Table S1. Bacterial strains, plasmids and primers used in this study.

Name	Description	Origin
Strain		
Pectobacterium atrosepticum SCRI1043 (*Pba*)	Wild type	[1]
Pectobacterium atrosepticum SCRI1043*ΔentA*	Mutant strain of SCRI1043 containing the kanamycin cassette in the chromosome; *Km^R*	This study
Pectobacterium atrosepticum SCRI1043comp*ΔentA*	Mutant strain of SCRI1043 containing the kanamycin cassette in the chromosome and the entA complementation construct on pGEM:entA; *Amp^R*	This study
Escherichia coli cc118	Host for suicidal vector pKNG101; *Δ(ara, leu) araD ΔlacX 74 galE galK PhoA20 thi-1 rpsE rpoB argE (am) recA1, Sm^R*	[2]
Escherichia coli HH26/pNJ5000	Mobilizing strain for conjugative transfer of the suicide vector pKNG101 into *Pba* cells; *tra+; Tet^R*	[3]
Escherichia coli NovaBlue	*endA 1hsdR17 (rK12–mK12+) supE44 thi-1 recA1 gyrA96 relA1 lacF' [proA+ B+ lacIqZ ΔM15::Tn10 (Tet^R)]*	Novagen
Plasmids		
pGEM-T Easy	Linearized vector for cloning; *f1 ori Amp^R lacZ*	Promega
pGEM:*entA*	*f1 ori Amp^R lacZ entA*	This study
pGEM:*ΔentA*;Km^R	*f1 ori Amp^R lacZ Km^R*	This study
pKD4	Matrix for PCR amplification of kanamycin resistance cassette; *oriRγ rgnB bla Km^R*	[4]
pKNG101	Suicide mobilized vector for inactivation of target genes; *pir-ori R6K mobRK2 sacB Sm^R*	[5]
pKNG101:*ΔentA*;Km^R	Suicide plasmid carrying mutant locus *ΔentA*;Km^R; *Km^R SmR sacB*	This study
pGEM:*entA;* complementation construct	*f1 ori Amp^R lacZ entA*	This study

Primers	
Primer name	**Primer sequence 5'—3'**
Primers for mutagenesis	
up*entA*F	GCTGCGTACCGATGAAATGC
dn*entA*R	CAATGTCTTTCTCGCCGCTG
dn*entA*KmF	CCATGTCAGCCGTTAAGGGATGGCGGGGCAACGCTGACGGCCTGATTTG
up*entA*KmR	CAGCTCCAGCCTACACAATCGAGGTTGTGCCTTGTTCATCATTTTC
Km*entA*F	GATGAACAAGGCACAACCTCGATTGTGTAGGCTGGAGCTGCTTC
Km*entA*R	GCCGTCAGCGTTGCCCCGCCATCCCTTAACGGCTGACATGGGAATTAGC

Check*entA*F	CGAAAGGCGTGGAGATCGGC
Check*entA*R	GCCAGCGGGGAGCCAATC
Primers for complementation construct	
promF	GCTTTCATCCATGACAGGCTCACATTTAG
prom*entA*R	TGTGCCTTGTTCATCATAAGACAGCTCCTGCGCTCGCAAA
prom*entA*F	TTTGCGAGCGCAGGAGCTGTCTTATGATGAACAAGGCACA
entA*term*R	GATGCTGCCCTTTATCACTGTTCAGGCCGTCAGCGTTGCC
entA*term*F	GGCAACGCTGACGGCCTGAACAGTGATAAAGGGCAGCATC
termR	CCTAAGCCTGCTATCTCCTCAGCATAATTTC
Primers for qRT-PCR	
NtAOCF	CGGCTCAACTGATTCCTCTACAAC
NtAOCR	GTTTGTTGCTAAAGGGGACAAGATC
NtLOXF	CACGCATAATGAGTTTGATAGTTTTG
NtLOXR	CTGAGGAGTTGGGTACTTAAATAGGC
NtEFF	GCCCAACACTTCTTGATGCTC
NtEFR	GACACCAGTTTCCACACGACC
NtATPF	GGTCGATGGCTTGGGAGTACC
NtATPR	GCACAGATTTACGTTCAATAATACCAG

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