

Table S1. Devices development for Glioblastoma application

| Device | Matrix composition | Proposed mechanism | Study model | Major Findings | Reference |
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| | | | (In vitro, in vivo, ex vivo) | | |
| Hydrogel system | HA-functionalized Methacrylate gelatin/poly(ethylene glycol) (PEG)4A | Mimic the microenvironment matrix for GBM cells | 2-3D cultures, patient-derived GBM cells | HA-dose-dependent alterations in GBM malignancy-associated markers. HA can lead to a specific interaction in EGFR+ cells via CD44. | [Pedron et al., 2013] 10.1016/j.biomaterials.2013.06.024 |
| Composite hydrogel | Collagen-Hyaluronan | Mimic the microenvironment matrix for GBM cells | 3D culture, patient-derived GBM cells | Cell morphology, spreading and migration were influenced by collagen type and HA concentration. | [Rao et al., 2013] 10.1021/am402097j |
| Hydrogels | HA-Gelatin thiol-reactive PEG diacrylate (PEGDA)/PEG divinyl sulfone (PEGDVS) crosslinked | Mimic the microenvironment matrix for GBM cells and develop a better analytical analysis <i>in vitro</i> | Single-cells, 3D culture, U118, U87R cell lines | Potential to identify differences in the invasive behaviour of the cell and analyse the cellular response to biophysical signals in the extracellular environment. | [Heffernan et al., 2014] 10.1007/s10439-014-1223-1 |
| Composite scaffold | Chitosan-HA scaffold | Mimic the microenvironment matrix for GBM cells | Single-cells, 2D culture, GSC GBM6 cell lines | Better cellular morphology, growth patterns, and malignant behaviour than adherent monolayers. | [Wang et al., 2016] 10.1002/adhm.201600684 |
| Micelles | HA-Modified Micelles lauroyl-gemcitabine (Gem-C12) and honokiol (HNK) encapsulated | Nanocarriers for targeting drug delivery of combined chemotherapeutics | Single-cells, 3D culture, U87 cell line Subcutaneous glioma-bearing mice; orthotopic xenograft GBM | Deep penetration into spheroids, endocytosis mediated by the CD44 receptor, and increased cytotoxicity of the combined treatment. Enhanced animal survival rate. | [Liu et al., 2018] 10.1021/acs.molpharmaceut.7b01035 |

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| Thermosensitive hydrogel | Doxorubicin/paclitaxel embedded Pluronic F127/HA hydrogel system (freeze-dried micelles) | Intratumoral administration of combined chemotherapy | Degradation rate, drug release Injection force determination, Chicken meat model | Controlled release of both chemotherapeutics and great stability. The formulation also showed easy injectability. | [Rezazadeh et al., 2018] 10.4103/1735-5362.228918 |
| Hydrogel | HA/ poly(ethylene glycol) (PEG) | Mimic the microenvironment matrix for GBM cells | 3D culture, patient-derived GBM cells | Cell viability and proliferation rates are as good as or better than standard glioma spheres culture. Compatible results for use in molecular and cellular analyses. | [Xiao et al., 2018] 10.3791/58176 |
| Aerogel sponge | Stromal cell derived factor-1 α loaded in Silk fibroin-HA-Heparin | Chemoattractant loaded scaffold trap for brain tumor cells | Single-cells, NIH3T3 cell line | Cytocompatible material with adequate mean pore diameter (60 μ m) and connectivity for welcoming cells. Texture similar to brain tissue (6–13 kPa Young's Modulus), slower biodegradation and 93% retention of SDF-1 α . | [Najberg et al., 2019] 10.1016/j.carbpol.2020.116107 |
| Nanofibers scaffold | Polycaprolactone (PCL) /gelatin (Gel) / hyaluronic acid (HA) | Mimic GBM extracellular matrix | Single-cells, U251 cell line | The topography and surface chemistry (hydrophilic) played an important role in the biomechanical, proliferative, and morphological properties of the cells. | [Unal et al., 2020] 10.3390/ma13112661 |
| Drug loaded Hydrogel | Crosslinked-HA-adipic acid dihydrazide (ADH) loaded with human urotensin (hUII) peptide and doxorubicin (DOX) or temozolomide (TMZ) | Chemoattractant matrix to trap and eradicate GBM cells | Single-cells, U87MG cell line | Cell migration and invasion into the hydrogel in response to the hUII. Significant cytotoxicity of DOX-loaded hydrogels for U87MG cells. | [Kasapidou et al., 2021] 10.1039/d1sm01003d |
| Drug-loaded hydrogel | Peptide functionalized HA (HACF) cross-linked by | Drug delivery reservoir for resection cavity | Single-cells, patient-derived GBM cells | Biocompatible, adaptable, and adjustable to local tissue stiffness. 45% improvement in patient survival. | [Parkins et al., 2021] 10.1016/j.biomaterials.2021.120919 |

