



Review

Adenosine, Schizophrenia and Cancer: Does the Purinergic System Offer a Pathway to Treatment?

Abdul-Rizaq Hamoud ¹, Karen Bach ¹, Ojal Kakrecha ¹, Nicholas Henkel ¹, Xiaojun Wu ¹, Robert E. McCullumsmith ^{1,2} and Sinead M. O'Donovan ^{1,*}

¹ Department of Neurosciences, University of Toledo, Toledo, OH 43614, USA

² Neurosciences Institute, ProMedica, Toledo, OH 43606, USA

* Correspondence: sinead.odonovan@utoledo.edu

Abstract: For over a century, a complex relationship between schizophrenia diagnosis and development of many cancers has been observed. Findings from epidemiological studies are mixed, with reports of increased, reduced, or no difference in cancer incidence in schizophrenia patients. However, as risk factors for cancer, including elevated smoking rates and substance abuse, are commonly associated with this patient population, it is surprising that cancer incidence is not higher. Various factors may account for the proposed reduction in cancer incidence rates including pathophysiological changes associated with disease. Perturbations of the adenosine system are hypothesized to contribute to the neurobiology of schizophrenia. Conversely, hyperfunction of the adenosine system is found in the tumor microenvironment in cancer and targeting the adenosine system therapeutically is a promising area of research in this disease. We outline the current biochemical and pharmacological evidence for hypofunction of the adenosine system in schizophrenia, and the role of increased adenosine metabolism in the tumor microenvironment. In the context of the relatively limited literature on this patient population, we discuss whether hypofunction of this system in schizophrenia, may counteract the immunosuppressive role of adenosine in the tumor microenvironment. We also highlight the importance of studies examining the adenosine system in this subset of patients for the potential insight they may offer into these complex disorders.

Keywords: schizophrenia; cancer; adenosine; purinergic signaling; epidemiology



Citation: Hamoud, A.-R.; Bach, K.; Kakrecha, O.; Henkel, N.; Wu, X.; McCullumsmith, R.E.; O'Donovan, S.M. Adenosine, Schizophrenia and Cancer: Does the Purinergic System Offer a Pathway to Treatment? *Int. J. Mol. Sci.* **2022**, *23*, 11835. <https://doi.org/10.3390/ijms231911835>

Academic Editor: Ronald Sluyter

Received: 9 August 2022

Accepted: 29 September 2022

Published: 5 October 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Epidemiological observations dating back over one hundred years suggest that incidence rates of some cancers may be lower in patients with schizophrenia [1–5]. Numerous studies have been conducted in the decades since then but have failed to reach a consensus; there are reports of increased, reduced, or no difference in incidence rates of most cancers in schizophrenia patients compared to the general population (see reviews [2–5] and epidemiology studies [5–11]). However, as many of the risk factors for cancer, including smoking, alcohol use, increased body weight, physical inactivity, and reduced access to medical care, are also found in schizophrenia populations, it is surprising that cancer incidence rates are not higher amongst these patients [4,6,12–17]. As has previously been observed, this unexpected finding lends support to the idea that a diagnosis of schizophrenia may be protective against development of (some) cancers [18].

2. Cancer Incidence in Schizophrenia

A limitation of many epidemiological studies of cancer incidence in schizophrenia is that they do not control for cancer-related confounding variables. Age, for example, is a major confounding variable, as patients with schizophrenia typically have a shorter lifespan [19], making comparisons of cancer incidence with control populations a challenge. It is also crucial to precisely define control groups (general population, patients in inpatient

care or outpatient care, non-schizophrenia, non-psychiatric, etc.) and to match study subjects based on age and sex. However, when risk factors like age and smoking are considered in analyses, the reported cancer incidence rates are lower than would be expected in this patient population [6,7]. It is also important to note that cancer incidence rates, and not cancer mortality rates, are proposed to be lower for some cancers in schizophrenia patients. Survival rates of schizophrenia patients who develop cancer are typically lower than the general public [20,21]. This is especially interesting when considering separate biological mechanisms regulate cancer initiation, proliferation and metastasis [22]. In Table 1 we summarize the results of recent epidemiology meta-analyses reporting cancer rates in schizophrenia patients. Overall, a limited number of prospective cohort studies were available for inclusion in these meta-analyses, missing medication history and detailed demographic information is common, variable data collection methods and the use of inconsistently defined control groups contribute to challenges in interpreting the data and determining whether cancer incidence rates are significantly reduced in schizophrenia patients [3,23]. Interestingly, breast cancer incidence rates are consistently higher in studies of schizophrenia populations (Table 1). Recent genome wide association studies (GWAS) have identified a shared risk locus associated with schizophrenia and breast cancer [24] and increased risk for breast, ovarian and thyroid cancers, but not other cancers, in schizophrenia patients [25]. Conversely, lung, liver and prostate cancer incidence appears to be lower in schizophrenia populations (Table 1), suggesting that incidence rates of some cancers may be reduced, or unchanged, in these patients. Additional appropriately powered studies that consider different cancer types and account for potentially confounding variables (age, BMI, smoking, medications, genetics, and geography) will be required to ascertain whether a diagnosis of schizophrenia protects against the development of some cancers.

Table 1. Review of Meta-analyses examining cancer rates in schizophrenia patients.

Source	Sex	Participants (N)	Cancers	Major Findings
[5]	M/F	480,356 SCZ patients	All sites	<ul style="list-style-type: none"> Decreased overall cancer incidence (M/F) No change in females alone Sex and cancer type were confounding factors
[4]	M/F	279,938 patients across all 21 analyses	All sites	<ul style="list-style-type: none"> Colon, skin, and prostate cancer incidence decreased Pooled overall cancer rates were not significantly changed prior to adjusting for smoking Lung cancer rates were decreased following adjustment Breast cancer incidence was increased in female SCZ patients SCZ relatives' cancer risk was decreased
[26]	M/F	31 studies with a median of 33,372 psychotic patients	All sites	<ul style="list-style-type: none"> Cancer and sex specific incidence rates were reported Risk ratios increased for oesophageal, breast, testicular, cervical and endometrial cancers Risk ratios decreased for prostate, colon, skin, and thyroid cancers Authors note better study design, controlling for confounders and appropriate comparison groups are needed
[27]	M	218,076 men across 13 studies	Prostate	<ul style="list-style-type: none"> Decreased risk of prostate cancer across 13 cohort studies in SCZ men (SIR = 0.61).

Table 1. Cont.

Source	Sex	Participants (N)	Cancers	Major Findings
[11]	M/F	496,265 SCZ patients across 12 studies	Lung	<ul style="list-style-type: none"> No changes observed in lung cancer incidence Authors note only one study accounted for smoking and seven studies examined lung cancer incidence only after SCZ diagnosis while the rest did not
[10]	M/F	312,834 SCZ patients across seven studies	Liver	<ul style="list-style-type: none"> Significantly lower liver cancer incidence was observed in SCZ males (SIR=0.71) but not in females (SIR=0.83). SCZ patients have ~20% decreased risk of liver cancer Authors note confounders were not adjusted for
[23]	F	466,244 patients across 15 studies	Breast	<ul style="list-style-type: none"> Breast cancer incidence was increased in SCZ patients. Authors suggest morbidity (T2D, hyperprolactinemia, etc.) in female SCZ patients may contribute to higher breast cancer rates

M-Male, F-Female, SCZ-Schizophrenia, T2D-Type II Diabetes, SIR-Standardized Incidence Ratio.

Chronic antipsychotic use, genetic factors, and pathophysiological changes have been posited to explain the potentially reduced risk of cancer development in schizophrenia patients. For example, there is a growing body of evidence that antipsychotics may protect against the development of cancer; antipsychotics are now being repurposed as potential cancer therapies, with anticancer mechanisms of action in peripheral tissues still poorly understood [6,28–31]. Epidemiological studies have found that unaffected family members of patients with schizophrenia have reduced rates of cancer [4,32], suggesting that genetic factors contributing to schizophrenia risk may also reduce the risk of developing cancer. However, much less is known about how pathophysiological changes associated with schizophrenia may contribute to the reduced risk of cancer development in some patients [33,34].

The potential protective effects of the adenosine system against cancer development in schizophrenia patients was first suggested over 20 years ago in the “Purinergic Hypothesis of Schizophrenia” by Lara and Souza [35,36]. The authors noted the reduced rates of cancer in schizophrenia patients, and proposed a role for the purinergic system in this phenomenon, although little was known about the function of the adenosine system in cancer at that time [37,38]. Today, the purinergic hypothesis has evolved, with a growing body of evidence suggesting that reduced availability of extracellular adenosine, and the resulting effects on neuromodulation, and immune system and bioenergetic regulation, may contribute to the onset of schizophrenia [39]. Conversely, increased adenosine metabolism is now recognized as a characteristic of many types of cancer. Elevated adenosine, and its metabolic precursor adenosine triphosphate (ATP), promote cancer growth and play a role in host immunosuppression in the tumor microenvironment (TME) [36,40–43].

Herein, we discuss the biochemical and pharmacological evidence for hypofunction of the adenosine system in schizophrenia and hyperfunction in cancer (Figure 1). We draw primarily from postmortem and clinical studies to explore whether perturbation of the adenosine system may contribute to reduced risk for cancer development in some schizophrenia patients. We also review the literature showing that augmentation or attenuation of the adenosine system can have therapeutic benefits for these disorders.

Perturbations of the Adenosine system in Schizophrenia and Cancer

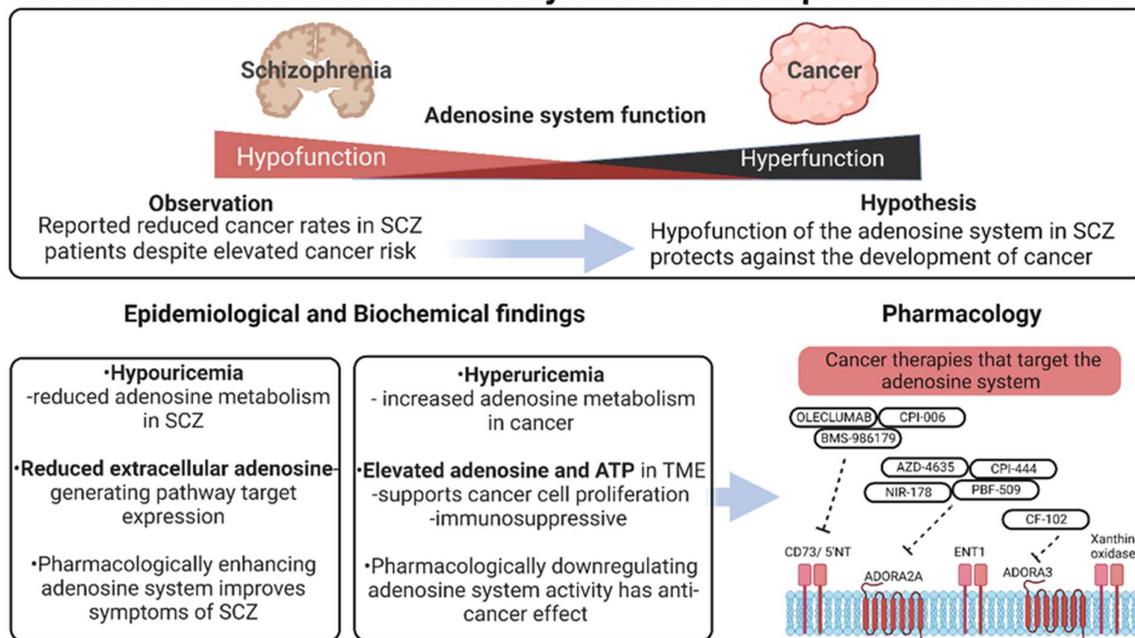


Figure 1. Perturbations of the adenosine system in schizophrenia and cancer. Overview of the population and biochemical evidence for perturbed adenosine system function in schizophrenia and cancer. Reduced cancer rates in schizophrenia patients despite elevated cancer risk implies some underlying biochemical protective mechanism. Reduced extracellular adenosine availability, and hypouricemia observed in these patients may explain the reduced cancer rates. Generated in Biorender.

3. The Adenosine System: Extracellular Adenosine Generating Pathways

The ratio of ATP:adenosine is tightly regulated by enzymatic pathways that modulate the generation of extracellular adenosine (Figure 2). Adenosine can be directly released from the cell by nucleoside transporters [44] where it is rapidly catabolized to inosine by the purine salvage pathway enzyme adenosine deaminase (ADA) [45]. Inosine is further metabolized via a series of enzymes to the end-product of adenosine metabolism, uric acid. Alternatively, adenosine can be generated from the sequential breakdown of ATP [46]. ATP can be released from the cell into the extracellular space via several different mechanisms including vesicular release, lysosomes, cell lysis, and nucleotide-permeable channels [47–51]. ATP is then rapidly catabolized to adenosine by the extracellular enzymes ectonucleoside triphosphate diphosphohydrolases (ENTPD 1–3, 8; Table 2) [52] which hydrolyze ATP to ADP/AMP. Ecto-5′nucleotidase (NT5E; Table 2) is the primary rate-limiting enzyme that converts AMP to adenosine [53–55]. During pathophysiological states, ATP and adenosine are released into the extracellular space to fine-tune the immune system [56]. ENTPD1 and NT5E play significant roles in the regulation of immunity and inflammation [57,58]. This enzyme cascade modulates purinergic signaling by driving a shift from a pro-inflammatory ATP state to an adenosine-induced anti-inflammatory environment, a role that is facilitated by their widespread expression on almost all tissue cell types [59]. Thus, adenosine and ATP levels are closely interrelated and the ratio of adenosine to ATP acts as a significant modulator of their biological effects.

