



Review

# From Pathogenesis to Therapy in Knee Osteoarthritis: Bench-to-Bedside

Elena Rezuş<sup>1,†</sup>, Alexandra Burlui<sup>1,\*</sup>, Anca Cardoneanu<sup>1,†</sup>, Luana Andreea Macovei<sup>1,†</sup> ,  
Bogdan Ionel Tamba<sup>2,\*</sup>  and Ciprian Rezuş<sup>3</sup>

<sup>1</sup> Department of Rheumatology and Physiotherapy, “Grigore T. Popa” University of Medicine and Pharmacy, 700115 Iași, Romania; elena.rezus@umfiasi.ro (E.R.); anca.cardoneanu@umfiasi.ro (A.C.); luana.macovei@umfiasi.ro (L.A.M.)

<sup>2</sup> Advanced Center for Research and Development in Experimental Medicine (CEMEX), “Grigore T. Popa” University of Medicine and Pharmacy, 700454 Iași, Romania

<sup>3</sup> Department of Internal Medicine, “Grigore T. Popa” University of Medicine and Pharmacy, 700115 Iași, Romania; ciprian.rezus@umfiasi.ro

\* Correspondence: maria-alexandra.burlui@umfiasi.ro (A.B.); bogdan.tamba@umfiasi.ro (B.I.T.)

† These authors contributed equally to this work.

**Abstract:** Osteoarthritis (OA) is currently the most widespread musculoskeletal condition and primarily affects weight-bearing joints such as the knees and hips. Importantly, knee OA remains a multifactorial whole-joint disease, the appearance and progression of which involves the alteration of articular cartilage as well as the synovium, subchondral bone, ligaments, and muscles through intricate pathomechanisms. Whereas it was initially depicted as a predominantly aging-related and mechanically driven condition given its clear association with old age, high body mass index (BMI), and joint malalignment, more recent research identified and described a plethora of further factors contributing to knee OA pathogenesis. However, the pathogenic intricacies between the molecular pathways involved in OA prompted the study of certain drugs for more than one therapeutic target (amelioration of cartilage and bone changes, and synovial inflammation). Most clinical studies regarding knee OA focus mainly on improvement in pain and joint function and thus do not provide sufficient evidence on the possible disease-modifying properties of the tested drugs. Currently, there is an unmet need for further research regarding OA pathogenesis as well as the introduction and exhaustive testing of potential disease-modifying pharmacotherapies in order to structure an effective treatment plan for these patients.

**Keywords:** osteoarthritis; knee joint; disease modifying drugs; cartilage; bone remodeling; inflammation; platelet-rich plasma; mesenchymal stem cells; ozone; hyaluronic acid



**Citation:** Rezuş, E.; Burlui, A.; Cardoneanu, A.; Macovei, L.A.; Tamba, B.I.; Rezuş, C. From Pathogenesis to Therapy in Knee Osteoarthritis: Bench-to-Bedside. *Int. J. Mol. Sci.* **2021**, *22*, 2697. <https://doi.org/10.3390/ijms22052697>

Academic Editor: Masato Sato

Received: 31 January 2021

Accepted: 4 March 2021

Published: 7 March 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

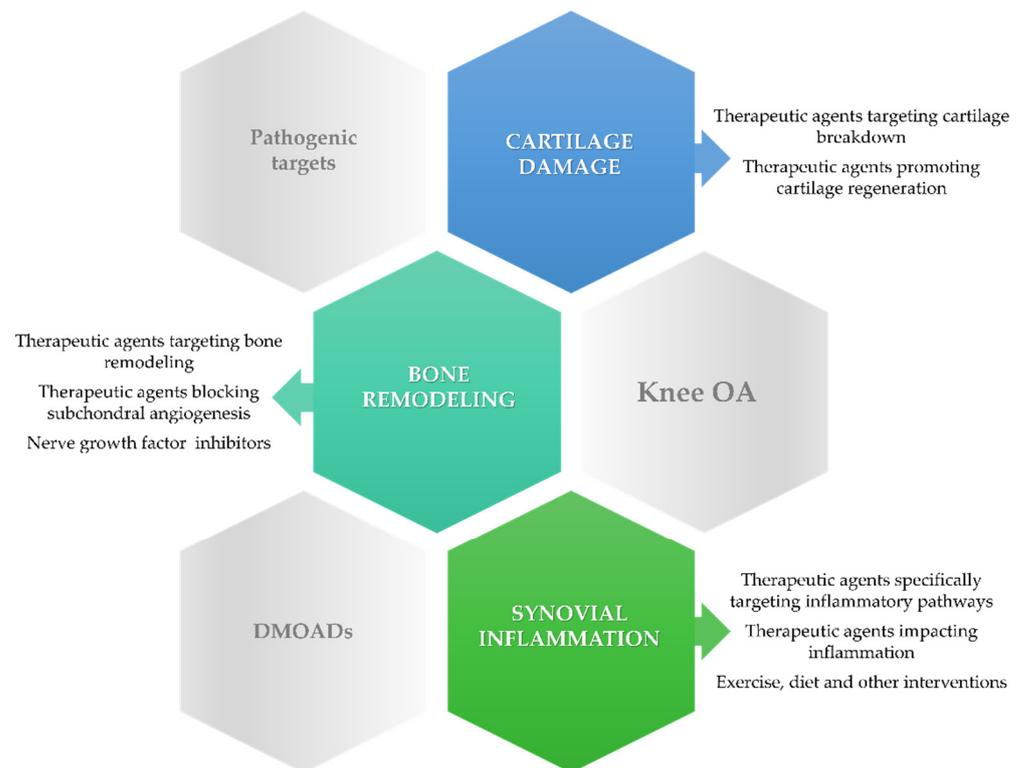
## 1. Introduction

Osteoarthritis (OA) is a chronic musculoskeletal condition that primarily affects weight-bearing joints (such as the knees, hips, and spine) yet may involve the hands as well as other non-weight-bearing articular sites [1–5]. Genetic predisposition has been deemed relevant, however, more so in the hands and hips rather than in knee OA [1–6]. Moreover, certain racial and gender-related differences were also reported [6,7]. Nevertheless, OA remains a multifactorial whole-joint disease, the appearance and progression of which involves the alteration of articular cartilage as well as the synovium, subchondral bone, ligaments, and muscles through intricate pathogenic mechanisms [1–3].

Whereas it was initially depicted as a predominantly aging-related and mechanically driven condition given its clear association with old age, high body mass index (BMI), and joint malalignment, more recent research identified and described a plethora of further factors contributing to knee OA pathogenesis [6–10].

Expert opinion in OA proposes case stratification, describing four phenotypes of the disease largely based on pathogenesis: mechanical, metabolic, osteoporotic, and inflam-

matory. Nonetheless, patient stratification could lead to more precise identification of the potential therapeutic targets yet demands a comprehensive evaluation pretreatment [5]. Novel findings on the mechanisms underlying the development of knee OA prompted the search for potential disease-modifying OA drugs (DMOADs) able to counteract the molecular pathways involved in cartilage degradation, inflammation, and bone remodeling (Figure 1). However, most therapeutic agents with potential disease-modifying properties have not yet proven their efficacy in slowing the progression of knee OA in clinical trials [6–8].

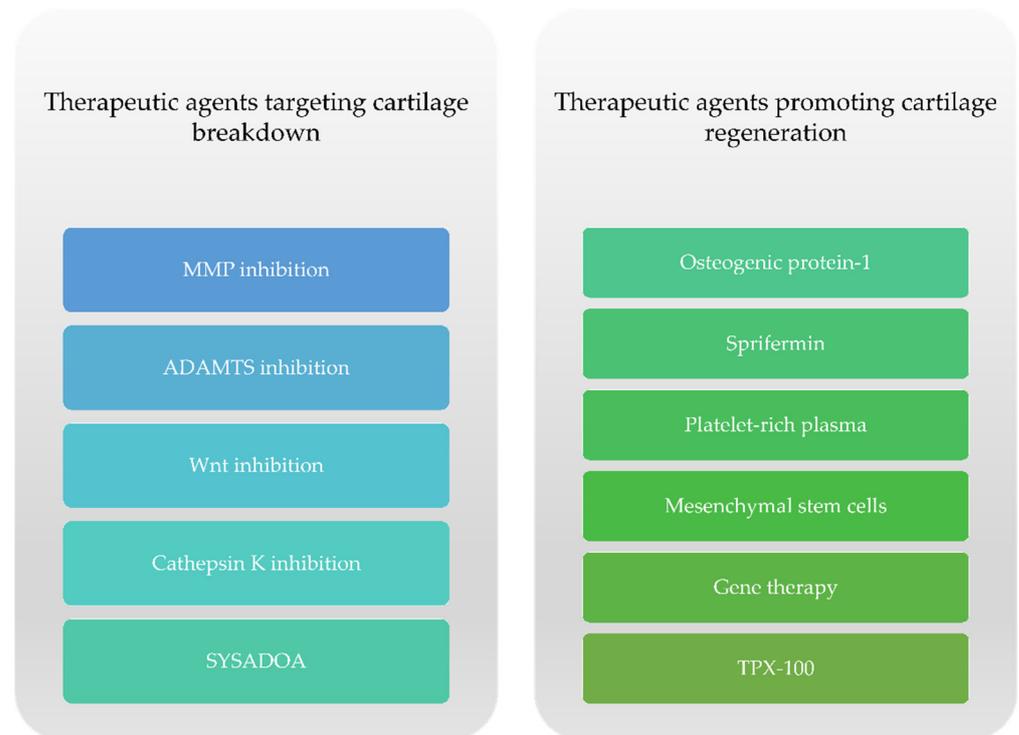


**Figure 1.** Therapeutic interventions targeting cartilage breakdown, bone remodeling, and inflammation in knee osteoarthritis (OA).

The present review aims to discuss the potentially disease-modifying therapeutic options targeting cartilage destruction, subchondral bone remodeling, and synovial inflammation in knee OA according to recent findings.

## 2. Therapeutic Approach to Cartilage Damage in Knee Osteoarthritis

The approval of DMOADs, according to regulatory guidelines from the United States Food and Drug Administration (FDA) and the European Medicines Agency (EMA) would need to meet the following conditions: slower loss in knee or hip joint space width (JSW) on x-ray and an appropriate symptomatic improvement [11]. Presently, there is no approved DMOAD despite the large amount of research conducted on the subject [12]. The currently available literature describes two subsets of potential DMOADs regarding cartilage damage in knee OA with respect to their specific actions: therapeutic agents targeting cartilage degeneration and drugs that support cartilage regeneration (Figure 2).



**Figure 2.** Therapeutic options targeting cartilage damage in knee OA.

### 2.1. Matrix Metalloproteinase Inhibition

Matrix metalloproteinases (MMPs) and aggrecanases are known as the main proteases responsible for matrix degradation in OA [13,14]. MMPs, a family of zinc-dependent enzymes, are recognized for their involvement in the degeneration of the extracellular matrix (ECM) [15]. They can be classified into the following groups: collagenases (MMP-1 and MMP-13), gelatinases (MMP-2 and MMP-9), stromelysins (MMP-3), metalloelastase (MMP-12), matrilysin (MMP-7), and membrane-type MMPs (MT-MMPs) [16].

In 2007, Krzeski et al. published a study of PG-116,800 (PG-530,742), an MMP-inhibitor, but no significant changes were observed in joint space width (JSW) of the knee or Western Ontario and McMaster Universities Osteoarthritis Index (WOMAC) scores at 1 year [17], yet important musculoskeletal side effects (named the “musculoskeletal syndrome” (MSS)) such as pain, loss of range of motion in large joints, joint swelling, stiffness, soft tissue pain, and Dupuytren’s contracture were observed in the treatment group. The exact cause of these reactions is not yet fully understood, but it is thought that chelation of zinc is involved in the process, seeing as the medication contains zinc-binding groups [18].

Given its notable pathogenic role in both OA as well as cartilage homeostasis, MMP-13 has been studied as a potential therapeutic target. ALS 1-0635 is a molecule that is free from zinc-chelating functional groups. Baragi et al. found a 67% deceleration in cartilage decline compared to the placebo in experimental animals in vivo [19].

Doxycycline is a broad-spectrum tetracycline antibiotic that also inhibits MMP activity, especially collagenase and gelatinase [20]. Nevertheless, a Cochrane review from 2012 stipulated that “the small benefit in terms of joint space narrowing is of questionable clinical relevance and outweighed by safety problems” [21].

### 2.2. ADAMTS Inhibition

In addition to MMPs, aggrecanases are also implicated in cartilage metabolism, acting as blockers of the aggrecan, the main proteoglycan of articular cartilage [22]. Aggrecan is a proteoglycan that incorporates chondroitin sulphate and keratan sulphate and is connected to a protein core. It links to another molecule, hyaluronan, thus creating stable large-molecular-weight molecules binding with a separate globular link [23]. In OA, the loss

of aggrecan is an early event in the degradation of articular cartilage and results in the decrement of functional and structural ECM integrity followed by an irreversible loss of collagen [24].

Based on the fact that ADAMTS-4 (aggrecanase 1) and ADAMTS-5 (aggrecanase 2) are able to cleave proteoglycans (such as aggrecan—the main component of articular cartilage), both of these molecules have been studied as potentially disease-modifying targets in OA. A humanized anti-ADAMTS-5 antibody (GSK2394002) was assessed in a study conducted by Larkin et al. The authors demonstrated the efficacy of this drug in preventing constitutional destruction as well as in decreasing mechanical pain. Nevertheless, systemic administration resulted in significant side effects (increased mean arterial pressure and cardiac ischemia) and therefore was not approved as a DMOAD [25]. CRB007, a chimeric murine/human IgG4 anti-ADAMTS-5 monoclonal antibody, displayed disease-modifying properties in animal models of OA in the study published by Chiusaroli et al. However, further extensive research is needed [26].

Huang et al. described several small molecular aggrecanase inhibitors that demonstrated chondroprotective activity in patients with OA. One of these compounds is AGG-523, a per os ADAMTS-4 and -5 inhibitor [27]. Animal models of OA demonstrated a reduced level of aggrecan fragments in joints [28]. AGG-523 was part of two Phase I studies, but these trials were discontinued due to unknown reasons [27].

A derivative of 5-(1H-pyrazol-4-yl) methylene)-2-thioxothiazolidin-4-one has been suggested to exhibit important activity against ADAMTS-5. Together with a hyaluronic acid hydrogel (HAX), this molecule was injected into the knees of rats. The results of the study indicate that ADAMTS-5 could be a promising target for disease-modifying OA treatment [29]. A 2017 study reported the effects of per os GLPG1972/S201086, a potent inhibitor of ADAMTS-5. GLPG1972/S201086 confirmed the important protective effect on cartilage and subchondral bone in posttraumatic OA by remarkably diminishing cartilage proteoglycan loss, cartilage impairment, and subchondral bone sclerosis. Furthermore, the safety parameters evaluated indicated that the drug was well tolerated [30]. A Phase II trial assessing the efficacy and safety of 3 doses of per os GLPG1972/ S201086 once daily in patients with knee OA was completed in July 2020, yet the final results have yet to be reported [31].

### 2.3. Wnt Inhibition

Wang et al. described Wnt as a glycoprotein with extracellular position, for which signaling engages 19 Wnt genes and receptors that are capable of managing canonical  $\beta$ -catenin-dependent and noncanonical  $\beta$ -catenin-independent signaling pathways [32]. The abovementioned pathways are responsible for various biological aspects involving the cartilage, thus confirming their pathological role in OA [33,34].

Small molecules such as XAV-939 and SM04690 were discovered and studied as potential inhibitors of Wnt signaling in patients with knee OA. Certain Phase I, II, and IIb studies highlighted the positive effects of SM04690 in terms of pain reduction, functional impairment, and JSW in addition to a seemingly good safety profile. These data suggest that SM04690 could be considered for use as a disease-modifying agent in patients with moderate to severe symptomatic knee OA [35–37].

Takada et al. and Grossmann et al., in their respective studies of StAx-35R (a staple  $\beta$ -catenin-binding domain of Axin) and SAH-Bcl9 (a staple peptide derived from the Bcl9 homology domain-2), described the involvement and role in  $\beta$ -catenin transcriptional activity [38].

