhou, H.; Zheng, Y.; Chang, Q.; Wang, M.; Wu, J.; et al. S100A11 Promotes Liver Steatosis via FOXO1-Mediated Autophagy and Lipogenesis. *Cell Mol. Gastroenterol. Hepatol.* **2021**, *11*, 697–724.
394. Brijmohan, A.S.; Batchu, S.N.; Majumder, S.; Alghamdi, T.A.; Thieme, K.; McGaugh, S.; Liu, Y.; Advani, S.L.; Bowskill, B.B.; Kabir, M.G.; et al. HDAC6 Inhibition Promotes Transcription Factor EB Activation and Is Protective in Experimental Kidney Disease. *Front. Pharmacol.* **2018**, *9*, 34.
395. Chang, P.; Li, H.; Hu, H.; Li, Y.; Wang, T. The Role of HDAC6 in Autophagy and NLRP3 Inflammasome. *Front. Immunol.* **2021**, *12*, 763831.
396. Mazzetti, S.; De Leonardis, M.; Gagliardi, G.; Calogero, A.M.; Basellini, M.J.; Madaschi, L.; Costa, I.; Cacciatore, F.; Spinello, S.; Bramerio, M.; et al. Phospho-HDAC6 Gathers Into Protein Aggregates in Parkinson's Disease and Atypical Parkinsonisms. *Front. Neurosci.* **2020**, *14*, 624.
397. Li, E.; Choi, J.; Sim, H.R.; Kim, J.; Jun, J.H.; Kyung, J.; Ha, N.; Kim, S.; Ryu, K.H.; Chung, S.S.; et al. A novel HDAC6 inhibitor, CKD-504, is effective in treating preclinical models of huntington's disease. *BMB Rep.* **2023**, *56*, 178–183.
398. Guan, J.S.; Haggarty, S.J.; Giacometti, E.; Dannenberg, J.H.; Joseph, N.; Gao, J.; Nieland, T.J.; Zhou, Y.; Wang, X.; Mazitschek, R.; et al. HDAC2 negatively regulates memory formation and synaptic plasticity. *Nature* **2009**, *459*, 55–60.
399. Graff, J.; Rei, D.; Guan, J.S.; Wang, W.Y.; Seo, J.; Hennig, K.M.; Nieland, T.J.; Fass, D.M.; Kao, P.F.; Kahn, M.; et al. An epigenetic blockade of cognitive functions in the neurodegenerating brain. *Nature* **2012**, *483*, 222–226.
400. Jawerka, M.; Colak, D.; Dimou, L.; Spiller, C.; Lagger, S.; Montgomery, R.L.; Olson, E.N.; Wurst, W.; Gottlicher, M.; Gotz, M. The specific role of histone deacetylase 2 in adult neurogenesis. *Neuron Glia Biol.* **2010**, *6*, 93–107.

401. Zhu, X.; Wang, S.; Yu, L.; Jin, J.; Ye, X.; Liu, Y.; Xu, Y. HDAC3 negatively regulates spatial memory in a mouse model of Alzheimer's disease. *Aging Cell* **2017**, *16*, 1073–1082.
402. Bardai, F.H.; D'Mello, S.R. Selective toxicity by HDAC3 in neurons: regulation by Akt and GSK3beta. *J. Neurosci.* **2011**, *31*, 1746–1751.
403. Fitzsimons, H.L.; Schwartz, S.; Given, F.M.; Scott, M.J. The histone deacetylase HDAC4 regulates long-term memory in *Drosophila*. *PLoS ONE* **2013**, *8*, e83903.
404. Aleksandrova, Y.; Munkuev, A.; Mozhaitsev, E.; Suslov, E.; Tsypyshev, D.; Chaprov, K.; Begunov, R.; Volcho, K.; Salakhutdinov, N.; Neganova, M. Elaboration of the Effective Multi-Target Therapeutic Platform for the Treatment of Alzheimer's Disease Based on Novel Monoterpene-Derived Hydroxamic Acids. *Int. J. Mol. Sci.* **2023**, *24*, 9743.
405. Neganova, M.E.; Klochkov, S.G.; Aleksandrova, Y.R.; Osipov, V.N.; Avdeev, D.V.; Pukhov, S.A.; Gromyko, A.V.; Aliev, G. New Spirocyclic Hydroxamic Acids as Effective Antiproliferative Agents. *Anticancer. Agents Med. Chem.* **2021**, *21*, 597–610.
406. Neganova, M.; Aleksandrova, Y.; Suslov, E.; Mozhaitsev, E.; Munkuev, A.; Tsypyshev, D.; Chicheva, M.; Rogachev, A.; Su-kocheva, O.; Volcho, K.; et al. Novel Multitarget Hydroxamic Acids with a Natural Origin CAP Group against Alzheimer's Disease: Synthesis, Docking and Biological Evaluation. *Pharmaceutics* **2021**, *13*, 1893.
407. Neganova, M.E.; Klochkov, S.G.; Aleksandrova, Y.R.; Aliev, G. The Hydroxamic Acids as Potential Anticancer and Neuroprotective Agents. *Curr. Med. Chem.* **2021**, *28*, 8139–8162.
408. Mann, B.S.; Johnson, J.R.; Cohen, M.H.; Justice, R.; Pazdur, R. FDA approval summary: vorinostat for treatment of advanced primary cutaneous T-cell lymphoma. *Oncologist* **2007**, *12*, 1247–1252.
409. Siegel, D.; Hussein, M.; Belani, C.; Robert, F.; Galanis, E.; Richon, V.M.; Garcia-Vargas, J.; Sanz-Rodriguez, C.; Rizvi, S. Vorinostat in solid and hematologic malignancies. *J. Hematol. Oncol.* **2009**, *2*, 31.
410. Bradley, D.; Rathkopf, D.; Dunn, R.; Stadler, W.M.; Liu, G.; Smith, D.C.; Pili, R.; Zwiebel, J.; Scher, H.; Hussain, M. Vorinostat in advanced prostate cancer patients progressing on prior chemotherapy (National Cancer Institute Trial 6862): trial results and interleukin-6 analysis: a study by the Department of Defense Prostate Cancer Clinical Trial Consortium and University of Chicago Phase 2 Consortium. *Cancer* **2009**, *115*, 5541–5549.
411. Quinn, D.I.; Tsao-Wei, D.D.; Twardowski, P.; Aparicio, A.M.; Frankel, P.; Chatta, G.; Wright, J.J.; Groshen, S.G.; Khoo, S.; Lenz, H.J.; et al. Phase II study of the histone deacetylase inhibitor vorinostat (Suberoylanilide Hydroxamic Acid; SAHA) in recurrent or metastatic transitional cell carcinoma of the urothelium—an NCI-CTEP sponsored: California Cancer Consortium trial, NCI 6879. *Investig. New Drugs* **2021**, *39*, 812–820.
412. DuBois, S.G.; Granger, M.M.; Groshen, S.; Tsao-Wei, D.; Ji, L.; Shamirian, A.; Czarnecki, S.; Goodarzi, F.; Berkovich, R.; Shimada, H.; et al. Randomized Phase II Trial of MIBG Versus MIBG, Vincristine, and Irinotecan Versus MIBG and Vorinostat for Patients With Relapsed or Refractory Neuroblastoma: A Report From NANT Consortium. *J. Clin. Oncol.* **2021**, *39*, 3506–3514.