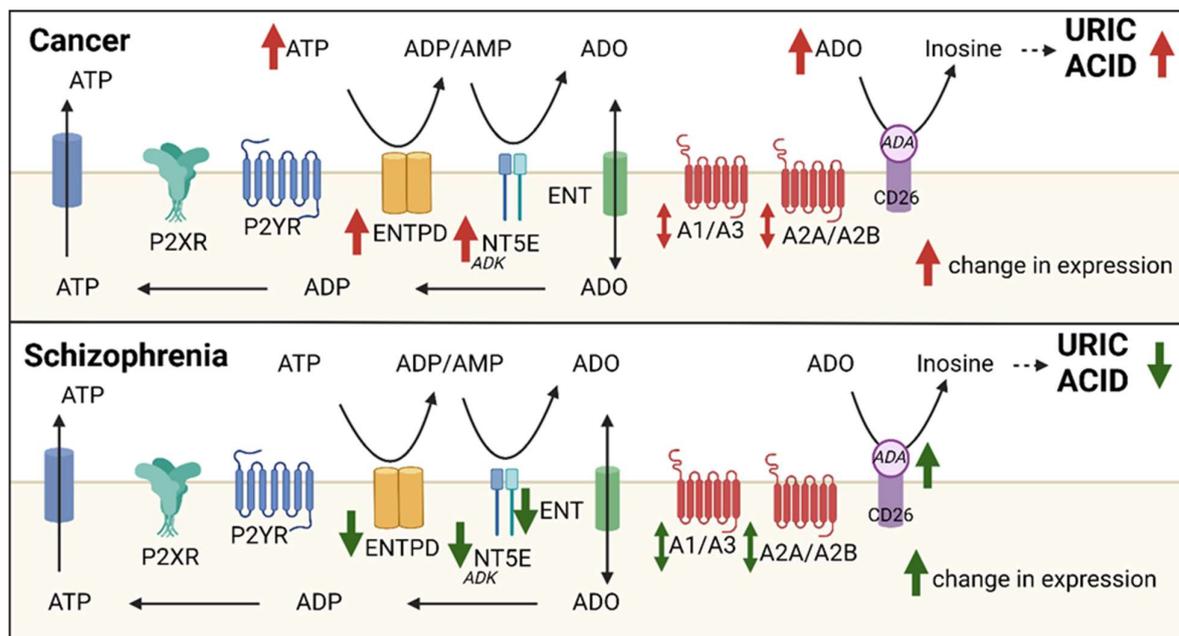


Figure 2. ATP and adenosine metabolism and signaling in cancer and schizophrenia. Perturbations in extracellular adenosine generating pathways are implicated in schizophrenia (green arrows) and cancer (red arrows). Differential purinergic (P1 and P2 receptors) receptor expression is reported in schizophrenia and cancer. ADA adenosine deaminase, ADK adenosine kinase, ADO adenosine, ATP adenosine triphosphate, ADP adenosine diphosphate AMP adenosine monophosphate, ENTDP ectonucleoside triphosphate diphosphohydrolases, NT5E ecto-5′ nucleotidase. Generated in Biorender.

Table 2. Nomenclature of ENTDP family enzymes.

Gene Name	Protein Name	Additional Names
ENTPD1	NTPDase1	CD39, ATPase, ecto-apyrase
ENTPD2	NTPDase2	CD39L1, ecto-ATPase
ENTPD3	NTPDase3	CD39L3
ENTPD8	NTPDase8	liver canalicular ecto-ATPase
NT5E	Ecto-5′ nucleotidase	5′NT, CD73, NT5E

ENTPD/NTPDase: Ectonucleoside triphosphate diphosphohydrolase, CD39: Cluster of Differentiation 39, adapted from [54,55].

4. The Adenosine System: Purinergic Receptors

Adenosine and ATP activate two main families of receptors; the purinergic P1 and P2 receptors; respectively (see Supplementary Table 1 for receptor nomenclature; ligands; G-protein; and ions) [60–68]. The G-protein coupled P1 adenosine receptors are inhibitory (A_1 and A_3) and excitatory (A_{2A} R and A_{2B} R); with high affinity (A_1 R and A_{2A} R) (0.1–0.3 μ M) and low affinity (A_{2B} R; A_3 R) (15–25 μ M) for adenosine [36,69,70], although some studies using functional assays report that adenosine is approximately equipotent at its receptors [71–73]. Purinergic receptor activation is a tightly regulated process. A_1 Rs are typically tonically activated by endogenous extracellular adenosine whereas A_{2A} Rs are selectively recruited by adenosine produced through extracellular catabolism of ATP by ectonucleotidases [74–77].

Due to the widespread constitutive expression of purinergic receptors in the body and the ability of P1Rs to form homo- and heterodimers with other neurotransmitter receptor types [68], adenosine and ATP serve significant roles in modulating neural processes, immune response, energy metabolism, and sleep [69–71]. Presynaptic A_1 receptor activation inhibits excitatory synaptic transmission by reducing calcium influx and glutamate re-

lease, while post-synaptic A₁ receptor activation reduces ionotropic glutamate receptor and voltage-sensitive calcium channel activation [78] leading to a decrease in excitatory synapse activity. Conversely, the actions of facilitatory A_{2A}Rs increase the release of glutamate and ionotropic glutamate receptor function in different brain regions. Their expression on astrocytes regulates glutamate uptake and Na⁺/K⁺ ATPase, contributing to the role of A_{2A}Rs as selective mediators of synaptic plasticity [78]. Increasing the complexity of adenosine's neuromodulatory action [79,80], adenosine receptors can also form heterodimers with other G-protein coupled receptors [81]. Activation of antagonistic A_{2A}-D₂ receptor heteromers, first identified in striatal membrane preparations [82] and found in GABAergic striatopallidal neurons [83], results in reduced D₂ receptor affinity for agonists. A_{2A}-D₂ receptors expressed on striatal astrocytes also functionally interact to modulate glutamate gliotransmitter release [84]. Inhibitory A₁ receptors dimerize with excitatory dopamine D₁ receptors to form receptor heteromers that modulate striatonigral neuronal function [83]. Adenosine's ability to modulate glutamatergic and dopaminergic pathways is of relevance in the pathophysiology of schizophrenia.

ATP and adenosine also regulate immune responses via activation of purinergic receptors, leading to immune cell activation, signal amplification, chemotaxis, and phagocytosis [56,85–87]. ATP acts as a pro-inflammatory danger associated molecular pattern (DAMP) when it is increased in extracellular fluids, usually indicating inflammation, ischemia, or injury [36,88]. In contrast, adenosine displays potent anti-inflammatory effects [89–91]. Modulation of purinergic receptors and ectonucleotidases allows for complex feedback regulation. During periods of injury or stress, cells respond by increasing ATP release into the extracellular space, inducing a pro-inflammatory response upon binding to P2 receptors. Most immune cells express several types of P2XRs and P2YRs [92]. Injured or stressed cells release ATP to recruit immune cells for phagocytosis and clearance. Immune cells can also release ATP to amplify cellular activation and regulate chemotaxis [93]. Prolonged states of inflammation can be toxic to tissues and cells. Thus, to prevent excessive inflammation and tissue damage, ATP is hydrolyzed by ectonucleotidases to generate adenosine. The decreased extracellular ratio of ATP:adenosine reduces P2 signaling and increases P1 signaling, specifically the upregulation and activation of A_{2A}Rs and A_{2B}Rs. Engagement of A_{2A}Rs allows for the inhibition of lymphocyte activation [94] while activation of A_{2B}R increases the production of vascular endothelial growth factor (VEGF) and interleukin-6 (IL-6) [95]. A_{2B}Rs, which are mainly expressed on macrophages, dendritic cells, and mast cells, may then contribute to the resolution of inflammation by promoting healing [96]. The endogenous anti-inflammatory role of adenosine is utilized in therapies for acute and chronic inflammatory diseases like chronic obstructive pulmonary disease and rheumatoid arthritis [97,98].

5. The Adenosine System: Perturbed Adenosine Metabolism in Disease: Uric Acid

The final product of adenosine metabolism, uric acid, is used clinically to assess purine catabolism. Altered levels of uric acid in the blood of schizophrenia patients implicates impairment of the adenosine system in the pathophysiology of this disorder. Reduced plasma uric acid levels are reported in clinically stable and relapsed schizophrenia patients [99] and in first-episode neuroleptic-naïve schizophrenia patients [100] compared to healthy controls. Although schizophrenia patients are not hypouricemic (generally defined as <2 mg/dL), they report lower uric acid plasma levels (schizophrenia male—5.1 mg/dL, schizophrenia female—4.4 mg/dL; healthy control male—6 mg/dL, healthy control female—5.1 mg/dL), suggesting perturbed purinergic signaling [100]. In contrast, higher levels of uric acid are reported during the acute phase in schizophrenia patients [101]. Similarly, during psychosis relapse, elevated uric acid levels have been found in schizophrenia patients that were as high as in bipolar disorder patients during manic episodes and higher than in the healthy control group [102]. These inconsistent findings of uric acid levels may be attributed to the relatively small study sizes, differences in uric acid levels at different stages of this heterogeneous disorder, or the effects of antipsychotic medications. A recent meta-

analysis of 17 studies addressed these limitations, and in subgroup analyses He et al., found that uric acid levels are significantly reduced in first episode psychosis (Weighted mean difference $-40.61 \mu\text{mol/L}$, $\text{CI}_{95\%} -59.47--21.76$, $p < 0.0001$) in schizophrenia patients but not in those with chronic schizophrenia (Weighted mean difference = $-3.09 \mu\text{mol/L}$, $\text{CI}_{95\%} -35.21--29.04$, $p = 0.85$) [103]. They also reported uric acid levels are reduced in male (Weighted mean difference = $-34.83 \mu\text{mol/L}$, $\text{CI}_{95\%} -54.97--14.69$, $p = 0.0007$) and American schizophrenia patients (Weighted mean difference = $-39.90 \mu\text{mol/L}$, $\text{CI}_{95\%} -61.47--18.33$, $p = 0.0003$). These findings suggest disease-stage and sex-dependent differences in uric acid levels indicating an alteration in purine catabolism early in the course of the disease. Hypofunction of the adenosine system in schizophrenia lends one possible explanation for these decreased uric acid levels in patients, as a lowered availability of adenosine restricts uric acid production.

Uric acid also plays a dual role in the body's antioxidant defense system [104]. Low levels of uric acid decrease the body's ability to prevent free radical damage, leading to cellular damage and death [105]. Conversely, high levels of uric acid (hyperuricemia, generally defined as $>6 \text{ mg/dL}$) contribute to an inflammatory response, as seen in disorders such as gout and metabolic syndrome [106,107]. Uric acid acts as a powerful antioxidant to neutralize free radicals, such as peroxy nitrite and hydroxyl, making its levels in the blood a strong indicator of oxidative stress. There is robust evidence supporting an increased oxidative stress status in schizophrenia [108,109], while the use of antioxidant supplementation, like vitamin E and C, improves some of the psychopathological symptoms of schizophrenia [110–112]. Thus, as well as reflecting changes in adenosine metabolism, baseline levels of purine metabolites are a good predictor of clinical and neurological symptoms in schizophrenia [113].

Hyperuricemia is associated with cancer incidence and mortality [114,115]. Elevated serum uric acid (6.8 mg/dL or $404 \mu\text{M}$ without the presence of gout) is positively correlated with risk for gastrointestinal cancers and an increased overall mortality risk for cancers [116]. High serum uric acid ($>6 \text{ mg/dL}$) is used as a prognostic predictor of colorectal cancer, pancreatic cancer, large B-cell lymphoma, and esophageal squamous cell carcinoma [117–120]. A study examining hyperuricemia in cancer development showed spontaneous hepatocellular carcinoma and hepatomegaly in mice following knockout of the enzyme, *Urah^{Plt2/Plt2}*, responsible for uric acid metabolism [121]. These findings point to altered adenosine catabolism as contributing to cancer incidence and mortality. Adenosine levels are increased in the TME but not in plasma of patients with cancer [122,123]. Equally, expression and activity of the enzyme ADA, which metabolizes adenosine to inosine, is also increased in tumors [124], but similar increases in serum may originate from other sources [125]. Increased uric acid levels likely reflect the increased abundance of adenosine in the TME and are an indicator of oxidative stress in response to cancer.

We posit that the adenosine system is hypofunctional in patients with schizophrenia resulting in reduced uric acid generation, in line with the reports of lower peripheral uric acid levels, which potentially mitigates the increased risk for cancer associated with hyperuricemia. Hypo- and hyperuricemia are associated with disease and although animal models indicate a potential causative role for hyperuricemia in hepatic cancers [121], the etiology of dysregulated uric acid levels in these diseases is not yet known. Rather, uric acid acts as a marker of purine catabolism, suggesting significant perturbation of the adenosine system in both schizophrenia and cancer.

6. Schizophrenia: Evidence for Adenosine System Perturbation

Schizophrenia is a severe, debilitating mental illness characterized by paranoid delusional thinking, hallucinations and alterations in volition, neurocognition, and affective regulation. While the global incidence of schizophrenia is relatively small, estimated at 0.30–0.66%, there is a substantial burden to the individual afflicted with the disease and to society [126,127]. One evolving theory describing the etiology of schizophrenia is dysfunction of the adenosine system. As a neuromodulator of both the glutamate and dopamine

neurotransmitter systems, perturbation of adenosine may reconcile the hyperdopaminergic and hypoglutamatergic states that underlie the positive, negative, and cognitive symptoms found in patients with schizophrenia. This hypofunction of adenosine hypothesis developed from observation of several phenomena: (1) presynaptic activation of adenosine A₁ receptors inhibits the release of glutamate, (2) postsynaptic activation reduces NMDA-receptor functioning, and (3) adenosine signaling through postsynaptic A_{2A} receptors inhibits D₂ receptor signaling through a heteromeric complex, as reviewed [128]. Both dysfunction of the glutamate and the dopamine systems have been attributed to the etiology of schizophrenia.