Another molecule involved in Wnt/ $\beta$ -catenin signaling is LRP5 (lipoprotein receptor-associated protein 5), which stimulates catabolic factors responsible for OA cartilage destruction and inhibits the anabolic factor type II collagen. A study on LRP5-knockdown mice confirmed the benefits on cartilage [39,40].

Lorecivint inhibits the Wnt signaling pathway while also suppressing CLK2 (CDC-like kinase 2) and DYRK1A-mediated (dual-specificity tyrosine phosphorylation-regulated

kinase 1A) phosphorylation of SIRT1 and FOXO1, both involved in Wnt/ $\beta$ -catenin activity. The drug has been shown to be relatively safe and well-tolerated, with important outcomes regarding cartilage destruction and inflammation, by decreasing catabolic enzymes and blocking the inflammatory process [41]. Studies on patients with knee OA have demonstrated the favorable effects of lorecivivint on reducing symptoms (notably pain), and improving physical function and patient global assessment [42].

#### 2.4. Cathepsin K Inhibition

Cathepsin K is a lysosomal cysteine protease found in activated osteoclasts and chondrocytes (as well as other cell types) and has been shown to be involved in cartilage degradation (by destroying types I and II collagen and aggrecan found in the cartilage) and bone resorption [43,44].

Balicatib (AAE581), a cathepsin K inhibitor, was investigated in the treatment of patients with knee OA. The important side effects (skin rashes and dermal fibrosis) surpassed the beneficial outcome on both cartilage and bone, therefore leading to suspension of the study [45].

MIV-711 is described as being a selective and reversible agent that inhibits cathepsin K activity. MIV-711 holds an important role in decreasing serum CTX-I (a marker of bone turnover) and CTX-II levels (a marker of cartilage turnover), thus being implicated in both bone resorption and cartilage impairment [46]. MIV-711 was investigated in a multicenter, randomized, placebo-controlled, double-blind, three-arm parallel, Phase IIa study in patients with knee OA. The results showed that the reduction in medial femoral cartilage thickness was considerably reduced at 26 weeks. However, the medial tibia cartilage loss was not significantly changed. There was no substantial difference in pain reduction and quality of life scores or biomarker CTX-I and CTX-II values. The most important adverse events described were musculoskeletal symptoms, rashes, and infections. During the 6-month follow-up period, Conaghan et al. reported not finding significant benefits regarding OA-related symptoms. However, there is a need for further confirmation of MIV-711 as a DMOAD in OA through long-term trials [47].

#### 2.5. Osteogenic Protein-1

BMP-7 (bone morphogenetic protein-7 or osteogenic protein-7), a member of the (TGF)- $\beta$  superfamily, is a growth factor and is considered a possible therapeutic target in the process of restoration of damaged cartilage. Research analyzed BMP-7 (eptoterminal- $\alpha$ ) in the treatment of knee OA. A Phase I trial reported the amelioration of clinical symptoms and no dose-limiting toxicity. A Phase II trial and a Phase I trial also investigated BMP-7, but results have not yet been published [48].

#### 2.6. Sprifermin

Sprifermin (AS902330) is a recombinant human fibroblast growth factor-18 (rhFGF18). According to published data, sprifermin has positive effects on cartilage by stimulating cell multiplication and ECM constituent production [49,50]. On intraarticular administration, it binds FGFR3 receptors from the cartilage [51,52].

The study of Dahlberg et al. (first in-human trial, randomized, double-blind, placebo-controlled study) investigated patients who proposed joint arthroplasty due to severe OA of the knee. However, no significant differences in terms of symptom improvement were described between the treatment group and the placebo cohort [53].

Preliminary results at 3 years of an important study of sprifermin were presented by Hochberg et al. in 2019. The FORWARD study, a 5-year Phase II, dose-ranging, randomized trial investigated the effect of intraarticular injections of sprifermin administered every 6 or 12 months versus the placebo. The consequence of drug injection was a lower mean cartilage thickness loss compared to natural evolution (total femorotibial joint as well as in the medial, lateral, central medial, and central-lateral regions). Importantly, the FORWARD

study followed specifically structural progression, not other clinical issues associated with knee OA development and progression [54].

### 2.7. Platelet-Rich Plasma

Platelet-rich plasma (PRP) incorporates granules containing growth factors (transforming growth factor- $\beta$  (TGF- $\beta$ ), platelet-derived growth factor (PDGF), insulin-like growth factor (IGF), vascular endothelial growth factor (VEGF), and fibroblast growth factor (FGF)), cytokines, chemokines, and other mediators implicated in mesenchymal stem cell (MSC) proliferation and production of ECM and collagen, thus playing an important part in cartilage restoration [55,56].

Several randomized clinical trials have been completed in patients with knee OA, with intraarticular PRP being compared with hyaluronic acid. In terms of efficacy, the results of these studies revealed a favorable outcome in reducing symptoms and increasing mobility in these patients (with a mean duration of 12 months). Furthermore, no serious adverse effects were reported, suggesting a good safety profile for PRP [57–59]. However, there were a lot of discrepancies between these studies specifically in the protocols, methods, and patient characteristics; thus, it is challenging to compare all the data obtained [57].

### 2.8. Mesenchymal Stem Cells

Another promising treatment for cartilage repair seems to be MSCs derived from bone marrow, adipose tissue, or the umbilical cord. Studies have demonstrated that, compared with MSC injection, cell implantation has better results in patients with knee OA. Furthermore, there is a similar safety profile between autologous and allogenic MSCs [60,61].

A systematic review published by Jevotovsky et al. in 2018 evaluating 61 studies with MSCs as OA treatment concluded that MSC therapy has a favorable effect on OA patients but that there is limited high-quality evidence as well as a lack of long-term follow-up [62].

In 2015, Vega et al. presented a 1-year clinical trial that included patients with knee OA treated with intraarticular injections of allogeneic bone marrow MSCs. The results of this study showed considerable improvement in pain and functional indices, with a significant decrease in poor-quality cartilage areas and sustained augmentation of cartilage quality in the affected regions (quantified by magnetic resonance imaging (MRI) T2 mapping) [63].

An interesting aspect was represented by the possibility of treating MSCs before administration in order to boost the modulatory effect in OA. An example is combining MSCs with an inhibitor of signal transducer and activator of transcription 3. Under these conditions, the pro-inflammatory state was shown to be improved and the results of the treatment were more encouraging [64].

### 2.9. Gene Therapy

Gene therapy could be a promising alternative to OA treatment, mainly because of its long-term results on OA cartilage. Studies underline that intraarticular gene transfer treatment is a feasible option for patients suffering from OA compared to systemic administration with respect to safety, bioavailability, and direct targeting of the pathological site [65,66].

### 2.10. TPX-100

TPX-100, a peptide derived from matrix extracellular phosphoglycoprotein (MEPE), has expressed positive function when inducing articular cartilage regeneration in animal models. It is considered that this molecule facilitates cartilage formation only in areas with defects, without ectopic bone tissue formation [30].

A Phase II clinical study evaluating TPX-100 in patients with patellofemoral OA indicated significant amelioration and clinically efficient improvement in KOOS (Knee injury and Osteoarthritis Outcome Score) and WOMAC scores. Furthermore, promising effects were obtained concerning tibiofemoral cartilage thickness and volume at 6 and 12 months. Importantly, this performance was preserved for 30 months. The use of

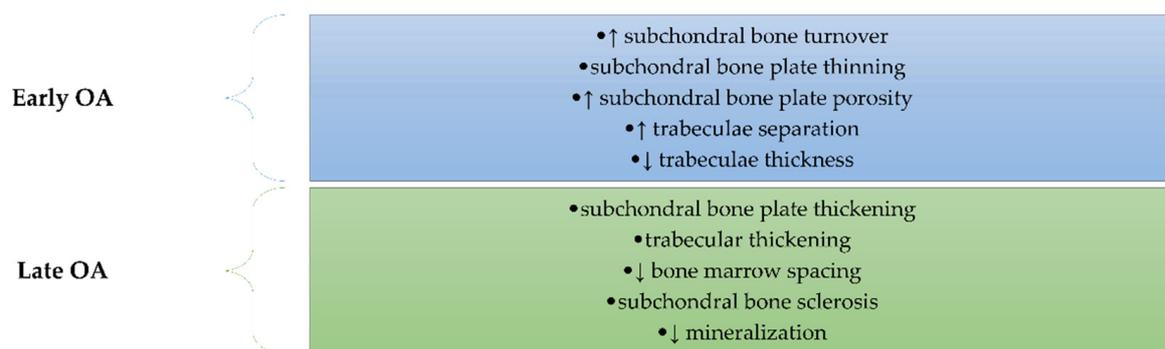
symptomatic treatment (nonsteroidal anti-inflammatory drugs (NSAIDs) and analgesics) diminished significantly across the study [67].

### 2.11. Symptomatic Slow-Acting Drugs for Osteoarthritis

A group of drugs known as SYSADOA (symptomatic slow-acting drugs for OA) have been used in patients with OA in order to improve clinical symptoms (pain and morning stiffness) as well as to slow progression of the disease. Among SYSADOA, chondroitin sulphate, glucosamine, and diacerein are the most widely studied. Using quantitative MRI (qMRI), clinical trials have demonstrated the contribution of chondroitin sulphate in decreasing cartilage volume loss [7,8]. Chondroitin sulphate influences proteoglycan metabolism and plays an important role in the balance of anabolic and catabolic processes at the ECM level. Chondroitin sulphate also decreases a series of proinflammatory factors with destructive properties in subchondral bone osteoblasts [9,10].

### 3. Therapeutic Approach to Bone Remodeling in Knee Osteoarthritis

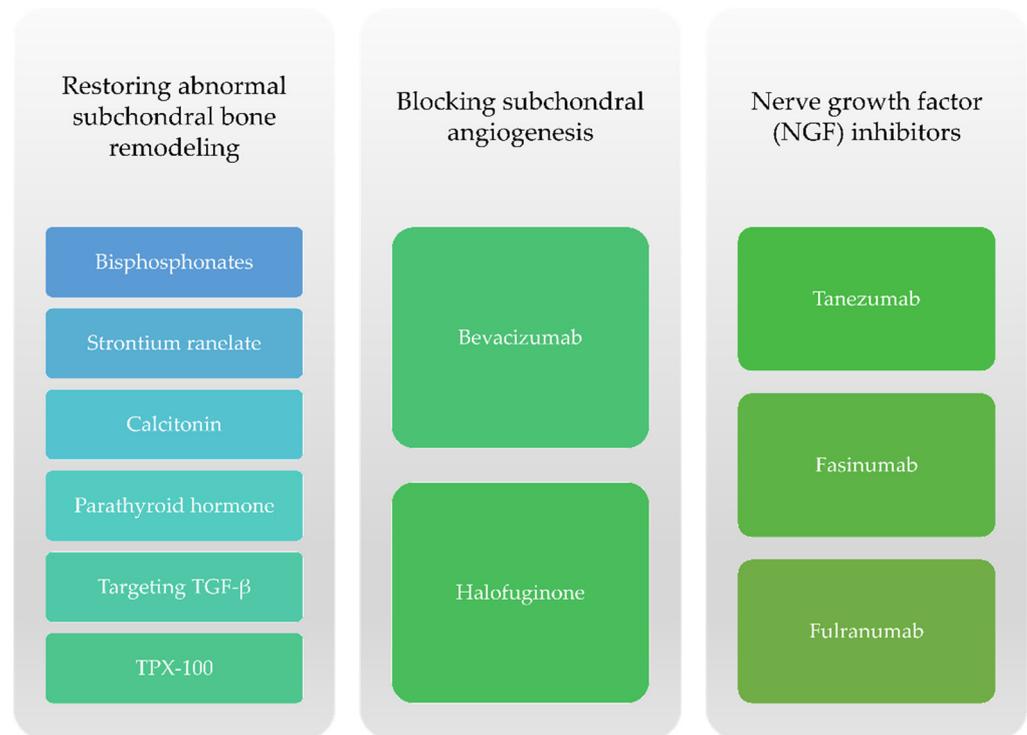
Subchondral bone remodeling plays a very important role in OA, mediating and preceding cartilage damage [68]. The structural changes in the subchondral bone are different depending on the stages of OA (Figure 3). Thus, in the early stage, there is an increase in subchondral bone turnover characterized by thinning of the subchondral bone plate and by increasing porosity associated with impairment of the trabeculae: thickness decreasing and separation increasing [69]. The late stage of OA is characterized by thickening of the plate and trabecular layers, by decreasing bone marrow spacing, and by sclerosis of the subchondral bone. Even if bone thickening occurs, there is insufficient bone mineralization due to a decreased calcium and collagen ratio followed by increased bone turnover [70–75].



**Figure 3.** The main structural changes in the subchondral bone occurs in OA depending on the stage of the disease (early or late). In the early stage, there is an increase in subchondral bone turnover characterized by thinning of the subchondral bone plate and by increasing porosity associated with impairment of the trabeculae: thickness decreasing and separation increasing. The late stage of OA is characterized by thickening of the plate and trabecular layers, by decreasing bone marrow spacing, by sclerosis of the subchondral bone, and by decreased mineralization.

Furthermore, the changes in subchondral bone are related to several signaling pathways such as the Wnt/ $\beta$ -Catenin, TGF- $\beta$ /Smad, RANK/RANKL/OPG (Receptor activator of nuclear factor kappa-B, RANK ligand, and osteoprotegerin), and MAPK (mitogen-activated protein kinase) signaling pathways [76–84]. Moreover, identification of the molecular pathways involved in OA-related bone remodeling has led to certain therapeutic agents targeting the mechanisms considered as possible DMOADs [85–90].

Regarding subchondral bone changes, therapies aim to block bone resorption caused by osteoclasts as well as other mechanisms such as novel angiogenesis or particular neuronal factors responsible for the onset of pain (Figure 4).



**Figure 4.** Therapeutic options targeting subchondral bone remodeling for knee OA.

### 3.1. Bisphosphonates

Bisphosphonates are antiresorptive drugs that can slow bone remodeling through the inhibition of osteoclasts. There are many published data that have shown pain relief, reduced bone destruction, and an improvement in joint structure following these treatments, but the clinical results are not very clear [91,92].

Alendronate, by inhibiting subchondral bone loss, caused an improvement in the structure of articular cartilage [93]. Other data showed an improvement of WOMAC pain score, an increase in bone mineral density, and a decrease in markers of bone destruction after alendronate administration for 2 years in patients with OA [91].

Risedronate use has not been shown to be clinically and radiologically effective in a group of 2483 patients with knee OA after 2 years of follow-up [94]. On the other hand, Spector et al. showed an improvement in pain and a decrease in subchondral remodeling following risedronate treatment [95].

The favorable effect of zoledronic acid was highlighted in a study that included 59 patients with knee OA. After 6 months of treatment, a significant reduction in pain was observed as well as a reduction in bone marrow lesions detected by magnetic resonance imaging [96]. Less favorable results (in relieving pain and loss of cartilage volume) after 24 months of treatment with zoledronic acid in knee OA cases were published by Aitken [91,92].

Clodronate, a non-amino bisphosphonate, appears to have a favorable effect in patients with knee OA, acting by increasing the secretion of SOX9, the transcription factor responsible for progenitor stem cell chondrogenic commitment. A 12-week, randomized, placebo-controlled study that included 80 cases of knee OA showed an improvement in pain and decreases in WOMAC score and in the need for analgesic medication after once weekly intraarticular injection of 2 mg clodronate [97].

### 3.2. Strontium Ranelate

Strontium ranelate can restore the abnormal subchondral bone remodeling by decreasing the activity of osteoclasts and by favoring the mineralization of new bone. After 3 years of treatment, patients with knee OA had an improvement in joint space narrowing

compared to placebo [98]. At a dose of 1800 mg/day, there was a decrease in chondrocyte apoptosis and an improvement in cartilage matrix [99]. At a higher dose—2000 mg/day—strontium ranelate showed better effects in terms of loss of cartilage volume and regarding the improvement of bone marrow lesions [100].

