



Article

DHEA Induces Sex-Associated Differential Patterns in Cytokine and Antibody Levels in Mice Infected with *Plasmodium berghei* ANKA

Fidel Orlando Buendía-González ^{1,2}, Luis Antonio Cervantes-Candelas ¹, Jesús Aguilar-Castro ^{1,2}, Omar Fernández-Rivera ^{1,2}, Teresita de Jesús Nolasco-Pérez ^{1,2}, Monserrat Sofía López-Padilla ^{1,2}, David Roberto Chavira-Ramírez ³, Armando Cervantes-Sandoval ⁴ and Martha Legorreta-Herrera ^{1,*}

- ¹ Laboratorio de Inmunología Molecular, Unidad de Investigación Química Computacional, Síntesis y Farmacología de Moléculas de Interés Biológico, Facultad de Estudios Superiores Zaragoza, Universidad Nacional Autónoma de México, Ciudad de México 09230, Mexico; fidelbuendia2@gmail.com (F.O.B.-G.); cervantescandelasluis@gmail.com (L.A.C.-C.); jesus_aguilar_castro@yahoo.com.mx (J.A.-C.); qbfbdz@gmail.com (O.F.-R.); teresyta.qfb@gmail.com (T.J.N.-P.); sofia.lpz100@gmail.com (M.S.L.-P.)
 - ² Posgrado en Ciencias Biológicas, Unidad de Posgrado, Circuito de Posgrados, Ciudad Universitaria, Universidad Nacional Autónoma de México, Ciudad de México 04510, Mexico
 - ³ Departamento de Biología de la Reproducción, Instituto Nacional de Ciencias Médicas y Nutrición Salvador Zubirán, Ciudad de México 14080, Mexico; roberto.chavira@incmnsz.mx
 - ⁴ Laboratorio de Aplicaciones Computacionales, Facultad de Estudios Superiores Zaragoza, Universidad Nacional Autónoma de México, Ciudad de México 09230, Mexico; arpacer@unam.mx
- * Correspondence: marthal@unam.mx; Tel.: +55-56230700 (ext. 83207)



Citation: Buendía-González, F.O.; Cervantes-Candelas, L.A.; Aguilar-Castro, J.; Fernández-Rivera, O.; Nolasco-Pérez, T.J.; López-Padilla, M.S.; Chavira-Ramírez, D.R.; Cervantes-Sandoval, A.; Legorreta-Herrera, M. DHEA Induces Sex-Associated Differential Patterns in Cytokine and Antibody Levels in Mice Infected with *Plasmodium berghei* ANKA. *Int. J. Mol. Sci.* **2023**, *24*, 12549. <https://doi.org/10.3390/ijms241612549>

Academic Editors: Jean-Paul Coutelier and Sylviane Pied

Received: 1 July 2023

Revised: 31 July 2023

Accepted: 2 August 2023

Published: 8 August 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Malaria is the most lethal parasitic disease worldwide; the severity of symptoms and mortality are higher in men than in women, exhibiting an evident sexual dimorphism in the immune response; therefore, the contribution of 17 β -estradiol and testosterone to this phenomenon has been studied. Both hormones differentially affect several aspects of innate and adaptive immunity. Dehydroepiandrosterone (DHEA) is the precursor of both hormones and is the sexual steroid in higher concentrations in humans, with immunomodulatory properties in different parasitic diseases; however, the involvement of DHEA in this sexual dimorphism has not been studied. In the case of malaria, the only information is that higher levels of DHEA are associated with reduced *Plasmodium falciparum* parasitemia. Therefore, this work aims to analyze the DHEA contribution to the sexual dimorphism of the immune response in malaria. We assessed the effect of modifying the concentration of DHEA on parasitemia, the number of immune cells in the spleen, cytokines, and antibody levels in plasma of CBA/Ca mice infected with *Plasmodium berghei* ANKA (*P. berghei* ANKA). DHEA differentially affected the immune response in males and females: it decreased IFN- γ , IL-2 and IL-4 concentrations only in females, whereas in gonadectomized males, it increased IgG2a and IgG3 antibodies. The results presented here show that DHEA modulates the immune response against *Plasmodium* differently in each sex, which helps to explain the sexual dimorphism present in malaria.

Keywords: DHEA; *Plasmodium*; gonadectomy; sexual dimorphism; IL-2; IL-4; IFN- γ ; IgG

1. Introduction

Malaria is the deadliest parasitic disease; in 2021, it caused 627,000 deaths [1] and is a clear example of sexual dimorphism in which males develop higher mortality than females [2,3]. In general, women exhibit more robust innate and adaptive immune responses against infectious agents and eliminate *Plasmodium* more efficiently than men. Although this sexual condition is accepted, few articles have analyzed this phenomenon in both sexes [4]. 17 β -estradiol and testosterone are involved in the sexual dimorphism that occurs in the immune response in malaria [5–7]. Both hormones modify survival

and cellular and humoral immune responses in *Plasmodium* infections [2,8,9]. DHEA is the precursor hormone of testosterone and 17 β -estradiol and has important immunopotentiating properties [10]. DHEA modulates immune response, cell activity and cytokine concentration [11], and this hormone increases macrophage and natural killer (NK) cell activity [12,13] and promotes the maturation of CD4⁺ T cells and their secretion of interferon (IFN) [14]. Furthermore, DHEA boosts the immune response in the bacterial [15], viral and parasitic diseases [16]. In parasitic diseases, DHEA has been shown to promote the immune response in patients infected with *Trypanosoma cruzi*, decreasing parasitemia and TNF- α levels [17]. Furthermore, increased DHEA concentration has been associated with reduced infection by *Schistosoma japonicum* and *Schistosoma mansoni* [18,19]. Additionally, DHEA promotes the Th1-type response and consequently promotes recovery in patients with *Leishmania mexicana* [20]. In the *Plasmodium berghei* infection, the DHEA analog (16 α -bromoepiandrosterone) decreases parasitemia in vitro and in vivo [21]. These findings show that DHEA modulates the immune response in parasitic diseases, and all of them exhibit sexual dimorphism [22]. Unfortunately, DHEA participation in the immune response against *Plasmodium*-infected individuals of both sexes has not yet been studied. Sexual dimorphism is a highly complex phenomenon involving sex hormones, genetic differences and even behavior between the sexes [4]. Due to the immunomodulatory properties of DHEA, it is likely that DHEA also contributes to sexual dimorphism in the immune response to *Plasmodium* because together with its sulfated form (DHEA-S), DHEA constitutes the steroid hormone at the highest concentration [23]. Moreover, DHEA positively regulates the immune response [24]. Particularly in malaria, it has been associated with lower parasitemia in *Plasmodium falciparum*-infected individuals and high hemoglobin concentrations [25,26]. However, the mechanisms that explain these findings are unknown. The purpose of this work is to analyze whether DHEA is involved in the sexual dimorphism of the immune response in malaria. To this end, we modified the concentration of this hormone in vivo in male and female mice that were infected with *P. berghei* ANKA; we evaluated the impact on parasitemia, immune cells in the spleen (CD3⁺, CD4⁺, CD8⁺, CD19⁺, macrophages and NK cells), pro- and anti-inflammatory cytokines and IgG and IgM levels.

To our knowledge, this work is the first report documenting the dimorphic participation of DHEA in the immune response to malaria and contributes to a better understanding of the differences between sexes that could support the future development of sex-specific, more efficient, anti-malarial therapies.

2. Results

2.1. DHEA Administration Dramatically Increased DHEA Levels in Mice Infected with *P. berghei* ANKA

We used a previously calibrated dose that increased DHEA concentration in vivo in mice [27]. All DHEA-treated groups showed markedly increased DHEA concentrations regardless of whether they were infected or gonadectomized (Figure 1A,B); furthermore, we detected that infection generated a dimorphic pattern by decreasing DHEA concentrations in intact females compared to males under the same conditions (Figure 1B).

2.2. Effects of Modifying DHEA Concentration on Temperature and Hemoglobin Concentration in Mice Infected with *P. berghei* ANKA

Quantifying weight loss, hypothermia and decreased hemoglobin levels provides insight into disease severity in CBA/Ca mice infected with *P. berghei* ANKA [6]. We did not detect differences depending on DHEA concentration on body weight; we also did not detect a dimorphic pattern in temperature or hemoglobin concentration (Supplementary Figures S1 and S2).

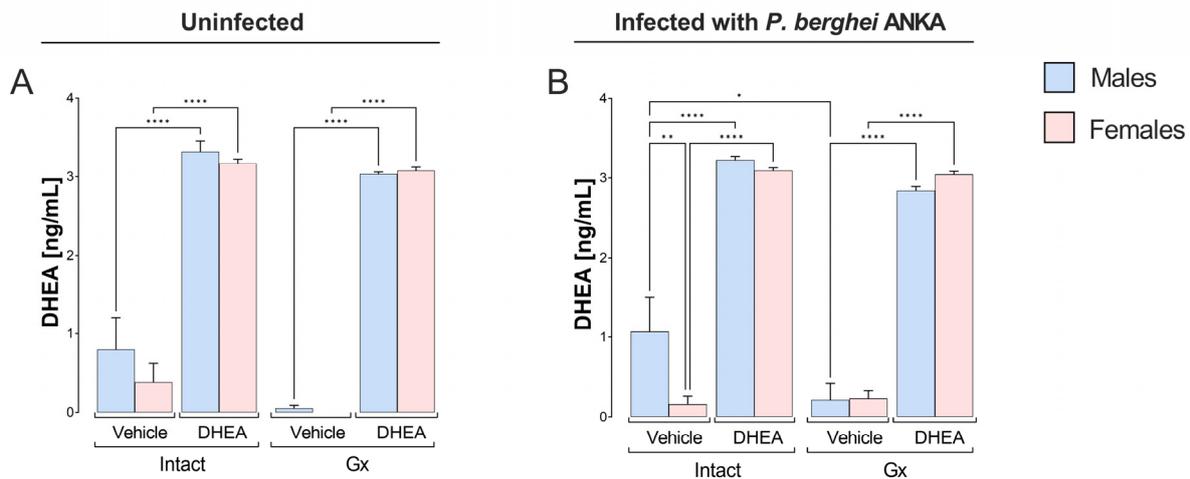


Figure 1. Effect of modifying DHEA levels on plasma steroid concentrations in male and female mice infected with *P. berghei* ANKA. Eight groups of male mice and eight groups of female mice were used; four groups of each sex were gonadectomized (Gx), and the other four groups remained intact as controls. Four weeks after surgery, two Gx and 2 intact groups were treated with DHEA for 5 days. The day after the last administration, one group of Gx and one of the intact mice of each sex were infected with *P. berghei* ANKA, and the remaining groups were injected with PBS as infection control groups. On day 8 post infection or on the equivalent day in uninfected groups, the mice were sacrificed, heart blood was extracted, and plasma was separated to determine the plasma concentration of DHEA. Uninfected groups are represented in (A) and infected groups in (B). The histogram represents the mean of each group \pm SEM mean standard error ($n = 10$). Lines above the histogram represent significant differences between groups. Asterisks (*) indicate statistical significance between 2 groups * ($p \leq 0.05$), ** ($p \leq 0.01$), and **** ($p \leq 0.0001$). The significance between groups was calculated with two-way ANOVA with a Bonferroni post hoc test. The whole experiment was performed twice.

2.3. DHEA Administration to Intact Male Mice Increased Parasitemia

Intact females exhibited higher parasitemia than intact males, particularly on days 5 and 6 post infection, inducing a dimorphic pattern (Figure 2A). In contrast to our expectations, DHEA administration to intact mice eliminated the differences between sexes by increasing parasitemia only in intact males (Figure 2B,E). Additionally, gonadectomy did not modify parasitemia in either sex, maintaining the dimorphic pattern (Figure 2C,E). Furthermore, the reconstitution of Gx mice with DHEA increased parasitemia in both sexes on day 6 postinfection (Figure 2D); in addition, the reconstitution of Gx mice with DHEA prior to infection eliminated the dimorphic pattern (Figure 2E). Finally, increasing DHEA administration to 40 mg/kg did not modify parasitemia (Figure S3).

2.4. Reducing the DHEA Concentration Generated a Dimorphic Pattern in the CD3⁺, CD19⁺ and NK Cell Populations of Uninfected Mice

Because the spleen is the central organ where the immune system eliminates *Plasmodium*, infection with this parasite induces significant changes in size, architecture and cellular composition [28]. In this work, we evaluated whether modifying DHEA levels affects the central cell populations of the immune response in the spleen. No differences were detected in the CD3⁺ population between uninfected intact males and females; gonadectomy generated a dimorphic pattern in which females had a higher number of CD3⁺ cells than males in the absence of infection. Additionally, the reconstitution of Gx mice with DHEA decreased the number of CD3⁺ cells, which eliminated the dimorphic pattern (Figure 3A). Interestingly, infection decreased this population in all intact groups, and gonadectomy reduced the number of CD3⁺ cells in the infected groups (Figure 3A). Regarding the CD4⁺ cells, gonadectomy reduced this population in uninfected females; however, reconstituting

Gx mice with DHEA did not restore their CD4⁺ population, and no changes were detected in infected mice (Figure 3B). Concerning CD8⁺ cells, we detected that intact uninfected females exhibited higher numbers of CD8⁺ cells than males under the same conditions, generating a sex-associated pattern, and DHEA administration further increased this difference. In addition, gonadectomy decreased this population, which eliminated the dimorphic pattern. Moreover, infection completely depleted this population only in the intact groups; in contrast, all Gx and infected groups dramatically increased CD8⁺ numbers regardless of DHEA concentration (Figure 3C).

