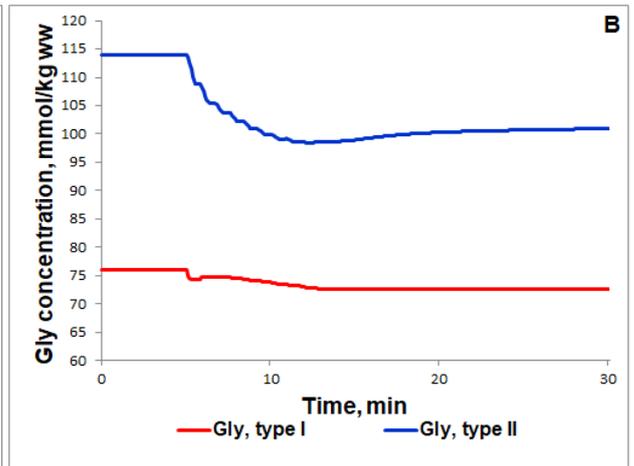
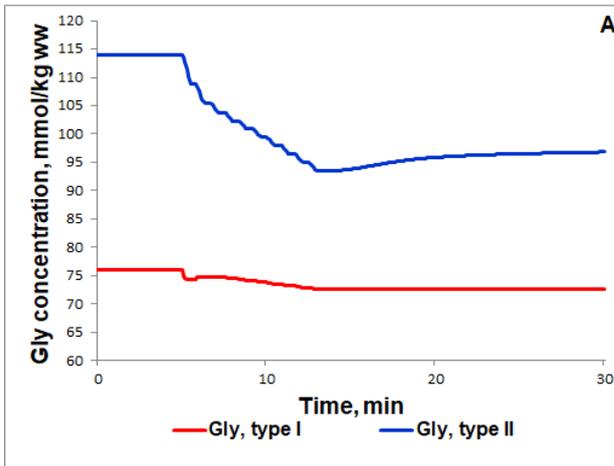
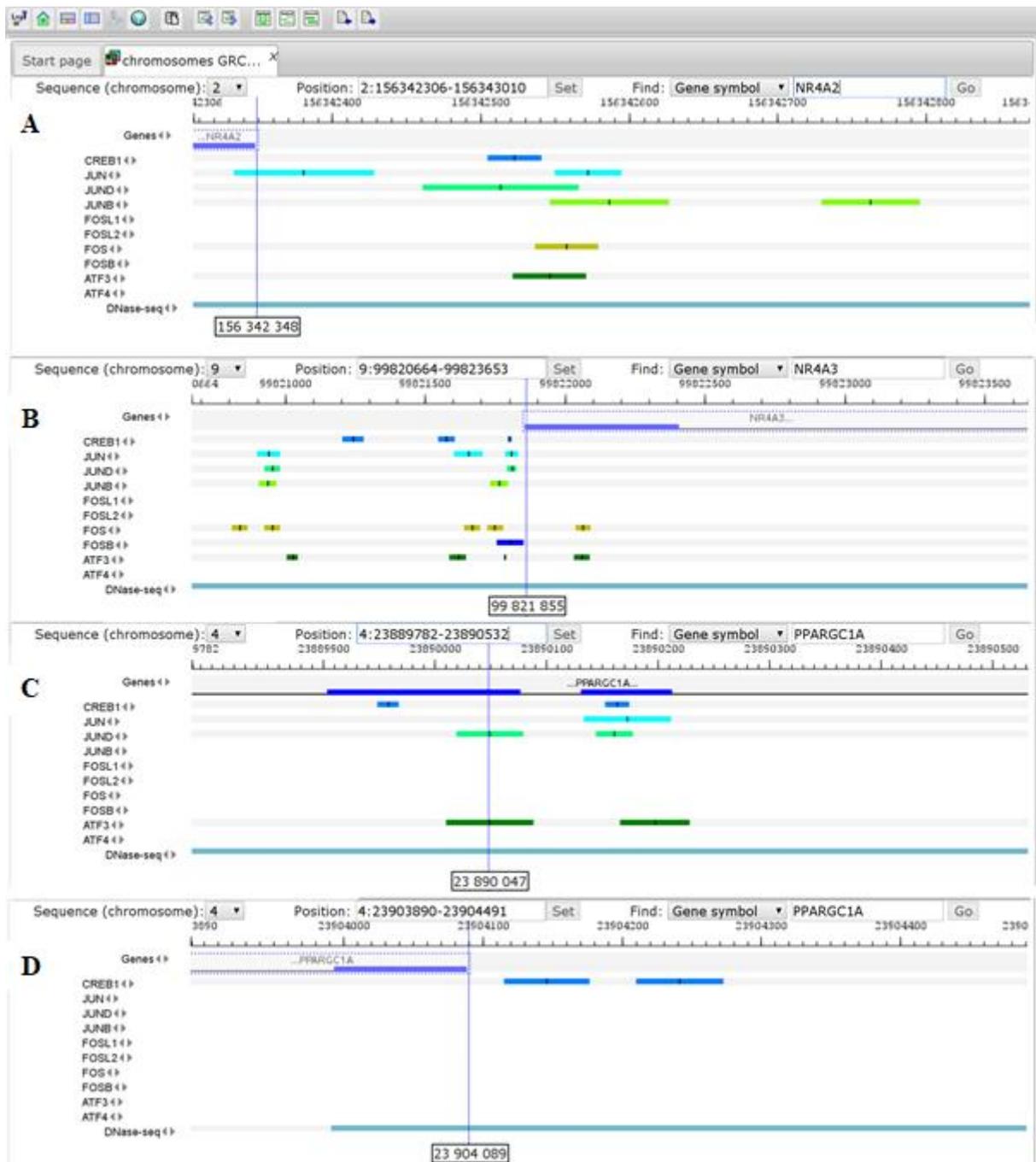