### 3.3. Calcitonin

Calcitonin, a hormone secreted by parafollicular thyroid cells, binds to specific receptors on osteoclasts and inhibits their activity, thus reducing damage to the subchondral bone in OA [101,102]. In patients with knee OA, the use of oral calcitonin has been shown to have an effect in relieving pain and in increasing cartilage volume compared to placebo [103]. Due to the high risk of neoplasms, the use of calcitonin for short periods of time is recommended [104].

### 3.4. Cathepsin K Inhibition

Cathepsin K inhibitors are antiresorptive drugs that inhibit osteolytic protease of osteoclasts. There are several animal studies that have shown the beneficial role of these molecules in stopping the progression of OA [105,106]. In studies, there are 2 tested molecules: an oral inhibitor called MIV-711, for which structural changes in the joints were not considered, and balicatib (AAE581) used in the knee OA, in which narrowing of the articular space and the cartilage volume followed [107,108].

### 3.5. Parathyroid Hormone

Teriparatide, a recombinant human parathyroid hormone (PTH), is a bone anabolic therapy that regulates endochondral ossification and inhibits chondrocyte hypertrophy [109]. Systemic administration of teriparatide can inhibit cartilage degradation and abnormal chondrocyte differentiation, can stimulate regeneration of the matrix, and can improve the structure of the subchondral bone [110,111]. There are several ongoing studies that aim to highlight the beneficial effects of this molecule on both subchondral bone and joint cartilage, being assimilated as chondroregenerative therapy.

### 3.6. Transforming Growth Factor $\beta$ Inhibition

TGF- $\beta$  gives positive feedback with the Wnt signaling pathway, favoring the differentiation of chondrocytes and osteoblasts [111]. In OA, TGF- $\beta$  is secreted in excess by osteoblasts and causes the development of osteophytes [112]. Using a murine model, studies have highlighted the role of neutralizing TGF- $\beta$  antibodies in stopping OA progression [113]. Other data have shown that implantation of an anti-TGF- $\beta$  antibody (1D11) in alginate acid microbeads in the subchondral bone or deletion of the TGF- $\beta$  type II receptor (T $\beta$ RII) inhibits Smad2/3 phosphorylation in osteoblastic precursors, protecting the osteochondral unit [114].

### 3.7. TPX-100

MEPE is a bone protein secreted by osteocytes, with a negative role in regulating bone mineralization and involved in remodeling the subchondral bone [115]. A 23-aminoacid derived from MEPE (TPX-100) has been shown to be effective in cases of moderate femuropatellar OA, leading to significant improvement in WOMAC scores. However, there was no evidence of structural improvement after 12 months of treatment [116].

### 3.8. Subchondral Angiogenesis Inhibition

Subchondral angiogenesis acts as a bridge between the articular cartilage and the subchondral bone. Blocking neoangiogenesis, the data showed a decrease in subchondral bone loss and a reduction in cartilage degradation [117]. Numerous factors such as VEGF (vascular endothelial growth factor), TGF- $\beta$ 1, PDGF-BB (platelet-derived growth factor BB monomer), and SLIT3 participate in the process of angiogenesis in the subchondral bone [117]. Halofuginone stops the action of TGF- $\beta$ 1 by inhibiting Smad2/3, thus blocking

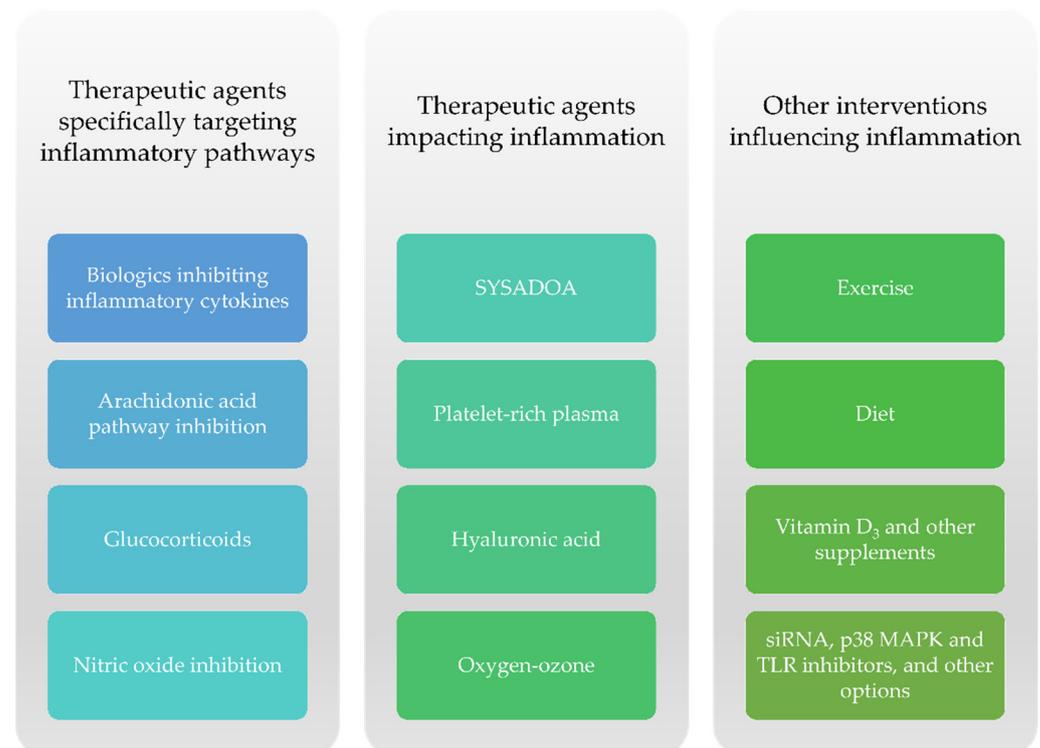
subchondral angiogenesis [118]. Bevacizumab, an antibody against VEGF, has been shown to be effective in reducing the formation of subchondral blood vessels, thereby inhibiting chondrocyte hypertrophy [119].

### 3.9. Nerve Growth Factor (NGF) Inhibition

Abnormal remodeling of the subchondral bone is associated with the presence of particular neural factors that cause innervation of sensory nerve structures in cases of OA [120]. NGFs are secreted by preosteoclasts, being triggers of subchondral bone innervation in OA [121]. NGF inhibitors such as tanezumab, fasinumab, or fulranumab have been shown to be effective in ameliorating joint pain and function in patients with OA [122,123]. For tanezumab, the data showed a significant improvement in pain [122,123], while fasinumab is still under investigation [124].

## 4. Therapeutic Approach to Synovial Inflammation in Knee Osteoarthritis

The appearance of low-grade inflammation remains one of the most prominent features in OA pathogenesis. Proinflammatory cytokines such as IL-1, IL-6, and TNF $\alpha$  may be upregulated and may promote the release of ROS (reactive oxygen species), MMPs, and ADAMTS [125]. Though numerous emerging drugs were tested in degenerative joint conditions, a few studies focused specifically on their impact on knee OA-related inflammation (systemic or synovial fluid levels of inflammatory markers, the presence of joint effusion, or prolonged morning stiffness) [126–130]. Nevertheless, several drugs and dietary supplements as well as lifestyle interventions (diet and physical exercise) have been shown to exhibit disease-modifying properties in knee OA (Figure 5).



**Figure 5.** Drugs, dietary supplements, and other interventions contributing to a reduction in inflammation in knee OA.

### 4.1. Biologics Targeting Proinflammatory Cytokines

Whereas they are more commonly used in immune-inflammatory diseases such as rheumatoid arthritis, psoriasis, ankylosing spondylitis, juvenile idiopathic arthritis, and inflammatory bowel diseases, certain TNF $\alpha$  inhibitors have also been studied in knee OA [131]. An open-label study including 56 patients with knee OA compared the efficacy

of a single intraarticular injection with 10 mg adalimumab (ADA) versus 25 mg hyaluronic acid (HA), obtaining better results in terms of pain and functional impairment in the TNF inhibitor subgroup compared to their counterparts (Table 1) [132]. However, the follow-up period was short (4 weeks) and all patients were under concomitant treatment with systemic NSAIDs (celecoxib 200 mg daily) [132].

Another research conducted on 39 patients suggested that intraarticular injections with etanercept (ETN) could be superior to hyaluronic acid for pain relief in moderate to severe knee OA. While this was true for WOMAC pain score improvement at 4 weeks, the differences in mean VAS (Visual Analogue Scale) levels between the two study groups failed to reach statistical significance at the end of the follow-up period [133].

A multicenter, randomized, double-blind, placebo-controlled study of 50 mg and 150 mg intraarticular anakinra (ANR) versus placebo found no significant benefits for ANR over 12 weeks [134]. Wang et al. conducted a placebo-controlled, randomized, double-blind, ascending dose research focused on anti-interleukin-1 $\alpha$ / $\beta$  dual variable domain immunoglobulin ABT-981 in patients with symptomatic knee OA, identifying lower levels of high-sensitivity C-reactive protein (hsCRP) as well as IL-1 $\alpha$  and IL-1 $\beta$  in the treatment group [135].

**Table 1.** Proinflammatory cytokine blockers adalimumab, etanercept, anakinra, and ABT-981 in knee OA.

Reference	Compound	Intervention	Patients	Duration	Results
Wang [132]	Adalimumab (ETA)	10 mg intraarticular ADA + Celecoxib 200 mg/day versus 25 mg HA + Celecoxib 200 mg/day	ADA + Celecoxib (N = 28) HA + Celecoxib (N = 28)	4 weeks	The authors found a significant improvement in pain and functionality in the ADA group.
Ohtori et al. [133]	Etanercept (ETN)	10 mg intraarticular ETN versus 25 mg HA	ETN (N = 19) HA (N = 20)	4 weeks	An initial significant amelioration was obtained in the ETN group compared to HA (weeks 1 and 2), yet the results were not maintained at week 4. A significant pain improvement was observed in the 150 mg ANR group compared to 50 mg ANR at day 4. Overall, intraarticular ANR did not demonstrate notable benefits, irrespective of the dose.
Chevalier et al. [134]	Anakinra (ANR)	150 mg intraarticular ANR versus 50 mg intraarticular ANR versus Placebo	ANR 150 mg (N = 67) ANR 50 mg (N = 34) Placebo (N = 69)	12 weeks	Mean hsCRP decreased through week 2 irrespective of ABT-981 dose/administration interval. While IL-1 $\alpha$ and IL-1 $\beta$ were lower in the treatment group, serum vascular endothelial growth factor and MMP-9 did not demonstrate significant changes.
Wang et al. [135]	ABT-981	ABT-981 (various doses) versus Placebo	ABT-981 0.3 mg/kg fortnightly (N = 7) 1 mg/kg fortnightly (N = 7) 3 mg/kg fortnightly (N = 7) 3 mg/kg every 4 weeks (N = 7) Placebo (N = 8)	113 days (cohorts 1, 2, and 3) 127 days (cohort 4)	

#### 4.2. Arachidonic Acid Pathway Inhibition

The arachidonic acid pathway employs cyclooxygenase enzymes (COX) and prompts the release of proinflammatory prostaglandins. The latter are also involved in pain mediation, which is why NSAIDs are widely prescribed as symptomatic treatment in OA [136]. However, certain NSAIDs have additionally been considered for use as disease-modifying agents [137,138]. A study performed on knee OA patients examined the impact of cele-

coxib, ibuprofen, and diclofenac on synovial fluid proinflammatory cytokine (TNF $\alpha$ , IL-6, and IL-8) and VEGF expression [139]. Aside from significant pain relief, NSAID therapy (particularly higher doses) showed notable beneficial effects in reducing inflammatory markers in patients' synovial fluid, thus suggesting that these drugs could be regarded as having disease-modifying properties [139]. Nevertheless, the numerous adverse events associated with prolonged administration of systemic NSAIDs limit their use primarily in older individuals and persons with comorbid conditions [140].

The dual hindrance of COX2 and 5-lipoxygenase (LOX) diminished the release of prostaglandin E2, leukotriene B4, collagenase-1, cathepsin, MMP-13, and IL-1 $\beta$  [141]. Moreover, COX/LOX blockade diminished OA-related cartilage degradation and synovial hypertrophy in preclinical studies [138,141]. Licofelone is a COX/LOX inhibitor that displayed a protective effect on knee cartilage volume loss in a multicenter clinical trial [142].

#### 4.3. Glucocorticoids

Glucocorticoids exhibit potent anti-inflammatory effects and have been shown to reduce pain and pain-related functional impairment in patients with OA. However, intraarticular or systemic corticosteroid administration has seldom been investigated with respect to its effect on clinical or paraclinical indicators of inflammation in knee OA (joint effusion and stiffness, synovial fluid inflammatory markers, and imaging tests) [143–145]. Furthermore, the risk–benefit ratio for using intraarticular glucocorticoids in knee OA remains a matter of discussion among researchers, clinicians, and medical organizations alike [146].

Nevertheless, a study evaluating a combination of triamcinolone acetonide and sodium hyaluronate was superior to intraarticular saline in improving joint stiffness, pain, and function in patients with knee OA with no or minimal to moderate joint space narrowing (Kellgren–Lawrence grades 1–3) [147]. A recent study investigating the effect of intraarticular injections with triamcinolone found that the latter reduced joint effusion in patients with knee OA [148].

#### 4.4. Symptomatic Slow-Acting Drugs for Osteoarthritis

The DISSCO study examined the efficacy of diacerein (diacetylrhein, an anthraquinone derivative, and SYSADOA) compared to celecoxib on knee osteoarthritis. The multicenter randomized trial concluded that diacerein was non-inferior to celecoxib. The percentage of patients experiencing joint effusion diminished by half over time, however, without notable discrepancies between the two treatment groups [149]. In rat synovial cells, diacerein reduced the mRNA levels of inflammatory cytokines (TNF $\alpha$ , IL-1, and IL-6), COX2, cartilage-degrading enzymes (MMP-3 and MMP-13), and ADAMTS-5 in a dose-dependent manner while also raising those of certain anti-inflammatory factors (IL-4 and IL-10). Moreover, the analysis of biopsy samples taken from rats with monosodium iodoacetate-induced OA as well as imaging tests (computed tomography) showed that the administration of diacerein-loaded nanoparticles may protect against joint destruction [150].

Other SYSADOAs such as glucosamine and chondroitin sulphate have also been thought to display anti-inflammatory properties together with benefits regarding cartilage degradation in patients with knee OA, yet results remain discrepant across studies [7–10].

#### 4.5. Nitric Oxide Inhibition

Nitric oxide (NO) together with inducible NO synthase (iNOS, an enzyme that participates in NO synthesis) are involved in OA-related inflammation, cartilage damage, and cellular injury. The release of proinflammatory cytokines such as TNF $\alpha$ , IL-1 $\beta$ , IL-17, and interferon  $\gamma$  (IFN $\gamma$ ) has been shown to upregulate the expression of iNOS, thus emphasizing the link between inflammation and oxidative stress [151,152].

Scientific evidence highlighting the involvement of iNOS in OA pathogenesis prompted testing of iNOS inhibitors in human subjects. A randomized double-blind placebo-

controlled trial of the iNOS inhibitor cindunistat (50 or 200 mg daily) was conducted on patients with symptomatic knee OA. In the Kellgren–Lawrence grade 2 subgroup, cindunistat 50 mg daily was associated with a delay in joint space narrowing at 48 weeks compared to the placebo. Nevertheless, these results were not maintained at 96 weeks [153]. Furthermore, the Kellgren–Lawrence grade 3 cohort did not show a significant improvement with respect to slowing knee OA progression with iNOS inhibition, irrespective of cindunistat dose [153].

#### 4.6. Platelet-Rich Plasma

PRP is an autologous formulation containing concentrated blood-derived platelets and growth factors that promotes synoviocyte differentiation and proliferation, with studies reporting significant benefits in OA of the knee [154,155]. Moreover, PRP is thought to be rich in bioactive molecules involved in tissue repair and regeneration and to wield anti-inflammatory properties by lowering synovial fluid proinflammatory marker levels [156–159]. Notably, some authors support the use of leukocyte-poor PRP (LP-PRP), arguing that the presence of white blood cells in intraarticular formulation may generate pro-inflammatory effects [160]. PRP has been deemed safe for use and has demonstrated promising results in different chronic rheumatic conditions and injury-related joint damage [161]. In knee OA, PRP displayed positive effects in improving pain, joint function, and quality of life according to recently published meta-analyses [157,162].

