

Plasticity of Naturally Occurring Regulatory T Cells in Allergic Airway Disease is Modulated by the Transcriptional Activity of *Il-6*

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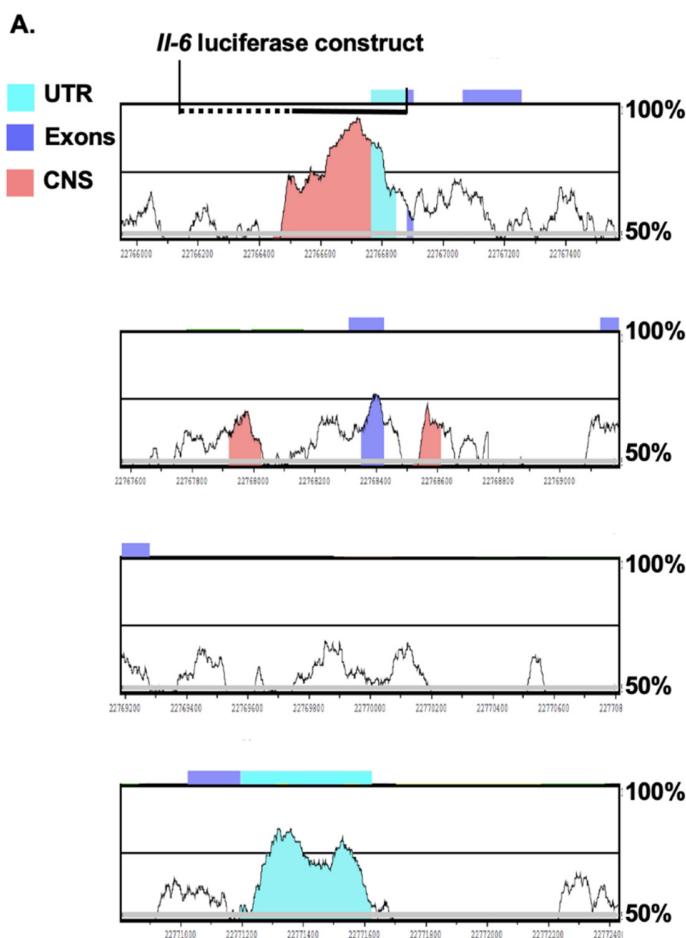
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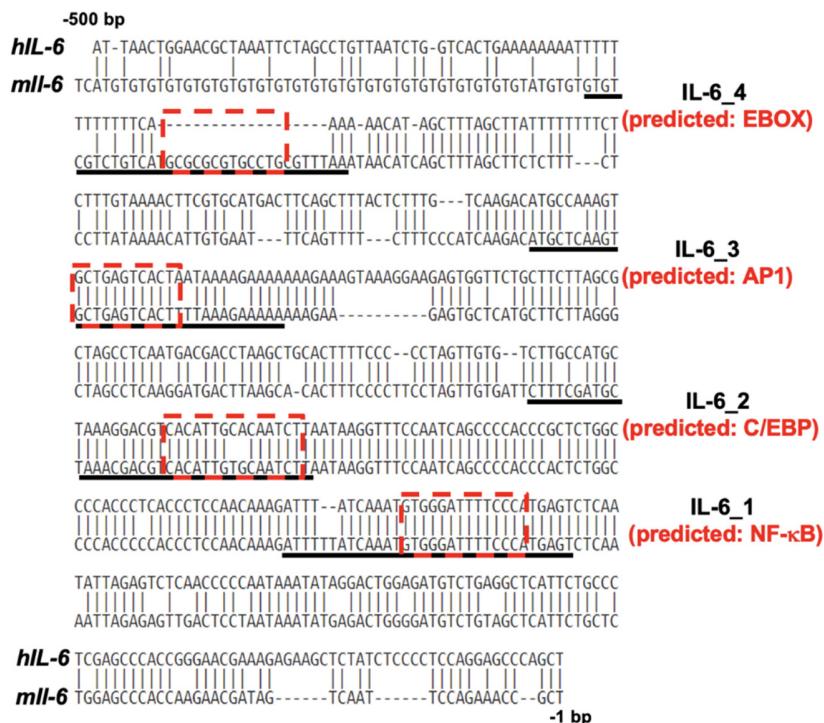


Online

Supplemental Figure 1A. The promoter regions of the human *IL-6* and mouse *Il-6* promoter share high homology. Percentage of alignment between the human *IL-6* and mouse *Il-6* gene using the VISTA genome browser software. Location of the luciferase construct (dotted and solid line) including the region of sequence homology in humans and mice as shown in Online Supplemental Figure 1B (solid line) are

depicted. Red = conserved non-coding sequence (CNS), turquoise = untranslated region (UTR), blue = exons.

B.



Online Supplemental Figure 1B. Sequence homology between the human (upper panel) and mouse (lower panel) *IL-6* promoter region. The sequence of both species (500 bp upstream of the translation start site) are shown including the four analyzed regions by electrophoretic mobility shift assay, predicted transcription factor binding to each of the studied *Il-6* loci, and the GT repeat. Relative distance to the translation start site was included.

