

Calcium Signalling in Heart and Vessels: Role of Calmodulin and Downstream Calmodulin-Dependent Protein Kinases

Sofia Beghi ^{1,2,*}, Malgorzata Furmanik ¹, Armand Jaminon ¹, Rogier Veltrop ¹, Nikolas Rapp ¹, Kanin Wichapong ¹, Elham Bidar ³, Annamaria Buschini ² and Leon J. Schurgers ¹

- ¹ Cardiovascular Research Institute Maastricht (CARIM), Department of Biochemistry, Maastricht University, P.O. Box 616, 6200 MD Maastricht, The Netherlands
- ² Department of Chemistry, Life Sciences and Environmental Sustainability, University of Parma, Parco Area Delle Scienze 11A, 43124 Parma, Italy
- ³ Department of Cardiothoracic Surgery, Heart and Vascular Centre, Maastricht University Medical Centre+, 6229 HX Maastricht, The Netherlands
- * Correspondence: sofia.beghi@unipr.it; Tel.: +39-3408473527

Abstract: Cardiovascular disease is the major cause of death worldwide. The success of medication and other preventive measures introduced in the last century have not yet halted the epidemic of cardiovascular disease. Although the molecular mechanisms of the pathophysiology of the heart and vessels have been extensively studied, the burden of ischemic cardiovascular conditions has risen to become a top cause of morbidity and mortality. Calcium has important functions in the cardiovascular system. Calcium is involved in the mechanism of excitation–contraction coupling that regulates numerous events, ranging from the production of action potentials to the contraction of cardiomyocytes and vascular smooth muscle cells. Both in the heart and vessels, the rise of intracellular calcium is sensed by calmodulin, a protein that regulates and activates downstream kinases involved in the regulation of cardiac functions. In this review, we present the current literature regarding the role of calcium/calmodulin pathways in the heart and vessels with the aim to summarize our mechanistic understanding of this process and to open novel avenues for research.

Keywords: cardiovascular disease; calcium signalling; calmodulin; calcium calmodulin dependent protein kinases

1. Introduction

Cardiovascular disease (CVD) is a complex disease with many factors and events involved. The disbalanced regulation of physiology, mechanobiology, and molecular biology results in heart and vessel disease. Indeed, different factors coexist and cooperate, together increasing the risk of initiation and progression of CVD [1–3]. CVD is the leading cause of death all over the world, and more people die annually from this type of disease than from any other cause. According to the World Health Organization, every year 17 million people die of CVD globally. Additionally, in 2020, in which the death rate increased due to COVID-19, CVD remained the principal cause of death [4]. Among CVD, atherosclerosis is the most common cause, which is associated with the growth and build-up of cholesterol-rich plaques within the inner wall of the vessel. Arterial occlusion can limit or block blood flow to the heart or other organs, causing myocardial infarction, ischaemic cardiomyopathy, stroke and peripheral arterial disease [5].

Among the different pathways involved in cardiovascular homeostasis, the regulation of calcium signalling plays a crucial role. One of the major pathways in which calcium is involved is excitation contraction coupling. Here, the rise of intracellular calcium (Ca²⁺) in cardiomyocytes and vascular smooth muscle cells (VSMCs) is sensed by calmodulin (CaM), a multifunctional intermediate target of the secondary messenger Ca²⁺. CaM ensures



Citation: Beghi, S.; Furmanik, M.; Jaminon, A.; Veltrop, R.; Rapp, N.; Wichapong, K.; Bidar, E.; Buschini, A.; Schurgers, L.J. Calcium Signalling in Heart and Vessels: Role of Calmodulin and Downstream Calmodulin-Dependent Protein Kinases. *Int. J. Mol. Sci.* **2022**, *23*, 16139. https://doi.org/10.3390/ ijms232416139

Academic Editors: Giuseppe Zanotti and Tito Calì

Received: 9 October 2022 Accepted: 13 December 2022 Published: 17 December 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). activation of downstream calcium calmodulin kinase proteins (CaMKs), including CaMKK1, CAMKK2, CaMKI, CaMKII, and CaMKIV. CaMKs are kinases that upon phosphorylation result in multiple cellular functions. This signalling pathway is called the Ca²⁺-CaM-dependent kinase cascade [6,7] and is involved in many cellular processes, including glucose homeostasis, apoptosis, hematopoietic stem cell maintenance, normal immune cell function and cell proliferation [8–12].

Atherosclerosis as well as aortic aneurysm are aggravated by dysregulated calcium metabolism, causing vascular calcification (VC). VC is a pathological condition caused by calcium-phosphate precipitation leading to vascular stiffness. VC is an actively regulated process with a key role for VSMCs, the major cell type in the media layer of arteries. VSMCs are involved in regulating vascular tone and remodelling processes in the vessel wall upon injury [13–15]. VC is driven by an imbalance in inhibitors (e.g., matrix gla protein) and promotors (e.g., Ca^{2+}/PO_4^{3-} , cell death, VSMC dedifferentiation and extracellular matrix remodelling) [16]. Calcium deposition leading to calcification takes place extracellularly. However, its nidus may be derived from intracellular signalling leading to extracellular vesicle release from VSMCs [17]. Similarly, calcium-phosphate deposition in aortic heart valves causes aortic valve insufficiency and is mediated by vascular interstitial cells that create the nidus for calcium-phosphate crystals in the extracellular space. Aortic valve calcification narrows the opening of the valve, reducing blood flow and is linked to aortic valve stenosis [18–20].

Current knowledge of CVD and the underlying mechanisms is insufficient to decrease the burden of disease. The aim of our review is to summarize the state-of-the-art knowledge on the role of calcium signalling in cardiac and vascular tissue, with a focus on calmodulin and its downstream calcium kinases.

The Role of Calcium in the Cardiovascular System

Calcium is an abundant element in nature and the most abundant mineral in the human body. Calcium constitutes 1.43% of the human body, whilst phosphate makes up 1.11% of the human body. All other minerals and metals accumulate to a combined 0.73% of the human body. Some 99% of calcium in the body is found in bones and teeth, whilst the other 1% is found in blood and soft tissues. Calcium plays a crucial role in the heart and vessels, both in physiological and pathological conditions [7,21–24].

Cells in the heart and blood vessels need calcium to contract and to perform their function. The concentration of calcium within the cytosol is some 100 nM. The calcium concentration outside the cell is some 2 mM, thus a factor of 20,000 higher. Therefore, the influx of calcium into the cell needs to be tightly regulated. This calcium entry is regulated by calcium-sensing receptors (CaSR), which are essential to sense Ca^{2+} in the extracellular fluid and facilitate active and orchestrated calcium uptake [25–27]. Upon calcium entry, this activates a series of downstream pathways that directly regulate cellular functions [21,24]. The concentration of calcium inside cardiac cells has a heterogeneous distribution thanks to the presence of different microdomains in the cytosol, the spaces between intracellular organelles, and the delimited compartments such as the sarcoplasmic reticulum. Calcium homeostasis and signalling within the cell are regulated by calcium channels that come in many forms and flavours with diverse functions and structures. The main role of calcium channels is to balance both the cellular influx and the efflux of calcium [28,29]. Calcium channels can be stimulated by numerous and diverse stimuli such as membrane depolarization, extracellular and intracellular signalling molecules, and physical forces such as stretching or temperature. Among the calcium channels on plasma membranes, the voltage-gated calcium channels are one of the most important as they rapidly transport calcium into the cytoplasm. These channels respond to a change in voltage across the cell membrane. The plasma membrane also harbours calcium transport mechanisms involved in removing calcium from the cell to provide a low cytosolic concentration. These calcium transport mechanisms include, amongst others, the plasma membrane calcium ATPase (PMCA) pump and the Na⁺/Ca²⁺ exchanger (NCX) [24,30,31].

Calcium is an essential central player in the heart because it is involved in the core function of excitation–contraction coupling (ECC) [23]. ECC is a number of events starting at the production of the action potential (electrical impulse) to the contraction of the heart muscles. Additionally, calcium is a co-factor for many enzymes, including proteins involved in blood coagulation. Platelets and several coagulation factors are activated by Ca²⁺, which is responsible for maintaining blood clot architecture and strength [32,33]. Calcium is also involved in the regulation of long-term processes, such as gene regulation, in order to change protein expression. Moreover, calcium has a central role in adaptive tissue regulation, such as cardiac hypertrophy [22].

In VSMC physiology, calcium fulfils a pivotal role in physiology as well as in pathology. Here, calcium is a key element in regulating blood flow and vascular luminal diameter. Calcium induces contraction of VSMCs by creating a complex with the ubiquitous calcium binding protein calmodulin and increasing the activity of myosin light-chain kinase [34]. VSMCs are characterized by a high degree of plasticity, having the capability to switch phenotype. Indeed, under physiological conditions, VSMCs are in a quiescent contractile phenotype. However, upon pathobiological or mechanical stress, VSMCs switch towards a synthetic phenotype, characterized by a higher degree of migration and proliferation [14,35,36]. In both VSMC phenotypes, intracellular calcium plays a different but pivotal role. The influx of calcium in VSMC with contractile phenotype is regulated mainly via voltage-dependent L-type calcium channels, responsible for excitation-contraction coupling. Upon mechanical or biological stress, VSMCs switch phenotype to a reparative and synthetic VSMC. Synthetic VSMCs shed extracellular vesicles that contain endogenous calcification inhibitors to prevent mineralization of the extracellular matrix. Upon long-term stress and mineral disbalance, the content of these extracellular vesicles changes, turning them from anti-calcific into pro-calcific vesicles [37]. These extracellular vesicles form the nidus for VC and their uptake by synthetic VSMCs causes a rise in intracellular calcium and subsequently cellular stress, eventually leading to cell death. In synthetic VSMCs, calcium entry is less dependent on voltage-dependent L-type calcium channels [34]. Instead, calcium entry is in part via clathrin-mediated extracellular vesicles and may contribute to vascular calcification [36]. Intracellular calcium levels in VSMCs are very dynamic, especially in contractile VSMC, where calcium changes both in spatial and temporal domains. These oscillations of intracellular calcium are more pronounced in contractile VSMCs compared to VSMCs with a synthetic phenotype. The higher oscillation frequencies in contractile VSMC can increase kinase activity in a frequency-dependent manner through a trapping phenomenon consisting of the acquisition of autonomous kinase activity of certain CaM kinases [34].

Below we describe the role of calmodulin and its signalling pathways in more detail in relation to CVD.

2. Calmodulin

Calmodulin (CaM) is involved in the regulation and transduction of calcium signalling. CaM is an important primary sensor of intracellular calcium levels in eukaryotic cells, playing a pivotal role in the transduction and deciphering of calcium signalling [38,39]. CaM is a low-molecular-weight protein of 16 kDa, composed of 149 amino acid residues. There are three different human genes (*CaM1, CaM2, CaM3*) that encode for three highly conserved proteins (differing only at the nucleotide level) [40–43]. Each gene is characterized by distinct promoter elements and by unique 5'- and 3'-untranslated regions that may permit regulation of CaM expression at discrete cellular sites during differentiation [44]. CaM is an α -helical protein composed of N- and C-terminal lobes, each containing two calcium-binding EF-hands. Thus, a total of four Ca²⁺ can be bound per CaM (Figure 1) [38–40]. Calcium binding to CaM induces a conformational change, which is unequivocal for activation and modulation of second messenger downstream proteins such as adenylate cyclase, serine/threonine kinases, nitric oxide synthase, serine threonine protein phosphatases (i.e., calcineurin), which are all involved in the transduction of intracellular calcium sig-

nalling (Figure 2). CaM also regulates calcium transport via plasma membrane ATPase, a high-affinity calcium pump that contributes to the maintenance of intracellular calcium homeostasis [45]. CaM also binds to several structural proteins such as spectrin, neuromodulin, and caldesmon [43,44]. These proteins have been shown to play important roles in the maintenance of plasma membrane integrity [46] and cytoskeletal structure [47], postsynaptic function, and in the regulation of smooth muscle cell contraction, respectively [48]. As a primary sensor of intracellular calcium levels, CaM plays a key role in a multitude of processes, such as the regulation of cell cycle, fertilization, intracellular signalling, differentiation, cell death, and cell contraction [39,49].



