

Lack of GABARAP-type proteins is accompanied by altered Golgi morphology and surfaceome composition

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Supplementary Methods are related to Figures S1, S4

Supplementary Figure S1 is related to Figure 1

Supplementary Figure S2 is related to Figure 1

Supplementary Figure S3 is related to Figure 1

Supplementary Figure S4 is related to Figure 3

Supplementary Figure S5 is related to Figure S4

Supplementary Table S1 is related to Figure 3B

Supplementary Methods

Live-cell imaging with fluorescently labelled ceramide

HEK293 Flp-In T-REx cells (3×10^5) were seeded into fibronectin coated 35 mm imaging dishes (Cat. No. 81158, ibidi) and cultured for 24 h in DMEM supplemented with 10 % FCS. Staining of the cells (WT and as GABARAP(s) SKO, DKO, or TKO) with BODIPY-FL C5-ceramide (Cat. No. B-22650, Life Technologies) was conducted according to the manufacturer's instructions. Briefly, cells were rinsed in HBSS and incubated for 30 min at 4 °C with 5 μ M BODIPY-FL C5-ceramide. Then, the cells were rinsed three times in ice-cold HBSS and incubated 30 min in phenol red-free DMEM supplemented with 10 % FCS. Finally, the cells were rinsed once in HBSS and stored in phenol red-free DMEM supplemented with 10 % FCS. Cells were incubated with Hoechst 33342 (Cat. No. R37605, Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions for nuclei staining. BODIPY-FL C5-ceramide was visualised by LSM using the 488 nm channel (MBS 488).