-1139 AGACCTTGAGC ATTGGAGGGG TTATTCAGAG TGAGACGTAC CACCTTCAGA TTCAAATCCT GTCATCCAGT AGAAGGGAGC TTCAAACACA AGCTAGCTAA
TCTGAACTCG TAACCTCCCCC AATAAGTCTC ACTCTGCATG GTGGAAGTCT AAGTTAGGA CAGTAGGTCA TCTTCCCTCG AAGTTTGTTG TCGATCGATT
 luciferase construct 5' start
 -1039 GATACAATGA GGTCTTCTT CGATATCTT ATCTTCATA TACCATGAAT CAAAGAAACT TCAACAAACAT GAGGACTGCA ACAGACCTTC AAGCCTCCTT
 CTATGTTACT CCAGGAAGAA GCTATAGAAA TAGAAGGTAT ATGGTACTTA GTTCTTGTGA AGTTGTTGTA CTCCTGACGT TGTCTGGAAG TTCGGAGGA
 -939 GCATGACCTC GAAATGTTT GGGGTGCTT GGCAGCAGTG GGATCAGCAC TAACAGATAA GGGCAACTCT CACAGAGACT AAAGGTCTTA ACTAAGAAGA
 CGTACTGGAC CTTTACAAAA CCCACAGGA CGGTGTCAC CCTAGTCGTG ATTGTCATT CCCGTTGAGA GTGTCTCTGA TTTCCAGAAT TGATTCTCT
 -839 TAGCCAAGAG ACCACTGGGG AGAATGCAGA GAATAGGCTT GGACTTGGAA GCCAAGATTG CTTGACAAACAGA GACAGAGAGT ATTTCTGTAC TTCACCCACT
 ATCGTTCTC TGGTACCCCCC TCTTACGTCT CTTATCCGAA CCTGAAACCTT CGGTCTAAC GAACTGTGTG CTGTCCTCTA TAAAGACATG AAGTGGTGA
 -739 TTACCCACCT GGCAACTCCT GGAAACAACT GCACAAAATT TGGAGGTGAA CAAACCATTA GAAACAACTG GTCTGACAA GACACAGGA AAACAGCAA
 AAATGGTGGG CGGTTGAGGA CTTTGTGTA CGTGTGTTAA ACCTCCACTT GTTGGTAAAT CTTTGTGAC CAGGACTGTT CTGTCCTT TTTGTCCTT
 -639 TATGCAACAT TACTGTCGTG TGTCAGGTT GGGTGCTGGG GGTGGGAGAG GGAGTGTG TGCTTTGTATG ATCTGAAAAAA ACTCAGGTCA GAACATCTGT
 ATACGTTGTA ATGACAGACA ACAGGTCACCA CCCACGACCC CCACCCCTCTC CCTCACACAC AGAACACATAC TAGACTTT TGAGTCCAGT CTTGTTAGACA
 GT-repeat
 -539 AGATCCTTAC AGACATACAA AAGAARTCTA GCCTCTTATT CATGTGTGTC TGTGTGTG TGTTGTG TGTTGTG TGTTGTG TGTTGTG TGTTGTG
 TCTAGGAATG TCTGTATGTT TTCTTAGGAT CGGAGAATAA GTACACACAC ACACACACAC ACACACACAC ACACACACAT ACACACACAG
II-6_4 (predicted: EBOX)
 -439 GTCTGTCACTG CGCGCGTCCC TCGGTTAA TAACATCAGC TTTAGCTTCT CTTTCTCCTT ATAAAACATT GTGAATTCTA GTTTCTCTT CCATCAAGAC
CAGACAGTAC CGCGCGACCG ACGCAAATT ATTGTAGTCG AAATCGAAGA GAAAGAGGAA TATTTGTAA CACTTAAAGT CAAAAGAAAG GGTAGTTCTG

II-6_3 (predicted: AP1)

-339 ATGCTCAAGT GCTGACTCAC TTTAAAGAA AAAAAAGAAG AGTGCTCATG CTTCTTAGGG CTAGCCTCAA GGATGACTTA AGCACACATT CCCCTTCTCA
TACGAGTTCA GCACACTCAGT AAAATTCTT TTTTCTCTC TCACGAGTAC GAAGAATCCC GATCGGAGTT CCTACTGAAT TGTTGTGAAA GGGGAAGGAT

transcription start site

II-6_2 (predicted: C/EBP)

-239 GTTGTGATTTC TTTCGATGCT AAACGACGTC ACATTGTGCA ATCTTAATAA GGTTTCCAAT CAGCCCCACC CACTCTGCC CCACCCCCAC CCTCCAACAA
 CAACACTAAG AAAGCTACGA TTGCTGCAG TGTAACACGT TAGAATTATT CCAAAGGTTA GTCGGGGTGG GTGAGACCGG GGTGGGGTGG