413. Leary, S.E.S.; Kilburn, L.; Geyer, J.R.; Kocak, M.; Huang, J.; Smith, K.S.; Hadley, J.; Ermoian, R.; MacDonald, T.J.; Goldman, S.; et al. Vorinostat and isotretinoin with chemotherapy in young children with embryonal brain tumors: A report from the Pediatric Brain Tumor Consortium (PBTC-026). *Neuro Oncol.* **2022**, *24*, 1178–1190.
414. Arora, S.P.; Tenner, L.; Sarantopoulos, J.; Morris, J.; Liu, Q.; Mendez, J.A.; Curiel, T.; Michalek, J.; Mahalingam, D. Modulation of autophagy: a Phase II study of vorinostat plus hydroxychloroquine versus regorafenib in chemotherapy-refractory metastatic colorectal cancer (mCRC). *Br. J. Cancer* **2022**, *127*, 1153–1161.
415. Patel, S.; Hurez, V.; Nawrocki, S.T.; Goros, M.; Michalek, J.; Sarantopoulos, J.; Curiel, T.; Mahalingam, D. Vorinostat and hydroxychloroquine improve immunity and inhibit autophagy in metastatic colorectal cancer. *Oncotarget* **2016**, *7*, 59087–59097.
416. Wang, Y.; Janku, F.; Piha-Paul, S.; Hess, K.; Broaddus, R.; Liu, L.; Shi, N.; Overman, M.; Kopetz, S.; Subbiah, V.; et al. Phase I studies of vorinostat with ixazomib or pazopanib imply a role of antiangiogenesis-based therapy for TP53 mutant malignancies. *Sci. Rep.* **2020**, *10*, 3080.
417. Teknos, T.N.; Grecula, J.; Agrawal, A.; Old, M.O.; Ozer, E.; Carrau, R.; Kang, S.; Rocco, J.; Blakaj, D.; Diavolitsis, V.; et al. A phase 1 trial of Vorinostat in combination with concurrent chemoradiation therapy in the treatment of advanced staged head and neck squamous cell carcinoma. *Investig. New Drugs* **2019**, *37*, 702–710.
418. Chan, E.; Arlinghaus, L.R.; Cardin, D.B.; Goff, L.; Berlin, J.D.; Parikh, A.; Abramson, R.G.; Yankeelov, T.E.; Hiebert, S.; Merchant, N.; et al. Phase I trial of vorinostat added to chemoradiation with capecitabine in pancreatic cancer. *Radiother. Oncol.* **2016**, *119*, 312–318.
419. Galanis, E.; Anderson, S.K.; Miller, C.R.; Sarkaria, J.N.; Jaeckle, K.; Buckner, J.C.; Ligon, K.L.; Ballman, K.V.; Moore, D.F., Jr.; Nebozhyn, M.; et al. Phase I/II trial of vorinostat combined with temozolomide and radiation therapy for newly diagnosed glioblastoma: results of Alliance N0874/ABTC 02. *Neuro Oncol.* **2018**, *20*, 546–556.
420. Chen, S.H.; Wu, H.M.; Ossola, B.; Schendzielorz, N.; Wilson, B.C.; Chu, C.H.; Chen, S.L.; Wang, Q.; Zhang, D.; Qian, L.; et al. Suberoylanilide hydroxamic acid, a histone deacetylase inhibitor, protects dopaminergic neurons from neurotoxin-induced damage. *Br. J. Pharmacol.* **2012**, *165*, 494–505.
421. Nuutinen, T.; Suuronen, T.; Kauppinen, A.; Salminen, A. Valproic acid stimulates clusterin expression in human astrocytes: Implications for Alzheimer's disease. *Neurosci. Lett.* **2010**, *475*, 64–68.
422. Kilgore, M.; Miller, C.A.; Fass, D.M.; Hennig, K.M.; Haggarty, S.J.; Sweatt, J.D.; Rumbaugh, G. Inhibitors of class 1 histone deacetylases reverse contextual memory deficits in a mouse model of Alzheimer's disease. *Neuropsychopharmacology* **2010**, *35*, 870–880.

423. Hanson, J.E.; La, H.; Plise, E.; Chen, Y.H.; Ding, X.; Hanania, T.; Sabath, E.V.; Alexandrov, V.; Brunner, D.; Leahy, E.; et al. SAHA enhances synaptic function and plasticity in vitro but has limited brain availability in vivo and does not impact cognition. *PLoS ONE* **2013**, *8*, e69964.
424. C, S.K.; Kakoty, V.; Krishna, K.V.; Dubey, S.K.; Chitkara, D.; Taliyan, R. Neuroprotective Efficacy of Co-Encapsulated Rosiglitazone and Vorinostat Nanoparticle on Streptozotocin Induced Mice Model of Alzheimer Disease. *ACS Chem. Neurosci.* **2021**, *12*, 1528–1541.
425. Kakoty, V.; C, S.K.; Dubey, S.K.; Yang, C.H.; Marathe, S.A.; Taliyan, R. Epigenetic regulation and autophagy modulation debilitates insulin resistance associated Alzheimer's disease condition in rats. *Metab. Brain Dis.* **2022**, *37*, 927–944.
426. Cuadrado-Tejedor, M.; Garcia-Barroso, C.; Sanchez-Arias, J.; Mederos, S.; Rabal, O.; Ugarte, A.; Franco, R.; Pascual-Lucas, M.; Segura, V.; Perea, G.; et al. Concomitant histone deacetylase and phosphodiesterase 5 inhibition synergistically prevents the disruption in synaptic plasticity and it reverses cognitive impairment in a mouse model of Alzheimer's disease. *Clin. Epigenetics* **2015**, *7*, 108.
427. More, S.S.; Itsara, M.; Yang, X.; Geier, E.G.; Tadano, M.K.; Seo, Y.; Vanbrocklin, H.F.; Weiss, W.A.; Mueller, S.; Haas-Kogan, D.A.; et al. Vorinostat increases expression of functional norepinephrine transporter in neuroblastoma in vitro and in vivo model systems. *Clin. Cancer Res.* **2011**, *17*, 2339–2349.
428. DuBois, S.G.; Groshen, S.; Park, J.R.; Haas-Kogan, D.A.; Yang, X.; Geier, E.; Chen, E.; Giacomini, K.; Weiss, B.; Cohn, S.L.; et al. Phase I Study of Vorinostat as a Radiation Sensitizer with <sup>131</sup>I-Metaiodobenzylguanidine (<sup>131</sup>I-MIBG) for Patients with Relapsed or Refractory Neuroblastoma. *Clin. Cancer Res.* **2015**, *21*, 2715–2721.
429. Sinha, S.; Cheng, K.; Schaffer, A.A.; Aldape, K.; Schiff, E.; Ruppin, E. In vitro and in vivo identification of clinically approved drugs that modify ACE2 expression. *Mol. Syst. Biol.* **2020**, *16*, e9628.
430. Pinto, N.; DuBois, S.G.; Marachelian, A.; Diede, S.J.; Taraseviciute, A.; Glade Bender, J.L.; Tsao-Wei, D.; Groshen, S.G.; Reid, J.M.; Haas-Kogan, D.A.; et al. Phase I study of vorinostat in combination with isotretinoin in patients with refractory/recurrent neuroblastoma: A new approaches to Neuroblastoma Therapy (NANT) trial. *Pediatr. Blood Cancer* **2018**, *65*, e27023.
431. Mahalingam, D.; Mita, M.; Sarantopoulos, J.; Wood, L.; Amaravadi, R.K.; Davis, L.E.; Mita, A.C.; Curiel, T.J.; Espitia, C.M.; Nawrocki, S.T.; et al. Combined autophagy and HDAC inhibition: a phase I safety, tolerability, pharmacokinetic, and pharmacodynamic analysis of hydroxychloroquine in combination with the HDAC inhibitor vorinostat in patients with advanced solid tumors. *Autophagy* **2014**, *10*, 1403–1414.