Figure 1. Schematic representation of Ca^{2+} -CaM activation. After binding 4 ions of Ca^{2+} , calmodulin undergoes conformational change that leads to activation of downstream proteins.

The main function of CaM in the heart is modulation of the action potential, leading to rapid contraction of a distinct group of cardiac cells through the regulation of different channels [22,38], such as the voltage-gated sodium channels [50], the voltage gated potassium channels [51], and the voltage-gated calcium channels [52]. CaM is also involved in the regulation of the sarcoplasmic reticulum calcium release channel (RyR2), the main source of intracellular calcium necessary to induce contraction [53,54]. Regulation of RyRs by CaM is isoform specific. For RyR1, CaM exhibits biphasic regulation that depends on Ca²⁺ concentration. It acts as a weak activator at nanomolar concentrations of Ca²⁺ (apo-CaM) and as an inhibitor at micromolar concentrations of Ca^{2+} (Ca^{2+} -CaM). On the other hand, for RyR2, Ca^{2+} -CaM only inhibits the channel, without activating effects [55]. Moreover, CaM interacts with secondary pathway effectors of cardiomyocyte contraction, such as the beta-adrenergic pathway [56,57] and cyclic nucleotide signalling [38]. In the context of the modulation of cyclic nucleotide signalling by CaM, it is important to consider the role of the phosphodiesterases (PDEs), which are functional antagonists of the adenyly cyclases (ACs) as they degrade cyclic nucleotides and subsequently lower, for example, the cAMP-dependent protein kinase-A (PKA) activity. Among the 21 mammalian genes of PDE, 3 PDE1 gene products (PDE1A–C) appear particularly interesting as PDE1A–C are all CaM regulated and found in heart tissue. The binding of Ca²⁺-CaM to N-terminal sites in PDE1 stimulates their activity and thus increases cell cAMP and cGMP degradation [58]. Degradation of cyclic nucleotides generally signals an easing of cardiomyocyte contractility. Finally, CaM interacts with downstream calcium CaM dependent kinases, supporting calcium recycling in cardiomyocytes necessary to maintain calcium homeostasis in preparation for a new excitation event [38].



Figure 2. The 3D illustrations show the flexibility of calmodulin in binding to 4 Ca^{2+} and undergoing conformational changes for its reaction with a specific downstream protein, such as the CaM kinase proteins. Specifically, the binding of the calcium–calmodulin complex to auto-inhibitory target protein regions leads this conformational change and the target protein activation. Ca²⁺ is displayed as yellow balls and peptide is shown as magenta ribbon.

During evolution, the number of functions for CaM has significantly increased with the need for sequence conservation [59]. In fact, CaM is highly conserved, and it has been shown that most of the human calmodulin mutations are deleterious and associated with life-threatening conditions during early infancy. Defects in CaM function disrupt important calcium signalling events, possibly resulting in fatal heart disease [44,45,59–61] (Table 1). Indeed, deleterious variants in CaM genes (CaM1, CaM2, and CaM3) cause calmodulinopathy, a rare life-threatening arrhythmia syndrome [59,61]. The term specifically refers to a wide spectrum of clinical manifestations [61], such as long-QT syndrome (LQTS) [59,62–65], catecholaminergic polymorphic ventricular tachycardia (CPVT) [66], or idiopathic ventricular fibrillation (IVF) [67]. CaM mutations were also identified in autopsy-negative sudden unexplained deaths (SUD) in young individuals [61,68]. Three heterozygous de novo mutations in either CaM1 or CaM2 were identified that caused an alteration of residues near the calcium binding loops in the calmodulin carboxyl-terminal domain. These alterations were observed in infants who exhibited life-threatening ventricular arrhythmias combined variably with epilepsy and delayed neurodevelopment [59]. Moreover, a common mutation in a highly conserved residue (Phe90) in CaM1 resulted in underlying IVF manifestation in childhood and adolescence [67]. Further, a novel missense mutation in CaM1 was identified in a Moroccan family with a history of ventricular tachycardia and sudden death. This missense mutation was associated with exercise-induced QT prolongation, ventricular tachycardia, and sudden cardiac death during childhood [69].

Gene	Nucleotide Change	Amino Acid Change	Associated Phenotype	Refs.
CALM1	c.389A>G	p.D130G	LQTS	[59,61]
CALM1	c.426C>G	p.F142L	LQTS	[59,61]
CALM1	c.161A>T	p.N54I	CPVT	[61,66]
CALM1	c.293A>G	p.N98S	LQTS, CPVT	[61,66]
CALM1	c.268T>C	p.F90L	IVF, SUD	[67,69]
CALM1	c.395A>T	p.D132V	LQTS	[64]
CALM2	c.293A>G	p.N98S	LQTS, SUD	[61,62,68]
CALM2	c.287A>T	p.D96V	LQTS	[59,61]
CALM2	c.293A>T	p.N98I	LQTS	[61,62]
CALM2	c.400G>C	p.D134H	LQTS	[38,39]
CALM2	c.389A>G	p.D130G	LQTS	[61]
CALM2	c.396T>G	p.D132E	LQTS, CPVT	[61,62]
CALM2	c.394G>C	p.D132H	LQTS	[64]
CALM2	c.407A>C	p.Q136P	LQTS, CPVT	[62]
CALM3	c.389A>G	p.D130G	LQTS	[61,63]
CALM3	c.308C>T	p.A103V	CPVT	[61]
CALM3	c.286G>C	p.D96H	LQTS	[61,65]
CALM3	c.426T>G	p.F142L	LQTS	[61,65]

Table 1. List of the main human CaMs gene polymorphisms. CaM polymorphisms are associated with a spectrum of arrhythmic phenotypes. Adapted from Kotta et al. [61].

In VSMCs, CaM is a critical calcium sensor which regulates different downstream proteins. The Ca²⁺-CaM complex is necessary to phosphorylate and activate myosin light-chain (MLC) actin–myosin interactions and VSMC contractions [70–72]. Specifically, CaMKII phosphorylates and inactivates MLC kinase, a process that may be essential in the regulation of VSM contraction [70]. CaM-calcium binding also controls the interaction between alpha-smooth muscle actin (α SMA) and myosin, affecting VSMC elasticity and adhesion processes [73]. Further, Ca²⁺-CaM activates the serine/threonine (Ser/Thr) phosphatase calcineurin as well as the family of Ca²⁺-CaM-dependent protein kinases, all involved in the regulation of cell cycle progression [70]. The role of CaM in VSMC-driven vascular disease (i.e., atherosclerosis, aneurysms, calcification) has not been studied in detail thus far.

Taken together, these findings highlight the role and function of CaM in the cardiovascular system.

3. Calcium Calmodulin Kinases

The transduction and amplification of intracellular calcium signals sensed by CaM results in regulation of downstream effectors involved in phosphorylation. Specifically, proteins involved in this pathway are of the calcium calmodulin kinase protein family (CaMK), a category of enzymes all classified as Ser/Thr kinases (Figure 3). CaMKs catalyse the transfer of phosphate from the gamma position of ATP to the hydroxyl group of Ser or Thr residues in proteins [39,49,74]. CaMKs can be divided in two different groups. The first group consists of CaMKK1, CAMKK2, CaMKI, CaMKII, and CaMKIV, which are multifunctional proteins. The second group consists of CaMKIII, phosphorylase kinase, and myosin light-chain kinase and are substrate-specific kinases with only one downstream target [74]. All CaMK proteins share a common structure with a bi-lobed catalytic domain followed by a regulatory domain containing both an autoinhibitory as well as a CaMbinding site (Figure 3) [74,75]. In the case of basal intracellular calcium levels, CaMKs are in an inactivated state due to the autoinhibitory domain that interacts and blocks the CaM binding or catalytic site. When intracellular calcium levels increase, CaM binds four Ca²⁺ to become saturated, creating a conformational change. This conformational change results in interaction with the CaM binding domain in CaMKs, resulting in activation. The activation of some CaMKs (i.e., CaMKI) requires both binding of calcium as well as an additional modification, such as phosphorylation. Moreover, the activity of certain

kinases (i.e., CaMKIV and CaMKII) can become independent of CaM, allowing them to be functional beyond the duration of a transient elevation in intracellular calcium [76–78]. These differences in regulation are important for the control of the many cellular functions of CaM.



Figure 3. The general structure shared by the CaM-kinases. The N-terminal catalytic domain is represented in light blue, followed by a regulatory region containing in white the autoinhibitory domain and in dark blue the CaM-binding domain. The black block shows the C-terminal association domain of CaMKII. Adapted from Beghi S, et al., 2020 [39].

Ca²⁺-CaM protein kinases (CaMKs) are involved in several process, controlling several different functions [75,79–93]. In the following sections we describe in more detail the role of multifunctional CAMKs in the heart and vasculature.

3.1. CAMKII

CaMKII is the most abundant calcium calmodulin kinase in the heart [7,94,95]. There are four different isoforms, namely CAMKII α , β , γ , and δ , encoded by four different genes. All four CaMKII isoforms are involved in cellular functions such as calcium homeostasis, membrane potential, the cell cycle, cytoskeletal organization, cell contraction, learning, memory, and gene expression and secretion [74,78,95]. It is important to note that the four isoforms of CaMKII have a different tissue distribution [96]. CaMKII γ and δ are mainly present in cardiac tissue whereas CaMKII α and β are expressed mainly in the neuronal system [7,57,97]. Specifically, CaMKII δ is the predominant isoform in the heart and plays a crucial role in the regulation of several cardiac functions. CaMKII δ is involved in the integration of calcium signalling through a variety of pathways to maintain cardiac homeostasis as it is predominantly involved in the excitation coupling (ECC) in the heart [38,78,94,97,98].

The effect of CaMKII on ECC is crucial because it phosphorylates calcium handling proteins including sarcoplasmic reticulum (SR) calcium release channels, such as ryanodine receptors (RyR) [99], phospholamban (PLB) [94], and L-type calcium channels [100]. These downstream proteins of CaMKII are responsible for the regulation of cellular calcium influx, calcium release from the SR, as well as calcium uptake into the SR [7]. The phosphorylation of myofilament proteins (cMyBP-C: cardiac myosin binding protein-C) and myosin regulatory light-chain 2 by CaMKII contributes to myofilament calcium sensitization and interactions [101,102]. Tong et al. showed that the phosphorylation of cMyBP-C due to CaMKIIS and PKA is a principal mediator of increased contractility observed with β -adrenergic stimulation or increased pacing. Moreover, its phosphorylation increases the force and kinetics of twitches in living cardiac muscle [101]. CaMKII also phosphorylates titin, which stabilizes the contractile machinery, causing a lower cardiomyocyte passive force likely associated with improved diastolic filling [103–105]. Meanwhile, the phosphorylation by CaMKII of troponin I enacts the calcium desensitization; this process is required to accelerate diastolic myofilament relaxation with increased heart rate [105,106]. Additionally, CaMKII targets and activates cardiac Na^+ [107,108] and K⁺ channels [54,109–111], which are involved in the regulation of calcium homeostasis, through phosphorylation.

The activation of CaMKII depends on Ca²⁺-CaM binding, subsequently activating all 12 subunits in separate holoenzymes (a biochemically active compound consisting of an

enzyme with its cofactor) forming the dodecameric structure of CaMKII [112]. Additionally, Ca²⁺-CaM bound to CaMKII ensures that one subunit is auto-phosphorylated at Thr286 by a neighbouring activated CaMKII subunit. This event generates autonomous activity, increasing the affinity of CaMKII for Ca²⁺-CaM some 1000-fold, an event known as "CaM-trapping" [74,77,78].