↑

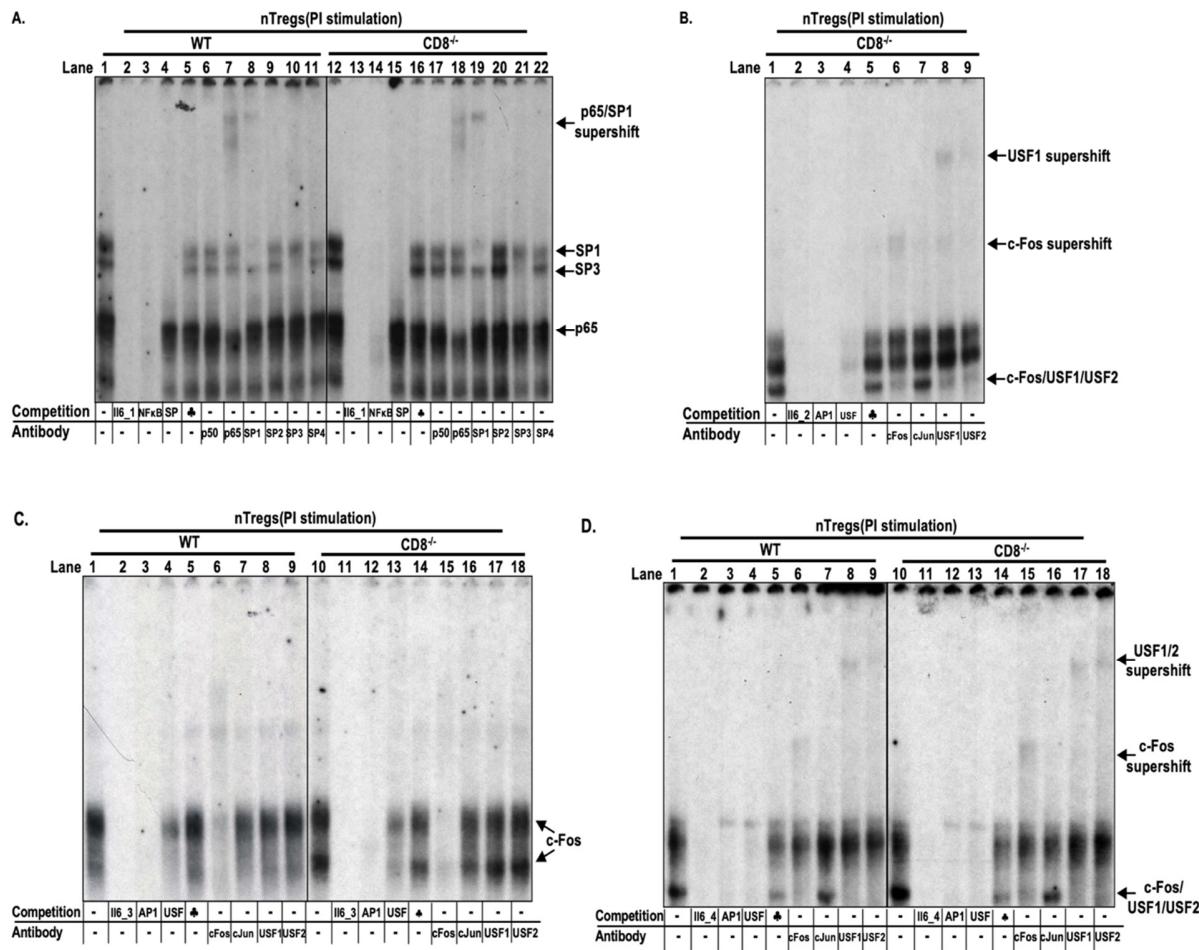
II-6_1 (predicted: NF-κB)

-139 AGATTTTAT CAAATGTGGG ATTTCCCAT GAGTCTCAA ATTAGAGAGT TGACTCTAA TAAATATGAG ACTGGGGATG TCTGTAGCTC ATTCTGCTCT
TCTAAAGATA GTTTACACCC TAAAGGGTA CTCAGAGTTT TAATCTCTCA ACTGAGGATT ATTTATACTC TGACCCCTAC AGACATCGAG TAAGACGAGA

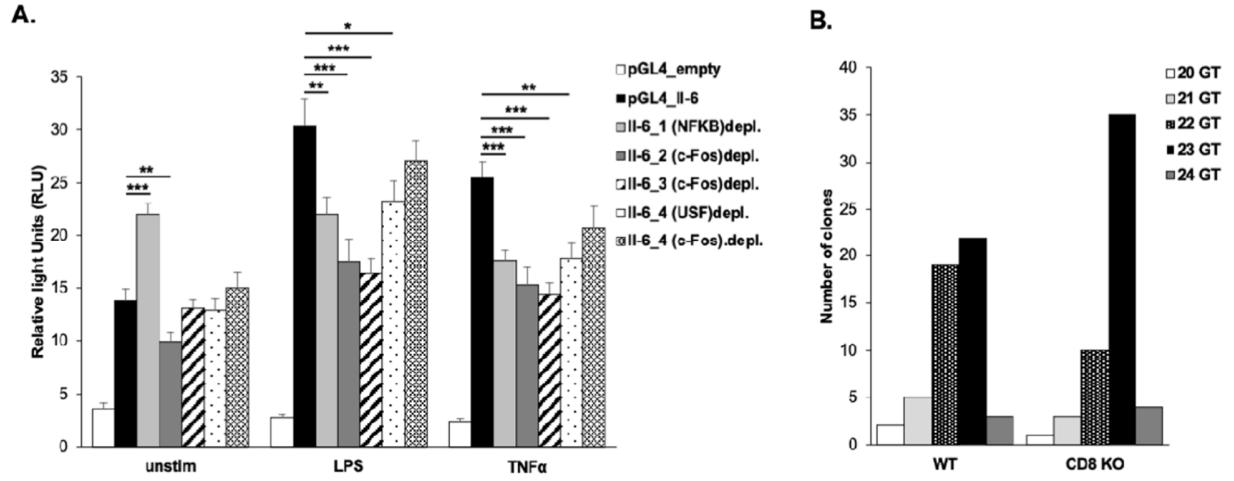
translation start site

-39 GGAGCCCCACC AAGAACGATA GTCAATTCCA GAAACCGCTA TGAAGTCTT CTCTGCAAGT AAGTGAAGGC AGTTCCCTGC CCTCTGGCGG AGCTATTGAG
 CCTCGGGTGG TTCTTGTCT CAGTTAAGGT CTTTGGCGAT ACTTCAAGA GAGACGTTCA TTCACTCCG TCAAGGAACG GGAGACCGCC TCGATAACTC
 luciferase construct 3' end

Online Supplemental Figure 2. Sequence of the mouse *Il-6* promoter region. The four analyzed regions by electrophoretic mobility shift assay and the GT repeat are underlined. The location of the cloned *Il-6* promoter region is depicted. Transcription and translation start site of *Il-6* are marked. Relative distance to the translation start site is shown.