In young adults (18–30 years of age) diagnosed with knee OA, intraarticular PRP therapy led to a statistically significant decrease in IL-1 $\beta$ , IL-6, TNF $\alpha$ , IL-17A, RANKL, and IFN $\gamma$  levels (Table 2) [163]. A study performed on eutrophic individuals (BMI between 22–25 kg/m<sup>2</sup>) aged between 42 and 79 years comparing intraarticular PRP, HA, and the combination between PRP and HA found that IL-1 $\beta$ , TNF $\alpha$ , TIMP1, and MMP-3 values were diminished in the PRP group at 6 months posttreatment [164]. Nevertheless, the association of PRP and HA held the best results in reducing proinflammatory markers [164].

**Table 2.** Platelet-rich plasma (PRP) treatment in knee OA: anti-inflammatory properties.

Reference	Intervention	Patients	Duration	Results
Huang et al. [163]	2–14 mL intraarticular PRP weekly versus Placebo (10 mL saline solution) weekly	PRP (N = 310) Placebo (N = 56)	8 weeks	Significant improvements in plasma IL-1 $\beta$ , IL-6, TNF $\alpha$ , IL-17A, RANKL, and IFN $\gamma$ were observed in the treatment group. IL-1 $\beta$ , TNF $\alpha$ , TIMP1, and MMP-3 demonstrated
Xu et al. [164]	4 mL intraarticular PRP, 3 injections per knee, half-month interval 2 mL intraarticular HA, 3 injections per knee, half-month interval 4 mL PRP + 2 mL HA, 3 injections per knee, half-month interval	PRP (N = 40 knees) HA (N = 34 knees) PRP + HA (48 knees) N = 78 patients total (122 knees) N = 44 patients received bilateral injections	24 months	significant decreases in the PRP group at 6 months posttreatment. Nevertheless, the PRP + HA group showed better results in this respect. Additionally, the PRP + HA cohort displayed IL-1 $\beta$ , TNF $\alpha$ , TIMP1, and MMP-3 inhibition at 12 months post-injection.

#### 4.7. Hyaluronic Acid

A randomized, double-blind, placebo-controlled study of obese patients with knee OA examined the outcomes of a per os preparation containing HA (70%) and other glycosaminoglycans (GAGs) [165]. At 3 months, the treatment group exhibited a significant decrement in TNF $\alpha$ , IL-1 $\alpha$  and IL-1 $\beta$ , IL-6, IL-17 $\alpha$ , IFN, and GM-CSF (granulocyte-macrophage colony-stimulating factor) levels, while the placebo cohort demonstrated notably higher synovial fluid concentrations of proinflammatory cytokines as well as leptin [165].

A recent research comparing the efficacy of intraarticular HA to ozone (O<sub>3</sub>) found that both reduced joint stiffness in patients with knee OA [166]. Other published studies aimed to investigate the possible differences between intraarticular HA and biologics, PRP, or O<sub>3</sub> (Table 3) [132,133,164–166]. Xu et al. reported positive results for a PRP + HA combination with respect to the reduction in proinflammatory cytokine values and MMP-3 [164].

**Table 3.** Hyaluronic acid (HA) treatment in knee OA: anti-inflammatory properties.

Reference	Intervention	Patients	Duration	Results
Nelson et al. [165]	80 mg oral preparation of HA (70%) + other GAGs versus Placebo	Oral preparation of HA + other GAGs (N = 21) Placebo (N = 19)	12 weeks	The HA-treated cohort demonstrated a notable decrease in TNF $\alpha$ , IL-1 $\alpha$ , IL-1 $\beta$ , IL-6, IL-17 $\alpha$ , IFN, and GM-CSF values. Furthermore, the placebo group exhibited significantly higher synovial fluid concentrations of inflammatory cytokines as well as leptin (a proinflammatory adipokine).
Wang [132]	25 mg HA + Celecoxib 200 mg/day versus 10 mg intraarticular ADA + Celecoxib 200 mg/day	HA + Celecoxib (N = 28) ADA + Celecoxib (N = 28)	4 weeks	Changes in joint stiffness did not exhibit statistically significant differences between HA + Celecoxib and ADA + Celecoxib.
Ohtori et al. [133]	25 mg HA versus 10 mg intraarticular ETN	HA (N = 20) ETN (N = 19)	4 weeks	The HA group displayed significantly weaker results in terms of joint stiffness improvement compared to ETN during the follow-up period.
Xu et al. [164]	2 mL intraarticular HA, 3 injections per knee, half-month interval 4 mL intraarticular PRP, 3 injections per knee, half-month interval 2 mL HA + 4 mL PRP, 3 injections per knee, half-month interval 3 weekly intraarticular injections of 20 mg/2 mL HA	HA (N = 34 knees) PRP (N = 40 knees) HA + PRP (48 knees)	24 months	The cohort that received the HA + PRP combination demonstrated IL-1 $\beta$ , TNF $\alpha$ , TIMP1, and MMP-3 inhibition at one-year posttreatment.
Raeissadat et al. [166]	3 weekly intraarticular injections of 30 $\mu$ g/mL O <sub>3</sub> (10 mL) versus 3 weekly intraarticular injections of 20 mg/2 mL HA	HA (N = 74) O <sub>3</sub> (N = 67)	24 weeks	There was a significant reduction in joint stiffness in the group treated with HA. However, these results were not significantly different from the O <sub>3</sub> -treated cohort.

#### 4.8. Oxygen–Ozone

A study of intraarticular injections of oxygen–ozone (O<sub>2</sub>–O<sub>3</sub>) versus triamcinolone in knee OA determined that joint effusion was notably reduced in both treatment arms [148]. Whereas the decrement in WOMAC and VAS values were in favor of the O<sub>2</sub>–O<sub>3</sub> group, the differences compared to corticosteroid-treated patients did not reach statistical significance with respect to the reduction in joint effusion on ultrasound [148]. Other published research reported finding a reduction in joint stiffness in knee OA patients following treatment with O<sub>3</sub> (Table 4) [148,166,167].

**Table 4.** O<sub>3</sub> and O<sub>2</sub>–O<sub>3</sub> treatment in knee OA: anti-inflammatory properties.

Reference	Intervention	Patients	Duration	Results
Babaei-Ghazani et al. [148]	15 µg/mL intraarticular O <sub>2</sub> –O <sub>3</sub> (10 mL) versus 40 mg intraarticular triamcinolone (1 mL)	O <sub>2</sub> –O <sub>3</sub> (N = 31) Triamcinolone (N = 31)	12 weeks	The authors found an important reduction in joint effusion on ultrasound in both treatment arms at 3 months post-injection.
Lopes de Jesus et al. [167]	20 µg/mL intraarticular O <sub>3</sub> (10 mL) versus Placebo (10 mL air) 3 weekly injections of 30 µg/mL intraarticular O <sub>3</sub> (10 mL)	O <sub>3</sub> (N = 61) Placebo (N = 35)	8 weeks	There was a significant improvement in joint stiffness at 8 weeks in the treatment arm. Both groups demonstrated a notable amelioration in joint stiffness yet without significant discrepancies between the two treatment arms.
Raeissadat et al. [166]	3 weekly injections of 20 mg/2 mL HA	O <sub>3</sub> (N = 67) HA (N = 74)	24 weeks	

#### 4.9. Exercise, Diet, and Supplements

Diet and lifestyle changes have also been shown to reduce inflammation in knee OA. The IDEA (Intensive Diet and Exercise for Arthritis) study enrolled over 400 overweight and obese individuals (BMI between 27–40.5 kg/m<sup>2</sup>) with symptomatic knee OA. The research subjects were randomized to three intervention groups (exercise, diet, or diet and exercise) and followed the program for 18 months [168]. Both diet as well as the combination of diet and exercise significantly lowered serum inflammatory marker levels [168].

Another research performed on women with knee OA examining the role of resistance training and 1000 mg daily nanocurcumin (turmeric) showed that the latter reduced synovial fluid NO concentrations by over 26% and collagenase II by circa 4.7% [169]. However, the authors commented that the short duration of the study (16 weeks) could explain the lack of statistically significant differences between the 3 intervention groups (resistance exercise, curcumin supplementation, and exercise and supplementation) [169].

Vitamin D-deficient individuals were found to be at risk of developing knee OA [160]. The impact of vitamin D supplementation on pain relief, the improvement in functional parameters, and cartilage loss remain matters of debate [170,171]. However, an MRI study analyzing the effect of 50,000 IU monthly vitamin D<sub>3</sub> supplementation in patients with knee OA found that the placebo group exhibited a significant increase in effusion-synovitis volume over two years compared to the treatment cohort [172].

It has been stated that old age may be associated with intestinal dysbiosis and increased gut permeability, which could promote a “leakage” of gut microbes into tissues, thus leading to a chronic proinflammatory state [173,174]. Tsai et al. aimed to characterize the immunological signature of the microbiome in osteoarthritic knee synovium, finding marked discrepancy in microbial abundance in patients compared to healthy controls [175]. Whereas intestinal dysbiosis has been linked to synovial inflammation in knee OA, the impact of probiotic use on disease progression have not been studied extensively in clinical trials [176,177]. Certain experimental studies reported promising results with respect to

the administration of *Lactobacillus acidophilus* in rats. In this regard, *Lactobacillus acidophilus* prompted the diminishment in proinflammatory marker expression together with an increase in anti-inflammatory cytokines in monosodium iodoacetate-induced OA. Moreover, *Lactobacillus acidophilus* exhibited anti-nociceptive effects in experimental animals [178].

It has been stated that dietary polyphenols and herbal medicine may also exhibit anti-inflammatory properties and reduce iNOS expression in OA [151,179]. Scoparone is a compound extracted from *Artemisia capillaris* that may exhibit potent anti-inflammatory effects. A recent study proved that scoparone could repress IL-1 $\beta$ -induced activation of the PI3K/Akt/NF- $\kappa$ B pathway as well as COX2 and iNOS [180]. In murine models of OA, acteoside (withdrawn from *Ligustrum purpurascens* kudingcha tea) reduced the expression of TNF $\alpha$ , IL-6, and IFN $\gamma$  and hindered the IL-1 $\beta$ -related activation of the JAK/STAT (janus kinase/signal transducer and activator of transcription) pathway [181].

Avocado soybean unsaponifiables were studied in knee OA, showing positive results. Moreover, avocado soybean unsaponifiables diminished the expression of iNOS, MMP-13, and TNF $\alpha$  in rat monosodium iodoacetate-induced OA [182].

#### 4.10. Other Therapeutic Options Exhibiting Antiinflammatory Properties

The inhibition of TNF $\alpha$  and pan-cytokine small interference RNA (siRNA) led to a decrease in proinflammatory cytokines (IL-1, IL-6, IL-8, and TNF $\alpha$ ), with promising findings being recorded in collagenase-induced murine models of OA [183–185].

P38 mitogen-activated protein kinase (MAPK), toll-like receptors (TLR2 and TLR4), and leptin have been considered potential targets for the development of new DMOAD options based on their experimentally shown involvement in OA-related inflammation [138]. Though in vitro and animal studies reported certain encouraging results, the efficacy of siRNA, p38 MAPK, TLR, and inflammatory adipokine blockade remains in need of further testing in clinical trials of knee OA [138].

Carboxymethyl chitosan (a chitosan soluble derivative) displays a significant physicochemical likeness to cartilage proteoglycans. In rat chondrocytes, increasing doses of carboxymethyl chitosan demonstrated iNOS inhibition and upregulated anti-inflammatory cytokine expression (IL-10) [186].

## 5. Conclusions

OA is a chronic and pathogenically multifaceted disease that remains a major cause of disability worldwide. Cartilage degradation, inflammation, and bone remodeling are presently regarded as therapeutic targets for the development of DMOADs. However, the pathogenic intricacies between the molecular pathways involved in OA prompted the study of certain drugs for more than one therapeutic target (amelioration of cartilage and bone changes, and inflammation). Most clinical studies regarding knee OA focus mainly on improvement in pain or joint function and thus do not provide sufficient evidence on the possible disease-modifying properties of the tested drugs. Currently, there is an unmet need for further research regarding OA pathogenesis as well as the introduction and exhaustive testing of potential disease-modifying pharmacotherapies in order to structure an effective treatment plan for these patients.

**Funding:** The present research received no external funding.

**Data Availability Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Roemer, F.W.; Kwoh, C.K.; Hayashi, D.; Felson, D.T.; Guermazi, A. The role of radiography and MRI for eligibility assessment in DMOAD trials of knee OA. *Nat. Rev. Rheumatol.* **2018**, *14*, 372–380. [[CrossRef](#)]
2. Malfait, A.-M.; Tortorella, M.D. The “elusive DMOAD”: Aggrecanase inhibition from laboratory to clinic. *Clin. Exp. Rheumatol.* **2019**, *37*, 130–134.