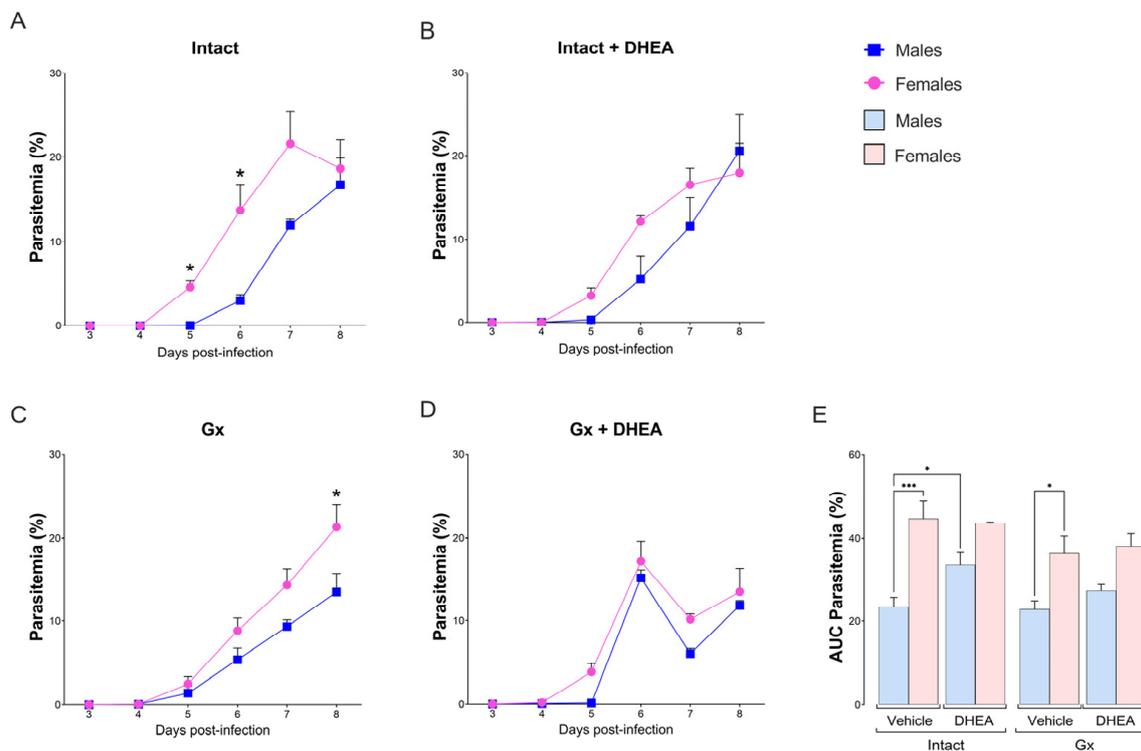


Figure 2. Effect of DHEA on parasitemia in male and female mice infected with *P. berghei* ANKA. Four groups of male mice and four groups of female mice were used; two groups of each sex were gonadectomized (Gx), and the other two groups remained intact as controls. Four weeks after surgery, two Gx and 2 intact groups were treated with DHEA for 5 days. The day after the last administration, one group of Gx and one intact group of each sex were infected with *P. berghei* ANKA (day zero), and the remaining groups were injected with PBS as infection control groups. Parasitemia was assessed daily from day 3 post infection. Parasitemia of intact male and female mice are shown in (A); parasitemia of DHEA-treated mice in (B); parasitemia of Gx mice in (C); parasitemia of Gx DHEA-treated mice in (D); and the area under the curve (AUC) of the above groups is shown in (E). Graphs represent the mean of each group \pm SEM ($n = 10$). Asterisk (*) indicates statistical significance between 2 groups ($p \leq 0.05$) and *** ($p \leq 0.001$) calculated with two-way ANOVA and Bonferroni post hoc test. The whole experiment was performed twice.

Regarding the macrophage (CD107b⁺) population, uninfected intact females had a higher number of this population than males under the same conditions, exhibiting a dimorphic pattern. When DHEA was administered to uninfected mice, the dimorphic pattern was maintained; in addition, gonadectomy decreased this cell population only in females, eliminating the dimorphic pattern (Figure 4A). Furthermore, infection increased macrophage numbers in intact vehicle-treated males and DHEA-treated Gx females (Figure 4A).

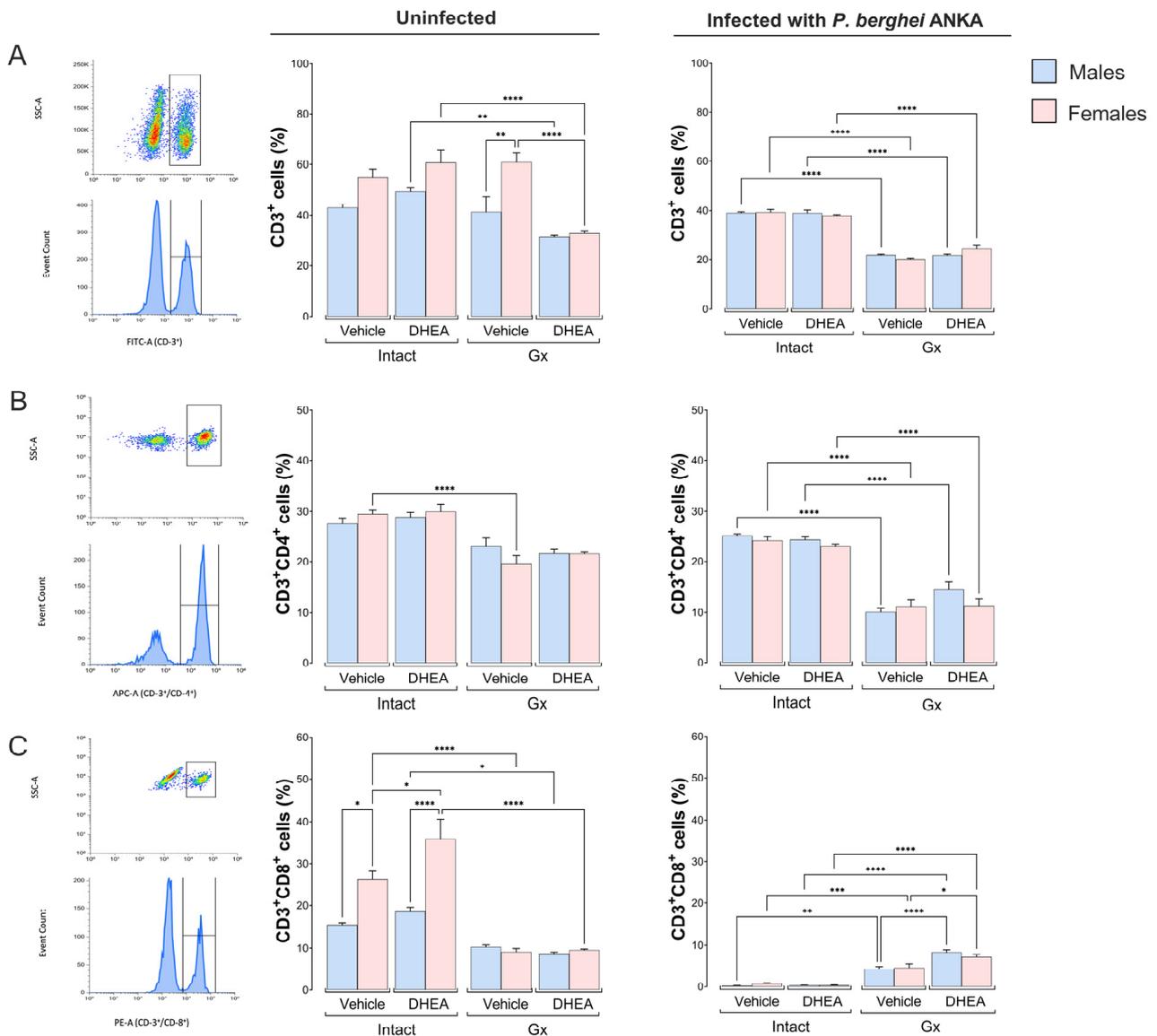


Figure 3. Effect of modifying DHEA levels on CD3⁺, CD3⁺/CD4⁺ and CD3⁺/CD8⁺ cell populations of male and female mice, uninfected and infected with *P. berghei* ANKA. Eight groups of male mice and eight groups of female mice were used; four groups of each sex were gonadectomized (Gx), and the other four groups remained intact. Four weeks after surgery, two Gx and 2 intact groups were administered DHEA for 5 days. The day after the last administration, one group of Gx and one group of intact mice of each sex were infected with *P. berghei* ANKA, and the remaining groups were injected with PBS as infection control groups. On day 8 postinfection, mice were sacrificed, spleens were removed and immune response cells were evaluated by flow cytometry. In the dot plot, the rectangle frames 100% of the cells positive for CD3⁺, CD4⁺ or CD8⁺ markers. The bar graphs represent the average number of cells in each group ± SEM. CD3⁺ lymphocytes are represented in (A), helper T cells (CD4⁺) in (B), cytotoxic T cells (CD8⁺) in (C). Lines above histograms represent significant differences between groups ($n = 10$). Asterisks (*) indicate statistical significance between groups * ($p \leq 0.05$), ** ($p \leq 0.01$), *** ($p \leq 0.001$) and **** ($p \leq 0.0001$). The significance between groups was calculated with two-way ANOVA with a Bonferroni post hoc test. The whole experiment was performed twice.