The enzymes responsible for the generation and regulation of extracellular adenosine are significantly altered in the brain in schizophrenia. Protein expression of the equilibrative nucleoside transporter (ENT1), which transports adenosine, is reduced in SCZ in the superior temporal gyrus [129]. Protein expression and enzyme activity of ENTPD1 and NT5E are significantly reduced in the striatum in schizophrenia subjects [130]. ENTPD1 and ENTPD2 transcript levels were also significantly reduced in enriched populations of astrocytes, but not neurons, in the dorsolateral prefrontal cortex (DLPFC) in schizophrenia [131]. Conversely, transcript levels of adenosine deaminase (ADA), the enzyme responsible for metabolizing adenosine to inosine, were significantly increased in an enriched population of pyramidal neurons in the same study, suggesting region- and cell-subtype specific dysregulation in adenosine metabolism. Although it has been proposed that increased expression of the enzyme adenosine kinase may drive adenosine hypofunction [39,132], recent findings show no significant changes in adenosine kinase expression in the brain in schizophrenia [133]. This suggests that hypofunction of ectonucleotidase and ADA pathways are likely responsible for the altered availability of extracellular adenosine.

Purinergic receptors are also significantly dysregulated in schizophrenia. Polymorphisms in the A₁ receptor, but not the A_{2A} receptor, are associated with schizophrenia [128]. In the striatum, a reduction in the transcript and the protein expression of the A_{2A} receptor, attributed to hypermethylation of the A_{2A} coding region, was found in schizophrenia [134]. Increases in striatal A_{2A} receptor density [135] and caudate nucleus A_{2A} and D₂ receptor protein expression [136] are reported in schizophrenia patients. Conversely, A_{2A}-D₂ receptor heterodimers were significantly reduced (almost 60%) in the same subjects [136]. These results lend support to the adenosine hypothesis of schizophrenia and deficits in the neuromodulatory action of adenosine on the dopamine system via the A_{2A}-D₂ receptor heteromers. No changes were found in A₁ receptor expression in the striatum [133], but there was a reduction in the A₁ receptor transcript in an enriched population of pyramidal neurons from the DLPFC in schizophrenia subjects [131]. Little is known about A_{2B} and A₃ receptor expression in the brain in schizophrenia, likely due to their low levels of expression in this tissue. As a modulator of the dopamine and glutamate neurotransmitter systems, dysfunction of the adenosine system is hypothesized to contribute to the dopaminergic hyperfunction and glutamatergic hypofunction that contributes to the onset of schizophrenia symptoms.

7. Schizophrenia: The Adenosine System as a Therapeutic Target

The clinical treatment for schizophrenia is mainly centered on antipsychotics, specifically dopamine D₂ receptor antagonists. However, a number of small clinical studies demonstrate the utility of the adenosine modulators allopurinol, a xanthine oxidase inhibitor [137,138], and dipyridole, an inhibitor of the adenosine transporter (ENT) [139]. Used as add-on therapies to antipsychotics, they improved both positive (hallucinations, delusions, mania) and negative symptoms (avolition, flat affect, asociality, alogia) of schizophrenia, as assessed by the Positive and Negative Syndrome Scale (PANSS) [140,141]. However, others report no differences between adjuvant allopurinol and placebo [142]. A recent study found that administering adenosine modulators as add-on therapies significantly reduced the rate of psychiatric rehospitalization and all-cause mortality among schizophrenia patients in a Finnish cohort, particularly in younger patients (<45 years) [143].

In animal models of schizophrenia-like behaviors, augmenting adenosine levels in mice that overexpress the enzyme ADK [37] and antagonism of the P2X7R in the sub-chronic phencyclidine-induced model of schizophrenia [144], significantly improved cognitive deficits. Further investigation of the adenosine system, including therapies that directly target purinergic receptors and ATP-adenosine metabolic pathway components, may offer new avenues for the treatment of schizophrenia [145].

8. Cancer: Extracellular Adenosine Generation and Cancer

The TME is a multicellular complex composed of various immune cells, cancer associated fibroblasts, endothelial cells and cancer stem cells [146]. The TME suppresses immune response and supports a complex network of signals between multiple cell types that work in concert to permit and sustain tumor growth and vascularization [147–149]. Accumulation of extracellular ATP is observed in the TME and under hypoxic, chronic and acute inflammation conditions [122,123]. Extracellular ATP levels are low (nanomolar range) in healthy tissue however upon cellular stress, damage or cell death, extracellular ATP can increase significantly (micromolar range) [123,150]. As ATP is released from cancer cells, it is metabolized to immunosuppressive AMP and adenosine, which act as key modulators of immune cells [36]. As in the brain, adenosine in the TME is primarily produced by extracellular metabolism of ATP via a cascade of ectonucleotidase enzymes (Figure 2) [151]. Studies examining *ex vivo* adenosine levels in rodent models found adenosine to be twice as high in tumors as compared to other tissues and at least 30% higher in the tumor core [123]. The increased presence of adenosine in the TME serves to regulate cancer cell growth and immune cell activity following adenosine receptor activation [123,152,153].

Enzymatic pathways that regulate the production of extracellular adenosine are also of importance in cancer development. The purine salvage pathway enzyme, ADA, modulates ATP:adenosine levels. ADA activity is increased in breast, kidney, and colorectal cancers as well as lymphocytes, a potential compensatory mechanism to offset elevated adenosine production [125,154–156]. Conversely, ADA activity was decreased in prostate and gastric tumors, as well as Hodgkin's lymphoma [157–159]. This highlights the dynamic metabolomic demands of cancer tissues and perturbation of adenosine metabolic processes in cancer.

Increased production of extracellular adenosine promotes both cancer cell proliferation and suppression of immune cells in the TME. There is abundant evidence that ENTPD1 and NT5E expression are increased in cancer including in gastric cancer, non-small cell lung cancer and prostate cancer [160–163]. ENTPD1 is elevated in intratumoral immune cells of non-small cell lung cancer [164]. Adult T-Cell leukemia/lymphoma cells that express high levels of ENTPD1 effectively evade antitumor immunity [165]. Similar findings have been reported in human follicular lymphoma, glioblastoma multiforme, breast cancer, rectal adenocarcinoma, non-small cell lung cancer, head and neck cancer, and medulloblastoma [166–172]. In immunocompetent rats, a NT5E siRNA was used to assess NT5E contribution to immune cell evasion and tumor growth in glioblastoma. In this model, decreased T lymphocyte infiltration and tumor cell apoptosis was observed [173]. Similarly, knockdown of ENTPD1 (CD39) in cancer cells and in mice yielded lower ATP consumption and increased T-Cell infiltration in tumors [174]. Additionally, a meta-analysis of 13 studies analyzing the prognostic value of NT5E, indicates NT5E expression correlates with poor overall survival and disease-free survival in solid-tumor cancer patients [175]. ENTPD1 and NT5E, as important regulators of extracellular adenosine availability, facilitate evasion of the immune system by cancer cells. These findings illustrate widespread upregulation of ATP-adenosine levels and metabolism by ectonucleotidases, resulting in cancer cell proliferation and immunosuppression in cancer. Targeting elevated adenosine levels and ATP-adenosine metabolism, may increase immune response in cancer.

Extracellular adenosine binds to the A_{2A} receptor expressed on macrophages, monocytes, dendritic cells, natural killer cells and T-Cells [41,123,176–178] and the A_{2B} receptor on macrophages and dendritic cells [96,179]. This increases intracellular cAMP leading to

cAMP/PKA signaling that suppresses T-cell response [180]. However, adenosine receptor activation has pleiotropic effects on other immune cells. It prevents dendritic cell activation but also increases the release of VEGF, a signaling protein that promotes angiogenesis thereby promoting tumor growth [96,181]. In tissues where adenosine concentrations are low, adenosine activation of A₁Rs can enhance neutrophil activity [85]. Adenosine can also induce chemotaxis in immature dendritic cells via A₁R activation [182], whereas mature dendritic cells are sensitive to adenosine-mediated inhibition through the A₁ receptor [181]. Overall, adenosine receptor activation has an immunosuppressive effect on different mature (and immature) immune cells and aids the progression of cancer by facilitating immune-surveillance evasion.

All adenosine receptors subtypes seem to contribute to cancer cell proliferation via activation of different kinase signaling pathways including AKT, ERK1/2, p38, JNK and PKC- δ pathways [42,183–189]. Colon cancer cells, where A_{2A} and A₃ are the most abundantly expressed receptor subtypes, undergo increased cell proliferation when treated with high (micromolar) concentrations of adenosine [189]. The A_{2A}R contributes to angiogenesis and wound healing indicating that the downstream targets of adenosine receptors, including ERK, JNK, p38, and AKT, contribute to tumor growth via multiple signaling pathways [190,191]. Bladder urothelial carcinoma tissues and cells express A_{2B}R at higher levels than other adenosine receptor subtypes. High expression of A_{2B}R is correlated with poor prognosis, and suppression of A_{2B}R inhibited proliferation, invasion, and migration of bladder urothelial carcinoma cells with arrest at the G1 phase [188]. Oral squamous cell carcinoma cells (OSCC) are inhibited following knockdown of A_{2B}R. Interestingly, the P2 purinergic receptor P2X7, which is expressed on tumor-associated immune cells, also contributes to immunosuppression of cancer in the tumor microenvironment [192–195]. Additionally, knockdown of the P2X7 receptor suppresses TGF β -1 induced cell migration in A549 lung cancer cells and actin remodeling, illustrating that activation of P2X7 contributes to cell migration [196].

In summary, purinergic signaling is important to cancer cell progression and immune response through a variety of pathways. Elevated extracellular ATP acts on P2 receptors, which are abundantly expressed in many cancer subtypes and immune cells which contribute to increased cancer cell proliferation and immune evasion. ATP is metabolized to adenosine which acts on P1 receptors, whose immunosuppressive role similarly helps tumors evade immune cells.

9. Cancer: Adenosine System as a Therapeutic Target in Cancer

A number of different anti-cancer therapies that target the adenosine system are currently in development (Table 3). Small molecules (See drug formulas Table 3 and structures in Supplementary Figure 1) have been developed to target the adenosine receptors to improve cancer immune response and inhibit proliferation and metastasis of cancer cells. A_{2A} receptor antagonists currently in clinical trials include compounds CPI-444, MK-3814, NIR178, and AZD4635 that are administered as monotherapies or in combination with PD-1 and PD-L1 antagonists [197]. A_{2A}R antagonists like AZD-4635, CPI-444, and NIR-178 are under investigation for their ability to improve T-Cell infiltration in tumors. Lesser studied A_{2B} and A₃ receptor antagonists, like PBF-1129 and CF-102, respectively, are also undergoing clinical trials (Table 3, Figure 1).

A promising but relatively understudied strategy is targeting adenosine-generating enzymes. Due to their higher specificity and longer half-life, monoclonal antibody-directed chemotherapies are used to target the adenosine generating enzyme NT5E (see Table 3) [198,199]. Internalization of NT5E following treatment with monoclonal antibody therapies prevents adenosine-mediated action on tumors, as well as preventing metastasis [199]. Therapies targeting ENTPD1 are predominately still exploratory. By reducing adenosine availability in the extracellular space, ENTPD1-targeted therapies seek to prevent metastasis and improve cancer immune response by restoring T-reg function [200–202]. In breast cancer, reducing

ENTPD1 activity and adenosine generation prevents A_{2B} receptor activation, reducing metastasis-inducing transcription factor FRA1/FOSL1 expression [203].

Adenosine system targeting cancer therapies currently being investigated in clinical trials highlight the importance of elucidating the role of adenosine system dysfunction in the development of novel cancer therapeutics.

Table 3. Drugs targeting the adenosine system in clinical trials for cancer and schizophrenia.

Target	Drug Name	Phase	Indication	Case Number	Formulas	Combination Therapy
NT5E	BMS-986179	I, II	Advanced solid tumors	NCT02754141	Monocolonal Ab	Nivolumab, rHuPH20
	CPI-006	I, Ib	Advanced solid tumors	NCT03454451	Monocolonal Ab	Ciforadenant, Pembrolizumab
	MEDI-9447 (OLECLUMAB)	I, II	Advanced solid tumors	NCT03611556	Monocolonal Ab	durvalumab, gemcitabine, nab-paclitaxel, oxaliplatin, leucovorin, 5-FU
		I, II	Advanced solid tumors	NCT03381274		Osimertinib, AZD4635
		I, II	Advanced solid tumors	NCT03616886		Durvalumab, Carboplatin, Paclitaxel
		II	Advanced solid tumors	NCT03267589		Durvalumab, Tremelimumab, MEDI0562
		II	Breast cancer	NCT03875573		Durvalumab, Oleclumab
		I, II	TNBC	NCT03742102		Durvalumab, Capivasertib, Oleclumab, Paclitaxel, Trastuzumab deruxtecan
	II	NCSLC	NCT03334617		Durvalumab, AZD9150, AZD6738, Vistusertib, Olaparib, Oleclumab, Trastuzumab deruxtecan, cediranib	
A2AR	AZD-4635	II	Prostate, mCRPC NCSLC	NCT04089553 NCT03381274	C15H11CIFN5	Oleclumab, Durvalumab Osimertinib, MEDI9447
	CPI-444	I, II	NCSLC	NCT03337698	C20H21N7O3	Atezolizumab, Cobimetinib, RO6958688, Pemetrexed, Carboplatin, Linagliptin, Tocilizumab, Ipatasertib,
	NIR-178	I, II	Solid tumors	NCT03207867	C10H8BrN7	Idasanutlin PDR001
	Caffeine Pentoxifylline	N/A I	Schizophrenia Schizophrenia	NCT02403193 NCT02832401 NCT04094207	C8H10N4O2 C13H18N4O3	N/A N/A
A3R	CF-102	Complete	Hepatocellular Carcinoma	NCT00790218	C18H18CIIN6O4	CI-IB-MECA
		II	NASH	NCT02927314		Placebo
		I, II	Hepatitis C	NCT00790673		Placebo
		II	Hepatocellular Carcinoma	NCT02128958		Placebo

mCRPC Metastatic Castration-Resistant Prostate Cancer, NASH Non-alcoholic steatohepatitis NCSLC Non-Small Cell Lung Cancer, TNBC Triple-Negative Breast Cancer.