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| | cucurbit[8]uril (CB[8]) | | Patient-derived xenograft (PDX) rats | | |
| Hydrogel | Methacryloyl-modified HA (HAMA) | Tissue engineering application | - | Suitable structure, porosity, and stiffness for tissue engineering purposes. Also, it showed to be resistant to collagenase biodegradation. | [Velasco-Rodriguez et al., 2021] 10.3390/ijms22136758 |
| Composite scaffold | HA/gelatin (Gel) modified bacterial cellulose (BC) | Mimic GBM extracellular matrix | 3D culture, U251 cell line | Good cell vitality, forming multi-layers and cell clusters in the scaffolds. | [Unal et al., 2021] 10.1007/s10570-020-03528-5 |
| Drug loaded Nanogel | Lactoferrin (Lf)/phenylboronic acid (PBA)-functionalized HA nanogels crosslinked with disulfide-bond and doxorubicin hydrochloride (DOX)-loaded | Drug delivery and dual-targeting GBM | Single-cells and coculture, G422, bEnd.3 cell lines SD rats - pharmacokinetics, ICR mice - biodistribution | Controlled release of the drug, effective BBB penetration, and precise glioma dual-targeting (PBA and HA), presenting strong cytotoxicity against G422 cells. | [Zhang et al., 2021] 10.1016/j.cjche.2021.08.029 |
| Polymer-drug conjugate | HA-Hydrazone-DOX | GBM local treatment | 2-3D culture GL261, U87MG, B16F10 cell lines GL261-bearing mice | Increased tumor cell mortality and inhibition of the tumor growth. Bio-responsive linker was crucial for the observed bioperformance. | [Malfanti et al., 2022] 10.3390/pharmaceutics14010124 |

Table S2. Systems developed for CNS injuries application

| Device | Matrix composition | Proposed mechanism | Porosity connectivity diameter | Mechanical properties | Study model | Major Findings | Reference |
|----------|--|--|--|-----------------------|--|---|---|
| | | | | | (In vitro, in vivo, ex vivo) | | |
| Hydrogel | HA -PDL hydrogel | Scaffold material for the repair of defects brain tissue | Good connectivity and adherence | - | <ul style="list-style-type: none"> Human DRG cell line and NSCs 3 months old Sprague Dawley rats | After implantation, the polymer hydrogel rightly bridged the tissue defect, establishing a permissive interface with the host tissue to aid cell ingrowth and angiogenesis. | [Tian et al.2005] 10.1089/ten.2005.11.513 |
| Hydrogel | HA-arginine-glycine-aspartic acid (RGD) | Tissue engineering for brain regeneration | - | - | <ul style="list-style-type: none"> Sprague-Dawley rats, cortex implantation | HA-RGD hydrogel provided a structural, 3D continuity across the cortex and favoured reorganisation of local wound-repair cells, angiogenesis, and axonal growth. | [Cui et al., 2006] 10.1007/s10856-006-0615-7 |
| Hydrogel | HA- Poly-D-lysine (PLL)/ nogo-66 receptor antibody (antiNgR) | Tissue engineering for nerve regeneration | 10 to 100 μ m interconnected network | - | <ul style="list-style-type: none"> Sprague-Dawley rats, lateral hemisection of spinal cord | Inhibited the glial scar formation, support angiogenesis, and promote axonal extension. | [Wei et al., 2007] 10.1002/jbm.b.31689 |

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| Sponge-like scaffold | Collagen type I and II/HA | Neurogenic induction of cells seeded into a construct for brain regeneration | 80-200µm pore size(75% to 91%) | 1-6.3kPa Young modulus | <ul style="list-style-type: none"> • 2-3D, primary cell culture | The scaffolds favour the differentiation of neural stem cells (NSCs) to neuronal cells in vitro. | [Wang et al., 2009] 10.1016/j.actbio.2009.03.033 |
| Gel | HA | Matrix to reduce the marginal glial scarring | 120 to 182 µm porosity | Tensile strength: 4.1-6.3 N/mm2 | <ul style="list-style-type: none"> • Sprague-Dawley rats | Inhibited the glial scar formation by decreasing the thickness of gliosis and by reducing the number of the glial cells. | [Lin et al., 2009] 10.1016/j.wneu.2009.09.004 |
| Hydrogel-microspheres composite | HA hydrogel / recombinant human brain-derived neurotrophic factor (BDNF) and vascular endothelial growth factor (VEGF)-poly(lactic-co-glycolic acid) microspheres loaded | Delivery system for NSCs | - | - | <ul style="list-style-type: none"> • 2-3D, primary and single-cells cultures (Sprague-Dawley embryos forebrain cells) | Stable releasing of the biofactors, promoting cells survival and growth. The soft property was suitable for CNS tissue. Biocompatible material, with great adhesion and cell proliferation. | [Wang et al., 2011] 10.1007/s11095-011-0452-3 |
| Hydrogel | HA-Modified methylcellulose | NSPCs cells delivery vehicle | - | - | <ul style="list-style-type: none"> • Rat NSPCs cells | Increased oligodendrocytes differentiation. | [Tam et al.,2012] 10.1039/c2jm33680d |