3. Castrogiovanni, P.; Di Rosa, M.; Ravalli, S.; Castorina, A.; Guglielmino, C.; Imbesi, R.; Vecchio, M.; Drago, F.; Szychlinska, M.A.; Musumeci, G. Moderate Physical Activity as a Prevention Method for Knee Osteoarthritis and the Role of Synovocytes as Biological Key. *Int. J. Mol. Sci.* **2019**, *20*, 511. [CrossRef]
4. Yeap, S.S. Current DMOAD options for the treatment of osteoarthritis. *Clin. Exp. Rheumatol.* **2020**, *38*, 802.
5. Roman-Blas, J.A.; Bizzi, E.; Largo, R.; Migliore, A.; Herrero-Beaumont, G. An update on the up and coming therapies to treat osteoarthritis, a multifaceted disease. *Expert Opin. Pharmacother.* **2016**, *17*, 1745–1756. [CrossRef]
6. Rezuş, E.; Cardoneanu, A.; Burlui, A.; Luca, A.; Codreanu, C.; Tamba, B.I.; Stanciu, G.-D.; Dima, N.; Bădescu, C.; Rezuş, C. The Link Between Inflammation and Degenerative Joint Diseases. *Int. J. Mol. Sci.* **2019**, *20*, 614. [CrossRef]
7. Martel-Pelletier, J.; Raynauld, J.-P.; Mineau, F.; Abram, F.; Paiement, P.; Delorme, P.; Pelletier, J.-P. Levels of serum biomarkers from a two-year multicentre trial are associated with treatment response on knee osteoarthritis cartilage loss as assessed by magnetic resonance imaging: An exploratory study. *Arthritis Res.* **2017**, *19*, 169. [CrossRef]
8. Vreju, F.A.; Ciurea, P.L.; Rosu, A.; Chisalau, B.A.; Parvanescu, C.D.; Firulescu, S.C.; Stiolica, A.T.; Barbulescu, A.L.; Dinescu, S.C.; Dumitrescu, C.I.; et al. The Effect of glucosamine, chondroitin and harpagophytum procumbens on femoral hyaline cartilage thickness in patients with knee osteoarthritis—An MRI versus ultrasonography study. *J. Mind Med. Sci.* **2019**, 162–168. [CrossRef]
9. Reginster, J.; Veronese, N. Highly purified chondroitin sulfate: A literature review on clinical efficacy and pharmaco-economic aspects in osteoarthritis treatment. *Aging Clin. Exp. Res.* **2020**, *33*, 37–47. [CrossRef] [PubMed]
10. Lane, N.; Shidara, K.; Wise, B. Osteoarthritis year in review 2016: Clinical. *Osteoarthr. Cartil.* **2017**, *25*, 209–215. [CrossRef] [PubMed]
11. United States Food and Drugs Administration. Available online: <https://www.fda.gov/regulatory-information/search-fda-guidance-documents/osteoarthritis-structural-endpoints-development-drugs> (accessed on 11 January 2021).
12. European Medicines Agency. Available online: [https://www.ema.europa.eu/en/documents/scientific-guideline/guideline-clinical-investigation-medicinal-products-used-treatment-osteoarthritis\\_en.pdf](https://www.ema.europa.eu/en/documents/scientific-guideline/guideline-clinical-investigation-medicinal-products-used-treatment-osteoarthritis_en.pdf) (accessed on 3 January 2021).
13. Fosang, A.J.; Little, C.B. Drug Insight: Aggrecanases as therapeutic targets for osteoarthritis. *Nat. Clin. Pract. Rheumatol.* **2008**, *4*, 420–427. [CrossRef] [PubMed]
14. Murphy, G.; Nagase, H. Reappraising metalloproteinases in rheumatoid arthritis and osteoarthritis: Destruction or re-pair? *Nat. Clin. Pract. Rheumatol.* **2008**, *4*, 128–135. [CrossRef]
15. Chow, Y.Y.; Chin, K.-Y. The Role of Inflammation in the Pathogenesis of Osteoarthritis. *Mediat. Inflamm.* **2020**, *2020*, 1–19. [CrossRef] [PubMed]
16. Bauvois, B. New facets of matrix metalloproteinases MMP-2 and MMP-9 as cell surface transducers: Outside-in signaling and relationship to tumor progression. *Biochim. Biophys. Acta BBA Bioenerg.* **2012**, *1825*, 29–36. [CrossRef]
17. Krzeski, P.; Buckland-Wright, C.; Bálint, G.; Cline, G.A.; Stoner, K.; Lyon, R.; Beary, J.; Aronstein, W.S.; Spector, T.D. Development of musculoskeletal toxicity without clear benefit after administration of PG-116800, a matrix metalloproteinase inhibitor, to patients with knee osteoarthritis: A randomized, 12-month, double-blind, placebo-controlled study. *Arthritis Res. Ther.* **2007**, *9*, R109. [CrossRef] [PubMed]
18. Drake, C.G. Book Review of “Tumor Immunology and Cancer Vaccines”. *Cancer Investig.* **2006**, *24*, 657. [CrossRef]
19. Baragi, V.M.; Becher, G.; Bendele, A.M.; Biesinger, R.; Bluhm, H.; Boer, J.; Deng, H.; Dodd, R.; Essers, M.; Feuerstein, T.; et al. A new class of potent matrix metalloproteinase 13 inhibitors for potential treatment of osteoarthritis: Evidence of histologic and clinical efficacy without musculoskeletal toxicity in rat models. *Arthritis Rheum.* **2009**, *60*, 2008–2018. [CrossRef]
20. Pinney, J.R.; Taylor, C.; Doan, R.; Burghardt, A.J.; Li, X.; Kim, H.T.; Ma, C.B.; Majumdar, S. Imaging longitudinal changes in articular cartilage and bone following doxycycline treatment in a rabbit anterior cruciate ligament transection model of osteoarthritis. *Magn. Reson. Imaging* **2012**, *30*, 271–282. [CrossRef]
21. da Costa, B.; Nüesch, E.; Reichenbach, S.; Jüni, P.; Rutjes, A. Doxycycline for osteoarthritis of the knee or hip. *Cochrane Database Syst. Rev.* **2012**. [CrossRef]
22. Dubail, J.; Apte, S.S. Insights on ADAMTS proteases and ADAMTS-like proteins from mammalian genetics. *Matrix Biol.* **2015**, *44–46*, 24–37. [CrossRef]
23. Kaushal, G.P.; Shah, S.V. The new kids on the block: ADAMTSs, potentially multifunctional metalloproteinases of the ADAM family. *J. Clin. Investig.* **2000**, *105*, 1335–1337. [CrossRef] [PubMed]
24. Little, C.B.; Meeker, C.T.; Golub, S.B.; Lawlor, K.E.; Farmer, P.J.; Smith, S.M.; Fosang, A.J. Blocking aggrecanase cleavage in the aggrecan interglobular domain abrogates cartilage erosion and promotes cartilage repair. *J. Clin. Investig.* **2008**, *118*, 3812. [CrossRef]
25. Larkin, J.; Lohr, T.A.; Elefante, L.; Shearin, J.; Matico, R.; Su, J.-L.; Xue, Y.; Liu, F.; Genell, C.; Miller, R.E.; et al. Translational development of an ADAMTS-5 antibody for osteoarthritis disease modification. *Osteoarthr. Cartil.* **2015**, *23*, 1254–1266. [CrossRef]
26. Chiusaroli, R.; Visentini, M.; Galimberti, C.; Casseler, C.; Mennuni, L.; Covaceuszach, S.; Lanza, M.; Ugolini, G.; Caselli, G.; Rovati, L.; et al. Targeting of ADAMTS5’s ancillary domain with the recombinant mAb CRB0017 ameliorates disease progression in a spontaneous murine model of osteoarthritis. *Osteoarthr. Cartil.* **2013**, *21*, 1807–1810. [CrossRef]
27. Huang, Z.; Ding, C.; Li, T.; Yu, S. Current status and future prospects for disease modification in osteoarthritis. *Rheumatology* **2017**, *57* (Suppl. S4), iv108–iv123. [CrossRef]

28. Chockalingam, P.; Sun, W.; Rivera-Bermudez, M.; Zeng, W.; Dufield, D.; Larsson, S.; Lohmander, L.; Flannery, C.; Glasson, S.; Georgiadis, K.; et al. Elevated aggrecanase activity in a rat model of joint injury is attenuated by an aggrecanase specific inhibitor. *Osteoarthr. Cartil.* **2011**, *19*, 315–323. [[CrossRef](#)]
29. Chen, P.; Zhu, S.; Wang, Y.; Mu, Q.; Wu, Y.; Xia, Q.; Zhang, X.; Sun, H.; Tao, J.; Hu, H.; et al. The amelioration of cartilage degeneration by ADAMTS-5 inhibitor delivered in a hyaluronic acid hydrogel. *Biomaterials* **2014**, *35*, 2827–2836. [[CrossRef](#)] [[PubMed](#)]
30. Clement-Lacroix, P.; Little, C.; Meurisse, S.; Blanqué, R.; Mollat, P.; Brebion, F.; Gosmini, R.; De Ceuninck, F.; Botez, I.; Lepescheux, L.; et al. GLPG1972: A potent, selective, orally available adams-5 inhibitor for the treatment of OA. *Osteoarthr. Cartil.* **2017**, *25*, S58–S59. [[CrossRef](#)]
31. Alcaraz, M.J.; Guillén, M.I.; Ferrándiz, M.L. Emerging therapeutic agents in osteoarthritis. *Biochem. Pharmacol.* **2019**, *165*, 4–16. [[CrossRef](#)]
32. Wang, Y.; Fan, X.; Xing, L.; Tian, F. Wnt signaling: A promising target for osteoarthritis therapy. *Cell Commun. Signal.* **2019**, *17*, 1–14. [[CrossRef](#)]
33. Liu, S.; Yang, H.; Shuan, L.; Zhang, M. Sirt1 regulates apoptosis and extracellular matrix degradation in resveratrol-treated osteoarthritis chondrocytes via the Wnt/ $\beta$ -catenin signaling pathways. *Exp. Ther. Med.* **2017**, *14*, 5057–5062. [[CrossRef](#)]
34. Zhou, Y.; Wang, T.; Hamilton, J.L.; Chen, D. Wnt/ $\beta$ -catenin Signaling in Osteoarthritis and in Other Forms of Arthritis. *Curr. Rheumatol. Rep.* **2017**, *19*, 1–8. [[CrossRef](#)] [[PubMed](#)]
35. Lietman, C.; Wu, B.; Lechner, S.; Shinar, A.; Sehgal, M.; Rossomacha, E.; Datta, P.; Sharma, A.; Gandhi, R.; Kapoor, M.; et al. Inhibition of Wnt/ $\beta$ -catenin signaling ameliorates osteoarthritis in a murine model of experimental osteoarthritis. *JCI Insight* **2018**, *3*. [[CrossRef](#)]
36. Deshmukh, V.; Hu, H.; Barroga, C.; Bossard, C.; Kc, S.; Dellamary, L.; Stewart, J.; Chiu, K.; Ibanez, M.; Pedraza, M.; et al. A small-molecule inhibitor of the Wnt pathway (SM04690) as a potential disease modifying agent for the treatment of osteoarthritis of the knee. *Osteoarthr. Cartil.* **2018**, *26*, 18–27. [[CrossRef](#)]
37. Seo, T.; Deshmukh, V.; Yazici, Y. AB0069 LORECEVIVINT (SM04690), an intra-articular, small-molecule CLK/DYRK1A inhibitor that modulates the WNT pathway, as a potential treatment for meniscal injuries. *Ann. Rheum. Dis.* **2020**, *79* (Suppl. S1), 1335. [[CrossRef](#)]
38. Takada, K.; Zhu, D.; Bird, G.; Sukhdeo, K.; Zhao, J.; Mani, M.; Lemieux, M.; Carrasco, D.; Ryan, J.; Horst, D.; et al. Targeted disruption of the BCL9/-catenin complex inhibits oncogenic WNT signaling. *Sci. Transl. Med.* **2012**, *4*, 148ra117. [[CrossRef](#)]
39. Shin, Y.; Huh, Y.H.; Kim, K.; Kim, S.; Park, K.H.; Koh, J.-T.; Chun, J.-S.; Ryu, J.-H. Low-density lipoprotein receptor-related protein 5 governs Wnt-mediated osteoarthritic cartilage destruction. *Arthritis Res. Ther.* **2014**, *16*, R37. [[CrossRef](#)] [[PubMed](#)]
40. Gao, S.-G.; Zeng, C.; Liu, J.-J.; Tian, J.; Cheng, C.; Zhang, F.-J.; Xiong, Y.-L.; Pan, D.; Xiao, Y.-B.; Lei, G.-H. Association between Wnt inhibitory factor-1 expression levels in articular cartilage and the disease severity of patients with osteoarthritis of the knee. *Exp. Ther. Med.* **2016**, *11*, 1405–1409. [[CrossRef](#)] [[PubMed](#)]
41. Deshmukh, V.; O'Green, A.; Bossard, C.; Seo, T.; Lamangan, L.; Ibanez, M.; Ghias, A.; Lai, C.; Do, L.; Cho, S.; et al. Modulation of the Wnt pathway through inhibition of CLK2 and DYRK1A by lorecevivint as a novel, potentially disease-modifying approach for knee osteoarthritis treatment. *Osteoarthr. Cartil.* **2019**, *27*, 1347–1360. [[CrossRef](#)]
42. Yazici, Y.; McAlindon, T.; Gibofsky, A.; Lane, N.; Swearingen, C.; Tambiah, J. Radiographic outcomes were concordant with outcome measures in rheumatology-osteoarthritis research society international (OMERACT-OARSI) strict response: Post-hoc analysis from a phase 2 study of a WNT pathway inhibitor, SM04690, for knee osteoarthritis treatment. *Osteoarthr. Cartil.* **2018**, *26*, S244–S245. [[CrossRef](#)]
43. Ghouri, A.; Conaghan, P.G. Update on novel pharmacological therapies for osteoarthritis. *Ther. Adv. Musculoskelet. Dis.* **2019**, *11*, 1759720–1986449. [[CrossRef](#)] [[PubMed](#)]
44. Ghouri, A.; Conaghan, P.G. Prospects for Therapies in Osteoarthritis. *Calcif. Tissue Int.* **2020**, 1–12. [[CrossRef](#)] [[PubMed](#)]
45. Manicourt, D.; Beaulieu, A.; Garnero, P.; Peterfy, C.; Bouisset, F.; Haemmerle, S.; Mindeholm, L. 229 Effect of treatment with the Cathepsin-K inhibitor, balicatib, on cartilage volume and biochemical markers of bone and cartilage degradation in patients with painful knee osteoarthritis. *Osteoarthr. Cartil.* **2007**, *15*, C130. [[CrossRef](#)]
46. Lindström, E.; Rzoska, B.; Henderson, I.; Terelius, Y.; Jerling, M.; Edenius, C.; Grabowska, U. Nonclinical and clinical pharmacological characterization of the potent and selective cathepsin K inhibitor MIV-711. *J. Transl. Med.* **2018**, *16*, 1–14. [[CrossRef](#)] [[PubMed](#)]
47. Conaghan, P.G.; Bowes, M.A.; Kingsbury, S.R.; Brett, A.; Guillard, G.; Tunblad, K.; Rzoska, B.; Larsson, T.; Holmgren, Å.; Manninen, A.; et al. Six months' treatment with MIV-711, a novel Cathepsin K inhibitor induces osteoarthritis structure modification: Results from a randomized double-blind placebo-controlled phase IIA trial. *Osteoarthr. Cartil.* **2018**, *26*, S25–S26. [[CrossRef](#)]
48. Hunter, D.; Pike, M.; Jonas, B.; Kissin, E.; Krop, J.; McAlindon, T. Phase 1 safety and tolerability study of BMP-7 in symptomatic knee osteoarthritis. *BMC Musculoskelet. Disord.* **2010**, *11*, 232. [[CrossRef](#)]
49. Reker, D.; Kjelgaard-Petersen, C.F.; Siebuhr, A.S.; Michaelis, M.; Gigout, A.; Karsdal, M.A.; Ladel, C.; Bay-Jensen, A.C. Sprifermin (rhFGF18) modulates extracellular matrix turnover in cartilage explants ex vivo. *J. Transl. Med.* **2017**, *15*, 1–12. [[CrossRef](#)]