This autonomous activity of CaMKII can also be mediated by other post-translational modifications such as oxidation, nitrosylation, and glycosylation [105] The altered hyperactivate regulation of CaMKII is increased in myocardial disease, contributing to apoptosis, arrhythmias [113], defective ECC, and ETC (excitation–transcription coupling) [53,114,115], favouring pathological hypertrophy [96] and contractile dysfunction specifically during heart failure (HF) [54,96,114,115]. CaMKII is a pro-arrhythmogenic protein increased during CVD, i.e., myocardial injury [116], atrial fibrillation [117], cardiac hypertrophy, and ischaemia/reperfusion injury [118], and inhibition of CaMKII might thus be a treatment option for cardiac pathologies [7,119,120].

Additionally, in blood vessels CaMKII plays a crucial role. In VSMCs, CaMKII is activated by Ca²⁺-CaM to regulate activation of myosin light-chain kinase (MLCK) by phosphorylation [70]. In VSMCs, CaMKII inhibits CREB (cAMP response element-binding protein) through different phosphorylation sites. CREB is an important transcription factor regulating expression of contractile and proliferative and migratory genes during VSMC phenotype switching. The rise of intracellular Ca²⁺ in VSMCs promotes nuclear translocation of Ca²⁺-CaMKII. γ CaMKII acts as a carrier of Ca²⁺/CaM, transporting it into the nucleus. In the nucleus Ca²⁺/CaM activates CaMKK and its substrate CaMKIV, which in turn activates CREB by phosphorylation [34,121]. Moreover, it has been shown that Ca²⁺-CaMKII and Erk1/2 activation play a crucial role in the regulation of VSMC proliferation by α -adrenergic receptor agonists [122]. Finally, it was recently shown that CAMKII δ and calponin 3 play critical roles in circRNA CDR1as/miR-7-5p-induced human pulmonary VSMC calcification under hypoxic conditions, which may contribute to pulmonary hypertension [123].

3.2. Ca²⁺-CaM-Dependent Kinase Cascade

3.2.1. CaMKK Family

The Ca²⁺-CaM-dependent kinase cascade consists of Ca²⁺-CaM kinase-kinase family (CaMKK1 and CaMKK2), CaMKI, and CaMKIV (Figure 4). This cascade is involved in different processes, such as cell proliferation, apoptosis, immune cell function, stem cell maintenance, as well as glucose homeostasis [7]. Dysregulation of kinases in the Ca²⁺-CaM-dependent kinase cascade is associated with a variety of diseases such as cancer, obesity, diabetes, neuronal defects, and CVD [12,124].

CaMKKs support transduction of calcium to other downstream kinases such as CaMKI and CaMKIV [32,62]. They are present both in the cytoplasm as well as in the nucleus of the cell. There are two isoforms of CaMKKs encoded by the genes CAMKK1 and CAMKK2 [39,74,125]. Both CaMKKs share a high sequence homology and the same common domain structure typical for all kinase proteins (Figure 3) [125,126]. The main difference between both CaMKKs is that CaMKK1 is kept in an inactive state until binding of Ca²⁺-CaM relieves the autoinhibitory mechanism, whilst autophosphorylation allows a partially autonomous activity of CaMKK2 in the absence of Ca²⁺-CaM [74,75,126]. Both kinases can be partially inhibited by PKA that phosphorylates CaMKKs at different Ser or Thr residues within the CaM kinase binding domain and within the ATP binding region in the catalytic domain. Specifically, when PKA phosphorylates CaMKK1 at Ser458, it prevents binding to Ca²⁺-CaM, thereby inhibiting CaMKK1 from responding to increased intracellular calcium levels [75,127]. Moreover, the principal site of autophosphorylation for CaMKK2 is Thr482, which generates a partial autonomous activity independent of Ca^{2+} -CaM, disrupting the autoinhibitory mechanism of CaMKK2. Since CaMKK2 is not dependent on rapid fluxes of intracellular calcium for basal activity, it can respond to other stimuli for longer durations [7,74,75]. The temporal sequence of these two distinct



signalling events, PKA phosphorylation on CaMKK and its ability to block Ca²⁺-CaM binding, plays an important role in directing signalling through CaMKK1.

Figure 4. Ca²⁺-CaM kinases downstream of calmodulin. Binding of calcium to calmodulin leads to activation of the Ca²⁺-CaM-dependent kinase cascade, consisting of CaMKK1 and CaMKK2, CaMKI, and CaMKIV, as well as activation of CaMKII. The activation of CaMKKs regulates crucial cellular functions. Calcium calmodulin mediates downstream signalling of CaMKI, CaMKIV, AMPK, Akt, and mTOR.

A common characteristic of both CaMKKs is that they phosphorylate CaMKI and CaMKIV at residues Thr177 and Thr196, respectively [39,74,75]. In addition, both CAMKKs activate AMP-activated protein kinase (AMPK), a key regulator of cellular energy balance [128] (Figure 4). Sallé-Lefort et al. showed that the CaMKK/AMPK complex is involved in the activation of hypoxia-inducible factor-1 α (HIF-1 α). HIF-1 α controls metastasis-associated lung adenocarcinoma transcript 1 (Malat1), enhancing its transcription upon low oxygen conditions. Their findings suggest that Malat1 expression is upregulated under hypoxic conditions by the CaMKK/AMPK/HIF-1 α axis [129]. Another downstream substrate of CaMKKs is AKT (also known as protein kinase B or PKB), an important oncology target. Activated AKT promotes phosphorylation of cellular substrates, e.g., TSC1/2, FOXO, GSK3, and mdm2 [130], thereby controlling cell proliferation, metabolism, cell growth, and survival [75,131] (Figure 3).

Below we discuss the roles of CaMKK1, CaMKK2, CaMKI, and CaMKIV in CVD in more detail.

CAMKK1

The main role of CaMKK1 is phosphorylation of downstream CaMKI and CaMKIV, whereas the activation of CaMKK1 fully depends on CaM [39,74,75,126]. Recently, the

crystal structure of CaMKK1 was revealed using two ATP-competitive inhibitors. The structural differences between CaMKK1 and CaMKK2 therefore can result in the generation of CaMKK-specific inhibitors [126]. Research supports the importance of CaMKKs in cardiovascular biology [126,132] via mTOR (mechanistic target of rapamycin) [133]. The mTOR pathway plays a key regulatory function in cardiovascular physiology and pathology (Figure 4) [134]. More recently it was shown that overexpression of CaMKK1 regulates the mesenchymal stem cell (MSC) secretome [135]. The overexpression of CAMKK1 in either MSC or upon direct injection of its encoding DNA into infarcted tissue resulted in improved cardiac function and increased vasculogenesis. In rats, injections in the heart of conditioned media from MSCs overexpressing CaMKK1 showed improvement in cardiac function after acute myocardial infarction, with increased vascular density and decreased scar tissue [135]. Moreover, the direct overexpression of CaMKK1 in infarcted tissue using a CaMKK1-encoding plasmid also significantly improved ejection fraction and decreased infarct size after acute myocardial infarction. Although the precise mechanism by which CAMKK1 affects regeneration is not known, these data put forward a novel role of CaMKK1 derived from the MSC secretome, indicating a potential therapeutic target for infarcted heart tissue [135].

Recently, the genetic variant rs7214723 of CaMKK1 was shown to be associated with a higher risk of developing CVD [136]. This genetic variant, which was shown to be associated with increased lung cancer risk [137,138], is a single-nucleotide polymorphism (SNP) that causes an amino acid change from glutamic acid (E) to glycine (G) at position 375 inside the catalytic domain of CaMKK1. This variation creates a charge change influencing substrate specificity, thereby inhibiting downstream CaMKI and CaMKIV [39,137,138]. This polymorphism appears to be highly represented in the population (MAF index: C = 0.3954 (1980/5008 1000 Genome)) [139]. Results from a cross-sectional study conducted on 300 cardiac patients showed a statistical difference between cardiopathic patients and European reference populations between the genotype and allele frequencies for rs7214723. These data suggest a potential role of rs7214723 that could be used as genetic biomarker in predicting CVD susceptibility [136].

Whilst interest in the function of CaMKK1 in the heart is growing, there is still lack of information regarding the role of CaMKK1 in vasculature and related vascular diseases.

CAMKK2

More research has been published on CAMKK2 compared to CAMKK1. Specifically, the interaction between CaMKK2 and AMP-activated protein kinase (AMPK) is well studied as it regulates many physiological processes such as hypothalamic control of feeding behaviour, hepatic gluconeogenesis, adipocyte differentiation and macroautophagy [128,131,140]. AMPK is involved in a variety of cellular processes, including sensing intracellular ATP levels, regulation of autophagy and mitochondrial fission [141]. Since AMPK activity depends on CaMKK2, CaMKK2 has a possible role in cardiac energy production to protect the heart against calcium overload induced by sustained pressure loads [38]. Moreover, AMPK signalling is repressed during cardiac hypertrophy, and cardiac-specific knockout of AMPK promotes stress-induced cardiac hypertrophy [38,142]. Thus, the inactivation of CaMKK2 might indirectly result in the development of metabolic dysfunction and cardiac hypertrophy (Figure 4). Histone demethylase JMJD1C, an important epigenetic factor, represses the activation of AMPK during cardiac hypertrophy through the reduction of CaMKK2 expression. This confirms that AMPK signalling is involved in JMJD1C-mediated cardiac hypertrophy [143]. Absence of functional CaMKK2 in mice resulted in increased left ventricular dilatation and dysfunction and subsequent mortality [144]. This demonstrates that CaMKK2 can exert beneficial effects against pressureoverload-induced heart failure, thereby providing a therapeutic target for treatment of heart failure [144]. Additionally, CaMKK2 plays an important role in GLUT4 translocation through AMPK activation in cardiomyocytes. GLUT4, after metabolic stress, translocates from intracellular vesicles to the plasma membrane protecting cardiac myocytes from

oxidative stress and ischaemic injury. During ischaemia, GLUT4 translocation represents a peculiar mechanism by which the heart increases glucose uptake increasing cell energy status. The pan-CaMKK inhibitor STO-609 as well as overexpression of a dominant-negative form of CaMKK2 in cardiomyocytes inhibited hydrogen-peroxide-mediated translocation of GLUT4 [143–145] (Figure 4). Further, CaMKK2 is involved in CaMKK2-AMPK-VASP/MLC signalling for migration of cells and for assembly of contractile actin stress fibres [146]. Additionally, it was shown that the regulation of mechanosensing is promoted by myosin-18 [135]. Exerted cellular forces are transduced via the cytoskeleton and are therefore of key importance in mechanosensing. Stress fibres are composed of actin (microfilaments) and non-muscle myosin II. Myosin-18B co-localizes with myosin II motor domains in these stress fibres. Moreover, myosin-18B knockout cells displayed the ability to exert forces to the environment, hence their involvement and importance in mechanostansduction [147,148].

Diabetes is an important risk factor for the development of CVD. In type 2 diabetic mice, endocrine hormone fibroblast growth factor 21 (FGF21) activates CaMKK2/AMPK α , thereby suppressing oxidative stress and enhancing endothelial nitric oxide synthase (eNOS) signalling, improving vessel relaxation [149]. This suggests the possible use of FGF21 to activate CaMKK2/AMPK α for the apeutic use of endothelial dysfunction in diabetes. Additionally, store-operated calcium entry (SOCE) is a central mechanism in cellular calcium signalling and in maintaining cellular calcium balance. Intracellular Ca²⁺ flux, driven by release of Ca²⁺ from intracellular stores can activate CaMKK2 via Ca²⁺/CaM [150]. This signalling pathway of SOCE-Ca²⁺/CaM-CAMKK2 is important for the regulation of autophagy and in maintaining proliferation and promoting the survival capability of endothelial progenitor cells (EPCs) [151]. This pathway is active at basal levels in cells of cardiovascular origin as an important homeostatic mechanism [152]. CaMKK2 activation and mTOR deactivation is associated with autophagy modulation (Figure 4). Here, a novel signalling pathway of SOCE-CAMKK2 in the regulation of autophagy was identified, revealing new insights in maintaining proliferation and survival capability of endothelial progenitor cells (EPCs). This offers ways to improve EPCs transplantation efficacy to enhance vascular re-endothelialization in patients with hypercholesterolaemia [151]. Moreover, tetrahydrobiopterin (BH4), a multifunctional cofactor implicated in regulation of nervous, immune, and cardiovascular systems, is a new potential endogenous activator of CaMKK2. BH4 targets CaMKK2 and promotes recovery of mitochondria in diabetic cardiomyopathy [153].

