



Supplementary materials

Induction by Phenobarbital of Phase I and II Xenobiotic-Metabolizing Enzymes in Bovine Liver: An Overall Catalytic and Immunochemical Characterization

Michela Cantiello ^{1,†,‡}, Monica Carletti ^{1,†,§}, Mery Giantin ², Giulia Gardini ¹, Francesca Capolongo ², Paolo Cascio ¹, Marianna Pauletto ², Flavia Girolami ¹, Mauro Dacasto ^{2,*} and Carlo Nebbia ^{1,*}

¹ Department of Veterinary Sciences, University of Turin, 10095 Grugliasco, Italy; michela.cantiello@gmail.com (M.C.); monica.carletti@irta-ricerche.it (M.C.); giulia.gardini@unito.it (G.G.); paolo.cascio@unito.it (P.C.); flavia.girolami@unito.it (F.G.)

² Department of Comparative Biomedicine and Food Science, University of Padua, 35020 Agripolis Legnaro, Italy; mery.giantin@unipd.it (M.G.); francesca.capolongo@unipd.it (F.C.); marianna.pauletto@unipd.it (M.P.)

* Correspondence: mauro.dacasto@unipd.it (M.D.); carlo.nebbia@unito.it (C.N.)

† These authors contributed equally to this work.

‡ Current address: Eurofins Biopharma Services, 33000 Bordeaux, France.

§ Current address: IRTA Ricerche s.r.l., 10093 Collegno, Italy.

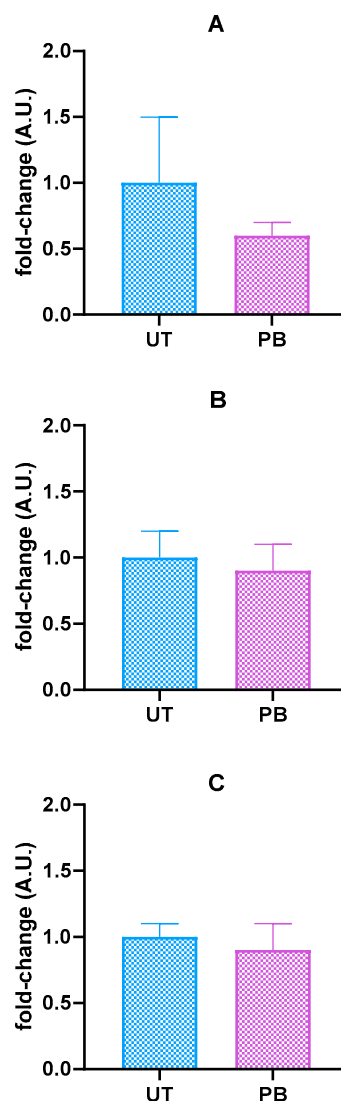


Figure S1. Hepatic nuclear receptors mRNA levels in untreated control (UT, n=3) and phenobarbital-treated cattle (PB, n=4). Total RNA was extracted from cattle liver aliquots; then, constitutive androstane receptor (CAR) (A), pregnane X receptor (PXR) (B), and retinoid X-receptor alpha (RXR α) (C) mRNA levels were measured by using a quantitative real-time RT-PCR (qPCR) approach. Data (arithmetic means \pm SD) are expressed as n-fold change (arbitrary units, A.U.), normalized to $\Delta\Delta C_t$ mean value of β -actin (ACTB, the chosen internal control gene, ICG), to whom an arbitrary value of 1 was assigned.

Table S1. Cytochromes P450 and b_5 content, NADPH cytochrome c (P450) reductase and NADH cytochrome b_5 reductase activities, and extent of metyrapone binding in untreated control (UT, $n = 3$) and phenobarbital-treated (PB, $n = 4$) cattle.