50. Lohmander, L.S.; Hellot, S.; Dreher, D.; Krantz, E.F.W.; Kruger, D.S.; Guermazi, A.; Eckstein, F. Intraarticular Sprifermin (Recombinant Human Fibroblast Growth Factor 18) in Knee Osteoarthritis: A Randomized, Double-Blind, Placebo-Controlled Trial. *Arthritis Rheumatol.* **2014**, *66*, 1820–1831. [[CrossRef](#)]
51. Roemer, F.W.; Aydemir, A.; Lohmander, L.S.; Crema, M.D.; Marra, M.D.; Muurahainen, N.; Felson, D.T.; Eckstein, F.; Guermazi, A. Structural effects of sprifermin in knee osteoarthritis: A post-hoc analysis on cartilage and non-cartilaginous tissue alterations in a randomized controlled trial. *BMC Musculoskelet. Disord.* **2016**, *17*, 1–7. [[CrossRef](#)]
52. Gigout, A.; Guehring, H.; Froemel, D.; Meurer, A.; Ladel, C.; Reker, D.; Bay-Jensen, A.; Karsdal, M.; Lindemann, S. Sprifermin (rhFGF18) enables proliferation of chondrocytes producing a hyaline cartilage matrix. *Osteoarthr. Cartil.* **2017**, *25*, 1858–1867. [[CrossRef](#)]
53. Dahlberg, L.; Flechsenhar, K.; Dreher, D.; Gouéteux, S.; Jurvelin, J. A randomized, double-blind, placebo-controlled, multicenter study of Rhfgf18 administered intraarticularly using single or multiple ascending doses in patients with primary knee osteoarthritis (OA), scheduled for total knee replacement. *Osteoarthr. Cartil.* **2011**, *19*, S143. [[CrossRef](#)]
54. Hochberg, M.C.; Guermazi, A.; Guehring, H.; Aydemir, A.; Wax, S.; Fleuranceau-Morel, P.; Bihlet, A.R.; Byrjalsen, I.; Andersen, J.R.; Eckstein, F. Effect of intra-articular sprifermin vs placebo on femorotibial joint cartilage thickness in patients with Osteoarthritis. *JAMA* **2019**, *322*, 1360–1370. [[CrossRef](#)] [[PubMed](#)]
55. Pers, Y.-M.; Rackwitz, L.; Ferreira, R.; Pullig, O.; Delfour, C.; Barry, F.; Sensebe, L.; Casteilla, L.; Fleury, S.; Bourin, P.; et al. Adipose Mesenchymal Stromal Cell-Based Therapy for Severe Osteoarthritis of the Knee: A Phase I Dose-Escalation Trial. *STEM CELLS Transl. Med.* **2016**, *5*, 847–856. [[CrossRef](#)]
56. Muchedzi, T.A.; Roberts, S.B. A systematic review of the effects of platelet rich plasma on outcomes for patients with knee osteoarthritis and following total knee arthroplasty. *Surgeon* **2018**, *16*, 250–258. [[CrossRef](#)] [[PubMed](#)]
57. Bennell, K.L.; Hunter, D.J.; Paterson, K.L. Platelet-Rich Plasma for the Management of Hip and Knee Osteoarthritis. *Curr. Rheumatol. Rep.* **2017**, *19*, 24. [[CrossRef](#)]
58. Huebner, K.; Frank, R.M.; Getgood, A. Ortho-Biologics for Osteoarthritis. *Clin. Sports Med.* **2019**, *38*, 123–141. [[CrossRef](#)] [[PubMed](#)]
59. Davies, P.S.E.; Graham, S.M.; Macfarlane, R.J.; Leonidou, A.; Mantalaris, A.; Tsiridis, E. Disease-modifying osteoarthritis drugs: in vitro and in vivo data on the development of DMOADs under investigation. *Expert Opin. Investig. Drugs* **2013**, *22*, 423–441. [[CrossRef](#)] [[PubMed](#)]
60. Toh, W.S.; Lai, R.C.; Hui, J.H.P.; Lim, S.K. MSC exosome as a cell-free MSC therapy for cartilage regeneration: Implications for osteoarthritis treatment. *Semin. Cell Dev. Biol.* **2017**, *67*, 56–64. [[CrossRef](#)]
61. Kim, Y.; Kwon, O.; Choi, Y.; Suh, D.; Heo, D.; Koh, Y. Comparative matched-pair analysis of the injection versus implantation of mesenchymal stem cells for knee osteoarthritis. *Am. J. Sports Med.* **2015**, *43*, 2738–2746. [[CrossRef](#)]
62. Jevotovsky, D.; Alfonso, A.; Einhorn, T.; Chiu, E. Osteoarthritis and stem cell therapy in humans: A systematic review. *Osteoarthr. Cartil.* **2018**, *26*, 711–729. [[CrossRef](#)] [[PubMed](#)]
63. Vega, A.; Martín-Ferrero, M.A.; Del Canto, F.; Alberca, M.; García, V.; Munar, A.; Orozco, L.; Soler, R.; Fuertes, J.J.; Huguet, M.; et al. Treatment of knee osteoarthritis with allogeneic bone marrow mesenchymal stem cells. *Transplantation* **2015**, *99*, 1681–1690. [[CrossRef](#)] [[PubMed](#)]
64. Lee, S.; Lee, S.H.; Na, H.S.; Kwon, J.Y.; Kim, G.; Jung, K.; Cho, K.; Kim, S.A.; Go, E.J.; Park, M.; et al. The therapeutic effect of STAT3 signaling-suppressed MSC on pain and articular cartilage damage in a rat model of monosodium iodoacetate-induced osteoarthritis. *Front. Immunol.* **2018**, *9*, 2881. [[CrossRef](#)]
65. Madry, H.; Cucchiari, M. Advances and challenges in gene-based approaches for osteoarthritis. *J. Gene Med.* **2013**, *15*, 343–355. [[CrossRef](#)] [[PubMed](#)]
66. Wu, Y.; Lu, X.; Shen, B.; Zeng, Y. The Therapeutic Potential and Role of miRNA, lncRNA, and circRNA in Osteoarthritis. *Curr. Gene Ther.* **2019**, *19*, 255–263. [[CrossRef](#)] [[PubMed](#)]
67. Zhang, W.; Robertson, W.B.; Zhao, J.; Chen, W.; Xu, J. Emerging Trend in the Pharmacotherapy of Osteoarthritis. *Front. Endocrinol.* **2019**, *10*, 431. [[CrossRef](#)] [[PubMed](#)]
68. Neogi, T. Clinical significance of bone changes in osteoarthritis. *Ther. Adv. Musculoskelet. Dis.* **2012**, *4*, 259–267. [[CrossRef](#)]
69. Hügler, T.; Geurts, J. What drives osteoarthritis? synovial versus subchondral bone pathology. *Rheumatology* **2017**, *56*, 1461–1471.
70. Burr, D.B. Anatomy and physiology of the mineralized tissues: Role in the pathogenesis of osteoarthrosis. *Osteoarthr. Cartil.* **2004**, *12*, 20–30. [[CrossRef](#)]
71. Day, J.S.; Ding, M.; Van Der Linden, J.C.; Hvid, I.; Sumner, D.R.; Weinans, H. A decreased subchondral trabecular bone tissue elastic modulus is associated with pre-arthritis cartilage damage. *J. Orthop. Res.* **2001**, *19*, 914–918. [[CrossRef](#)]
72. Ni, G.-X.; Zhan, L.-Q.; Gao, M.-Q.; Lei, L.; Zhou, Y.-Z.; Pan, Y.-X. Matrix metalloproteinase-3 inhibitor retards treadmill running-induced cartilage degradation in rats. *Arthritis Res. Ther.* **2011**, *13*, R192. [[CrossRef](#)]
73. Sanchez, C.; DeBerg, M.; Piccardi, N.; Msika, P.; Reginster, J.-Y.; Henrotin, Y. Osteoblasts from the sclerotic subchondral bone downregulate aggrecan but upregulate metalloproteinases expression by chondrocytes. This effect is mimicked by interleukin-6, -1 $\beta$  and oncostatin M pre-treated non-sclerotic osteoblasts. *Osteoarthr. Cartil.* **2005**, *13*, 979–987. [[CrossRef](#)] [[PubMed](#)]
74. Prasad, I.; Crawford, R.; Xiao, Y. Aggravation of ADAMTS and Matrix Metalloproteinase Production and Role of ERK1/2 Pathway in the Interaction of Osteoarthritic Subchondral Bone Osteoblasts and Articular Cartilage Chondrocytes—Possible Pathogenic Role in Osteoarthritis. *J. Rheumatol.* **2012**, *39*, 621–634. [[CrossRef](#)] [[PubMed](#)]

75. Bianco, D.; Todorov, A.; Čengić, T.; Pagenstert, G.; Schären, S.; Netzer, C.; Hügler, T.; Geurts, J. Alterations of Subchondral Bone Progenitor Cells in Human Knee and Hip Osteoarthritis Lead to a Bone Sclerosis Phenotype. *Int. J. Mol. Sci.* **2018**, *19*, 475. [[CrossRef](#)]
76. Zhang, R.-K.; Li, G.-W.; Zeng, C.; Lin, C.-X.; Huang, L.-S.; Huang, G.-X.; Zhao, C.; Feng, S.-Y.; Fang, H. Mechanical stress contributes to osteoarthritis development through the activation of transforming growth factor beta 1 (TGF- $\beta$ 1). *Bone Jt. Res.* **2018**, *7*, 587–594. [[CrossRef](#)] [[PubMed](#)]
77. Jung, Y.-K.; Han, M.-S.; Park, H.-R.; Lee, E.-J.; Jang, J.-A.; Kim, G.-W.; Lee, S.-Y.; Moon, D.; Han, S. Calcium-phosphate complex increased during subchondral bone remodeling affects early stage osteoarthritis. *Sci. Rep.* **2018**, *8*, 487. [[CrossRef](#)]
78. Loeser, R.F.; Shanker, G. Autocrine stimulation by insulin-like growth factor 1 and insulin-like growth factor 2 mediates chondrocyte survival in vitro. *Arthritis Rheum.* **2000**, *43*, 1552–1559. [[CrossRef](#)]
79. Kennedy, O.D.; Laudier, D.M.; Majeska, R.J.; Sun, H.B.; Schaffler, M.B. Osteocyte apoptosis is required for production of osteoclastogenic signals following bone fatigue in vivo. *Bone* **2014**, *64*, 132–137. [[CrossRef](#)] [[PubMed](#)]
80. Li, J.; Xue, J.; Jing, Y.; Wang, M.; Shu, R.; Xu, H.; Xue, C.; Feng, J.; Wang, P.; Bai, D. SOST Deficiency Aggravates Osteoarthritis in Mice by Promoting Sclerosis of Subchondral Bone. *BioMed Res. Int.* **2019**, *2019*, 1–8. [[CrossRef](#)]
81. Ganesh, T.; Laughrey, L.E.; Niroobakhsh, M.; Lara-Castillo, N. Multiscale finite element modeling of mechanical strains and fluid flow in osteocyte lacunocanalicular system. *Bone* **2020**, *137*, 115328. [[CrossRef](#)]
82. Dai, G.; Xiao, H.; Liao, J.; Zhou, N.; Zhao, C.; Xu, W.; Xu, W.; Liang, X.; Huang, W. Osteocyte TGF $\beta$ 1–Smad2/3 is positively associated with bone turnover parameters in sub-chondral bone of advanced osteoarthritis. *Int. J. Mol. Med.* **2020**, *46*, 167–178.
83. Zhou, X.; Cao, H.; Yuan, Y.; Wu, W. Biochemical Signals Mediate the Crosstalk between Cartilage and Bone in Osteoarthritis. *BioMed Res. Int.* **2020**, *2020*, 5720360. [[CrossRef](#)]
84. Hu, S.; Mao, G.; Zhang, Z.; Wu, P.; Wen, X.; Liao, W.; Zhang, Z. MicroRNA-320c inhibits development of osteoarthritis through downregulation of canonical Wnt signaling pathway. *Life Sci.* **2019**, *228*, 242–250. [[CrossRef](#)] [[PubMed](#)]
85. De Santis, M.; Di Matteo, B.; Chisari, E.; Cincinelli, G.; Angele, P.; Lattermann, C.; Filardo, G.; Vitale, N.D.; Selmi, C.; Kon, E. The Role of Wnt Pathway in the Pathogenesis of OA and Its Potential Therapeutic Implications in the Field of Regenerative Medicine. *BioMed Res. Int.* **2018**, *2018*, 1–8. [[CrossRef](#)]
86. Zhao, W.; Wang, T.; Luo, Q.; Chen, Y.; Leung, V.Y.L.; Wen, C.; Shah, M.F.; Pan, H.; Chiu, K.; Cao, X.; et al. Cartilage degeneration and excessive subchondral bone formation in spontaneous osteoarthritis involves altered TGF- $\beta$  signaling. *J. Orthop. Res.* **2015**, *34*, 763–770. [[CrossRef](#)]
87. Zhang, L.; Wang, P.-E.; Ying, J.; Jin, X.; Luo, C.; Xu, T.; Xu, S.; Dong, R.; Xiao, L.; Tong, P.; et al. Yougui Pills Attenuate Cartilage Degeneration via Activation of TGF- $\beta$ /Smad Signaling in Chondrocyte of Osteoarthritic Mouse Model. *Front. Pharmacol.* **2017**, *8*, 611. [[CrossRef](#)]
88. Upton, A.R.; Holding, C.A.; Dharmapatni, A.A.S.S.K.; Haynes, D.R. The expression of RANKL and OPG in the various grades of osteoarthritic cartilage. *Rheumatol. Int.* **2012**, *32*, 535–540. [[CrossRef](#)]
89. Zhong, M.; Carney, D.H.; Jo, H.; Boyan, B.D.; Schwartz, Z. Inorganic Phosphate Induces Mammalian Growth Plate Chondrocyte Apoptosis in a Mitochondrial Pathway Involving Nitric Oxide and JNK MAP Kinase. *Calcif. Tissue Int.* **2011**, *88*, 96–108. [[CrossRef](#)]
90. Nishii, T.; Tamura, S.; Shiomi, T.; Yoshikawa, H.; Sugano, N. Alendronate treatment for hip osteoarthritis: Prospective randomized 2-year trial. *Clin. Rheumatol.* **2013**, *32*, 1759–1766. [[CrossRef](#)]
91. Aitken, D.; Laslett, L.L.; Cai, G.; Hill, C.; March, L.; Wluka, A.E.; Wang, Y.; Blizzard, L.; Cicuttini, F.; Jones, G. A protocol for a multicentre, randomised, double-blind, placebo-controlled trial to compare the effect of annual infusions of zoledronic acid to placebo on knee structural change and knee pain over 24 months in knee osteoarthritis patients—ZAP2. *BMC Musculoskelet. Disord.* **2018**, *19*, 217. [[CrossRef](#)] [[PubMed](#)]
92. Zhu, S.; Chen, K.; Lan, Y.; Zhang, N.; Jiang, R.; Hu, J. Alendronate protects against articular cartilage erosion by inhibiting subchondral bone loss in ovariectomized rats. *Bone* **2013**, *53*, 340–349. [[CrossRef](#)] [[PubMed](#)]
93. Bingham, C.O., III; Buckland-Wright, J.C.; Garnero, P.; Cohen, S.B.; Dougados, M.; Adami, S.; Clauw, D.J.; Spector, T.D.; Pelletier, J.-P.; Raynauld, J.-P.; et al. Risedronate decreases biochemical markers of cartilage degradation but does not decrease symptoms or slow radiographic progression in patients with medial compartment osteoarthritis of the knee: Results of the two-year multinational knee osteoarthritis structural arthritis study. *Arthritis Rheum.* **2006**, *54*, 3494–3507. [[CrossRef](#)] [[PubMed](#)]
94. Spector, T.D.; Conaghan, P.G.; Buckland-Wright, J.C.; Garnero, P.; Cline, G.A.; Beary, J.F.; Valent, D.J.; Meyer, J.M. Effect of risedronate on joint structure and symptoms of knee osteoarthritis: Results of the BRISK randomized, controlled trial [ISRCTN01928173]. *Arthritis Res. Ther.* **2005**, *7*, R625–R633. [[CrossRef](#)] [[PubMed](#)]
95. Laslett, L.L.; Doré, D.A.; Quinn, S.J.; Boon, P.; Ryan, E.; Winzenberg, T.M.; Jones, G. Zoledronic acid reduces knee pain and bone marrow lesions over 1 year: A randomised controlled trial. *Ann. Rheum. Dis.* **2012**, *71*, 1322–1328. [[CrossRef](#)] [[PubMed](#)]
96. Rossini, M.; Adami, S.; Fracassi, E.; Viapiana, O.; Orsolini, G.; Povino, M.R.; Idolazzi, L.; Gatti, D. Effects of intra-articular clodronate in the treatment of knee osteoarthritis: Results of a double-blind, randomized placebo-controlled trial. *Rheumatol. Int.* **2015**, *35*, 255–263. [[CrossRef](#)]
97. Reginster, J.-Y.; Badurski, J.; Bellamy, N.; Bensen, W.; Chapurlat, R.; Chevalier, X.; Christiansen, C.; Genant, H.; Navarro, F.; Nasonov, E.; et al. Efficacy and safety of strontium ranelate in the treatment of knee osteoarthritis: Results of a double-blind, randomised placebo-controlled trial. *Ann. Rheum. Dis.* **2013**, *72*, 179–186. [[CrossRef](#)] [[PubMed](#)]