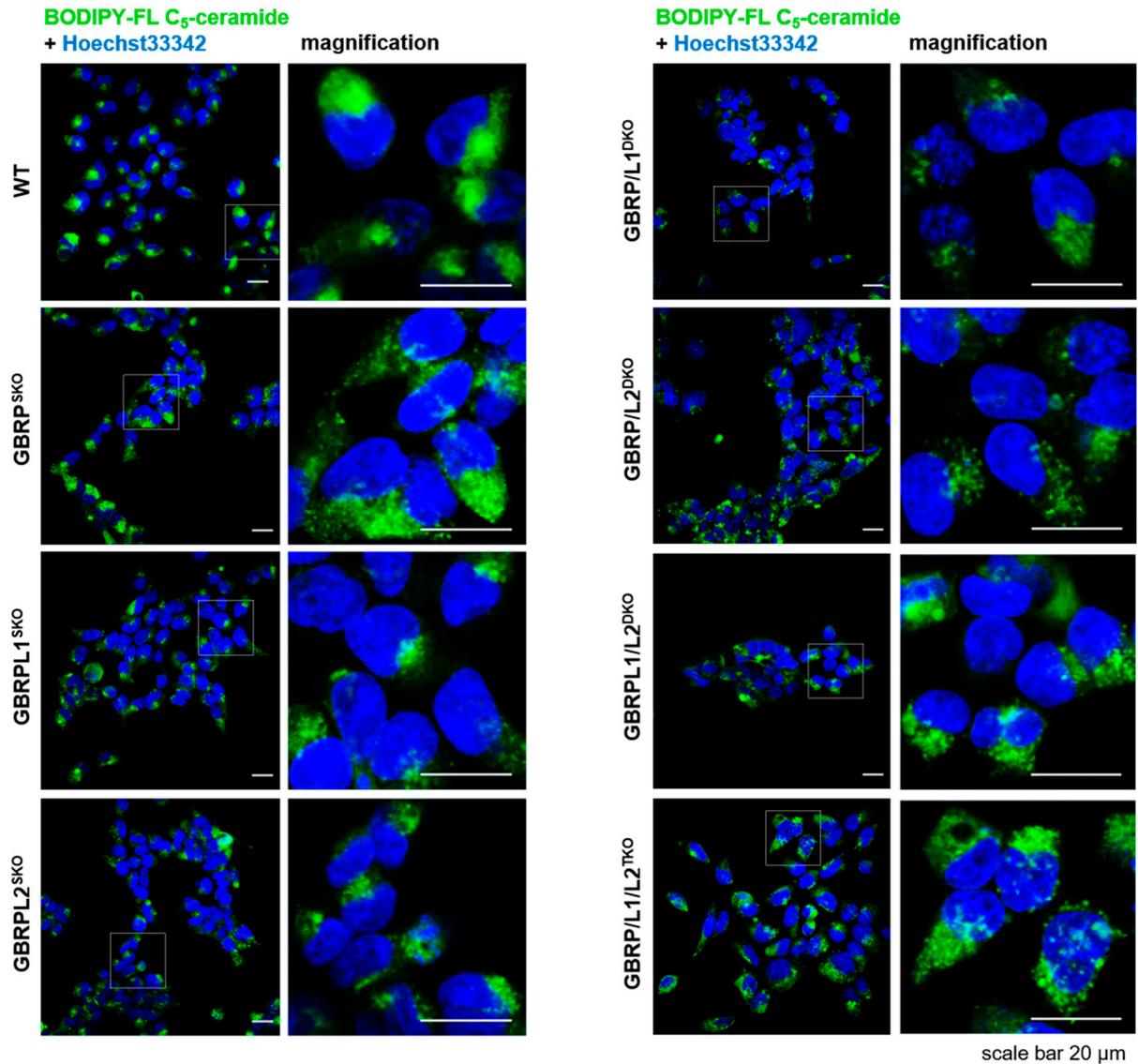
Fluorescence-activated cell sorting

Surface levels of HLA-A were analysed under steady-state conditions in HEK293 WT and GABARAP/L1/L2^{TKO} cells. In brief, 1×10^6 cells of 90 % confluent cells were washed twice with ice-cold PBS and resuspended in 100 μ l ice-cold FACS buffer (2 mM EGTA, 1 % FBS, 25 mM HEPES in PBS) containing 1:20 diluted HLA-A3 antibody conjugated to PE (ThermoFisher Scientific, 12-5754-42) or APC (ThermoFisher Scientific, 17-5754-42). Isotype controls (ThermoFisher Scientific, PE: Mouse IgG2a K, 12-4724-81, APC: Mouse IgG2a K, 17-4724-81) After 30 min incubation on ice in the dark, cells were washed thrice with ice-cold FACS buffer, resuspended in 0.5 ml FACS buffer and analysed by flow cytometry (Aria III, BD Bioscience, Franklin Lakes, USA). Cells were gated according to their size (SSC-A \times FSC-A) and being single cells (FSC-A \times FSC-H). Median fluorescence intensity of GABARAP/L1/L2^{TKO} cells was calculated relative to HEK293 WT intensity. Statistical significance was inferred as calculated by Welch's t-test using GraphPad Prism (version 8).