Parameter	UT	PB
total CYP [†]	0.80 ± 0.10	1.63 ± 0.24**
total cytochrome b_5 [†]	0.35 ± 0.13	0.45 ± 0.08
NADPH cytochrome c (P450) reductase [‡]	81.67 ± 10.10	87.38 ± 13.29
NADH cytochrome b_5 reduc- tase [‡]	1077.00 ± 108.67	383.50 ± 75.79***
metyrapone binding [†]	0.12 ± 0.03	0.96 ± 0.23**

Data are expressed as means ± SD. CYP: cytochrome P450. ** $p < 0.01$; *** $p < 0.001$ (unpaired t -test).

[†] nmoles/mg protein. [‡] nmoles/min·mg protein⁻¹.

Table S2. Cytochrome P450 (CYP) 1A, CYP2A, and CYP2E1 protein expression and *in vitro* metabolism of marker substrates in untreated control (UT, n = 3) and PB-treated (PB, n = 4) cattle.

Parameter	UT	PB
CYP1A expression [†]	214.00 ± 33.50	139.50 ± 25.60*
7-ethoxyresorufin deethylation [‡]	209.72 ± 40.71	138.15 ± 33.01*
7-methoxyresorufin deethylation [‡]	43.60 ± 1.18	165.57 ± 36.59**
benzo[a]pyrene hydroxylation [§]	1.22 ± 0.12	2.19 ± 0.35**
7-ethoxycoumarin deethylation [#]	0.71 ± 0.15	1.41 ± 0.29*
CYP2A expression [†]	N.A.	N.A.
coumarin hydroxylation [‡]	104.00 ± 41.00	95.00 ± 75.00
CYP2E1 expression [†]	125.50 ± 34.40	363.30 ± 192.80
Aniline hydroxylation [#]	0.65 ± 0.10	1.37 ± 0.33*
4-Aminophenol hydroxylation [#]	0.28 ± 0.05	0.79 ± 0.14**

Data are expressed as means ± SD. * $p < 0.05$; ** $p < 0.01$ (unpaired *t*-test). [†] arbitrary units (A.U.). [‡] pmoles/min·mg protein⁻¹. [§] µg quinine/mL. [#] nmoles/min·mg protein⁻¹. N.A. not available.