AMPK, among others downstream to CaMKK2, is a master sensor of cellular energy status involved in the progression of vascular calcification [154]. It is known that AMPK activators are associated with reduced calcification deposits [154], indicating a potential therapeutic role of AMPK in vascular calcification [155]. Vascular calcification is in part driven by the trans-differentiation of VSMC into cells with osteoblast characteristics, such as increased alkaline phosphatase activity and collagen II deposition, governed by the upregulation of Runt-related transcription factor 2 [156]. A recent study in mice showed that exogenous omentin-1 attenuates osteogenic differentiation of VSMCs through the activation of AMPK/Akt signalling [157]. On the contrary, the inhibition of AMPK and Akt signalling reverses the anti-calcific effect induced by omentin-1 both in vitro and in vivo [157]. Moreover, Lai and colleagues demonstrated that during calcification KMUP-3 (the xanthine derivative 7-[2-[4-(4-nitrobenzene)-piperazinyl]ethyl]–1,3-dimethylxanthine) inhibits both mTOR, downstream to CaMKK2, and β -catenin upregulation, essential for VSMC osteogenic phenotypic switching as well as enhancing AMPK activation inhibiting VSMC osteogenic differentiation [158].

3.2.2. CaMKI and CaMKIV

Ca²⁺-CaM dependent kinase I and IV (CaMKI and CaMKIV) share the same common kinase structure (Figure 3), and their activation requires both binding of Ca²⁺-CaM and subsequent phosphorylation by Ca²⁺-CaM kinase proteins (CaMKKs) at Thr 177 and Thr 196, respectively [7,74,75]. The difference in activation between CaMKI and CaMKIV

is that CaMKI remains entirely Ca²⁺-CaM-dependent, whereas CaMKIV can undergo an intra-subunit autophosphorylation of the Ser/Thr-rich N-terminus, generating Ca²⁺-CaM independent activity. This allows CaMKIV to maintain its functionality beyond increased intracellular calcium [39,74]. CaMKI is a monomeric kinase that is expressed by three different genes encoding α , β , and γ isoforms [159,160]. CaMKI is a cytosolic protein, which serves several functions including transcription activator activity, cell cycle control, hormone production, cell differentiation, actin filament organization, and neurite outgrowth (Figure 4). However, the precise role of CaMKI in CVD is not completely understood [39].

CaMKIV is a monomeric kinase and can be expressed as two isoforms (α and β), encoded by a single gene [161]. CaMKIV is considered important in the nervous system [162,163], but recently it received attention for its role in cardiovascular pathophysiology [164]. Several studies showed an association of the genetic variant rs10491334 in CaMKIV with elevated diastolic blood pressure [162–164] (Figure 4). Furthermore, rs10491334 was also associated with a reduction in the expression level of CaMKIV in hypertensive patients [164]. Moreover, CaMKK/CaMKIV has been shown to be a key endogenous protective pathway in ischaemia and an important regulator of blood–brain barrier integrity [165]. The deletion of CaMKK or CaMKIV in mice exacerbates stroke outcome, such as infarct volume, oedema formation and behavioural deficits. Liu et al. described furthermore that the CaMKK pathway is involved in the immune response to brain injury, increased blood brain barrier impairment, transcriptional inactivation of cAMP, and inflammatory responses in females after stroke. The CaMKK signalling pathway might therefore be a potential target for stroke treatment [166].

In the vasculature, it has been shown that the interaction between CaMKKs and CaMKIV is important in the regulation of CREB during phenotype switching of VSMCs [34] (Figure 4). Indeed, nuclear CaMKIV plays a crucial role in the activation of CREB, whereas CaMKII inhibits CREB by phosphorylating it [167,168]. Previous studies demonstrated that both CaMKII and CaMKIV can phosphorylate Ser¹³³ of CREB. However, CaMKII also phosphorylates Ser¹⁴² in the transcriptional activation domain of CREB. This event has been reported to inhibit CREB activity by interfering with CREB dimerization and protein interactions to form an active promoter complex [169,170]. It has been shown that VSMC phenotype depends on the activity and form of different calcium-dependent transcription factors, such as CREB [34,167]. Part of the VSMC phenotype regulation is therefore under the control of the Ca²⁺-CaM kinase pathway, thus making this an interesting therapeutic option to regulate VSMC phenotype switching.

4. Conclusions

In this review, we have summarized the current state of the literature on Ca²⁺-CaM -dependent kinase cascade functions, with a focus on the heart and vessels (Table 2). Our review clearly points towards an important role for calmodulin and calcium kinase proteins in CVD. CaMKKs are involved in regulating calcium signalling in cardiovascular metabolism resulting in calcium-induced autophagy, an important homeostatic mechanism active at basal levels in cells of cardiovascular origin [151,152]. Moreover, the CaMKK/CaMKIV phosphorylation cascade has been shown to be a key endogenous protective mechanism in ischemia and an important regulator of blood-brain barrier integrity [165]. In addition, genetic variants in Ca²⁺-CaM dependent kinases were associated with CVD [39,136,162,164]. Further, it is important to underline the role of Ca²⁺-CaM-dependent kinases in VSMC integrity. Whilst it is known that the interaction CaMKK2/CaMKIV is involved in phenotype switching of VSMCs [34], data are lacking as to the role of CaM and CAMKK1 in vessel wall homeostasis. As the Ca^{2+} -CaM-dependent kinase cascade regulates many cellular processes in both cardiac and vascular tissue, further investigation of this signalling pathway is necessary to understand this process and to open up novel avenues for the treatment of CVD.

Kinase	In Heart Pathology	In Vasculature Pathology	
CaM	Polymorphisms linked to calmodulinopathy, arrhythmia [44,45,59–69].	Unknown	
CaMKII	Implicated in myocardial injury [116], atrial fibrillation [117], cardiac hypertrophy, ischaemia/reperfusion injury [118], heart failures, contributing to apoptosis, arrhythmias [113], defective ECC and ETC [53,114,115], pathological hypertrophy [96] and contractile dysfunction during heart failure [54,96,114,115].	Regulation of VSMCs phenotype switching through the inhibition of CREB [34].	
CaMKK1	Polymorphism linked to the higher risk to develop CVD [136].	Regulation of VSMCs phenotype switching (CaMKK-CaMKIV-CREB) [34].	
CaMKK2	Inactivation of CaMKK2 indirectly results in the development of metabolic dysfunction and cardiac hypertrophy [143].	Regulation of VSMCs phenotype switching (CaMKK-CaMKIV-CREB) [34].	
САМКІ	Unknown	Unknown	
CAMKIV	Polymorphisms linked to elevated diastolic blood pressure [162–164].	Regulation of VSMCs phenotype switching (CaMKK-CaMKIV-CREB) [34].	

Table 2. Summary of the involvement of the CaM cascade in cardiovascular disease.

Author Contributions: Conceptualization, S.B., A.B. and L.J.S.; writing—original draft preparation, S.B., M.F., A.J., R.V., N.R., E.B., A.B. and L.J.S.; writing—review and editing, S.B., M.F., N.R., E.B., A.B. and L.J.S.; visualization of the figures and tables, S.B., M.F. and K.W.; supervision, L.J.S.; project administration, L.J.S.; funding acquisition, L.J.S. All authors have read and agreed to the published version of the manuscript.

Funding: This work was in part funded by the European Union's Horizon 2020 research and innovation programmes under the Marie Sklodowska-Curie grant agreement No 764474.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data are available upon reasonable request from the corresponding author.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Poulter, N. Coronary Heart Disease Is a Multifactorial Disease. Am. J. Hypertens. 1999, 12, 92S–95S. [CrossRef]
- Dhingra, R.; Vasan, R.S. Biomarkers in Cardiovascular Disease: Statistical Assessment and Section on Key Novel Heart Failure Biomarkers. *Trends Cardiovasc. Med.* 2017, 27, 123–133. [CrossRef] [PubMed]
- Soler-Botija, C.; Gálvez-Montón, C.; Bayés-Genís, A. Epigenetic Biomarkers in Cardiovascular Diseases. Front. Genet. 2019, 10, 950. [CrossRef] [PubMed]
- 4. Ortaliza, J.; Orgera, K.; Amin, K.; Cox, C. COVID-19 Continues to Be a Leading Cause of Death in the U.S. in June 2021. Available online: https://www.healthsystemtracker.org (accessed on 27 August 2021).
- 5. Libby, P. The Changing Landscape of Atherosclerosis. *Nature* 2021, 592, 524–533. [CrossRef] [PubMed]
- 6. Tokumitsu, H.; Soderling, T.R. Requirements for Calcium and Calmodulin in the Calmodulin Kinase Activation Cascade. J. Biol. Chem. **1996**, 271, 5617–5622. [CrossRef] [PubMed]
- Junho, C.V.C.; Caio-Silva, W.; Trentin-Sonoda, M.; Carneiro-Ramos, M.S. An Overview of the Role of Calcium/Calmodulin-Dependent Protein Kinase in Cardiorenal Syndrome. *Front. Physiol.* 2020, 11, 735. [CrossRef]
- Yu, X.; Murao, K.; Sayo, Y.; Imachi, H.; Cao, W.M.; Ohtsuka, S.; Niimi, M.; Tokumitsu, H.; Inuzuka, H.; Wong, N.C.W.; et al. The Role of Calcium/Calmodulin-Dependent Protein Kinase Cascade in Glucose Upregulation of Insulin Gene Expression. *Diabetes* 2004, 53, 1475–1481. [CrossRef]
- 9. Soderling, T.R. The Ca²⁺–Calmodulin-Dependent Protein Kinase Cascade. Trends Biochem. Sci. 1999, 24, 232–236. [CrossRef]
- 10. Kitsos, C.M.; Sankar, U.; Illario, M.; Colomer-Font, J.M.; Duncan, A.W.; Ribar, T.J.; Reya, T.; Means, A.R. Calmodulin-Dependent Protein Kinase IV Regulates Hematopoietic Stem Cell Maintenance. *J. Biol. Chem.* **2005**, *280*, 33101–33108. [CrossRef]