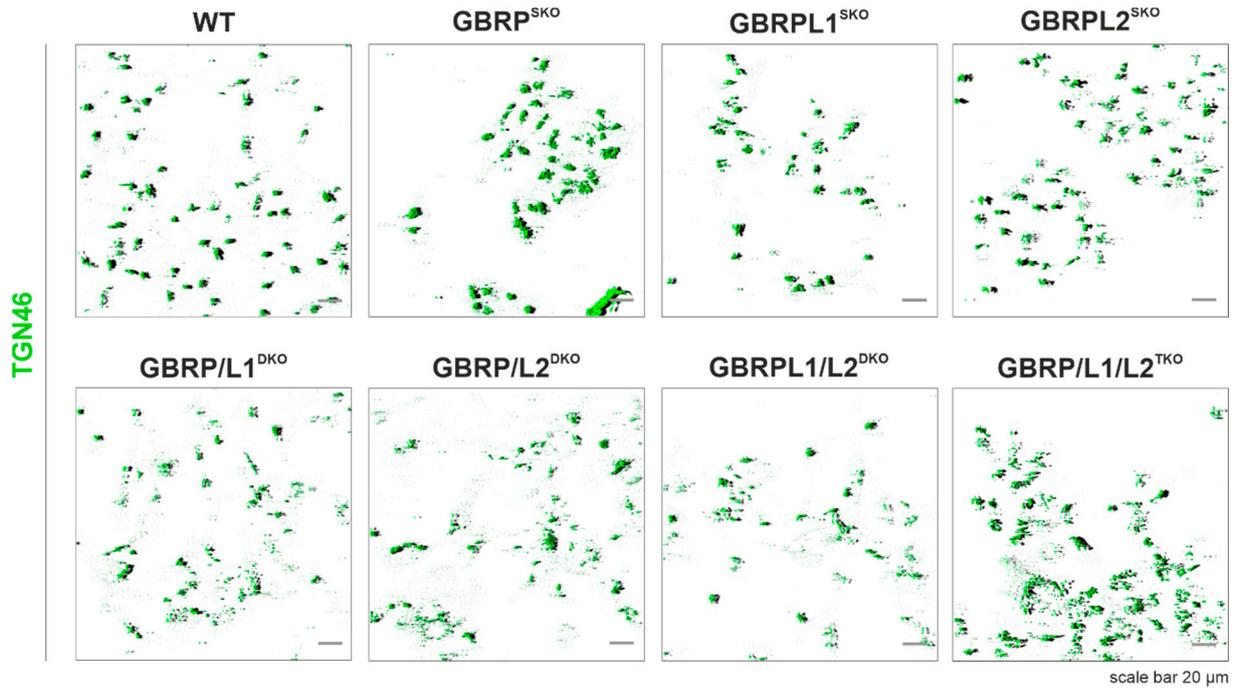
Immunoblotting

For proof-of-method immunoblotting, 10 μ g of surface-enriched proteins were diluted with 4 \times Lämmli's buffer (250 mM Tris-HCl pH 6.8, 40 % glycerol, 5 % SDS, 0.005 % bromophenol blue) containing 8 % fresh 2-mercaptoethanol. After samples were boiled for 5 min at 95 °C, surface fraction lysates were applied on 8 % SDS-PAA gel. After gel electrophoresis, semi-dry blotting of proteins onto 0.4 μ m polyvinylidene fluoride membrane was performed for 1 h 30 min at 77 mA constant current. Unspecific binding sites were blocked for 1 h at RT with 5 % BSA in TBS-T (TBS, 0.1 % Tween-20) and membrane incubated with primary antibody for TFRC (#13208, Cell Signaling Technologies, Danvers, USA) at 1:1000 dilution overnight at 4 °C. After washing (three times with TBS-T) and incubation with 1:5000 diluted fluorescently labelled secondary antibody (ab150083, abcam) for 1 h at RT, target protein was

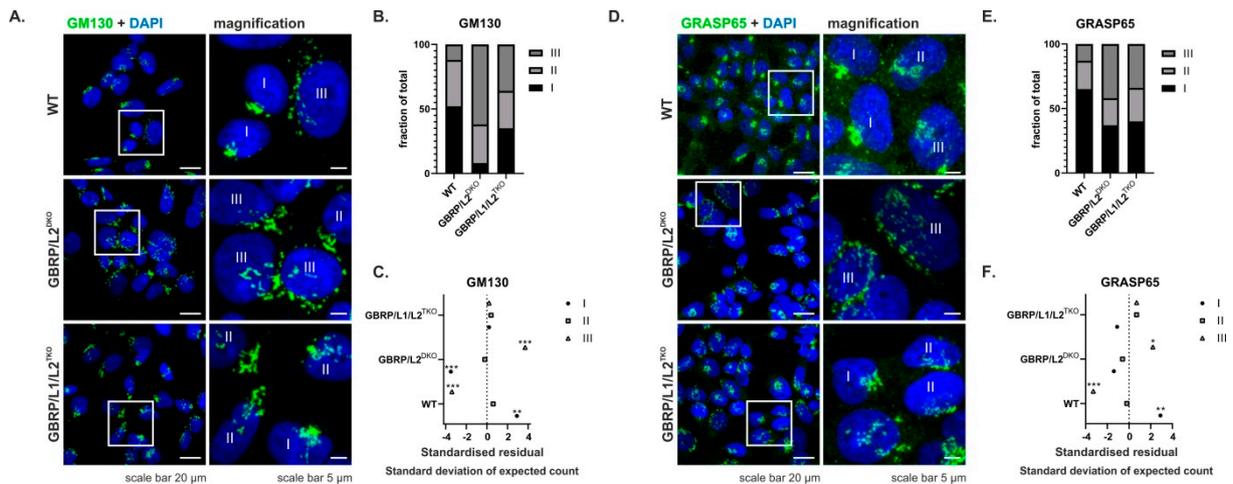
visualised using BioRad Imager. Statistical significance was inferred as calculated by Student's t-test using GraphPad Prism (version 8).



