



Review Clinical Potential of Kinase Inhibitors in Combination with Immune Checkpoint Inhibitors for the Treatment of Solid Tumors

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Abstract: Oncogenic kinases contribute to immunosuppression and modulate the tumor microenvironment in solid tumors. Increasing evidence supports the fundamental role of oncogenic kinase signaling networks in coordinating immunosuppressive tumor microenvironments. This has led to numerous studies examining the efficacy of kinase inhibitors in inducing anti-tumor immune responses by increasing tumor immunogenicity. Kinase inhibitors are the second most common FDA-approved group of drugs that are deployed for cancer treatment. With few exceptions, they inevitably lead to intrinsic and/or acquired resistance, particularly in patients with metastatic disease when used as a monotherapy. On the other hand, cancer immunotherapies, including immune checkpoint inhibitors, have revolutionized cancer treatment for malignancies such as melanoma and lung cancer. However, key hurdles remain to successfully incorporate such therapies in the treatment of other solid cancers. Here, we review the recent literature on oncogenic kinases that regulate tumor immunogenicity, immune suppression, and anti-tumor immunity. Furthermore, we discuss current efforts in clinical trials that combine kinase inhibitors and immune checkpoint inhibitors to treat breast cancer and other solid tumors.

Keywords: kinase signaling; breast cancer; solid tumors; anti-tumor immunity; immunosuppression; kinase inhibitors; cancer immunotherapy

1. Introduction

The host immune system has multiple cellular machineries to eradicate malignant lesions. However, tumors develop multiple mechanisms to escape the host anti-tumor immune response. Immunotherapy, which re-engages immune surveillance pathways, has become one of the pillars of cancer treatment today. However, the ability of tumors to perpetuate an immunosuppressive microenvironment, combined with their ability to avoid being recognized as 'non-self', continues to impede the success of immunotherapy for many solid malignancies. Small-molecule kinase inhibitors represent an opportunity to overcome these key hurdles.

Over the past decades, studies have established that oncogenic kinases are fundamental in driving tumorigenesis and shaping the immune milieu to affect cancer progression and responsiveness to therapy. As of February 2021, at least 53 inhibitors (small-molecule or antibody-based) targeting more than 24 tyrosine/serine/threonine kinases have been approved by the FDA to treat various solid cancers. In line with the tumorigenic role of these kinases, many of these kinase inhibitors elicit anti-tumor immune responses, enhance tumor immunogenicity by regulating antigen processing and presentation and reduce immune suppression, which ultimately improves tumor killing. Many critical kinases are



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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). shared by tumor cells and immune cells, and the genetic or pharmacological inhibition of these kinases affects the function of both cell types. This has important implications in the clinical success of said inhibitors and warrants deeper understanding and attention as the research community explores ways to co-opt host immunity for cancer treatment.

Here, we provide a brief overview of the host immune responses mounted against malignant lesions, tumor immune evasion mechanisms, and challenges associated with immunotherapy and kinase inhibitors. We then review the mechanisms by which tumorintrinsic oncogenic kinases shape the immune microenvironment, with a specific focus on the role of receptor and non-receptor kinases and their immediate molecular effectors. Preclinical studies and clinical trial results demonstrating that genetic and chemical perturbations of kinases can elicit anti-tumor immune responses to eradicate tumors, especially in combination with immunotherapy, are summarized. We spotlight on breast cancer as a model of recent advancements. Based on the evidence provided in this review, we posit that a rational, evidence-based combination of kinase inhibitors and immunotherapy may overcome some of the hurdles faced by both therapeutic modalities and improve the treatment of cancer patients.

2. Cancer Immunosuppression and Anti-Tumor Immunity

The true appreciation of the immune response in suppressing tumor formation came when mice lacking adaptive immunity (RAG2 knock-out) showed an increased tumor incidence upon carcinogen exposure [1]. Numerous immune cell types collaboratively mediate tumor cell killing through multiple mechanisms, including the recognition of tumorassociated or tumor-specific antigens by the adaptive immune system and non-antigendependent killing by the innate immune system. Cytotoxic T lymphocytes (CD8+ CTL) are part of the adaptive immune system and induce tumor cell apoptosis through (1) recognition of antigens presented in the context of MHC class I leading to perforin/granzyme B secretion and (2) recognition of Fas on tumors via FasL, leading to caspase-mediated cell death. CTLs also release cytokines such as TNF α and IFN γ to promote cell cycle arrest in tumors [2-6]. While CTLs mount their attacks via T cell receptor (TCR) mediated recognition of antigen-MHC complexes, natural killer (NK) cells induce similar cytotoxic responses, but in an antigen-independent manner. Instead, the cytotoxic potential of NK cells is controlled by the net balance of stimulatory versus inhibitory receptors on the surface of NK cells themselves, combined with particular classes of ligands expressed by the target cells [7,8]. In oncology, immunogenic cell death (ICD) represents a type of non-microbial death that can be initiated, either due to endoplasmic reticulum stress or in response to cytotoxic treatments, such as anthracyclines or radiotherapy. These various stressors lead to the release of damage-associated molecular patterns (DAMPs), which prime the innate immune system to educate potent anti-tumor immune responses. B cells of the adaptive immune system have also emerged as playing anti-tumorigenic roles through (1) release of tumor antigen-specific antibodies (once B cells differentiate into plasma cells) that trigger antibody-dependent cellular cytotoxicity (ADCC) by NK cells or complement-dependent cytotoxicity (CDC), and (2) B cell receptor-mediated antigen presentation to CD8+ or naïve CD4+ cells for tumor killing [9–11]. During ADCC, membrane-bound antigens on the surface of tumor cells are recognized by specific antibodies. NK cells expressing Fc receptors then bind the Fc portion of these antibodies, leading to NK cell activation and subsequent release of cytotoxic granules that elicit tumor cell lysis [12]. Finally, beyond immune cells with direct tumoricidal properties, infiltrating dendritic cells into tumors are central for activating CTL-driven anti-tumor immunity, as professional antigen-presenting cells that educate both Th and CTL immune responses (reviewed in [13]).

The ability of malignant cells to escape from such anti-tumor immune responses through promoting immunosuppression has been established as a critical hallmark of cancer [14]. Malignant cells achieve this by organizing into a complex structure composed of diverse cell types, including stromal cells, immune cells, and endothelial cells, all of which are in constant communication [15]. Tumors develop multiple tiers of immunosuppressive mechanisms to escape the host immune response, which has been extensively reviewed [16,17]. Briefly, this involves (1) secretion of immunosuppressive cytokines that inhibit anti-tumor adaptive (e.g., CTLs) and innate immune cells (e.g., NK cells) and polarize immune cells to pro-tumorigenic subtypes (e.g., T regulatory cells; T_{reg}), (2) secretion of chemokines that recruit immunosuppressive stromal and immune cells (e.g., myeloid-derived suppressor cells, tumor-associated fibroblasts, and macrophages) that in turn secrete immunosuppressive cytokines (e.g., IL-10, TGF β), (3) promotion of anergy and tolerance in anti-tumor immune cells through expression of surface inhibitory ligands (e.g., PD-L1) and persistent self-antigen presentation, (4) suppression of antigen presentation through, e.g., epigenetic mechanisms [18], to avoid detection by adaptive immunity and (5) upregulation of signaling pathways that reduce necessary metabolites (e.g., ATP) for immune cell (e.g., immature DC) activation in the tumor microenvironment [16].

Tumors can be classified as (1) immune cold (lack of immune infiltration) due to a lack of tumor antigens, deficiency in antigen presentation, absence of T cell priming, and impaired T cell trafficking, or (2) immune hot (high immune infiltration and increased mutational burden providing an abundance of tumor-specific neoantigens) [19]. Furthermore, not only the number of tumor-infiltrating lymphocytes (TIL; especially CD8+ T cells) but their spatial organization contribute to prognostic and predictive stratification in breast cancer [20–24]. This is also observed in ovarian cancer [25] and early-stage non-small cell lung cancer [26]. Thus, spatial, quantitative, and qualitative differences in the type of immune infiltrates in the tumor microenvironment are prognostic of disease outcome.

3. Cancer Immunotherapy and Challenges

In recent years, cancer immunotherapy has revolutionized the treatment of cancer. It aims to reinstate immune surveillance, turning cold tumors into hot tumors to eradicate cancer [19]. Numerous cancer immunotherapy modalities have been developed, and they are extensively reviewed [27]. Many solid tumors establish immune suppression by upregulating the expression of key immune checkpoint receptors (e.g., PD-1, CTLA4) on infiltrating T cells as well as immunosuppressive ligands (e.g., PD-L1, PD-L2, B7-H4), either on tumor cells or other cell types in the tumor microenvironment. [28,29]. Normally, PD-1 expressed on T cells restricts peripheral tissue damage and inflammation through limiting TCR-mediated effector T cell function and maintaining self-tolerance [29]. Tumors co-opt this pathway and promote immunosuppression by expressing PD-L1 downstream of oncogenic signaling pathways (e.g., STAT3, STAT1, Myc, 9p24.1 amplification, CD44 [30]) or following exposure to IFNγ secreted by T cells [29].

In this regard, the advent of immune checkpoint inhibitors (ICIs) targeting either CTLA-4, PD-1, or PD-L1 has revolutionized the field of cancer therapy, resulting in sustained clinical remissions in patients otherwise refractory to standard of cancer therapies in many cancer types, including melanomas and lung cancers. Indeed, there are currently one CTLA-4 and six PD-1/PD-L1 inhibitors approved across ten tumor types and numerous stages of cancer [31]. Unfortunately, while checkpoint inhibitors have resulted in strong and durable responses in some cancers, a large proportion of tumor types remain refractory to this treatment modality [32]. While a high mutational burden and increased density of infiltrating TILs are often predictive or response to ICIs, key questions remain. First, what other cell types dictate immunotherapeutic responsiveness (e.g., composition and the landscape of immune cells in the tumor microenvironment, unique tumor-specific signaling mechanisms) or lack thereof? Second, what are the predictive biomarkers for therapeutic responsiveness beyond the drug target itself? Third, what are the resistance mechanisms, conferred either by tumor cells or those within the local microenvironment? Based on the molecular understanding of how tumor-intrinsic signaling alters the immune response, a rational combination of chemotherapy, targeted therapy, and cancer immunotherapy methods need to be explored. Indeed, chemotherapy has already been shown to improve the sensitivity of immune checkpoint inhibitors (as reviewed in [33]).

In breast cancer, immune checkpoint inhibitors as single-agent or in combination with other therapies have been explored [34,35]. Stanton et al. examined 13,914 patient samples and determined that 5–26% of breast cancers have high infiltration of lymphocytes while 16% of cancers showed no infiltration. A median of 20% triple-negative (TNBC), 16% HER2+, and 6% ER+/PR+/HER2- (HR+) breast cancers show predominant lymphocyte infiltration (defined as >50% lymphocytic infiltrate) [36]. CD8+ CTLs indicative of antitumor immune responses, as well as FOXP3+ Treg cells indicative of a tumorigenic immune response, were most prominent in TNBC (60% infiltrated with CTLs and 70% infiltrated with T_{reg}) and HER2+ (61% and 67%) tumors compared to HR+ breast cancers (43% and 38%) [36]. These data indicate that subsets of breast cancers, especially within the TNBC and HER2+ subtypes, are relatively more immunogenic and contain high TILs [37,38]. Importantly, high infiltration of TILs has been associated with improved prognosis in earlystage TNBCs and HER2+ breast cancers [21,39–41], while the opposite is true in luminal breast cancer [42]. Based on these findings, immunotherapy approaches have been explored mostly in TNBC and HER2+ subtypes. As of January 2021, two immune checkpoint inhibitors have been approved by the FDA for breast cancer: (1) atezolizumab (PD-L1 inhibitor) with protein-bound paclitaxel for locally advanced, non-removable TNBC or metastatic TNBCs that are PD-L1 positive (IMpassion130 trial) [43], and (2) pembrolizumab (PD-1 inhibitor) for locally recurrent unresectable or metastatic TNBCs that are PD-L1 positive (KEYNOTE-355) [44]. Several important insights have been made from these and other preclinical studies. First, tumor mutational burden predicts prolonged survival associated with high (and not low) immune infiltration in breast cancer [45], especially in TNBC and HER2+ subtypes [34]. This is consistent with observations made in lung cancer and melanoma [46]. Second, only a subset of metastatic breast cancer patients, especially those expressing high tumor PD-L1, benefit from immune checkpoint blockade therapy [47]. Third, multiple layers of immunosuppressive mechanisms exist in the tumor microenvironment to impede treatment responsiveness [48]. Indeed, the single-agent activity of avelumab (PD-L1 inhibitor) [49], atezolizumab [50], and pembrolizumab [51] have shown limited activity in breast cancer patients, underscoring the need for rationally designed combination approaches [35].

4. Tumor-Intrinsic Kinase Signaling That Coordinate Cancer Immunosuppression

Since the proposal that cancer is a wound that never heals [52], studies in the 1990s demonstrated that tumors co-opt inflammation for survival [53–56]. Malignant progression not only relies on intrinsic signaling (loss of tumor suppressors and gain of oncogenes due to genetic aberrations) but also on extrinsic cellular players from the local microenvironment [57]. Today, it is fully established that tumor cell-intrinsic mechanisms continuously shape the tumor immune landscape to favor cancer progression and therapeutic resistance [57,58]. Oncogenic receptor and non-receptor kinases are crucial contributors of tumor-intrinsic signaling that coordinate cancer immunosuppression through diverse mechanisms (Figure 1).



Figure 1. Pharmacological inhibition of kinase signaling relieves immunosuppression in solid malignancies. Deregulated activation of numerous kinases belonging to receptor tyrosine kinase, non-receptor tyrosine kinases, and serine/threonine kinase families induce gene expression changes that potentiate the growth and metastatic spread of solid tumors. In addition to influencing tumor cell-intrinsic processes that are essential for malignant progression, kinase signaling networks allow solid tumors to evade anti-tumor immune responses through multiple mechanisms. These include: decreasing antigen processing and presentation, increased secretion of immunosuppressive molecules, increased expression of immune checkpoint ligands, and stimulation of chronic inflammation. As such, pharmacological inhibitors of kinase signaling networks can relieve immune suppression and improve the sensitivity of solid malignancies to immune checkpoint inhibitors. Permission to use adapted figure elements originally published by Elsevier Press (Hallmarks of Cancer: The Next Generation) was obtained: License #5017191244575.