10. Summary and Conclusions

One of the most surprising findings from epidemiological studies of schizophrenia is the reportedly low incidence of cancer relative to the general population. Conflicting reports of cancer rates in schizophrenia have been published for decades, even though this population are considered at increased risk of developing cancer, due to the prevalence of risk factors like smoking and access to medical care [4]. Such controversial findings lend support for the idea that understanding the pathophysiology of schizophrenia may offer unique insight into the etiology of cancer.

Overall, there is a paucity of data available on cancer-related markers in schizophrenia patients who develop cancer. The available literature on this unique population has breadth but lacks depth, as the publicly available epidemiological data sets are limited to diagnoses and procedures (e.g., National Inpatient Sample). Several studies consider genetic risk variants that may be relevant to schizophrenia and cancer, but there is little data available from clinical laboratory tests reporting on biological markers for cancer in schizophrenia patients [204–206]. Thus, we consider reports from postmortem and clinical studies that assess the purinergic system in these disorders.

The adenosine system is hyperfunctional in cancer, facilitating tumor growth, survival and proliferation while evidence suggests this system is hypofunctional in schizophrenia. While it is challenging to determine if hypofunction of the adenosine system in the brain in

schizophrenia extends systemically (and may impact cancer development throughout the body), corresponding changes in peripheral uric acid levels in a subset of the schizophrenia population suggest that this may be the case. Perturbation of ATP and adenosine metabolism, resulting in reduced availability of extracellular adenosine and dysregulated purinergic receptor activation, may attenuate the pathological oncogenic processes driven by elevated purine levels. Adenosine and ATP contribute to tumorigenesis through P1 and P2 receptor mediated signaling. Importantly, several P1 receptor antagonists are under investigation as therapies for different cancers. In schizophrenia, modulating adenosine catabolism, rather than directly targeting P1 receptors (due to associated side effects) is a moderately successful strategy at alleviating symptoms of schizophrenia. This approach is also used in cancer treatment, and drugs targeting ENTPD and NT5E are under exploration.

Much work is required to determine why and how a schizophrenia diagnosis appears to protect against the development of cancer. Schizophrenia and cancer are complex, heterogeneous disorders; multiple genetic and environmental factors interact leading to the onset of these disorders. Perturbation of neurobiological processes, like the adenosine system, may underlie the lower incidence of cancer reported in some schizophrenia patients, and lends support for targeting this system therapeutically in these disorders.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ijms231911835/s1>.

Author Contributions: Conceptualization, A.-R.H. and S.M.O.; writing—original draft preparation, A.-R.H., K.B., O.K., N.H., X.W. and S.M.O.; writing—review and editing, A.-R.H., K.B., R.E.M. and S.M.O.; supervision, S.M.O. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by RO1 MH107487 and MH121102 (REM). This project was supported by Grant YIG-1-139-20 awarded to SMOD from the American Foundation for Suicide Prevention. The content is solely the responsibility of the authors and does not necessarily represent the official views of the American Foundation for Suicide Prevention.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. HMSO; The Board of Control of the Commissioners in Lunacy. *Sixty-Fourth Report of the Commissioners in Lunacy to the Lord Chancellor*; His Majesty's Stationery Office: London, UK, 1910.
2. Berchuck, J.E.; Meyer, C.S.; Zhang, N.; Berchuck, C.M.; Trivedi, N.N.; Cohen, B.; Wang, S. Association of Mental Health Treatment with Outcomes for US Veterans Diagnosed with Non-Small Cell Lung Cancer. *JAMA Oncol.* **2020**, *6*, 1055–1062. [[CrossRef](#)] [[PubMed](#)]
3. Bushe, C.J.; Hodgson, R. Schizophrenia and cancer: In 2010 do we understand the connection? *Can. J. Psychiatry* **2010**, *55*, 761–767. [[CrossRef](#)] [[PubMed](#)]
4. Catts, V.S.; Catts, S.V.; O'Toole, B.I.; Frost, A.D. Cancer incidence in patients with schizophrenia and their first-degree relatives—A meta-analysis. *Acta Psychiatr. Scand.* **2008**, *117*, 323–336. [[CrossRef](#)] [[PubMed](#)]
5. Li, H.; Li, J.; Yu, X.; Zheng, H.; Sun, X.; Lu, Y.; Zhang, Y.; Li, C.; Bi, X. The incidence rate of cancer in patients with schizophrenia: A meta-analysis of cohort studies. *Schizophr. Res.* **2018**, *195*, 519–528. [[CrossRef](#)]
6. Mortensen, P.B. The occurrence of cancer in first admitted schizophrenic patients. *Schizophr. Res.* **1994**, *12*, 185–194. [[CrossRef](#)]
7. Mortensen, P.B. The incidence of cancer in schizophrenic patients. *J. Epidemiol. Community Health* **1989**, *43*, 43–47. [[CrossRef](#)]
8. Goldacre, M.J.; Kurina, L.M.; Wotton, C.J.; Yeates, D.; Seagroatt, V. Schizophrenia and cancer: An epidemiological study. *Br. J. Psychiatry* **2005**, *187*, 334–338. [[CrossRef](#)]
9. Lichtermann, D.; Ekelund, J.; Pukkala, E.; Tanskanen, A.; Lonnqvist, J. Incidence of cancer among persons with schizophrenia and their relatives. *Arch. Gen. Psychiatry* **2001**, *58*, 573–578. [[CrossRef](#)]
10. Xu, D.; Chen, G.; Kong, L.; Zhang, W.; Hu, L.; Chen, C.; Li, J.; Zhuo, C. Lower risk of liver cancer in patients with schizophrenia: A systematic review and meta-analysis of cohort studies. *Oncotarget* **2017**, *8*, 102328–102335. [[CrossRef](#)]
11. Zhuo, C.; Zhuang, H.; Gao, X.; Triplett, P.T. Lung cancer incidence in patients with schizophrenia: Meta-analysis. *Br. J. Psychiatry* **2019**, *215*, 704–711. [[CrossRef](#)]

12. Hippisley-Cox, J.; Vinogradova, Y.; Coupland, C.; Parker, C. Risk of Malignancy in Patients with Schizophrenia or Bipolar Disorder: Nested Case-Control Study. *Arch. Gen. Psychiatry* **2007**, *64*, 1368–1376. [[CrossRef](#)]
13. Osborn, D.P.; Limburg, H.; Walters, K.; Petersen, I.; King, M.; Green, J.; Watson, J.; Nazareth, I. Relative incidence of common cancers in people with severe mental illness. Cohort study in the United Kingdom THIN primary care database. *Schizophr. Res.* **2013**, *143*, 44–49. [[CrossRef](#)]
14. Lasser, K.; Boyd, J.W.; Woolhandler, S.; Himmelstein, D.U.; McCormick, D.; Bor, D.H. Smoking and mental illness: A population-based prevalence study. *J. Am. Med. Assoc.* **2000**, *284*, 2606–2610. [[CrossRef](#)]
15. Shinozaki, Y.; Nakao, M.; Takeuchi, T.; Yano, E. Smoking rates among schizophrenia patients in Japan. *Psychiatry Res.* **2011**, *186*, 165–169. [[CrossRef](#)]
16. Campo-Arias, A.; Diaz-Martinez, L.A.; Rueda-Jaimes, G.E.; Rueda-Sanchez, M.; Farelo-Palacin, D.; Diaz, F.J.; de Leon, J. Smoking is associated with schizophrenia, but not with mood disorders, within a population with low smoking rates: A matched case-control study in Bucaramanga, Colombia. *Schizophr. Res.* **2006**, *83*, 269–276. [[CrossRef](#)]
17. Murphy, K.A.; Stone, E.M.; Presskreischer, R.; McGinty, E.E.; Daumit, G.L.; Pollack, C.E. Cancer Screening Among Adults with and without Serious Mental Illness: A Mixed Methods Study. *Med. Care* **2021**, *59*, 327–333. [[CrossRef](#)]
18. Hodgson, R.; Wildgust, H.J.; Bushe, C.J. Cancer and schizophrenia: Is there a paradox? *J. Psychopharmacol.* **2010**, *24* (Suppl. 4), 51–60. [[CrossRef](#)]
19. Hjorthoj, C.; Sturup, A.E.; McGrath, J.J.; Nordentoft, M. Years of potential life lost and life expectancy in schizophrenia: A systematic review and meta-analysis. *Lancet Psychiatry* **2017**, *4*, 295–301. [[CrossRef](#)]
20. Kisely, S.; Crowe, E.; Lawrence, D. Cancer-related mortality in people with mental illness. *JAMA Psychiatry* **2013**, *70*, 209–217. [[CrossRef](#)]
21. Ni, L.; Wu, J.; Long, Y.; Tao, J.; Xu, J.; Yuan, X.; Yu, N.; Wu, R.; Zhang, Y. Mortality of site-specific cancer in patients with schizophrenia: A systematic review and meta-analysis. *BMC Psychiatry* **2019**, *19*, 323. [[CrossRef](#)]
22. Fares, J.; Fares, M.Y.; Khachfe, H.H.; Salhab, H.A.; Fares, Y. Molecular principles of metastasis: A hallmark of cancer revisited. *Signal Transduct. Target Ther.* **2020**, *5*, 28. [[CrossRef](#)]
23. Xiping, Z.; Shuai, Z.; Feijiang, Y.; Bo, C.; Shifeng, Y.; Qihui, C. Meta-analysis of the Correlation Between Schizophrenia and Breast Cancer. *Clin. Breast Cancer* **2019**, *19*, e172–e185. [[CrossRef](#)]
24. Lu, D.; Song, J.; Lu, Y.; Fall, K.; Chen, X.; Fang, F.; Landen, M.; Hultman, C.M.; Czene, K.; Sullivan, P.; et al. A shared genetic contribution to breast cancer and schizophrenia. *Nat. Commun.* **2020**, *11*, 4637. [[CrossRef](#)]
25. Yuan, K.; Song, W.; Liu, Z.; Lin, G.N.; Yu, S. Mendelian Randomization and GWAS Meta Analysis Revealed the Risk-Increasing Effect of Schizophrenia on Cancers. *Biology* **2022**, *11*, 1345. [[CrossRef](#)]
26. Wootten, J.C.; Wiener, J.C.; Blanchette, P.S.; Anderson, K.K. Cancer incidence and stage at diagnosis among people with psychotic disorders: Systematic review and meta-analysis. *Cancer Epidemiol.* **2022**, *80*, 102233. [[CrossRef](#)]
27. Ge, F.; Huo, Z.; Liu, Y.; Du, X.; Wang, R.; Lin, W.; Wang, R.; Chen, J.; Lu, Y.; Wen, Y.; et al. Association between schizophrenia and prostate cancer risk: Results from a pool of cohort studies and Mendelian randomization analysis. *Compr. Psychiatry* **2022**, *115*, 152308. [[CrossRef](#)]
28. Durand, N.; Sirmsir, M.; Signetti, L.; Labbal, F.; Ballotti, R.; Mus-Veteau, I. Methiothepin Increases Chemotherapy Efficacy against Resistant Melanoma Cells. *Molecules* **2021**, *26*, 1867. [[CrossRef](#)]
29. Matteoni, S.; Matarrese, P.; Ascione, B.; Buccarelli, M.; Ricci-Vitiani, L.; Pallini, R.; Villani, V.; Pace, A.; Paggi, M.G.; Abbruzzese, C. Anticancer Properties of the Antipsychotic Drug Chlorpromazine and Its Synergism with Temozolomide in Restraining Human Glioblastoma Proliferation In Vitro. *Front. Oncol.* **2021**, *11*, 635472. [[CrossRef](#)]
30. Weissenrieder, J.S.; Reed, J.L.; Moldovan, G.L.; Johnson, M.T.; Trebak, M.; Neighbors, J.D.; Mailman, R.B.; Hohl, R.J. Antipsychotic drugs elicit cytotoxicity in glioblastoma multiforme in a calcium-dependent, non-D2 receptor-dependent, manner. *Pharmacol. Res. Perspect.* **2021**, *9*, e00689. [[CrossRef](#)]
31. Zhuo, C.; Xun, Z.; Hou, W.; Ji, F.; Lin, X.; Tian, H.; Zheng, W.; Chen, M.; Liu, C.; Wang, W.; et al. Surprising Anticancer Activities of Psychiatric Medications: Old Drugs Offer New Hope for Patients with Brain Cancer. *Front. Pharmacol.* **2019**, *10*, 1262. [[CrossRef](#)]
32. Ji, J.; Sundquist, K.; Ning, Y.; Kendler, K.S.; Sundquist, J.; Chen, X. Incidence of cancer in patients with schizophrenia and their first-degree relatives: A population-based study in Sweden. *Schizophr. Bull.* **2013**, *39*, 527–536. [[CrossRef](#)] [[PubMed](#)]
33. Platten, M.; Nollen, E.A.A.; Rohrig, U.F.; Fallarino, F.; Opitz, C.A. Tryptophan metabolism as a common therapeutic target in cancer, neurodegeneration and beyond. *Nat. Rev. Drug Discov.* **2019**, *18*, 379–401. [[CrossRef](#)] [[PubMed](#)]
34. Sarkar, C.; Chakraborty, D.; Basu, S. Neurotransmitters as regulators of tumor angiogenesis and immunity: The role of catecholamines. *J. Neuroimmune Pharmacol.* **2013**, *8*, 7–14. [[CrossRef](#)] [[PubMed](#)]
35. Lara, D.R.; Souza, D.O. Schizophrenia: A purinergic hypothesis. *Med. Hypotheses* **2000**, *54*, 157–166. [[CrossRef](#)]
36. Vigano, S.; Alatzoglou, D.; Irving, M.; Menetrier-Caux, C.; Caux, C.; Romero, P.; Coukos, G. Targeting Adenosine in Cancer Immunotherapy to Enhance T-Cell Function. *Front. Immunol.* **2019**, *10*, 925. [[CrossRef](#)]
37. Shen, H.Y.; Singer, P.; Lytle, N.; Wei, C.J.; Lan, J.Q.; Williams-Karnesky, R.L.; Chen, J.F.; Yee, B.K.; Boison, D. Adenosine augmentation ameliorates psychotic and cognitive endophenotypes of schizophrenia. *J. Clin. Investig.* **2012**, *122*, 2567–2577. [[CrossRef](#)]
38. Dunwiddie, T.V.; Masino, S.A. The role and regulation of adenosine in the central nervous system. *Annu. Rev. Neurosci.* **2001**, *24*, 31–55. [[CrossRef](#)]