Supplementary Figure S1. Loss of GABARAP leads to a redistribution of Golgi-localised ceramide. HEK^{WT} (WT) or HEK cells with deficiency for one (SKO), two (DKO), or all three (TKO) of the GABARAPs (GBRPs) were cultured for 24 h at 37 °C and 5 % CO₂ in fibronectin coated 35 mm imaging dishes, stained with BODIPY-FL C₅-ceramide according to the manufacturer's instructions (Cat. No. B-22650, Life Technologies), and visualised by confocal fluorescence microscopy. Nuclei were counterstained with Hoechst 33342. Cells were recorded as z-stacks and the slices of each stack were combined in ImageJ by applying the function „SUMSLICES“. For each condition, a representative image of five frames is shown. Scale bar, 20 μm.

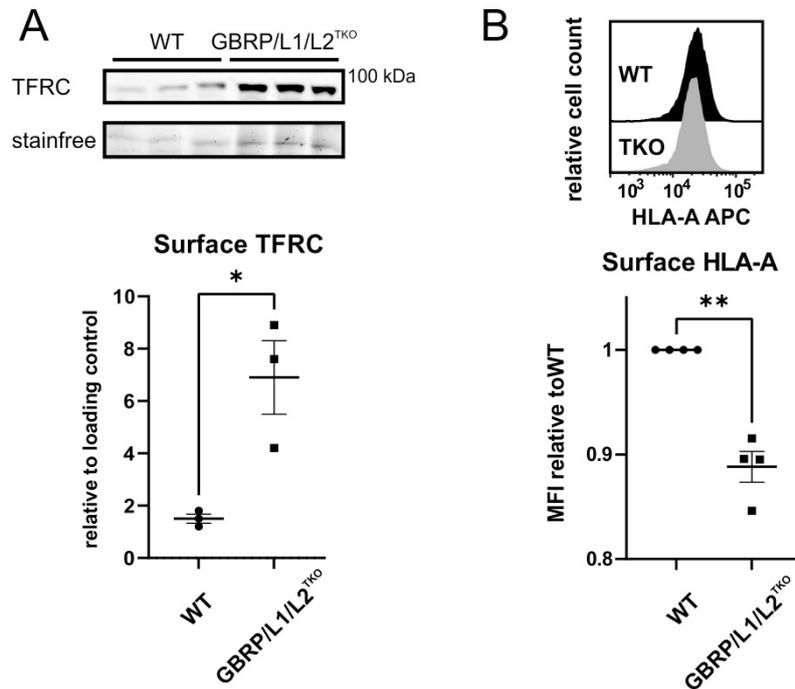


Supplementary Figure S2. 3D visualisation of the *trans*-Golgi morphology in HEK293 WT cells and under various GABARAP-type protein deficiencies. Individual planes of each stack were displayed as 3D image in ZEN 2.3 SP1 FP1 (black edition). The reconstructions relate to the respective images in Figure 1A. Scale bar, 20 μm .