5. Non-Receptor Kinases in Immunosuppression

One of the first demonstrations of an oncogenic kinase directly impacting the immune landscape came in 2004, whereby the H-RasG12V oncogene was shown to induce CXCL8 transcription in various cancer cell lines to promote macrophage infiltration and vascularization in vivo [59]. Numerous other studies have further linked deregulated activation of serine/threonine kinases, including the Ras/Raf/Mek/Erk and PI3K/AKT pathways as well as cyclin-dependent kinases, to the establishment of chronic inflammation in solid tumors. For example, BRAFV600E, a constitutively active form of the BRAF serine/threonine kinase, drives melanoma and has been shown to promote IL-6, IL-10, and VEGF secretion in a STAT3 dependent manner [60]. This, in turn, could suppress LPS induced inflammation by dendritic cells. Indeed, combination therapies that include MEK inhibitors with immunostimulatory agonists induce profound immunogenic responses in preclinical models of K-Ras positive pancreatic cancer by limiting the activation of immunosuppressive cell subsets, including M2 macrophages, myeloid-derived suppressor cells, and regulatory T cells [61]. In melanoma, the loss of PTEN (tumor suppressor that negatively regulates AKT/PI3K activity) leads to decreased numbers of TILs, reduced responsiveness to PD-1 checkpoint inhibition, and increased secretion of immunosuppressive cytokines [62]. In breast cancer patients, inhibition of cyclin-dependent kinases 4 and 6 (CDK4/6), which are fundamental drivers of cell cycle progression downstream of oncogenic signaling pathways, induces significant anti-tumor immune responses [63]. Non-receptor tyrosine kinases also contribute to cancer immune suppression. Indeed, nasopharyngeal carcinoma cells secrete ISG15, an IFN-responsive gene, which activates Src family kinases in macrophages to promote their M2 polarization and immunosuppressive properties [64]. In squamous cell

carcinoma, nuclear focal adhesion kinase (FAK) regulates transcription of CCL5 to promote T regulatory cell recruitment and exhaustion of CD8+ T cells to promote tumor growth, and treatment with FAK inhibitors reactivates anti-tumor immune responses [65]. Together, these studies suggest that targeting non-receptor kinase signaling networks may represent a promising therapeutic strategy to relieve tumor immune suppression, particularly as part of a rationally designed combination therapy.

6. Receptor Tyrosine Kinases in Immunosuppression

Overexpression or activating mutations in receptor tyrosine kinases leading to their aberrant activation are critical drivers of various solid tumors [66]. Receptor tyrosine kinases (RTKs) are single-pass transmembrane proteins expressed on the surface of various cell types that regulate proliferation, differentiation, survival, metabolism, migration, and invasion in cancer [66]. There are 58 known RTKs that fall into 20 subfamilies [66]. Upon binding to their cognate ligands, RTKs undergo conformational changes, leading to receptor dimerization and activation of their tyrosine kinase domains. These activated kinases then trans-phosphorylate key tyrosine residues in their cytoplasmic tails, leading to the recruitment of adaptor proteins that initiate downstream signaling pathways such as the mitogen-activated protein kinase (MAPK) and phosphoinositide 3- kinase (PI3K)/AKT pathways [67].

Studies have reported essential roles for several RTKs, including CSF1R, VEGFR, RON, and the TAM family, in promoting cancer immune suppression (reviewed in [68–71]). More recent studies have further elucidated the prominent examples of EGFR, HER2, and AXL signaling contributing directly to tumor-driven immunosuppression. For example, the high activation status of EGFR and HER2 is associated with increased PD-L1 expression in gastric cancer cells and patient tumor tissues [72]. Indeed, EGFR signaling in lung cancer activates the PD-1 immune checkpoint to promote immune evasion [73]. Constitutively active EGFR variants can induce immune suppression in lung cancers through their ability to shed mutant EGFR-containing exosomes into infiltrating dendritic cells, abrogating their ability to present tumor antigens [74]. Finally, increased Axl signaling stimulates NF-κB signaling to potentiate chronic inflammation and subsequent immune evasion in pancreatic cancer cells [75]. Together, these studies highlight an emerging role for RTKs in the establishment and maintenance of immune suppression and highlight the therapeutic potential for combining tyrosine kinase inhibitors with immunomodulatory agents (as discussed below).

7. Adaptor Proteins in Immunosuppression

Adaptor proteins are critical integrators of downstream tyrosine kinases to initiate oncogenic signaling cascades. They do so by nucleating signaling complexes through their ability to engage in both phospho-tyrosine dependent and—independent interactions. For example, the Shc1 adaptor protein is recruited to activated oncogenic RTKs and tyrosine kinases (TKs), such as EGFR, ERBB2, ERBB3, ERBB4, INSR, IGF1R, VEGFR3, FGFR1, TrkA, RET, MET, FGFR2, VEGFR2, c-kit, JAK3, EphA2, Src family kinases, Alk, PDGFRa, Ron, TrkB, and Axl through phospho-tyrosine binding domains such as PTB or SH2 domains [76–83]. This subsequently allows Shc1 to become phosphorylated on key tyrosine residues (Y239/240 and Y317, equivalent to Y313 in mice), which nucleates downstream signaling complexes to activate PI3K/AKT and Ras/MAPK oncogenic pathways [84]. Indeed, Shc1 signaling downstream of ErbB2 and polyomavirus middle T antigen (PyMT) oncogenes leads to suppression of CTL infiltration and IFN γ -driven immunity during the early stages of mammary tumorigenesis [85]. Importantly, mammary tumor progression of Shc1 deficient hyperplasias is significantly accelerated in athymic mice compared to immunocompetent animals, suggesting that Shc1 signaling suppresses T cell immune responses, ultimately facilitating tumor progression [85]. This is partially mediated by tyrosine 239/240 and tyrosine 313 residues of Shc1, which activate STAT3-dependent immunosuppression and inhibit STAT1-induced immune surveillance in breast cancer

cells, respectively [86]. Loss of phospho-Y313-Shc1 signaling is associated with a STAT1 dependent increase of MHC class I surface expression. It is plausible that numerous tumor intrinsic TKs may be implicated in regulating immunosuppression through Shc1. In line with this, targeting the pathways closely upstream or downstream of Shc1 (e.g., ErbB2, EGFR, MEK, ERK, PTEN, PI3K) by drugs or molecular manipulation has been shown to enhance anti-tumor immune responses in various studies, as discussed later. Moreover, Crk, another adaptor protein primarily known for its role in cell adhesion and growth factor signaling, has been demonstrated to promote immunosuppression in the 4T1 murine model of breast cancer [87]. Deletion of Crk enhances anti-tumor immune responses and secretion of cytokines that favor immune surveillance, leading to reduced tumor growth and metastasis. Loss of Crk also enhanced tumor clearance upon PD-1 checkpoint inhibition [87]. Taken together, these studies demonstrate the immunosuppressive role of oncogenic signaling pathways and how they may be targeted to elicit anti-tumor immunity.

8. Kinase Inhibitors Potentiate the Tumoricidal Responses of Immunotherapy

Oncogenic kinases drive immunosuppression, and their inhibition elicits anti-tumor immune responses, yet with inevitable resistance and recurrence. Despite the success of immunotherapy in oncology, only select patients experience durable responses due, in part, to persistent immunosuppression in the tumor microenvironment. Thus, a concomitant reduction of immunosuppression by kinase inhibitors while unlocking the therapeutic potential of tumoricidal immune cells has gained attention as a strategy to extend the clinical benefit of the current standard of therapies. To this end, roughly 53 small molecule kinase inhibitors and 12 antibody-based kinase inhibitors have been approved by the FDA for cancer treatment, 11 of which are for being studied in breast cancer (Supplementary Table S1). Kinase inhibitors have provided significant improvement in clinical outcomes in various cancers. However, they often lead to acquired resistance and recurrence after long-term exposure due to functional redundancy of the kinome and emergence of mutant variants that are resistant to the kinase inhibitor. Clinical trials have been initiated for various cancers to evaluate the efficacy of kinase inhibitors in combination with cancer immunotherapy (summarized in Supplementary Table S1).

9. Tyrosine Kinase Inhibitors

Cancer cells rely on a large family of cytoplasmic and receptor tyrosine kinases to initiate tumorigenic signaling pathways that ultimately activate downstream signaling molecules, including serine/threonine kinases, which are effectors of their immunosuppressive properties. In particular, pharmacological inhibitors targeting the ErbB2 RTK family (EGFR, ErbB2, ErbB3, and ErbB4) are standard of care for subsets of breast cancer and NSCLC patients and have been shown to elicit potent anti-tumor immune responses in the tumor microenvironment, thereby increasing the clinical impact of immune checkpoint inhibitors (Supplementary Table S1 and as reviewed in [88]). Several other tyrosine kinase inhibitors (TKI) also induce CTL-driven anti-tumor immune responses, including sunitinib, a broad spectrum RTK inhibitor that targets VEGFR, PDGFR α , Ret, and Kit, which has been shown to reverse immune suppression by inhibiting STAT3 signaling in renal cell carcinoma [89]. BMS-777607/ASLAN002, a Ron-selective kinase inhibitor, reduces breast cancer lung metastasis of breast cancer by promoting anti-tumor immune responses [90].

Recent studies have further elucidated the mechanisms of action by which smallmolecule tyrosine kinase inhibitors targeting the ErbB2 family evokes tumoricidal immune responses. The treatment of EGFR and HER2 overexpressing gastric cancer cells with Afatinib (a pan-ErbB2 family TKI) and lapatinib (EGFR, ErbB2 inhibitor) reduced PD-L1 expression levels [72]. Lapatinib treatment further decreased the secretion of immunosuppressive cytokines (e.g., CCL2, VEGF) from HER2-amplified tumor cells [72]. An ErbB2-driven breast cancer mouse model (MMTV/Neu) also responded to lapatinib treatment with increased IFN γ driven anti-tumor adaptive immune responses in a STAT1 dependent manner [91]. Other solid tumors undergoing clinical trials to test the efficacy of kinase inhibitors in combination with immunotherapy include non-small cell lung cancer [92], lung squamous cell carcinoma [93], and advanced renal cell carcinoma [94,95], and breast cancer (Supplementary Table S2). In hepatocellular carcinoma, immunotherapy has shown efficacy in the second-line setting, while tyrosine kinase inhibitors have shown benefit both in the first-(Sorafenib) and second-line (regorafenib, cabozantinib, and ramucirumab) settings [96–98]. Based on this, preliminary trials are ongoing in hepatocellular carcinoma patients to determine how the combination of tyrosine kinase inhibitors and immunotherapy prolongs survival compared to each as a single agent [99]. Additionally, promising safety profiles and results were seen in NSCLC (erlotinib and nivolumab) [100], while high toxicities have halted some trials [101]. In May 2019, a breakthrough randomized clinical trial in renal cell carcinoma (JAVELIN Renal 101) showed avelumab in combination with axitinib resulted in a significant survival benefit compared to standard of care sunitinib, leading to the FDA approval of this regimen [102].

Monoclonal antibody-based therapies targeting ErbB2 family signaling are also employed in cancer patients and rely, in part, on an intact immune system to achieve maximal clinical benefit. EGFR-neutralizing antibodies combined with chemotherapy depend on immunogenic cell death (ICD) to clear colorectal cancers [103]. Trastuzumab, a recombinant humanized monoclonal antibody directed against the human HER2 receptor tyrosine kinase, mediates tumor-killing partially by inducing antibody-dependent cell-mediated cytotoxicity (ADCC) against HER2 overexpressing tumor cells. Accordingly, the therapeutic effect of trastuzumab is diminished in mice that lack NK cells or those that have macrophages disabled to bind the Fc region of trastuzumab [104]. Similarly, NK cell-derived IFN γ induced PD-L1 expression in tumors and enhanced cetuximab (EGFR inhibitor)-mediated ADCC [105].

Finally, evidence suggests that cross-talk between cancer cells and immune cells influences the therapeutic responsiveness of these targeted therapies. One high-throughput immuno-oncology screen identified the EGFR inhibitor erlotinib as a potent enhancer of antigen-specific CTL tumor cell killing, synergizing with anti-PD-1 checkpoint inhibition to suppress colon cancer growth [106]. Preclinical studies in lung cancer show that the EGFR pathway enhances immunosuppression through increased engagement of PD-1/PD-L1 and CTLA4 in an ERK and NFkB dependent manner [107]. In line with this, erlotinib treatment in transgenic mice that develop EGFR^{L858R}-driven lung cancers induced the infiltration of T cells, B cells, NK cells, and CD11c+MHC-II+ cells as well as immunosuppressive CD11b+Gr1+ MDSCs [108]. Unlike the initial anti-tumorigenic immune infiltrates that are induced by EGFR TKIs, these therapy-induced increases in MDSC populations persist and are accompanied by increased levels of circulating immunosuppressive cytokines (IL-10, CCL2) in serum [109]. Thus, identifying strategies to prevent and/or overcome such TKIinduced immunosuppressive adaptive responses are required to achieve durable clinical benefit with these classes of drugs, particularly in combination with immune checkpoint inhibitors.

10. Serine/Threonine Kinase Inhibitors

Numerous signal transduction pathways, downstream of tyrosine kinases, bifurcate on key serine/threonine kinases, which phosphorylate effector molecules to potentiate the emergence of aggressive cancers and the establishment of an immunosuppressive tumor microenvironment.

10.1. CDK4/6 Inhibitors

This family of inhibitors, including abemaciclib, palbociclib, and ribociclib, targets CDK4 and CDK6, two serine/threonine kinases that are required for cell cycle progression. There are currently over 200 clinical trials ongoing with CDK4/6 inhibitors across multiple tumor types, including breast, lung, ovarian, colorectal, and prostate cancers. These inhibitors have shown particular promise in treating metastatic breast cancers, particu-

larly in ER-positive tumors in combination with hormonal therapies. Indeed, abemaciclib has been approved by the FDA for the treatment of advanced HR+HER2- breast cancer, based on the MONARCH 3 trial [110]. Abemaciclib, in combination with anastrozole (an aromatase inhibitor), results in increased adaptive immune response signatures that are phenotypic of increased T cell activation and antigen presentation even in early-stage HR+HER2- breast cancer, providing optimism for ongoing clinical trials in HR2+HER2disease (Supplementary Table S2) [111]. Preclinical studies further show that CDK4/6 inhibitors delay colorectal cancer growth in syngeneic mice, in part, by stimulating tumoricidal immune responses. This includes increased T cell infiltration, T cell effector function, antigen processing and presentation, macrophage and dendritic cell activation, combined with a potent inflammatory response [112,113]. Indeed, abemaciclib further potentiated the therapeutic benefit of PD-L1 inhibitors in colorectal tumors [112]. Unexpectedly, however, CDK4/6 inhibition increased PD-L1 protein stability in tumor cells by preventing proteasome-mediated PD-L1 degradation [114]. These studies highlight the mechanistic basis for combining CDK4/6 inhibitors and PD-1/PD-L1 inhibitors as this combinatorial approach stimulates an immunogenic tumor microenvironment and primes cancers for PD1/PD-L1 immune checkpoint blockade.