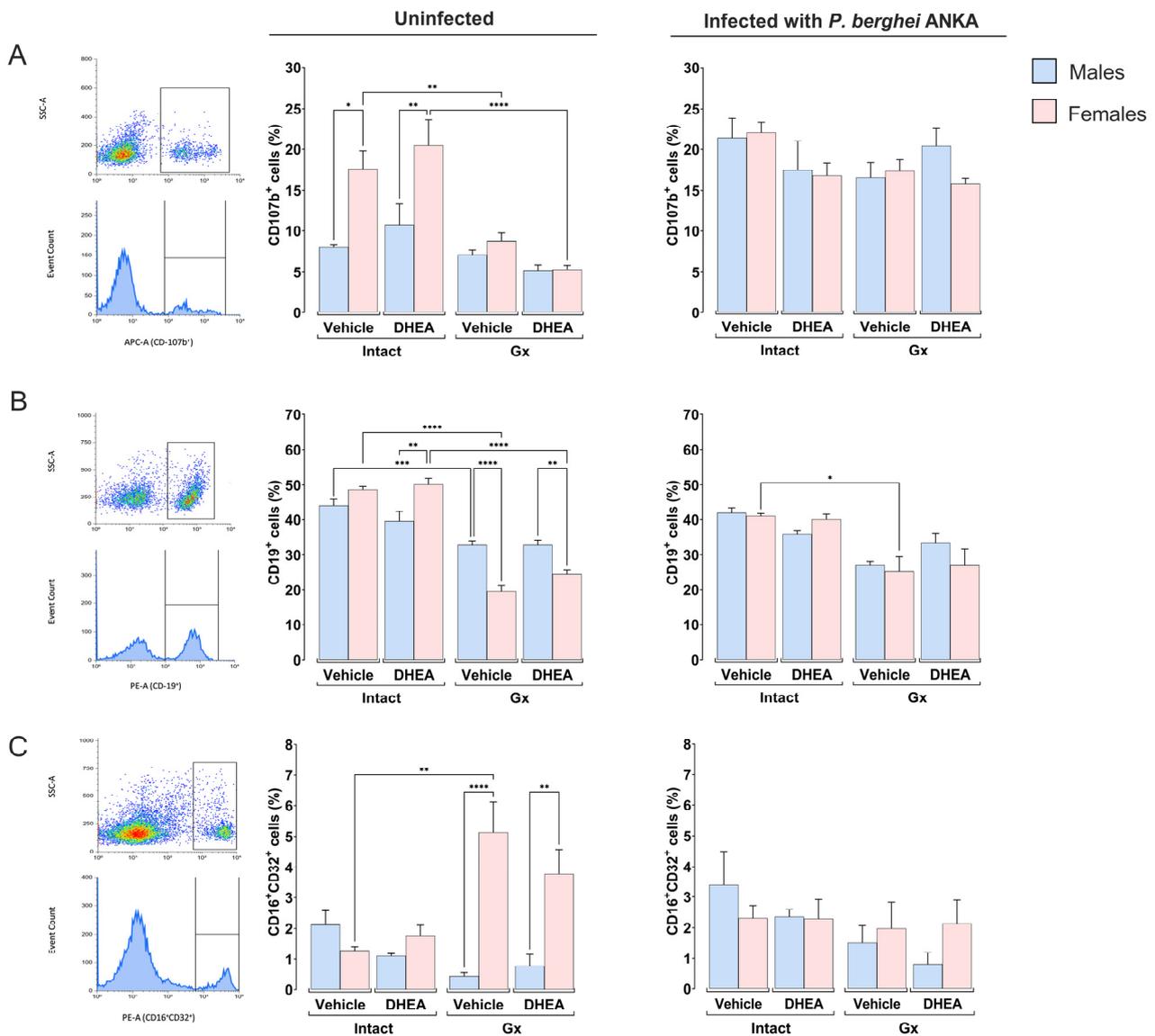


Figure 4. Effect of modifying DHEA levels on macrophage (CD107b⁺), B-cell (CD19⁺) and NK cell (CD3⁻CD16⁺/CD32⁺) populations of male and female mice, uninfected and infected with *P. berghei* ANKA. Eight groups of male mice and eight groups of female mice were used; four groups of each sex were gonadectomized (Gx), and the other four groups remained intact. Four weeks after surgery, two Gx and 2 intact groups were administered DHEA for 5 days. The day after the last administration, one group of Gx and one group of intact mice of each sex were infected with *P. berghei* ANKA, and the remaining groups were injected with PBS as infection control groups. On day 8 post infection, mice were sacrificed, spleens were removed and immune response cells were evaluated by flow cytometry. In the dot plot, the rectangle frames 100% of the cells positive for CD107b⁺ or CD19⁺ or CD16⁺/CD32⁺ markers. The bar graphs represent the average number of cells in each group ± SEM. Macrophages (CD107b⁺) are represented in (A), B cells (CD19⁺) in (B), and NK cells (CD3⁻/CD19⁻/CD16⁺/CD32⁺) in (C). Lines above histograms represent significant differences between groups ($n = 10$). Asterisks (*) indicate statistical significance between 2 groups ($p \leq 0.05$), ** ($p \leq 0.01$), *** ($p \leq 0.001$) and **** ($p \leq 0.0001$). The significance between groups was calculated with two-way ANOVA with a Bonferroni post hoc test. The whole experiment was performed twice.

Regarding B cells (CD19⁺), DHEA administration to intact uninfected mice generated a dimorphic pattern; females exhibited higher numbers of CD19⁺ cells than males under the same conditions; additionally, gonadectomy considerably decreased this population,

particularly in females, which also generated a dimorphic pattern that did not change after DHEA reconstitution (Figure 4B). Gonadectomy decreased the number of CD19⁺ cells exclusively in infected females (Figure 4B).

Finally, gonadectomy increased the NK population only in uninfected females, which induced a dimorphic pattern that did not change with DHEA administration. In infected mice, changing the DHEA concentration did not affect this population (Figure 4C).

2.5. DHEA Generated a Dimorphic Pattern in the Concentrations of IFN- γ , IL-2 and IL-4

Given that immune response cells produce cytokines and malaria severity is associated with elevated concentrations of TNF- α , IL-6 and IL-10 [29,30]. Additionally, DHEA decreases IFN- γ synthesis and increases IL-10 levels in vitro [31]. Therefore, we assessed the concentrations of pro- and anti-inflammatory cytokines. No differences in IFN- γ concentration were detected between intact male and female mice or Gx mice treated with the vehicle without infection; furthermore, DHEA administration increased IFN- γ concentration only in uninfected females (Figure 5A). In general, infection increased the IFN- γ concentration in intact animals treated with vehicle or DHEA. Administering DHEA decreased the IFN- γ concentration only in intact infected females, which induced a dimorphic pattern (Figure 5A). Regarding TNF- α , in general, males exhibited higher levels than females, although the increase was not significant; the infection increased the TNF- α concentration in the intact groups; however, DHEA administration did not significantly modify the concentration of this cytokine (Figure 5B). Concerning IL-2, gonadectomy in uninfected mice resulted in a dimorphic pattern, with males having higher IL-2 concentrations than females under the same conditions. Reconstituting Gx mice with DHEA dramatically decreased the IL-2 concentration in uninfected Gx males, eliminating the dimorphic pattern. In infected mice, gonadectomy increased the IL-2 concentration only in vehicle-treated females, and reconstitution of Gx mice with DHEA significantly decreased the IL-2 concentration exclusively in females, resulting in a dimorphic pattern (Figure 5C).

Regarding IL-10, modifying the concentration of DHEA did not change the concentration of this cytokine in uninfected mice. In addition, infection increased the concentration of IL-10 in intact females, generating a dimorphic pattern in which females exhibited a higher concentration of IL-10 than males. DHEA administration eliminated this dimorphic pattern. Furthermore, gonadectomy decreased the concentration of IL-10 only in infected females, and reconstitution of Gx females with DHEA did not restore the concentration of this cytokine in infected females (Figure 5D).

Related to IL-4, DHEA-reconstituted Gx mice in the absence of infection had decreased IL-4 concentrations, whereas, in infected mice, DHEA-reconstituted male Gx mice had higher IL-4 concentrations than females under the same conditions, resulting in a sex-dependent pattern (Figure 6A). Concerning IL-6, the reconstitution of Gx mice with DHEA without infection increased IL-6 concentrations in both sexes. However, neither gonadectomy nor infection changed the IL-6 concentration (Figure 6B). Finally, infection or DHEA administration did not change the IL-17 concentration (Figure 6C).

2.6. Reconstitution of Gx Males with DHEA Increased the Concentration of the IgG Subclass

Antibodies are required to eliminate the *Plasmodium* parasite [32]; in addition, DHEA has been reported to regulate antibody synthesis in mice [33]; thus, we tested whether modifying DHEA concentration affects antibody levels. DHEA administration did not significantly modify the IgM concentration or total IgG in the intact or Gx mice (Figure 7A,B). Nevertheless, when we evaluated IgG subclasses, we detected that the reconstitution of gonadectomized mice with DHEA increased the levels of IgG1, IgG2a, IgG2b and IgG3 exclusively in males (Figure 7C–F). To determine whether the immune response was polarized to a Th1 or Th2 type, the IgG1/IgG2a ratio was assessed. As the ratio in all groups was lower than one, DHEA promoted a Th1 response (Figure 7G).

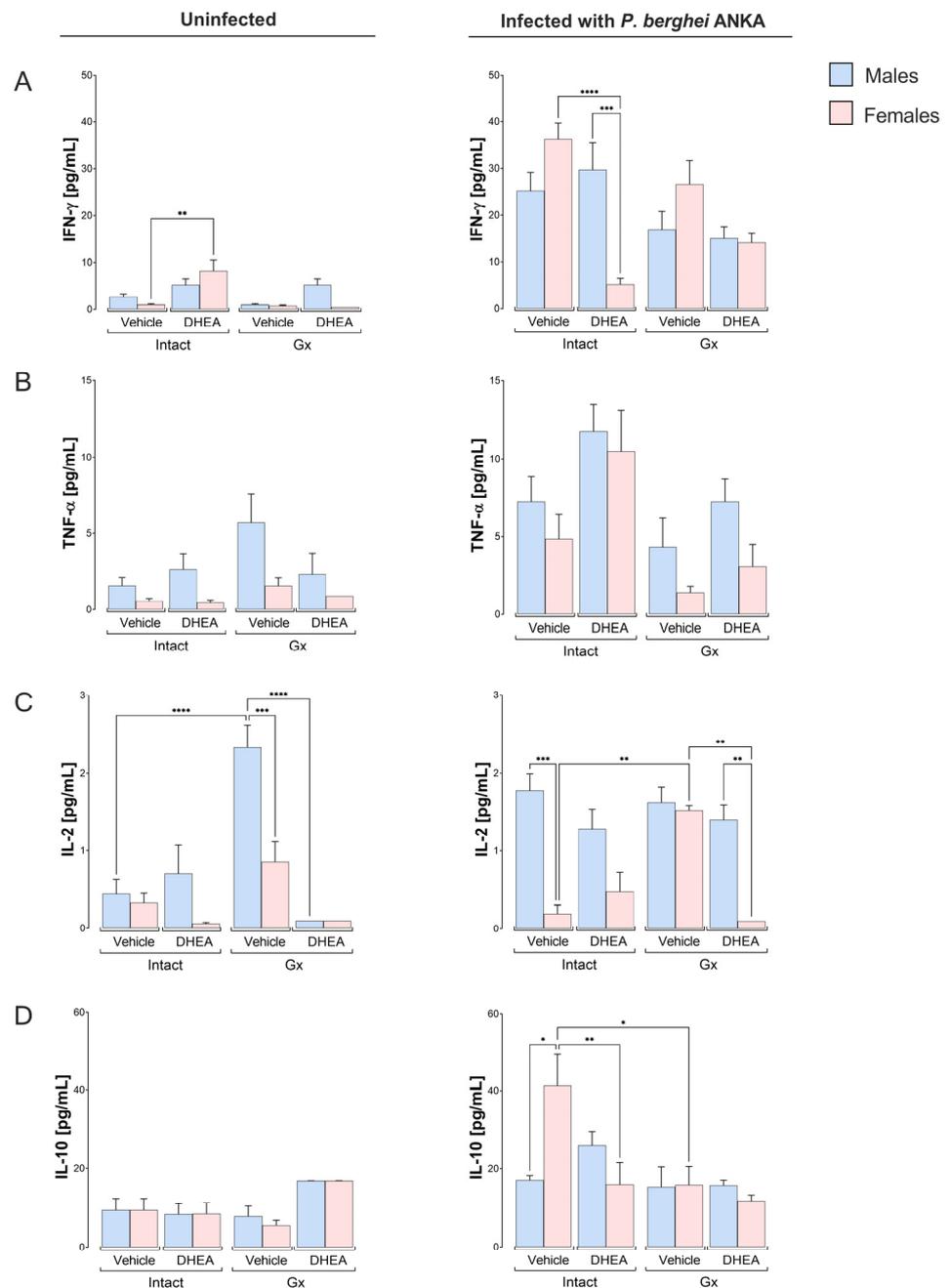


Figure 5. Effect of modifying DHEA levels on the plasma concentration of cytokines: IFN- γ , TNF- α , IL-2 and IL-10 in uninfected and infected male and female mice with *P. berghei* ANKA. Eight groups of male mice and eight groups of female mice were used; four groups of each sex were gonadectomized (Gx), and the other four groups remained intact. Four weeks after surgery, two Gx and two intact groups were reconstituted with DHEA for 5 days. On the day following the last administration, one group of Gx and one of the intact mice of each sex were infected with *P. berghei* ANKA, and the remaining groups were injected with PBS as infection control groups. On day 8 post infection, mice were sacrificed and plasma was obtained to quantify the levels of the cytokines IFN- γ (A), TNF- α (B), IL-2 (C) and IL-10 (D). Determination was performed by flow cytometry. Histograms represent the mean of each group \pm SEM ($n = 10$). The lines above histograms represent significant differences between groups. Asterisks (*) indicate statistical significance between 2 groups * ($p \leq 0.05$), ** ($p \leq 0.01$), *** ($p \leq 0.001$) and **** ($p \leq 0.0001$). The significance between groups was calculated with two-way ANOVA with a Bonferroni post hoc test. The whole experiment was performed twice.