- Racioppi, L.; Lento, W.; Huang, W.; Arvai, S.; Doan, P.L.; Harris, J.R.; Marcon, F.; Nakaya, H.I.; Liu, Y.; Chao, N. Calcium/Calmodulin-Dependent Kinase Kinase 2 Regulates Hematopoietic Stem and Progenitor Cell Regeneration. *Cell Death Dis.* 2017, *8*, e3076. [CrossRef]
- Colomer, J.; Means, A.R. Physiological Roles of the Ca²⁺/CaM-Dependent Protein Kinase Cascade in Health and Disease. *Calcium Signal. Dis.* 2007, 45, 169–214. [CrossRef]
- 13. Durham, A.L.; Speer, M.Y.; Scatena, M.; Giachelli, C.M.; Shanahan, C.M. Role of Smooth Muscle Cells in Vascular Calcification: Implications in Atherosclerosis and Arterial Stiffness. *Cardiovasc. Res.* **2018**, *114*, 590–600. [CrossRef] [PubMed]
- 14. Jaminon, A.; Reesink, K.; Kroon, A.; Schurgers, L. The Role of Vascular Smooth Muscle Cells in Arterial Remodeling: Focus on Calcification-Related Processes. *Int. J. Mol. Sci.* 2019, 20, 5694. [CrossRef] [PubMed]
- 15. Proudfoot, D. Molecular Mechanisms of Arterial Calcification. Artery Res. 2009, 3, 1423–1430. [CrossRef]
- 16. Lee, S.J.; Lee, I.K.; Jeon, J.H. Vascular Calcification-New Insights into Its Mechanism. Int. J. Mol. Sci. 2020, 21, 2685. [CrossRef]
- Krohn, J.B.; Hutcheson, J.D.; Martínez-Martínez, E.; Aikawa, E. Extracellular Vesicles in Cardiovascular Calcification: Expanding Current Paradigms. J. Physiol. 2016, 594, 2895–2903. [CrossRef]
- Christensen, J.L.; Tan, S.; Chung, H.E.; Ghosalkar, D.S.; Qureshi, R.; Chu, A.; Yu, W.; Berus, J.; Shah, N.R.; Wu, W.C.; et al. Aortic Valve Calcification Predicts All-Cause Mortality Independent of Coronary Calcification and Severe Stenosis. *Atherosclerosis* 2020, 307, 16–20. [CrossRef]
- Veulemans, V.; Piayda, K.; Maier, O.; Bosbach, G.; Polzin, A.; Hellhammer, K.; Afzal, S.; Klein, K.; Dannenberg, L.; Zako, S.; et al. Aortic Valve Calcification Is Subject to Aortic Stenosis Severity and the Underlying Flow Pattern. *Heart Vessel.* 2021, 36, 242–251. [CrossRef]
- Di Lullo, L.; Tripepi, G.; Ronco, C.; D'Arrigo, G.; Barbera, V.; Russo, D.; di Iorio, B.R.; Uguccioni, M.; Paoletti, E.; Ravera, M.; et al. Cardiac Valve Calcification and Use of Anticoagulants: Preliminary Observation of a Potentially Modifiable Risk Factor. *Int. J. Cardiol.* 2019, 278, 243–249. [CrossRef]
- 21. Terrar, D.A. Calcium Signaling in the Heart. In *Advances in Experimental Medicine and Biology;* Springer: Berlin/Heidelberg, Germany, 2020; Volume 1131. [CrossRef]
- Fearnley, C.J.; Llewelyn Roderick, H.; Bootman, M.D. Calcium Signaling in Cardiac Myocytes. *Cold Spring Harb. Perspect Biol.* 2011, 3, a004242. [CrossRef]
- Reid, I.R.; Birstow, S.M.; Bolland, M. Calcium and Cardiovascular Disease. *Endocrinol. Metab.* 2017, 32, 339–349. [CrossRef] [PubMed]
- 24. Clapham, D.E. Calcium Signaling. Cell 2007, 131, 1047–1058. [CrossRef] [PubMed]
- 25. Schreckenberg, R.; Schlüter, K.-D. Calcium sensing receptor expression and signalling in cardiovascular physiology and disease. *Vasc. Pharmacol.* **2018**, *107*, 35–42. [CrossRef]
- Sundararaman, S.S.; van der Vorst, E.P.C. Calcium-Sensing Receptor (CaSR), Its Impact on Inflammation and the Consequences on Cardiovascular Health. Int. J. Mol. Sci. 2021, 22, 2478. [CrossRef] [PubMed]
- Alfadda, T.I.; Saleh, A.M.A.; Houillier, P.; Geibel, J.P. Calcium-sensing receptor 20 years later. Am. J. Physiol. Cell Physiol. 2014, 307, C221–C231. [CrossRef] [PubMed]
- Raffaello, A.; Mammucari, C.; Gherardi, G.; Rizzuto, R. Calcium at the Center of Cell Signaling: Interplay between Endoplasmic Reticulum, Mitochondria, and Lysosomes. *Trends Biochem. Sci.* 2016, *41*, 1035–1049. [CrossRef]
- Zhang, C.; Miller, C.L.; Gorkhali, R.; Zou, J.; Huang, K.; Brown, E.M.; Yang, J.J. Molecular Basis of the Extracellular Ligands Mediated Signaling by the Calcium Sensing Receptor. *Front. Physiol.* 2016, *7*, 441. [CrossRef]
- Bagur, R.; Hajnóczky, G. Intracellular Ca²⁺ Sensing: Its Role in Calcium Homeostasis and Signaling. *Mol. Cell* 2017, 66, 780–788. [CrossRef]
- 31. Cooper, D.; Dimri, M. Biochemistry, Calcium Channels; StatPearls: Treasure Island, FL, USA, 2022; PMID: 32965869.
- Muszbek, L.; Bereczky, Z.; Bagoly, Z.; Komáromi, I.; Katona, É. Factor XIII: A Coagulation Factor with Multiple Plasmatic and Cellular Functions. *Physiol. Rev.* 2011, 91, 931–972. [CrossRef]
- Singh, S.; Dodt, J.; Volkers, P.; Hethershaw, E.; Philippou, H.; Ivaskevicius, V.; Imhof, D.; Oldenburg, J.; Biswas, A. Structure functional insights into calcium binding during the activation of coagulation factor XIII A. *Sci. Rep.* 2019, 9, 11324. [CrossRef]
- Matchkov, V.V.; Kudryavtseva, O.; Aalkjaer, C. Intracellular Ca²⁺ Signalling and Phenotype of Vascular Smooth Muscle Cells. Basic Clin. Pharmacol. Toxicol. 2012, 110, 42–48. [CrossRef] [PubMed]
- Petsophonsakul, P.; Furmanik, M.; Forsythe, R.; Dweck, M.; Schurink, G.W.; Natour, E.; Reutelingsperger, C.; Jacobs, M.; Mees, B.; Schurgers, L. Role of Vascular Smooth Muscle Cell Phenotypic Switching and Calcification in Aortic Aneurysm Formation Involvement of Vitamin K-Dependent Processes. *Arter. Thromb. Vasc. Biol.* 2019, *39*, 1351–1368. [CrossRef] [PubMed]
- Furmanik, M.; Chatrou, M.; van Gorp, R.; Akbulut, A.; Willems, B.; Schmidt, H.; van Eys, G.; Bochaton-Piallat, M.-L.; Proudfoot, D.; Biessen, E.; et al. Reactive Oxygen-Forming Nox5 Links Vascular Smooth Muscle Cell Phenotypic Switching and Extracellular Vesicle-Mediated Vascular Calcification. *Circ. Res.* 2020, 127, 911–927. [CrossRef] [PubMed]
- Yang, W.; Zou, B.; Hou, Y.; Yan, W.; Chen, T.; Qu, S. Extracellular vesicles in vascular calcification. *Clin. Chim. Acta* 2019, 499, 118–122. [CrossRef] [PubMed]
- 38. Sorensen, A.B.; Søndergaard, M.T.; Overgaard, M.T. Calmodulin in a Heartbeat. FEBS J. 2013, 280, 5511–5532. [CrossRef]
- Beghi, S.; Cavaliere, F.; Buschini, A. Gene polymorphisms in calcium-calmodulin pathway: Focus on cardiovascular disease. Mutat. Res. Mutat. Res. Rev. 2020, 786, 108325. [CrossRef]

- Babu, Y.S.; Sack, J.S.; Greenhough, T.J.; Bugg, C.E.; Means, A.R.; Cook, W.J. Three-dimensional structure of calmodulin. *Nature* 1985, 315, 37–40. [CrossRef]
- 41. Finn, B.; Forsén, S. The evolving model of calmodulin structure, function and activation. Structure 1995, 3, 7–11. [CrossRef]
- 42. Ye, Q.; Wei, Y.; Fischer, R.; Borner, C.; Berchtold, M.W. Expression of calmodulin and calmodulin binding proteins in rat fibroblasts stably transfected with protein kinase C and oncogenes. *Biochim. Biophys. Acta Mol. Cell Res.* **1997**, 1359, 89–96. [CrossRef]
- 43. Halling, D.B.; Liebeskind, B.J.; Hall, A.W.; Aldrich, R.W. Conserved properties of individual Ca²⁺ -binding sites in calmodulin. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, E1216–E1225. [CrossRef]
- 44. Friedberg, F.; Rhoads, A.R.; Friedberg, A.R.R.F. Evolutionary Aspects of Calmodulin. *IUBMB Life* 2001, *51*, 215–221. [CrossRef] [PubMed]
- 45. Brini, M. Plasma membrane Ca²⁺-ATPase: From a housekeeping function to a versatile signaling role. *Pflugers Arch.* **2009**, 457, 657–664. [CrossRef] [PubMed]
- Zhang, R.; Zhang, C.; Zhao, Q.; Li, D. Spectrin: Structure, function and disease. Sci. China Life Sci. 2013, 56, 1076–1085. [CrossRef] [PubMed]
- 47. Hartl, M.; Schneider, R. A Unique Family of Neuronal Signaling Proteins Implicated in Oncogenesis and Tumor Suppression. *Front. Oncol.* **2019**, *9*, 289. [CrossRef] [PubMed]
- 48. Brozovich, F.; Nicholson, C.; Degen, C.; Gao, Y.Z.; Aggarwal, M.; Morgan, K. Mechanisms of Vascular Smooth Muscle Contraction and the Basis for Pharmacologic Treatment of Smooth Muscle Disorders. *Pharmacol. Rev.* **2016**, *68*, 476–532. [CrossRef]
- 49. Means, A.R.; VanBerkum, M.F.; Bagchi, I.; Lu, K.P.; Rasmussen, C.D. Regulatory functions of calmodulin. *Pharmacol. Ther.* **1991**, 50, 255–270. [CrossRef]
- 50. Abriel, H. Cardiac sodium channel Nav1.5 and interacting proteins: Physiology and pathophysiology. *J. Mol. Cell. Cardiol.* 2010, 48, 2–11. [CrossRef]
- 51. Kang, P.W.; Westerlund, A.M.; Shi, J.; White, K.M.F.; Dou, A.K.; Cui, A.H.; Silva, J.R.; Delemotte, L.; Cui, J. Calmodulin acts as a state-dependent switch to control a cardiac potassium channel opening. *Sci. Adv.* **2020**, *6*, eabd6798. [CrossRef]
- Muir, W.W. Cardiovascular Physiology. In Veterinary Anesthesia and Analgesia: The Fifth Edition of Lumb and Jones; Wiley: Hoboken, NJ, USA, 2017; pp. 415–472. [CrossRef]
- 53. Yang, D.; Song, L.-S.; Zhu, W.-Z.; Chakir, K.; Wang, W.; Wu, C.; Wang, Y.; Xiao, R.-P.; Chen, S.R.W.; Cheng, H. Calmodulin Regulation of Excitation-Contraction Coupling in Cardiac Myocytes. *Circ. Res.* **2003**, *92*, 659–667. [CrossRef]
- 54. Pitt, G.S. Calmodulin and CaMKII as molecular switches for cardiac ion channels: Fig. 1. *Cardiovasc. Res.* **2007**, *73*, 641–647. [CrossRef]
- 55. Ogawa, H.; Kurebayashi, N.; Yamazawa, T.; Murayama, T. Regulatory mechanisms of ryanodine receptor/Ca²⁺ release channel revealed by recent advancements in structural studies. *J. Muscle Res. Cell Motil.* **2021**, *42*, 291–304. [CrossRef] [PubMed]
- 56. Bers, D.M. Cardiac excitation-contraction coupling. Nature 2002, 415, 198–205. [CrossRef] [PubMed]
- 57. Wang, W.; Zhu, W.; Wang, S.; Yang, D.; Crow, M.T.; Xiao, R.-P.; Cheng, H. Sustained B1 -Adrenergic Stimulation Modulates Cardiac Contractility by Ca²⁺ /Calmodulin Kinase Signaling Pathway. *Circ. Res.* **2004**, *95*, 798–806. [CrossRef] [PubMed]
- 58. Omori, K.; Kotera, J. Overview of PDEs and Their Regulation. Circ. Res. 2007, 100, 309–327. [CrossRef] [PubMed]
- Crotti, L.; Johnson, C.N.; Graf, E.; de Ferrari, G.M.; Cuneo, B.F.; Ovadia, M.; Papagiannis, J.; Feldkamp, M.D.; Rathi, S.G.; Kunic, J.D.; et al. Calmodulin Mutations Associated with Recurrent Cardiac Arrest in Infants. *Circulation* 2013, 127, 1009–1017. [CrossRef]
- 60. Baba, M.L.; Goodman, M.; Berger-Cohn, J.; Demaille, J.G.; Matsuda, G. The early adaptive evolution of calmodulin. *Mol. Biol. Evol.* **1984**, *1*, 442–455. [CrossRef]
- 61. Kotta, M.-C.; Sala, L.; Ghidoni, A.; Badone, B.; Ronchi, C.; Parati, G.; Zaza, A.; Crotti, L. Calmodulinopathy: A Novel, Life-Threatening Clinical Entity Affecting the Young. *Front. Cardiovasc. Med.* **2018**, *5*, 175. [CrossRef]
- Makita, N.; Yagihara, N.; Crotti, L.; Johnson, C.N.; Beckmann, B.-M.; Roh, M.S.; Shigemizu, D.; Lichtner, P.; Ishikawa, T.; Aiba, T.; et al. Novel Calmodulin Mutations Associated with Congenital Arrhythmia Susceptibility. *Circ. Cardiovasc. Genet.* 2014, 7, 466–474. [CrossRef]
- Reed, G.J.; Boczek, N.J.; Etheridge, S.P.; Ackerman, M.J. CALM3 mutation associated with long QT syndrome. *Heart Rhythm.* 2015, 12, 419–422. [CrossRef]
- 64. Pipilas, D.C.; Johnson, C.N.; Webster, G.; Schlaepfer, J.; Fellmann, F.; Sekarski, N.; Wren, L.M.; Ogorodnik, K.V.; Chazin, D.M.; Chazin, W.J.; et al. Novel calmodulin mutations associated with congenital long QT syndrome affect calcium current in human cardiomyocytes. *Heart Rhythm.* **2016**, *13*, 2012–2019. [CrossRef]
- 65. Chaix, M.-A.; Koopmann, T.T.; Goyette, P.; Alikashani, A.; Latour, F.; Fatah, M.; Hamilton, R.M.; Rioux, J.D. Novel CALM3 mutations in pediatric long QT syndrome patients support a CALM3 -specific calmodulinopathy. *Heart Rhythm Case Rep.* **2016**, *2*, 250–254. [CrossRef] [PubMed]
- Nyegaard, M.; Overgaard, M.T.; Søndergaard, M.T.; Vranas, M.; Behr, E.R.; Hildebrandt, L.L.; Lund, J.; Hedley, P.L.; Camm, A.J.; Wettrell, G.; et al. Mutations in Calmodulin Cause Ventricular Tachycardia and Sudden Cardiac Death. *Am. J. Hum. Genet.* 2012, 91, 703–712. [CrossRef] [PubMed]
- Marsman, R.F.; Barc, J.; Beekman, L.; Alders, M.; Dooijes, D.; van den Wijngaard, A.; Ratbi, I.; Sefiani, A.; Bhuiyan, Z.A.; Wilde, A.A.; et al. A Mutation in CALM1 Encoding Calmodulin in Familial Idiopathic Ventricular Fibrillation in Childhood and Adolescence. J. Am. Coll. Cardiol. 2014, 63, 259–266. [CrossRef] [PubMed]