10.2. RAF/MEK Inhibitors

There has been significant interest in exploring RAF/MEK inhibitors as enhancers of anti-tumor immune responses based on early preclinical studies showing that MEK inhibitors potentiate CD8+ T cell responses by preventing an exhausted phenotype in cancer models, leading to durable responses in combination with PD-L1 inhibitors [115]. Vemurafenib and dabrafenib are two selective V600 mutant BRAF inhibitors that have been deployed for the treatment of V600E+ tumors, including melanoma, colorectal cancer, and non-small cell lung cancers. Cobimetinib and trametinib are MEK1/2 inhibitors that function downstream of BRAF and are indicated for treating both BRAF wild-type and mutant tumors. Numerous clinical trials have reported on the efficacy of this family of inhibitors in overcoming immune suppression. Recent clinical trials have evaluated the safety and efficacy of triple therapies that combine a BRAF and MEK inhibitor together with an immune checkpoint inhibitor. For example, a recent phase Ib study tested vemurafenib and cobimetinib, combined with a PD-L1 inhibitor (atezolizumab), in patients with BRAF V600E metastatic melanoma (NCT1656642). Durable anti-tumor responses were observed in 40% of patients even 30 months following combination treatment, which was associated with increased recruitment of activated TILs [116]. This is further supported by a recent phase 3 COMBI-I clinical trial (NCT02967692) that examined the efficacy of a novel PD-1 inhibitor (spartalizumab) in combination with dabrafenib and trametinib in patients with unresectable and/or metastatic BRAF V600E mutant melanomas. A 78% objective response rate was observed, whereby 44% of patients showed a complete pathological response. Although 72% of patients experienced an immune-related adverse event, this study suggested that combined inhibition of BRAF and MEK signaling together could increase the efficacy of immune checkpoint inhibitors [117]. Preclinical studies in mouse models of BRAF V600E mutant melanoma have begun to elucidate the mechanistic basis for improved immunological responses to combined treatment with BRAF and MEK inhibitors. Combined dabrafenib and trametinib treatment increased the infiltration and cytotoxicity of TILs, which was associated with decreased recruitment of tumor-associated macrophages, T regulatory cells and increased antigen processing and presentation of melanosomal antigens [118]. Trametinib has been associated with increased HLA expression, increased CD8+ T cell infiltration, and improved tumor control in combination with checkpoint inhibitors [119–122]. In addition, dual BRAF/MEK inhibition further decreased the production of immunosuppressive adenosine in BRAF mutant melanoma cells by reducing the expression of components within the CD73 adenosinergic pathway [123].

10.3. PI3K/mTOR Inhibitors

The PI3K/AKT/mTOR signaling pathway is commonly activated in solid tumors and is critical for tumor progression, metastatic spread, and therapeutic resistance [124,125]. PI3K proteins are phosphatidylinositol 3' lipid kinases and are composed of heterodimers with regulatory (p85) and catalytic subunits (p110). Four catalytic isoforms (p110 α , p110 β , p110 γ , p110 δ) are activated downstream of tyrosine kinases, and G-protein coupled receptors, leading to the activation of multiple serine/threonine kinases, including several AKT family members (AKT1, AKT2, AKT3) and mechanistic target of rapamycin (mTOR), which functions within two distinct protein complexes (mTORC1 and mTORC2). Together, these serine/threonine kinases phosphorylate hundreds of target proteins with oncogenic functions. Indeed, PI3K/AKT/mTOR functions as a nutrient sensor to induce cancer cell proliferation and survival under nutrient replete conditions [126]. It does so, in part, by increasing the rate of protein synthesis by promoting the assembly of the eIF4F complex (eIF4E/4A/4G), which potentiates cap-dependent translation initiation. Indeed, mRNA translation of genes with oncogenic properties is selectively controlled by eIF4E availability, the rate-limiting step for eIF4F complex assembly. Moreover, phosphorylation of eIF4E by MNK1/2, another class of serine/threonine kinases, further potentiates the rate of mRNA translation of eIF4E-sensitive genes [127].

Dysregulation of the PI3K/AKT/mTOR signaling pathway in solid tumors has been implicated in the establishment of immunosuppression by numerous studies by stimulating the production of immunosuppressive cytokines, recruitment of immunosuppressive cell types, and induction of immune checkpoint ligands on tumor cells (as reviewed in [128,129]). Recent literature suggests that PTEN-deficient tumors show increased expression of immunosuppressive cytokines and genetic silencing of PTEN expression in melanoma cells attenuated anti-tumorigenic T cell responses in vivo, leading to resistance to PD-1 immune checkpoint blockade [62]. Intriguingly, another study found that PTEN was essential to induce IFN-driven innate immunity and potentiate anti-viral responses independent of its role in negatively regulating PI3K/AKT signaling. Instead, PTEN protein phosphatase activity was shown to control IRF3 nuclear import, a key transcription factor that regulates type I IFN responses [130]. This study highlights potential novel and unappreciated roles for PTEN in promoting immune surveillance in solid tumors. Having said this, other preclinical studies clearly suggest the mTOR/MNK/eIF4E activation in solid tumors promotes an immunosuppressive microenvironment. For example, mTOR signaling in breast cancer cells increases G-CSF expression to stimulate the recruitment of immunosuppressive MDSCs [131]. Deregulated PI3K signaling in tumor cells further establishes an immunosuppressive niche by inducing activation of pro-inflammatory mediators, including nitric oxide synthase and lipoxygenase, in the tumor microenvironment [132]. Furthermore, drugs targeting the mRNA translation machinery, including eFT508 (MNK1/2 inhibitor) and silvestrol (eIF4A inhibitor), reduce PD-L1 expression by tumor cells in models of liver cancer and melanoma, sensitizing tumors to T cell-dependent immune responses [133,134]. Finally, early preclinical studies suggested that combining a pan-PI3K inhibitor with an immune adjuvant induces production of IFN γ and IL-17producing inflammatory T cells, leading to profound anti-tumorigenic immune responses in mouse models of lung cancer and melanoma, paving the way for future studies looking at combination therapy with immunotherapies [135].

In light of these observations, significant efforts by the pharmaceutical sector have yielded numerous inhibitors that selectively target PI3K/AKT/mTOR signaling [136,137]. These include pan-PI3K and/or dual PI3K/mTOR inhibitors (buparlisib, bimiralisib, co-panlisib, dactolisib, idelalisib, apitolisib, gedatolisib, tenalisib), isoform-specific PI3K inhibitors (taselisib, alpelisib, parsaclisib, serabelisib, umbralisib), AKT inhibitors (AZD5363, MK2206), mTORC1-selective inhibitors (rapalogs, including temsirolimus, everolimus), dual mTORC1/mTORC2 inhibitors (asTORi, including INK128, AZD8055, LXI-15029) and MNK inhibitors (tomisvosertib). Several recent phase I clinical trials with these inhibitors demonstrate their therapeutic potential in reversing immune suppression. For

example, a phase I study testing taselisib, a p110 α -specific inhibitor, in women with triplenegative breast cancer showed increased expression of genes associated with activated T cell and NK cell responses, co-incident with anti-tumor responses [138]. Moreover, SF2523, a pan-PI3K and dual BRD4 inhibitor, inhibited the growth of lung, melanoma, and colorectal cancers in syngeneic models, which was associated with reduced infiltration of MDSCs and restoration of CD8+ T cell function [139]. Indeed, in murine models of metastatic breast cancer, buparlisib induced an inflammatory response and synergized with PD-1 neutralizing antibodies [140]. Temsirolimus, an mTOR inhibitor, could enhance anti-tumor immune responses in melanoma and renal cell cancer mouse models when used with cancer vaccines [141]. In contrast, everolimus, a mTORC1-specific inhibitor, has been shown to suppresses CTL and NK cell function and upregulate the presence of regulatory T cells [142,143], which is in line with its use as an immunosuppressant in organ transplantation [144,145]. These negative effects could be alleviated by combining cyclophosphamide with everolimus, leading to the depletion of Tregs and MDSCs, co-incident with an increased level of CD8+ effector T cells in the blood of patients with metastatic renal cell carcinoma [146]. This highlights the need for careful consideration into drug combinations. Together, these studies establish that pharmacological targeting of the PI3K/AKT/mTOR signaling pathway may synergize with immune checkpoint inhibitors in eliciting tumoricidal immune responses in solid tumors.

11. Combination Strategies Targeting Kinase Inhibitors Improve the Efficacy of Immune Checkpoint Inhibitors in Breast Cancer

Durable clinical responses to immune checkpoint inhibitors have only been observed in cancer types, including melanoma and NSCLCs, which exhibit a high degree of genomic instability and are immunologically "hot" tumors. In contrast, most breast cancers are not infiltrated by abundant TILs and have low levels of microsatellite instability, resulting in the paucity of available tumor antigens. In this regard, clinical trials examining whether rationally designed combination therapies can increase immunological responses are ongoing. Many breast cancer clinical trials using immunotherapy combined with chemotherapy or other targeted agents have mostly been in TNBC and HER2+ tumors as these subtypes display the highest immunogenicity (PD-L1+ TIL, PD-L1+ tumor, mutation, neoantigen load, and MHC expression) [34]. Other kinase inhibitors that have shown promise include Raf/MEK inhibitors, PI3K/mTOR inhibitors, CDK4/6 inhibitors, and HER2 inhibitors [35]. Current clinical trials combining kinase inhibitors with checkpoint inhibitors in breast cancer are summarized (Supplementary Table S2). Biomarkers associated with immunomodulation are being assessed with a renewed focus for dual treatment of lapatinib and trastuzumab (NCT02213042) or trastuzumab and pertuzumab (NCT03144947) in the neoadjuvant setting in advanced HER2+ breast cancer patients, which may further guide future combination strategies.

Beyond checkpoint inhibitors, cancer vaccines have also been tested in breast cancer clinical trials, albeit with limited success. Metastatic, trastuzumab-refractory HER2+ breast cancer patients were treated with lapatinib and a HER2-based cancer vaccine (a recombinant protein with extracellular domain and part of the intracellular domain of HER2 combined with an adjuvant) concurrently based on success in a preclinical model. However, no objective clinical responses were seen [147]. Previously, a single-arm, non-randomized feasibility study in HER2+ metastatic breast cancer patients (n = 20) was done using a HER2+ whole-cell breast cancer vaccine and weekly trastuzumab. This showed a 6-month clinical benefit of 55%, which was supported by mouse model studies with control groups [148]. While the results were encouraging, further studies with larger cohorts and control arms are necessary to determine the true benefit of cancer vaccines to treat HER2+ breast cancers.

12. Kinase Inhibitors and Tumor-Intrinsic Antigen Processing and Presentation

Intriguingly, multiple studies have linked the efficacy of kinase inhibitors to high MHC class I antigen presentation by tumor cells. Cabozantinib (targets RET and MET)

has been shown to increase MHC class I (H-2Db) and Fas expression in colon cancer cell lines [149]. In NSCLC, mutation or overexpression of EGFR promotes immunosuppression, and the inhibition of EGFR using gefitinib or erlotinib can restore MHC class I expression, reduce PD-L1 expression or upregulate the expression of NKG2D ligands for NK cell-mediated tumor killing [107]. Furthermore, CDK4/6 inhibitor abemaciclib induces upregulation of antigen presentation in the context of MHC class I, leading to breast tumor regression [63,112]. A similar increase in tumor cell surface MHC-I expression upon abemaciclib treatment was observed in a mouse colorectal tumor model [112] and RB positive Ewing sarcoma preclinical model [113]. BRAF inhibitor vemurafenib has been shown to upregulate MHC in BRAFV600E homozygous melanoma [150]. High-throughput shRNA screens revealed that MEK, EGFR, and RET negatively regulate antigen processing and presentation machinery and MHC class I expression in an ERK-dependent manner [151]. In line with this, pharmacological inhibition of these kinases led to improved T cell-mediated killing through antigen-MHC recognition [151]. FDA-approved tyrosine kinases that alter antigen presentation pathways are noted in Supplementary Table S1. It was recently shown that palbociclib (CDK4/6 inhibitor), in addition to increasing the MHC class I expression, can alter the peptide-MHC repertoire in melanoma cell lines to reflect the intracellular response to CDK4/6 inhibition [152]. How the quality and quantity of tumor-associated antigen repertoires are impacted by kinase inhibitors will have important implications for therapeutic cancer vaccine development and other cell-based immunotherapy modalities that target tumor-specific antigens [153]. Together, these studies provide the basis for a strategic combination of kinase inhibitors (small molecule inhibitor and antibody-based) with synthetic immune-based therapy (engineered TCR-based or antibody-based) for the treatment of cancer.

13. Kinase Inhibitors Impact JAK/STAT-Mediated Tumor Immunity

The JAK/STAT signaling pathway is critical for the initiation and subsequent resolution of inflammatory responses. Janus kinases (JAK) are a family of tyrosine kinases (JAK1, JAK2, JAK3, and TYK2) that are activated downstream of multiple cytokine receptors, both in tumor cells and immune cells. They are activated by growth factors (e.g., EGF) as well as inflammatory (e.g., IFN α , IFN β , IFN γ , IL-6, IL-23) and immunosuppressive (e.g., IL-10, IL-27, IL-35) cytokines. Once activated, JAKs phosphorylate specific members of the Signal Transducer and Activator of Transcription (STAT1-6) transcription factor family. Tyrosine phosphorylation allows the formation of STAT family homo- and hetero-dimers, which translocate to the nucleus where they induce the expression of hundreds of anti-viral, inflammatory, or immunosuppressive genes [154].

In solid cancers, STAT1 and STAT3 have pleiotropic roles during cancer development and in the establishment of immune responses. Type I (IFN α/β) and type II (IFN γ) interferons stimulate the formation of STAT1/STAT1 homodimers or STAT1/STAT2 heterodimers to induce inflammatory responses [155]. In contrast, the delayed formation of STAT1/STAT3 heterodimers downstream of IFN receptors negatively regulate STAT1dependent inflammatory responses to induce immune suppression [156]. By the same token, multiple cytokines, including IL-6 and IL-10, induce the formation of STAT3/STAT3 homodimers, which increase the expression of genes that recruit and activate immunosuppressive macrophages (M2) and regulatory T cells [157]. Coupled with the observation that the STAT3 pathway also induces tumor growth, angiogenesis, and metastasis, several STAT3 inhibitors are in development to treat solid malignancies [158]. Indeed, we previously showed that an increased ratio of active STAT1/STAT3 in breast tumors is associated with improved immune surveillance, increased production of inflammatory cytokines, and enhanced sensitivity to immune checkpoint inhibitors in breast cancer [86].

The IFN/STAT1 pathway functions as a double-edged sword in cancer development [159]. Several studies have shown that it coordinates tumor-suppressive transcriptional responses, both in the tumor cells themselves and in cells within the tumor microenvironment through multiple mechanisms. IFN/STAT1 signaling potentiates cell cycle arrest and apoptosis in tumor cells, induces angiostatic responses, increases antigen processing and presentation by tumor cells, and primes the innate and adaptive immune cells to activate immune cell subsets (Th1/CTL; NK) that promote anti-tumor immunity. While STAT1 is required for immune surveillance, its chronic activation paradoxically potentiates and maintains tumor immune evasion by increasing the expression of immunosuppressive mediators (PD-L1, IDO1) [159]. Indeed, whereas STAT1 signaling in NK and T cells potentiates their effector functions [160,161], sustained STAT1 signaling in T cells protects them from NK-cell mediated cytotoxicity, preventing their elimination in inflamed tissues [162].

