

Supplementary Materials

SAXS Examinations of the Redox-Dependent Formation of a DNA-SOD1 Complex

Huiling Wang, Mingfang Wang, Zefeng Nie, Shuang Qiu, Xiaoping Huang, Xiang Li, Yanfang Cui, Chunrong Liu and Changlin Liu *

Key Laboratory of Pesticide and Chemical Biology of Ministry of Education, School of Chemistry, Central China Normal University, Wuhan 430079, China

* Correspondence: liuchl@mail.ccnu.edu.cn

Supplementary Figures

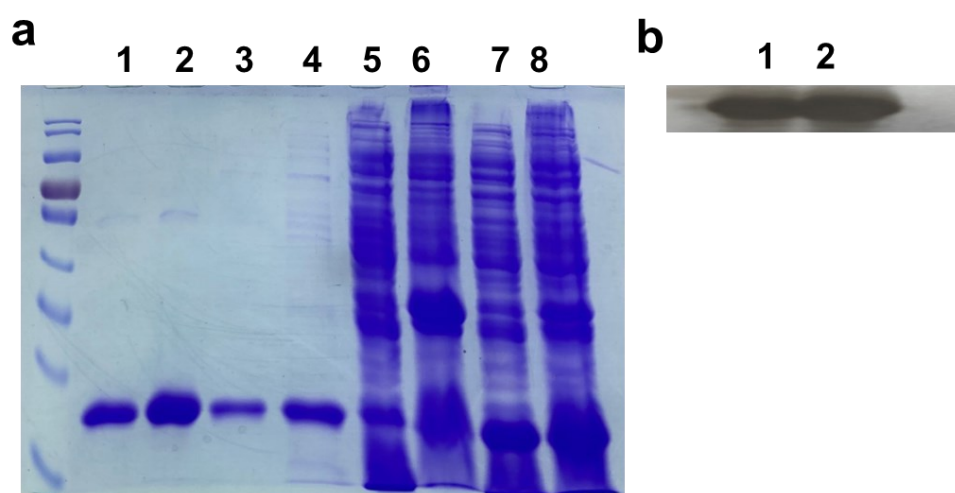


Figure S1. SDS-PAGE and western blot assay of recombinant human SOD1. (a) SDS-PAGE of human SOD1, line 1: elution buffer average, line 2: elution buffer max, line 3: wash buffer 2, line 4: wash buffer 1, line 5 flow solution, line 6: precipitation, line 7: supernate, line 8: bacteria solution. (b) western blot of human SOD1, line 1: 4 μ M SOD1, line 2: 5 μ M SOD1.