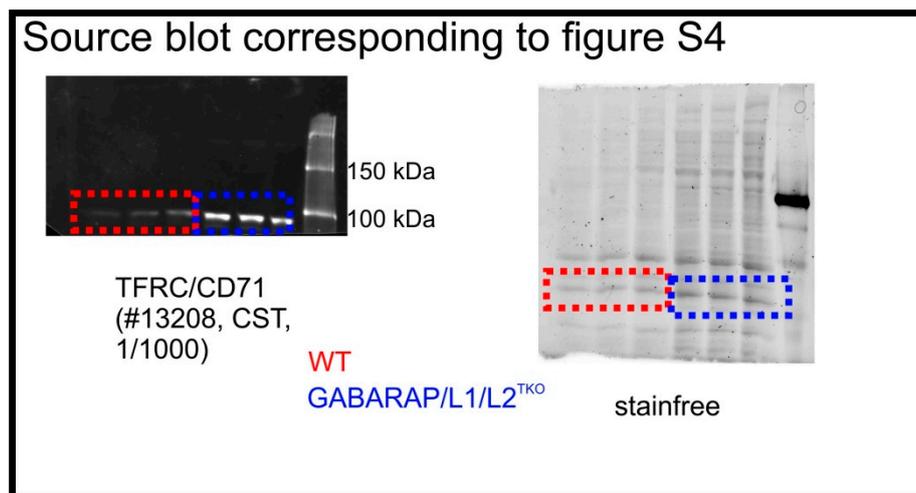


Supplementary Figure S3. Influence of GABARAP-type protein deficiency on *cis*-Golgi morphology. (A) WT, GABARAP/L2^{DKO} or GABARAP/L1/L2^{TKO} cells were fixed (4 % PFA), immunolabelled with anti-human GM130 antibody, and visualised by confocal fluorescence microscopy. Nuclei were counterstained with DAPI. Scale bar total = 20 μm , scale bar zoom-in = 5 μm . (B) Distribution of percentage of cells per cell type assigned to Golgi structure category I (compact), II (partly compact), and III (dispersed) according to GM130 staining. (C) Standardised residual distribution of analysed genotypes. Per cell type, in total ≥ 63 cells from three individual experiments were analysed and categorised by visual judgement. Asterisks indicate significant differences from the mean based on the standardised residual distribution with: $|z| \geq 2.58$ ** ($p \leq 0.01$), $|z| \geq 3.29$ *** ($p \leq 0.001$). (D) WT, GABARAP/L2^{DKO} or GABARAP/L1/L2^{TKO} cells were fixed (4 % PFA), immunolabelled with anti-human GRASP65 antibody, and visualised by confocal fluorescence microscopy. Nuclei were counterstained with DAPI. Scale bar total = 20 μm , scale bar zoom-in = 5 μm . (E) Distribution of percentage of cells per cell type assigned to Golgi structure category I (compact), II (partly compact), and III (dispersed) according to GRASP65 staining. (F) Standardised residual distribution of analysed genotypes. Per cell type, in total ≥ 103 cells were analysed and categorised by visual judgement. Asterisks indicate

significant differences from the mean based on the standardised residual distribution with: $|z| \geq 1.96$ * ($p \leq 0.05$), $|z| \geq 2.58$ ** ($p \leq 0.01$), $|z| \geq 3.29$ *** ($p \leq 0.001$).



Supplementary Figure S4. Validation of surface proteome mass-spectrometry analysis by two independent methods. (A) Cell surface protein-enriched fractions were analysed by immunoblot. Surface levels of TFRC are shown for three independent experiments. Asterisk marks significant differences between GABARAP/L1/L2^{TKO} and WT cells as calculated using independent t-test. $p \leq 0.05$ = *. (B) Surface levels of MHC-I subtype HLA-A were analysed by fluorescence-activated cell sorting. Representative histograms of four independent experiments of single cells stained with HLA-A antibody are shown. Significant differences of median fluorescence intensities (MFI) between WT and GABARAP/L1/L2^{TKO} cells are marked by asterisk as calculated by Welch's t-test. $p \leq 0.01$ = **. GBRP = GABARAP. APC = Allophycocyanin.