Several studies highlight this complex relationship between JAK/STAT1 signaling, tumor immunity, and sensitivity to immune checkpoint inhibitors in oncology. For example, whole exome and transcriptomic analysis of >1000 tumors treated with immune checkpoint inhibitors (across seven tumor types) showed that an elevated clonal tumor mutational burden, coupled with increased expression of CXCL9 (an IFN-inducible gene), is predictive of superior response [163]. In an independent study, macrophage-derived CXCL9 and CXCL10 were significantly elevated in response to immune checkpoint inhibition and were required to mount effective CTL-driven anti-tumor immune responses [164]. In high-grade serous ovarian carcinomas, elevated PD-L1 levels, indicative of increased STAT1 activation, is associated with elevated numbers of tumor-infiltrating lymphocytes and good outcome [165]. Finally, tumors with a stem-like phenotype possess decreased type I IFN/STAT1 signaling and are highly immunosuppressive, despite their high mutational burden [166]. Indeed, glioma stem cells evade immune surveillance by downregulating STAT1 expression at the epigenetic level [167].

In contrast, some studies point to IFN/STAT1 activation in the inferior response to kinase and/or immune checkpoint inhibition. For example, two neo-adjuvant clinical trials, including palbociclib (CDK4/6i) plus endocrine therapy, showed that increased IFN/STAT1 signaling in ER+ breast cancers were associated with elevated immune checkpoint levels, endocrine resistance, and poor outcome [168]. In pancreatic tumors, dinaciclib (a pan CDK inhibitor targeting CDK2/5/9) reversed immune suppression by inhibiting IFNγ-induced expression of immunosuppressive mediators, including IDO1 and PD-L1 [169]. Indeed, IFN/STAT1 activation in individual HER2+ breast cancers can exert opposing effects on their sensitivity to HER2 kinase inhibitors. Whereas Th1 cytokines, including IFNs, sensitized HER2+ tumors to lapatinib [170], trastuzumab increased PD-L1 expression in breast tumors, contributing to trastuzumab resistance [171].

Combined, these studies demonstrate a complex role for IFN/STAT1 signaling in tumor development and immune evasion, highlighting the need for further research to identify whether activation of this pathway will potentiate and/or suppress sensitivity to immune checkpoint inhibitors, alone or in combination with kinase inhibitors.

14. Kinase Inhibitors Target Immune Cells in the Tumor Microenvironment

Immune cells depend on multiple kinases to function. As such, kinase inhibitors used to treat cancer also directly regulate immune cell signaling and activity in the tumor microenvironment (reviewed in [172]), leading both to immune-related adverse events as well as improved sensitivity to immune-based therapies.

For example, skin inflammation is a prominent adverse effect of some EGFR inhibitors through off-target inhibition of Ste-10-like (STK10) serine/threonine kinase, leading to enhanced lymphocyte migration and secretion of IL-2 [173]. Moreover, immune checkpoint inhibitors are frequently associated with immune-related adverse events in cancer patients, and emerging evidence suggests that kinase inhibitors may be employed to modulate these toxicities. Indeed, the treatment of a melanoma patient experiencing anti-PD1 induced colitis with an mTOR inhibitor (sirolimus) could dampen systemic inflammatory responses and relieve this toxicity while sparing the anti-tumorigenic effects of PD-1 blockade [174].

Perturbation of signaling pathways in immune cells within the tumor microenvironment by kinase inhibitors also affects the efficacy of targeted and immune based-therapies, altering patient outcomes. Several RTKs, including Tyro3, Mer, and Axl (TAM-family), are expressed on APCs (macrophage and dendritic cells) and negatively regulate their activation and antigen presentation capabilities [175]. Inhibition of these TAM receptors on NK cells also leads to rejection of breast cancer metastasis in mouse models [176]. Given that intra-tumoral Tyro3, Mer, and Axl signaling contributes to tumor growth, immune evasion, drug resistance, proliferation, and metastasis [177–179], TAM RTKs represent an important drug candidate to simultaneously target both malignant cells and immune cells in order to enhance anti-tumor immunity. Similarly, other kinase inhibitors that induce tumor-intrinsic effects also target immune cells to simultaneously relieve immune suppression as part of their mechanism of action. While trametinib is used to treat many solid tumors, this MEK inhibitor also suppresses naïve CD8+ T cell priming and protects CD8+ T cells from chronic T cell receptor activation, leading to synergy with anti-PD-L1 inhibitors [115]. CDK4/6 inhibition also elicits increased antigen presentation in tumor cells and suppresses the proliferation of regulatory T cells to result in CTL-mediated anti-tumor immunity [60].

PI3K/AKT/mTOR signaling is critical in controlling immune cell function and has been linked to immunosuppressive immune cell function in the tumor milieu. $PI3K\gamma$, which is expressed specifically in immune cells, is an important drug target for pan PI3K inhibitors in modulating anti-tumor immune responses. Indeed, PI3K γ is required in Tregs and MDSCs to stimulate their infiltration into tumor tissue [180]. Moreover, PI3K γ expression in macrophages negatively regulates the pro-inflammatory TLR4/NFkB signaling pathway while positively regulating IL-4 and C/EBPb signaling [181], polarizing macrophages toward immunosuppressive types. In line with this, PI3K γ inhibitors can relieve macrophage-driven immunosuppression on T cells and synergize with PD-1 inhibitors to impede tumor growth [181]. Similarly, Mnk signaling in macrophages also increases their immunosuppressive properties [182]. On the other hand, dactolisib, a panspecific PI3K inhibitor, reduced mRNA translation initiation in granulocytic MDSCs in a preclinical model of prostate cancer, inhibiting their immunosuppressive properties [183]. Indeed, dactolisib synergized with immune checkpoint blockade to induce durable tumoricidal responses in prostate cancer models [184]. Beyond this, mTOR signaling in multiple immune cell types contributes to immune suppression. For example, glioblastoma cells upregulate mTOR signaling in microglia, tissue-resident macrophages, which increases their immunosuppressive properties [185]. Furthermore, inhibition of mTOR signaling in T cells allows for their spontaneous activation into effector T cells, suggesting that this pathway is important for T cell tolerance. Indeed, mTOR function is required for the generation of regulatory T cells through metabolic reprogramming [186]. Similarly, mTOR activation increases fatty acid synthesis in dendritic cells, which indirectly reduces acetyl CoA pools, leading to reduced histone acetylation. This epigenetic reprogramming minimizes the ability of DCs to activate cytotoxic T lymphocytes [187].

While some studies indicate kinase inhibitors as promising agents to reverse immunosuppression in combination with cancer immunotherapy, others show that kinase inhibitors may dampen anti-tumor immune responses, potentially contributing to their lack of efficacy in cancer treatment [188]. For example, mTOR signaling plays a complex role in regulating NK cell function whereby mTORC1 signaling stimulates NK cell cytolytic function, whereas mTORC2 activity favors immunosuppressive NK cells [189]. Finally, mTORC2 deletion in macrophages stimulates a pro-inflammatory microenvironment that potentiates colitis-induced colon cancer [190]. These studies suggest that rapalogs (mTORC1-specific) and active site dual-specificity mTOR inhibitors (asTORi) may differentially impact the tumor immune microenvironment and, potentially, sensitivity to combination immunotherapies. Combined, these studies demonstrate that kinase inhibition impacts both tumor cells and immune cells in the tumor microenvironment to modulate treatment response and sensitivity to immunotherapy. Thus, it is critical to investigate the impact of individually targeted therapies on immune cells to maximize their ability to synergize with immunotherapies.

15. Conclusions

Collectively, a combination of kinase inhibitors and immunotherapy holds promise in cancer treatment. It remains to be seen whether kinase inhibitors combined with immunotherapy will be effective in different subtypes of cancers. Where there is a lack of response, its mechanisms should be investigated. The intrinsic kinase signaling pathways tumors employ to adapt and resist cancer immunotherapy need to be investigated. Where efficacy is seen, it must be established if a combination approach—over a single-agent approach—is preferred. In addition to duration, the order and timing of the combined approach, whether phased or sequential use of the drugs is as effective need to be tested [112]. Mechanistic studies that inform rational combination of kinase inhibitors and immunotherapeutic modalities for clinical trials are in need. Given all the evidence presented above, combinatorial use of kinase inhibitors and cancer immunotherapy may help to combat drug resistance and broaden responsiveness.

Supplementary Materials: The following are available online at https://www.mdpi.com/1422-0 067/22/5/2608/s1, Table S1: Summary of kinase inhibitors approved by the FDA to treat solid tumors, their molecular targets and the studies demonstrating their regulatory roles on tumor immunogenicity or immune responses, Table S2: Ongoing clinical trials of kinase inhibitors approved for breast cancer treatment in combination with immunotherapy. Only the kinase inhibitors and the immunotherapeutic agents that are combined are listed.

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References

- Shankaran, V.; Hiroaki, I.; Allen, T.; Bruce, J.M.; White, P.E.; Swanson, L.J.; Schreiber, R.D. IFN [Gamma] and Lymphocytes Prevent Primary Tumour Development and Shape Tumour Immunogenicity. *Nature* 2001, 410, 1107–1111. [CrossRef]
- Dunn Gavin, P.; Allen, T.; Bruce, H.I.; Lloyd, J.; Robert, D.S. Cancer Immunoediting: From Immunosurveillance to Tumor Escape. Nat. Immunol. 2002, 3, 991–998. [CrossRef]
- Dunn, G.P.; Old, L.J.; Schreiber, R.D. The Immunobiology of Cancer Immunosurveillance and Immunoediting. *Immunity* 2004, 21, 137–148. [CrossRef]
- Kim, H.-J.; Cantor, H. The Path to Reactivation of Antitumor Immunity and Checkpoint Immunotherapy. *Cancer Immunol. Res.* 2014, 2, 926–936. [CrossRef]
- DeNardo, D.G.; Andreu, P.; Coussens, L.M. Interactions between lymphocytes and myeloid cells regulate pro-versus anti-tumor immunity. *Cancer Metastasis Rev.* 2010, 29, 309–316. [CrossRef] [PubMed]
- 6. Zitvogel, L.; Tesniere, A.; Kroemer, G. Cancer Despite Immunosurveillance: Immunoselection and Immunosub-version. *Nat. Rev. Immunol.* 2006, *6*, 715–727. [CrossRef]
- Voskoboinik, I.; Whisstock, J.C.; Trapani, J.A. Perforin and Granzymes: Function, Dysfunction and Human Pathology. *Nat. Rev. Immunol.* 2015, 15, 388–400. [CrossRef]
- 8. Wang, W.; Erbe, A.K.; Hank, J.A.; Zachary, S.M.; Sondel, P.M. Nk Cell-Mediated Anti-Body-Dependent Cellular Cytotoxicity in Cancer Immunotherapy. *Front. Immunol.* 2015, *6*, 368. [CrossRef] [PubMed]
- 9. Tsou, P.; Katayama, H.; Ostrin, E.J.; Hanash, S.M. The Emerging Role of B Cells in Tumor Immunity. *Cancer Res.* 2016, 76, 5597–5601. [CrossRef] [PubMed]
- 10. Mahmoud, S.M.A.; Lee, A.H.S.; Paish, E.C.; Macmillan, R.D.; Ellis, I.O.; Green, A.R. The prognostic significance of B lymphocytes in invasive carcinoma of the breast. *Breast Cancer Res. Treat.* **2011**, *132*, 545–553. [CrossRef]
- 11. Wouters, M.C.A.; Nelson, B.H. Prognostic Significance of Tumor-Infiltrating B Cells and Plasma Cells in Human Cancer. *Clin. Cancer Res.* **2018**, *24*, 6125–6135. [CrossRef]
- 12. Boero, S.A.; Morabito, B.; Banelli, B.; Cardinali, B.; Dozin, G.; Lunardi, P.; Piccioli, S.; Lastraioli, R.; Carosio, S.S.; Levaggi, F.A.; et al. Analysis of in Vitro Adcc and Clinical Response to Trastuzumab: Possible Relevance of Fcgammariiia/Fcgammariia Gene Polymorphisms and Her-2 Expression Levels on Breast Cancer Cell Lines. *J. Transl. Med.* **2015**, *13*, 324. [CrossRef]