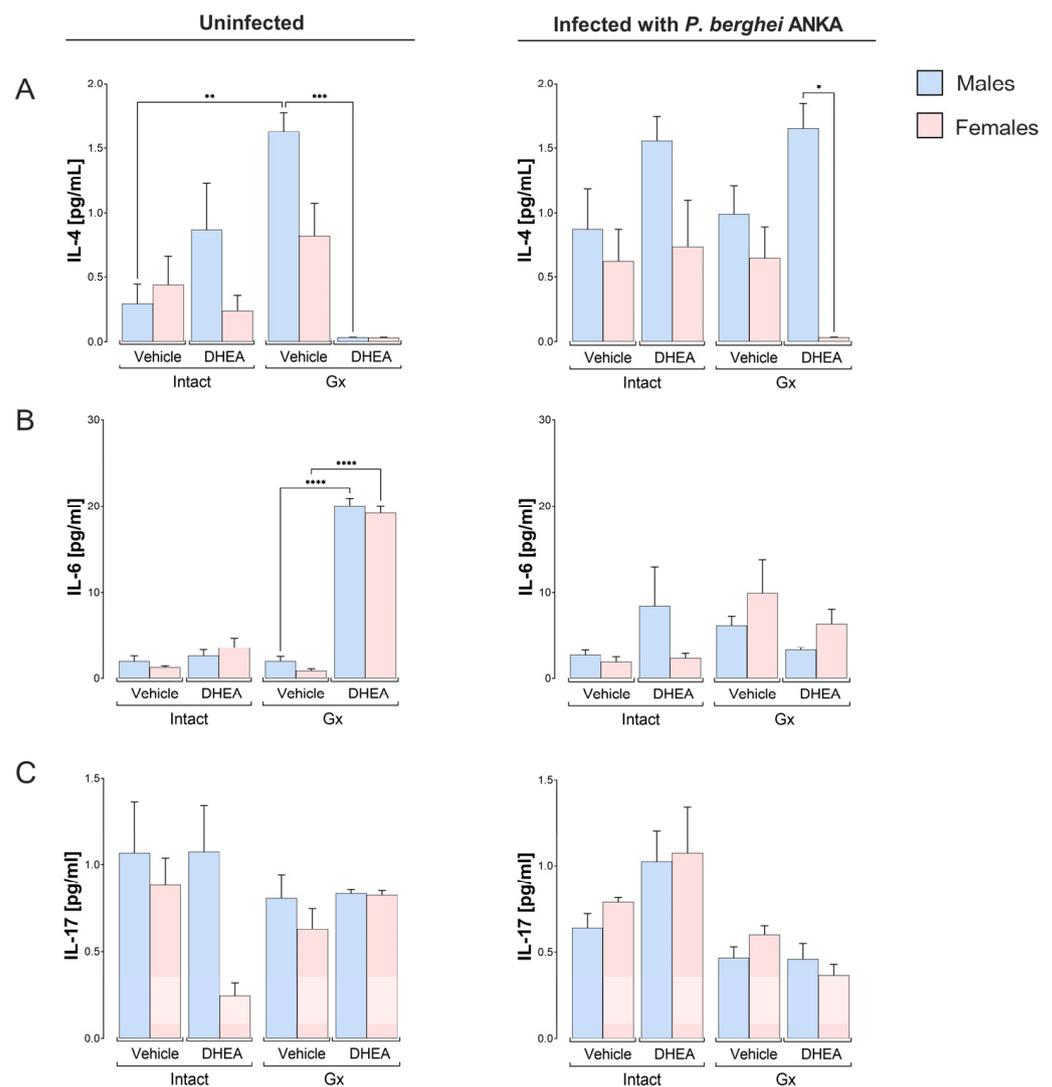


Figure 6. Effect of modifying DHEA levels on the plasma concentration of the cytokines IL-4, IL-6 and IL-17 in uninfected and infected male and female mice with *P. berghei* ANKA. Eight groups of male mice and eight groups of female mice were used; four groups of each sex were gonadectomized (Gx), and the other four groups remained intact. Four weeks after surgery, two Gx and two intact groups were reconstituted with DHEA for 5 days. On the day following the last administration, one group of Gx and one of the intact mice of each sex were infected with *P. berghei* ANKA, and the remaining groups were injected with PBS as infection control groups. On day 8 post infection, mice were sacrificed, and plasma was obtained to quantify the levels of the cytokines IL-4 (A), IL-6 (B) and IL-17 (C). Determination was performed by flow cytometry. Histograms represent the mean of each group \pm SEM ($n = 10$). Lines above histograms represent significant differences between groups. Asterisks (*) indicate statistical significance between 2 groups * ($p \leq 0.05$), ** ($p \leq 0.01$), *** ($p \leq 0.001$) and **** ($p \leq 0.0001$). The significance between groups was calculated with two-way ANOVA with a Bonferroni post hoc test. The whole experiment was performed twice.

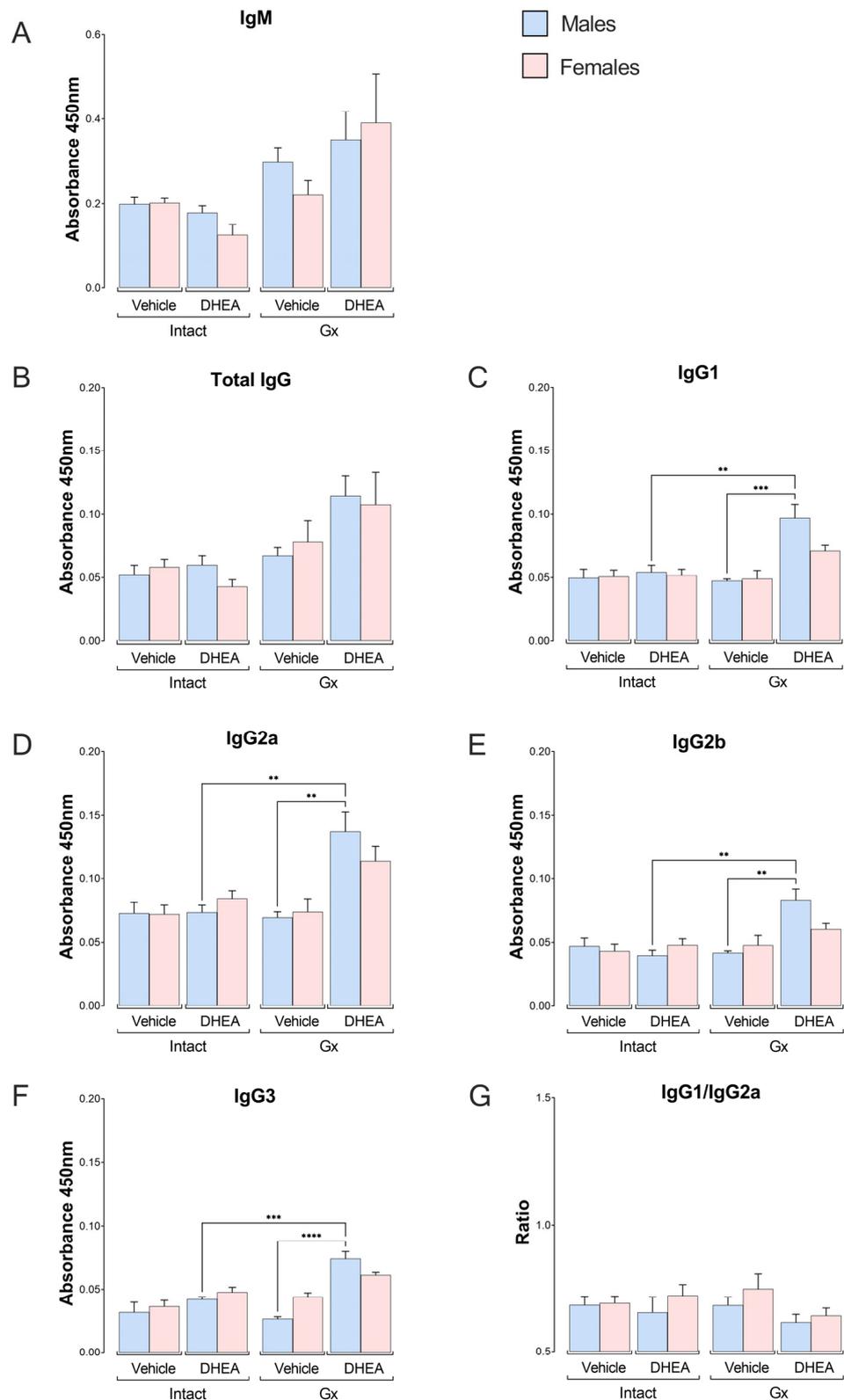


Figure 7. Effect of modifying DHEA levels on plasma concentration of total IgM and IgG, IgG1, IgG2a, IgG2b and IgG3 antibodies of male and female mice infected with *P. berghei* ANKA. Eight groups of male mice and eight groups of female mice were used; four groups of each sex were gonadectomized (Gx), and the other four groups remained intact. Four weeks after surgery, two Gx and 2 intact groups were treated with DHEA for 5 days. The day after the last administration, one group of Gx and one of the intact mice of each sex were infected with *P. berghei* ANKA, and the

remaining groups were injected with PBS as infection control groups. On day 8 post infection, all mice were sacrificed, and plasma was separated to determine the levels of antibodies: IgM (A), total IgG (B), IgG1 (C), IgG2a (D), IgG2b (E), and IgG3 (F). Additionally, the IgG1/IgG2a ratio (G) was calculated. Histograms represent the mean of each group \pm SEM, ($n = 10$). Lines above histograms represent significant differences between groups. Asterisks (*) indicate statistical significance between groups ** ($p \leq 0.01$), *** ($p \leq 0.001$) and **** ($p \leq 0.0001$). The significance between groups was calculated with two-way ANOVA and a Bonferroni post hoc test. The whole experiment was performed twice.

3. Discussion

To evaluate the participation of DHEA in the sexual dimorphism that occurs in the immune response against *P. berghei* ANKA, we modified the concentration of DHEA in male and female mice. We decreased its levels by gonadectomy or increased it by exogenous administration. After treatments, mice were infected with *P. berghei* ANKA. We found that gonadectomy, infection and DHEA administration generated sex-associated patterns.

We first demonstrated that the dose of DHEA we administered increased its concentration in plasma. This effect is consistent with that described by Medina et al., who reported that subcutaneous DHEA administration increases the concentration of this steroid up to 24 times on day 7 post treatment [34]. However, in our work, gonadectomy only decreased DHEA concentration by a trend; a likely explanation for this finding is that since gonadectomy was performed four weeks prior to infection, the mouse censured its deficiency and was able to synthesize DHEA at extragonadal sites [35]. In addition, we found that DHEA did not change body weight, temperature, or hemoglobin concentration. However, gonadectomy decreased the temperature in the uninfected groups in both sexes. It is possible that this surgery, by decreasing the concentration of 17 β -estradiol [7], deregulated thermogenesis in adipose tissue [36]. In addition, DHEA increases mitochondrial FADH₂ synthesis, resulting in heat generation [37]. In a further finding, gonadectomy increased hemoglobin concentration, which is an unexpected result because high levels of DHEA are associated with increased hemoglobin concentration [26]. This increase is likely to be a consequence of the reduction in 17 β -estradiol concentration, as it negatively regulates the erythropoietin synthesis [38]. In addition, gonadectomy in male rats has been reported to increase erythropoietin levels [39]. We also analyzed the effect of modifying DHEA concentration on parasitemia, detecting that intact and Gx-infected females treated with vehicle had higher parasitemia than males; interestingly, this dimorphic pattern was eliminated when DHEA was administered, as it increased parasitemia in intact males without affecting female mice; then, we tried a higher dose (40 mg/kg), but the result was the same (Supplementary Figure S3). To our knowledge, ours is the first report that DHEA increases parasite load. In contrast, Freilich et al. reported that the DHEA analog 16 α -bromoepiandrosterone decreases parasitemia in *P. berghei* ANKA-infected rats [21], and it is likely that this difference is due to DHEA presenting higher affinity for the androgen receptor than 16 α -bromoepiandrosterone [40]. Our result also contrasts with that described by dos Santos et al. in *Trypanosoma cruzi* infection [41]. A probable explanation is the parasite used, as we experimented with *P. berghei* ANKA in an in vivo study. In addition, it is likely that these discrepancies are because immune response cells have different numbers of receptors for DHEA in males than in females [42], which also vary depending on whether the mice are intact or gonadectomized [43].

Because the spleen is the leading site of *Plasmodium* destruction, we analyzed the effect of modifying DHEA levels on the central cells of the immune response in the spleen. We found that uninfected Gx females exhibited higher numbers of CD3⁺ cells than males under the same conditions; this result corresponds with the dimorphic pattern found in IL-2 concentration in this group. Most likely, IL-2, in addition to promoting CD3⁺ cell proliferation, it also regulates the process of apoptosis mediated by FASL expression in T cells [44]. This explanation is supported by the disappearance of the dimorphic pattern in T cells when DHEA was administered to Gx mice. In contrast, Cao et al. described that administering DHEA increases both T-lymphocyte proliferation and IL-2 concentra-

tion [13]. This difference could be explained because the Cao group administered a dose of 14.42 µg/kg DHEA orally, while we used 8 mg/kg DHEA subcutaneously. Furthermore, infection decreased the number of CD3⁺ cells in intact vehicle- or DHEA-treated females and vehicle-treated Gx females compared to their uninfected controls. A likely explanation is that *Plasmodium* infection induces apoptosis in this cell population [45].