- 68. Anderson, J.; Tester, D.J.; Will, M.L.; Ackerman, M.J. Whole-Exome Molecular Autopsy After Exertion-Related Sudden Unexplained Death in the Young. *Circ. Cardiovasc. Genet.* **2016**, *9*, 259–265. [CrossRef] [PubMed]
- 69. Marsman, R.F.; Barc, J.G.; Beekman, L.; Bhuijan, Z.A.; Wilde, A.A.M.; Bezzina, C.R. A mutation in CALM1 encoding calmodulin causes sudden cardiac death in childhood and adolescence. *Eur. Heart J.* **2013**, *34*, P2298. [CrossRef]
- Koledova, V.V.; Khalil, R.A. Ca²⁺, Calmodulin, and Cyclins in Vascular Smooth Muscle Cell Cycle. *Circ. Res.* 2006, 98, 1240–1243. [CrossRef]
- 71. Hughes, A. Calcium Channels in Vascular Smooth Muscle Cells. J. Vasc. Res. 1995, 32, 353–370. [CrossRef]
- 72. Clunn, G.F.; Sever, P.S.; Hughes, A.D. Calcium channel regulation in vascular smooth muscle cells: Synergistic effects of statins and calcium channel blockers. *Int. J. Cardiol.* 2010, 139, 2–6. [CrossRef]
- 73. Zhu, Y.; Qu, J.; He, L.; Zhang, F.; Zhou, Z.; Yang, S.; Zhou, Y. Calcium in Vascular Smooth Muscle Cell Elasticity and Adhesion: Novel Insights into the Mechanism of Action. *Front. Physiol.* **2019**, *10*, 852. [CrossRef]
- 74. Swulius, M.T.; Waxham, M.N. Ca²⁺/Calmodulin-dependent Protein Kinases. Cell. Mol. Life Sci. 2008, 65, 2637–2657. [CrossRef]
- Brzozowski, J.S.; Skelding, K.A. The Multi-Functional Calcium/Calmodulin Stimulated Protein Kinase (CaMK) Family: Emerging Targets for Anti-Cancer Therapeutic Intervention. *Pharmaceuticals* 2019, 12, 8. [CrossRef] [PubMed]
- Westphal, R.S.; Anderson, K.A.; Means, A.R.; Wadzinski, B.E. A Signaling Complex of Ca²⁺-Calmodulin-Dependent Protein Kinase IV and Protein Phosphatase 2A. *Science* 1998, 280, 1258–1261. [CrossRef] [PubMed]
- Meyer, T.; Hanson, P.I.; Stryer, L.; Schulman, H. Calmodulin Trapping by Calcium-Calmodulin-Dependent Protein Kinase. *Science* 1992, 256, 1199–1202. [CrossRef] [PubMed]
- Beckendorf, J.; van den Hoogenhof, M.M.G.; Backs, J. Physiological and unappreciated roles of CaMKII in the heart. *Basic Res. Cardiol.* 2018, 113, 29. [CrossRef]
- Joseph, J.D.; Means, A.R. Identification and Characterization of Two Ca²⁺/CaM-dependent Protein Kinases Required for Normal Nuclear Division in Aspergillus nidulans. *J. Biol. Chem.* 2000, 275, 38230–38238. [CrossRef]
- Skelding, K.A.; Rostas, J.A.P.; Verrills, N.M. Controlling the cell cycle: The role of calcium/calmodulin-stimulated protein kinases I and II. Cell Cycle 2011, 10, 631–639. [CrossRef]
- Wayman, G.A.; Kaech, S.; Grant, W.F.; Davare, M.; Impey, S.; Tokumitsu, H.; Nozaki, N.; Banker, G.; Soderling, T.R. Regulation of Axonal Extension and Growth Cone Motility by Calmodulin-Dependent Protein Kinase I. J. Neurosci. 2004, 24, 3786–3794. [CrossRef]
- 82. Condon, J.C.; Pezzi, V.; Drummond, B.M.; Yin, S.; Rainey, W.E. Calmodulin-Dependent Kinase I Regulates Adrenal Cell Expression of Aldosterone Synthase. *Endocrinology* **2002**, *143*, 3651–3657. [CrossRef]
- 83. Schmitt, J.M.; Guire, E.S.; Saneyoshi, T.; Soderling, T.R. Calmodulin-Dependent Kinase Kinase/Calmodulin Kinase I Activity Gates Extracellular-Regulated Kinase-Dependent Long-Term Potentiation. *J. Neurosci.* 2005, 25, 1281–1290. [CrossRef]
- Ang, E.S.M.; Zhang, P.; Steer, J.H.; Tan, J.W.-Y.; Yip, K.; Zheng, M.H.; Joyce, D.A.; Xu, J. Calcium/calmodulin-dependent kinase activity is required for efficient induction of osteoclast differentiation and bone resorption by receptor activator of nuclear factor kappa B ligand (RANKL). J. Cell. Physiol. 2007, 212, 787–795. [CrossRef]
- Soderling, T.R.; Derkach, V.A. Postsynaptic protein phosphorylation and LTP. *Trends Neurosci.* 2000, 23, 75–80. [CrossRef] [PubMed]
- Von Hertzen, L.S.J.; Giese, K.P. Alpha-isoform of Ca²⁺/calmodulin-dependent kinase II autophosphorylation is required for memory consolidation-specific transcription. *NeuroReport* 2005, *16*, 1411–1414. [CrossRef] [PubMed]
- Wang, Z.-W. Regulation of Synaptic Transmission by Presynaptic CaMKII and BK Channels. *Mol. Neurobiol.* 2008, 38, 153–166.
 [CrossRef] [PubMed]
- 88. Fink, C.C.; Bayer, K.-U.; Myers, J.W.; Ferrell, J.E.; Schulman, H.; Meyer, T. Selective Regulation of Neurite Extension and Synapse Formation by the β but not the α Isoform of CaMKII. *Neuron* 2003, *39*, 283–297. [CrossRef] [PubMed]
- Rostas, J.A.P.; Spratt, N.J.; Hoffman, A.; Murtha, L.A.; Pepperall, D.; McLeod, D.D.; Dickson, P.W.; Skelding, K.A. The Role of Ca²⁺-Calmodulin Stimulated Protein Kinase II in Ischaemic Stroke—A Potential Target for Neuroprotective Therapies. *Neurochem. Int.* 2017, 107, 33–42. [CrossRef] [PubMed]
- Rostas, J.A.P.; Hoffman, A.; Murtha, L.A.; Pepperall, D.; McLeod, D.D.; Dickson, P.W.; Spratt, N.J.; Skelding, K.A. Ischaemia- and excitotoxicity-induced CaMKII-Mediated neuronal cell death: The relative roles of CaMKII autophosphorylation at T286 and T253. *Neurochem. Int.* 2017, 104, 6–10. [CrossRef]
- Liu, W.; Xu, C.; Ran, D.; Wang, Y.; Zhao, H.; Gu, J.; Liu, X.; Bian, J.; Yuan, Y.; Liu, Z. CaMKII mediates cadmium induced apoptosis in rat primary osteoblasts through MAPK activation and endoplasmic reticulum stress. *Toxicology* 2018, 406–407, 70–80. [CrossRef] [PubMed]
- Shin, M.K.; Kim, M.-K.; Bae, Y.-S.; Jo, I.; Lee, S.-J.; Chung, C.-P.; Park, Y.-J.; Min, D.S. A novel collagen-binding peptide promotes osteogenic differentiation via Ca²⁺/calmodulin-dependent protein kinase II/ERK/AP-1 signaling pathway in human bone marrow-derived mesenchymal stem cells. *Cell. Signal.* 2008, 20, 613–624. [CrossRef] [PubMed]
- Takemura, M.; Mishima, T.; Wang, Y.; Kasahara, J.; Fukunaga, K.; Ohashi, K.; Mizuno, K. Ca²⁺/Calmodulin-dependent Protein Kinase IV-mediated LIM Kinase Activation Is Critical for Calcium Signal-induced Neurite Outgrowth. *J. Biol. Chem.* 2009, 284, 28554–28562. [CrossRef]
- 94. Mattiazzi, A.; Kranias, E.G. The role of CaMKII regulation of phospholamban activity in heart disease. *Front. Pharmacol.* **2014**, *5*, 5. [CrossRef]