- 13. Lucarini, V.; Melaiu, O.; Tempora, P.; D'Amico, S.; Locatelli, F.; Fruci, D. Dendritic Cells: Behind the Scenes of T-Cell Infiltration into the Tumor Microenvironment. *Cancers* **2021**, *13*, 433. [CrossRef]
- 14. Hanahan, D.; Weinberg, R.A. Hallmarks of Cancer: The Next Generation. Cell 2011, 144, 646–674. [CrossRef]
- Ott, P.A.; Adams, S. Small-Molecule Protein Kinase Inhibitors and Their Effects on the Immune System: Implications for Cancer Treatment. *Immunotherapy* 2011, 3, 213–227. [CrossRef]
- Kroemer, G.; Senovilla, L.; Galluzzi, L.; André, F.; Zitvogel, L. Natural and therapy-induced immunosurveillance in breast cancer. *Nat. Med.* 2015, 21, 1128–1138. [CrossRef]
- 17. Gabrilovich, D.I.; Ostrand-Rosenberg, S.; Bronte, V. Coordinated regulation of myeloid cells by tumours. *Nat. Rev. Immunol.* 2012, 12, 253–268. [CrossRef] [PubMed]
- 18. Hãc Ninger, E.; Krueger, T.E.G.; Lang, J.M.; Heninger, E. Augmenting Antitumor Immune Responses with Epigenetic Modifying Agents. *Front. Immunol.* **2015**, *6*, 29. [CrossRef]
- 19. Bonaventura, P.; Shekarian, T.; Alcazer, V.; Valladeau-Guilemond, J.; Valsesia-Wittmann, S.; Amigorena, S.; Caux, C.; Depil, S. Cold Tumors: A Therapeutic Challenge for Immunotherapy. *Front. Immunol.* **2019**, *10*, 168. [CrossRef]
- Joel, S.; Gupta, R.; Hou, L.; Kurc, T.; Singh, P.; Nguyen, V.; Samaras, D.; Shroyer, K.R.; Zhao, T.; Batiste, R.; et al. Spatial Organization and Molecular Correlation of Tumor-Infiltrating Lymphocytes Using Deep Learning on Pathology Images. *Cell Rep.* 2018, 23, 23181.
- Li, X.; Gruosso, T.; Zuo, D.; Omeroglu, A.; Meterissian, S.; Guiot, M.-C.; Salazar, A.; Park, M.; Levine, H. Infiltration of CD8+ T cells into tumor cell clusters in triple-negative breast cancer. *Proc. Natl. Acad. Sci. USA* 2019, *116*, 3678–3687. [CrossRef] [PubMed]
- König, L.; Mairinger, F.D.; Hoffmann, O.; Bittner, A.-K.; Schmid, K.W.; Kimmig, R.; Kasimir-Bauer, S.; Bankfalvi, A. Dissimilar patterns of tumor-infiltrating immune cells at the invasive tumor front and tumor center are associated with response to neoadjuvant chemotherapy in primary breast cancer. *BMC Cancer* 2019, *19*, 1–13. [CrossRef]
- Heindl, A.; Sestak, I.; Naidoo, K.; Cuzick, J.; Dowsett, M.; Yuan, Y. Relevance of Spatial Heterogeneity of Immune Infiltration for Predicting Risk of Recurrence After Endocrine Therapy of ER+ Breast Cancer. J. Natl. Cancer Inst. 2018, 110, 166–175. [CrossRef]
- 24. Nawaz, S.; Heindl, A.; Koelble, K.; Yuan, Y. Beyond immune density: Critical role of spatial heterogeneity in estrogen receptornegative breast cancer. *Mod. Pathol.* 2015, *28*, 766–777. [CrossRef]
- 25. Zhang, A.W.; McPherson, A.; Milne, K.; Kroeger, D.R.; Hamilton, P.T.; Miranda, A.; Funnell, T.; Little, N.; De Souza, C.P.; Laan, S.; et al. Interfaces of Malignant and Immunologic Clonal Dynamics in Ovarian Cancer. *Cell* **2018**, *173*, 1755–1769. [CrossRef]
- Corredor, G.; Wang, X.; Zhou, Y.; Lu, C.; Fu, P.; Syrigos, K.N.; Rimm, D.L.; Yang, M.; Romero, E.; Schalper, K.A.; et al. Spatial Architecture and Arrangement of Tumor-Infiltrating Lymphocytes for Predicting Likelihood of Recurrence in Early-Stage Non–Small Cell Lung Cancer. *Clin. Cancer Res.* 2019, 25, 1526–1534. [CrossRef]
- 27. Riley, R.S.; June, C.H.; Langer, R.; Mitchell, M.J. Delivery technologies for cancer immunotherapy. *Nat. Rev. Drug Discov.* 2019, 18, 175–196. [CrossRef]
- 28. Topalian, S.L.; Drake, C.G.; Pardoll, D.M. Immune Checkpoint Blockade: A Common Denominator Approach to Cancer Therapy. *Cancer Cell* **2015**, *27*, 450–461. [CrossRef]
- 29. Xia, Y.; Medeiros, L.J.; Young, K.H. Immune checkpoint blockade: Releasing the brake towards hematological malignancies. *Blood Rev.* **2016**, *30*, 189–200. [CrossRef]
- 30. Kong, T.; Ahn, R.; Yang, K.; Zhu, X.; Fu, Z.; Morin, G.; Bramley, R.; Cliffe, N.C.; Xue, Y.; Kuasne, H.; et al. CD44 Promotes PD-L1 Expression and Its Tumor-Intrinsic Function in Breast and Lung Cancers. *Cancer Res.* **2019**, *80*, 444–457. [CrossRef] [PubMed]
- 31. Himmel, M.E.; Saibil, S.D.; Saltman, A.P. Immune checkpoint inhibitors in cancer immunotherapy. *Can. Med. Assoc. J.* 2020, 192, e651. [CrossRef]
- Sade-Feldman, M.; Yizhak, K.; Bjorgaard, S.L.; Ray, J.P.; De Boer, C.G.; Jenkins, R.W.; Lieb, D.J.; Chen, J.H.; Frederick, D.T.; Barzily-Rokni, M.; et al. Defining T Cell States Associated with Response to Checkpoint Immunotherapy in Melanoma. *Cell* 2018, 175, 998–1013. [CrossRef] [PubMed]
- 33. Galluzzi, L.; Humeau, J.; Buqué, A.; Zitvogel, L.; Kroemer, G. Immunostimulation with Chemotherapy in the Era of Immune Checkpoint Inhibitors. *Nat. Rev. Clin. Oncol.* 2020, *17*, 725–741. [CrossRef]
- Makhoul, I.; Atiq, M.; Alwbari, A.; Kieber-Emmons, T. Breast Cancer Immunotherapy: An Update. Breast Cancer Basic Clin. Res. 2018, 12, 34. [CrossRef] [PubMed]
- 35. Emens, L.A. Breast Cancer Immunotherapy: Facts and Hopes. Clin. Cancer Res. 2018, 24, 511-520. [CrossRef]
- Stanton, S.E.; Adams, S.; Disis, M.L. Variation in the Incidence and Magnitude of Tumor-Infiltrating Lymphocytes in Breast Cancer Subtypes: A Systematic Review. JAMA Oncol. 2016, 2, 1354–1360. [CrossRef]
- Cimino-Mathews, A.; Thompson, E.; Taube, J.M.; Ye, X.; Lu, Y.; Meeker, A.; Xu, H.; Sharma, R.; Lecksell, K.; Cornish, T.C.; et al. PD-L1 (B7-H1) expression and the immune tumor microenvironment in primary and metastatic breast carcinomas. *Hum. Pathol.* 2016, 47, 52–63. [CrossRef]
- Loi, S.N.; Sirtaine, F.; Piette, R.; Salgado, G.; Viale, F.; Van Eenoo, G.; Rouas, P.; Francis, J.P.; Crown, E.; Hitre, E.; et al. Prognostic and Predictive Value of Tumor-Infiltrating Lymphocytes in a Phase Iii Randomized Adjuvant Breast Cancer Trial in Node-Positive Breast Cancer Com-paring the Addition of Docetaxel to Doxorubicin with Doxorubicin-Based Chemotherapy: Big 02-98. J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol. 2013, 31, 860–867. [CrossRef]
- 39. Savas, P.P.; Salgado, R.; Denkert, C.; Sotiriou, C.; Darcy, P.K.P.; Smyth, M.J.M.; Loi, S. Clinical relevance of host immunity in breast cancer: From TILs to the clinic. *Nat. Rev. Clin. Oncol.* **2016**, *13*, 228–241. [CrossRef]

- Ali, H.R.; Provenzano, E.; Dawson, S.-J.; Blows, F.M.; Liu, B.; Shah, M.; Earl, H.M.; Poole, C.J.; Hiller, L.; Dunn, J.A.; et al. Association between CD8+ T-cell infiltration and breast cancer survival in 12 439 patients. *Ann. Oncol.* 2014, 25, 1536–1543. [CrossRef]
- Burstein, M.D.; Tsimelzon, A.; Poage, G.M.; Covington, K.R.; Contreras, A.; Fuqua, S.A.; Savage, M.I.; Osborne, C.K.; Hilsenbeck, S.G.; Chang, J.C.; et al. Comprehensive Genomic Analysis Identifies Novel Subtypes and Targets of Triple-Negative Breast Cancer. *Clin. Cancer Res.* 2015, *21*, 1688–1698. [CrossRef]
- 42. Denkert, C.; Von Minckwitz, G.; Darb-Esfahani, S.; Lederer, B.; Heppner, B.I.; Weber, K.E.; Budczies, J.; Huober, J.; Klauschen, F.; Furlanetto, J.; et al. Tumour-infiltrating lymphocytes and prognosis in different subtypes of breast cancer: A pooled analysis of 3771 patients treated with neoadjuvant therapy. *Lancet Oncol.* **2018**, *19*, 40–50. [CrossRef]
- Emens, A.L.; Molinero, L.; Loi, S.; Rugo, H.S.; Schneeweiss, A.; Diéras, V.; Iwata, H.; Barrios, C.H.; Nechaeva, M.; Duc, A.N.; et al. Atezolizumab and nab-Paclitaxel in Advanced Triple-Negative Breast Cancer: Biomarker Evaluation of the IMpassion130 Study. J. Natl. Cancer Inst. 2021, 379, 2108–2121. [CrossRef]
- Cortes, J.; Cescon, D.W.; Rugo, H.S.; Nowecki, Z.; Im, S.-A.; Yusof, M.; Gallardo, C.; Lipatov, O.; Barrios, C.H.; Holgado, E.; et al. Pembrolizumab Plus Chemotherapy Versus Placebo Plus Chemotherapy for Previously Untreated Locally Recurrent Inoperable or Metastatic Triple-Negative Breast Cancer (Key-note-355): A Randomised, Placebo-Controlled, Double-Blind, Phase 3 Clinical Trial. *Lancet* 2020, 396, 1817–1828. [CrossRef]
- Thomas, A.; Routh, E.D.; Pullikuth, A.; Jin, G.; Chou, J.W.; Hoadley, K.A.; Print, C.; Knowlton, N.; Black, M.A.; Demaria, S.; et al. Tumor Mutational Burden is a Determinant of Immune-Mediated Survival in Breast Cancer. *Oncoimmunology* 2018, 7, e1490854. [CrossRef] [PubMed]
- Goodman, A.M.; Kato, S.; Bazhenova, L.; Patel, S.P.; Frampton, G.M.; Miller, V.; Stephens, P.J.; Daniels, G.A.; Kurzrock, R. Tumor Mutational Burden as an Independent Predictor of Response to Immunotherapy in Diverse Cancers. *Mol. Cancer Ther.* 2017, 16, 2598–2608. [CrossRef]
- 47. Zou, Y.; Zou, X.; Zheng, S.; Tang, H.; Zhang, L.; Liu, P.; Xie, X. Efficacy and predictive factors of immune checkpoint inhibitors in metastatic breast cancer: A systematic review and meta-analysis. *Ther. Adv. Med. Oncol.* **2020**, *12*. [CrossRef]
- 48. Hegde, P.S.; Chen, D.S. Top 10 Challenges in Cancer Immunotherapy. Immunity 2020, 52, 17–35. [CrossRef]
- 49. Heery, C.R.; O'Sullivan-Coyne, G.; Madan, R.A.; Cordes, L.; Rajan, A.; Rauckhorst, M.; Lamping, E.; Oyelakin, I.; Marté, J.L.; Lepone, L.M.; et al. Avelumab for Meta-static or Locally Advanced Previously Treated Solid Tumours (Javelin Solid Tumor): A Phase 1a, Multicohort, Dose-Escalation Trial. *Lancet Oncol.* **2017**, *18*, 587–598. [CrossRef]
- 50. Schmid, P.; Cruz, C.; Braiteh, F.S.; Eder, J.P.; Tolaney, S.; Kuter, I.; Nanda, R.; Chung, C.; Cassier, P.; Delord, J.P.; et al. Abstract 2986: Atezolizumab in Metastatic Thbc (Mthbc): Long-Term Clinical Outcomes and Biomarker Analyses. *Cancer Res.* 2017, 77, 2986.
- Adams, S.; Schmid, P.; Rugo, H.; Winer, E.; Loirat, D.; Awada, A.; Cescon, D.; Iwata, H.; Campone, M.; Nanda, R.; et al. Pembrolizumab monotherapy for previously treated metastatic triple-negative breast cancer: Cohort A of the phase II KEYNOTE-086 study. Ann. Oncol. 2019, 30, 397–404. [CrossRef] [PubMed]
- 52. Flier, J.S.; Underhill, L.H.; Dvorak, H.F. Tumors: Wounds That Do Not Heal. N. Engl. J. Med. 1986, 315, 1650–1659. [CrossRef]
- 53. Hudson, J.D.; Shoaibi, M.A.; Maestro, R.; Carnero, A.; Hannon, G.J.; Beach, D.H. A Proinflammatory Cytokine Inhibits P53 Tumor Suppressor Activity. J. Exp. Med. 1999, 190, 1375–1382. [CrossRef]
- 54. Cordon-Cardo, C.; Prives, C. At the Crossroads of Inflammation and Tumorigenesis. J. Exp. Med. 1999, 190, 1367–1370. [CrossRef]
- 55. Coussens, L.M.; Tinkle, C.L.; Hanahan, D.; Werb, Z. Mmp-9 Supplied by Bone Mar-row-Derived Cells Contributes to Skin Carcinogenesis. *Cell* **2000**, *103*, 481–490. [CrossRef]
- Coussens, L.M.; Raymond, W.W.; Bergers, G.; Laig-Webster, M.; Behrendtsen, O.; Werb, Z.; Caughey, G.H.; Hanahan, D. Inflammatory mast cells up-regulate angiogenesis during squamous epithelial carcinogenesis. *Genes Dev.* 1999, 13, 1382–1397. [CrossRef]
- 57. Spranger, S.; Gajewski, T.F. Impact of oncogenic pathways on evasion of antitumour immune responses. *Nat. Rev. Cancer* **2018**, *18*, 139–147. [CrossRef]
- 58. Wellenstein, M.D.; De Visser, K.E. Cancer-Cell-Intrinsic Mechanisms Shaping the Tumor Immune Landscape. *Immunity* **2018**, *48*, 399–416. [CrossRef] [PubMed]
- 59. Sparmann, A.; Bar-Sagi, D. Ras-induced interleukin-8 expression plays a critical role in tumor growth and angiogenesis. *Cancer Cell* **2004**, *6*, 447–458. [CrossRef]
- 60. Sumimoto, H.; Imabayashi, F.; Iwata, T.; Kawakami, Y. The Braf-Mapk Signaling Pathway is Essential for Cancer-Immune Evasion in Human Melanoma Cells. *J. Exp. Med.* 2006, 203, 1651–1656. [CrossRef]
- 61. Natali, P.G.; Nicotra, M.R.; Nuti, M.; Bigotti, G.; Calabrò, A.; Schlom, J.; Giacomini, P. Molecular profile, tissue distribution and prognostic evaluation of a human melanoma-carcinoma antigen recognized by the murine monoclonal antibody B1.1. *Int. J. Biol. Mark.* **1988**, *3*, 211–220. [CrossRef]
- 62. Peng, W.; Chen, J.Q.; Liu, C.; Malu, S.; Creasy, C.; Tetzlaff, M.T.; Xu, C.; McKenzie, J.A.; Zhang, C.; Liang, X.; et al. Loss of PTEN Promotes Resistance to T Cell–Mediated Immunotherapy. *Cancer Discov.* **2016**, *6*, 202–216. [CrossRef]
- 63. Goel, S.; DeCristo, M.J.; Watt, A.C.; Brin, J.H.; Sceneay, J.; Li, B.B.; Khan, N.; Ubellacker, J.M.; Xie, S.O.; Metzger-Filho, J.; et al. Cdk4/6 Inhibition Triggers Anti-Tumour Immunity. *Nature* **2017**, *548*, 471–475. [CrossRef]