In this study, uninfected intact females had higher numbers of CD3⁺/CD8⁺ cells than males under the same conditions, which corroborates the findings of Arsenovic et al., who demonstrated that female rats have higher numbers of CD8⁺ T lymphocytes than males [46]. We show that DHEA administration increased the number of CD3⁺/CD8⁺ cells in uninfected female mice, further accentuating the dimorphic pattern of this population. A probable explanation is that DHEA at high concentrations binds to ERα and Erβ (estrogen receptors) [47], and this interaction induces the proliferation of T-lymphocytes [48]. However, this possibility requires experimental demonstration in our model. Moreover, infection decreased the number of CD3⁺/CD8⁺ cells in intact males and females, corresponding with the decrease in total CD3⁺ cells during infection. This result is likely due to *Plasmodium* infection inducing apoptosis of CD3⁺/CD8⁺ T cells in the spleen [49]. Another possibility is that TCD3⁺/CD8⁺ lymphocytes migrate to lymphoid nodules or to the brain [50–52]. In addition, modifying the DHEA concentration did not change the number of macrophages in any group, which is probably why DHEA did not decrease parasitemia in our malaria model. Regarding CD19⁺ cells, DHEA administration did not modify this population; nevertheless, gonadectomy decreased it in both uninfected and infected females treated with vehicle. This effect is probably due to the loss of estrogen regulation of CD19⁺ cells by removing the gonads, as previously shown [6,53].

Finally, gonadectomy induced a dimorphic pattern in the NK cell population, with Gx-uninfected male mice showing higher numbers of NK cells than females under the same conditions. This dimorphic pattern may be associated with Gx males developing higher IL-2 concentrations than Gx females, as IL-2 is one of the main factors promoting NK cell proliferation [54]. In summary, DHEA administration decreased the concentration of IL-2 in Gx females, which increased the number of CD3⁺/CD8⁺ T cells.

Immune cells modulate immunity by secreting cytokines. In this work, we detected that infection with *P. berghei* ANKA increased the concentrations of IFN-γ and TNF-α, which corroborates what has been previously described [55]. In addition, DHEA administration decreased the concentration of IFN-γ in intact infected females compared to intact infected males, resulting in a sex-associated pattern. This pattern corresponds to the lower concentration of IFN-γ in DHEA-treated female rats infected with *Trypanosoma cruzi* compared to male rats under the same conditions [56], which indicates that DHEA modulates cytokine production differently in males and females. Furthermore, DHEA inhibits the inflammatory response by preventing NF-κB translocation and PI3K-mediated signaling [57], which promotes the activation of lymphocytes. Finally, it is also likely that DHEA decreases IFN-γ secretion by modulating NK cells, which are the main IFN-γ expressing cells in the spleen [49].

On the other hand, gonadectomy increased the IL-2 concentration in males compared to females without infection, resulting in a dimorphic pattern. Interestingly, this sex-associated pattern was eliminated when DHEA was administered, which corroborates the findings of Pratschke et al. who reported that DHEA administration decreases IL-2 synthesis in lymphocytes from patients with abdominal surgery. [58]. In contrast, in the infected groups, the reconstitution of Gx mice with DHEA decreased the IL-2 concentration in females only, resulting in a sex-dependent pattern.

Furthermore, infection increased the IL-10 concentration exclusively in intact females, resulting in a dimorphic pattern that was eliminated by DHEA administration. Interestingly, this group had the lowest DHEA concentration and the highest IL-10 levels. This finding corresponds with that described in patients with systemic lupus erythematosus in whom DHEA administration decreases their IL-10 concentration [59]. Another possible explanation is the reciprocal regulation between the concentration of the cytokines IFN-γ

and IL-10 in *Plasmodium* infection [60], which is relevant given that IL-10 decreases the incidence of cerebral malaria and modulates the effects of IFN- γ [61,62]. Moreover, Omer and Riley demonstrated that TGF- β , in addition to increasing IL-10 concentration, improves survival and decreases parasitemia in *Plasmodium berghei* infected mice [63]. Given that Treg cells are important for regulating inflammation by producing TGF- β [64] and that DHEA modulates the proliferation of Tregs [65], it would be important to evaluate the effect of DHEA on TGF- β concentration as well as to assess the population of Tregs to understand the immunoregulation exerted by DHEA in this disease.

An interesting finding is that modifying the concentration of DHEA in infected mice only affected the concentration of IL-2 and IL-4 in Gx females, which generated dimorphic patterns depending on sex, a likely explanation being that IL-2 is required for IL-4 synthesis [66]. In addition, DHEA administration decreased the IL-4 concentration in uninfected Gx males, which was expected given that this steroid suppresses the Th2-type response [13,67]. Regarding IL-6, DHEA administration increased the levels of this cytokine in uninfected Gx male and female mice, an effect that differs from that reported by Sudo et al., who described that DHEA administration suppressed IL-6 levels [33]. The discrepancy in this result is probably because we performed the determination in Gx mice, and hormones produced in the gonads, such as testosterone, also upregulate the synthesis of this cytokine [68].

Related to the antibody levels, reconstituting Gx and infected males with DHEA increased the concentration of IgG1, IgG2a, IgG2b and IgG3 antibodies in plasma. This finding corroborates the results of Cheng and Tseng, who described that DHEA administration increases the concentrations of IgM and IgG [69]. In addition, regardless of treatment or sex, the IgG1/IgG2a ratio was less than one, suggesting a predominance of the Th1-type response characteristic of *P. berghei* infections [70]. However, only DHEA-treated and infected Gx males exhibited increased IgG1, IgG2a, IgG2b and IgG3 levels. A probable explanation is that DHEA decreased the IL-4 concentration in Gx females under the same conditions, and this cytokine is a growth and differentiation factor for B lymphocytes that promotes their differentiation into antibody-producing cells [71].

The increase in antibody concentration induced by DHEA is important because it is associated with reduced disease severity [72]. It is critical to note that gonadectomy and DHEA administration could also affect testosterone and 17 β -estradiol levels, which could influence the interpretation of the results.

While the molecular mechanisms by which DHEA modifies immune function in malaria remain to be revealed, Vargas-Villavicencio et al. have shown that DHEA administration increases androgen receptor (AR) expression in the spleen [27]. In addition, DHEA also binds estrogen receptors ER α and ER β [40], suggesting that DHEA interacts with the AR, ER α and ER β present in immune response cells and acts through mechanisms involving a classical nuclear receptor in the immune system, although the affinity for these receptors is low [73]. In addition, DHEA may also be transformed into androgens, estrogens or other metabolites in peripheral tissues [74].

4. Materials and Methods

4.1. Mice

Four-week-old male or female CBA/Ca mice were used. The mice were a generous gift from Dr. William Jarra (National Institute for Medical Research, London, UK). The animals were bred and maintained in a specific pathogen-free environment in the animal house of the Facultad de Estudios Superiores Zaragoza, Universidad Nacional Autónoma de México. All protocols used in animal handling were authorized by the animal care and use committee, registration number 24/08/SO/3.4.1, respecting the official national standard NOM-062-ZOO-1999.

4.2. Gonadectomy

4.2.1. Orchidectomy

Male mice were gonadectomized (Gx) as previously described [8]. In brief, 4-week-old CBA/Ca male mice were anesthetized with a mixture of ketamine: xylazine 80 mg/kg: 8 mg/kg (Phoenix Pharmaceutical Inc., St. Joseph, MO, USA). The testes and epididymis were removed via scrotal excision under sterile conditions, and the efferent ducts were sealed by electrocautery.

4.2.2. Ovariectomy

Four-week-old female CBA/Ca mice were anesthetized with the mixture (ketamine:xylazine), incisions were made in the lower abdomens under aseptic conditions, ovaries were removed, and muscles were sutured. Male and female mice were used 4 weeks after surgery to allow for their recovery.

4.3. DHEA Administration

DHEA (Sigma-Aldrich, St. Louis, MO, USA) was diluted in vehicle (sweet almond oil LASA, México City, Mexico) and injected subcutaneously (8 mg/kg body weight) daily for 5 days prior to infection as previously described [27]. Control groups received 50 μ L of vehicle.

4.4. Parasite, Infection and Parasitemia

P. berghei ANKA was kindly donated by Dr. William Jarra (National Institute for Medical Research, London, UK). Parasites were expanded in mice and stored in cryovials under liquid nitrogen. To activate the parasite, a vial was thawed at room temperature and immediately intraperitoneally inoculated into a 4-week-old mouse. When parasitemia reached approximately 20%, a blood sample was drawn in cold PBS, the total number of erythrocytes was counted in a Neubauer chamber and blood was diluted with PBS to obtain a suspension with 1×10^4 parasitized red cells/mL. For infection, each mouse was inoculated intravenously with 100 μ L of the above suspension (1×10^3 parasitized red cells).

To evaluate parasitemia, blood smears were prepared, fixed with absolute methanol and stained with Giemsa (Sigma-Aldrich, St. Louis, MO, USA) diluted 1:10 in phosphate buffer. To quantify the number of parasites, the 100 \times objective of a Carl Zeiss Standard 20 Microscope (Carl Zeiss Ltd., Welwyn Garden City, UK) was used. Fifty fields were evaluated when parasitemia was less than 2%, and 200 erythrocytes were evaluated when parasitemia exceeded this number. Since *P. berghei* ANKA is lethal and most mice die around day 9 post infection, we sacrificed all mice on day 8 post infection [8]. Parasitemia in each group was presented as the geometric mean of parasitized red cells \pm SEM.

4.5. Body Weight and Temperature

Daily from day 0 to day 8 postinfection, all mice were weighed on a semianalytical balance (Ohaus, Parsippany, NJ, USA). Body temperature was also assessed daily using an infrared thermometer (Thermofocus, 01500A/H1N1, Vedano Olona-Varse, Italy).

4.6. Hemoglobin Concentration

Two microliters of mouse tail blood were collected daily and mixed with 498 μ L of Drabkin solution (1 mM potassium ferrocyanide, 7.6 mM potassium cyanide and 11.9 mM sodium bicarbonate (Sigma-Aldrich)). The mixture was incubated at room temperature for 5 min. The absorbance was measured at 540 nm on a plate reader (Multiskan GO, Thermo Fisher Scientific, Inc., Waltham, MA, USA). To calculate the hemoglobin concentration, a curve was prepared with a rat hemoglobin standard (Sigma-Aldrich).

4.7. Extraction of Sex Hormones from Plasma Samples

On day 8 post infection, mice were sacrificed, and blood was collected into heparinized tubes and centrifuged at $2000 \times g$ for 15 min. Plasma was stored in aliquots at $-70\text{ }^{\circ}\text{C}$ until use. For extraction of sex steroids, 100 μL of plasma, 900 μL of PBS and 5 mL of ethyl ether (JT Baker, Fisher Scientific SL, Phillipsburg, NJ, USA) were mixed vigorously for 5 min. The aqueous phase was frozen in a dry ice bath with 96% ethanol. The organic phase was transferred to a glass tube and evaporated in a water bath at $37\text{ }^{\circ}\text{C}$ for 48 h. Samples were diluted with 1000 μL of 0.1% PBS/gelatin (Sigma-Aldrich).

4.8. Quantification of DHEA in Plasma

The commercial DHEA kit EIA-3415 (DRG International, Springfield NJ, USA) was used. Ten microliters of plasma extract were mixed with 200 μL of DHEA-conjugated horseradish peroxidase and incubated for one hour at room temperature. The plates were washed 4 times. and 100 μL of tetramethylbenzidine (H_2O_2 -TMB 0.26 g/L) was added. The plates were incubated for 15 min at room temperature protected from light. The reaction was halted with stop solution, and the plates were gently shaken and read at 450 nm on a Multiskan Ascent 96 plate reader (Thermo Fisher Scientific, Waltham, MA, USA). The DHEA concentration was calculated using a standard curve included in the kit.

4.9. Quantification of Cell Populations in the Spleen

Quantification of cell populations was performed by flow cytometry as previously described [7]. Briefly, on day 8 post infection, mice were sacrificed, the spleen was disaggregated on nylon mesh, cells were washed with PBS and erythrocytes were removed with a lysis solution (Beckton and Dickinson, Franklin Lakes, NJ, USA). Cells were washed with sterile staining buffer (PBS, 1% bovine albumin, 0.1% NaN_2), and the cells were stained with the following dilutions of fluorochrome-coupled anti-mouse antibodies: CD3-FITC clone 17A2 (1:250), CD4-APC clone RM4-5 (1:1000), CD8-PE clone 53-6.7 (1:1000), CD107b-PE clone M3/84 (1:125), CD19-APC clone 1D3/CD19 (1:1600) and CD16/32-PE clone 93 (1:150). We used three antibody combinations: the first for total T cells (CD3^+), T-helper cells ($\text{CD3}^+/\text{CD4}^+$) and cytotoxic T cells ($\text{CD3}^+/\text{CD8}^+$). The second combination identified B cells and macrophages (CD19^+ and CD107b), respectively, and the third ($\text{CD19}^- \text{CD3}^- \text{CD16}^+/\text{CD32}^+$) identified NK cells. The cells were incubated for 30 min at room temperature and protected from light. The cells were then washed with PBS and suspended in 100 μL of FACS solution and acquired on a FACS Aria II cytometer (BD Biosciences, San Jose, CA, USA). From this region, we selected the CD3^+ population by plotting SSC vs FITC- CD3^+ and calculated the percentage positive for the first gate (to exclude CD19^+ and NK cells). Two dot plots corresponding to CD4^+ and CD8^+ cells were generated from the first region (SSC vs. APC- CD4^+ and SSC vs. PE- CD8^+), and the percentages were calculated according to the lymphocytes in the first gate. The sum of CD4^+ and CD8^+ lymphocyte percentages is the total CD3^+ lymphocyte count. In the second combination, CD19^+ and CD107b cells were identified with APC-anti CD19 and PE-anti CD107b , respectively. The CD19^+ cells were then selected using the SSC dot plot vs APC- CD19 . Macrophages were selected based on the FSC and SSC dot blot, which corresponded to 100%. This procedure excludes CD3^+ , CD19^+ and NK cells from macrophages that were selected by plotting SSC vs. PE-anti CD107b . Finally, the third combination was used to identify NK cells and four dot plots were used: in the first, we selected lymphocytes and NK cells by drawing SSC vs. FSC; in the second dot blot we used FITC-anti CD3 to select the CD3^- population; in the third dot blot, we plotted APC-anti CD19 and selected CD19^- cells; and in the fourth dot blot, we plotted SSC vs PE- $\text{CD16}^+/\text{CD32}^+$. The first step to analyze the results was to define the negative and positive regions of the fluorescence in each dot plot, using the control of cells stained with a single antibody. Subsequently, the fluorescence of each tube containing only one fluorochrome and the isotype controls were used to determine the positive region for each antibody. For each mixture, 10,000 cells were

acquired. All antibodies were purchased from BioLegend (San Diego, CA, USA). Data were analyzed using FlowJo 2.5.1 software (Beckton and Dickinson, Ashland, OR, USA).