Online Supplemental Figure 3. Distinct affinity of transcription factor binding to promoter regions within the *Il-6* gene in nTregs from wild-type (WT) and CD8^{-/-} mice. A-D. EMSA analyses of four different *Il-6* promoter loci upstream of the transcription start site of the *Il-6* gene were performed including A. Il-6_1, B. Il-6_2, C. Il-6_3, and D. Il-6_4 using nuclear extract isolated from nTregs of WT or CD8^{-/-} (KO) mice (5 µg) stimulated with PMA/ionomycin (50 ng/ml/1 µM) for 3 hours. Competitors (100-fold molar excess) and supershift antibodies (4 µg) for each experiment are noted below the respective lanes. ♦: competition experiment using an unrelated oligo.



Online Supplemental Figure 4. Four defined regions are essential for transcription factor binding and a novel variable length STR is found in the *Il-6* promoter **A.** NIH3T3 cells were transiently transfected with the empty pGL4 luciferase vector (pGL4 empty) or with different reporter constructs of the *Il-6* promoter (1,138 bp). In the construct *Il-6_1* (NF-κB)depl., the NF-κB binding site as identified by EMSA within *Il-6_1* was depleted by site-directed mutagenesis. Correspondingly, the c-Fos binding site within *Il-6_2* [*Il-6_2*(c-Fos)depl.], the c-Fos binding site within *Il-6_3* [*Il-6_3*(c-Fos)depl.], the USF binding site within *Il-6_4* [*Il-6_4*(USF)depl.], and the c-Fos binding site within *Il-6_4* [*Il-6_4*(c-Fos)depl.] were mutated (4-5 independent experiments, 2 technical replicates). Cells were left in medium or stimulated with 10 ng/ml LPS or 0.5 ng/ml TNF α . Luciferase activity was normalized for transfection efficiency using the control plasmid pRL-TK. The relative luciferase activity is presented in relative light units (RLU). Paired, two tailed students t-test was performed. * $p\leq 0.05$, ** $p\leq 0.01$, *** $p\leq 0.001$. **B.** *Il-6* luciferase promoter constructs were generated using DNA isolated from wild-type (WT) or CD8 $^{-/-}$ (CD8 KO) nTregs. To determine the distribution of a GT short tandem repeat in nTregs of both strains, a total of 104 clones (WT=51, CD8 $^{-/-}$ =53) were sequenced. Distribution of number of clones in nTregs from each strain containing 20-24 GT repeats are depicted.

Supplemental Table 1: *In silico* transcription factor (TF) binding analyses of the *Il-6* promoter region including 600 bp upstream of the translation start site.
 Relative distance upstream of the translation start site and the chromosomal strand (+ or -) of the predicted binding of the TF are depicted. Capital letters in the sequence indicate the core sequence of a predicted TF binding matrix with the highest degree of conservation. Letters marked in red reflect the conserved sequences (>60%) between the TF matrix and *Il-6*. Marked in bold are predicted TF binding regions further evaluated by electrophoretic mobility shift assay.

TF	TF family	Name of TF family	Relative distance to TLSS (bp)		Strand	Core similarity	Matrix similarity	Sequence
BTEB3	KLFS	Krueppel like transcription factors	-595	-579	+	1.000	0.939	ggagag GGAGt tgttgt
GAGA	GABF	GA-boxes	-598	-574	+	1.000	0.81	gtggg AGAGggag tgtgtctttg
TIEG	SP1F	GC-Box factors SP1/GC	-591	-577	+	0.750	0.879	ag GGAGt gtgtgtct
TAXCREB	CREB	cAMP-responsive element binding proteins	-591	-571	-	0.750	0.713	atacaa AGACacaca ctccct
GATA3	GATA	GATA binding factors	-575	-563	-	1.000	0.932	ttc A GATcataca
AREB6	ZFHX	Two-handed zinc finger homeodomain transcription factors	-559	-547	-	1.000	0.945	ttctg ACCT gagt
REV-ERBA	RORA	v-ERB and RAR-related orphan receptor alpha	-563	-541	+	1.000	0.895	aaaaactcag GTCA gaacatctg
FHXB	FKHD	Fork head domain factors	-556	-540	+	0.818	0.853	cagg t AGAAcatctgt
TAL1ALPHAE47	HAND	Twist subfamily of class B bHLH transcription factors	-554	-534	-	1.000	0.952	ggatcta CAGA tgtctgacc
NGN_NEUROD	NEUR	NeuroD, Beta2, HLH domain	-550	-538	+	1.000	0.985	agaa CATC tgttag
RP58	RP58	RP58 (ZFP238) zinc finger protein	-550	-538	+	1.000	0.954	agaa CATC tgttag
PRE	GREF	Glucocorticoid responsive and related elements	-552	-534	-	1.000	0.851	ggatctacaga TGTT tctga
POU3F3	OCT1	Octamer binding protein	-510	-494	-	0.784	0.823	cacat GAAT aagaggct
PIT1	PIT1	GHF-1 pituitary specific pou domain transcription factor	-509	-495	+	1.000	0.93	gcctc TATT catgt
PAX4_PD	PAX6	PAX-4/PAX-6 paired domain binding sites	-436	-418	-	1.000	0.919	cag GCA gcgcgcatgaca
ZF5	ZF5F	ZF5 POZ domain zinc finger	-432	-422	+	1.000	0.967	at gc GCC gtgt
HELT	HESF	Vertebrate homologues of enhancer of split complex	-432	-418	-	1.000	0.959	cagg CACG gcgcatt
AHRARNT	AHRR	AHR-arnt heterodimers and AHR-related factors	-436	-412	+	1.000	0.952	tgtcatgcgc GCGT gcctgcgttta
MYCMAX	EBOX	E-box binding factors	-430	-418	-	1.000	0.921	caggcaCGCGcg c
OTP	CART	Cart-1 (cartilage homeoprotein 1)	-423	-403	-	0.751	0.841	gatgt tt ATTaaacgcaggca