Supplementary Figure S1. Model validation. The model simulations were compared to experimental data. (A) Model-predicted dynamic changes of PCr concentration in the muscle during incremental (blue line), low (green line) and moderate intensity (red line) exercises (axis X - time of the exercise in minutes) and experimental measurements of PCr concentration changes during incremental (blue square dots, [1]), low-intensity (green diamond dots, [2]), green circle dots, [3]), moderate intensity (red square dots, [4]) and high-intensity (magenta square dots, [3]) exercises; (B) Model-predicted dynamic changes of lactate concentration in the muscle during low (green line) and moderate intensity (red line) exercises (axis X - time of the exercise in minutes) and experimental measurements of lactate concentration changes during low-intensity (green square dots, [2]) exercise; (C) Model-predicted dynamic changes of ATP concentration in the muscle during low (green line) and moderate intensity (red line) exercises (axis X - time of the exercise in minutes) and experimental measurements of ATP concentration changes during low-intensity exercise (green square dots, [2]) and all-out exercise (magenta square dots, [5]). The set of experimental data for ATP demonstrates lower [2] and upper [5] boundaries for the concentration; (D) Model-predicted dynamic changes of pH in the muscle during low (green line) and moderate intensity (red line) exercises (axis X - time of the exercise in minutes) and experimental measurements of ATP concentration changes during low-intensity (green square dots, [2]) exercise. Experimental data are shown as mean \pm SD.



Supplementary Figure S2. Simulation results for high-intensity intermittent exercise (each bout of 30-s exercise separated by 20-s recovery, [6]). (F,G) Glycogen concentration in type I (red) and type II fibers (blue) with constant power generated by type II fibers (Figure 10A) and with successive decline of the power generated by type II fibers (Figure 10C), correspondingly.