- 95. Luczak, E.D.; Anderson, M.E. CaMKII oxidative activation and the pathogenesis of cardiac disease. *J. Mol. Cell. Cardiol.* **2014**, *73*, 112–116. [CrossRef] [PubMed]
- 96. Anderson, M.E.; Brown, J.H.; Bers, D.M. CaMKII in myocardial hypertrophy and heart failure. *J. Mol. Cell. Cardiol.* 2011, 51, 468–473. [CrossRef] [PubMed]
- Maier, L.S.; Bers, D.M. Role of Ca²⁺/calmodulin-dependent protein kinase (CaMK) in excitation–contraction coupling in the heart. *Cardiovasc. Res.* 2007, 73, 631–640. [CrossRef] [PubMed]
- 98. Lars, S.M. Role of CaMKII for signaling and regulation in the heart. Front. Biosci. 2009, 14, 486–496. [CrossRef]
- Chen, H.H.; Zhang, C.; Lyu, R. The Role of RyR2 Phosphorylation in Heart Failure. *Fudan Univ. J. Med. Sci.* 2017, 44. [CrossRef]
 Bers, D.M.; Morotti, S. Ca²⁺ current facilitation is CaMKII-dependent and has arrhythmogenic consequences. *Front. Pharmacol.* 2014, 5, 144. [CrossRef]
- Tong, C.W.; Wu, X.; Liu, Y.; Rosas, P.C.; Sadayappan, S.; Hudmon, A.; Muthuchamy, M.; Powers, P.A.; Valdivia, H.H.; Moss, R.L. Phosphoregulation of Cardiac Inotropy via Myosin Binding Protein-C During Increased Pacing Frequency or B₁-Adrenergic Stimulation. *Circ. Heart Fail.* 2015, *8*, 595–604. [CrossRef]
- Eikemo, H.; Moltzau, L.R.; Hussain, R.I.; Nguyen, C.H.; Qvigstad, E.; Levy, F.O.; Skomedal, T.; Osnes, J.-B. CaMKII in addition to MLCK contributes to phosphorylation of regulatory light chain in cardiomyocytes. *Biochem. Biophys. Res. Commun.* 2016, 471, 219–225. [CrossRef]
- 103. Perkin, J.; Slater, R.; Del Favero, G.; Lanzicher, T.; Hidalgo, C.; Anderson, B.; Smith, J.E.; Sbaizero, O.; Labeit, S.; Granzier, H. Phosphorylating Titin's Cardiac N2B Element by ERK2 or CaMKIIδ Lowers the Single Molecule and Cardiac Muscle Force. *Biophys. J.* 2015, 109, 2592–2601. [CrossRef]
- 104. Hamdani, N.; Krysiak, J.; Kreusser, M.M.; Neef, S.; dos Remedios, C.G.; Maier, L.S.; Krüger, M.; Backs, J.; Linke, W.A. Crucial Role for Ca²⁺ /Calmodulin-Dependent Protein Kinase-II in Regulating Diastolic Stress of Normal and Failing Hearts via Titin Phosphorylation. *Circ. Res.* 2013, 112, 664–674. [CrossRef]
- 105. Bussey, C.; Erickson, J. Physiology and pathology of cardiac CaMKII. Curr. Opin. Physiol. 2018, 1, 52-58. [CrossRef]
- 106. Guilbert, A.; Lim, H.J.; Cheng, J.; Wang, Y. CaMKII-dependent myofilament Ca²⁺ desensitization contributes to the frequencydependent acceleration of relaxation. *Cell Calcium* 2015, *58*, 489–499. [CrossRef] [PubMed]
- 107. Ashpole, N.M.; Herren, A.W.; Ginsburg, K.S.; Brogan, J.D.; Johnson, D.E.; Cummins, T.R.; Bers, D.M.; Hudmon, A. Ca²⁺/Calmodulin-dependent Protein Kinase II (CaMKII) Regulates Cardiac Sodium Channel NaV1.5 Gating by Multiple Phosphorylation Sites. J. Biol. Chem. 2012, 287, 19856–19869. [CrossRef] [PubMed]
- Maier, L.S. CaMKII regulation of voltage-gated sodium channels and cell excitability. *Heart Rhythm.* 2011, 8, 474–477. [CrossRef]
 [PubMed]
- 109. Mustroph, J.; Maier, L.S.; Wagner, S. CaMKII regulation of cardiac K channels. Front. Pharmacol. 2014, 5, 20. [CrossRef]
- Alday, A.; Ahyayauch, H.; Fernández-López, V.; Echeazarra, L.; Urrutia, J.; Casis, O.; Gallego, M. CaMKII Modulates the Cardiac Transient Outward K+ Current through its Association with Kv4 Channels in Non-Caveolar Membrane Rafts. *Cell. Physiol. Biochem.* 2020, 54, 27–39. [CrossRef]
- 111. Bers, D.M.; Grandi, E. CaMKII Regulation of Cardiac Ion Channels. J. Cardiovasc. Pharmacol. 2009, 54, 180–187. [CrossRef]
- 112. Myers, J.B.; Zaegel, V.; Coultrap, S.J.; Miller, A.P.; Bayer, K.U.; Reichow, S.L. The CaMKII holoenzyme structure in activationcompetent conformations. *Nat. Commun.* 2017, *8*, 15742. [CrossRef]
- 113. Wu, Y.; Temple, J.; Zhang, R.; Dzhura, I.; Zhang, W.; Trimble, R.; Roden, D.M.; Passier, R.; Olson, E.N.; Colbran, R.J.; et al. Calmodulin Kinase II and Arrhythmias in a Mouse Model of Cardiac Hypertrophy. *Circulation* **2002**, *106*, 1288–1293. [CrossRef]
- 114. Li, L.; Satoh, H.; Ginsburg, K.S.; Bers, D.M. The effect of Ca²⁺-calmodulin-dependent protein kinase II on cardiac excitationcontraction coupling in ferret ventricular myocytes. *J. Physiol.* **1997**, *501*, 17–31. [CrossRef]
- 115. Bers, D.M. Calcium Cycling and Signaling in Cardiac Myocytes. Annu. Rev. Physiol. 2008, 70, 23–49. [CrossRef] [PubMed]
- Ren, G.; Dewald, O.; Frangogiannis, N.G. Inflammatory mechanisms in myocardial infarction. *Curr. Drug Targets Inflamm. Allergy* 2003, 2, 242–256. [CrossRef] [PubMed]
- 117. Liu, Z.; Finet, J.E.; Wolfram, J.A.; Anderson, M.E.; Ai, X.; Donahue, J.K. Calcium/calmodulin-dependent protein kinase II causes atrial structural remodeling associated with atrial fibrillation and heart failure. *Heart Rhythm.* 2019, 16, 1080–1088. [CrossRef] [PubMed]
- 118. Rajtik, T.; Carnicka, S.; Szobi, A.; Giricz, Z.; O-Uchi, J.; Hassova, V.; Svec, P.; Ferdinandy, P.; Ravingerova, T.; Adameova, A. Oxidative activation of CaMKIIδ in acute myocardial ischemia/reperfusion injury: A role of angiotensin AT1 receptor-NOX2 signaling axis. *Eur. J. Pharmacol.* 2016, 771, 114–122. [CrossRef]
- Cipolletta, E.; Rusciano, M.R.; Maione, A.S.; Santulli, G.; Sorriento, D.; Del Giudice, C.; Ciccarelli, M.; Franco, A.; Crola, C.; Campiglia, P.; et al. Targeting the CaMKII/ERK Interaction in the Heart Prevents Cardiac Hypertrophy. *PLoS ONE* 2015, 10, e0130477. [CrossRef]
- Rokita, A.G.; Anderson, M.E. New Therapeutic Targets in Cardiology: Arrhythmias and Ca²⁺/Calmodulin- Dependent Kinase II (CaMKII). *Circulation* 2012, 126, 2125–2139. [CrossRef]
- 121. Ma, H.; Groth, R.D.; Cohen, S.M.; Emery, J.F.; Li, B.; Hoedt, E.; Zhang, G.; Neubert, T.A.; Tsien, R.W. ΓCaMKII Shuttles Ca²⁺/CaM to the Nucleus to Trigger CREB Phosphorylation and Gene Expression. *Cell* **2014**, *159*, 281–294. [CrossRef]

- 122. Cipolletta, E.; Monaco, S.; Maione, A.S.; Vitiello, L.; Campiglia, P.; Pastore, L.; Franchini, C.; Novellino, E.; Limongelli, V.; Bayer, K.U.; et al. Calmodulin-Dependent Kinase II Mediates Vascular Smooth Muscle Cell Proliferation and Is Potentiated by Extracellular Signal Regulated Kinase. *Endocrinology* 2010, 151, 2747–2759. [CrossRef]
- 123. Ma, C.; Gu, R.; Wang, X.; He, S.; Bai, J.; Zhang, L.; Zhang, J.; Li, Q.; Qu, L.; Xin, W.; et al. circRNA CDR1as Promotes Pulmonary Artery Smooth Muscle Cell Calcification by Upregulating CAMK2D and CNN3 via Sponging miR-7-5p. *Mol. Ther. Nucleic Acids* 2020, 22, 530–541. [CrossRef]
- Skelding, K.A.; Rostas, J.A.P. Regulation of Multifunctional Calcium/Calmodulin Stimulated Protein Kinases by Molecular Targeting. Adv. Exp. Med. Biol. 2020, 1131, 649–679. [CrossRef]
- Tokumitsu, H.; Wayman, G.A.; Muramatsu, M.; Soderling, T.R. Calcium/Calmodulin-Dependent Protein Kinase Kinase: Identification of Regulatory Domains. *Biochemistry* 1997, 36, 12823–12827. [CrossRef] [PubMed]
- 126. Santiago, A.D.S.; Couñago, R.M.; Ramos, P.Z.; Godoi, P.H.C.; Massirer, K.B.; Gileadi, O.; Elkins, J.M. Structural Analysis of Inhibitor Binding to CAMKK1 Identifies Features Necessary for Design of Specific Inhibitors. *Sci. Rep.* 2018, *8*, 14800. [CrossRef] [PubMed]
- 127. Wayman, G.A.; Tokumitsu, H.; Soderling, T.R. Inhibitory Cross-talk by cAMP Kinase on the Calmodulin-dependent Protein Kinase Cascade. J. Biol. Chem. 1997, 272, 16073–16076. [CrossRef] [PubMed]
- Racioppi, L.; Means, A.R. Calcium/Calmodulin-dependent Protein Kinase Kinase 2: Roles in Signaling and Pathophysiology. J. Biol. Chem. 2012, 287, 31658–31665. [CrossRef] [PubMed]
- Sallé-Lefort, S.; Miard, S.; Nolin, M.-A.; Boivin, L.; Paré, M.-È.; Debigaré, R.; Picard, F. Hypoxia upregulates Malat1 expression through a CaMKK/AMPK/HIF-1α axis. Int. J. Oncol. 2016, 49, 1731–1736. [CrossRef]
- 130. He, Y.; Sun, M.M.; Zhang, G.G.; Yang, J.; Chen, K.S.; Xu, W.W.; Li, B. Targeting PI3K/Akt signal transduction for cancer therapy. *Signal Transduct. Target. Ther.* **2021**, *6*, 425. [CrossRef]
- O'Byrne, S.N.; Scott, J.W.; Pilotte, J.R.; da Santiago, A.S.; Langendorf, C.G.; Oakhill, J.S.; Eduful, B.J.; Couñago, R.M.; Wells, C.I.; Zuercher, W.J.; et al. In Depth Analysis of Kinase Cross Screening Data to Identify CAMKK2 Inhibitory Scaffolds. *Molecules* 2020, 25, 325. [CrossRef]
- 132. Takemoto-Kimura, S.; Suzuki, K.; Horigane, S.-I.; Kamijo, S.; Inoue, M.; Sakamoto, M.; Fujii, H.; Bito, H. Calmodulin kinases: Essential regulators in health and disease. *J. Neurochem.* **2017**, *141*, 808–818. [CrossRef]
- 133. Ferey, J.L.A.; Brault, J.J.; Smith, C.A.S.; Witczak, C.A. Constitutive activation of CaMKKα signaling is sufficient but not necessary for mTORC1 activation and growth in mouse skeletal muscle. Am. J. Physiol. Endocrinol. Metab. 2014, 307, E686–E694. [CrossRef]
- 134. Sciarretta, S.; Forte, M.; Frati, G.; Sadoshima, J. New Insights into the Role of mTOR Signaling in the Cardiovascular System. *Circ. Res.* **2018**, *122*, 489–505. [CrossRef]
- Dong, F.; Patnaik, S.; Duan, Z.-H.; Kiedrowski, M.; Penn, M.S.; Mayorga, M.E. A Novel Role for CAMKK1 in the Regulation of the Mesenchymal Stem Cell Secretome. *Stem Cells Transl. Med.* 2017, *6*, 1759–1766. [CrossRef] [PubMed]
- 136. Beghi, S.; Cavaliere, F.; Manfredini, M.; Ferrarese, S.; Corazzari, C.; Beghi, C.; Buschini, A. Polymorphism rs7214723 in CAMKK1: A new genetic variant associated with cardiovascular diseases. *Biosci. Rep.* **2021**, *41*, BSR20210326. [CrossRef] [PubMed]
- 137. Zhang, Y.-H.; Xu, Q.; Zhao, Z.; Wu, J.; Liu, W.-X.; Wang, H.; Jin, L.; Wang, J.-C. Polymorphism rs7214723 in CAMKK1 and lung cancer risk in Chinese population. *Tumor Biol.* 2013, 34, 3147–3152. [CrossRef]
- 138. Chen, D.; Zhong, F.; Chen, Y. Association of calcium/calmodulin-dependent protein kinase kinase1 rs7214723 polymorphism with lung cancer risk in a Chinese population. *Biosci. Rep.* 2017, 37, BSR2017076. [CrossRef]
- 139. Auton, A.; Abecasis, G.R.; Altshuler, D.M.; Durbin, R.M.; Bentley, D.R.; Chakravarti, A.; Clark, A.G.; Donnelly, P.; Eichler, E.E.; Flicek, P.; et al. A Global Reference for Human Genetic Variation. *Nature* **2015**, *526*, 68–74. [CrossRef]
- Hurley, R.L.; Anderson, K.A.; Franzone, J.M.; Kemp, B.E.; Means, A.R.; Witters, L.A. The Ca²⁺/Calmodulin-dependent Protein Kinase Kinases Are AMP-activated Protein Kinase Kinases. J. Biol. Chem. 2005, 280, 29060–29066. [CrossRef]
- Herzig, S.; Shaw, R.J. AMPK: Guardian of metabolism and mitochondrial homeostasis. *Nat. Rev. Mol. Cell Biol.* 2018, 19, 121–135. [CrossRef] [PubMed]
- Salt, I.; Hardie, D.G. AMP-Activated Protein Kinase: An Ubiquitous Signaling Pathway with Key Roles in the Cardiovascular System. Circ. Res. 2017, 120, 1825–1841. [CrossRef] [PubMed]
- 143. Yu, S.; Li, Y.; Zhao, H.; Wang, Q.; Chen, P. The Histone Demethylase JMJD1C Regulates CAMKK2-AMPK Signaling to Participate in Cardiac Hypertrophy. *Front. Physiol.* **2020**, *11*, 539. [CrossRef]
- 144. Watanabe, S.; Horie, T.; Nagao, K.; Kuwabara, Y.; Baba, O.; Nishi, H.; Sowa, N.; Narazaki, M.; Matsuda, T.; Takemura, G.; et al. Cardiac-Specific Inhibition of Kinase Activity in Calcium/Calmodulin-Dependent Protein Kinase Kinase-β Leads to Accelerated Left Ventricular Remodeling and Heart Failure after Transverse Aortic Constriction in Mice. *PLoS ONE* 2014, 9, e108201. [CrossRef]
- 145. Horie, T.; Ono, K.; Nagao, K.; Nishi, H.; Kinoshita, M.; Kawamura, T.; Wada, H.; Shimatsu, A.; Kita, T.; Hasegawa, K. Oxidative stress induces GLUT4 translocation by activation of PI3-K/Akt and dual AMPK kinase in cardiac myocytes. *J. Cell. Physiol.* 2008, 215, 733–742. [CrossRef] [PubMed]
- Tojkander, S.; Ciuba, K.; Lappalainen, P. CaMKK2 Regulates Mechanosensitive Assembly of Contractile Actin Stress Fibers. *Cell Rep.* 2018, 24, 11–19. [CrossRef] [PubMed]
- 147. Zhao, S.; Shi, X.; Zhang, Y.; Wen, Z.; Cai, J.; Gao, W.; Xu, J.; Zheng, Y.; Ji, B.; Cui, Y.; et al. Myosin-18B Promotes Mechanosensitive CaMKK2-AMPK-VASP Regulation of Contractile Actin Stress Fibers. *iScience* **2020**, *23*, 100975. [CrossRef] [PubMed]