98. Yu, D.-G.; Ding, H.-F.; Mao, Y.-Q.; Liu, M.; Yu, B.; Zhao, X.; Wang, X.-Q.; Li, Y.; Liu, G.-W.; Nie, S.-B.; et al. Strontium ranelate reduces cartilage degeneration and subchondral bone remodeling in rat osteoarthritis model. *Acta Pharmacol. Sin.* **2013**, *34*, 393–402. [CrossRef] [PubMed]
99. Pelletier, J.-P.; Roubille, C.; Raynauld, J.-P.; Abram, F.; Dorais, M.; Delorme, P.; Martel-Pelletier, J. Disease-modifying effect of strontium ranelate in a subset of patients from the Phase III knee osteoarthritis study SEKIOA using quantitative MRI: Reduction in bone marrow lesions protects against cartilage loss. *Ann. Rheum. Dis.* **2015**, *74*, 422–429. [CrossRef]
100. Granholm, S.; Lundberg, P.; Lerner, U.H. Expression of the calcitonin receptor, calcitonin receptor-like receptor, and receptor activity modifying proteins during osteoclast differentiation. *J. Cell. Biochem.* **2008**, *104*, 920–933. [CrossRef]
101. Nielsen, R.; Bay-Jensen, A.-C.; Byrjalsen, I.; Karsdal, M. Oral salmon calcitonin reduces cartilage and bone pathology in an osteoarthritis rat model with increased subchondral bone turnover. *Osteoarthr. Cartil.* **2011**, *19*, 466–473. [CrossRef]
102. Karsdal, M.; Byrjalsen, I.; Alexandersen, P.; Bihlet, A.; Andersen, J.; Riis, B.; Bay-Jensen, A.; Christiansen, C. Treatment of symptomatic knee osteoarthritis with oral salmon calcitonin: Results from two phase 3 trials. *Osteoarthr. Cartil.* **2015**, *23*, 532–543. [CrossRef]
103. EMA. European Medicines Agency Recommends Limiting Long-Term Use of Calcitonin Medicines 2012. Available online: [https://www.ema.europa.eu/en/documents/press-release/european-medicines-agency-recommends-limiting-long-term-use-calcitonin-medicines\\_en.pdf](https://www.ema.europa.eu/en/documents/press-release/european-medicines-agency-recommends-limiting-long-term-use-calcitonin-medicines_en.pdf) (accessed on 3 January 2021).
104. McDougall, J.J.; Schuelert, N.; Bowyer, J. Cathepsin K inhibition reduces CTXII levels and joint pain in the guinea pig model of spontaneous osteoarthritis. *Osteoarthr. Cartil.* **2010**, *18*, 1355–1357. [CrossRef] [PubMed]
105. Hayami, T.; Zhuo, Y.; Wesolowski, G.A.; Pickarski, M.; Duong, L.T. Inhibition of cathepsin K reduces cartilage degeneration in the anterior cruciate ligament transection rabbit and murine models of osteoarthritis. *Bone* **2012**, *50*, 1250–1259. [CrossRef]
106. Lindström, E.; Rzoska, B.; Tunblad, K.; Edenius, C.; Bendele, A.M.; Maul, D.; Larson, M.; Shah, N.; Otto, V.Y.; Jerome, C.; et al. The selective cathepsin K inhibitor MIV-711 attenuates joint pathology in experimental animal models of osteoarthritis. *J. Transl. Med.* **2018**, *16*, 56. [CrossRef] [PubMed]
107. Karsdal, M.; Michaelis, M.; Ladel, C.; Siebuhr, A.; Bihlet, A.; Andersen, J.; Guehring, H.; Christiansen, C.; Bay-Jensen, A.; Kraus, V. Disease-modifying treatments for osteoarthritis (DMOADs) of the knee and hip: Lessons learned from failures and opportunities for the future. *Osteoarthr. Cartil.* **2016**, *24*, 2013–2021. [CrossRef] [PubMed]
108. Vortkamp, A.; Lee, K.; Lanske, B.; Segre, G.V.; Kronenberg, H.M.; Tabin, C.J. Regulation of Rate of Cartilage Differentiation by Indian Hedgehog and PTH-Related Protein. *Science* **1996**, *273*, 613–622. [CrossRef]
109. Sampson, E.R.; Hilton, M.J.; Tian, Y.; Chen, D.; Schwarz, E.M.; Mooney, R.A.; Bukata, S.V.; O’Keefe, R.J.; Awad, H.; Puzas, J.E.; et al. Teriparatide as a Chondroregenerative Therapy for Injury-Induced Osteoarthritis. *Sci. Transl. Med.* **2011**, *3*, 101ra93. [CrossRef] [PubMed]
110. Bellido, M.; Lugo, L.; Roman-Blas, J.A.; Castañeda, S.; Calvo, E.; Largo, R.; Herrero-Beaumont, G. Improving subchondral bone integrity reduces progression of cartilage damage in experimental osteoarthritis preceded by osteoporosis. *Osteoarthr. Cartil.* **2011**, *19*, 1228–1236. [CrossRef]
111. Wu, M.; Chen, G.; Li, Y.-P. TGF- $\beta$  and BMP signaling in osteoblast, skeletal development, and bone formation, homeostasis and disease. *Bone Res.* **2016**, *4*, 16009. [CrossRef]
112. Shen, J.; Li, S.; Chen, D. TGF- $\beta$  signaling and the development of osteoarthritis. *Bone Res.* **2014**, *2*, 14002. [CrossRef]
113. Xie, L.; Tintani, F.; Wang, X.; Li, F.; Zhen, G.; Qiu, T.; Wan, M.; Crane, J.; Chen, Q.; Cao, X. Systemic neutralization of TGF- $\beta$  attenuates osteoarthritis. *Ann. N. Y. Acad. Sci.* **2016**, *1376*, 53–64. [CrossRef]
114. Zhen, G.; Wen, C.; Jia, X.; Li, Y.; Crane, J.L.; Mears, S.C.; Askin, F.B.; Frassica, F.J.; Chang, W.; Yao, J.; et al. Inhibition of TGF- $\beta$  signaling in mesenchymal stem cells of subchondral bone attenuates osteoarthritis. *Nat. Med.* **2013**, *19*, 704–712. [CrossRef] [PubMed]
115. Rowe, P.; Kumagai, Y.; Gutierrez, G.; Garrett, I.; Blacher, R.; Rosen, D.; Cundy, J.; Navvab, S.; Chen, D.; Drezner, M.; et al. MEPE has the properties of an osteoblastic phosphatonin and minihibin. *Bone* **2004**, *34*, 303–319. [CrossRef] [PubMed]
116. McGuire, D.; Lane, N.; Segal, N.; Metyas, S.; Barthel, H.; Miller, M.; Rosen, D.; Kumagai, Y. TPX-100 leads to marked, sustained improvements in subjects with knee osteoarthritis: Pre-clinical rationale and results of a controlled clinical trial. *Osteoarthr. Cartil.* **2018**, *26*, S243. [CrossRef]
117. Nagai, T.; Sato, M.; Kutsuna, T.; Kokubo, M.; Ebihara, G.; Ohta, N.; Mochida, J. Intravenous administration of anti-vascular endothelial growth factor humanized monoclonal antibody bevacizumab improves articular cartilage repair. *Arthritis Res. Ther.* **2010**, *12*, R178. [CrossRef] [PubMed]
118. Cui, Z.; Crane, J.; Xie, H.; Jin, X.; Zhen, G.; Li, C.; Xie, L.; Wang, L.; Bian, Q.; Qiu, T.; et al. Halofuginone attenuates osteoarthritis by inhibition of TGF- $\beta$  activity and H-type vessel formation in subchondral bone. *Ann. Rheum. Dis.* **2015**, *75*, 1714–1721. [CrossRef]
119. Lu, J.; Zhang, H.; Cai, D.; Zeng, C.; Lai, P.; Shao, Y.; Fang, H.; Li, D.; Ouyang, J.; Zhao, C.; et al. Positive-Feedback Regulation of Subchondral H-Type Vessel Formation by Chondrocyte Promotes Osteoarthritis Development in Mice. *J. Bone Miner. Res.* **2018**, *33*, 909–920. [CrossRef] [PubMed]
120. Walsh, D.A.; McWilliams, D.F.; Turley, M.J.; Dixon, M.R.; Fransès, R.E.; Mapp, P.I.; Wilson, D. Angiogenesis and nerve growth factor at the osteochondral junction in rheumatoid arthritis and osteoarthritis. *Rheumatology* **2010**, *49*, 1852–1861. [CrossRef]
121. Xie, H.; Cui, Z.; Wang, L.; Xia, Z.; Hu, Y.; Xian, L.; Li, C.; Xie, L.; Crane, J.; Wan, M.; et al. PDGF-BB secreted by preosteoclasts induces angiogenesis during coupling with osteogenesis. *Nat. Med.* **2014**, *20*, 1270–1278. [CrossRef]

122. Schnitzer, T.J.; Ekman, E.F.; Spierings, E.L.H.; Greenberg, H.S.; Smith, M.D.; Brown, M.T.; West, C.R.; Verburg, K.M. Efficacy and safety of tanezumab monotherapy or combined with non-steroidal anti-inflammatory drugs in the treatment of knee or hip osteoarthritis pain. *Ann. Rheum. Dis.* **2015**, *74*, 1202–1211. [[CrossRef](#)]
123. Hochberg, M.C.; Tive, L.A.; Abramson, S.B.; Vignon, E.; Verburg, K.M.; West, C.R.; Smith, M.D.; Hungerford, D.S. When Is Osteonecrosis Not Osteonecrosis?: Adjudication of Reported Serious Adverse Joint Events in the Tanezumab Clinical Development Program. *Arthritis Rheumatol.* **2016**, *68*, 382–391. [[CrossRef](#)]
124. Clinicaltrials.gov. Study to Determine the Safety and the Efficacy of Fasinumab Compared to Placebo and Nonsteroidal Anti-Inflammatory Drugs (NSAIDs) for Treatment of Adults with Pain from Osteoarthritis of the Knee or Hip (FACT OA2). Re-generon Pharmaceuticals. 2018. Available online: <https://clinicaltrials.gov/ct2/show/NCT03304379> (accessed on 5 January 2021).
125. Yang, C.Y.; Chanalaris, A.; Troeberg, L. ADAMTS and ADAM metalloproteinases in osteoarthritis—Looking beyond the ‘usual suspects’. *Osteoarthr. Cartil.* **2017**, *25*, 1000–1009. [[CrossRef](#)]
126. Atukorala, I.; Kwok, C.K.; Guermazi, A.; Roemer, F.W.; Boudreau, R.M.; Hannon, M.J.; Hunter, D.J. Synovitis in knee osteoarthritis: A precursor of disease? *Ann. Rheum. Dis.* **2016**, *75*, 390–395. [[CrossRef](#)]
127. Schnitzer, T.J.; Easton, R.; Pang, S.; Levinson, D.J.; Pixton, G.; Viktrup, L.; Davignon, I.; Brown, M.T.; West, C.R.; Verburg, K.M. Effect of tanezumab on joint pain, physical function, and patient global assessment of osteoarthritis among patients with osteoarthritis of the hip or knee: A randomized clinical trial. *JAMA* **2019**, *322*, 37–48. [[CrossRef](#)]
128. Dakin, P.; DiMartino, S.J.; Gao, H.; Maloney, J.; Kivitz, A.J.; Schnitzer, T.J.; Stahl, N.; Yancopoulos, G.D.; Geba, G.P. The Efficacy, Tolerability, and Joint Safety of Fasinumab in Osteoarthritis Pain: A Phase IIb/III Double-Blind, Placebo-Controlled, Randomized Clinical Trial. *Arthritis Rheumatol.* **2019**, *71*, 1824–1834. [[CrossRef](#)]
129. Kim, J.-R.; Yoo, J.J.; Kim, H.A. Therapeutics in Osteoarthritis Based on an Understanding of Its Molecular Pathogenesis. *Int. J. Mol. Sci.* **2018**, *19*, 674. [[CrossRef](#)]
130. Yu, S.P.-C.; Hunter, D.J. Emerging drugs for the treatment of knee osteoarthritis. *Expert Opin. Emerg. Drugs* **2015**, *20*, 361–378. [[CrossRef](#)] [[PubMed](#)]
131. Siebuhr, A.S.; Bay-Jensen, A.C.; Jordan, J.M.; Kjelgaard-Petersen, C.F.; Christiansen, C.; Abramson, S.B.; Attur, M.; Berenbaum, F.; Kraus, V.; Karsdal, M.A. Inflammation (or synovitis)-driven osteoarthritis: An opportunity for personalizing prognosis and treatment? *Scand. J. Rheumatol.* **2015**, *45*, 87–98. [[CrossRef](#)] [[PubMed](#)]
132. Wang, J. Efficacy and safety of adalimumab by intra-articular injection for moderate to severe knee osteoarthritis: An open-label randomized controlled trial. *J. Int. Med Res.* **2017**, *46*, 326–334. [[CrossRef](#)]
133. Ohtori, S.; Orita, S.; Yamauchi, K.; Eguchi, Y.; Ochiai, N.; Kishida, S.; Kuniyoshi, K.; Aoki, Y.; Nakamura, J.; Ishikawa, T.; et al. Efficacy of Direct Injection of Etanercept into Knee Joints for Pain in Moderate and Severe Knee Osteoarthritis. *Yonsei Med. J.* **2015**, *56*, 1379–1383. [[CrossRef](#)]
134. Chevalier, X.; Goupille, P.; Beaulieu, A.D.; Burch, F.X.; Bensen, W.G.; Conrozier, T.; Loeuille, D.; Kivitz, A.J.; Silver, D.; Appleton, B.E. Intraarticular injection of anakinra in osteoarthritis of the knee: A multicenter, randomized, double-blind, placebo-controlled study. *Arthritis Rheum.* **2009**, *61*, 344–352. [[CrossRef](#)] [[PubMed](#)]
135. Wang, S.X.; Abramson, S.B.; Attur, M.; Karsdal, M.A.; Preston, R.A.; Lozada, C.J.; Kosloski, M.P.; Hong, F.; Jiang, P.; Saltarelli, M.J.; et al. Safety, tolerability, and pharmacodynamics of an anti-interleukin-1 $\alpha$ / $\beta$  dual variable domain immuno-globulin in patients with osteoarthritis of the knee: A randomized phase 1 study. *Osteoarthr. Cartil.* **2017**, *25*, 1952–1961. [[CrossRef](#)]
136. Migliore, A.; Paoletta, M.; Moretti, A.; Liguori, S.; Iolascon, G. The perspectives of intra-articular therapy in the management of osteoarthritis. *Expert Opin. Drug Deliv.* **2020**, *17*, 1213–1226. [[CrossRef](#)]
137. Gibofsky, A.; Hochberg, M.C.; Jaros, M.J.; Young, C.L. Efficacy and safety of low-dose submicron diclofenac for the treatment of osteoarthritis pain: A 12 week, phase 3 study. *Curr. Med Res. Opin.* **2014**, *30*, 1883–1893. [[CrossRef](#)] [[PubMed](#)]
138. Philp, A.M.; Davis, E.T.; Jones, S.W. Developing anti-inflammatory therapeutics for patients with osteoarthritis. *Rheumatology* **2016**, *56*, 869–881. [[CrossRef](#)]
139. Gallelli, L.; Galasso, O.; Falcone, D.; Southworth, S.; Greco, M.; Ventura, V.; Romualdi, P.; Corigliano, A.; Terracciano, R.; Savino, R.; et al. The effects of nonsteroidal anti-inflammatory drugs on clinical outcomes, synovial fluid cytokine concentration and signal transduction pathways in knee osteoarthritis. A randomized open label trial. *Osteoarthr. Cartil.* **2013**, *21*, 1400–1408. [[CrossRef](#)] [[PubMed](#)]
140. Lim, C.C.; Ang, A.T.W.; Kadir, H.B.A.; Lee, P.H.; Goh, B.Q.; Harikrishnan, S.; Kwek, J.L.; Gan, S.S.W.; Choo, J.C.J.; Tan, N.C. Short-Course Systemic and Topical Non-Steroidal Anti-Inflammatory Drugs: Impact on Adverse Renal Events in Older Adults with Co-Morbid Disease. *Drugs Aging* **2021**, *38*, 147–156. [[CrossRef](#)]
141. Jovanovic, D.V.; Fernandes, J.C.; Martel-Pelletier, J.; Jolicœur, F.C.; Reboul, P.; Laufer, S.; Tries, S.; Pelletier, J.P. In vivo dual inhibition of cyclooxygenase and lipoxygenase by ML-3000 reduces the progression of experimental osteoarthritis: Suppression of collagenase 1 and interleukin-1 $\beta$  synthesis. *Arthritis Rheum.* **2001**, *44*, 2320–2330. [[CrossRef](#)]
142. Raynauld, J.P.; Martel-Pelletier, J.; Bias, P.; Laufer, S.; Haroui, B.; Choquette, D.; Beaulieu, A.D.; Abram, F.; Dorais, M.; Vignon, E.; et al. Protective effects of licofelone, a 5-lipoxygenase and cy-clo-oxygenase inhibitor, versus naproxen on cartilage loss in knee osteoarthritis: A first multicentre clinical trial using quanti-tative MRI. *Ann. Rheum. Dis.* **2009**, *68*, 938–947. [[CrossRef](#)] [[PubMed](#)]
143. Vaishya, R.; Pandit, R.; Agarwal, A.K.; Vijay, V. Intra-articular hyaluronic acid is superior to steroids in knee osteoarthritis: A comparative, randomized study. *J. Clin. Orthop. Trauma* **2017**, *8*, 85–88. [[CrossRef](#)]