Supplementary Figure S5. Source blot corresponding to Figure S4. Regions used for display and analysis are indicated.

- 1 **Supplementary Table S1.** Raw data of mass-spectrometry analysis of surface-enriched proteomes of HEK293 WT
- 2 and GABARAP/L1/L2^{TKO} cells can be found in the attached excel file Table S1 Mass-spectrometry data set.

Supplementary Table S2. Significantly different expressed surface-annotated proteins. For consistency, proteins are sorted as depicted in Figure 3B. Respective raw data are given in Supplementary Table S1.

| GENE SYMBOL | GENE NAME | GENE ID | UNIPROT | CSPA CONFIDENCE LEVEL | DIFFERENCE TKO-WT |
|--------------------|--|----------------|----------------|------------------------------|--------------------------|
| PODXL2 | podocalyxin like 2 | 50512 | Q9NZ53 | 1 | 1.95089 |
| CNNM4 | cyclin and CBS domain divalent metal cation transport mediator 4 | 26504 | Q6P4Q7 | 1 | 0.801802 |
| ITGA7 | integrin subunit alpha 7 | 3679 | Q13683 | 1 | 0.516933 |
| ADAM15 | ADAM metallopeptidase domain 15 | 8751 | Q13444 | 1 | 0.575086 |
| KIRREL | Kin of IRRE-like protein 1 | 55243 | Q96J84 | 1 | 0.514684 |
| CD59 | CD59 molecule (CD59 blood group) | 966 | P13987 | 1 | 0.559599 |
| ITPRIP | inositol 1,4,5-trisphosphate receptor interacting protein | 85450 | Q8IWB1 | 1 | 0.417335 |
| GGCX | gamma-glutamyl carboxylase | 2677 | P38435 | 2 | 0.703851 |
| ECE1 | endothelin converting enzyme 1 | 1889 | P42892 | 1 | 1.0075 |
| CNNM2 | cyclin and CBS domain divalent metal cation transport mediator 2 | 54805 | Q9H8M5 | 1 | 0.859415 |
| INSR | Insulin receptor | 3643 | P06213 | 1 | 0.912155 |
| SMPDL3B | sphingomyelin phosphodiesterase acid like 3B | 27293 | Q92485 | 1 | 1.01647 |
| HNRNPK | heterogeneous nuclear ribonucleoprotein K | 3190 | P61978 | 1 | 0.694312 |
| SLC4A7 | solute carrier family 4 member 7 | 9497 | Q9Y6M7 | 1 | 0.592821 |
| TPBG | trophoblast glycoprotein | 7162 | Q13641 | 1 | 0.787366 |
| NCSTN | nicastrin | 23385 | Q92542 | 1 | 0.772031 |
| M6PR | mannose-6-phosphate receptor, cation dependent | 4074 | P20645 | 1 | 0.557621 |
| TFRC | transferrin receptor | 7037 | P02786 | 1 | 0.422732 |
| PTPRF | protein tyrosine phosphatase receptor type F | 5792 | P10586 | 1 | 0.287566 |
| TMED7 | transmembrane p24 trafficking protein 7 | 51014 | Q9Y3B3 | 1 | 0.367156 |
| ASIC1 | acid sensing ion channel subunit 1 | 41 | P78348 | 1 | 0.541489 |
| SEL1L | SEL1L adaptor subunit of ERAD E3 ubiquitin ligase | 6400 | Q9UBV2 | 1 | 0.604953 |
| ABCC1 | ATP binding cassette subfamily C member 1 | 4363 | P33527 | 1 | 0.578134 |
| ANO6 | anoctamin 6 | 196527 | Q4KMQ2 | 1 | 0.478563 |
| TYRO3 | TYRO3 protein tyrosine kinase | 7301 | Q06418 | 1 | 0.544295 |
| SLC39A14 | solute carrier family 39 member 14 | 23516 | Q15043 | 1 | 0.594535 |
| NPTN | neuroplastin | 27020 | Q9Y639 | 1 | 0.499769 |