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| Electrospun nanofiber scaffold | HA-laminin-coated polycaprolactone (PCL) nanofibers | Neural tissue engineering for restore damaged tissue | Fiber diameters: 600–1200 nm | - | <ul style="list-style-type: none"> • 2-3D cell culture, eGFP neuronal SH-SY5Y cell line | The aligned nanofibers permitted the guidance of neurite outgrowth. | [McMurtrey, 2014] 10.1088/1741-2560/11/6/066009 |
| Hydrogel | HA-laminin/SDF-1 α loaded | Matrix to enhance neural transplant retention and migration delivery system | - | - | <ul style="list-style-type: none"> • 3D, primary cell culture; NPSCs • C57BL/6 mice, brain injection | SDF-1 α signalling critically mediates NPSC transplant chemotactic migration, and the matrix increases NPSCs transplant retention. | [Addington et al., 2017] 10.1016/j.matbio.2016.09.007 |
| Microporous annealed particle hydrogel | HA/K-peptide/Q-peptide/RGD | Matrix to promote brain tissue repair after stroke | - | - | <ul style="list-style-type: none"> • C57BL/6, stroke-induced | Reduced inflammatory response following stroke and increased peri-infarct vascularization. | [Nih et al., 2017] 10.1002/adma.201606471 |
| Drug delivery scaffold | SA and HA loaded device | Microenvironnement to embed stem cells in traumatic brain injury | 200 μ m to tens of μ m | 0.1 kPa to 1 kPa Young Modulus | <ul style="list-style-type: none"> • HUC-MSCs • SD rats | NSCs embedded in HA collagen biomaterials may promote reinnervation of damaged facial nerves, and artificial conduction of NSCs may provide | [Zhang et al., 2008] 10.1039/C7TB03213G |

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| | | | | | | a potential treatment for peripheral nerve injury. | |
| Hydrogel | Glycidyl methacrylate-HA and Methacrylic anhydride-HA/Collagen I and laminin functionalized | Engineered scaffold for tissue regeneration | 1 μ m pore size | 2-6kPa Young modulus | <ul style="list-style-type: none"> • Schwann Cells and dorsal root ganglia (DRGs) cell lines • Rat isolated tissue (sciatic nerve, brain and spinal cord) | The material can be tuned to numerous soft tissues, and supported 3D axonal elongation of DRGs cultures. | [Spearman et al., 2020] 10.1002/jbm.a.36814. |
| Hydrogel | HA-galactose oxidase (GalOx)-horseradish peroxidase (HRP)/bone mesenchymal stem cells (BMSC) and nerve growth factors (NGF) encapsulated | Implant or cell/drug delivery vehicle for tissue repair | <100um pore size loose and porous structure | Low elastic modulus <0.1kPa | <ul style="list-style-type: none"> • 3D and single-cells models; BMSC cells • Traumatic Brain Injury (TBI) model, C57BL/6 mice | Good biocompatibility, stability, and biodegradability. Suitable for cell survival and proliferation, accelerated brain repair process. | [Wang et al., 2022] 10.1016/j.mtbio.2021.100201 |
| Hydrogel | HA-poly(N-isopropylacrylamide) | Hybrid interpenetrating polymer network as cell carrier for | - | 0.1kPa-8kPa Elastic modulus | <ul style="list-style-type: none"> • Primary culture, NP cells • Cartilage and intervertebral | The matrix supported the cell viability and the phenotype expression of the encapsulated cells. | [Guo et al., 2022] 10.1016/j.carbpol.2021.118828 |

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| | | nucleus pulposus (NP) repair | | | disc (IVD) organ culture | | |
| Nanogel | Iron oxide (Fe ₃ O ₄)-HA | Theranostic material for Alzheimer's disease | - | - | <ul style="list-style-type: none"> Single-cells culture, C8-D1A cell line | Non-cytotoxic matrix. Efficient generation of negative contrast in MRI. | [Chen et al., 2022] 10.1016/j.arabjc.2022.103748 |