- 64. Chen, R.-H.; Xiao, Z.-W.; Yan, X.-Q.; Han, P.; Liang, F.-Y.; Wang, J.-Y.; Yu, S.-T.; Zhang, T.-Z.; Chen, S.-Q.; Zhong, Q.; et al. Tumor Cell-Secreted ISG15 Promotes Tumor Cell Migration and Immune Suppression by Inducing the Macrophage M2-Like Phenotype. *Front. Immunol.* **2020**, *11*, 64. [CrossRef]
- Serrels, A.; Lund, T.; Serrels, B.; Byron, A.; McPherson, R.C.; Von Kriegsheim, A.; Gómez-Cuadrado, L.; Canel, M.; Muir, M.; Ring, J.E.; et al. Nuclear FAK Controls Chemokine Transcription, Tregs, and Evasion of Anti-tumor Immunity. *Cell* 2015, *163*, 160–173. [CrossRef]
- 66. Lemmon, M.A.; Schlessinger, J. Cell Signaling by Receptor Tyrosine Kinases. Cell 2010, 141, 1117–1134. [CrossRef] [PubMed]
- 67. Butti, R.; Das, S.; Gunasekaran, V.P.; Yadav, A.S.; Kumar, D.; Kundu, G.C. Receptor tyrosine kinases (RTKs) in breast cancer: Signaling, therapeutic implications and challenges. *Mol. Cancer* **2018**, *17*, 1–18. [CrossRef]
- 68. Myers, K.V.; Amend, S.R.; Pienta, K.J. Targeting Tyro3, Axl and Mertk (Tam Receptors): Implications for Macrophages in the Tumor Microenvironment. *Mol. Cancer* 2019, *18*, 94. [CrossRef] [PubMed]
- 69. Xun, Q.; Wang, Z.; Hu, X.; Ding, K.; Lu, X. Small-Molecule CSF1R Inhibitors as Anticancer Agents. *Curr. Med. Chem.* **2020**, *27*, 3944–3966. [CrossRef]
- Hato, T.; Zhu, A.X.; Duda, D.G. Rationally Combining Anti-Vegf Therapy with Checkpoint Inhibitors in Hepatocellular Carcinoma. *Immunotherapy* 2016, *8*, 299–313. [CrossRef] [PubMed]
- Faham, N.; Welm, A.L. RON Signaling is a Key Mediator of Tumor Progression in Many Human Cancers. *Cold Spring Harb. Symp. Quant. Biol.* 2016, *81*, 177–188. [CrossRef] [PubMed]
- 72. Suh, K.J.; Sung, J.H.; Kim, J.W.; Han, S.-H.; Lee, H.S.; Min, A.; Kang, M.H.; Kim, J.E.; Kim, S.H.; Lee, J.-O.; et al. EGFR or HER2 inhibition modulates the tumor microenvironment by suppression of PD-L1 and cytokines release. *Oncotarget* 2017, *8*, 63901–63910. [CrossRef] [PubMed]
- Akbay, E.A.; Koyama, S.; Carretero, J.; Altabef, A.; Tchaicha, J.H.; Christensen, C.L.; Mikse, O.R.; Cherniack, A.D.; Beauchamp, E.M.; Pugh, T.J.; et al. Activation of the PD-1 Pathway Contributes to Immune Escape in EGFR-Driven Lung Tumors. *Cancer Discov.* 2013, *3*, 1355–1363. [CrossRef]
- 74. Yu, S.; Sha, H.; Qin, X.; Chen, Y.; Li, X.; Shi, M.; Feng, J. Egfr E746-A750 Deletion in Lung Cancer Represses Anti-Tumor Immunity through the Exosome-Mediated Inhibition of Dendritic Cells. *Oncogene* 2020, *39*, 2643–2657. [CrossRef]
- Ludwig, K.F.; Du, W.; Sorrelle, N.B.; Wnuk-Lipinska, K.; Topalovski, M.; Toombs, J.E.; Cruz, V.H.; Yabuuchi, S.; Rajesh Kumar, N.; Maitra, A.; et al. Small-Molecule Inhibition of Axl Targets Tumor Immune Suppression and Enhances Chemotherapy in Pancreatic Cancer. *Cancer Res.* 2018, *78*, 246–255. [CrossRef]
- 76. Ishihara, H.; Sasaoka, T.; Ishiki, M.; Takata, Y.; Imamura, T.; Usui, I.; Langlois, W.J.; Sawa, T.; Kobayashi, M. Functional Importance of Shc Tyrosine 317 on Insulin Signaling in Rat1 Fibroblasts Expressing Insulin Receptors. J. Biol. Chem. 1997, 272, 9581–9586. [CrossRef]
- Sasaoka, T.; Kobayashi, M. The Functional Significance of Shc in Insulin Signaling as a Substrate of the Insulin Receptor. *Endocr. J.* 2000, 47, 373–381. [CrossRef]
- 78. Galvagni, F.; Pennacchini, S.; Salameh, A.; Rocchigiani, M.; Neri, F.; Orlandini, M.; Petraglia, F.; Gotta, S.; Sardone, G.L.; Matteucci, G.; et al. Endothelial Cell Adhesion to the Extracellular Matrix Induces c-Src–Dependent VEGFR-3 Phosphorylation Without the Activation of the Receptor Intrinsic Kinase Activity. *Circ. Res.* 2010, *106*, 1839–1848. [CrossRef] [PubMed]
- Smith, M.J.; Hardy, W.R.; Murphy, J.M.; Jones, N.; Pawson, T. Screening for PTB Domain Binding Partners and LigandSpecificity Using Proteome-Derived NPXY Peptide Arrays. *Mol. Cell. Biol.* 2006, 26, 8461–8474. [CrossRef]
- 80. Foster, B.M.; Zaidi, D.; Young, T.R.; Mobley, M.E.; Kerr, B.A. CD117/c-kit in Cancer Stem Cell-Mediated Progression and Therapeutic Resistance. *Biomedicines* **2018**, *6*, 31. [CrossRef] [PubMed]
- Ha, J.R.; Siegel, P.M.; Ursini-Siegel, J. The Tyrosine Kinome Dictates Breast Cancer Heterogeneity and Therapeutic Responsiveness. J. Cell. Biochem. 2016, 117, 1971–1990. [CrossRef]
- Mishra, J.; Kumar, N. Adapter Protein Shc Regulates Janus Kinase 3 Phosphorylation. J. Biol. Chem. 2014, 289, 15951–15956. [CrossRef] [PubMed]
- 83. Klint, P.; Kanda, S.; Claesson-Welsh, L. Shc and a Novel 89-kDa Component Couple to the Grb2-Sos Complex in Fibroblast Growth Factor-2-stimulated Cells. *J. Biol. Chem.* **1995**, 270, 23337–23344. [CrossRef]
- 84. Ursini-Siegel, J.; Muller, W.J. The ShcA Adaptor Protein is a Critical Regulator of Breast Cancer Progression. *Cell Cycle* 2008, 7, 1936–1943. [CrossRef]
- 85. Ursini-Siegel, J.; Cory, S.; Zuo, D.; Hardy, W.R.; Rexhepaj, E.; Lam, S.; Schade, B.; Jirstrom, K.; Bjur, E.; Piccirillo, C.A.; et al. Receptor Tyrosine Kinase Signaling Favors a Protumorigenic State in Breast Cancer Cells by Inhibiting the Adaptive Immune Response. *Cancer Res.* **2010**, *70*, 7776–7787. [CrossRef]
- 86. Ahn, R.; Sabourin, V.; Bolt, A.M.; Hébert, S.; Totten, S.; De Jay, N.; Festa, M.C.; Young, Y.K.; Im, Y.K.; Pawson, T.; et al. The Shc1 adaptor simultaneously balances Stat1 and Stat3 activity to promote breast cancer immune suppression. *Nat. Commun.* 2017, 8. [CrossRef]
- 87. Kumar, S.; Davra, V.; Obr, A.E.; Geng, K.; Wood, T.L.; De Lorenzo, M.S.; Birge, R.B. Crk adaptor protein promotes PD-L1 expression, EMT and immune evasion in a murine model of triple-negative breast cancer. *Oncolmmunology* **2017**, *7*, e1376155. [CrossRef]
- Kumagai, S.; Koyama, S.; Nishikawa, H. Antitumour immunity regulated by aberrant ERBB family signalling. *Nat. Rev. Cancer* 2021, 1–17. [CrossRef]

- 89. Xin, H.; Zhang, C.; Herrmann, A.; Du, Y.; Figlin, R.; Yu, H. Sunitinib Inhibition of Stat3 Induces Renal Cell Carcinoma Tumor Cell Apoptosis and Reduces Immunosuppressive Cells. *Cancer Res.* **2009**, *69*, 2506–2513. [CrossRef]
- Eyob, H.; Ekiz, H.A.; Derose, Y.S.; Waltz, S.E.; Williams, M.A.; Welm, A.L. Inhibition of Ron Kinase Blocks Conversion of Micrometastases to Overt Metastases by Boosting Antitumor Immunity. *Cancer Discov.* 2013, *3*, 751–760. [CrossRef]
- Hannesdóttir, L.; Tymoszuk, P.; Parajuli, N.; Wasmer, M.-H.; Philipp, S.; Daschil, N.; Datta, S.; Koller, J.-B.; Tripp, C.H.; Stoitzner, P.; et al. Lapatinib and doxorubicin enhance the Stat1-dependent antitumor immune response. *Eur. J. Immunol.* 2013, 43, 2718–2729. [CrossRef] [PubMed]
- 92. Yang, Z.; Tam, K.Y. Combination Strategies Using EGFR-TKi in NSCLC Therapy: Learning from the Gap between Pre-Clinical Results and Clinical Outcomes. *Int. J. Biol. Sci.* 2018, 14, 204–216. [CrossRef] [PubMed]
- Levy, B.; Paz-Ares, L.; Bennouna, J.; Felip, E.; Rodríguez Abreu, D.; Isla, D.; Barlesi, F.; Molinier, O.; Madelaine, J.; Audigier-Valette, C.; et al. Afatinib with Pembrolizumab for Treatment of Patients with Locally Advanced/Metastatic Squamous Cell Carcinoma of the Lung: The Lux-Lung Io/Keynote 497 Study Protocol. *Clin. Lung Cancer* 2019, 20, e407–e412. [CrossRef]
- 94. Atkins, M.B.; Tannir, N.M. Current and emerging therapies for first-line treatment of metastatic clear cell renal cell carcinoma. *Cancer Treat. Rev.* 2018, 70, 127–137. [CrossRef] [PubMed]
- 95. Vanneman, M.; Dranoff, G. Combining immunotherapy and targeted therapies in cancer treatment. *Nat. Rev. Cancer* 2012, 12, 237–251. [CrossRef]
- 96. Zhu, A.X.; Finn, R.S.; Galle, P.R.; Llovet, J.M.; Blanc, J.F.; Okusaka, T.; Chau, I.; Cella, D.; Girvan, A.; Gable, J.; et al. 622pd-Ramucirumab as Second-Line Treatment in Patients with Advanced Hepatocellular Carcinoma (Hcc) and Elevated Alpha-Fetoprotein (Afp) Following First-Line Sorafenib: Patient Reported Outcome Results across Two Phase Iii Studies (Reach-2 and Reach). Ann. Oncol. 2018, 29, viii208. [CrossRef]
- 97. Bruix, J.; Qin, S.; Merle, P.; Granito, A.; Huang, Y.-H.; Bodoky, G.; Pracht, M.; Yokosuka, O.; Rosmorduc, O.; Breder, V.; et al. Regorafenib for patients with hepatocellular carcinoma who progressed on sorafenib treatment (RESORCE): A randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet* 2017, 389, 56–66. [CrossRef]
- 98. Abou-Alfa, G.K.; Meyer, T.; Cheng, A.-L.; El-Khoueiry, A.B.; Rimassa, L.; Ryoo, B.-Y.; Cicin, I.; Merle, P.; Chen, Y.; Park, J.-W.; et al. Cabozantinib in Patients with Advanced and Progressing Hepatocellular Carcinoma. *N. Engl. J. Med.* **2018**, *379*, 54–63. [CrossRef]
- 99. Marino, D.; Zichi, C.; Audisio, M.; Sperti, E.; Di Maio, M. Second-line treatment options in hepatocellular carcinoma. *Drugs Context* **2019**, *8*, 1–13. [CrossRef]
- 100. Ma, B.; Rudin, C.; Cervantes, A.; Dowlati, A.; Costa, D.; Schmid, P.; Heist, R.; Villaflor, V.; Sarkar, I.; Huseni, M.; et al. 4410 Preliminary safety and clinical activity of erlotinib plus atezolizumab from a Phase Ib study in advanced NSCLC. *Ann. Oncol.* 2016, 27. [CrossRef]
- 101. Ahn, M.-J.; Yang, J.; Yu, H.; Saka, H.; Ramalingam, S.; Goto, K.; Kim, S.-W.; Yang, L.; Walding, A.; Oxnard, G. 136O: Osimertinib combined with durvalumab in EGFR-mutant non-small cell lung cancer: Results from the TATTON phase Ib trial. *J. Thorac. Oncol.* 2016, 11, S115. [CrossRef]
- 102. Motzer, R.J.; Penkov, K.; Haanen, J.; Rini, B.; Albiges, L.; Campbell, M.T.; Venugopal, B.; Kollmannsberger, C.; Negrier, S.; Uemura, M. Avelumab plus Axitinib versus Sunitinib for Advanced Renal-Cell Carcinoma. N. Engl. J. Med. 2019, 380, 1103–1115. [CrossRef] [PubMed]
- 103. Pozzi, C.; Cuomo, A.; Spadoni, I.; Magni, E.; Silvola, A.; Conte, A.; Sigismund, S.; Ravenda, P.S.; Bonaldi, T.; Zampino, M.G.; et al. The EGFR-specific antibody cetuximab combined with chemotherapy triggers immunogenic cell death. *Nat. Med.* 2016, 22, 624–631. [CrossRef] [PubMed]
- 104. Muntasell, A.; Cabo, M.; Servitja, S.; Tusquets, I.; Martínez-García, M.; Rovira, A.; Rojo, F.; Albanell, J.; López-Botet, M. Interplay between Natural Killer Cells and Anti-HER2 Antibodies: Perspectives for Breast Cancer Immunotherapy. *Front. Immunol.* 2017, 8, 1544. [CrossRef] [PubMed]
- 105. Concha-Benavente, F.; Ferris, R. Jak2 Inhibition Prevents Nk-Released Ifnγ-Mediated Pd-L1 Upreg-ulation and Enhances Cetuximab Mediated Adcc of Hnc Cells (Tum2p.1014). J. Immunol. 2015, 194, 69.
- 106. Lizotte, P.H.; Hong, R.L.; Luster, T.A.; Cavanaugh, M.E.; Taus, L.J.; Wang, S.; Dhaneshwar, A.; Mayman, N.; Yang, A.; Kulkarni, M.L.; et al. A High-Throughput Immune-Oncology Screen Identifies Egfr Inhibitors as Potent Enhancers of Antigen-Specific Cytotoxic T-Lymphocyte Tumor Cell Killing. *Cancer Immunol. Res.* 2018, *6*, 1511–1523. [CrossRef]
- 107. Liang, H.; Liu, X.; Wang, M. Immunotherapy combined with epidermal growth factor receptor tyrosine kinase inhibitors in non-small-cell lung cancer treatment. *Oncol. Targets Ther.* **2018**, *11*, 6189–6196. [CrossRef]
- 108. Venugopalan, A.; Lee, M.-L.; Niu, G.; Medina-Echeverz, J.; Tomita, Y.; Lizak, M.J.; Cultraro, C.M.; Simpson, R.M.; Chen, X.; Trepel, J.B.; et al. Egfr-Targeted Therapy Results in Dramatic Early Lung Tumor Regression Accompanied by Imaging Response and Immune Infiltration in Egfr Mutant Transgenic Mouse Models. *Oncotarget* 2016, 7, 54137. [CrossRef] [PubMed]
- 109. Jia, Y.; Li, X.; Jiang, T.; Zhao, S.; Zhao, C.; Zhang, L.; Liu, X.; Shi, J.; Qiao, M.; Luo, J.; et al. EGFR-targeted therapy alters the tumor microenvironment in EGFR-driven lung tumors: Implications for combination therapies. *Int. J. Cancer* 2019, 145, 1432–1444. [CrossRef]
- 110. Goetz, M.P.; Toi, M.; Campone, M.; Sohn, J.; Paluch-Shimon, S.; Huober, J.; Park, I.H.; Trédan, O.; Chen, S.-C.; Manso, L.; et al. MONARCH 3: Abemaciclib as Initial Therapy for Advanced Breast Cancer. *J. Clin. Oncol.* **2017**, *35*, 3638–3646. [CrossRef]