Supplementary Figure S3. Binding sites of transcription factors of CREB, FOS and JUN families in promoter loci of the target genes *NR4A2*, *NR4A3* and *PPARGC1A*, known from ChIP-seq experiments, as seen in the GTRD genome browser [7]. Transcription start sites are shown with a vertical dashed line; the exact coordinate of each transcription start is given in a box below: A. Promoter region of the *NR4A2* gene (ENSG00000153234). B. Promoter region of the *NR4A3* gene (ENSG00000119508). C. The region of the canonical promoter of the *PPARGC1A* gene (transcript RefSeq ID NM_013261, the coordinate of the transcription start is given according to the NCBI annotation, version 109.20190905). D. The region of the alternative promoter of the *PPARGC1A* gene (transcript RefSeq ID XM_005248132, the coordinate of the transcription start is given according to the NCBI annotation, version 109.20190905)

Supplementary Table S1. Binding sites of transcription factors CREB, FOS and JUN in promoter regions of the target genes *NR4A2*, *NR4A3* and *PPARGC1A*, known from ChIP-seq experiments (data from the GTRD database).

Transcription factor (TF)	Target gene	Transcription start site (TSS)	TF site location, relative to TSS	Reference
pCREB1	<i>NR4A2</i> (<i>ENSG00000153234</i>)	chr2:156342348	-175	[7]
	<i>NR4A3</i> (<i>ENSG00000119508</i>)	chr9:99821855	612	[7]
			-279	
	<i>PPARGC1A</i> * (<i>XM_005248132</i>)	chr4:23904089	58	[7]
-153				
<i>PPARGC1A</i> # (<i>NM_013261</i>)	chr4:23890047	89	[7]	
		-117		
JUN	<i>NR4A2</i>	chr2:156342348	-224	[7]
			-33	
	<i>NR4A3</i>	chr9:99821855	915	[7]
			-200	
<i>PPARGC1A</i> *	chr4:23904089			
<i>PPARGC1A</i> #	chr4:23890047	-126	[7]	
JUND	<i>NR4A2</i>	chr2:156342348	-166	[7]
			-901	[7]
	<i>NR4A3</i>	chr9:99821855	-45	
<i>PPARGC1A</i> *	chr4:23904089			
<i>PPARGC1A</i> #	chr4:23890047	-3	[7]	
		-114		
JUNB	<i>NR4A2</i>	chr2:156342348	-239	[7]
			-415	
	<i>NR4A3</i>	chr9:99821855	-917	[7]
			-90	
<i>PPARGC1A</i> *	chr4:23904089			
<i>PPARGC1A</i> #	chr4:23890047			
FOSL1	<i>NR4A2</i>	chr2:156342348		
	<i>NR4A3</i>	chr9:99821855		
	<i>PPARGC1A</i> *	chr4:23904089		
	<i>PPARGC1A</i> #	chr4:23890047		
FOSL2	<i>NR4A2</i>	chr2:156342348		
	<i>NR4A3</i>	chr9:99821855		
	<i>PPARGC1A</i> *	chr4:23904089		
	<i>PPARGC1A</i> #	chr4:23890047		
FOS	<i>NR4A2</i>	chr2:156342348	-210	[7]
	<i>NR4A3</i>	chr9:99821855	-1017	[7]
-901				

			-188	
			-105	
			-210	
	<i>PPARGC1A</i> *	chr4:23904089		
	<i>PPARGC1A</i> #	chr4:23890047		
FOSB	<i>NR4A2</i>	chr2:156342348		
	<i>NR4A3</i>	chr9:99821855	50	[7]
	<i>PPARGC1A</i> *	chr4:23904089		
	<i>PPARGC1A</i> #	chr4:23890047		
ATF3	<i>NR4A2</i>	chr2:156342348	-198	[7]
	<i>NR4A3</i>	chr9:99821855	-828	[7]
			-237	
			-69	
			-205	
	<i>PPARGC1A</i> *	chr4:23904089		
<i>PPARGC1A</i> #	chr2:156342348	-3	[7]	
		-151		
ATF4	<i>NR4A2</i>	chr2:156342348		
	<i>NR4A3</i>	chr9:99821855		
	<i>PPARGC1A</i> *	chr4:23904089		
	<i>PPARGC1A</i> #	chr4:23890047		

* – alternative promoter of the *PPARGC1A*; # – canonical promoter of the *PPARGC1A*.

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