<i>E4BP4</i>	<i>CREB</i>	cAMP-responsive element binding proteins	-421	-401	-	0.758	0.852	ctgatgttatTTAAacgcagg
<i>TEF_HLF</i>	<i>PARF</i>	PAR/bZIP family	-419	-403	+	0.784	0.82	tgcgtTTAAataacatc
<i>E4BP4</i>	<i>CREB</i>	cAMP-responsive element binding proteins	-420	-400	+	0.769	0.803	ctgcgtttaaATAAcatcagc
<i>LHX3</i>	<i>LHXF</i>	Lim homeodomain factors	-421	-399	-	0.751	0.812	agctgatgttATTaaacgcagg
<i>TEF</i>	<i>PARF</i>	PAR/bZIP family	-418	-402	-	0.773	0.888	tgatgttatTTAAacgc
<i>MTBF</i>	<i>HMTB</i>	Human muscle-specific Mt binding site	-413	-405	-	1.000	0.902	tgttATTTa
<i>PROP1</i>	<i>CART</i>	Cart-1 (cartilage homeoprotein 1)	-418	-398	+	0.753	0.844	gcgtttAAATaaacatcagctt
<i>CREB</i>	<i>CREB</i>	cAMP-responsive element binding proteins	-416	-396	-	0.862	0.863	taaagcTGATgtttaaac
<i>BARBIE</i>	<i>BARB</i>	Barbiturate-inducible element box from pro+eukaryotic genes	-407	-393	-	1.000	0.91	agctAAAGctgatgt
<i>EVI1</i>	<i>EVI1</i>	EVI1-myleoid transforming protein	-403	-387	-	0.750	0.841	aagagAAGCtaaagctg
<i>IRF7</i>	<i>IRFF</i>	Interferon regulatory factors	-401	-381	-	1.000	0.892	aggaGAAAgagaagctaagc
<i>LTATA</i>	<i>VTBP</i>	Vertebrate TATA binding protein factor	-390	-374	-	1.000	0.84	tttTATAaggagaaga
<i>HOXD13</i>	<i>ABDB</i>	Abdominal-B type homeodomain transcription factors	-386	-370	+	1.000	0.919	tcccttaTAAAacatt
<i>CDX2</i>	<i>CDXF</i>	Vertebrate caudal related homeodomain protein	-386	-368	-	1.000	0.852	acaatgtTTTAtaaggaga
<i>PBX_HOXA9</i>	<i>HOXC</i>	HOX - PBX complexes	-384	-368	-	0.750	0.849	acaatGTtataagga
<i>VTATA</i>	<i>VTBP</i>	Vertebrate TATA binding protein factor	-383	-367	+	1.000	0.941	cctaTAAAacattgt
<i>FHXB</i>	<i>FKHD</i>	Fork head domain factors	-378	-362	-	0.909	0.855	aaattcACAAtgttta
<i>FAST1</i>	<i>FAST</i>	FAST-1 SMAD interacting proteins	-375	-359	+	1.000	0.884	aacaTGTGaattcag
<i>IRF1</i>	<i>IRFF</i>	Interferon regulatory factors	-371	-351	-	1.000	0.901	aaagaaaactGAAAatcacaa
<i>HMGY</i>	<i>SORY</i>	SOX/SRY-sex/testis determinig and related HMG box factors	-372	-350	+	1.000	0.93	attgtgAATTtcagtttcttc
<i>DINR</i>	<i>INRE</i>	Core promoter initiator elements	-364	-354	+	1.000	0.965	ttTCAGtttcc
<i>RBPJK</i>	<i>RBPF</i>	RBPJ - kappa	-357	-343	-	1.000	0.946	ttgaTGGGaaagaaaa
<i>PAX1</i>	<i>PAX1</i>	PAX-1 binding sites	-355	-337	+	0.750	0.689	tCTTcccataagacatg
<i>OCT3_4</i>	<i>OCT1</i>	Octamer binding protein	-342	-326	+	1.000	0.824	gacATGCtcaagtgcgt
<i>P53</i>	<i>P53F</i>	p53 tumor suppressor	-344	-322	-	1.000	0.935	gactcagcacttgatCATGtctt
<i>DEC2</i>	<i>HESF</i>	Vertebrate homologues of enhancer of split complex	-339	-325	-	0.871	0.972	ttagcaCTTGagcat
<i>NKX25</i>	<i>NKXH</i>	NKX homeodomain factors	-339	-321	+	1.000	1	atgctcAAGTgctgagtca