- 111. Hurvitz, S.A.; Martin, M.; Press, M.F.; Chan, D.; Fernandez-Abad, M.; Petru, E.; Rostorfer, R.; Guarneri, V.; Huang, C.-S.; Barriga, S.; et al. Potent Cell-Cycle Inhibition and Upregulation of Immune Response with Abemaciclib and Anastrozole in Neomonarch, Phase Ii Neoadjuvant Study in Hr⁺/Her2⁻ Breast Cancer. *Clin. Cancer Res.* 2020, *26*, 566–580. [CrossRef] [PubMed]
- 112. Schaer, D.A.; Beckmann, R.P.; Dempsey, J.A.; Huber, L.; Forest, A.; Amaladas, N.; Li, Y.; Wang, Y.C.; Rasmussen, E.R.; Chin, D.; et al. The Cdk4/6 Inhibitor Abemaciclib Induces a T Cell Inflamed Tumor Microenvironment and Enhances the Efficacy of Pd-L1 Checkpoint Blockade. *Cell Rep.* 2018, 22, 2978–2994. [CrossRef] [PubMed]
- 113. Dowless, M.S.; Lowery, C.D.; Shackleford, T.J.; Renschler, M.; Stephens, J.R.; Flack, R.; Blosser, W.; Gupta, S.; Stewart, J.; Webster, Y.; et al. Abemaciclib is Active in Preclinical Models of Ewing Sarcoma via Multipronged Regulation of Cell Cycle, DNA Methylation, and Interferon Pathway Signaling. *Clin. Cancer Res.* 2018, 24, 6028–6039. [CrossRef]
- 114. Zhang, J.; Bu, X.; Wang, H.; Zhu, Y.; Geng, Y.; Nihira, N.T.; Tan, Y.; Ci, Y.; Wu, F.; Dai, X.; et al. Cyclin D–CDK4 kinase destabilizes PD-L1 via cullin 3–SPOP to control cancer immune surveillance. *Nat. Cell Biol.* **2018**, 553, 91–95. [CrossRef]
- 115. Ebert, P.J.; Cheung, J.; Yang, Y.; McNamara, E.; Hong, R.; Moskalenko, M.; Gould, S.E.; Maecker, H.; Irving, B.A.; Kim, J.M.; et al. MAP Kinase Inhibition Promotes T Cell and Anti-tumor Activity in Combination with PD-L1 Checkpoint Blockade. *Immunity* 2016, 44, 609–621. [CrossRef]
- 116. Sullivan, R.J.; Hamid, O.; Gonzalez, R.; Infante, R.; Patel, M.R.; Hodi, F.S.; Lewis, K.D.; Tawbi, H.A.; Hernandez, G.; Wongchenko, M.J.; et al. Atezolizumab Plus Cobimetinib and Vemurafenib in Braf-Mutated Melanoma Patients. *Nat. Med.* 2019, 25, 929–935. [CrossRef]
- 117. Dummer, R.; Lebbé, C.; Atkinson, V.; Mandalà, M.; Nathan, P.D.; Arance, A.; Richtig, E.; Yamazaki, N.; Robert, C.; Schadendorf, D.; et al. Combined PD-1, BRAF and MEK Inhibition in Advanced Braf-Mutant Melanoma: Safety Run-in and Biomarker Cohorts of COMBI-I. *Nat. Med.* 2020, 26, 1557–1563. [CrossRef] [PubMed]
- 118. Hu-Lieskovan, S.; Mok, S.; Moreno, B.H.; Tsoi, J.; Robert, L.; Goedert, L.; Pinheiro, E.M.; Koya, R.C.; Graeber, T.G.; Comin-Anduix, B.; et al. Improved Antitumor Activity of Immunotherapy with Braf and Mek Inhibitors in Braf (V600e) Melanoma. *Sci. Transl. Med.* 2015, *7*, 279ra41. [CrossRef]
- 119. Liu, L.; Mayes, P.A.; Eastman, S.; Shi, H.; Yadavilli, S.; Zhang, T.; Yang, J.; See-staller-Wehr, L.; Zhang, S.-Y.; Hopson, C.; et al. The Braf and Mek Inhibitors Dabrafenib and Trametinib: Effects on Immune Function and in Combination with Immuno-modulatory Antibodies Targeting PD-1, PD-L1, and CTLA-4. *Clin. Cancer Res.* 2015, *21*, 1639–1651. [CrossRef]
- 120. Reinhard, D.; Ramelyte, E.; Schindler, S.; Thürigen, O.; Levesque, M.P.; Koelblinger, P. Mek Inhibition and Immune Responses in Advanced Melanoma. *Oncoimmunology* **2017**, *6*, e1335843.
- 121. Frederick, D.T.; Piris, A.; Cogdill, A.P.; Cooper, Z.A.; Lezcano, C.; Ferrone, C.R.; Mitra, D.; Boni, A.; Newton, L.P.; Liu, C.; et al. Braf Inhibition is Associated with Enhanced Melanoma Antigen Expression and a More Favorable Tumor Microenvironment in Patients with Metastatic Melanoma. *Clin. Cancer Res.* **2013**, *19*, 1225–1231. [CrossRef]
- 122. Deken, M.A.; Gadiot, J.; Jordanova, E.S.; Lacroix, R.; Van Gool, M.; Kroon, P.; Pineda, C.; Foppen, M.H.G.; Scolyer, R.; Song, J.-Y.; et al. Targeting the MAPK and PI3K pathways in combination with PD1 blockade in melanoma. *Oncoimmunology* 2016, *5*, e1238557. [CrossRef]
- 123. Young, A.; Ngiow, S.F.; Madore, J.; Reinhardt, J.; Landsberg, J.; Chitsazan, A.; Rautela, J.; Bald, T.; Barkauskas, D.S.; Ahern, E.; et al. Targeting Adenosine in BRAF-Mutant Melanoma Reduces Tumor Growth and Metastasis. *Cancer Res.* 2017, 77, 4684–4696. [CrossRef]
- 124. Hoxhaj, G.; Manning, B.D. The PI3K–AKT network at the interface of oncogenic signalling and cancer metabolism. *Nat. Rev. Cancer* 2019, 20, 74–88. [CrossRef]
- 125. Liu, R.; Chen, Y.; Liu, G.; Li, C.; Song, Y.; Cao, Z.; Li, W.; Hu, J.; Lu, C.; Liu, Y. PI3K/AKT pathway as a key link modulates the multidrug resistance of cancers. *Cell Death Dis.* **2020**, *11*, 1–12. [CrossRef]
- 126. Saxton, R.A.; Sabatini, D.M. mTOR Signaling in Growth, Metabolism, and Disease. Cell 2017, 168, 960–976. [CrossRef]
- 127. Chu, J.; Cargnello, M.; Topisirovic, I.; Pelletier, J. Translation Initiation Factors: Reprogramming Protein Synthesis in Cancer. *Trends Cell Biol.* **2016**, *26*, 918–933. [CrossRef]
- 128. O'Donnell, J.S.; Massi, D.; Teng, M.W.L.; Mandala, M. Pi3k-Akt-mTOR Inhibition in Cancer Immunotherapy, Re-dux. *Semin. Cancer Biol.* **2018**, *48*, 91–103. [CrossRef]
- 129. Lastwika, K.J.; Wilson, W., 3rd; Li, Q.K.; Norris, J.; Xu, H.; Ghazarian, S.R.; Kitagawa, H.; Kawabata, S.; Taube, J.M.; Yao, S.; et al. Control of Pd-L1 Expression by Oncogenic Activation of the Akt-mTOR Pathway in Non-Small Cell Lung Cancer. *Cancer Res.* 2016, 76, 227–238. [CrossRef] [PubMed]
- 130. Li, S.; Zhu, M.; Pan, R.; Fang, T.; Cao, Y.-Y.; Chen, S.; Zhao, X.; Lei, C.-Q.; Guo, L.; Chen, Y.; et al. The tumor suppressor PTEN has a critical role in antiviral innate immunity. *Nat. Immunol.* **2016**, *17*, 241–249. [CrossRef]
- Welte, T.; Kim, I.S.; Tian, L.; Gao, X.; Wang, H.; Li, J.; Holdman, X.B.; Herschkowitz, J.I.; Pond, A.; Xie, G.; et al. Erratum: Oncogenic mTOR signalling recruits myeloid-derived suppressor cells to promote tumour initiation. *Nat. Cell Biol.* 2016, 18, 822. [CrossRef] [PubMed]
- 132. Villegas, S.N.; Gombos, R.; García-López, L.; Gutiérrez-Pérez, I.; García-Castillo, J.; Vallejo, D.M.; Da Ros, V.G.; Balles-ta-Illán, E.; Mihály, J.; Dominguez, M. Pi3k/Akt Cooperates with Oncogenic Notch by Inducing Nitric Oxide-Dependent Inflammation. *Cell Rep.* 2018, 22, 2541–2549. [CrossRef]
- 133. Xu, Y.; Poggio, M.; Jin, H.Y.; Shi, Z.; Forester, C.M.; Wang, Y.; Stumpf, C.R.; Xue, L.; Devericks, E.; So, L.; et al. Translation control of the immune checkpoint in cancer and its therapeutic targeting. *Nat. Med.* **2019**, *25*, 301–311. [CrossRef] [PubMed]

- 134. Cerezo, M.; Guemiri, R.; Druillennec, S.; Girault, I.; Malka-Mahieu, H.; Shen, S.; Allard, D.; Martineau, S.; Welsch, C.; Agoussi, S.; et al. Translational Control of Tumor Immune Escape Via the Eif4f-Stat1-Pd-L1 Axis in Mela-noma. *Nat. Med.* 2018, 24, 1877–1886. [CrossRef] [PubMed]
- 135. Marshall, N.A.; Galvin, K.C.; Corcoran, A.-M.B.; Boon, L.; Higgs, R.; Mills, K.H. Immunotherapy with PI3K Inhibitor and Toll-Like Receptor Agonist Induces IFN-γ+IL-17+ Polyfunctional T Cells That Mediate Rejection of Murine Tumors. *Cancer Res.* 2012, 72, 581–591. [CrossRef]
- Yang, J.; Nie, J.; Ma, X.; Wei, Y.; Peng, Y.; Wei, X. Targeting PI3K in cancer: Mechanisms and advances in clinical trials. *Mol. Cancer* 2019, 18, 1–28. [CrossRef]
- 137. Prabhu, S.A.; Moussa, O.; Miller, W.H., Jr.; Del Rincón, S.V. The Mnk1/2-Eif4e Axis as a Potential Therapeutic Target in Melanoma. *Int. J. Mol. Sci.* 2020, *21*, 4055. [CrossRef]
- 138. Lehmann, B.D.; Abramson, V.G.; Sanders, M.E.; Mayer, E.L.; Haddad, T.C.; Nanda, R.; Van Poznak, C.; Storniolo, A.M.; Nangia, J.R.; Gonzalez-Ericsson, P.I.; et al. TBCRC 032 IB/II Multicenter Study: Molecular Insights to AR Antagonist and PI3K Inhibitor Efficacy in Patients with AR+ Metastatic Triple-Negative Breast Cancer. *Clin. Cancer Res.* 2020, *26*, 2111–2123. [CrossRef]
- Joshi, S.; Singh, A.R.; Liu, K.X.; Pham, T.V.; Zulcic, M.; Skola, D.; Chun, H.B.; Glass, C.K.; Morales, G.A.; Garlich, J.R.; et al. SF2523: Dual PI3K/BRD4 Inhibitor Blocks Tumor Immunosuppression and Promotes Adaptive Immune Responses in Cancer. *Mol. Cancer Ther.* 2019, *18*, 1036–1044. [CrossRef]
- 140. Sai, J.; Owens, P.; Novitskiy, S.V.; Hawkins, O.E.; Vilgelm, A.E.; Yang, J.; Sobolik, T.; Lavender, N.; Johnson, A.C.; McClain, C.; et al. PI3K Inhibition Reduces Mammary Tumor Growth and Facilitates Antitumor Immunity and Anti-PD1 Responses. *Clin. Cancer Res.* 2017, 23, 3371–3384. [CrossRef] [PubMed]
- 141. Wang, Y.; Wang, X.Y.; Subjeck, J.R.; Shrikant, P.A.; Kim, H.L. Temsirolimus, an mTOR Inhibitor, Enhances An-ti-Tumour Effects of Heat Shock Protein Cancer Vaccines. *Br. J. Cancer* 2011, *104*, 643–652. [CrossRef]
- 142. Templeton, A.J.; Dutoit, V.; Cathomas, R.; Rothermundt, C.; Bärtschi, D.; Dröge, C.; Gautschi, O.; Borner, M.; Fechter, E.; Stenner, F.; et al. Phase 2 Trial of Single-Agent Everolimus in Chemotherapy-Naive Patients with Castration-Resistant Prostate Cancer (Sakk 08/08). *Eur. Urol.* 2013, *64*, 150–158. [CrossRef]
- 143. Amandine, P.; Papaserafeim, M.; Anke Rietveld, N.L.; Kaestel, C.; Gruaz, L.; Vonarburg, C.; Spirig, R.; Puga Yung, G.L.; Seebach, J.G. Small-Molecule Immunosuppressive Drugs and Therapeutic Immu-noglobulins Differentially Inhibit Nk Cell Effector Functions in Vitro. *Front. Immunol.* 2019, 10, 556.
- 144. Pascual, J.; Berger, S.P.; Witzke, O.; Tedesco, H.; Mulgaonkar, S.; Qazi, Y.; Chadban, S.; Oppenheimer, F.; Sommerer, C.; Oberbauer, R.; et al. Everolimus with Reduced Calcineurin Inhibitor Exposure in Renal Transplantation. *J. Am. Soc. Nephrol.* 2018, 29, 1979–1991. [CrossRef] [PubMed]
- 145. Liu, J.; Liu, D.; Li, J.; Zhu, L.; Zhang, C.; Lei, K.; Xu, Q.; You, R. Efficacy and Safety of Everolimus for Maintenance Immunosuppression of Kidney Transplantation: A Meta-Analysis of Randomized Controlled Trials. *PLoS ONE* 2017, 12, e0170246. [CrossRef]
- 146. Huijts, C.M.; Lougheed, S.M.; Bodalal, Z.; Van Herpen, C.M.; Hamberg, P.; Tascilar, M.; Haanen, J.B.; Verheul, H.M.; De Gruijl, T.D. The effect of everolimus and low-dose cyclophosphamide on immune cell subsets in patients with metastatic renal cell carcinoma: Results from a phase I clinical trial. *Cancer Immunol. Immunother.* 2019, 68, 503–515. [CrossRef]
- 147. Hamilton, E.; Blackwell, K.; Hobeika, A.C.; Clay, T.M.; Broadwater, G.; Ren, X.-R.; Chen, W.; Castro, H.; Lehmann, F.; Spector, N.; et al. Phase I clinical trial of HER2-specific immunotherapy with concomitant HER2 kinase inhibition. *J. Transl. Med.* **2012**, *10*, 28. [CrossRef]
- 148. Chen, G.; Gupta, R.; Petrik, S.; Laiko, M.; Leatherman, J.M.; Asquith, J.M.; Daphtary, M.M.; Garrett-Mayer, E.; Davidson, N.E.; Hirt, K.; et al. A Feasibility Study of Cyclophosphamide, Trastuzumab, and an Allogeneic GM-CSF–Secreting Breast Tumor Vaccine for HER2+ Metastatic Breast Cancer. *Cancer Immunol. Res.* **2014**, *2*, 949–961. [CrossRef]
- 149. Kwilas, A.R.; Ardiani, A.; Donahue, R.N.; Aftab, D.T.; Hodge, J.W. Dual effects of a targeted small-molecule inhibitor (cabozantinib) on immune-mediated killing of tumor cells and immune tumor microenvironment permissiveness when combined with a cancer vaccine. *J. Transl. Med.* **2014**, *12*, 1–15. [CrossRef]
- 150. Sapkota, B.; Hill, C.E.; Pollack, B.P. Vemurafenib Enhances Mhc Induction in Braf (V600e) Homozygous Melanoma Cells. Oncoimmunology **2013**, 2, e22890. [CrossRef]
- 151. Brea, E.J.; Oh, C.Y.; Manchado, E.; Budhu, S.; Gejman, R.S.; Mo, G.; Mondello, P.; Han, J.E.; Jarvis, C.A.; Ulmert, D.; et al. Kinase Regulation of Human MHC Class I Molecule Expression on Cancer Cells. *Cancer Immunol. Res.* **2016**, *4*, 936–947. [CrossRef]
- Stopfer, L.E.; Mesfin, J.M.; Joughin, B.A.; Lauffenburger, D.A.; White, F.M. Multiplexed Relative and Absolute Quantitative Immunopeptidomics Reveals Mhc I Repertoire Alterations Induced by Cdk4/6 Inhibition. *Nat. Commun.* 2020, 11, 2760. [CrossRef] [PubMed]
- 153. Kessler, J.H.; Melief, C.J.M. Identification of T-cell epitopes for cancer immunotherapy. *Leukemia* 2007, 21, 1859–1874. [CrossRef] [PubMed]
- 154. Babon, J.J.; Lucet, I.S.; Murphy, J.M.; Nicola, N.A.; Varghese, L.N. The Molecular Regulation of Janus Kinase (Jak) Activation. *Biochem. J.* 2014, 462, 1–13. [CrossRef]
- 155. Stark, G.R.; Darnell, J.E., Jr. The JAK-STAT Pathway at Twenty. Immunity 2012, 36, 503–514. [CrossRef] [PubMed]
- 156. Ho, H.H.; Ivashkiv, L.B. Role of Stat3 in Type I Interferon Responses. Negative Regulation of Stat1-Dependent Inflammatory Gene Activation. *J. Biol. Chem.* 2006, 281, 14111–14118. [CrossRef] [PubMed]