432. Fu, S.; Hou, M.M.; Naing, A.; Janku, F.; Hess, K.; Zinner, R.; Subbiah, V.; Hong, D.; Wheler, J.; Piha-Paul, S.; et al. Phase I study of pazopanib and vorinostat: a therapeutic approach for inhibiting mutant p53-mediated angiogenesis and facilitating mutant p53 degradation. *Ann. Oncol.* **2015**, *26*, 1012–1018.
433. Yang, J.; Zhou, R.; Ma, Z. Autophagy and Energy Metabolism. *Adv. Exp. Med. Biol.* **2019**, *1206*, 329–357.
434. Judge, A.; Dodd, M.S. Metabolism. *Essays Biochem.* **2020**, *64*, 607–647.
435. Martinez-Reyes, I.; Chandel, N.S. Cancer metabolism: looking forward. *Nat. Rev. Cancer* **2021**, *21*, 669–680.
436. Kroemer, G.; Pouyssegur, J. Tumor cell metabolism: cancer's Achilles' heel. *Cancer Cell* **2008**, *13*, 472–482.
437. Pascale, R.M.; Calvisi, D.F.; Simile, M.M.; Feo, C.F.; Feo, F. The Warburg Effect 97 Years after Its Discovery. *Cancers* **2020**, *12*, 2819.
438. Sun, X.; Peng, Y.; Zhao, J.; Xie, Z.; Lei, X.; Tang, G. Discovery and development of tumor glycolysis rate-limiting enzyme inhibitors. *Bioorg Chem.* **2021**, *112*, 104891.
439. Fernie, A.R.; Carrari, F.; Sweetlove, L.J. Respiratory metabolism: glycolysis, the TCA cycle and mitochondrial electron transport. *Curr. Opin. Plant Biol.* **2004**, *7*, 254–261.
440. Ward, P.S.; Thompson, C.B. Metabolic reprogramming: a cancer hallmark even warburg did not anticipate. *Cancer Cell* **2012**, *21*, 297–308.
441. Yoshida, G.J. Metabolic reprogramming: the emerging concept and associated therapeutic strategies. *J. Exp. Clin. Cancer Res.* **2015**, *34*, 111.
442. Jia, D.; Park, J.H.; Jung, K.H.; Levine, H.; Kaiparettu, B.A. Elucidating the Metabolic Plasticity of Cancer: Mitochondrial Reprogramming and Hybrid Metabolic States. *Cells* **2018**, *7*, 21.
443. Pfeiffer, T.; Schuster, S.; Bonhoeffer, S. Cooperation and competition in the evolution of ATP-producing pathways. *Science* **2001**, *292*, 504–507.
444. Locasale, J.W.; Cantley, L.C. Altered metabolism in cancer. *BMC Biol.* **2010**, *8*, 88.
445. Srinivasan, S.; Guha, M.; Kashina, A.; Avadhani, N.G. Mitochondrial dysfunction and mitochondrial dynamics-The cancer connection. *Biochim. Biophys. Acta Bioenerg.* **2017**, *1858*, 602–614.
446. Shi, Y.; Wang, Y.; Jiang, H.; Sun, X.; Xu, H.; Wei, X.; Wei, Y.; Xiao, G.; Song, Z.; Zhou, F. Mitochondrial dysfunction induces radioresistance in colorectal cancer by activating [Ca<sup>2+</sup>]<sub>i</sub>(m)-PDP1-PDH-histone acetylation retrograde signaling. *Cell Death Dis.* **2021**, *12*, 837.
447. Ma, J.; Zhang, Q.; Chen, S.; Fang, B.; Yang, Q.; Chen, C.; Miele, L.; Sarkar, F.H.; Xia, J.; Wang, Z. Mitochondrial dysfunction promotes breast cancer cell migration and invasion through HIF1α accumulation via increased production of reactive oxygen species. *PLoS ONE* **2013**, *8*, e69485.
448. Lee, H.C.; Huang, K.H.; Yeh, T.S.; Chi, C.W. Somatic alterations in mitochondrial DNA and mitochondrial dysfunction in gastric cancer progression. *World J. Gastroenterol.* **2014**, *20*, 3950–3959.

449. Roth, K.G.; Mambetsariev, I.; Kulkarni, P.; Salgia, R. The Mitochondrion as an Emerging Therapeutic Target in Cancer. *Trends Mol. Med.* **2020**, *26*, 119–134.
450. Almaguel, F.A.; Sanchez, T.W.; Ortiz-Hernandez, G.L.; Casiano, C.A. Alpha-Enolase: Emerging Tumor-Associated Antigen, Cancer Biomarker, and Oncotherapeutic Target. *Front. Genet.* **2020**, *11*, 614726.
451. Kohnken, R.; Kodigepalli, K.M.; Wu, L. Regulation of deoxynucleotide metabolism in cancer: novel mechanisms and therapeutic implications. *Mol. Cancer* **2015**, *14*, 176.
452. Le, T.M.; Poddar, S.; Capri, J.R.; Abt, E.R.; Kim, W.; Wei, L.; Uong, N.T.; Cheng, C.M.; Braas, D.; Nikanjam, M.; et al. ATR inhibition facilitates targeting of leukemia dependence on convergent nucleotide biosynthetic pathways. *Nat. Commun.* **2017**, *8*, 241.
453. Efimova, E.V.; Takahashi, S.; Shamsi, N.A.; Wu, D.; Labay, E.; Ulanovskaya, O.A.; Weichselbaum, R.R.; Kozmin, S.A.; Kron, S.J. Linking Cancer Metabolism to DNA Repair and Accelerated Senescence. *Mol. Cancer Res.* **2016**, *14*, 173–184.
454. Backos, D.S.; Franklin, C.C.; Reigan, P. The role of glutathione in brain tumor drug resistance. *Biochem. Pharmacol.* **2012**, *83*, 1005–1012.
455. Traverso, N.; Ricciarelli, R.; Nitti, M.; Marengo, B.; Furfaro, A.L.; Pronzato, M.A.; Marinari, U.M.; Domenicotti, C. Role of glutathione in cancer progression and chemoresistance. *Oxid. Med. Cell Longev.* **2013**, *2013*, 972913.
456. Thorens, B.; Mueckler, M. Glucose transporters in the 21st Century. *Am. J. Physiol. Endocrinol. Metab.* **2010**, *298*, E141–5.
457. Deng, D.; Yan, N. GLUT, SGLT, and SWEET: Structural and mechanistic investigations of the glucose transporters. *Protein Sci.* **2016**, *25*, 546–558.
458. Lu, Y.Y.; Wu, C.H.; Hong, C.H.; Chang, K.L.; Lee, C.H. GLUT-1 Enhances Glycolysis, Oxidative Stress, and Fibroblast Proliferation in Keloid. *Life* **2021**, *11*, 505.
459. Reckzeh, E.S.; Waldmann, H. Small-Molecule Inhibition of Glucose Transporters GLUT-1-4. *Chembiochem* **2020**, *21*, 45–52.
460. Carvalho, K.C.; Cunha, I.W.; Rocha, R.M.; Ayala, F.R.; Cajaiba, M.M.; Begnami, M.D.; Vilela, R.S.; Paiva, G.R.; Andrade, R.G.; Soares, F.A. GLUT1 expression in malignant tumors and its use as an immunodiagnostic marker. *Clinics* **2011**, *66*, 965–972.