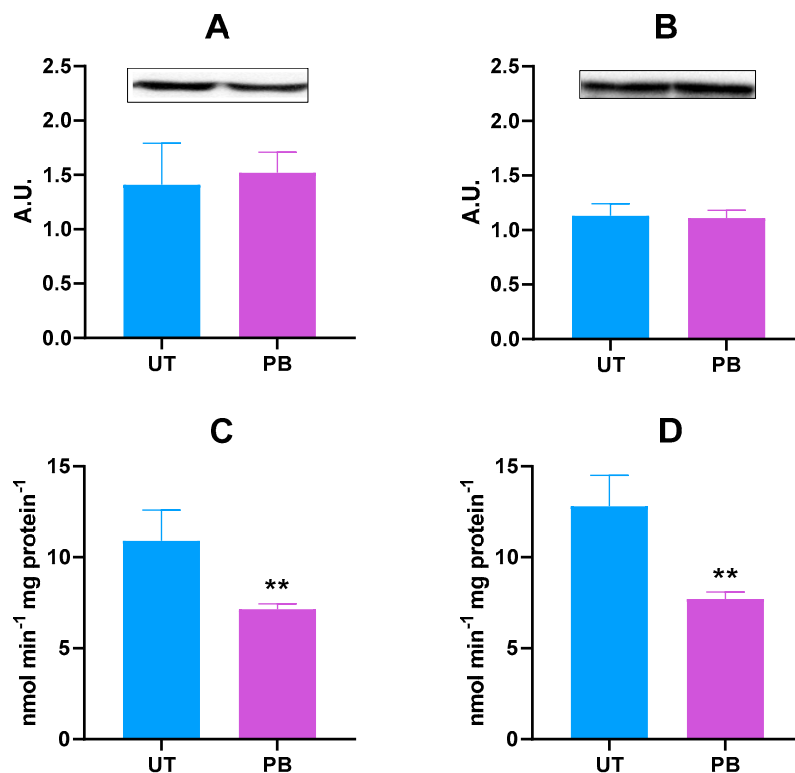


Figure S2. Hepatic flavin monooxygenase 1 and 3 (FMO1 and FMO3) protein expression (A-B) and *in vitro* metabolism of marker substrates ethylenethiourea and methimazole (C-D) in untreated control (UT, n=3) and phenobarbital-treated (PB, n=4) cattle. A.U., arbitrary units; ETU, ethylenethiourea; MTZ, methimazole. Data are expressed as arithmetic means \pm SD. ** $p < 0.01$; *** $p < 0.001$ (unpaired *t*-test).

Table S3. *In vitro* metabolism of carboxylesterase and epoxide hydrolase marker substrates in untreated control (UT, n = 3) and phenobarbital-treated (PB, n = 4) cattle.

Parameter	UT	PB
APA hydrolysis [∞]	5.55 ± 0.78	8.18 ± 1.08*
IPA hydrolysis [#]	334.87 ± 83.54	368.72 ± 18.56
PNP hydrolysis [∞]	5.65 ± 2.39	6.69 ± 0.90
<i>trans</i> -stilbene oxide epoxide hydrolysis [#]	2.46 ± 0.08	1.95 ± 0.36

APA: α -naphthylacetate; IPA: indophenyl acetate; PNP: *p*-nitrophenylacetate. Data are expressed as means \pm SD. * $p < 0.05$ (unpaired *t*-test). [∞] μ moles/min·mg protein⁻¹. [#] nmoles/min·mg protein⁻¹.

Table S4. Gene acronyms, GenBank accession numbers, qPCR primers (references and oligonucleotide sequences) and amplicon size.

Gene	GeneBank ID	Primer reference	5' → 3' primer sequence	Amplicon size (bp)
<i>ACTB</i>	NM_173979	Giantin et al., 2008	F: GTCGACACCGCAACCAGTT R: AAGCCGGCCTTGACAT	85
<i>CYP2B22</i>	NM_001075173	Giantin et al., 2008	F: GCGGACCTCATCCCCATT R: GTGCCCTTGGAAGGATGT	80
<i>CYP2C18</i>	NM_001076051	Giantin et al., 2008	F: ATGTTAAGAACATTGGCAAATCCTT R: GGCCATAGGTGTTTGAGAGATTG	51
<i>CYP2C31</i>	XM_600421	Giantin et al., 2008	F: TCCCAAGGGCACAACCATA R: CCTTGCCATCGTGCAGG	56
<i>CYP2C42</i>	XM_612374	Giantin et al., 2008	F: TCCCTGGACATGAACAACCC R: TTGTGCTTTTCCTGTTCCATCTT	71
<i>CYP3A</i>	NM_174531	Cantiello et al., 2009	F: GCCAGAGCCCGAGGAGTT R: GCAGGTAGACGTAAGGATTTATGCT	77
<i>PXR</i>	NM_001103226	Cantiello et al. 2009	F: TGAAGGCCTACATCGAGTTCAAC R: GGCCATGATCTTCAGGAACAA	68
<i>CAR</i>	NM_001079768	Cantiello et al. 2009	F: GAAGGACATGATCCTATCGACAGA R: CGTCGCTGGGCCTGTCT	63
<i>RXRα</i>	XM_881943	Cantiello et al. 2009	F: GCCTCAATGGTGTCTCAAAG R: AGCTGTACACCCCGTAGTGCTT	120

ACTB: β-actin; bp: base pairs; *CAR*, constitutive androstane receptor; *CYP2B22*, cytochrome P450 2B22; *CYP2C18*, cytochrome P450 2C18; *CYP2C31*, cytochrome P450 2C31; *CYP2C42*, cytochrome P450 2C42; *CYP3A*, cytochrome P450 3A; *PXR*, pregnane X receptor; qPCR, quantitative Real time RT-PCR; *RXRα*, retinoid X-receptor alpha.