4.10. Plasma Th1/Th2/Th17 Cytokine Quantification

Groups of intact males or females or Gx mice were treated with DHEA and infected with *P. berghei* ANKA. On day 8 post infection, blood was drawn and plasma was separated and frozen at $-70\text{ }^{\circ}\text{C}$ until use. The cytokine levels of IFN- γ , TNF- α , IL-2, IL-4, IL6, IL-10 and IL-17 were quantified in plasma using the Cytometric Bead Array (CBA) Mouse (BD Mouse Th1/Th2/Th17 cytokine CBA Kit Biosciences-Pharmingen, Heidelberg, Germany). Briefly, 25 μL of plasma or 25 μL of the standards were incubated with 50 μL of beads coated with antibodies directed against IFN- γ , TNF- α , IL-2, IL-4, IL-6, IL-10 and IL-17. Each sample was incubated with 50 μL of detection reagent (phycoerythrin) for 2 h at room temperature and protected from light. One milliliter of wash solution was added, and samples were centrifuged at $200\times g$ for 5 min. The supernatant was removed, and the beads were suspended in 100 μL of wash solution. Finally, the samples were evaluated in a FACSAria II cytometer, and the results were analyzed with the FCAP array program. A standard curve was used to quantify the concentration of each cytokine. The detection levels in the kit used were as follows: IL-2 (0.1 pg/mL), IL-4 (0.03 pg/mL), IL-6 (1.4 pg/mL), IL-17 (0.8 pg/mL), IFN- γ (0.5 pg/mL) and TNF (0.9 pg/mL).

4.11. Quantification of IgM and IgG Antibody Concentrations

Antibody levels were assessed as described previously [75]. Briefly, 96-well plates (Corning, NY, USA) were incubated with 100 μL of *P. berghei* ANKA antigen solution (10 $\mu\text{g}/\text{mL}$) and blocked with 10% serum at $4\text{ }^{\circ}\text{C}$ overnight. The plate was washed and incubated with the test plasma samples for 2 h at $37\text{ }^{\circ}\text{C}$. Pre calibrated dilutions of biotinylated antibodies specific for IgM, IgG, IgG1, IgG2a, IgG2b and IgG3 (Zymed, South San Francisco, CA, USA) diluted in PBS tween 0.05% milk 0.02% were added and incubated for 1 h at $37\text{ }^{\circ}\text{C}$. The plate was washed and incubated with streptavidin–peroxidase solution (Sigma-Aldrich) diluted 1:2500 in PBS tween 0.05% for 1 h at $37\text{ }^{\circ}\text{C}$. After washing, the plate was incubated with 100 μL orthophenylenediamine (OPD) and 0.03% H_2O_2 diluted in citrate buffer (Sigma-Aldrich). Finally, the reaction was halted with 100 μL of 0.1 N H_2SO_4 , and the absorbance at 450 nm was measured on a Multiskan GO plate spectrophotometer (Thermo-Fisher Scientific, Inc.). Since there are no commercial standards to be used as positive controls, optical density at 450 nm was used to denote antibody levels.

4.12. Statistical Analysis

Parasitemia, body weight, temperature, hemoglobin concentration, steroid concentration, cell populations, cytokine concentration and antibodies, for calculations two-way ANOVA was performed using Statgraphics XVI (The Plains, VA, USA) $p \leq 0.05$ with a Bonferroni post hoc test $p \leq 0.05$.

5. Conclusions

In contrast to our expectations, DHEA increased parasitemia in intact males, and its immunomodulatory effects depended on sex, parasite infection and DHEA concentration. Our results explain, at least in part, the sexual dimorphism in the immune response in malaria since DHEA administration decreased IFN- γ and IL-10 concentrations only in intact females. In contrast, in gonadectomized males, DHEA increased the number of CD8^+ T cells without infection. Interestingly, in infected Gx males, DHEA increased IgG1, IgG2a, IgG2b, IgG3 and IL-4 levels, corresponding to both Th1 and Th2 responses (Figure 8). However, the molecular mechanisms involved are unknown. Therefore, it is essential to explore in depth the protective mechanisms of DHEA in cerebral malaria, where a Th2-type response could counteract the inflammatory process of this complication. Finally, this work helps to explain at least in part the sexual dimorphism of the immune response in malaria,

which will be useful for the future development of therapies that are specific to men and women, as they respond differently to DHEA administration.

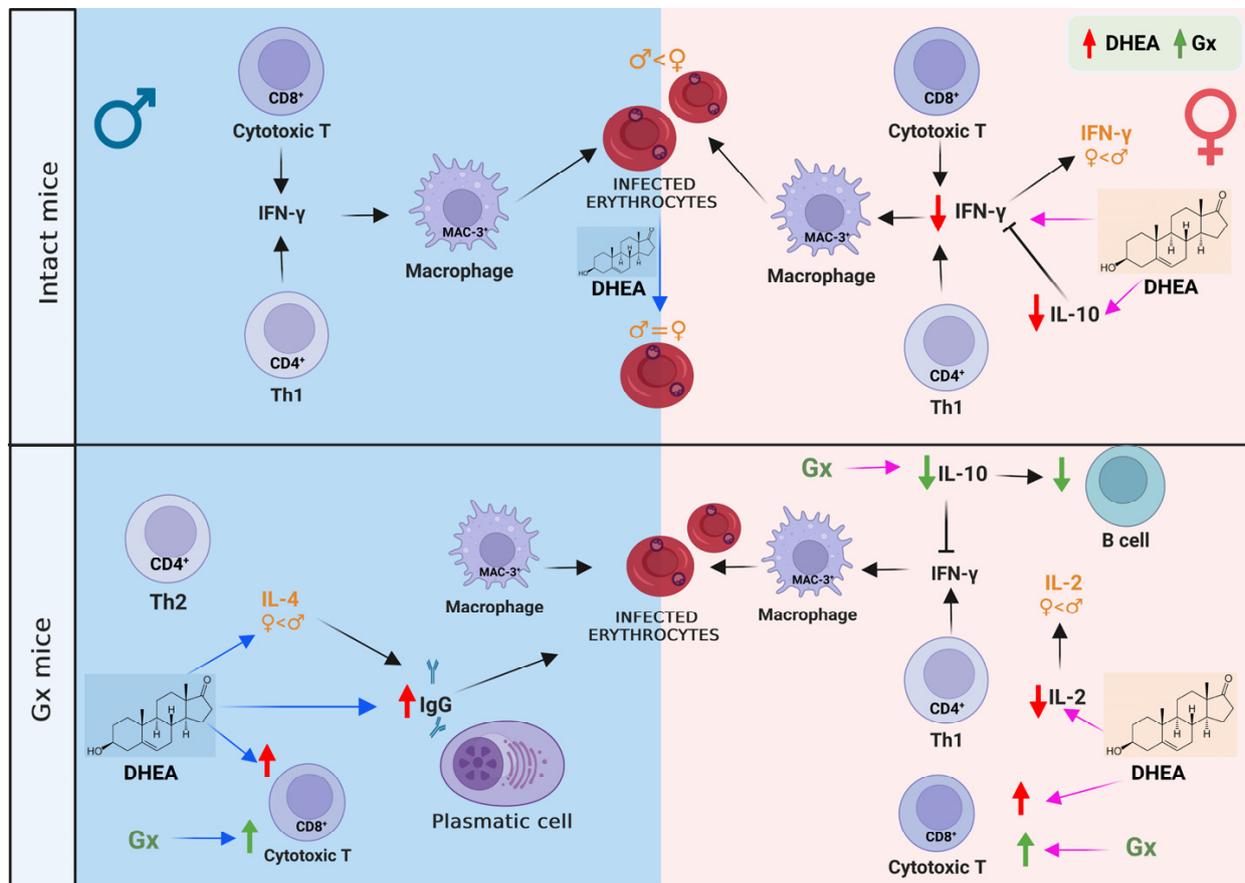


Figure 8. Effect of DHEA concentration on sexual dimorphism in the immune response of male and female CBA/Ca mice infected with *P. berghei* ANKA. The blue box represents DHEA concentration-induced changes in males, while the pink box represents changes in females. The green arrows indicate the effects of lowering DHEA concentration by gonadectomy and red arrows represent the effect of increasing DHEA concentration on the corresponding variables. Administering DHEA to intact males increased their parasitemia, which eliminated the dimorphic pattern present in intact mice treated with vehicle. In contrast, in the group of intact female mice, the administration of DHEA decreased the concentration of IFN- γ , which generated a dimorphic pattern and intact males presented a higher concentration of INF- γ than females under the same conditions. In addition, DHEA decreased IL-10 concentration in intact females; possibly, this is associated with IL-10 modulating IFN- γ secretion in *Plasmodium* infections. Interestingly, DHEA administration generated a dimorphic pattern in IL-2 and IL-4 concentrations in which Gx males presented a higher concentration of these cytokines than Gx females. Additionally, gonadectomy increased the number of CD8⁺ lymphocytes in both sexes; likewise, DHEA administration increased the number of this cell population. Furthermore, gonadectomy decreased the number of B cells in females, possibly by decreasing the IL-10 concentration. Reconstituting Gx females with DHEA did not restore B-cell number or IL-10 concentration. In contrast, reconstituting Gx males with DHEA increased the levels of IgG antibodies, corresponding with the higher IL-4 concentration in Gx males. The figure was created with BioRender.

Supplementary Materials: The supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ijms241612549/s1>.

Author Contributions: All authors significantly contributed to this work. F.O.B.-G. conducted the study, data processing, discussion of results and wrote the draft of manuscript; L.A.C.-C. quantification of immune cells, data processing and discussion of results; J.A.-C. graphics design and result discussion; T.J.N.-P. data obtention, tables design and discussion of results; O.F.-R. cytokine quantification and result discussion; M.S.L.-P. data obtention and discussion of results; D.R.C.-R. steroids quantification and discussion of results; A.C.-S. statistical analysis and result discussion; M.L.-H. conceived the project, designed experiments, performed analysis and discussion of results and wrote the paper. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported by the Programa de Apoyo a Proyectos de Investigación e Innovación Tecnológica, Universidad Nacional Autónoma de México, PAPIIT IN228620 awarded to Martha Legorreta-Herrera.

Institutional Review Board Statement: The research protocol was reviewed and approved by the Local Ethics Committee of the Facultad de Estudios Superiores Zaragoza, UNAM. Registration number 28/04/SO/3.4.1 in strict compliance with the Mexican Official Standard NOM-94-062-ZOO-1999.

Informed Consent Statement: Not applicable.

Data Availability Statement: The raw data supporting the findings of this paper will be made available by the authors without undue reservation.

Acknowledgments: Fidel O. Buendía-González belongs to the Posgrado en Ciencias Biológicas, UNAM is a fellow from the Programa de Becas Nacionales CONACyT: 743453, and this work is part of the requirements for his PhD degree from the Posgrado en Ciencias Biológicas, UNAM. Jesús Aguilar-Castro, Teresita de Jesús Nolasco-Pérez, Omar Fernández-Rivera and Monserrat S. López-Padilla are CONACyT fellows from the Posgrado en Ciencias Biológicas, UNAM. The authors would like to thank María Isabel Soto Cruz, Abraham Landa Piedra and Víctor Hugo Salazar-Castañón UNAM for the helpful discussion of results. In addition, we thank Adriana Altamirano Bautista, Román Hernández Meza and Dolores Elizabeth Guzmán Andrade, UNAM, for help with gonadectomy and taking care of mice. We also thank Guadalupe García Gómez for help with flow cytometry.