461. Yin, C.; Gao, B.; Yang, J.; Wu, J. Glucose Transporter-1 (GLUT-1) Expression is Associated with Tumor Size and Poor Prognosis in Locally Advanced Gastric Cancer. *Med. Sci. Monit. Basic. Res.* **2020**, *26*, e920778.
462. Higashi, T.; Tamaki, N.; Torizuka, T.; Nakamoto, Y.; Sakahara, H.; Kimura, T.; Honda, T.; Inokuma, T.; Katsushima, S.; Ohshio, G.; et al. FDG uptake, GLUT-1 glucose transporter and cellularity in human pancreatic tumors. *J. Nucl. Med.* **1998**, *39*, 1727–1735.
463. Gasinska, A.; Jaszczynski, J.; Rychlik, U.; Luczynska, E.; Pogodzinski, M.; Palaczynski, M. Prognostic Significance of Serum PSA Level and Telomerase, VEGF and GLUT-1 Protein Expression for the Biochemical Recurrence in Prostate Cancer Patients after Radical Prostatectomy. *Pathol. Oncol. Res.* **2020**, *26*, 1049–1056.
464. Carreno, D.; Corro, N.; Torres-Estay, V.; Veliz, L.P.; Jaimovich, R.; Cisternas, P.; San Francisco, I.F.; Sotomayor, P.C.; Tanasova, M.; Inestrosa, N.C.; et al. Fructose and prostate cancer: toward an integrated view of cancer cell metabolism. *Prostate Cancer Prostatic Dis.* **2019**, *22*, 49–58.
465. Mogi, A.; Koga, K.; Aoki, M.; Hamasaki, M.; Uesugi, N.; Iwasaki, A.; Shirakusa, T.; Tamura, K.; Nabeshima, K. Expression and role of GLUT-1, MCT-1, and MCT-4 in malignant pleural mesothelioma. *Virchows Arch.* **2013**, *462*, 83–93.
466. Khabaz, M.N.; Qureshi, I.A.; Al-Maghrabi, J.A. GLUT 1 expression is a supportive mean in predicting prognosis and survival estimates of endometrial carcinoma. *Ginekol. Pol.* **2019**, *90*, 582–588.
467. Canpolat, T.; Ersoz, C.; Uguz, A.; Vardar, M.A.; Altintas, A. GLUT-1 Expression in Proliferative Endometrium, Endometrial Hyperplasia, Endometrial Adenocarcinoma and the Relationship Between GLUT-1 Expression and Prognostic Parameters in Endometrial Adenocarcinoma. *Turk. Patoloji Derg.* **2016**, *32*, 141–147.
468. Kubo, T.; Shimose, S.; Fujimori, J.; Furuta, T.; Arihiro, K.; Ochi, M. Does expression of glucose transporter protein-1 relate to prognosis and angiogenesis in osteosarcoma? *Clin. Orthop. Relat. Res.* **2015**, *473*, 305–310.
469. van de Nes, J.A.; Griewank, K.G.; Schmid, K.W.; Grabellus, F. Immunocytochemical analysis of glucose transporter protein-1 (GLUT-1) in typical, brain invasive, atypical and anaplastic meningioma. *Neuropathology* **2015**, *35*, 24–36.
470. Luo, X.M.; Zhou, S.H.; Fan, J. Glucose transporter-1 as a new therapeutic target in laryngeal carcinoma. *J. Int. Med. Res.* **2010**, *38*, 1885–1892.
471. Xi, J.; Wang, Y.; Liu, H. GLUT-1 participates in the promotion of lncRNA CASC9 in proliferation and metastasis of laryngeal carcinoma cells. *Gene* **2020**, *726*, 144194.
472. Krzeslak, A.; Wojcik-Krowiranda, K.; Forma, E.; Jozwiak, P.; Romanowicz, H.; Bienkiewicz, A.; Brys, M. Expression of GLUT1 and GLUT3 glucose transporters in endometrial and breast cancers. *Pathol. Oncol. Res.* **2012**, *18*, 721–728.
473. Deng, Y.; Zou, J.; Deng, T.; Liu, J. Clinicopathological and prognostic significance of GLUT1 in breast cancer: A meta-analysis. *Medicine* **2018**, *97*, e12961.
474. Kang, S.S.; Chun, Y.K.; Hur, M.H.; Lee, H.K.; Kim, Y.J.; Hong, S.R.; Lee, J.H.; Lee, S.G.; Park, Y.K. Clinical significance of glucose transporter 1 (GLUT1) expression in human breast carcinoma. *Jpn. J. Cancer Res.* **2002**, *93*, 1123–1128.
475. Hussein, Y.R.; Bandyopadhyay, S.; Semaan, A.; Ahmed, Q.; Albashiti, B.; Jazaerly, T.; Nahleh, Z.; Ali-Fehmi, R. Glut-1 Expression Correlates with Basal-like Breast Cancer. *Transl. Oncol.* **2011**, *4*, 321–327.
476. Fukushi, A.; Kim, H.D.; Chang, Y.C.; Kim, C.H. Revisited Metabolic Control and Reprogramming Cancers by Means of the Warburg Effect in Tumor Cells. *Int. J. Mol. Sci.* **2022**, *23*, 10037.

477. Ciscato, F.; Ferrone, L.; Masgras, I.; Laquatra, C.; Rasola, A. Hexokinase 2 in Cancer: A Prima Donna Playing Multiple Characters. *Int. J. Mol. Sci.* **2021**, *22*, 4716.
478. Garcia, S.N.; Guedes, R.C.; Marques, M.M. Unlocking the Potential of HK2 in Cancer Metabolism and Therapeutics. *Curr. Med. Chem.* **2019**, *26*, 7285–7322.
479. Gottlob, K.; Majewski, N.; Kennedy, S.; Kandel, E.; Robey, R.B.; Hay, N. Inhibition of early apoptotic events by Akt/PKB is dependent on the first committed step of glycolysis and mitochondrial hexokinase. *Genes. Dev.* **2001**, *15*, 1406–1418.
480. Jiao, L.; Zhang, H.L.; Li, D.D.; Yang, K.L.; Tang, J.; Li, X.; Ji, J.; Yu, Y.; Wu, R.Y.; Ravichandran, S.; et al. Regulation of glycolytic metabolism by autophagy in liver cancer involves selective autophagic degradation of HK2 (hexokinase 2). *Autophagy* **2018**, *14*, 671–684.
481. Zhang, X.Y.; Zhang, M.; Cong, Q.; Zhang, M.X.; Zhang, M.Y.; Lu, Y.Y.; Xu, C.J. Hexokinase 2 confers resistance to cisplatin in ovarian cancer cells by enhancing cisplatin-induced autophagy. *Int. J. Biochem. Cell Biol.* **2018**, *95*, 9–16.
482. Tian, X.; Liu, D.; Zuo, X.; Sun, X.; Wu, M.; Li, X.; Teng, Y. Hexokinase 2 promoted cell motility and proliferation by activating Akt1/p-Akt1 in human ovarian cancer cells. *J. Ovarian Res.* **2022**, *15*, 92.
483. Yang, L.; Yan, X.; Chen, J.; Zhan, Q.; Hua, Y.; Xu, S.; Li, Z.; Wang, Z.; Dong, Y.; Zuo, D.; et al. Hexokinase 2 discerns a novel circulating tumor cell population associated with poor prognosis in lung cancer patients. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2012228118.