39. Boison, D.; Singer, P.; Shen, H.Y.; Feldon, J.; Yee, B.K. Adenosine hypothesis of schizophrenia—Opportunities for pharmacotherapy. *Neuropharmacology* **2012**, *62*, 1527–1543. [[CrossRef](#)]
40. Blay, J.; White, T.D.; Hoskin, D.W. The extracellular fluid of solid carcinomas contains immunosuppressive concentrations of adenosine. *Cancer Res.* **1997**, *57*, 2602–2605.
41. Cekic, C.; Day, Y.J.; Sag, D.; Linden, J. Myeloid expression of adenosine A2A receptor suppresses T and NK cell responses in the solid tumor microenvironment. *Cancer Res.* **2014**, *74*, 7250–7259. [[CrossRef](#)]
42. Gessi, S.; Bencivenni, S.; Battistello, E.; Vincenzi, F.; Colotta, V.; Catarzi, D.; Varano, F.; Merighi, S.; Borea, P.A.; Varani, K. Inhibition of A2A Adenosine Receptor Signaling in Cancer Cells Proliferation by the Novel Antagonist TP455. *Front. Pharmacol.* **2017**, *8*, 888. [[CrossRef](#)]
43. Huang, Y.; Gu, Z.; Fan, Y.; Zhai, G.; Zhao, X.; Sun, Q.; Shi, Y.; Lin, G. Inhibition of the adenosinergic pathway: The indispensable part of oncological therapy in the future. *Purinergic Signal.* **2019**, *15*, 53–67. [[CrossRef](#)]
44. Boswell-Casteel, R.C.; Hays, F.A. Equilibrative nucleoside transporters—A review. *Nucleosides Nucleotides Nucleic Acids* **2017**, *36*, 7–30. [[CrossRef](#)]
45. Cristalli, G.; Costanzi, S.; Lambertucci, C.; Lupidi, G.; Vittori, S.; Volpini, R.; Camaioni, E. Adenosine deaminase: Functional implications and different classes of inhibitors. *Med. Res. Rev.* **2001**, *21*, 105–128. [[CrossRef](#)]
46. Zimmermann, H.; Zebisch, M.; Strater, N. Cellular function and molecular structure of ecto-nucleotidases. *Purinergic Signal.* **2012**, *8*, 437–502. [[CrossRef](#)]
47. Li, A. Mechanisms of ATP release, the enabling step in purinergic dynamics. *Cell. Physiol. Biochem.* **2011**, *28*, 1135–1144. [[CrossRef](#)]
48. Lohman, A.W. Mechanisms of ATP release and signaling in the blood vessel wall. *Cardiovasc. Res.* **2012**, *95*, 269–280. [[CrossRef](#)]
49. Lazarowski, E.R. Molecular mechanisms of purine and pyrimidine nucleotide release. *Adv. Pharmacol.* **2011**, *61*, 2211–2261.
50. Chavan, V.; Willis, J.; Walker, S.K.; Clark, H.R.; Liu, X.; Fox, M.A.; Srivastava, S.; Mukherjee, K. Correction: Central Presynaptic Terminals Are Enriched in ATP but the Majority Lack Mitochondria. *PLoS ONE* **2017**, *12*, e0181140. [[CrossRef](#)]
51. Lindberg, D.; Shan, D.; Ayers-Ringler, J.; Oliveros, A.; Benitez, J.; Prieto, M.; McCullumsmith, R.; Choi, D.S. Purinergic signaling and energy homeostasis in psychiatric disorders. *Curr. Mol. Med.* **2015**, *15*, 275–295. [[CrossRef](#)]
52. Mizumoto, N.; Kumamoto, T.; Robson, S.C.; Sévigny, J.; Matsue, H.; Enjyoji, K.; Takashima, A. CD39 is the dominant Langerhans cell-associated ecto-NTPDase: Modulatory roles in inflammation and immune responsiveness. *Nat. Med.* **2002**, *8*, 358–365. [[CrossRef](#)] [[PubMed](#)]
53. Tozzi, M.G.; Pesi, R.; Allegrini, S. On the physiological role of cytosolic 5'-nucleotidase II (cN-II): Pathological and therapeutical implications. *Curr. Med. Chem.* **2013**, *34*, 4285–4291. [[CrossRef](#)] [[PubMed](#)]
54. Romio, M.; Reinbeck, B.; Bongardt, S.; Hüls, S.; Burghoff, S.; Schrader, J. Extracellular purine metabolism and signaling of CD73-derived adenosine in murine Treg and Teff cells. *Am. J. Physiol. Cell Physiol.* **2011**, *301*, 530–539. [[CrossRef](#)] [[PubMed](#)]
55. Robson, S. The E-NTPDase family of ectonucleotidases: Structure function relationships and pathophysiological significance. *Purinergic Signal.* **2006**, *2*, 409–430. [[CrossRef](#)]
56. Cekic, G. Purinergic regulation of the immune system. *Nature Rev. Immunol.* **2016**, *16*, 177–192. [[CrossRef](#)]
57. Friedman, D.J.; Kunzli, B.M.; Yi, A.R.; Sevigny, J.; Berberat, P.O.; Enjyoji, K.; Csizmadia, E.; Friess, H.; Robson, S.C. From the Cover: CD39 deletion exacerbates experimental murine colitis and human polymorphisms increase susceptibility to inflammatory bowel disease. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 16788–16793. [[CrossRef](#)]
58. Louis, N.A.; Robinson, A.M.; MacManus, C.F.; Karhausen, J.; Scully, M.; Colgan, S.P. Control of IFN- α A by CD73: Implications for mucosal inflammation. *J. Immunol.* **2008**, *180*, 4246–4255. [[CrossRef](#)]
59. Yegutkin, G.G. Nucleotide- and nucleoside-converting ectoenzymes: Important modulators of purinergic signalling cascade. *Biochim. Biophys. Acta—Mol. Cell Res.* **2008**, *1783*, 673–694. [[CrossRef](#)]
60. Kopp, R.; Krautloher, A.; Ramirez-Fernandez, A.; Nicke, A. P2 \times 7 Interactions and Signaling—Making Head or Tail of It. *Front. Mol. Neurosci.* **2019**, *12*, 183. [[CrossRef](#)]
61. Samways, D.S.; Li, Z.; Egan, T.M. Principles and properties of ion flow in P2X receptors. *Front. Cell. Neurosci.* **2014**, *8*, 6. [[CrossRef](#)]
62. Ormond, S.J.; Barrera, N.P.; Qureshi, O.S.; Henderson, R.M.; Edwardson, J.M.; Murrell-Lagnado, R.D. An uncharged region within the N terminus of the P2 \times 6 receptor inhibits its assembly and exit from the endoplasmic reticulum. *Mol. Pharmacol.* **2006**, *69*, 1692–1700. [[CrossRef](#)]
63. Jeong, Y.H.; Walsh, M.C.; Yu, J.; Shen, H.; Wherry, E.J.; Choi, Y. Mice Lacking the Purinergic Receptor P2 \times 5 Exhibit Defective Inflammasome Activation and Early Susceptibility to *Listeria monocytogenes*. *J. Immunol.* **2020**, *205*, 760–766. [[CrossRef](#)]
64. Stokes, L.; Layhadi, J.A.; Bibic, L.; Dhuna, K.; Fountain, S.J. P2 \times 4 Receptor Function in the Nervous System and Current Breakthroughs in Pharmacology. *Front. Pharmacol.* **2017**, *8*, 291. [[CrossRef](#)]
65. Fabbretti, E. ATP P2 \times 3 receptors and neuronal sensitization. *Front. Cell. Neurosci.* **2013**, *7*, 236. [[CrossRef](#)]
66. Richler, E.; Shigetomi, E.; Khakh, B.S. Neuronal P2 \times 2 receptors are mobile ATP sensors that explore the plasma membrane when activated. *J. Neurosci.* **2011**, *31*, 16716–16730. [[CrossRef](#)]
67. Mahaut-Smith, M.P.; Jones, S.; Evans, R.J. The P2 \times 1 receptor and platelet function. *Purinergic Signal.* **2011**, *7*, 341–356. [[CrossRef](#)]
68. Erb, L.; Weisman, G.A. Coupling of P2Y receptors to G proteins and other signaling pathways. *Wiley Interdiscip. Rev. Membr. Transp. Signal.* **2012**, *1*, 789–803. [[CrossRef](#)]
69. Olah, M.E. Adenosine receptor subtypes: Characterization and therapeutic regulation. *Annu. Rev. Pharmacol. Toxicol.* **1995**, *35*, 581–606. [[CrossRef](#)]