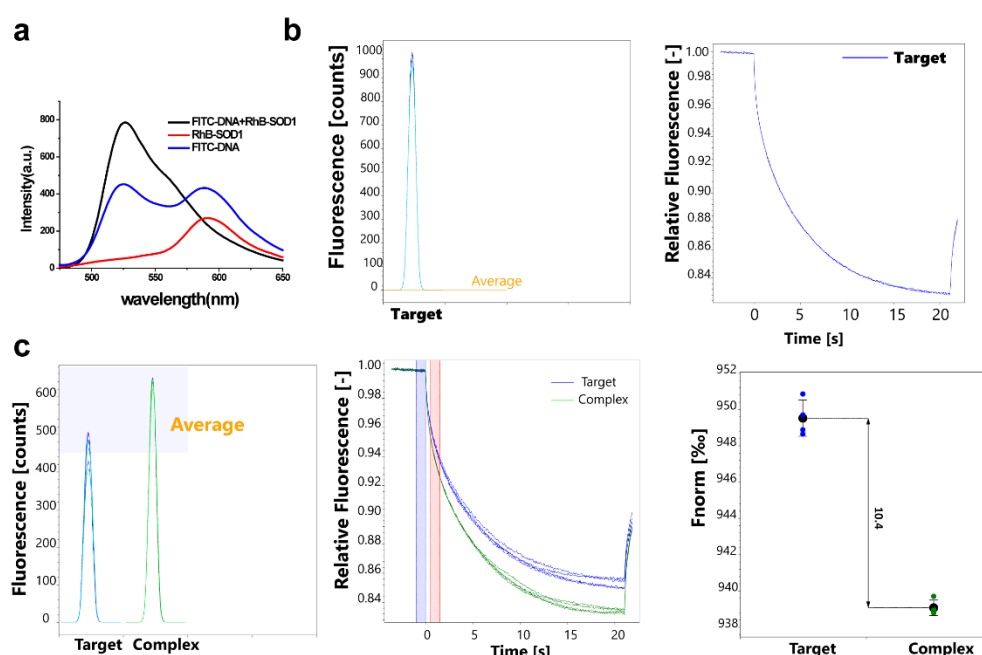


Figure S2. FRET and MST assay of the dsDNA and SOD1. (a) 100 μM FITC-dsDNA [1] was mixed with 200 μM RhB-SOD1 at 37 $^{\circ}\text{C}$ for 2 h in 20 mM Tris-HCl (pH 7.4), and the mixture was tested at the excitation wavelength of 450 nm. (b) NHS-SOD1 was first diluted to a final concentration of 3 μM by 10 mM pH 7.4 PBS containing 0.05% Tween, and then loaded into the Nano Temper Technologies glass capillaries for MST tests. (c) Binding check assays between NHS-SOD1 and dsDNA. 500 nM NHS-SOD1 was mixed with 20 μM dsDNA at room temperature for 5 min, and loaded into the Nano Temper Technologies glass capillaries. MST measurements were carried out using a 20% excitation power. NHS-SOD1 was recognized as the target.

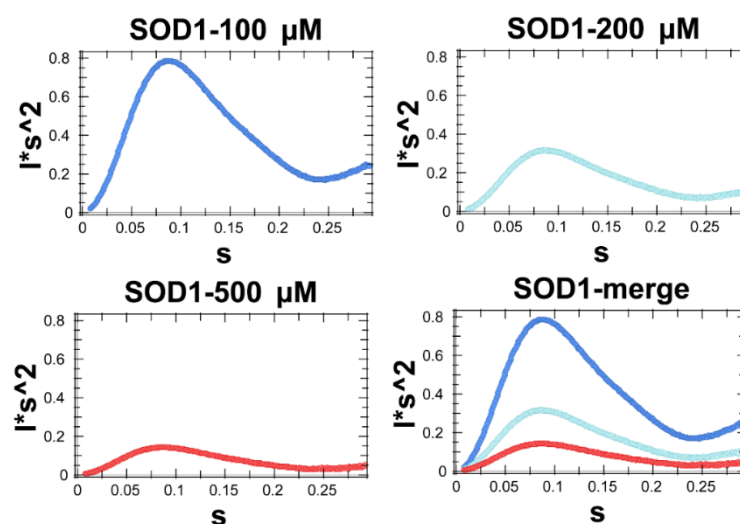


Figure S3. Kratky plot of bovine SOD1.

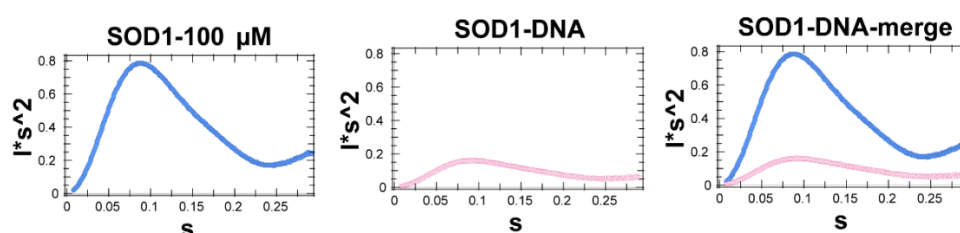


Figure S4. Kratky plot of SOD1 and SOD1-DNA complex.