484. Deng, Y.; Lu, J. Targeting hexokinase 2 in castration-resistant prostate cancer. *Mol. Cell Oncol.* **2015**, *2*, e974465.
485. Rho, M.; Kim, J.; Jee, C.D.; Lee, Y.M.; Lee, H.E.; Kim, M.A.; Lee, H.S.; Kim, W.H. Expression of type 2 hexokinase and mitochondria-related genes in gastric carcinoma tissues and cell lines. *Anticancer Res.* **2007**, *27*, 251–258.
486. Sato-Tadano, A.; Suzuki, T.; Amari, M.; Takagi, K.; Miki, Y.; Tamaki, K.; Watanabe, M.; Ishida, T.; Sasano, H.; Ohuchi, N. Hexokinase II in breast carcinoma: a potent prognostic factor associated with hypoxia-inducible factor-1 $\alpha$  and Ki-67. *Cancer Sci.* **2013**, *104*, 1380–1388.
487. Zhang, D.; Wang, H.; Yu, W.; Qiao, F.; Su, X.; Xu, H. Downregulation of hexokinase 2 improves radiosensitivity of breast cancer. *Transl. Cancer Res.* **2019**, *8*, 290–297.
488. Shanguan, X.; He, J.; Ma, Z.; Zhang, W.; Ji, Y.; Shen, K.; Yue, Z.; Li, W.; Xin, Z.; Zheng, Q.; et al. SUMOylation controls the binding of hexokinase 2 to mitochondria and protects against prostate cancer tumorigenesis. *Nat. Commun.* **2021**, *12*, 1812.
489. Wang, J.; Shao, F.; Yang, Y.; Wang, W.; Yang, X.; Li, R.; Cheng, H.; Sun, S.; Feng, X.; Gao, Y.; et al. A non-metabolic function of hexokinase 2 in small cell lung cancer: promotes cancer cell stemness by increasing USP11-mediated CD133 stability. *Cancer Commun.* **2022**, *42*, 1008–1027.
490. Thomas, G.E.; Egan, G.; Garcia-Prat, L.; Botham, A.; Voisin, V.; Patel, P.S.; Hoff, F.W.; Chin, J.; Nachmias, B.; Kaufmann, K.B.; et al. The metabolic enzyme hexokinase 2 localizes to the nucleus in AML and normal haematopoietic stem and progenitor cells to maintain stemness. *Nat. Cell Biol.* **2022**, *24*, 872–884.
491. Yi, M.; Ban, Y.; Tan, Y.; Xiong, W.; Li, G.; Xiang, B. 6-Phosphofructo-2-kinase/fructose-2,6-bisphosphatase 3 and 4: A pair of valves for fine-tuning of glucose metabolism in human cancer. *Mol. Metab.* **2019**, *20*, 1–13.
492. Yalcin, A.; Solakoglu, T.H.; Ozcan, S.C.; Guzel, S.; Peker, S.; Celikler, S.; Balaban, B.D.; Sevinc, E.; Gurpinar, Y.; Chesney, J.A. 6-phosphofructo-2-kinase/fructose 2,6-bisphosphatase-3 is required for transforming growth factor beta1-enhanced invasion of Panc1 cells in vitro. *Biochem. Biophys. Res. Commun.* **2017**, *484*, 687–693.
493. Lei, L.; Hong, L.L.; Ling, Z.N.; Zhong, Y.; Hu, X.Y.; Li, P.; Ling, Z.Q. A Potential Oncogenic Role for PFKFB3 Overexpression in Gastric Cancer Progression. *Clin. Transl. Gastroenterol.* **2021**, *12*, e00377.
494. Moon, J.S.; Jin, W.J.; Kwak, J.H.; Kim, H.J.; Yun, M.J.; Kim, J.W.; Park, S.W.; Kim, K.S. Androgen stimulates glycolysis for de novo lipid synthesis by increasing the activities of hexokinase 2 and 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase 2 in prostate cancer cells. *Biochem. J.* **2011**, *433*, 225–233.
495. Zadra, G.; Ribeiro, C.F.; Chetta, P.; Ho, Y.; Cacciatore, S.; Gao, X.; Syamala, S.; Bango, C.; Photopoulos, C.; Huang, Y.; et al. Inhibition of de novo lipogenesis targets androgen receptor signaling in castration-resistant prostate cancer. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 631–640.
496. Alvarez, R.; Mandal, D.; Chittiboina, P. Canonical and Non-Canonical Roles of PFKFB3 in Brain Tumors. *Cells* **2021**, *10*, 2913.
497. Chen, J.; Xie, J.; Jiang, Z.; Wang, B.; Wang, Y.; Hu, X. Shikonin and its analogs inhibit cancer cell glycolysis by targeting tumor pyruvate kinase-M2. *Oncogene* **2011**, *30*, 4297–4306.
498. Zhu, S.; Guo, Y.; Zhang, X.; Liu, H.; Yin, M.; Chen, X.; Peng, C. Pyruvate kinase M2 (PKM2) in cancer and cancer therapeutics. *Cancer Lett.* **2021**, *503*, 240–248.
499. Xia, Y.; Wang, X.; Liu, Y.; Shapiro, E.; Lepor, H.; Tang, M.S.; Sun, T.T.; Wu, X.R. PKM2 Is Essential for Bladder Cancer Growth and Maintenance. *Cancer Res.* **2022**, *82*, 571–585.
500. Yu, S.; Zang, W.; Qiu, Y.; Liao, L.; Zheng, X. Deubiquitinase OTUB2 exacerbates the progression of colorectal cancer by promoting PKM2 activity and glycolysis. *Oncogene* **2022**, *41*, 46–56.
501. Zhou, S.; Li, D.; Xiao, D.; Wu, T.; Hu, X.; Zhang, Y.; Deng, J.; Long, J.; Xu, S.; Wu, J.; et al. Inhibition of PKM2 Enhances Sensitivity of Olaparib to Ovarian Cancer Cells and Induces DNA Damage. *Int. J. Biol. Sci.* **2022**, *18*, 1555–1568.
502. Huang, Y.; Chen, L.M.; Xie, J.Y.; Han, H.; Zhu, B.F.; Wang, L.J.; Wang, W.J. High Expression of PKM2 Was Associated with the Poor Prognosis of Acute Leukemia. *Cancer Manag. Res.* **2021**, *13*, 7851–7858.
503. Wang, L.; Yang, L.; Yang, Z.; Tang, Y.; Tao, Y.; Zhan, Q.; Lei, L.; Jing, Y.; Jiang, X.; Jin, H.; et al. Glycolytic Enzyme PKM2 Mediates Autophagic Activation to Promote Cell Survival in NPM1-Mutated Leukemia. *Int. J. Biol. Sci.* **2019**, *15*, 882–894.

504. Cogley, J.N.; Fiorello, M.L.; Bailey, D.M. 13 reasons why the brain is susceptible to oxidative stress. *Redox Biol.* **2018**, *15*, 490–503.
505. Ardanaz, C.G.; Ramirez, M.J.; Solas, M. Brain Metabolic Alterations in Alzheimer's Disease. *Int. J. Mol. Sci.* **2022**, *23*, 3785.
506. Holper, L.; Ben-Shachar, D.; Mann, J.J. Multivariate meta-analyses of mitochondrial complex I and IV in major depressive disorder, bipolar disorder, schizophrenia, Alzheimer disease, and Parkinson disease. *Neuropsychopharmacology* **2019**, *44*, 837–849.
507. Holper, L.; Lan, M.J.; Brown, P.J.; Sublette, E.M.; Burke, A.; Mann, J.J. Brain cytochrome-c-oxidase as a marker of mitochondrial function: A pilot study in major depression using NIRS. *Depress. Anxiety* **2019**, *36*, 766–779.