70. Ralevic, V.; Burnstock, G. Receptors for purines and pyrimidines. *Pharmacol. Rev.* **1998**, *50*, 413–492.
71. Klotz, K.N.; Hessling, J.; Hegler, J.; Owman, C.; Kull, B.; Fredholm, B.B.; Lohse, M.J. Comparative pharmacology of human adenosine receptor subtypes—Characterization of stably transfected receptors in CHO cells. *Naunyn-Schmiedeberg Arch. Pharmacol.* **1998**, *357*, 1–9. [[CrossRef](#)]
72. Fredholm, B.B.; Irenius, E.; Kull, B.; Schulte, G. Comparison of the potency of adenosine as an agonist at human adenosine receptors expressed in Chinese hamster ovary cells. *Biochem. Pharmacol.* **2001**, *61*, 443–448. [[CrossRef](#)]
73. Schulte, G.; Fredholm, B.B. Human adenosine A(1), A(2A), A(2B), and A(3) receptors expressed in Chinese hamster ovary cells all mediate the phosphorylation of extracellular-regulated kinase 1/2. *Mol. Pharmacol.* **2000**, *58*, 477–482. [[CrossRef](#)] [[PubMed](#)]
74. Haskó, G. Adenosine receptor signaling in the brain immune system. *Trends Pharmacol. Sci.* **2005**, *26*, 511–516. [[CrossRef](#)] [[PubMed](#)]
75. Augusto, E.; Matos, M.; Seigny, J.; El-Tayeb, A.; Bynoe, M.S.; Muller, C.E.; Cunha, R.A.; Chen, J.F. Ecto-5'-nucleotidase (CD73)-mediated formation of adenosine is critical for the striatal adenosine A2A receptor functions. *J. Neurosci.* **2013**, *33*, 11390–11399. [[CrossRef](#)]
76. Rebola, N.; Lujan, R.; Cunha, R.A.; Mulle, C. Adenosine A2A receptors are essential for long-term potentiation of NMDA-EPSCs at hippocampal mossy fiber synapses. *Neuron* **2008**, *57*, 121–134. [[CrossRef](#)]
77. Cunha, R.A.; Correia-de-Sa, P.; Sebastiao, A.M.; Ribeiro, J.A. Preferential activation of excitatory adenosine receptors at rat hippocampal and neuromuscular synapses by adenosine formed from released adenine nucleotides. *Br. J. Pharmacol.* **1996**, *119*, 253–260. [[CrossRef](#)]
78. Cunha, R.A. How does adenosine control neuronal dysfunction and neurodegeneration? *J. Neurochem.* **2016**, *139*, 1019–1055. [[CrossRef](#)]
79. Cunha, R.A. Neuroprotection by adenosine in the brain: From A(1) receptor activation to A (2A) receptor blockade. *Purinergic Signal.* **2005**, *1*, 111–134. [[CrossRef](#)]
80. Lopes, C.R.; Lourenco, V.S.; Tome, A.R.; Cunha, R.A.; Canas, P.M. Use of knockout mice to explore CNS effects of adenosine. *Biochem. Pharmacol.* **2021**, *187*, 114367. [[CrossRef](#)]
81. Agnati, L.F.; Ferre, S.; Lluís, C.; Franco, R.; Fuxe, K. Molecular mechanisms and therapeutical implications of intramembrane receptor/receptor interactions among heptahelical receptors with examples from the striatopallidal GABA neurons. *Pharmacol. Rev.* **2003**, *55*, 509–550. [[CrossRef](#)]
82. Ferre, S.; von Euler, G.; Johansson, B.; Fredholm, B.B.; Fuxe, K. Stimulation of high-affinity adenosine A2 receptors decreases the affinity of dopamine D2 receptors in rat striatal membranes. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 7238–7241. [[CrossRef](#)]
83. Ferre, S.; Fredholm, B.B.; Morelli, M.; Popoli, P.; Fuxe, K. Adenosine-dopamine receptor-receptor interactions as an integrative mechanism in the basal ganglia. *Trends Neurosci.* **1997**, *20*, 482–487. [[CrossRef](#)]
84. Cervetto, C.; Venturini, A.; Passalacqua, M.; Guidolin, D.; Genedani, S.; Fuxe, K.; Borroto-Esqueda, D.O.; Cortelli, P.; Woods, A.; Maura, G.; et al. A2A-D2 receptor-receptor interaction modulates gliotransmitter release from striatal astrocyte processes. *J. Neurochem.* **2017**, *140*, 268–279. [[CrossRef](#)]
85. Haskó, G.; Cronstein, B.N. Adenosine: An endogenous regulator of innate immunity. *Trends Immunol.* **2004**, *25*, 33–39. [[CrossRef](#)]
86. Panther, E.; Corinti, S.; Idzko, M.; Herouy, Y.; Napp, M.; La Sala, A.; Girolomoni, G.; Norgauer, J. Adenosine affects expression of membrane molecules, cytokine and chemokine release, and the T-cell stimulatory capacity of human dendritic cells. *Blood* **2003**, *101*, 3985–3990. [[CrossRef](#)]
87. Chekeni, F.B.; Elliott, M.R.; Sandilos, J.K.; Walk, S.F.; Kinchen, J.M.; Lazarowski, E.R.; Armstrong, A.J.; Penuela, S.; Laird, D.W.; Salvesen, G.S.; et al. Pannexin 1 channels mediate 'find-me' signal release and membrane permeability during apoptosis. *Nature* **2010**, *467*, 863–867. [[CrossRef](#)]
88. Krysko, D.V.; Garg, A.D.; Kaczmarek, A.; Krysko, O.; Agostinis, P.; Vandenabeele, P. Immunogenic cell death and DAMPs in cancer therapy. *Nat. Rev. Cancer* **2012**, *12*, 860–875. [[CrossRef](#)]
89. Dosch, M. Mechanisms of ATP Release by Inflammatory Cells. *Int. J. Mol. Sci.* **2018**, *19*, 1222. [[CrossRef](#)]
90. Borea, P.A.; Gessi, S.; Merighi, S.; Varani, K. Adenosine as a multi-signalling guardian angel in human diseases: When, where and how does it exert its protective effects? *Trends Pharmacol. Sci.* **2016**, *37*, 419–434. [[CrossRef](#)]
91. Ernst, P.B.; Garrison, J.C.; Thompson, L.F. Much ado about adenosine: Adenosine synthesis and function in regulatory T cell biology. *J. Immunol.* **2010**, *185*, 1993–1998. [[CrossRef](#)]
92. Di Virgilio, F.; Vuerich, M. Purinergic signaling in the immune system. *Auton. Neurosci.* **2015**, *191*, 117–123. [[CrossRef](#)]
93. Wang, X.; Qin, W.; Xu, X.; Xiong, Y.; Zhang, Y.; Zhang, H.; Sun, B. Endotoxin-induced autocrine ATP signaling inhibits neutrophil chemotaxis through enhancing myosin light chain phosphorylation. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 4483–4488. [[CrossRef](#)]
94. Koshiba, M.; Kojima, H.; Huang, S.; Apasov, S.; Sitkovsky, M.V. Memory of extracellular adenosine A2A purinergic receptor-mediated signalling in murine T cells. *J. Biol. Chem.* **1997**, *272*, 25881–25889. [[CrossRef](#)]
95. Ernens, I. Adenosine up-regulates vascular endothelial growth factor in human macrophages. *Biochem. Biophys. Res. Commun.* **2010**, *392*, 351–356. [[CrossRef](#)]
96. Novitskiy, S.V. Adenosine receptors in regulation of dendritic cell differentiation and function. *Blood* **2008**, *112*, 1822–1831. [[CrossRef](#)]
97. Abbas, M.; Verville, J.; Mroue, K. Resolution of bronchoconstriction with positive airway pressure after intravenous adenosine. *Am. J. Emerg. Med.* **2021**, *44*, e481–e482. [[CrossRef](#)]

98. Shams, S.; Martinez, J.M.; Dawson, J.R.D.; Flores, J.; Gabriel, M.; Garcia, G.; Guevara, A.; Murray, K.; Pacifici, N.; Vargas, M.V.; et al. The Therapeutic Landscape of Rheumatoid Arthritis: Current State and Future Directions. *Front. Pharmacol.* **2021**, *12*, 680043. [[CrossRef](#)]
99. Yao, J.K.; Reddy, R.; van Kammen, D.P. Reduced level of plasma antioxidant uric acid in schizophrenia. *Psychiatry Res.* **1998**, *80*, 29–39. [[CrossRef](#)]
100. Reddy, R.; Keshavan, M.; Yao, J.K. Reduced plasma antioxidants in first-episode patients with schizophrenia. *Schizophr. Res.* **2003**, *62*, 205–212. [[CrossRef](#)]
101. Nagamine, T. Abnormal laboratory values during the acute and recovery phases in schizophrenic patients: A retrospective study. *Neuropsychiatr. Dis. Treat.* **2010**, *6*, 281–288. [[CrossRef](#)]
102. Gültekin, B.K.; Kesebir, S.; Kabak, S.G.; Ergün, F.F.; Yaylaci, E.T. Are Uric Acid Levels Different from Healthy Subjects in Bipolar Affective Disorder and Schizophrenia? Relationship Between Clinical Improvement and Episode Severity in Male Patients. *Nöro Psikiyat. Arşivi* **2014**, *51*, 229–232. [[CrossRef](#)] [[PubMed](#)]
103. He, Q.; You, Y.; Yu, L.; Yao, L.; Lu, H.; Zhou, X.; Wu, S.; Chen, L.; Chen, Y.; Zhao, X. Uric acid levels in subjects with schizophrenia: A systematic review and meta-analysis. *Psychiatry Res.* **2020**, *292*, 113305. [[CrossRef](#)] [[PubMed](#)]
104. Kang, D.H.; Ha, S.K. Uric Acid Puzzle: Dual Role as Anti-oxidant and Pro-oxidant. *Electrolyte Blood Press.* **2014**, *12*, 1–6. [[CrossRef](#)] [[PubMed](#)]
105. Ames, B.N.; Cathcart, R.; Schwiers, E.; Hochstein, P. Uric acid provides an antioxidant defense in humans against oxidant- and radical-caused aging and cancer: A hypothesis. *Proc. Natl. Acad. Sci. USA* **1981**, *78*, 6858–6862. [[CrossRef](#)] [[PubMed](#)]
106. Godin, O.; Leboyer, M.; Gaman, A.; Aouizerate, B.; Berna, F.; Brunel, L.; Capdevielle, D.; Chereau, I.; Dorey, J.M.; Dubertret, C.; et al. Metabolic syndrome, abdominal obesity and hyperuricemia in schizophrenia: Results from the FACE-SZ cohort. *Schizophr. Res.* **2015**, *168*, 388–394. [[CrossRef](#)]
107. Rajan, S.; Zalpuri, I.; Harrington, A.; Cimpeanu, C.; Song, X.; Fan, X. Relationship between serum uric acid level and cardiometabolic risks in nondiabetic patients with schizophrenia. *Int. Clin. Psychopharmacol.* **2016**, *31*, 51–56. [[CrossRef](#)]
108. Flatow, J.; Buckley, P.; Miller, B.J. Meta-analysis of oxidative stress in schizophrenia. *Biol. Psychiatry* **2013**, *74*, 400–409. [[CrossRef](#)]
109. Ermakov, E.A.; Dmitrieva, E.M.; Parshukova, D.A.; Kazantseva, D.V.; Vasilieva, A.R.; Smirnova, L.P. Oxidative Stress-Related Mechanisms in Schizophrenia Pathogenesis and New Treatment Perspectives. *Oxid. Med. Cell. Longev.* **2021**, *2021*, 8881770. [[CrossRef](#)]
110. Arvindakshan, M.; Ghatge, M.; Ranjekar, P.K.; Evans, D.R.; Mahadik, S.P. Supplementation with a combination of omega-3 fatty acids and antioxidants (vitamins E and C) improves the outcome of schizophrenia. *Schizophr. Res.* **2003**, *62*, 195–204. [[CrossRef](#)]
111. Mahadik, S.P.; Pillai, A.; Joshi, S.; Foster, A. Prevention of oxidative stress-mediated neuropathology and improved clinical outcome by adjunctive use of a combination of antioxidants and omega-3 fatty acids in schizophrenia. *Int. Rev. Psychiatry* **2006**, *18*, 119–131. [[CrossRef](#)]
112. Ng, F.; Berk, M.; Dean, O.; Bush, A.I. Oxidative stress in psychiatric disorders: Evidence base and therapeutic implications. *Int. J. Neuropsychopharmacol.* **2008**, *11*, 851–876. [[CrossRef](#)]
113. Yao, J.K.; Condray, R.; Dougherty, G.G., Jr.; Keshavan, M.S.; Montrose, D.M.; Matson, W.R.; McEvoy, J.; Kaddurah-Daouk, R.; Reddy, R.D. Associations between purine metabolites and clinical symptoms in schizophrenia. *PLoS ONE* **2012**, *7*, e42165. [[CrossRef](#)]
114. Ben Salem, C.; Slim, R.; Fathallah, N.; Hmouda, H. Drug-induced hyperuricaemia and gout. *Rheumatology* **2017**, *56*, 679–688. [[CrossRef](#)]
115. Mi, S.; Gong, L.; Sui, Z. Friend or Foe? An Unrecognized Role of Uric Acid in Cancer Development and the Potential Anticancer Effects of Uric Acid-lowering Drugs. *J. Cancer* **2020**, *11*, 5236–5244. [[CrossRef](#)]
116. Yang, J.; Wang, Y.; Zhao, Q.; Zhang, X.; Wang, X.; Qin, X.; Zhang, R.; Shen, L.; Jiang, X.; Jiang, H.; et al. Association of serum uric acid with increased risk of cancer among hypertensive Chinese. *Int. J. Cancer* **2017**, *141*, 112–120. [[CrossRef](#)]
117. Mao, L.; Guo, C.; Zheng, S. Elevated urinary 8-oxo-7,8-dihydro-2'-deoxyguanosine and serum uric acid are associated with progression and are prognostic factors of colorectal cancer. *Onco Targets Ther.* **2018**, *11*, 5895–5902. [[CrossRef](#)]
118. Chen, Y.F.; Li, Q.; Chen, D.T.; Pan, J.H.; Chen, Y.H.; Wen, Z.S.; Zeng, W.A. Prognostic value of pre-operative serum uric acid levels in esophageal squamous cell carcinoma patients who undergo R0 esophagectomy. *Cancer Biomark.* **2016**, *17*, 89–96. [[CrossRef](#)]
119. Yuan, C.; Xu, X.H.; Wang, X.L.; Xu, L.; Chen, Z.; Li, Y.Q. Relationship between serum uric acid and metastatic and nonmetastatic rectal cancer patients with undergoing no chemotherapy. *Medicine* **2016**, *95*, e5463. [[CrossRef](#)]
120. Stotz, M.; Szkandera, J.; Seidel, J.; Stojakovic, T.; Samonigg, H.; Reitz, D.; Gary, T.; Kornprat, P.; Schaberl-Moser, R.; Hoefler, G.; et al. Evaluation of uric acid as a prognostic blood-based marker in a large cohort of pancreatic cancer patients. *PLoS ONE* **2014**, *9*, e104730. [[CrossRef](#)]
121. Stevenson, W.S.; Hyland, C.D.; Zhang, J.G.; Morgan, P.O.; Willson, T.A.; Gill, A.; Hilton, A.A.; Viney, E.M.; Bahlo, M.; Masters, S.L.; et al. Deficiency of 5-hydroxyisourate hydrolase causes hepatomegaly and hepatocellular carcinoma in mice. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 16625–16630. [[CrossRef](#)]
122. Serra, S.; Horenstein, A.L.; Vaisitti, T.; Brusa, D.; Rossi, D.; Laurenti, L.; D'Arena, G.; Coscia, M.; Tripodo, C.; Inghirami, G.; et al. CD73-generated extracellular adenosine in chronic lymphocytic leukemia creates local conditions counteracting drug-induced cell death. *Blood* **2011**, *118*, 6141–6152. [[CrossRef](#)]