| | | | | | |
|-----------------|---|--------|--------|---|-----------|
| ALCAM | activated leukocyte cell adhesion molecule | 214 | Q13740 | 1 | 0.458814 |
| ATP1A1 | ATPase Na ⁺ /K ⁺ transporting subunit alpha 1 | 476 | P05023 | 1 | 0.340416 |
| NCR3LG1 | natural killer cell cytotoxicity receptor 3 ligand 1 | 374383 | Q68D85 | 1 | 0.578246 |
| ATP1B3 | ATPase Na ⁺ /K ⁺ transporting subunit beta3 | 483 | P54709 | 1 | 0.570178 |
| SLC3A2 | solute carrier family 3 member 2 | 6520 | P08195 | 1 | 0.643518 |
| ACTN1 | actinin alpha 1 | 87 | P12814 | 1 | 0.712156 |
| JAM3 | junctional adhesion molecule 3 | 83700 | Q9BX67 | 1 | 0.468754 |
| CD276 | CD276 molecule | 80381 | Q5ZPR3 | 1 | 0.428201 |
| CADM1 | cell adhesion molecule 1 | 23705 | Q9BY67 | 1 | 0.497293 |
| MIA3 | MIA SH3 domain ER export factor 3 | 375056 | 5JRA6 | 1 | -0.909785 |
| HLA-C | major histocompatibility complex, class I, C | 3107 | P10321 | 1 | -0.539395 |
| HLA-A | major histocompatibility complex, class I, A | 3105 | P04439 | 1 | -1.0196 |
| LGALS3BP | galectin 3 binding protein | 3959 | Q08380 | 1 | -0.991107 |
| CLPTM1L | cleft palate transmembrane protein 1-like | 81037 | Q96KA5 | 2 | -0.741051 |
| MFAP3 | microfibril associated protein 3 | 4238 | P55082 | 1 | -0.768297 |
| TOR1AIP1 | torsin 1A interacting protein 1 | 26092 | Q5JTV8 | 2 | -0.685934 |
| RPN1 | ribophorin 1 | 6184 | P04843 | 1 | -0.358063 |
| TMEM259 | transmembrane protein 259 | 91304 | Q4ZIN3 | 2 | -0.314032 |
| STT3A | STT3 oligosaccharyltransferase complex catalytic subunit A | 3703 | P46977 | 2 | -0.59739 |
| CACHD1 | cache domain containing 1 | 57685 | Q5VU97 | 1 | -0.580828 |
| CNTN1 | contactin 1 | 1272 | Q12860 | 1 | -0.711767 |
| EMC1 | ER membrane protein complex subunit 1 | 23065 | Q8N766 | 1 | -0.646965 |
| MCAM | melanoma cell adhesion molecule | 4162 | P43121 | 1 | -0.682648 |
| F11R | F11 receptor | 50848 | Q9Y624 | 1 | -0.478582 |
| ITGA4 | integrin subunit alpha 4 | 3676 | P13612 | 1 | -0.446824 |
| EPHA4 | EPH receptor A4 | 2043 | P54764 | 1 | -0.57585 |
| EFNB1 | ephrin B1 | 1947 | P98172 | 1 | -0.896389 |
| EPCAM | epithelial cell adhesion molecule | 4072 | P16422 | 1 | -0.78135 |
| BRI3BP | BRI3 binding protein | 140707 | Q8WY22 | 2 | -0.665538 |
| CD46 | CD46 molecule | 4179 | P15529 | 1 | -0.848529 |
| CAPNS1 | calpain small subunit 1 | 826 | P04632 | 1 | -0.562122 |