508. Bergman, O.; Karry, R.; Milhem, J.; Ben-Shachar, D. NDUFV2 pseudogene (NDUFV2P1) contributes to mitochondrial complex I deficits in schizophrenia. *Mol. Psychiatry* **2020**, *25*, 805–820.
509. Rice, M.W.; Smith, K.L.; Roberts, R.C.; Perez-Costas, E.; Melendez-Ferro, M. Assessment of cytochrome C oxidase dysfunction in the substantia nigra/ventral tegmental area in schizophrenia. *PLoS ONE* **2014**, *9*, e100054.
510. Gonzalez-Rodriguez, P.; Zampese, E.; Stout, K.A.; Guzman, J.N.; Ilijic, E.; Yang, B.; Tkatch, T.; Stavarahe, M.A.; Wokosin, D.L.; Gao, L.; et al. Disruption of mitochondrial complex I induces progressive parkinsonism. *Nature* **2021**, *599*, 650–656.
511. Yan, M.H.; Wang, X.; Zhu, X. Mitochondrial defects and oxidative stress in Alzheimer disease and Parkinson disease. *Free Radic. Biol. Med.* **2013**, *62*, 90–101.
512. Dong, Y.; Brewer, G.J. Global Metabolic Shifts in Age and Alzheimer's Disease Mouse Brains Pivot at NAD<sup>+</sup>/NADH Redox Sites. *J. Alzheimers Dis.* **2019**, *71*, 119–140.
513. Ohta, S.; Ohsawa, I. Dysfunction of mitochondria and oxidative stress in the pathogenesis of Alzheimer's disease: on defects in the cytochrome c oxidase complex and aldehyde detoxification. *J. Alzheimers Dis.* **2006**, *9*, 155–166.
514. Lunnun, K.; Ibrahim, Z.; Proitsi, P.; Lourdasamy, A.; Newhouse, S.; Sattlecker, M.; Furney, S.; Saleem, M.; Soininen, H.; Kloszewska, I.; et al. Mitochondrial dysfunction and immune activation are detectable in early Alzheimer's disease blood. *J. Alzheimers Dis.* **2012**, *30*, 685–710.
515. Jaroudi, W.; Garami, J.; Garrido, S.; Hornberger, M.; Keri, S.; Moustafa, A.A. Factors underlying cognitive decline in old age and Alzheimer's disease: the role of the hippocampus. *Rev. Neurosci.* **2017**, *28*, 705–714.
516. Fonseca, L.M.; Yokomizo, J.E.; Bottino, C.M.; Fuentes, D. Frontal Lobe Degeneration in Adults with Down Syndrome and Alzheimer's Disease: A Review. *Dement. Geriatr. Cogn. Disord.* **2016**, *41*, 123–136.
517. Berron, D.; van Westen, D.; Ossenkoppele, R.; Strandberg, O.; Hansson, O. Medial temporal lobe connectivity and its associations with cognition in early Alzheimer's disease. *Brain* **2020**, *143*, 1233–1248.
518. Ahullo-Fuster, M.A.; Ortiz, T.; Varela-Donoso, E.; Nacher, J.; Sanchez-Sanchez, M.L. The Parietal Lobe in Alzheimer's Disease and Blindness. *J. Alzheimers Dis.* **2022**, *89*, 1193–1202.
519. Perluigi, M.; Barone, E.; Di Domenico, F.; Butterfield, D.A. Aberrant protein phosphorylation in Alzheimer disease brain disturbs pro-survival and cell death pathways. *Biochim. Biophys. Acta* **2016**, *1862*, 1871–1882.
520. Francis, B.M.; Yang, J.; Song, B.J.; Gupta, S.; Maj, M.; Bazinet, R.P.; Robinson, B.; Mount, H.T. Reduced levels of mitochondrial complex I subunit NDUFB8 and linked complex I + III oxidoreductase activity in the TgCRND8 mouse model of Alzheimer's disease. *J. Alzheimers Dis.* **2014**, *39*, 347–355.
521. Adav, S.S.; Park, J.E.; Sze, S.K. Quantitative profiling brain proteomes revealed mitochondrial dysfunction in Alzheimer's disease. *Mol. Brain* **2019**, *12*, 8.
522. Emmerzaal, T.L.; Rodenburg, R.J.; Tanila, H.; Verweij, V.; Kiliaan, A.J.; Kozicz, T. Age-Dependent Decrease of Mitochondrial Complex II Activity in a Familial Mouse Model for Alzheimer's Disease. *J. Alzheimers Dis.* **2018**, *66*, 75–82.
523. Webster, S.J.; Bachstetter, A.D.; Van Eldik, L.J. Comprehensive behavioral characterization of an APP/PS-1 double knock-in mouse model of Alzheimer's disease. *Alzheimers Res. Ther.* **2013**, *5*, 28.
524. Zhang, C.; Rissman, R.A.; Feng, J. Characterization of ATP alterations in an Alzheimer's disease transgenic mouse model. *J. Alzheimers Dis.* **2015**, *44*, 375–378.
525. Manczak, M.; Calkins, M.J.; Reddy, P.H. Impaired mitochondrial dynamics and abnormal interaction of amyloid beta with mitochondrial protein Drp1 in neurons from patients with Alzheimer's disease: implications for neuronal damage. *Hum. Mol. Genet.* **2011**, *20*, 2495–2509.
526. Manczak, M.; Kandimalla, R.; Fry, D.; Sesaki, H.; Reddy, P.H. Protective effects of reduced dynamin-related protein 1 against amyloid beta-induced mitochondrial dysfunction and synaptic damage in Alzheimer's disease. *Hum. Mol. Genet.* **2016**, *25*, 5148–5166.
527. Deak, F.; Freeman, W.M.; Ungvari, Z.; Csiszar, A.; Sonntag, W.E. Recent Developments in Understanding Brain Aging: Implications for Alzheimer's Disease and Vascular Cognitive Impairment. *J. Gerontol. A Biol. Sci. Med. Sci.* **2016**, *71*, 13–20.
528. Kim, D.I.; Lee, K.H.; Oh, J.Y.; Kim, J.S.; Han, H.J. Relationship Between beta-Amyloid and Mitochondrial Dynamics. *Cell Mol. Neurobiol.* **2017**, *37*, 955–968.
529. Shi, C.; Zhu, X.; Wang, J.; Long, D. Intramitochondrial I $\kappa$ B/NF- $\kappa$ B signaling pathway is involved in amyloid beta peptide-induced mitochondrial dysfunction. *J. Bioenerg. Biomembr.* **2014**, *46*, 371–376.
530. Yamazaki, Y.; Fujii, S. Extracellular ATP modulates synaptic plasticity induced by activation of metabotropic glutamate receptors in the hippocampus. *Biomed. Res.* **2015**, *36*, 1–9.
531. Recuero, M.; Munoz, T.; Aldudo, J.; Subias, M.; Bullido, M.J.; Valdivieso, F. A free radical-generating system regulates APP metabolism/processing. *FEBS Lett.* **2010**, *584*, 4611–4618.

532. Leuner, K.; Schutt, T.; Kurz, C.; Eckert, S.H.; Schiller, C.; Occhipinti, A.; Mai, S.; Jendrach, M.; Eckert, G.P.; Kruse, S.E.; et al. Mitochondrion-derived reactive oxygen species lead to enhanced amyloid beta formation. *Antioxid. Redox Signal* **2012**, *16*, 1421–1433.