123. Ohta, A.; Gorelik, E.; Prasad, S.J.; Ronchese, F.; Lukashev, D.; Wong, M.K.; Huang, X.; Caldwell, S.; Liu, K.; Smith, P.; et al. A2A adenosine receptor protects tumors from antitumor T cells. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 13132–13137. [[CrossRef](#)]
124. Roberts, E.L.; Roberts, O.T. Plasma adenosine deaminase isoform 2 in cancer patients undergoing chemotherapy. *Br. J. Biomed. Sci.* **2012**, *69*, 11–13. [[CrossRef](#)]
125. Aghaei, M.; Karami-Tehrani, F.; Salami, S.; Atri, M. Adenosine deaminase activity in the serum and malignant tumors of breast cancer: The assessment of isoenzyme ADA1 and ADA2 activities. *Clin. Biochem.* **2005**, *38*, 887–891. [[CrossRef](#)] [[PubMed](#)]
126. McGrath, J.; Saha, S.; Welham, J.; El Saadi, O.; MacCauley, C.; Chant, D. A systematic review of the incidence of schizophrenia: The distribution of rates and the influence of sex, urbanicity, migrant status and methodology. *BMC Med.* **2004**, *2*, 13. [[CrossRef](#)] [[PubMed](#)]
127. Saha, S.; Chant, D.; Welham, J.; McGrath, J. A systematic review of the prevalence of schizophrenia. *PLoS Med.* **2005**, *2*, e141. [[CrossRef](#)] [[PubMed](#)]
128. Krugel, U. Purinergic receptors in psychiatric disorders. *Neuropharmacology* **2016**, *104*, 212–225. [[CrossRef](#)] [[PubMed](#)]
129. Shan, D.; Haroutunian, V.; Meador-Woodruff, J.H.; McCullumsmith, R.E. Expression of equilibrative nucleoside transporter type 1 protein in elderly patients with schizophrenia. *Neuroreport* **2012**, *23*, 224–227. [[CrossRef](#)]
130. Aliagas, E.; Villar-Menendez, I.; Sevigny, J.; Roca, M.; Romeu, M.; Ferrer, I.; Martin-Satue, M.; Barrachina, M. Reduced striatal ecto-nucleotidase activity in schizophrenia patients supports the “adenosine hypothesis”. *Purinergic Signal.* **2013**, *9*, 599–608. [[CrossRef](#)]
131. O'Donovan, S.M.; Sullivan, C.; Koene, R.; Devine, E.; Hasselfeld, K.; Moody, C.L.; McCullumsmith, R.E. Cell-subtype-specific changes in adenosine pathways in schizophrenia. *Neuropsychopharmacology* **2018**, *43*, 1667–1674. [[CrossRef](#)]
132. Boison, D. Adenosine kinase: Exploitation for therapeutic gain. *Pharmacol. Rev.* **2013**, *65*, 906–943. [[CrossRef](#)]
133. Villar-Menendez, I.; Diaz-Sanchez, S.; Blanch, M.; Albasanz, J.L.; Pereira-Veiga, T.; Monje, A.; Planchat, L.M.; Ferrer, I.; Martin, M.; Barrachina, M. Reduced striatal adenosine A2A receptor levels define a molecular subgroup in schizophrenia. *J. Psychiatr. Res.* **2014**, *51*, 49–59. [[CrossRef](#)]
134. Villar-Menendez, I.; Porta, S.; Buiira, S.P.; Pereira-Veiga, T.; Diaz-Sanchez, S.; Albasanz, J.L.; Ferrer, I.; Martin, M.; Barrachina, M. Increased striatal adenosine A2A receptor levels is an early event in Parkinson's disease-related pathology and it is potentially regulated by miR-34b. *Neurobiol. Dis.* **2014**, *69*, 206–214. [[CrossRef](#)]
135. Deckert, J.; Brenner, M.; Durany, N.; Zochling, R.; Paulus, W.; Ransmayr, G.; Tatschner, T.; Danielczyk, W.; Jellinger, K.; Riederer, P. Up-regulation of striatal adenosine A(2A) receptors in schizophrenia. *Neuroreport* **2003**, *14*, 313–316. [[CrossRef](#)]
136. Valle-Leon, M.; Callado, L.F.; Aso, E.; Cajiao-Manrique, M.M.; Sahlholm, K.; Lopez-Cano, M.; Soler, C.; Altafaj, X.; Watanabe, M.; Ferre, S.; et al. Decreased striatal adenosine A2A-dopamine D2 receptor heteromerization in schizophrenia. *Neuropsychopharmacology* **2021**, *46*, 665–672. [[CrossRef](#)]
137. Brunstein, M.G.; Ghisolfi, E.S.; Ramos, F.L.; Lara, D.R. A clinical trial of adjuvant allopurinol therapy for moderately refractory schizophrenia. *J. Clin. Psychiatry* **2005**, *66*, 213–219. [[CrossRef](#)]
138. Dickerson, F.B.; Stallings, C.R.; Origoni, A.E.; Sullens, A.; Khushalani, S.; Sandson, N.; Yolken, R.H. A double-blind trial of adjunctive allopurinol for schizophrenia. *Schizophr. Res.* **2009**, *109*, 66–69. [[CrossRef](#)]
139. Akhondzadeh, S.; Shasavand, E.; Jamilian, H.; Shabestari, O.; Kamalipour, A. Dipyridamole in the treatment of schizophrenia: Adenosine-dopamine receptor interactions. *J. Clin. Pharm. Ther.* **2000**, *25*, 131–137. [[CrossRef](#)]
140. Akhondzadeh, S.; Safarcherati, A.; Amini, H. Beneficial antipsychotic effects of allopurinol as add-on therapy for schizophrenia: A double blind, randomized and placebo controlled trial. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2005**, *29*, 253–259. [[CrossRef](#)]
141. Wonodi, I.; Gopinath, H.V.; Liu, J.; Adami, H.; Hong, L.E.; Allen-Emerson, R.; McMahon, R.P.; Thaker, G.K. Dipyridamole monotherapy in schizophrenia: Pilot of a novel treatment approach by modulation of purinergic signaling. *Psychopharmacology* **2011**, *218*, 341–345. [[CrossRef](#)]
142. Weiser, M.; Gershon, A.A.; Rubinstein, K.; Petcu, C.; Ladea, M.; Sima, D.; Podea, D.; Keefe, R.S.; Davis, J.M. A randomized controlled trial of allopurinol vs. placebo added on to antipsychotics in patients with schizophrenia or schizoaffective disorder. *Schizophr. Res.* **2012**, *138*, 35–38. [[CrossRef](#)]
143. Lintunen, J.; Lahteenvuo, M.; Tiihonen, J.; Tanskanen, A.; Taipale, H. Adenosine modulators and calcium channel blockers as add-on treatment for schizophrenia. *NPJ Schizophr.* **2021**, *7*, 1. [[CrossRef](#)]
144. Huang, H.; Zheng, S.; Chen, M.; Xie, L.; Li, Z.; Guo, M.; Wang, J.; Lu, M.; Zhu, X. The potential of the P2X7 receptor as a therapeutic target in a sub-chronic PCP-induced rodent model of schizophrenia. *J. Chem. Neuroanat.* **2021**, *116*, 101993. [[CrossRef](#)]
145. Wardas, J. Potential role of adenosine A2A receptors in the treatment of schizophrenia. *Front. Biosci.-Landmark* **2008**, *13*, 4071–4096. [[CrossRef](#)]
146. Balkwill, F.R.; Capasso, M.; Hagemann, T. The tumor microenvironment at a glance. *J. Cell Sci.* **2012**, *125*, 5591–5596. [[CrossRef](#)]
147. Lawson, D.A.; Kessenbrock, K.; Davis, R.T.; Pervolarakis, N.; Werb, Z. Tumour heterogeneity and metastasis at single-cell resolution. *Nat. Cell Biol.* **2018**, *20*, 1349–1360. [[CrossRef](#)]
148. Plaks, V.; Kong, N.; Werb, Z. The cancer stem cell niche: How essential is the niche in regulating stemness of tumor cells? *Cell Stem Cell* **2015**, *16*, 225–238. [[CrossRef](#)]
149. Takai, K.; Le, A.; Weaver, V.M.; Werb, Z. Targeting the cancer-associated fibroblasts as a treatment in triple-negative breast cancer. *Oncotarget* **2016**, *7*, 82889–82901. [[CrossRef](#)]
150. Di Virgilio, F.; Adinolfi, E. Extracellular purines, purinergic receptors and tumor growth. *Oncogene* **2017**, *36*, 293–303. [[CrossRef](#)]

151. Allard, B.; Longhi, M.S.; Robson, S.C.; Stagg, J. The ectonucleotidases CD39 and CD73: Novel checkpoint inhibitor targets. *Immunol. Rev.* **2017**, *276*, 121–144. [[CrossRef](#)]
152. Ohta, A.; Sitkovsky, M. Role of G-protein-coupled adenosine receptors in downregulation of inflammation and protection from tissue damage. *Nature* **2001**, *414*, 916–920. [[CrossRef](#)] [[PubMed](#)]
153. Vecchio, E.A.; Tan, C.Y.; Gregory, K.J.; Christopoulos, A.; White, P.J.; May, L.T. Ligand-Independent Adenosine A2B Receptor Constitutive Activity as a Promoter of Prostate Cancer Cell Proliferation. *J. Pharmacol. Exp. Ther.* **2016**, *357*, 36–44. [[CrossRef](#)] [[PubMed](#)]
154. Vannoni, D.; Bernini, A.; Carlucci, F.; Civitelli, S.; Di Pietro, M.C.; Leoncini, R.; Rosi, F.; Tabucchi, A.; Tanzini, G.; Marinello, E. Enzyme activities controlling adenosine levels in normal and neoplastic tissues. *Med. Oncol.* **2004**, *21*, 187–195. [[CrossRef](#)] [[PubMed](#)]
155. Mahajan, M.; Tiwari, N.; Sharma, R.; Kaur, S.; Singh, N. Oxidative stress and its relationship with adenosine deaminase activity in various stages of breast cancer. *Indian J. Clin. Biochem.* **2013**, *28*, 51–54. [[CrossRef](#)]
156. Ghaderi, B.; Amini, S.; Maroofi, F.; Jalali, C.; Javanmardi, M.; Roshani, D.; Abdi, M. Adenosine Deaminase Activity in Chronic Lymphocytic Leukemia and Healthy Subjects. *Iran. J. Cancer Prev.* **2016**, *9*, e5069. [[CrossRef](#)]
157. Biri, H.; Ozturk, H.S.; Kacmaz, M.; Karaca, K.; Tokucoglu, H.; Durak, I. Activities of DNA turnover and free radical metabolizing enzymes in cancerous human prostate tissue. *Cancer Investig.* **1999**, *17*, 314–319. [[CrossRef](#)]
158. Namiot, Z.; Stasiewicz, J.; Namiot, A.; Kemon, A.; Kralisz, M.; Gorski, J. Adenosine deaminase activity in patients with the intestinal type of gastric carcinoma. *Cancer Lett.* **1996**, *109*, 199–202. [[CrossRef](#)]
159. Murray, J.L.; Perez-Soler, R.; Bywaters, D.; Hersh, E.M. Decreased adenosine deaminase (ADA) and 5′ nucleotidase (5NT) activity in peripheral blood T cells in Hodgkin disease. *Am. J. Hematol.* **1986**, *21*, 57–66. [[CrossRef](#)]
160. Turcotte, M.; Spring, K.; Pommey, S.; Chouinard, G.; Cousineau, I.; George, J.; Chen, G.M.; Gendoo, D.M.; Haibe-Kains, B.; Karn, T.; et al. CD73 is associated with poor prognosis in high-grade serous ovarian cancer. *Cancer Res.* **2015**, *75*, 4494–4503. [[CrossRef](#)]
161. Cai, X.Y.; Wang, X.F.; Li, J.; Dong, J.N.; Liu, J.Q.; Li, N.P.; Yun, B.; Xia, R.L. Overexpression of CD39 and high tumoral CD39(+)/CD8(+) ratio are associated with adverse prognosis in resectable gastric cancer. *Int. J. Clin. Exp. Pathol.* **2015**, *8*, 14757–14764.
162. Inoue, Y.; Yoshimura, K.; Kurabe, N.; Kahyo, T.; Kawase, A.; Tanahashi, M.; Ogawa, H.; Inui, N.; Funai, K.; Shinmura, K.; et al. Prognostic impact of CD73 and A2A adenosine receptor expression in non-small-cell lung cancer. *Oncotarget* **2017**, *8*, 8738–8751. [[CrossRef](#)]
163. Leclerc, B.G.; Charlebois, R.; Chouinard, G.; Allard, B.; Pommey, S.; Saad, F.; Stagg, J. CD73 Expression Is an Independent Prognostic Factor in Prostate Cancer. *Clin. Cancer Res.* **2016**, *22*, 158–166. [[CrossRef](#)]
164. Tondell, A.; Wahl, S.G.F.; Sponaas, A.M.; Sorhaug, S.; Borset, M.; Haug, M. Ectonucleotidase CD39 and Checkpoint Signalling Receptor Programmed Death 1 are Highly Elevated in Intratumoral Immune Cells in Non-small-cell Lung Cancer. *Transl. Oncol.* **2020**, *13*, 17–24. [[CrossRef](#)]
165. Nagate, Y.; Ezoe, S.; Fujita, J.; Okuzakis, D.; Motooka, D.; Ishibashi, T.; Ichii, M.; Tanimura, A.; Kurashige, M.; Morii, E.; et al. Ectonucleotidase CD39 is highly expressed on ATLL cells and is responsible for their immunosuppressive function. *Leukemia* **2020**, *35*, 107–118. [[CrossRef](#)]
166. Mandapathil, M.; Boduc, M.; Roessler, M.; Guldner, C.; Walliczek-Dworschak, U.; Mandic, R. Ectonucleotidase CD39 expression in regional metastases in head and neck cancer. *Acta Otolaryngol.* **2018**, *138*, 428–432. [[CrossRef](#)]
167. Li, J.; Wang, L.; Chen, X.; Li, L.; Li, Y.; Ping, Y.; Huang, L.; Yue, D.; Zhang, Z.; Wang, F.; et al. CD39/CD73 upregulation on myeloid-derived suppressor cells via TGF-beta-mTOR-HIF-1 signaling in patients with non-small cell lung cancer. *Oncoimmunology* **2017**, *6*, e1320011. [[CrossRef](#)]
168. Zhang, B.; Cheng, B.; Li, F.S.; Ding, J.H.; Feng, Y.Y.; Zhuo, G.Z.; Wei, H.F.; Zhao, K. High expression of CD39/ENTPD1 in malignant epithelial cells of human rectal adenocarcinoma. *Tumour Biol.* **2015**, *36*, 9411–9419. [[CrossRef](#)]
169. Quezada, C.; Garrido, W.; Oyarzun, C.; Fernandez, K.; Segura, R.; Melo, R.; Casanello, P.; Sobrevia, L.; San Martin, R. 5′-ectonucleotidase mediates multiple-drug resistance in glioblastoma multiforme cells. *J. Cell. Physiol.* **2013**, *228*, 602–608. [[CrossRef](#)]
170. Loi, S.; Pommey, S.; Haibe-Kains, B.; Beavis, P.A.; Darcy, P.K.; Smyth, M.J.; Stagg, J. CD73 promotes anthracycline resistance and poor prognosis in triple negative breast cancer. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 11091–11096. [[CrossRef](#)]
171. Cappellari, A.R.; Rockenbach, L.; Dietrich, F.; Clarimundo, V.; Glaser, T.; Braganhol, E.; Abujamra, A.L.; Roesler, R.; Ulrich, H.; Battastini, A.M. Characterization of ectonucleotidases in human medulloblastoma cell lines: Ecto-5′NT/CD73 in metastasis as potential prognostic factor. *PLoS ONE* **2012**, *7*, e47468. [[CrossRef](#)]
172. Hilchey, S.P.; Kobie, J.J.; Cochran, M.R.; Secor-Socha, S.; Wang, J.C.; Hyrien, O.; Burack, W.R.; Mosmann, T.R.; Quataert, S.A.; Bernstein, S.H. Human follicular lymphoma CD39+ infiltrating T cells contribute to adenosine-mediated T cell hyporesponsiveness. *J. Immunol.* **2009**, *183*, 6157–6166. [[CrossRef](#)]
173. Azambuja, J.H.; Schuh, R.S.; Michels, L.R.; Iser, I.C.; Beckenkamp, L.R.; Roliano, G.G.; Lenz, G.S.; Scholl, J.N.; Sevigny, J.; Wink, M.R.; et al. Blockade of CD73 delays glioblastoma growth by modulating the immune environment. *Cancer Immunol. Immunother.* **2020**, *69*, 1801–1812. [[CrossRef](#)]