- 148. Jiu, Y.; Kumari, R.; Fenix, A.M.; Schaible, N.; Liu, X.; Varjosalo, M.; Krishnan, R.; Burnette, D.T.; Lappalainen, P. Myosin-18B Promotes the Assembly of Myosin II Stacks for Maturation of Contractile Actomyosin Bundles. *Curr. Biol.* 2019, 29, 81–92.e5. [CrossRef]
- 149. Ying, L.; Li, N.; He, Z.; Zeng, X.; Nan, Y.; Chen, J.; Miao, P.; Ying, Y.; Lin, W.; Zhao, X.; et al. Fibroblast growth factor 21 Ameliorates diabetes-induced endothelial dysfunction in mouse aorta via activation of the CaMKK2/AMPKα signaling pathway. *Cell Death Dis.* **2019**, *10*, 665. [CrossRef]
- 150. Jardin, I.; Lopez, J.J.; Sanchez-Collado, J.; Gomez, L.J.; Salido, G.M.; Rosado, J.A. Store-Operated Calcium Entry and Its Implications in Cancer Stem Cells. *Cells* **2022**, *11*, 1332. [CrossRef]
- 151. Yang, J.; Yu, J.; Li, D.; Yu, S.; Ke, J.; Wang, L.; Wang, Y.; Qiu, Y.; Gao, X.; Zhang, J.; et al. Store-operated calcium entryactivated autophagy protects EPC proliferation via the CAMKK2-MTOR pathway in ox-LDL exposure. *Autophagy* 2017, 13, 82–98. [CrossRef]
- 152. Bravo-San Pedro, J.M.; Kroemer, G.; Galluzzi, L. Autophagy and Mitophagy in Cardiovascular Disease. *Circ. Res.* 2017, 120, 1812–1824. [CrossRef]
- 153. Kim, H.K.; Ko, T.H.; Song, I.-S.; Jeong, Y.J.; Heo, H.J.; Jeong, S.H.; Kim, M.; Park, N.M.; Seo, D.Y.; Kha, P.T.; et al. BH4 activates CaMKK2 and rescues the cardiomyopathic phenotype in rodent models of diabetes. *Life Sci. Alliance* 2020, *3*, e201900619. [CrossRef]
- Zhan, J.-K.; Wang, Y.-J.; Wang, Y.; Tang, Z.-Y.; Tan, P.; Huang, W.; Liu, Y.-S. Adiponectin attenuates the osteoblastic differentiation of vascular smooth muscle cells through the AMPK/mTOR pathway. *Exp. Cell Res.* 2014, 323, 352–358. [CrossRef]
- 155. Lu, Y.; Yuan, T.; Min, X.; Yuan, Z.; Cai, Z. AMPK: Potential Therapeutic Target for Vascular Calcification. *Front. Cardiovasc. Med.* (*IF 3.915*) **2021**, *8*, 670222. [CrossRef] [PubMed]
- Tyson, J.; Bundy, K.; Roach, C.; Douglas, H.; Ventura, V.; Segars, M.; Schwartz, O.; Simpson, C. Mechanisms of the Osteogenic Switch of Smooth Muscle Cells in Vascular Calcification: WNT Signaling, BMPs, Mechanotransduction, and EndMT. *Bioengineering* 2020, 7, 88. [CrossRef] [PubMed]
- 157. Xu, F.; Li, F.-X.; Lin, X.; Zhong, J.-Y.; Wu, F.; Shan, S.-K.; Tan, C.-M.; Yuan, L.-Q.; Liao, X.-B. Adipose tissue-derived omentin-1 attenuates arterial calcification via AMPK/Akt signaling pathway. *Aging* **2019**, *11*, 8760–8776. [CrossRef] [PubMed]
- 158. Lai, C.-H.; Chang, C.-W.; Lee, F.-T.; Kuo, C.-H.; Hsu, J.-H.; Liu, C.-P.; Wu, H.-L.; Yeh, J.-L. Targeting vascular smooth muscle cell dysfunction with xanthine derivative KMUP-3 inhibits abdominal aortic aneurysm in mice. *Atherosclerosis* 2020, 297, 16–24. [CrossRef]
- Haribabu, B.; Hook, S.S.; Selbert, M.A.; Goldstein, E.G.; Tomhave, E.D.; Edelman, A.M.; Snyderman, R.; Means, A.R. Human calcium-calmodulin dependent protein kinase I: cDNA cloning, domain structure and activation by phosphorylation at threonine-177 by calcium-calmodulin dependent protein kinase I kinase. *EMBO J.* 1995, 14, 3679–3686. [CrossRef]
- Zha, M.; Zhong, C.; Ou, Y.; Han, L.; Wang, J.; Ding, J. Crystal Structures of Human CaMKIα Reveal Insights into the Regulation Mechanism of CaMKI. *PLoS ONE* 2012, 7, e44828. [CrossRef]
- 161. Bleier, J.; Toliver, A. Exploring the Role of CaMKIV in Homeostatic Plasticity. J. Neurosci. 2017, 37, 11520–11522. [CrossRef]
- 162. Chen, S.; Wang, Z.; Zhang, L.; Lü, G.; Zhou, C.; Wang, D.; Tang, Z.; Wang, L.; Qin, L.; Zhai, Z. CAMK4 gene variation is associated with hypertension in a Uygur population. *Genet. Mol. Res.* **2016**, *15*, gmr.15017207. [CrossRef]
- Levy, D.; Larson, M.G.; Benjamin, E.J.; Newton-Cheh, C.; Wang, T.J.; Hwang, S.-J.; Vasan, R.S.; Mitchell, G.F. Framingham Heart Study 100K Project: Genome-wide associations for blood pressure and arterial stiffness. *BMC Med Genet.* 2007, *8*, S3–S11. [CrossRef]
- 164. Santulli, G.; Cipolletta, E.; Sorriento, D.; Del Giudice, C.; Anastasio, A.; Monaco, S.; Maione, A.S.; Condorelli, G.; Puca, A.; Trimarco, B.; et al. CaMK4 Gene Deletion Induces Hypertension. *J. Am. Heart Assoc.* **2012**, *1*, e001081. [CrossRef]
- 165. McCullough, L.D.; Tarabishy, S.; Liu, L.; Benashski, S.; Xu, Y.; Ribar, T.; Means, A.; Li, J. Inhibition of Calcium/Calmodulin-Dependent Protein Kinase Kinase β and Calcium/Calmodulin-Dependent Protein Kinase IV Is Detrimental in Cerebral Ischemia. *Stroke* 2013, 44, 2559–2566. [CrossRef] [PubMed]
- 166. Liu, L.; McCullough, L.; Li, J. Genetic deletion of calcium/calmodulin-dependent protein kinase kinase β (CaMKK β) or CaMK IV exacerbates stroke outcomes in ovariectomized (OVXed) female mice. *BMC Neurosci.* **2014**, *15*, 118. [CrossRef] [PubMed]
- 167. Wamhoff, B.R.; Bowles, D.; Owens, G.K. Excitation–Transcription Coupling in Arterial Smooth Muscle. *Circ. Res.* 2006, *98*, 868–878. [CrossRef] [PubMed]
- Najwer, I.; Lilly, B. Ca²⁺/calmodulin-dependent protein kinase IV activates cysteine-rich protein 1 through adjacent CRE and CArG elements. *Am. J. Physiol. Cell Physiol.* 2005, 289, C785–C793. [CrossRef] [PubMed]
- Sun, P.; Enslen, H.; Myung, P.S.; Maurer, R.A. Differential activation of CREB by Ca²⁺/calmodulin-dependent protein kinases type II and type IV involves phosphorylation of a site that negatively regulates activity. *Genes Dev.* 1994, *8*, 2527–2539. [CrossRef]
- 170. Liu, Y.; Sun, L.-Y.; Singer, D.V.; Ginnan, R.; Singer, H.A. CaMKIIδ-dependent Inhibition of cAMP-response Element-binding Protein Activity in Vascular Smooth Muscle. J. Biol. Chem. 2013, 288, 33519–33529. [CrossRef]