533. Djordjevic, J.; Roy Chowdhury, S.; Snow, W.M.; Perez, C.; Cadonic, C.; Fernyhough, P.; Albensi, B.C. Early Onset of Sex-Dependent Mitochondrial Deficits in the Cortex of 3xTg Alzheimer's Mice. *Cells* **2020**, *9*, 1541.
534. Joh, Y.; Choi, W.S. Mitochondrial Complex I Inhibition Accelerates Amyloid Toxicity. *Dev. Reprod.* **2017**, *21*, 417–424.
535. Lasagna-Reeves, C.A.; Castillo-Carranza, D.L.; Sengupta, U.; Clos, A.L.; Jackson, G.R.; Kaye, R. Tau oligomers impair memory and induce synaptic and mitochondrial dysfunction in wild-type mice. *Mol. Neurodegener.* **2011**, *6*, 39.
536. David, D.C.; Hauptmann, S.; Scherping, I.; Schuessel, K.; Keil, U.; Rizzu, P.; Ravid, R.; Drose, S.; Brandt, U.; Muller, W.E.; et al. Proteomic and functional analyses reveal a mitochondrial dysfunction in P301L tau transgenic mice. *J. Biol. Chem.* **2005**, *280*, 23802–23814.
537. Gotz, J.; Chen, F.; Barmettler, R.; Nitsch, R.M. Tau filament formation in transgenic mice expressing P301L tau. *J. Biol. Chem.* **2001**, *276*, 529–534.
538. Rhein, V.; Song, X.; Wiesner, A.; Ittner, L.M.; Baysang, G.; Meier, F.; Ozmen, L.; Bluethmann, H.; Drose, S.; Brandt, U.; et al. Amyloid-beta and tau synergistically impair the oxidative phosphorylation system in triple transgenic Alzheimer's disease mice. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 20057–20062.
539. Yamada, E.S.; Respondek, G.; Mussner, S.; de Andrade, A.; Hollerhage, M.; Depienne, C.; Rastetter, A.; Tarze, A.; Friguet, B.; Salama, M.; et al. Annonacin, a natural lipophilic mitochondrial complex I inhibitor, increases phosphorylation of tau in the brain of FTDP-17 transgenic mice. *Exp. Neurol.* **2014**, *253*, 113–125.
540. Liu, Y.; Cao, Y.; Zhang, W.; Bergmeier, S.; Qian, Y.; Akbar, H.; Colvin, R.; Ding, J.; Tong, L.; Wu, S.; et al. A small-molecule inhibitor of glucose transporter 1 downregulates glycolysis, induces cell-cycle arrest, and inhibits cancer cell growth in vitro and in vivo. *Mol. Cancer Ther.* **2012**, *11*, 1672–1682.
541. Peng, Y.; Xing, S.N.; Tang, H.Y.; Wang, C.D.; Yi, F.P.; Liu, G.L.; Wu, X.M. Influence of glucose transporter 1 activity inhibition on neuroblastoma in vitro. *Gene* **2019**, *689*, 11–17.
542. Shibuya, K.; Okada, M.; Suzuki, S.; Seino, M.; Seino, S.; Takeda, H.; Kitanaka, C. Targeting the facilitative glucose transporter GLUT1 inhibits the self-renewal and tumor-initiating capacity of cancer stem cells. *Oncotarget* **2015**, *6*, 651–661.
543. Zhang, R.S.; Li, Z.K.; Liu, J.; Deng, Y.T.; Jiang, Y. WZB117 enhanced the anti-tumor effect of apatinib against melanoma via blocking STAT3/PKM2 axis. *Front. Pharmacol.* **2022**, *13*, 976117.
544. Zhao, F.; Ming, J.; Zhou, Y.; Fan, L. Inhibition of Glut1 by WZB117 sensitizes radioresistant breast cancer cells to irradiation. *Cancer Chemother. Pharmacol.* **2016**, *77*, 963–972.
545. Zhu, Y.; Wang, A.; Zhang, S.; Kim, J.; Xia, J.; Zhang, F.; Wang, D.; Wang, Q.; Wang, J. Paclitaxel-loaded ginsenoside Rg3 liposomes for drug-resistant cancer therapy by dual targeting of the tumor microenvironment and cancer cells. *J. Adv. Res.* **2023**, *49*, 159–173.
546. Klippel, S.; Jakubikova, J.; Delmore, J.; Ooi, M.; McMillin, D.; Kastritis, E.; Laubach, J.; Richardson, P.G.; Anderson, K.C.; Mitsiades, C.S. Methyljasmonate displays in vitro and in vivo activity against multiple myeloma cells. *Br. J. Haematol.* **2012**, *159*, 340–351.
547. Li, J.; Chen, K.; Wang, F.; Dai, W.; Li, S.; Feng, J.; Wu, L.; Liu, T.; Xu, S.; Xia, Y.; et al. Methyl jasmonate leads to necrosis and apoptosis in hepatocellular carcinoma cells via inhibition of glycolysis and represses tumor growth in mice. *Oncotarget* **2017**, *8*, 45965–45980.
548. Jiang, Y.X.; Siu, M.K.Y.; Wang, J.J.; Leung, T.H.Y.; Chan, D.W.; Cheung, A.N.Y.; Ngan, H.Y.S.; Chan, K.K.L. PFKFB3 Regulates Chemoresistance, Metastasis and Stemness via IAP Proteins and the NF-kappaB Signaling Pathway in Ovarian Cancer. *Front. Oncol.* **2022**, *12*, 748403.
549. Wang, Y.; Hao, F.; Nan, Y.; Qu, L.; Na, W.; Jia, C.; Chen, X. PKM2 Inhibitor Shikonin Overcomes the Cisplatin Resistance in Bladder Cancer by Inducing Necroptosis. *Int. J. Biol. Sci.* **2018**, *14*, 1883–1891.
550. Dai, Y.; Liu, Y.; Li, J.; Jin, M.; Yang, H.; Huang, G. Shikonin inhibited glycolysis and sensitized cisplatin treatment in non-small cell lung cancer cells via the exosomal pyruvate kinase M2 pathway. *Bioengineered* **2022**, *13*, 13906–13918.
551. Zhang, Q.; Liu, Q.; Zheng, S.; Liu, T.; Yang, L.; Han, X.; Lu, X. Shikonin Inhibits Tumor Growth of ESCC by suppressing PKM2 mediated Aerobic Glycolysis and STAT3 Phosphorylation. *J. Cancer* **2021**, *12*, 4830–4840.
552. Sancho, P.; Barneda, D.; Heeschen, C. Hallmarks of cancer stem cell metabolism. *Br. J. Cancer* **2016**, *114*, 1305–1312.
553. Yakisich, J.S.; Azad, N.; Kaushik, V.; Iyer, A.K.V. The Biguanides Metformin and Buformin in Combination with 2-Deoxy-glucose or WZB-117 Inhibit the Viability of Highly Resistant Human Lung Cancer Cells. *Stem Cells Int.* **2019**, *2019*, 6254269.
554. Li, Y.L.; Weng, H.C.; Hsu, J.L.; Lin, S.W.; Guh, J.H.; Hsu, L.C. The Combination of MK-2206 and WZB117 Exerts a Synergistic Cytotoxic Effect Against Breast Cancer Cells. *Front. Pharmacol.* **2019**, *10*, 1311.
555. Liu, W.; Fang, Y.; Wang, X.T.; Liu, J.; Dan, X.; Sun, L.L. Overcoming 5-Fu resistance of colon cells through inhibition of Glut1 by the specific inhibitor WZB117. *Asian Pac. J. Cancer Prev.* **2014**, *15*, 7037–7041.