<i>TCF11MAFG</i>	<i>AP1R</i>	MAF and AP1 related factors	-335	-315	-	1.000	0.957	taaaag TGAC ttagcacttga
<i>AP1</i>	<i>AP1F</i>	<i>AP1, Activating protein 1</i>	-329	-319	+	1.000	0.968	gctgAGTCact
<i>AP1</i>	<i>AP1F</i>	<i>AP1, Activating protein 1</i>	-329	-319	-	0.971	0.974	agTGACtcagc
<i>NFE2</i>	<i>AP1R</i>	MAF and AP1 related factors	-333	-313	+	1.000	1	aagtg CTGAg tcaattttaaa
<i>VTATA</i>	<i>VTBP</i>	Vertebrate TATA binding protein factor	-326	-310	-	1.000	0.927	ttctt TA AAagtgactc
<i>BARBIE</i>	<i>BARB</i>	Barbiturate-inducible element box from pro+eukaryotic genes	-310	-296	+	1.000	0.888	aaaa AAAG aagagtg
<i>EVI1.07</i>	<i>EVI1</i>	EVI1-myleoid transforming protein	-310	-294	+	1.000	0.916	aaaaaa AAGA agagtgtc
<i>PAX2</i>	<i>PAX8</i>	PAX-2/5/8 binding sites	-297	-283	+	0.923	0.94	tg CTCA tgcttctta
<i>HMX2</i>	<i>HOMF</i>	Homeodomain transcription factors	-272	-254	+	1.000	0.916	caaggatga CTT Aaggcaca
<i>CREB2</i>	<i>CREB</i>	cAMP-responsive element binding proteins	-272	-252	+	1.000	0.906	caagga TGAC ttaagcact
<i>ATATA</i>	<i>VTBP</i>	Vertebrate TATA binding protein factor	-269	-253	-	1.000	0.812	gtgt gct TAAGtcatcc
<i>HMX2</i>	<i>HOMF</i>	Homeodomain transcription factors	-267	-249	-	1.000	0.896	gaaagtgtg CTT Aaggcat
<i>MYT1</i>	<i>MYT1</i>	MYT1 C2HC zinc finger protein	-260	-248	-	1.000	0.799	gg AAGT gtgttt
<i>BLIMP1</i>	<i>PRDF</i>	Positive regulatory domain I binding factor	-261	-243	-	1.000	0.838	gaaggg GAAA gtgtgttta
<i>IRF7</i>	<i>IRFF</i>	Interferon regulatory factors	-259	-239	-	0.821	0.905	ctag GAA Gggaaagtgtgt
<i>MZF1</i>	<i>MZF1</i>	Myeloid zinc finger 1 factors	-254	-244	-	1.000	1	aa GGGG aaagt
<i>GABP</i>	<i>ETSF</i>	Human and murine ETS1 factors	-254	-234	-	1.000	0.868	cacaacta GGA Agggaaagt
<i>IRF4</i>	<i>IRFF</i>	Interferon regulatory factors	-236	-216	-	1.000	0.942	gttttagc atc GAAAgaatcac
<i>PAX2</i>	<i>PAX2</i>	PAX-2 binding sites	-236	-214	+	1.000	0.788	gtgattcttcgat gt AAACga
<i>CREB1</i>	<i>CREB</i>	cAMP-responsive element binding proteins	-223	-203	-	1.000	0.974	acaatg TGAC gtcgtttagca
<i>E4F</i>	<i>E4FF</i>	Ubiquitous GLI - Krueppel like zinc finger involved in cell cycle regulation	-217	-205	+	1.000	0.893	acg ACGT cacatt
<i>CREB</i>	<i>CREB</i>	cAMP-responsive element binding proteins	-220	-200	-	1.000	0.902	tgcacaatg TGAC gtcgttta
<i>SOX9</i>	<i>SORY</i>	SOX/SRY-sex/testis determining and related HMG box factors	-218	-196	-	1.000	0.944	agattgc ACA Atgtgacgtcgtt
<i>CEBPB</i>	<i>CEBP</i>	<i>Ccaat/Enhancer Binding Protein</i>	-210	-196	+	1.000	0.989	cacattgtGCAAtct