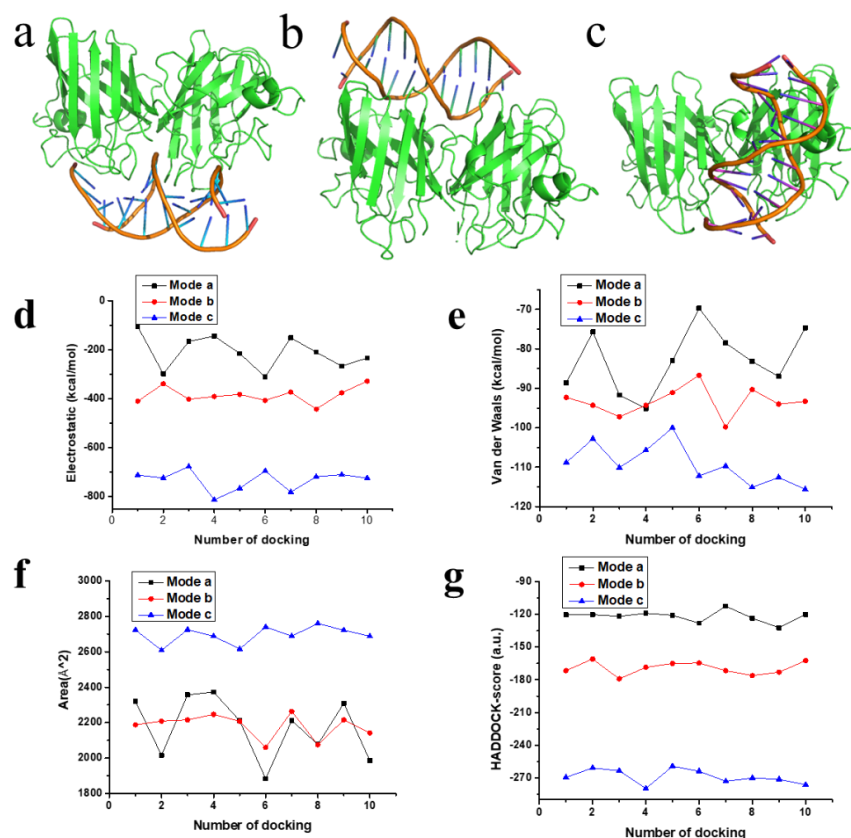


Figure S5. Three binding models (a,b,c) of SOD1 to dsDNA provided by HADDOCK 2.2 and comparisons among three models in the electrostatic and van der Waals energy, contact areas and HADDOCK score (d,e,f,g).

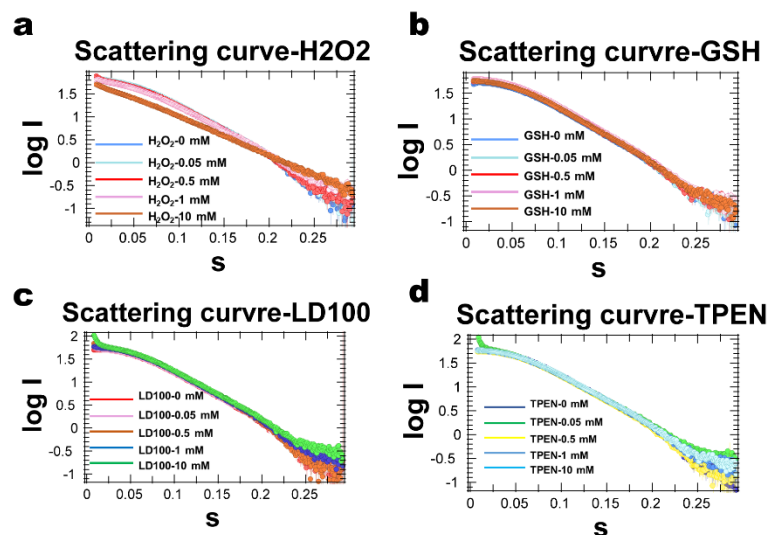


Figure S6. Scattering curves of dsDNA-SOD1 complexes. (a) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM H₂O₂. (b) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM GSH. (c) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM LD100. (d) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM TPEN.