Conflicts of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. World Health Organization. *World Malaria Report 2021*; World Health Organization: Geneva, Switzerland, 2021.
2. Landgraf, B.; Kollaritsch, H.; Wiedermann, G.; Wernsdorfer, W.H. Parasite density of *Plasmodium falciparum* malaria in Ghanaian schoolchildren: Evidence for influence of sex hormones? *Trans. R. Soc. Trop. Med. Hyg.* **1994**, *88*, 73–74. [[CrossRef](#)] [[PubMed](#)]
3. Wildling, E.; Winkler, S.; Kremsner, P.G.; Brandts, C.; Jenne, L.; Wernsdorfer, W.H. Malaria epidemiology in the province of Moyen Ogoov, Gabon. *Trop. Med. Parasitol.* **1995**, *46*, 77–82. [[PubMed](#)]
4. Klein, S.L. Hormonal and immunological mechanisms mediating sex differences in parasite infection. *Parasite Immunol.* **2004**, *26*, 247–264. [[CrossRef](#)]
5. Aguilar-Castro, J.; Cervantes-Candelas, L.A.; Buendia-Gonzalez, F.O.; Nolasco-Perez, T.J.; Lopez-Padilla, M.S.; Fernandez-Rivera, O.; Cervantes-Sandoval, A.; Legorreta-Herrera, M. Dimorphic effect of 17beta-oestradiol on pathology and oxidative stress in experimental malaria. *Immunobiology* **2020**, *225*, 151873. [[CrossRef](#)]
6. Cervantes-Candelas, L.A.; Aguilar-Castro, J.; Buendia-Gonzalez, F.O.; Fernandez-Rivera, O.; Nolasco-Perez, T.J.; Lopez-Padilla, M.S.; Chavira-Ramirez, D.R.; Legorreta-Herrera, M. 17beta-Estradiol Is Involved in the Sexual Dimorphism of the Immune Response to Malaria. *Front. Endocrinol.* **2021**, *12*, 643851. [[CrossRef](#)]
7. Aguilar-Castro, J.; Cervantes-Candelas, L.A.; Buendia-Gonzalez, F.O.; Fernandez-Rivera, O.; Nolasco-Perez, T.J.; Lopez-Padilla, M.S.; Chavira-Ramirez, D.R.; Cervantes-Sandoval, A.; Legorreta-Herrera, M. Testosterone induces sexual dimorphism during infection with *Plasmodium berghei* ANKA. *Front. Cell Infect. Microbiol.* **2022**, *12*, 968325. [[CrossRef](#)] [[PubMed](#)]
8. Legorreta-Herrera, M.; Mosqueda-Romo, N.A.; Nava-Castro, K.E.; Morales-Rodriguez, A.L.; Buendia-Gonzalez, F.O.; Morales-Montor, J. Sex hormones modulate the immune response to *Plasmodium berghei* ANKA in CBA/Ca mice. *Parasitol. Res.* **2015**, *114*, 2659–2669. [[CrossRef](#)] [[PubMed](#)]
9. Cernetich, A.; Garver, L.S.; Jedlicka, A.E.; Klein, P.W.; Kumar, N.; Scott, A.L.; Klein, S.L. Involvement of gonadal steroids and gamma interferon in sex differences in response to blood-stage malaria infection. *Infect. Immun.* **2006**, *74*, 3190–3203. [[CrossRef](#)] [[PubMed](#)]
10. Rutkowski, K.; Sowa, P.; Rutkowska-Talipska, J.; Kuryliszyn-Moskal, A.; Rutkowski, R. Dehydroepiandrosterone (DHEA): Hypes and hopes. *Drugs* **2014**, *74*, 1195–1207. [[CrossRef](#)]

11. Knoferl, M.W.; Angele, M.K.; Catania, R.A.; Diodato, M.D.; Bland, K.I.; Chaudry, I.H. Immunomodulatory effects of dehydroepiandrosterone in proestrus female mice after trauma-hemorrhage. *J. Appl. Physiol.* **2003**, *95*, 529–535. [[CrossRef](#)]
12. Solerte, S.B.; Fioravanti, M.; Vignati, G.; Giustina, A.; Cravello, L.; Ferrari, E. Dehydroepiandrosterone sulfate enhances natural killer cell cytotoxicity in humans via locally generated immunoreactive insulin-like growth factor I. *J. Clin. Endocrinol. Metab.* **1999**, *84*, 3260–3267. [[CrossRef](#)] [[PubMed](#)]
13. Cao, J.; Yu, L.; Zhao, J.; Ma, H. Effect of dehydroepiandrosterone on the immune function of mice in vivo and in vitro. *Mol. Immunol.* **2019**, *112*, 283–290. [[CrossRef](#)]
14. Leplina, O.Y.; Tikhonova, M.A.; Sakchno, L.V.; Tyrinova, T.V.; Ostanin, A.A.; Chernykh, E.R. Effect of dehydroepiandrosterone sulfate on maturation and functional properties of interferon-alpha-induced dendritic cells. *Bull. Exp. Biol. Med.* **2009**, *148*, 68–71. [[CrossRef](#)] [[PubMed](#)]
15. Ben-Nathan, D.; Padgett, D.A.; Loria, R.M. Androstenediol and dehydroepiandrosterone protect mice against lethal bacterial infections and lipopolysaccharide toxicity. *J. Med. Microbiol.* **1999**, *48*, 425–431. [[CrossRef](#)] [[PubMed](#)]
16. Loria, R.M.; Inge, T.H.; Cook, S.S.; Szakal, A.K.; Regelson, W. Protection against acute lethal viral infections with the native steroid dehydroepiandrosterone (DHEA). *J. Med. Virol.* **1988**, *26*, 301–314. [[CrossRef](#)]
17. Filipin Mdel, V.; Caetano, L.C.; Brazao, V.; Santello, F.H.; Toldo, M.P.; do Prado, J.C., Jr. DHEA and testosterone therapies in *Trypanosoma cruzi*-infected rats are associated with thymic changes. *Res. Vet. Sci.* **2010**, *89*, 98–103. [[CrossRef](#)] [[PubMed](#)]
18. Kurtis, J.D.; Friedman, J.F.; Leenstra, T.; Langdon, G.C.; Wu, H.W.; Manalo, D.L.; Su, L.; Jiz, M.; Jarilla, B.; Pablo, A.O.; et al. Pubertal development predicts resistance to infection and reinfection with *Schistosoma japonicum*. *Clin. Infect. Dis.* **2006**, *42*, 1692–1698. [[CrossRef](#)]
19. Fallon, P.G.; Richardson, E.J.; Jones, F.M.; Dunne, D.W. Dehydroepiandrosterone sulfate treatment of mice modulates infection with *Schistosoma mansoni*. *Clin. Diagn. Lab. Immunol.* **1998**, *5*, 251–253. [[CrossRef](#)]
20. Galindo-Sevilla, N.; Soto, N.; Mancilla, J.; Cerbulo, A.; Zambrano, E.; Chavira, R.; Huerto, J. Low serum levels of dehydroepiandrosterone and cortisol in human diffuse cutaneous leishmaniasis by *Leishmania mexicana*. *Am. J. Trop. Med. Hyg.* **2007**, *76*, 566–572. [[CrossRef](#)]
21. Freilich, D.; Ferris, S.; Wallace, M.; Leach, L.; Kallen, A.; Frincke, J.; Ahlem, C.; Hacker, M.; Nelson, D.; Hebert, J. 16alpha-bromoepiandrosterone, a dehydroepiandrosterone (DHEA) analogue, inhibits *Plasmodium falciparum* and *Plasmodium berghei* growth. *Am. J. Trop. Med. Hyg.* **2000**, *63*, 280–283. [[CrossRef](#)]
22. Bernin, H.; Lotter, H. Sex bias in the outcome of human tropical infectious diseases: Influence of steroid hormones. *J. Infect. Dis.* **2014**, *209* (Suppl. S3), S107–S113. [[CrossRef](#)]
23. Neunzig, J.; Bernhardt, R. Dehydroepiandrosterone sulfate (DHEAS) stimulates the first step in the biosynthesis of steroid hormones. *PLoS ONE* **2014**, *9*, e89727. [[CrossRef](#)]
24. Buendia-Gonzalez, F.O.; Legorreta-Herrera, M. The Similarities and Differences between the Effects of Testosterone and DHEA on the Innate and Adaptive Immune Response. *Biomolecules* **2022**, *12*, 1768. [[CrossRef](#)]
25. Kurtis, J.D.; Mtalib, R.; Onyango, F.K.; Duffy, P.E. Human resistance to *Plasmodium falciparum* increases during puberty and is predicted by dehydroepiandrosterone sulfate levels. *Infect. Immun.* **2001**, *69*, 123–128. [[CrossRef](#)] [[PubMed](#)]
26. Leenstra, T.; ter Kuile, F.O.; Kariuki, S.K.; Nixon, C.P.; Oloo, A.J.; Kager, P.A.; Kurtis, J.D. Dehydroepiandrosterone sulfate levels associated with decreased malaria parasite density and increased hemoglobin concentration in pubertal girls from western Kenya. *J. Infect. Dis.* **2003**, *188*, 297–304. [[CrossRef](#)] [[PubMed](#)]
27. Vargan-Villavicencio, J.A.; Larralde, C.; Morales-Montor, J. Treatment with dehydroepiandrosterone in vivo and in vitro inhibits reproduction, growth and viability of *Taenia crassiceps* metacystodes. *Int. J. Parasitol.* **2008**, *38*, 775–781. [[CrossRef](#)] [[PubMed](#)]
28. Ghosh, D.; Stumhofer, J.S. The spleen: “epicenter” in malaria infection and immunity. *J. Leukoc. Biol.* **2021**, *110*, 753–769. [[CrossRef](#)]
29. Lyke, K.E.; Burges, R.; Cissoko, Y.; Sangare, L.; Dao, M.; Diarra, I.; Kone, A.; Harley, R.; Plowe, C.V.; Doumbo, O.K.; et al. Serum levels of the proinflammatory cytokines interleukin-1 beta (IL-1beta), IL-6, IL-8, IL-10, tumor necrosis factor alpha, and IL-12(p70) in Malian children with severe *Plasmodium falciparum* malaria and matched uncomplicated malaria or healthy controls. *Infect. Immun.* **2004**, *72*, 5630–5637. [[CrossRef](#)]
30. Awandare, G.A.; Goka, B.; Boeuf, P.; Tetteh, J.K.; Kurtzhals, J.A.; Behr, C.; Akanmori, B.D. Increased levels of inflammatory mediators in children with severe *Plasmodium falciparum* malaria with respiratory distress. *J. Infect. Dis.* **2006**, *194*, 1438–1446. [[CrossRef](#)]
31. Powell, J.M.; Sonnenfeld, G. The effects of dehydroepiandrosterone (DHEA) on in vitro spleen cell proliferation and cytokine production. *J. Interferon Cytokine Res.* **2006**, *26*, 34–39. [[CrossRef](#)]
32. Langhorne, J.; Cross, C.; Seixas, E.; Li, C.; Von Der Weid, T. A role for B cells in the development of T cell helper function in a malaria infection in mice. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 1730–1734. [[CrossRef](#)]
33. Sudo, N.; Yu, X.N.; Kubo, C. Dehydroepiandrosterone attenuates the spontaneous elevation of serum IgE level in NC/Nga mice. *Immunol. Lett.* **2001**, *79*, 177–179. [[CrossRef](#)] [[PubMed](#)]
34. Medina, M.C.; Souza, L.C.; Caperuto, L.C.; Anhe, G.F.; Amanso, A.M.; Teixeira, V.P.; Bordin, S.; Carpinelli, A.R.; Britto, L.R.; Barbieri, R.L.; et al. Dehydroepiandrosterone increases beta-cell mass and improves the glucose-induced insulin secretion by pancreatic islets from aged rats. *FEBS Lett.* **2006**, *580*, 285–290. [[CrossRef](#)] [[PubMed](#)]