<i>HLF</i>	<i>PARF</i>	PAR/bZIP family	-211	-195	+	0.836	0.869	tcacattgt GCAA tctt
<i>HHEX</i>	<i>HOMF</i>	Homeodomain transcription factors	-206	-188	+	1.000	0.953	tttgcaatct TAAT aagg
<i>BRN3</i>	<i>BRNF</i>	Brn POU domain factors	-202	-184	+	1.000	0.84	gcaatct TAAT aaggttc
<i>VAX2</i>	<i>HBOX</i>	Homeobox transcription factors	-202	-184	-	1.000	0.852	gaaacc TTA agattgc
<i>BARX2</i>	<i>HOMF</i>	Homeodomain transcription factors	-202	-184	+	1.000	0.952	gcaatct TAAT aaggttc
<i>NKX61</i>	<i>NKX6</i>	NK6 homeobox transcription factors	-200	-186	+	1.000	0.915	aat TTA taagggtt
<i>BRN5</i>	<i>BRN5</i>	Brn-5 POU domain factors	-203	-181	-	1.000	0.922	ttggaaacc TTA agattgca
<i>OCT1</i>	<i>OCT1</i>	Octamer binding protein	-200	-184	-	1.000	0.861	gaaacc TTA Agatt
<i>NKX25</i>	<i>NKXH</i>	NKX homeodomain factors	-200	-182	+	1.000	0.899	aat TTA taaggttcca
<i>NFAT5</i>	<i>NFAT</i>	Nuclear factor of activated T-cells	-198	-180	-	1.000	0.888	att GGAA accttataaga
<i>BRN2</i>	<i>BRNF</i>	Brn POU domain factors	-195	-177	-	0.900	0.879	ctGATTggaaaccttatta
<i>NFY</i>	<i>CAAT</i>	CCAAT binding factors	-188	-174	+	1.000	0.951	gttt CCAAt cagcccc
<i>CDP</i>	<i>CLOX</i>	CLOX and CLOX homology (CDP) factors	-189	-171	+	1.000	0.969	ggttc CAAT cagcccc
<i>INSM1</i>	<i>INSM</i>	Insulinoma associated factors	-180	-168	-	1.000	0.907	tgggt GGGG ctga
<i>BKLF</i>	<i>KLFS</i>	Krueppel like transcription factors	-182	-166	-	1.000	0.99	agt GGGT gggctgatt
<i>MAZR</i>	<i>MAZF</i>	Myc associated zinc fingers	-179	-167	-	1.000	0.895	gtggg GGGG ctg
<i>GC</i>	<i>SP1F</i>	GC-Box factors SP1/GC	-180	-166	-	0.872	0.942	agt GGT Gggctga
<i>WT1</i>	<i>EGRF</i>	EGR/nerve growth factor induced protein C & related factors	-179	-163	-	1.000	0.932	cagag TGGG tgggctg
<i>GLI3</i>	<i>GLIF</i>	GLI zinc finger family	-178	-164	+	1.000	0.881	agcc CCAC ccactct
<i>NGFIC</i>	<i>EGRF</i>	EGR/nerve growth factor induced protein C & related factors	-177	-161	-	0.770	0.833	gcc GAGT gggtggggc
<i>PLAG1</i>	<i>PLAG</i>	Pleomorphic adenoma gene	-176	-156	-	0.833	0.881	GTGG ggccagagtgggtgggg
<i>BKLF</i>	<i>KLFS</i>	Krueppel like transcription factors	-167	-151	-	1.000	0.998	tgg GGGT ggggccagag
<i>MAZR</i>	<i>MAZF</i>	Myc associated zinc fingers	-164	-152	-	1.000	0.949	gggggt GGGG cca
<i>GC</i>	<i>SP1F</i>	GC-Box factors SP1/GC	-165	-151	-	0.872	0.945	tggg GGT Gggccag
<i>GKLF</i>	<i>KLFS</i>	Krueppel like transcription factors	-165	-149	-	1.000	0.984	ggtgtggg GTGG ggccag
<i>PLAG1</i>	<i>PLAG</i>	Pleomorphic adenoma gene	-166	-146	-	1.000	0.885	GAGG gtgggggtggggccaga
<i>ZBP89</i>	<i>ZBPF</i>	Zinc binding protein factors	-166	-144	+	1.000	0.976	tctggcccca CCCC accctcca
<i>WT1</i>	<i>EGRF</i>	EGR/nerve growth factor induced protein C & related factors	-162	-146	-	1.000	0.998	gaggg TGGG gtggggc

<i>RREB1</i>	<i>RREB</i>	Ras-responsive element binding protein	-161	-147	+	1.000	0.876	cCCCAcccccacccct
<i>BKLF</i>	<i>KLFS</i>	Krueppel like transcription factors	-161	-145	-	1.000	0.974	ggaGGTgggggtgggg
<i>GC</i>	<i>SP1F</i>	GC-Box factors SP1/GC	-159	-145	-	0.872	0.893	ggagGGTGgggggtgg
<i>ZNF219</i>	<i>ZBPF</i>	Zinc binding protein factors	-163	-141	+	1.000	0.941	ggcccaCCCCaccctccaaca
<i>SPZ1</i>	<i>SPZ1</i>	Testis-specific bHLH-Zip transcription factors	-154	-144	-	1.000	0.958	tGGAGggtggg
<i>LEF1</i>	<i>LEFF</i>	LEF1/TCF	-149	-133	+	1.000	0.876	cctccaaCAAAGatttt
<i>GFI1</i>	<i>GFI1</i>	Growth factor independence transcriptional repressor	-146	-132	-	1.000	0.905	aaaAATCtttgttgg
<i>SOX2</i>	<i>SORY</i>	SOX/SRY-sex/testis determinig and related HMG box factors	-150	-128	+	1.000	0.929	ccctccaACAAagattttatca
<i>CDX2</i>	<i>CDXF</i>	Vertebrate caudal related homeodomain protein	-141	-123	+	1.000	0.862	aaagatTTTAtcaaatgt
<i>GATA</i>	<i>GATA</i>	GATA binding factors	-137	-125	-	1.000	0.936	atttGATAaaaat
<i>SMARCA3</i>	<i>RUSH</i>	SWI/SNF related nucleophosphoproteins with a RING finger DNA binding motif	-131	-121	-	0.960	0.965	ccACATttgat
<i>NFAT5</i>	<i>NFAT</i>	Nuclear factor of activated T-cells	-128	-110	-	1.000	0.904	atGGAAtccacatt
<i>NFκB p65</i>	<i>NFKB</i>	Nuclear factor kappa B/c-rel	-124	-112	+	1.000	0.982	gtgggattTTCCc
<i>NFκB</i>	<i>NFKB</i>	Nuclear factor kappa B/c-rel	-122	-110	-	1.000	0.891	atGGGAAaatccc
<i>IK1</i>	<i>IKRS</i>	Ikaros zinc finger family	-120	-108	-	1.000	0.941	tcatGGGAaaatc
<i>RBPJK</i>	<i>RBPF</i>	RBPJ - kappa	-121	-107	-	1.000	0.965	ctcaTGGGaaatcc
<i>OLF1</i>	<i>NOLF</i>	Neuron-specific-olfactory factor	-121	-99	+	1.000	0.894	ggatttTCCCatgagtctaaaa
<i>PHOX2</i>	<i>CART</i>	Cart-1 (cartilage homeoprotein 1)	-111	-91	-	1.000	0.882	ctctcTAATttgagactcat
<i>MSX</i>	<i>HOMF</i>	Homeodomain transcription factors	-107	-89	-	1.000	0.989	aactctcTAATttgagac
<i>GSH1</i>	<i>HBOX</i>	Homeobox transcription factors	-104	-86	-	1.000	0.881	gtcaacttcTAATttgaa
<i>MYT1L</i>	<i>MYT1</i>	MYT1 C2HC zinc finger protein	-96	-84	+	1.000	0.945	agagAGTTgactc
<i>HOXC13</i>	<i>ABDB</i>	Abdominal-B type homeodomain transcription factors	-88	-72	+	1.000	0.949	gactcctaTAAAatgt
<i>BRN3</i>	<i>BRNF</i>	Brn POU domain factors	-88	-70	-	0.759	0.844	ctcatatTTATtaggagtc
<i>PHOX2</i>	<i>CART</i>	Cart-1 (cartilage homeoprotein 1)	-87	-67	+	1.000	0.881	actccTAATaaatatgagact