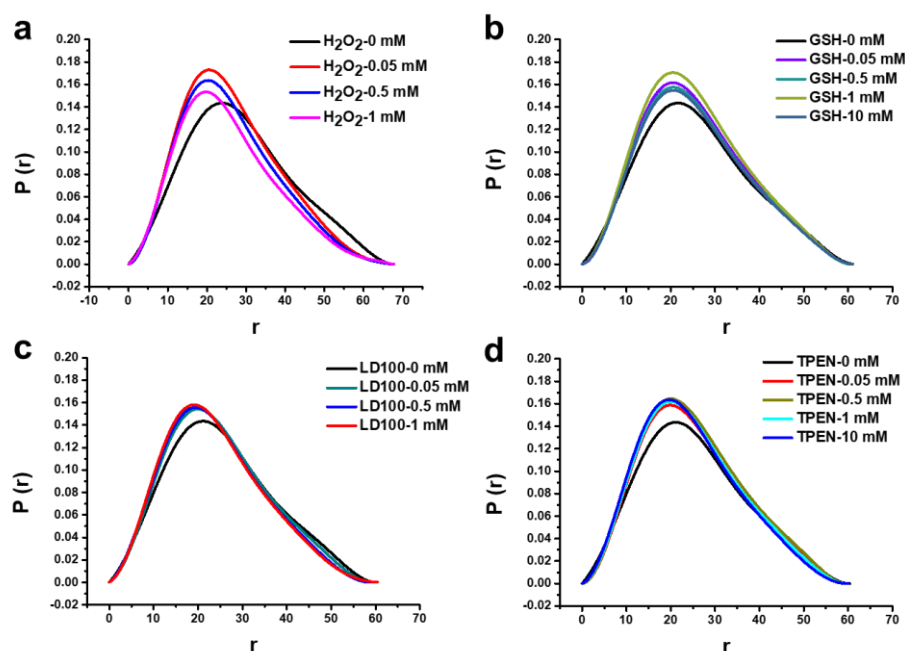
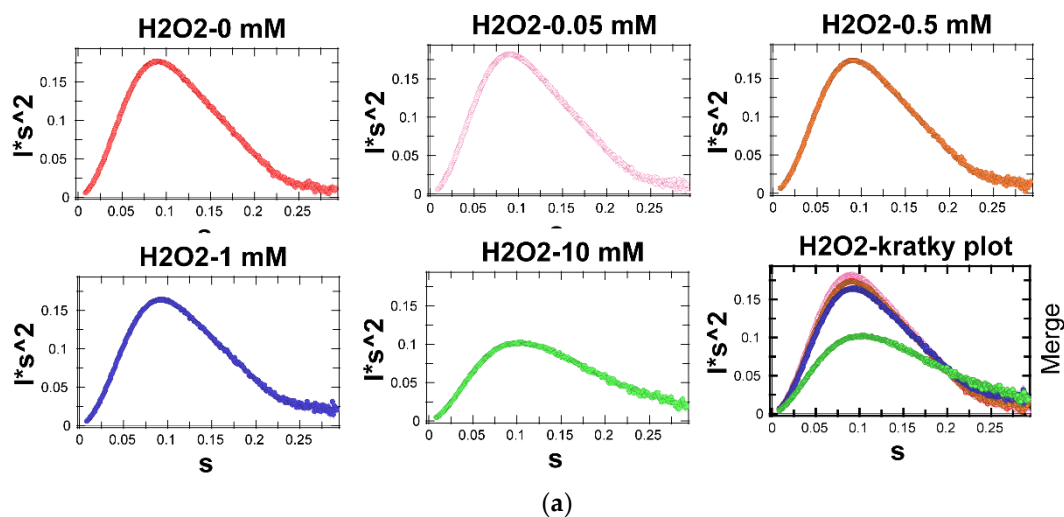


Figure S7. Pair distribution functions of dsDNA-SOD1 complexes computed by GNOM. (a) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM H_2O_2 . (b) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM GSH. (c) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM LD100. (d) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM TPEN. Samples were centrifuged at 10000 rpm, 4°C for 10 minutes to remove potential aggregates before data collection.