Table S3. Systems developed for PNS injuries application

| Device | Matrix composition | Proposed mechanism | Porosity and connectivity | Mechanical properties | Study model | Major Findings | Reference |
|--------------------------|--------------------|---|-----------------------------------|-----------------------|---|--|---|
| | | | | | (In vitro, in vivo, ex vivo) | | |
| Composite Conduit | HA / collagen | Tissue engineering for nerve regeneration | Diameter 1.2 mm Porosity 50 μm | - | <ul style="list-style-type: none"> Rat SCs and neurospheres NSCs from neural cortex of SD rat embryos | Cultured rat Schwann cells and neurospheres grow <i>in vitro</i> on new artificial HA-based nerve conduits. After 3 weeks of culture, conduits stayed circular with a round lumen, and cell-conduits conserved their original structure. | [Sakai et al., 2007] PMID: 17502696 |
| Composite conduit | HA / collagen | Tissue engineering for nerve regeneration | 7 mm in length | - | <ul style="list-style-type: none"> Normal adult New Zealand rabbits | NSC-embedded NT-3 HA-collagen composite scaffold, has shown that a number of nerve fibers were still unmyelinated. Degeneration and swelling of myelin lamellae were also visible. | [Zhang et al., 2008] 10.1186/1479-5876-6-67 |

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| Composite Conduit | HA-Silk Fibroin composite device | Tissue engineering for nerve regeneration | 79% of porosity | G'/G': 6 to 7 viscoelasticity | <ul style="list-style-type: none"> • Rat Schwann cells (rSCs) • Male CD-1 mice | Both HA and HA-SF scaffolds were received by the host with no residual immune response at 8 weeks. The results show that SF incorporation improves the mechanical properties of the material and leads to promising biocompatible conduits for tubing strategies. | [Gisbert et al., 2020] 10.1016/j.ijbiomac.2020.01.149 |
| Composite Conduit | PDLLA/ β -TCP/HA/CHS/NGF | Polymer based scaffold for controlled release of NGF | Semi-permeable, pore size less than 10 μ m | 13-60 kPa Elastic modulus | <ul style="list-style-type: none"> • PDLLA/β-TCP nerve conduits • Sciatic nerve defect in SD rat | Substantial enhancements in nerve regeneration were noticed after using the PDLLA/ β -TCP/HA/CHS/NGF NGCs based on the different assessments. In vivo studies show that the PDLLA/ β -TCP/HA/CHS/NGF sustained-release NGCs can significantly stimulate peripheral nerve regeneration, and the effect is comparable to that of autograft. | [Yan et al., 2021] 10.1007/s11595-021-2450-6 |
| Scaffold | PEDOT-doped HA NPs/ CS / Gel matrix | Tissue engineering for nerve regeneration | 200– 300 μ m pore size 83-92% porosity | 13-60 kPa Elastic modulus | <ul style="list-style-type: none"> • PC12 cell line | 8% PEDOT-HA/Cs/Gel scaffold had a higher cell adhesive efficiency and cell viability in comparison to the other conductive scaffolds. It also has shown a higher expression of synapse growth genes of GAP43 and SYP compared | [Wang et al., 2017] 10.1016/j.msec.2016.10.029 |

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| | | | | | | with the Cs/Gel control group. | |
| Hydrogel | HA-CS composite system | Hydrogels for nerve growth factor(NGF) sustained release and nerve regeneration | Porosity: 20 to 100 μ m | 2 to 4 kPa Elastic modulus | <ul style="list-style-type: none"> RSC96 rat SCs and PC12 cell lines Healthy adult Sprague-Dawley (SD) rats | <ul style="list-style-type: none"> The good mechanical, porous, and swelling properties play a synergistic role in enhancing nerve regeneration. The CS-HA/NGF hydrogels through a sustained release are favorable for promoting cell adhesion, spreading, and differentiation Chitosan and HA inhibited extraneural scarring, promoted nerve regeneration, raised the nerve conduction velocity and amended the recovery of nerve function. | <p>[Xu et al., 2016] 10.1177/0883911516662068</p> <p>[Li et al., 2018] 10.3892/mmr.2018.8388</p> |

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| | | | | | | Combined together they enhance neural regeneration and repair. | |
| Hydrogel | HA-CS/NGF hydrogel | Injectable hydrogels aimed to sustained release of nerve growth factor (NGF) | Pore size 53-73μm Porosity(82-87%) | 2.3-4.5kPa Elastic modulus | <ul style="list-style-type: none"> • BMMSCs, RSC96 rat SCs cells and PC12 cell line | <p>PDLLA/β-TCP nerve conduits formulated with CS-HA/NGF hydrogels improved the axon regeneration and myelination, contrasted to the PDLLA/β-TCP hollow nerve conduits and the autograft group. These findings imply that the CS-HA/NGF injectable hydrogel can successfully upgrade nerve regeneration, hence, it is a good candidate in the field of neural tissue engineering.</p> | <p>[Xu et al., 2022] 10.1016/j.compositesb.2021.109509</p> |