556. Chen, Q.; Meng, Y.Q.; Xu, X.F.; Gu, J. Blockade of GLUT1 by WZB117 resensitizes breast cancer cells to adriamycin. *Anticancer. Drugs* **2017**, *28*, 880–887.
557. Ancey, P.B.; Contat, C.; Boivin, G.; Sabatino, S.; Pascual, J.; Zangger, N.; Perentes, J.Y.; Peters, S.; Abel, E.D.; Kirsch, D.G.; et al. GLUT1 Expression in Tumor-Associated Neutrophils Promotes Lung Cancer Growth and Resistance to Radiotherapy. *Cancer Res.* **2021**, *81*, 2345–2357.

558. Sun, M.; Zhao, S.; Duan, Y.; Ma, Y.; Wang, Y.; Ji, H.; Zhang, Q. GLUT1 participates in tamoxifen resistance in breast cancer cells through autophagy regulation. *Naunyn Schmiedebergs Arch. Pharmacol.* **2021**, *394*, 205–216.
559. Shima, T.; Taniguchi, K.; Tokumaru, Y.; Inomata, Y.; Arima, J.; Lee, S.W.; Takabe, K.; Yoshida, K.; Uchiyama, K. Glucose transporter-1 inhibition overcomes imatinib resistance in gastrointestinal stromal tumor cells. *Oncol. Rep.* **2022**, *47*.
560. Zeng, Z.; Nian, Q.; Chen, N.; Zhao, M.; Zheng, Q.; Zhang, G.; Zhao, Z.; Chen, Y.; Wang, J.; Zeng, J.; et al. Ginsenoside Rg3 inhibits angiogenesis in gastric precancerous lesions through downregulation of Glut1 and Glut4. *Biomed. Pharmacother.* **2022**, *145*, 112086.
561. Chen, C.; Xia, J.; Ren, H.; Wang, A.; Zhu, Y.; Zhang, R.; Gan, Z.; Wang, J. Effect of the structure of ginsenosides on the in vivo fate of their liposomes. *Asian J. Pharm. Sci.* **2022**, *17*, 219–229.
562. Chen, X.; Zhao, Y.; He, C.; Gao, G.; Li, J.; Qiu, L.; Wang, X.; Gao, Y.; Qi, Y.; Sun, K.; et al. Identification of a novel GLUT1 inhibitor with in vitro and in vivo anti-tumor activity. *Int. J. Biol. Macromol.* **2022**, *216*, 768–778.
563. Nath, K.; Guo, L.; Nancolas, B.; Nelson, D.S.; Shestov, A.A.; Lee, S.C.; Roman, J.; Zhou, R.; Leeper, D.B.; Halestrap, A.P.; et al. Mechanism of antineoplastic activity of lonidamine. *Biochim. Biophys. Acta* **2016**, *1866*, 151–162.
564. Shutkov, I.A.; Okulova, Y.N.; Mazur, D.M.; Melnichuk, N.A.; Babkov, D.A.; Sokolova, E.V.; Spasov, A.A.; Milaeva, E.R.; Nazarov, A.A. New Organometallic Ru(II) Compounds with Lonidamine Motif as Antitumor Agents. *Pharmaceutics* **2023**, *15*, 1366.
565. Muhammad, N.; Tan, C.P.; Nawaz, U.; Wang, J.; Wang, F.X.; Nasreen, S.; Ji, L.N.; Mao, Z.W. Multi-action Platinum(IV) Prodrug Containing Thymidylate Synthase Inhibitor and Metabolic Modifier against Triple-Negative Breast Cancer. *Inorg. Chem.* **2020**, *59*, 12632–12642.
566. Goldin, N.; Arzoine, L.; Heyfets, A.; Israelson, A.; Zaslavsky, Z.; Bravman, T.; Bronner, V.; Notcovich, A.; Shoshan-Barmatz, V.; Flescher, E. Methyl jasmonate binds to and detaches mitochondria-bound hexokinase. *Oncogene* **2008**, *27*, 4636–4643.
567. Fricker, L.D. Proteasome Inhibitor Drugs. *Annu. Rev. Pharmacol. Toxicol.* **2020**, *60*, 457–476.
568. Clem, B.; Telang, S.; Clem, A.; Yalcin, A.; Meier, J.; Simmons, A.; Rasku, M.A.; Arumugam, S.; Dean, W.L.; Eaton, J.; et al. Small-molecule inhibition of 6-phosphofructo-2-kinase activity suppresses glycolytic flux and tumor growth. *Mol. Cancer Ther.* **2008**, *7*, 110–120.
569. Boyd, S.; Brookfield, J.L.; Critchlow, S.E.; Cumming, I.A.; Curtis, N.J.; Debreczeni, J.; Degorce, S.L.; Donald, C.; Evans, N.J.; Groombridge, S.; et al. Structure-Based Design of Potent and Selective Inhibitors of the Metabolic Kinase PFKFB3. *J. Med. Chem.* **2015**, *58*, 3611–3625.
570. Emimi Veseli, B.; Perrotta, P.; Van Wielendaele, P.; Lambeir, A.M.; Abdali, A.; Bellosta, S.; Monaco, G.; Bultynck, G.; Martinet, W.; De Meyer, G.R.Y. Small molecule 3PO inhibits glycolysis but does not bind to 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase-3 (PFKFB3). *FEBS Lett.* **2020**, *594*, 3067–3075.
571. Emimi Veseli, B.; Van Wielendaele, P.; Delibegovic, M.; Martinet, W.; De Meyer, G.R.Y. The PFKFB3 Inhibitor AZ67 Inhibits Angiogenesis Independently of Glycolysis Inhibition. *Int. J. Mol. Sci.* **2021**, *22*, 5970.
572. Mashouri, L.; Yousefi, H.; Aref, A.R.; Ahadi, A.M.; Molaei, F.; Alahari, S.K. Exosomes: composition, biogenesis, and mechanisms in cancer metastasis and drug resistance. *Mol. Cancer* **2019**, *18*, 75.
573. Wilkins, H.M.; Mahnken, J.D.; Welch, P.; Bothwell, R.; Koppel, S.; Jackson, R.L.; Burns, J.M.; Swerdlow, R.H. A Mitochondrial Biomarker-Based Study of S-Equol in Alzheimer's Disease Subjects: Results of a Single-Arm, Pilot Trial. *J. Alzheimers Dis.* **2017**, *59*, 291–300.
574. Tsai, M.C.; Lin, S.H.; Hidayah, K.; Lin, C.I. Equol Pretreatment Protection of SH-SY5Y Cells against Abeta (25-35)-Induced Cytotoxicity and Cell-Cycle Reentry via Sustaining Estrogen Receptor Alpha Expression. *Nutrients* **2019**, *11*, 2356.
575. Xu, X.; Sun, Y.; Cen, X.; Shan, B.; Zhao, Q.; Xie, T.; Wang, Z.; Hou, T.; Xue, Y.; Zhang, M.; et al. Metformin activates chaperone-mediated autophagy and improves disease pathologies in an Alzheimer disease mouse model. *Protein Cell* **2021**, *12*, 769–787.
576. Zhang, Q.Q.; Li, W.S.; Liu, Z.; Zhang, H.L.; Ba, Y.G.; Zhang, R.X. Metformin therapy and cognitive dysfunction in patients with type 2 diabetes: A meta-analysis and systematic review. *Medicine* **2020**, *99*, e19378.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.