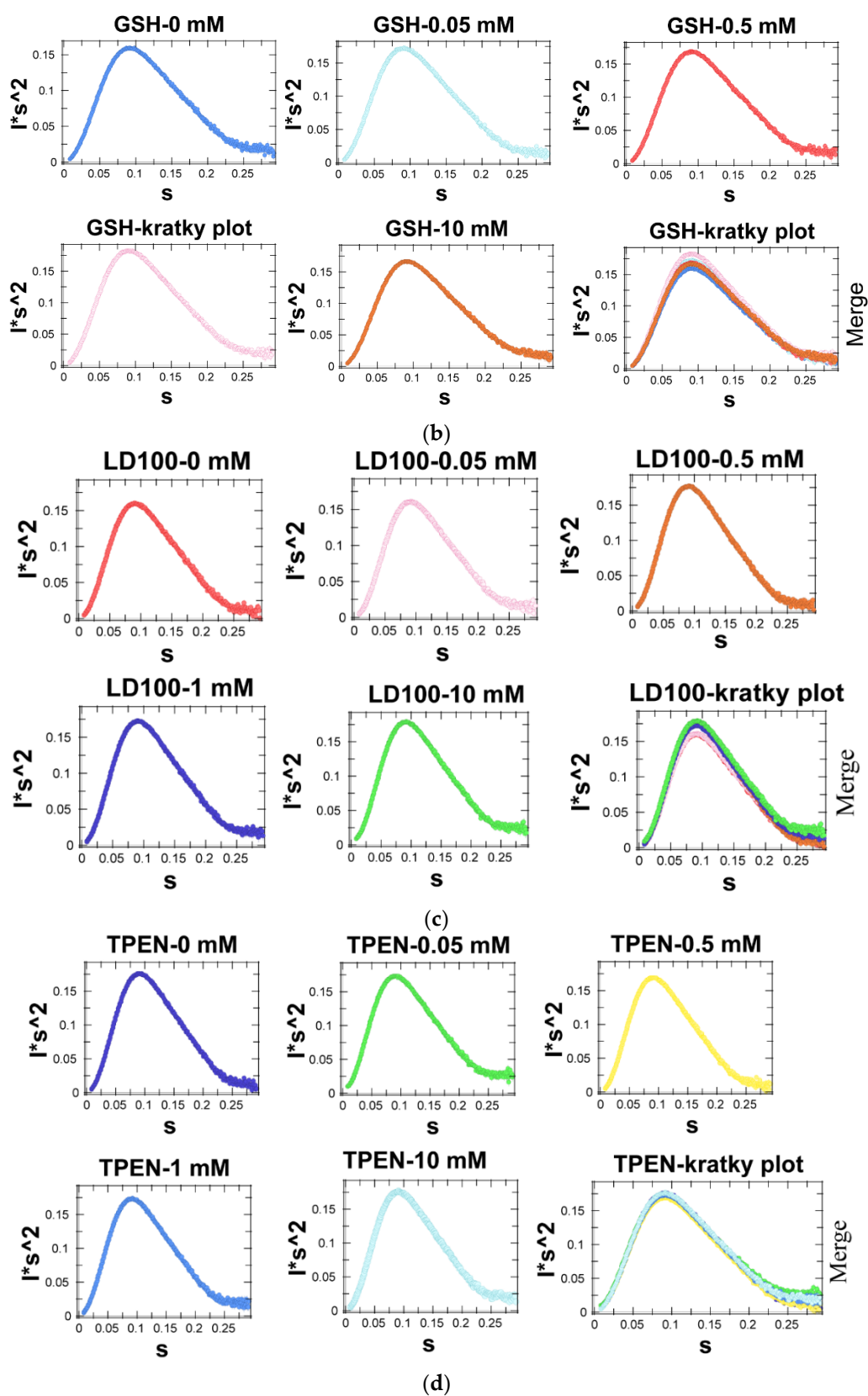


Figure S8. Kratky plot of dsDNA-SOD1 complexes under different conditions (a) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM H_2O_2 . (b) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM GSH. (c) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM LD100. (d) The mixture of SOD1 and dsDNA in the presence of 0, 0.05, 0.5, 1 mM TPEN.

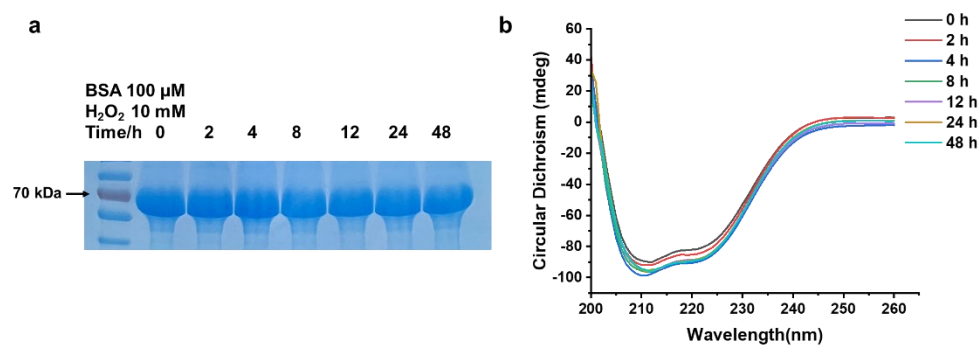


Figure S9. SDS-PAGE and CD spectra of BSA. (a) 100 μ M BSA was added with 10 mM H₂O₂ for varied time (0, 2, 4, 8, 12, 24 and 48 h) at 37 °C and tested using SDS-PAGE assay. (b) 100 μ M BSA was added with 10 mM H₂O₂ for varied time (0, 2, 4, 8, 12, 24 and 48 h) at 37 °C and tested by circular dichroism.

References

1. Roy, P.; Rajfur, Z.; Pomorski, P.; Jacobson, K. Microscope-based techniques to study cell adhesion and migration. *Nat. Cell. Biol.* **2002**, *4*, E91–E96.