144. He, W.-W.; Kuang, M.-J.; Zhao, J.; Sun, L.; Lu, B.; Wang, Y.; Ma, J.-X.; Ma, X.-L. Efficacy and safety of intraarticular hyaluronic acid and corticosteroid for knee osteoarthritis: A meta-analysis. *Int. J. Surg.* **2017**, *39*, 95–103. [[CrossRef](#)]
145. Mol, M.F.; Runhaar, J.; Bos, P.K.; Dorleijn, D.M.J.; Vis, M.; Gussekloo, J.; Bindels, P.J.E.; Bierma-Zeinstra, S.M.A. Effectiveness of intramuscular gluteal glucocorticoid injection versus intra-articular glucocorticoid injection in knee osteoarthritis: Design of a multicenter randomized, 24 weeks comparative parallel-group trial. *BMC Musculoskelet. Disord.* **2020**, *21*, 1–9. [[CrossRef](#)]
146. Samuels, J.; Pillinger, M.H.; Jevsevar, D.; Felson, D.; Simon, L.S. Critical appraisal of intra-articular glucocorticoid injections for symptomatic osteoarthritis of the knee. *Osteoarthr. Cartil.* **2021**, *29*, 8–16. [[CrossRef](#)]
147. Hangody, L.; Szody, R.; Lukasik, P.; Zgadzaj, W.; Lénárt, E.; Dokoupilova, E.; Bichovsk, D.; Berta, A.; Vasarhelyi, G.; Ficzer, A.; et al. Intraarticular injection of a cross-linked sodium hyaluronate combined with triamcinolone hexacetonide (Cingal) to provide symptomatic relief of osteoarthritis of the knee: A randomized, double-blind, placebo-controlled multicenter clinical trial. *Cartilage* **2018**, *9*, 276–283. [[CrossRef](#)] [[PubMed](#)]
148. Babaei-Ghazani, A.; Najarzadeh, S.; Mansoori, K.; Forogh, B.; Madani, S.P.; Ebadi, S.; Fadavi, H.R.; Eftekharsadat, B. The effects of ultrasound-guided corticosteroid injection compared to oxygen-ozone (O<sub>2</sub>-O<sub>3</sub>) injection in patients with knee osteo-arthritis: A randomized controlled trial. *Clin. Rheumatol.* **2018**, *37*, 2517–2527. [[CrossRef](#)]
149. Pelletier, J.-P.; Raynaud, J.-P.; Dorais, M.; Bessette, L.; Dokoupilova, E.; Morin, F.; Pavelka, K.; Paiement, P.; Martel-Pelletier, J.; DISSCO Trial Investigator Group. An international, multicentre, double-blind, randomized study (DISSCO): Effect of diacerein vs celecoxib on symptoms in knee osteoarthritis. *Rheumatology* **2020**, *59*, 3858–3868. [[CrossRef](#)]
150. Jung, J.H.; Kim, S.E.; Kim, H.-J.; Park, K.; Song, G.G.; Choi, S.J. A comparative pilot study of oral diacerein and locally treated diacerein-loaded nanoparticles in a model of osteoarthritis. *Int. J. Pharm.* **2020**, *581*, 119249. [[CrossRef](#)] [[PubMed](#)]
151. Leonidou, A.; Lepetsos, P.; Mintzas, M.; Kenanidis, E.; Macheras, G.; Tzetis, M.; Potoupnis, M.; Tsiridis, E. Inducible nitric oxide synthase as a target for osteoarthritis treatment. *Expert Opin. Ther. Targets* **2018**, *22*, 299–318. [[CrossRef](#)]
152. Ahmad, N.; Ansari, M.Y.; Haqqi, T.M. Role of iNOS in osteoarthritis: Pathological and therapeutic aspects. *J. Cell. Physiol.* **2020**, *235*, 6366–6376. [[CrossRef](#)]
153. Le Graverand, M.-P.H.; Clemmer, R.S.; Redifer, P.; Brunell, R.M.; Hayes, C.W.; Brandt, K.D.; Abramson, S.B.; Manning, P.T.; Miller, C.G.; Vignon, E. A 2-year randomised, double-blind, placebo-controlled, multicentre study of oral selective iNOS inhibitor, cindunistat (SD-6010), in patients with symptomatic osteoarthritis of the knee. *Ann. Rheum. Dis.* **2013**, *72*, 187–195. [[CrossRef](#)]
154. Jacob, G.; Shetty, V.; Shetty, S. A study assessing intra-articular PRP vs PRP with HMW HA vs PRP with LMW HA in early knee osteoarthritis. *J. Arthrosc. Jt. Surg.* **2017**, *4*, 65–71. [[CrossRef](#)]
155. Cook, C.S.; Smith, P.A. Clinical Update: Why PRP Should Be Your First Choice for Injection Therapy in Treating Osteoarthritis of the Knee. *Curr. Rev. Musculoskelet. Med.* **2018**, *11*, 583–592. [[CrossRef](#)]
156. Saif, D.S.; Hegazy, N.N.; Zahran, E.S. Evaluating the efficacy of intra-articular injections of platelet rich plasma (PRP) in rheumatoid arthritis patients and its impact on inflammatory cytokines, disease activity and quality of life. *Curr. Rheumatol. Rev.* **2020**, *16*, 1. [[CrossRef](#)] [[PubMed](#)]
157. Filardo, G.; Previtali, D.; Napoli, F.; Candrian, C.; Zaffagnini, S.; Grassi, A. PRP injections for the treatment of knee osteoarthritis: A meta-analysis of randomized controlled trials. *Cartilage* **2020**. [[CrossRef](#)]
158. O’Connell, B.; Wragg, N.M.; Wilson, S.L. The use of PRP injections in the management of knee osteoarthritis. *Cell Tissue Res.* **2019**, *376*, 143–152. [[CrossRef](#)] [[PubMed](#)]
159. Yin, W.; Xu, H.; Sheng, J.; Xu, Z.; Xie, X.; Zhang, C. Comparative evaluation of the effects of platelet rich plasma formulations on extracellular matrix formation and the NF- $\kappa$ B signaling pathway in human articular chondrocytes. *Mol. Med. Rep.* **2017**, *15*, 2940–2948. [[CrossRef](#)] [[PubMed](#)]
160. Simental-Mendía, M.; Vilchez-Cavazos, J.F.; Peña-Martínez, V.M.; Said-Fernández, S.; Lara-Arias, J.; Martínez-Rodríguez, H.G. Leukocyte-poor platelet-rich plasma is more effective than the conventional therapy with acetaminophen for the treatment of early knee osteoarthritis. *Arch. Orthop. Trauma Surg.* **2016**, *136*, 1723–1732. [[CrossRef](#)]
161. Tong, S.; Liu, J.; Zhang, C. Platelet-rich plasma inhibits inflammatory factors and represses rheumatoid fibroblast-like synoviocytes in rheumatoid arthritis. *Clin. Exp. Med.* **2017**, *17*, 441–449. [[CrossRef](#)]
162. Migliorini, F.; Driessen, A.; Quack, V.; Sippel, N.; Cooper, B.; El Mansy, Y.; Tingart, M.; Eschweiler, J. Comparison between intra-articular infiltrations of placebo, steroids, hyaluronic and PRP for knee osteoarthritis: A Bayesian network meta-analysis. *Arch. Orthop. Trauma Surg.* **2020**, 1–18. [[CrossRef](#)]
163. Huang, G.; Hua, S.; Yang, T.; Ma, J.; Yu, W.; Chen, X. Platelet-rich plasma shows beneficial effects for patients with knee osteoarthritis by suppressing inflammatory factors. *Exp. Ther. Med.* **2018**, *15*, 3096–3102. [[CrossRef](#)]
164. Xu, Z.; He, Z.; Shu, L.; Li, X.; Ma, M.; Ye, C. Intra-articular platelet-rich plasma combined with hyaluronic acid injection for knee osteoarthritis is superior to platelet-rich plasma or hyaluronic acid alone in inhibiting inflammation and improving pain and function. *Arthrosc. J. Arthrosc. Relat. Surg.* **2020**. [[CrossRef](#)]
165. Nelson, F.R.; Zvirbulis, R.A.; Zonca, B.; Li, K.W.; Turner, S.M.; Pasierb, M.; Wilton, P.; Martinez-Puig, D.; Wu, W. The effects of an oral preparation containing hyaluronic acid (Oralvisc<sup>®</sup>) on obese knee osteoarthritis patients determined by pain, function, bradykinin, leptin, inflammatory cytokines, and heavy water analyses. *Rheumatol. Int.* **2015**, *35*, 43–52. [[CrossRef](#)]
166. Raeissadat, S.A.; Rayegani, S.M.; Forogh, B.; Abadi, P.H.; Moridnia, M.; Dehghan, S.R. Intra-articular ozone or hyaluronic acid injection: Which one is superior in patients with knee osteoarthritis? A 6-month randomized clinical trial. *J. Pain Res.* **2018**, *11*, 111. [[CrossRef](#)] [[PubMed](#)]

167. Lopes de Jesus, C.C.; Dos Santos, F.C.; de Jesus, L.M.; Monteiro, I.; Sant'Ana, M.S.; Trevisani, V.F. Comparison between in-tra-articular ozone and placebo in the treatment of knee osteoarthritis: A randomized, double-blinded, placebo-controlled study. *PLoS ONE* **2017**, *12*, e0179185. [[CrossRef](#)]
168. Loeser, R.; Beavers, D.; Bay-Jensen, A.; Karsdal, M.; Nicklas, B.; Guermazi, A.; Hunter, D.; Messier, S. Effects of dietary weight loss with and without exercise on interstitial matrix turnover and tissue inflammation biomarkers in adults with knee osteoarthritis: The Intensive Diet and Exercise for Arthritis trial (IDEA). *Osteoarthr. Cartil.* **2017**, *25*, 1822–1828. [[CrossRef](#)]
169. Cheragh-Birjandi, S.; Moghbeli, M.; Haghighi, F.; Safdari, M.R.; Baghernezhad, M.; Akhavan, A.; Ganji, R. Impact of resistance exercises and nano-curcumin on synovial levels of collagenase and nitric oxide in women with knee osteoarthritis. *Transl. Med. Commun.* **2020**, *5*, 1–6. [[CrossRef](#)]
170. Manoy, P.; Yuktanandana, P.; Tanavalee, A.; Anomasiri, W.; Ngarmukos, S.; Tanpowpong, T.; Honsawek, S. Vitamin D Supplementation Improves Quality of Life and Physical Performance in Osteoarthritis Patients. *Nutrients* **2017**, *9*, 799. [[CrossRef](#)]
171. Gao, X.R.; Chen, Y.S.; Deng, W. The effect of vitamin D supplementation on knee osteoarthritis: A meta-analysis of randomized controlled trials. *Int. J. Surg.* **2017**, *46*, 14–20. [[CrossRef](#)]
172. Wang, X.; Cicuttini, F.; Jin, X.; Wluka, A.; Han, W.; Zhu, Z.; Blizzard, L.; Antony, B.; Winzenberg, T.; Jones, G.; et al. Knee effusion-synovitis volume measurement and effects of vitamin D supplementation in patients with knee osteoarthritis. *Osteoarthr. Cartil.* **2017**, *25*, 1304–1312. [[CrossRef](#)] [[PubMed](#)]
173. Thevaranjan, N.; Puchta, A.; Schulz, C.; Naidoo, A.; Szamosi, J.; Verschoor, C.P.; Loukov, D.; Schenck, L.P.; Jury, J.; Foley, K.P.; et al. Age-Associated Microbial Dysbiosis Promotes Intestinal Permeability, Systemic Inflammation, and Macrophage Dysfunction. *Cell Host Microbe* **2017**, *21*, 455–466.e4. [[CrossRef](#)] [[PubMed](#)]
174. Franceschi, C.; Campisi, J. Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases. *J. Gerontol. Ser. A Biol. Sci. Med. Sci.* **2014**, *69* (Suppl. S1), S4–S9. [[CrossRef](#)]
175. Tsai, J.C.; Casteneda, G.; Lee, A.; Dereschuk, K.; Li, W.T.; Chakladar, J.; Lombardi, A.F.; Ongkeko, W.M.; Chang, E.Y. Identification and Characterization of the Intra-Articular Microbiome in the Osteoarthritic Knee. *Int. J. Mol. Sci.* **2020**, *21*, 8618. [[CrossRef](#)]
176. Szychlińska, M.A.; Di Rosa, M.; Castorina, A.; Mobasher, A.; Musumeci, G. A correlation between intestinal microbiota dysbiosis and osteoarthritis. *Heliyon* **2019**, *5*, e01134. [[CrossRef](#)]
177. Silvestre, M.P.; Rodrigues, A.M.; Canhão, H.; Marques, C.; Teixeira, D.; Calhau, C.; Branco, J. Cross-Talk between Diet-Associated Dysbiosis and Hand Osteoarthritis. *Nutrients* **2020**, *12*, 3469. [[CrossRef](#)]
178. Lee, S.H.; Kwon, J.Y.; Jhun, J.; Jung, K.; Park, S.-H.; Yang, C.W.; Cho, Y.; Kim, S.J.; Cho, M.-L. Lactobacillus acidophilus ameliorates pain and cartilage degradation in experimental osteoarthritis. *Immunol. Lett.* **2018**, *203*, 6–14. [[CrossRef](#)]
179. Ahmad, N.; Ansari, M.Y.; Bano, S.; Haqqi, T.M. Imperatorin suppresses IL-1 $\beta$ -induced iNOS expression via inhibiting ERK-MAPK/AP1 signaling in primary human OA chondrocytes. *Int. Immunopharmacol.* **2020**, *85*, 106612. [[CrossRef](#)] [[PubMed](#)]
180. Lu, C.; Li, Y.; Hu, S.; Cai, Y.; Yang, Z.; Peng, K. Scoparone prevents IL-1 $\beta$ -induced inflammatory response in human osteoarthritic chondrocytes through the PI3K/Akt/NF- $\kappa$ B pathway. *Biomed. Pharmacother.* **2018**, *106*, 1169–1174. [[CrossRef](#)]
181. Qiao, Z.; Tang, J.; Wu, W.; Liu, M. Acteoside inhibits inflammatory response via JAK/STAT signaling pathway in osteoarthritic rats. *BMC Complement. Altern. Med.* **2019**, *19*, 1–8. [[CrossRef](#)]
182. Al-Afify, A.S.; El-Akabawy, G.; El-Sherif, N.M.; El-Safty, F.E.-N.A.; El-Habiby, M.M. Avocado soybean unsaponifiables ameliorates cartilage and subchondral bone degeneration in mono-iodoacetate-induced knee osteoarthritis in rats. *Tissue Cell* **2018**, *52*, 108–115. [[CrossRef](#)]
183. Shin, H.J.; Park, H.; Shin, N.; Kwon, H.H.; Yin, Y.; Hwang, J.A.; Kim, S.I.; Kim, S.R.; Kim, S.; Joo, Y.; et al. p47phox siR-NA-Loaded PLGA nanoparticles suppress ROS/oxidative stress-induced chondrocyte damage in osteoarthritis. *Polymers* **2020**, *12*, 443. [[CrossRef](#)] [[PubMed](#)]
184. Hoshi, H.; Akagi, R.; Yamaguchi, S.; Muramatsu, Y.; Akatsu, Y.; Yamamoto, Y.; Sasaki, T.; Takahashi, K.; Sasho, T. Effect of inhibiting MMP13 and ADAMTS5 by intra-articular injection of small interfering RNA in a surgically induced osteoarthritis model of mice. *Cell Tissue Res.* **2017**, *368*, 379–387. [[CrossRef](#)] [[PubMed](#)]
185. Chen, L.X.; Lin, L.; Wang, H.J.; Wei, X.L.; Fu, X.; Zhang, J.Y.; Yu, C.L. Suppression of early experimental osteoarthritis by in vivo delivery of the adenoviral vector-mediated NF-kappaBp65-specific siRNA. *Osteoarthr. Cartil.* **2008**, *16*, 174–184. [[CrossRef](#)] [[PubMed](#)]
186. Kong, Y.; Zhang, Y.; Zhao, X.; Wang, G.; Liu, Q. Carboxymethyl-chitosan attenuates inducible nitric oxide synthase and promotes interleukin-10 production in rat chondrocytes. *Exp. Ther. Med.* **2017**, *14*, 5641–5646. [[CrossRef](#)] [[PubMed](#)]