- 157. Yu, H.; Kortylewski, M.; Pardoll, D. Crosstalk between Cancer and Immune Cells: Role of Stat3 in the Tumour Microenvironment. *Nat. Rev. Immunol.* **2007**, *7*, 41–51. [CrossRef] [PubMed]
- 158. Johnson, D.E.; O'Keefe, R.A.; Grandis, J.R. Targeting the IL-6/JAK/STAT3 signalling axis in cancer. *Nat. Rev. Clin. Oncol.* 2018, 15, 234–248. [CrossRef] [PubMed]
- 159. Meissl, K.; Macho-Maschler, S.; Müller, M.; Strobl, B. The good and the bad faces of STAT1 in solid tumours. *Cytokine* **2017**, *89*, 12–20. [CrossRef] [PubMed]
- 160. Madera, S.; Rapp, M.; Firth, M.A.; Beilke, J.N.; Lanier, L.L.; Sun, J.C. Type I IFN promotes NK cell expansion during viral infection by protecting NK cells against fratricide. *J. Exp. Med.* **2016**, *213*, 225–233. [CrossRef]
- Meissl, K.; Simonović, N.; Amenitsch, L.; Witalisz-Siepracka, A.; Klein, K.; Lassnig, C.; Puga, A.; Vogl, C.; Poelzl, A.; Bosmann, M.; et al. STAT1 Isoforms Differentially Regulate NK Cell Maturation and Anti-tumor Activity. *Front. Immunol.* 2020, *11*, 2189. [CrossRef] [PubMed]
- Kang, Y.H.; Biswas, A.; Field, M.; Snapper, S.B. STAT1 signaling shields T cells from NK cell-mediated cytotoxicity. *Nat. Commun.* 2019, 10, 1–13. [CrossRef]
- 163. Consales, C.; Valentini, E.; Albas, A.; Mendonca, R.; Fuches, R.; Soares, M.; Pereira, C. The preparation of cultured rabies virus and the production of antiserum for human use. *J. Biol. Stand.* **1988**, *16*, 27–32. [CrossRef]
- 164. House, I.G.; Savas, P.; Lai, J.; Chen, A.X.Y.; Oliver, A.J.; Teo, Z.L.; Todd, K.L.; Henderson, M.A.; Giuffrida, L.; Petley, E.V.; et al. Macrophage-Derived CXCL9 and CXCL10 Are Required for Antitumor Immune Responses Following Immune Checkpoint Blockade. *Clin. Cancer Res.* 2020, 26, 487–504. [CrossRef] [PubMed]
- Webb, J.R.; Milne, K.; Kroeger, D.R.; Nelson, B.H. PD-L1 expression is associated with tumor-infiltrating T cells and favorable prognosis in high-grade serous ovarian cancer. *Gynecol. Oncol.* 2016, 141, 293–302. [CrossRef] [PubMed]
- 166. Miranda, A.; Hamilton, P.T.; Zhang, A.W.; Pattnaik, S.; Becht, E.; Mezheyeuski, A.; Bruun, J.; Micke, P.; De Reynies, A.; Nelson, B.H. Cancer stemness, intratumoral heterogeneity, and immune response across cancers. *Proc. Natl. Acad. Sci. USA* 2019, 116, 9020–9029. [CrossRef]
- 167. Zhan, X.; Guo, S.; Li, Y.; Ran, H.; Huang, H.; Mi, L.; Wu, J.; Wang, X.; Xiao, D.; Chen, L.; et al. Glioma Stem-Like Cells Evade Interferon Suppression through Mbd3/Nurd Complex-Mediated Stat1 Downregulation. *J. Exp. Med.* 2020, *4*, e20191340. [CrossRef] [PubMed]
- 168. De Angelis, C.; Fu, X.; Cataldo, M.L.; Nardone, A.; Pereira, R.; Veeraraghavan, J.; Nanda, S.; Qin, L.; Sethunath, V.; Wang, T.; et al. Activation of the IFN Signaling Pathway is Associated with Resistance to Cdk4/6 Inhibitors and Immune Checkpoint Activation in Er-Positive Breast Cancer. *Clin. Cancer Res.* **2021**. [CrossRef]
- 169. Huang, J.; Chen, P.; Liu, K.; Liu, J.; Zhou, B.; Wu, R.; Peng, Q.; Liu, Z.X.; Li, C.; Kroemer, G.; et al. Cdk1/2/5 Inhibition Overcomes Ifng-Mediated Adaptive Immune Resistance in Pancreatic Cancer. *Gut* **2020**. [CrossRef] [PubMed]
- Showalter, L.E.; Oechsle, C.; Ghimirey, N.; Steele, C.; Czerniecki, B.J.; Koski, G.K. Th1 cytokines sensitize HER-expressing breast cancer cells to lapatinib. *PLoS ONE* 2019, 14, e0210209. [CrossRef]
- 171. Chaganty, B.K.R.; Qiu, S.; Gest, A.; Lu, Y.; Ivan, C.; Calin, G.A.; Weiner, L.M.; Fan, Z. Trastuzumab Upregulates Pd-L1 as a Potential Mechanism of Trastuzumab Resistance through Engagement of Immune Effector Cells and Stimulation of Ifnγ Secretion. *Cancer Lett.* 2018, 430, 47–56. [CrossRef]
- 172. Tan, H.-Y.; Wang, N.; Lam, W.; Guo, W.; Feng, Y.; Cheng, Y.-C. Targeting tumour microenvironment by tyrosine kinase inhibitor. *Mol. Cancer* 2018, *17*, 1–15. [CrossRef]
- 173. Yamamoto, N.; Honma, M.; Suzuki, H. Off-Target Serine/Threonine Kinase 10 Inhibition by Erlotinib Enhances Lymphocytic Activity Leading to Severe Skin Disorders. *Mol. Pharm. Ther.* **2011**, *80*, 466–475. [CrossRef] [PubMed]
- 174. Esfahani, K.; Al-Aubodah, T.-A.; Thebault, P.; Lapointe, R.; Hudson, M.; Johnson, N.A.; Baran, D.; Bhulaiga, N.; Takano, T.; Cailhier, J.-F.; et al. Targeting the mTOR pathway uncouples the efficacy and toxicity of PD-1 blockade in renal transplantation. *Nat. Commun.* **2019**, *10*, 1–9. [CrossRef]
- 175. Lu, Q.; Lemke, G. Homeostatic Regulation of the Immune System by Receptor Tyrosine Kinases of the Tyro 3 Family. *Science* 2001, 293, 306–311. [CrossRef] [PubMed]
- 176. Paolino, M.; Choidas, A.; Wallner, S.; Pranjic, B.; Uribesalgo, I.; Loeser, S.; Jamieson, A.M.; Langdon, W.Y.; Ikeda, F.; Fededa, J.P.; et al. The E3 ligase Cbl-b and TAM receptors regulate cancer metastasis via natural killer cells. *Nat. Cell Biol.* 2014, 507, 508–512. [CrossRef] [PubMed]
- 177. Gay, C.M.; Balaji, K.; Byers, L.A. Giving AXL the axe: Targeting AXL in human malignancy. *Br. J. Cancer* 2017, *116*, 415–423. [CrossRef] [PubMed]
- Graham, D.K.; De Ryckere, D.; Davies, K.D.; Earp, H.S. The Tam Family: Phosphatidylserine-Sensing Receptor Tyrosine Kinases Gone Awry in Cancer. Nat. Rev. Cancer 2014, 14, 769–785. [CrossRef]
- 179. Burstyn-Cohen, T.; Maimon, A. TAM receptors, Phosphatidylserine, inflammation, and Cancer. *Cell Commun. Signal.* **2019**, 17, 1–9. [CrossRef]
- 180. Schmid, M.C.; Avraamides, C.J.; Dippold, H.C.; Franco, I.; Foubert, P.; Ellies, L.G.; Acevedo, L.M.; Manglicmot, J.R.; Song, X.; Wrasidlo, W.; et al. Receptor Tyrosine Kinases and Tlr/Il1rs Unexpectedly Activate Myeloid Cell Pi3kγ, a Single Convergent Point Promoting Tumor Inflammation and Progression. *Cancer Cell* **2011**, *19*, 715–727. [CrossRef]
- 181. Kaneda, M.M.; Messer, K.S.; Ralainirina, N.; Li, H.; Leem, C.J.; Gorjestani, S.; Woo, G.; Nguyen, A.V.; Figueiredo, C.C.; Foubert, P.; et al. PI3Kγ is a molecular switch that controls immune suppression. *Nature* **2016**, *539*, 437–442. [CrossRef] [PubMed]

- 182. Bartish, M.; Tong, D.; Pan, Y.; Wallerius, M.; Liu, H.; Ristau, J.; Ferreira, S.D.S.; Wallmann, T.; Van Hoef, V.; Masvidal, L.; et al. MNK2 governs the macrophage antiinflammatory phenotype. *Proc. Natl. Acad. Sci. USA* 2020, 117, 27556–27565. [CrossRef] [PubMed]
- Liu, G.; Jin, Z.; Lu, X. Differential Targeting of Gr-MDSCs, T Cells and Prostate Cancer Cells by Dactolisib and Dasatinib. *Int. J. Mol. Sci.* 2020, 21, 2337. [CrossRef] [PubMed]
- 184. Lu, X. 2025 Targeting immunosuppressive myeloid cells to enhance cancer immunotherapy. J. Clin. Transl. Sci. 2018, 2, 29. [CrossRef]
- 185. Dumas, A.A.; Pomella, N.; Rosser, G.; Guglielmi, L.; Vinel, C.; Millner, T.O.; Rees, J.; Aley, N.; Sheer, D.; Wei, J.; et al. Microglia Promote Glioblastoma Via mTOR-Mediated Immunosuppression of the Tumour Microenvironment. *EMBO J.* 2020, 39, e103790. [CrossRef]
- 186. Chapman, N.M.; Zeng, H.; Nguyen, T.M.; Wang, Y.; Vogel, P.; Dhungana, Y.; Liu, X.; Neale, G.; Locasale, J.W.; Chi, H. mTOR Coordinates Transcriptional Programs and Mitochondrial Metabolism of Activated T (Reg) Subsets to Protect Tissue Homeo-stasis. *Nat. Commun.* 2018, 9, 2095. [CrossRef] [PubMed]
- 187. Shi, L.; Chen, X.; Zang, A.; Li, T.; Hu, Y.; Ma, S.; Lü, M.; Yin, H.; Wang, H.; Zhang, X.; et al. TSC1/mTOR-controlled metabolicepigenetic crosstalk underpins DC control of CD8+ T-cell homeostasis. *PLoS Biol.* 2019, 17, e3000420. [CrossRef] [PubMed]
- 188. Polk, A.; Svane, I.-M.; Andersson, M.; Nielsen, D. Checkpoint inhibitors in breast cancer—Current status. *Cancer Treat. Rev.* 2018, 63, 122–134. [CrossRef]
- 189. Wang, F.; Meng, M.; Mo, B.; Yang, Y.; Ji, Y.; Huang, P.; Lai, W.; Pan, X.; You, T.; Luo, H.; et al. Crosstalks between mTORC1 and mTORC2 variagate cytokine signaling to control NK maturation and effector function. *Nat. Commun.* **2018**, *9*, 1–17. [CrossRef]
- 190. Katholnig, K.; Schütz, B.; Fritsch, S.D.; Schörghofer, D.; Linke, M.; Sukhbaatar, N.; Matschinger, J.M.; Unterleuthner, D.; Hirtl, M.; Lang, M.; et al. Inactivation of mTORC2 in macrophages is a signature of colorectal cancer that promotes tumorigenesis. *JCI Insight* 2019, 4. [CrossRef]