<i>BRN5</i>	<i>BRN5</i>	Brn-5 POU domain factors	-60	-38	+	0.751	0.844	gtctgttagtc ATTC tgctctgg
<i>PAX6</i>	<i>PAX6</i>	PAX-4/PAX-6 paired domain binding sites	-47	-29	-	1.000	0.886	tggtgggctC CAG agcaga
<i>HOMEZ</i>	<i>HZIP</i>	Homeodomain-leucine zipper transcription factors	-30	-16	-	1.000	0.839	tgact ATCG tcttg
<i>TH1E47</i>	<i>HAND</i>	Twist subfamily of class B bHLH transcription factors	-21	-1	+	1.000	0.958	tagtcaatt CCAG aaccgct
<i>STAT</i>	<i>STAT</i>	Signal transducer and activator of transcription	-20	-2	-	1.000	0.896	gcccgttct GGAA ttgact

Supplemental Table 2: Single-stranded oligonucleotides used for electrophoretic mobility shift assays (EMSA).

Oligonucleotide name	Oligonucleotide sequence (5'3')
Il-6_1_fwd	GATTTTATCAAATGTGGGATTTCCCATGAGT
Il-6_1_rev	ACTCATGGAAAATCCCACATTGATAAAAAATC
Il-6_2_fwd	CTTCGATGCTAACCGACGTACATTGTGCAATCTT
Il-6_2_rev	AAGATTGCACAATGTGACGTCGTTAGCATCGAAAG
Il-6_3_fwd	ATGCTCAAGTGCTGAGTCACTTAAAGAAAAA
Il-6_3_rev	TTTTCTTAAAAGTGA CTCAGCACTTGAGCAT
Il-6_4_fwd	TGTGCGTCTGTCATGCGCGCGCTGCCTGCGTTAA
IL6_4_rev	TTAAACGCAGGCACCGCGCGCATGACAGACGACACA
USF consensus sequence_fwd	CACCCGGTCACGTGGCCTACACC
USF consensus sequence_rev	GGTGTAGGCCACGTGACCGGGTG
SP consensus sequence_fwd	ATTCGATCGGGCGGGCGAGC
SP consensus sequence_rev	GCTCGCCCCGCCCCGATCGAAT
NF-κB consensus site fwd	TCAGAGGGGACTTCCGAGAGGCG
NF-κB consensus site rev	CGCCTCTCGAAAGTCCCCTCTGA
AP1 consensus sequence_fwd	TGAGTCATGAGTCATGAGTCA
AP1 consensus sequence_rev	TGACTCATGACTCATGACTCA

Supplemental Table 3: Sequence of primers used to amplify the *Il-6* promoter region for generating luciferease reporter constructs, to verify the correctness of the sequence, and to perform site-directed mutagenesis. Underlined basepairs represent mutated region of a potential transcription factor in the *Il-6* promoter region used for site-directed mutagenesis.

Primer name	Primer sequence (5`3`)	Application
Il-6 promoter_fwd	CTAACTGGCCGGTACCA <u>Gacttgaggcattggaggggttattc</u> *	PCR to amplify <i>Il-6</i> promoter
Il-6 promoter_rev	GGCCAGATCTGATATCCTCGAG <u>Agcggttctgaaattgactatc</u> G *	PCR to amplify <i>Il-6</i> promoter
RVprimer3	CTAGCAAAATAGGCTGTCCC	Sequencing of <i>Il-6</i> promoter construct
pGL4.10_rev	CGGTAAAGGCGATGGTGC	Sequencing of <i>Il-6</i> promoter construct
Il-6_1_Mut	GATTTTATCAAATGTGGAT <u>GTTCCC</u> CATGAGTCT	Site-directed mutagenesis of NF-κB in Il-6_1
Il-6_2_Mut	CTTCGATGCTAACG <u>A</u> TATCACATTGTGCAATC	Site-directed mutagenesis of cFos in Il-6_2
Il-6_3_Mut	CCATCAAGACATGCTCAAGTG <u>CTT</u> CACTTTAAAG	Site-directed mutagenesis of cFos in Il-6_3
Il-6_4_Mut (USF)	GTGTCGTCTGT <u>CATGCGCGT</u> ATGCCTGC <u>GT</u> AAATAAC	Site-directed mutagenesis of USF in Il-6_4
Il-6_4_Mut (cFos)	GTGTGTCGT <u>CTTACATGCGCGTGCCTGC</u>	Site-directed mutagenesis of cFos in Il-6_4

* Capital letters indicate homologous overlap between the empty pGL4.10 reporter vector the *Il-6* promoter region designed to build the seamless clone using the Gibson Assembly Cloning kit according to manufacturer's protocol (Thermofisher).