174. Kashyap, A.S.; Thelemann, T.; Klar, R.; Kallert, S.M.; Festag, J.; Buchi, M.; Hinterwimmer, L.; Schell, M.; Michel, S.; Jaschinski, F.; et al. Antisense oligonucleotide targeting CD39 improves anti-tumor T cell immunity. *J. Immunother. Cancer* **2019**, *7*, 67. [[CrossRef](#)]
175. Wang, R.; Zhang, Y.; Lin, X.; Gao, Y.; Zhu, Y. Prognostic value of CD73-adenosinergic pathway in solid tumor: A meta-analysis and systematic review. *Oncotarget* **2017**, *8*, 57327–57336. [[CrossRef](#)]
176. Waickman, A.T.; Alme, A.; Senaldi, L.; Zarek, P.E.; Horton, M.; Powell, J.D. Enhancement of tumor immunotherapy by deletion of the A2A adenosine receptor. *Cancer Immunol. Immunother.* **2012**, *61*, 917–926. [[CrossRef](#)]
177. Koroskenyi, K.; Kiss, B.; Szondy, Z. Adenosine A2A receptor signaling attenuates LPS-induced pro-inflammatory cytokine formation of mouse macrophages by inducing the expression of DUSP1. *Biochim. Biophys. Acta* **2016**, *1863 Pt A*, 1461–1471. [[CrossRef](#)]
178. Cekic, C.; Linden, J. Adenosine A2A receptors intrinsically regulate CD8+ T cells in the tumor microenvironment. *Cancer Res.* **2014**, *74*, 7239–7249. [[CrossRef](#)]
179. Hasko, G.; Pacher, P.; Deitch, E.A.; Vizi, E.S. Shaping of monocyte and macrophage function by adenosine receptors. *Pharmacol. Ther.* **2007**, *113*, 264–275. [[CrossRef](#)] [[PubMed](#)]
180. Wehbi, V.L.; Tasken, K. Molecular Mechanisms for cAMP-Mediated Immunoregulation in T cells—Role of Anchored Protein Kinase A Signaling Units. *Front. Immunol.* **2016**, *7*, 222. [[CrossRef](#)] [[PubMed](#)]
181. Desrosiers, M.D.; Cembrola, K.M.; Fakir, M.J.; Stephens, L.A.; Jama, F.M.; Shamel, A.; Mehal, W.Z.; Santamaria, P.; Shi, Y. Adenosine deamination sustains dendritic cell activation in inflammation. *J. Immunol.* **2007**, *179*, 1884–1892. [[CrossRef](#)] [[PubMed](#)]
182. Panther, E.; Idzko, M.; Herouy, Y.; Rheinen, H.; Gebicke-Haerter, P.J.; Mrowietz, U.; Dichmann, S.; Norgauer, J. Expression and function of adenosine receptors in human dendritic cells. *FASEB J.* **2001**, *15*, 1963–1970. [[CrossRef](#)]
183. Mirza, A.; Basso, A.; Black, S.; Malkowski, M.; Kwee, L.; Pachter, J.A.; Lachowicz, J.E.; Wang, Y.; Liu, S. RNA interference targeting of A1 receptor-overexpressing breast carcinoma cells leads to diminished rates of cell proliferation and induction of apoptosis. *Cancer Biol. Ther.* **2005**, *4*, 1355–1360. [[CrossRef](#)]
184. Lin, Z.; Yin, P.; Reierstad, S.; O'Halloran, M.; Coon, V.J.; Pearson, E.K.; Mutlu, G.M.; Bulun, S.E. Adenosine A1 receptor, a target and regulator of estrogen receptor α action, mediates the proliferative effects of estradiol in breast cancer. *Oncogene* **2010**, *29*, 1114–1122. [[CrossRef](#)]
185. Merighi, S.; Benini, A.; Mirandola, P.; Gessi, S.; Varani, K.; Leung, E.; Maclennan, S.; Borea, P.A. A3 adenosine receptor activation inhibits cell proliferation via phosphatidylinositol 3-kinase/Akt-dependent inhibition of the extracellular signal-regulated kinase 1/2 phosphorylation in A375 human melanoma cells. *J. Biol. Chem.* **2005**, *280*, 19516–19526. [[CrossRef](#)]
186. Merighi, S.; Mirandola, P.; Milani, D.; Varani, K.; Gessi, S.; Klotz, K.N.; Leung, E.; Baraldi, P.G.; Borea, P.A. Adenosine receptors as mediators of both cell proliferation and cell death of cultured human melanoma cells. *J. Investig. Dermatol.* **2002**, *119*, 923–933. [[CrossRef](#)]
187. Kasama, H.; Sakamoto, Y.; Kasamatsu, A.; Okamoto, A.; Koyama, T.; Minakawa, Y.; Ogawara, K.; Yokoe, H.; Shiiba, M.; Tanzawa, H.; et al. Adenosine A2b receptor promotes progression of human oral cancer. *BMC Cancer* **2015**, *15*, 563. [[CrossRef](#)]
188. Zhou, Y.; Chu, X.; Deng, F.; Tong, L.; Tong, G.; Yi, Y.; Liu, J.; Tang, J.; Tang, Y.; Xia, Y.; et al. The adenosine A2b receptor promotes tumor progression of bladder urothelial carcinoma by enhancing MAPK signaling pathway. *Oncotarget* **2017**, *8*, 48755–48768. [[CrossRef](#)]
189. Gessi, S.; Merighi, S.; Varani, K.; Cattabriga, E.; Benini, A.; Mirandola, P.; Leung, E.; Mac Lennan, S.; Feo, C.; Baraldi, S.; et al. Adenosine receptors in colon carcinoma tissues and colon tumoral cell lines: Focus on the A(3) adenosine subtype. *J. Cell. Physiol.* **2007**, *211*, 826–836. [[CrossRef](#)]
190. Koszalka, P.; Golunska, M.; Urban, A.; Stasiłojc, G.; Stanisławowski, M.; Majewski, M.; Skladanowski, A.C.; Bigda, J. Specific Activation of A3, A2A and A1 Adenosine Receptors in CD73-Knockout Mice Affects B16F10 Melanoma Growth, Neovascularization, Angiogenesis and Macrophage Infiltration. *PLoS ONE* **2016**, *11*, e0151420. [[CrossRef](#)]
191. Allen-Gipson, D.S.; Wong, J.; Spurzem, J.R.; Sisson, J.H.; Wyatt, T.A. Adenosine A2A receptors promote adenosine-stimulated wound healing in bronchial epithelial cells. *Am. J. Physiol. Lung Cell Mol. Physiol.* **2006**, *290*, L849–L855. [[CrossRef](#)]
192. Ghiringhelli, F.; Apetoh, L.; Tesniere, A.; Aymeric, L.; Ma, Y.; Ortiz, C.; Vermaelen, K.; Panaretakis, T.; Mignot, G.; Ullrich, E.; et al. Activation of the NLRP3 inflammasome in dendritic cells induces IL-1 β -dependent adaptive immunity against tumors. *Nat. Med.* **2009**, *15*, 1170–1178. [[CrossRef](#)] [[PubMed](#)]
193. Kan, L.K.; Williams, D.; Drummond, K.; O'Brien, T.; Monif, M. The role of microglia and P2X7 receptors in gliomas. *J. Neuroimmunol.* **2019**, *332*, 138–146. [[CrossRef](#)] [[PubMed](#)]
194. Adinolfi, E.; Capece, M.; Franceschini, A.; Falzoni, S.; Giuliani, A.L.; Rotondo, A.; Sarti, A.C.; Bonora, M.; Syberg, S.; Corigliano, D.; et al. Accelerated tumor progression in mice lacking the ATP receptor P2X7. *Cancer Res.* **2015**, *75*, 635–644. [[CrossRef](#)] [[PubMed](#)]
195. De Marchi, E.; Orioli, E.; Pegoraro, A.; Sangaletti, S.; Portararo, P.; Curti, A.; Colombo, M.P.; Di Virgilio, F.; Adinolfi, E. The P2X7 receptor modulates immune cells infiltration, ectonucleotidases expression and extracellular ATP levels in the tumor microenvironment. *Oncogene* **2019**, *38*, 3636–3650. [[CrossRef](#)]
196. Takai, E.; Tsukimoto, M.; Harada, H.; Sawada, K.; Moriyama, Y.; Kojima, S. Autocrine regulation of TGF- β 1-induced cell migration by exocytosis of ATP and activation of P2 receptors in human lung cancer cells. *J. Cell Sci.* **2012**, *125*, 5051–5060. [[CrossRef](#)]
197. Leone, R.D.; Emens, L.A. Targeting adenosine for cancer immunotherapy. *J. Immunother. Cancer* **2018**, *6*, 57. [[CrossRef](#)]

198. Levy, R. SD-101 and BMS-986178 in Treating Patients with Advanced or Metastatic Solid Malignancies. 2020. Available online: <https://clinicaltrials.gov/ct2/show/NCT03831295> (accessed on 9 August 2022).
199. Terp, M.G.; Olesen, K.A.; Arnsparang, E.C.; Lund, R.R.; Lagerholm, B.C.; Ditzel, H.J.; Leth-Larsen, R. Anti-human CD73 monoclonal antibody inhibits metastasis formation in human breast cancer by inducing clustering and internalization of CD73 expressed on the surface of cancer cells. *J. Immunol.* **2013**, *191*, 4165–4173. [[CrossRef](#)]
200. Yegutkin, G.G.; Marttila-Ichihara, F.; Karikoski, M.; Niemela, J.; Laurila, J.P.; Elima, K.; Jalkanen, S.; Salmi, M. Altered purinergic signaling in CD73-deficient mice inhibits tumor progression. *Eur. J. Immunol.* **2011**, *41*, 1231–1241. [[CrossRef](#)]
201. Stagg, J.; Divisekera, U.; Duret, H.; Sparwasser, T.; Teng, M.W.; Darcy, P.K.; Smyth, M.J. CD73-deficient mice have increased antitumor immunity and are resistant to experimental metastasis. *Cancer Res.* **2011**, *71*, 2892–2900. [[CrossRef](#)]
202. Wang, L.; Zhou, X.; Zhou, T.; Ma, D.; Chen, S.; Zhi, X.; Yin, L.; Shao, Z.; Ou, Z.; Zhou, P. Ecto-5'-nucleotidase promotes invasion, migration and adhesion of human breast cancer cells. *J. Cancer Res. Clin. Oncol.* **2008**, *134*, 365–372. [[CrossRef](#)]
203. Desmet, C.J.; Gallenne, T.; Prieur, A.; Reyat, F.; Visser, N.L.; Wittner, B.S.; Smit, M.A.; Geiger, T.R.; Laoukili, J.; Iskit, S.; et al. Identification of a pharmacologically tractable Fra-1/ADORA2B axis promoting breast cancer metastasis. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 5139–5144. [[CrossRef](#)]
204. Zuber, V.; Jonsson, E.G.; Frei, O.; Witoelar, A.; Thompson, W.K.; Schork, A.J.; Bettella, F.; Wang, Y.; Djurovic, S.; Smeland, O.B.; et al. Identification of shared genetic variants between schizophrenia and lung cancer. *Sci. Rep.* **2018**, *8*, 674. [[CrossRef](#)]
205. Sargazi, S.; Nia, M.H.; Mirinejad, S.; Moudi, M.; Shahroudi, M.J.; Saravani, R.; Valian-Borojeni, S. Association of a Novel KIF26B Gene Polymorphism with Susceptibility to Schizophrenia and Breast Cancer: A Case-Control Study. *Iran. J. Public Health* **2021**, *50*, 397–406. [[CrossRef](#)]
206. Ibanez, K.; Boullosa, C.; Tabares-Seisdedos, R.; Baudot, A.; Valencia, A. Molecular evidence for the inverse comorbidity between central nervous system disorders and cancers detected by transcriptomic meta-analyses. *PLoS Genet.* **2014**, *10*, e1004173. [[CrossRef](#)]