35. Dalla Valle, L.; Couet, J.; Labrie, Y.; Simard, J.; Belvedere, P.; Simontacchi, C.; Labrie, F.; Colombo, L. Occurrence of cytochrome P450c17 mRNA and dehydroepiandrosterone biosynthesis in the rat gastrointestinal tract. *Mol. Cell. Endocrinol.* **1995**, *111*, 83–92. [[CrossRef](#)] [[PubMed](#)]
36. Zhou, Z.; Moore, T.M.; Drew, B.G.; Ribas, V.; Wanagat, J.; Civelek, M.; Segawa, M.; Wolf, D.M.; Norheim, F.; Seldin, M.M.; et al. Estrogen receptor alpha controls metabolism in white and brown adipocytes by regulating Polg1 and mitochondrial remodeling. *Sci. Transl. Med.* **2020**, *12*, eaax8096. [[CrossRef](#)]
37. Bobyleva, V.; Kneer, N.; Bellei, M.; Battelli, D.; Lardy, H.A. Concerning the mechanism of increased thermogenesis in rats treated with dehydroepiandrosterone. *J. Bioenerg. Biomembr.* **1993**, *25*, 313–321. [[CrossRef](#)]
38. Peschle, C.; Rappaport, I.A.; Sasso, G.F.; Condorelli, M.; Gordon, A.S. The role of estrogen in the regulation of erythropoietin production. *Endocrinology* **1973**, *92*, 358–362. [[CrossRef](#)]
39. Voegeli, T.A.; Kurtz, A.; Grimm, M.O.; Effert, P.; Eckardt, K.U. Anemia under androgen deprivation: Influence of flutamide, cyproteroneacetate and orchietomy on the erythropoietin system. *Horm. Metab. Res.* **2005**, *37*, 89–93. [[CrossRef](#)]
40. Chen, F.; Knecht, K.; Birzin, E.; Fisher, J.; Wilkinson, H.; Mojena, M.; Moreno, C.T.; Schmidt, A.; Harada, S.; Freedman, L.P.; et al. Direct agonist/antagonist functions of dehydroepiandrosterone. *Endocrinology* **2005**, *146*, 4568–4576. [[CrossRef](#)]
41. dos Santos, C.D.; Toldo, M.P.A.; do Prado Júnior, J.C. *Trypanosoma cruzi*: The effects of dehydroepiandrosterone (DHEA) treatment during experimental infection. *Acta Trop.* **2005**, *95*, 109–115. [[CrossRef](#)]
42. Lu, S.F.; McKenna, S.E.; Cologer-Clifford, A.; Nau, E.A.; Simon, N.G. Androgen receptor in mouse brain: Sex differences and similarities in autoregulation. *Endocrinology* **1998**, *139*, 1594–1601. [[CrossRef](#)] [[PubMed](#)]
43. Azzi, L.; El-Alfy, M.; Labrie, F. Gender differences and effects of sex steroids and dehydroepiandrosterone on androgen and oestrogen alpha receptors in mouse sebaceous glands. *Br. J. Dermatol.* **2006**, *154*, 21–27. [[CrossRef](#)] [[PubMed](#)]
44. Refaeli, Y.; Van Parijs, L.; London, C.A.; Tschopp, J.; Abbas, A.K. Biochemical mechanisms of IL-2-regulated Fas-mediated T cell apoptosis. *Immunity* **1998**, *8*, 615–623. [[CrossRef](#)] [[PubMed](#)]
45. Sanchez-Torres, L.; Rodriguez-Ropon, A.; Aguilar-Medina, M.; Favila-Castillo, L. Mouse splenic CD4⁺ and CD8⁺ T cells undergo extensive apoptosis during a *Plasmodium chabaudi chabaudi* AS infection. *Parasite Immunol.* **2001**, *23*, 617–626. [[CrossRef](#)] [[PubMed](#)]
46. Arsenovic-Ranin, N.; Kosec, D.; Pilipovic, I.; Nacka-Aleksic, M.; Bufan, B.; Stojic-Vukanic, Z.; Leposavic, G. Sex and age as determinants of rat T-cell phenotypic characteristics: Influence of peripubertal gonadectomy. *Mol. Cell. Biochem.* **2017**, *431*, 169–185. [[CrossRef](#)]
47. Bruder, J.M.; Sobek, L.; Oettel, M. Dehydroepiandrosterone stimulates the estrogen response element. *J. Steroid Biochem. Mol. Biol.* **1997**, *62*, 461–466. [[CrossRef](#)]
48. Priyanka, H.P.; Krishnan, H.C.; Singh, R.V.; Hima, L.; Thyagarajan, S. Estrogen modulates in vitro T cell responses in a concentration- and receptor-dependent manner: Effects on intracellular molecular targets and antioxidant enzymes. *Mol. Immunol.* **2013**, *56*, 328–339. [[CrossRef](#)]
49. Legorreta-Herrera, M.; Rivas-Contreras, S.; Ventura-Gallegos, J.; Zentella-Dehesa, A. Nitric oxide is involved in the upregulation of IFN-gamma and IL-10 mRNA expression by CD8(+) T cells during the blood stages of *P. chabaudi* AS infection in CBA/Ca mice. *Int. J. Biol. Sci.* **2011**, *7*, 1401–1411.
50. Belnoue, E.; Potter, S.M.; Rosa, D.S.; Mauduit, M.; Gruner, A.C.; Kayibanda, M.; Mitchell, A.J.; Hunt, N.H.; Renia, L. Control of pathogenic CD8⁺ T cell migration to the brain by IFN-gamma during experimental cerebral malaria. *Parasite Immunol.* **2008**, *30*, 544–553. [[CrossRef](#)]
51. Druzd, D.; Matveeva, O.; Ince, L.; Harrison, U.; He, W.; Schmal, C.; Herzel, H.; Tsang, A.H.; Kawakami, N.; Leliavski, A. Lymphocyte circadian clocks control lymph node trafficking and adaptive immune responses. *Immunity* **2017**, *46*, 120–132. [[CrossRef](#)]
52. Francelin, C.; Paulino, L.C.; Gameiro, J.; Verinaud, L. Effects of *Plasmodium berghei* on thymus: High levels of apoptosis and premature egress of CD4⁽⁺⁾CD8⁽⁺⁾ thymocytes in experimentally infected mice. *Immunobiology* **2011**, *216*, 1148–1154. [[CrossRef](#)] [[PubMed](#)]
53. Grimaldi, C.M.; Cleary, J.; Dagtas, A.S.; Moussai, D.; Diamond, B. Estrogen alters thresholds for B cell apoptosis and activation. *J. Clin. Investig.* **2002**, *109*, 1625–1633. [[CrossRef](#)] [[PubMed](#)]
54. Sharma, R.; Das, A. IL-2 mediates NK cell proliferation but not hyperactivity. *Immunol. Res.* **2018**, *66*, 151–157. [[CrossRef](#)]
55. Legorreta-Herrera, M.; Nava-Castro, K.E.; Palacios-Arreola, M.I.; Hernandez-Cervantes, R.; Aguilar-Castro, J.; Cervantes-Candelas, L.A.; Morales-Montor, J. Sex-Associated Differential mRNA Expression of Cytokines and Its Regulation by Sex Steroids in Different Brain Regions in a *Plasmodium berghei* ANKA Model of Cerebral Malaria. *Mediators Inflamm.* **2018**, *2018*, 5258797. [[CrossRef](#)] [[PubMed](#)]
56. Santos, C.D.; Toldo, M.P.; Santello, F.H.; Filipin Mdel, V.; Brazao, V.; do Prado Junior, J.C. Dehydroepiandrosterone increases resistance to experimental infection by *Trypanosoma cruzi*. *Vet. Parasitol.* **2008**, *153*, 238–243. [[CrossRef](#)] [[PubMed](#)]
57. Zhao, J.; Cao, J.; Yu, L.; Ma, H. Dehydroepiandrosterone alleviates *E. Coli* O157:H7-induced inflammation by preventing the activation of p38 MAPK and NF-kappaB pathways in mice peritoneal macrophages. *Mol. Immunol.* **2019**, *114*, 114–122. [[CrossRef](#)]
58. Pratschke, S.; von Dossow-Hanfstingl, V.; Dietz, J.; Schneider, C.P.; Tufman, A.; Albertsmeier, M.; Winter, H.; Angele, M.K. Dehydroepiandrosterone modulates T-cell response after major abdominal surgery. *J. Surg. Res.* **2014**, *189*, 117–125. [[CrossRef](#)]
59. Chang, D.M.; Chu, S.J.; Chen, H.C.; Kuo, S.Y.; Lai, J.H. Dehydroepiandrosterone suppresses interleukin 10 synthesis in women with systemic lupus erythematosus. *Ann. Rheum. Dis.* **2004**, *63*, 1623–1626. [[CrossRef](#)]

60. Medina, T.S.; Costa, S.P.; Oliveira, M.D.; Ventura, A.M.; Souza, J.M.; Gomes, T.F.; Vallinoto, A.C.; Povoá, M.M.; Silva, J.S.; Cunha, M.G. Increased interleukin-10 and interferon-gamma levels in *Plasmodium vivax* malaria suggest a reciprocal regulation which is not altered by IL-10 gene promoter polymorphism. *Malar. J.* **2011**, *10*, 264. [[CrossRef](#)]
61. Claser, C.; De Souza, J.B.; Thorburn, S.G.; Grau, G.E.; Riley, E.M.; Rénia, L.; Hafalla, J.C. Host resistance to Plasmodium-induced acute immune pathology is regulated by interleukin-10 receptor signaling. *Infect. Immun.* **2017**, *85*, e00941-16. [[CrossRef](#)]
62. Ho, A.; Liu, Y.; Khan, T.A.; Hsu, D.-H.; Bazan, J.F.; Moore, K.W. A receptor for interleukin 10 is related to interferon receptors. *Proc. Natl. Acad. Sci. USA* **1993**, *90*, 11267–11271. [[CrossRef](#)] [[PubMed](#)]
63. Omer, F.M.; Riley, E.M. Transforming growth factor beta production is inversely correlated with severity of murine malaria infection. *J. Exp. Med.* **1998**, *188*, 39–48. [[CrossRef](#)] [[PubMed](#)]
64. Tran, D.Q. TGF-beta: The sword, the wand, and the shield of FOXP3(+) regulatory T cells. *J. Mol. Cell Biol.* **2012**, *4*, 29–37. [[CrossRef](#)]
65. Coles, A.J.; Thompson, S.; Cox, A.L.; Curran, S.; Gurnell, E.M.; Chatterjee, V.K. Dehydroepiandrosterone replacement in patients with Addison's disease has a bimodal effect on regulatory (CD4+CD25hi and CD4+FoxP3+) T cells. *Eur. J. Immunol.* **2005**, *35*, 3694–3703. [[CrossRef](#)] [[PubMed](#)]
66. Raju, K.; Rabinovich, B.A.; Radvanyi, L.G.; Spaner, D.; Miller, R.G. A central role for IL-2 in fate determination of mature T cells—I: Role in determining the T_H1/T_H2 profile in primary T cell cultures. *Int. Immunol.* **2001**, *13*, 1453–1459. [[CrossRef](#)] [[PubMed](#)]
67. Tabata, N.; Tagami, H.; Terui, T. Dehydroepiandrosterone may be one of the regulators of cytokine production in atopic dermatitis. *Arch. Dermatol. Res.* **1997**, *289*, 410–414. [[CrossRef](#)] [[PubMed](#)]
68. Son, B.K.; Kojima, T.; Ogawa, S.; Akishita, M. Testosterone inhibits aneurysm formation and vascular inflammation in male mice. *J. Endocrinol.* **2019**, *241*, 307–317. [[CrossRef](#)]
69. Cheng, G.F.; Tseng, J. Regulation of murine interleukin-10 production by dehydroepiandrosterone. *J. Interferon Cytokine Res.* **2000**, *20*, 471–478. [[CrossRef](#)]
70. Huang, B.; Huang, S.; Liu, M.; Chen, Y.; Wu, B.; Guo, H.; Su, X.-Z.; Lu, F. T cell Ig and mucin-1 and-3 in *Plasmodium berghei* ANKA infection. *Parasitol. Res.* **2013**, *112*, 2713. [[CrossRef](#)]
71. Hofman, F.M.; Brock, M.; Taylor, C.R.; Lyons, B. IL-4 regulates differentiation and proliferation of human precursor B cells. *J. Immunol.* **1988**, *141*, 1185–1190. [[CrossRef](#)]
72. Yone, C.L.; Kremsner, P.G.; Luty, A.J. Immunoglobulin G isotype responses to erythrocyte surface-expressed variant antigens of *Plasmodium falciparum* predict protection from malaria in African children. *Infect. Immun.* **2005**, *73*, 2281–2287. [[CrossRef](#)] [[PubMed](#)]
73. Miller, K.K.; Al-Rayyan, N.; Ivanova, M.M.; Mattingly, K.A.; Ripp, S.L.; Klinge, C.M.; Prough, R.A. DHEA metabolites activate estrogen receptors alpha and beta. *Steroids* **2013**, *78*, 15–25. [[CrossRef](#)] [[PubMed](#)]
74. Nephew, K.P.; Sheeler, C.Q.; Dudley, M.D.; Gordon, S.; Nayfield, S.G.; Khan, S.A. Studies of dehydroepiandrosterone (DHEA) with the human estrogen receptor in yeast. *Mol. Cell. Endocrinol.* **1998**, *143*, 133–142. [[CrossRef](#)] [[PubMed](#)]
75. Legorreta-Herrera, M.; Ventura-Ayala, M.L.; Licona-Chavez, R.N.; Soto-Cruz, I.; Hernandez-Clemente, F.F. Early treatment during a primary malaria infection modifies the development of cross immunity. *Parasite Immunol.* **2004**, *26*, 7–17. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.