

Electronic supplementary material

Nitric Oxide and Hydrogen Peroxide Signaling in Extractive *Shiraia* Fermentation by Triton X-100 for Hypocrellin A

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Table S1. Primers used in RT-qPCR. F: forward primer, R: reverse primer.

Gene symbol	Gene name	Sequence
<i>PKS</i>	Polyketide synthase	F: TGCTGAGGTAGCAGTCAAGC R: TTATGCTACGGTCGTCGCTC
<i>Mono</i>	Salicylate 1-monooxygenase	F: TCTCGGGAAATTATGGCACG R: ACAACCGTTCTCGCATCAGT
<i>OmeF</i>	O-methyltransferase	F: GAACTACCTGAAGGCACGCT R: GCTCGGAAGGATACTCGCTC
<i>MCO</i>	Multicopper oxidase	F: TATGGCGCTACGAGTGGAC R: ACTCCCTGGCGATAACGTA
<i>FAD</i>	FAD/FMN-containing dehydrogenase	F: TGTGACCGCCATCACCTTAC R: TTGTCGTATGGGTGGGAAGC
<i>MFS</i>	Major facilitator superfamily	F: TCCCCTAGCCTTGCTTCTG R: CCGGCTTCTTCTTGACGCTA
<i>ABC</i>	ATP-binding cassette	F: GACTTGAGCCTATCCGCCTC R: AGAGTCGCCTCTGTGATCCT
18S rRNA	18S rRNA gene	F: GAAAGTTAGGGATCGAAGA R: TAGTCGGCATAGTTACGGT

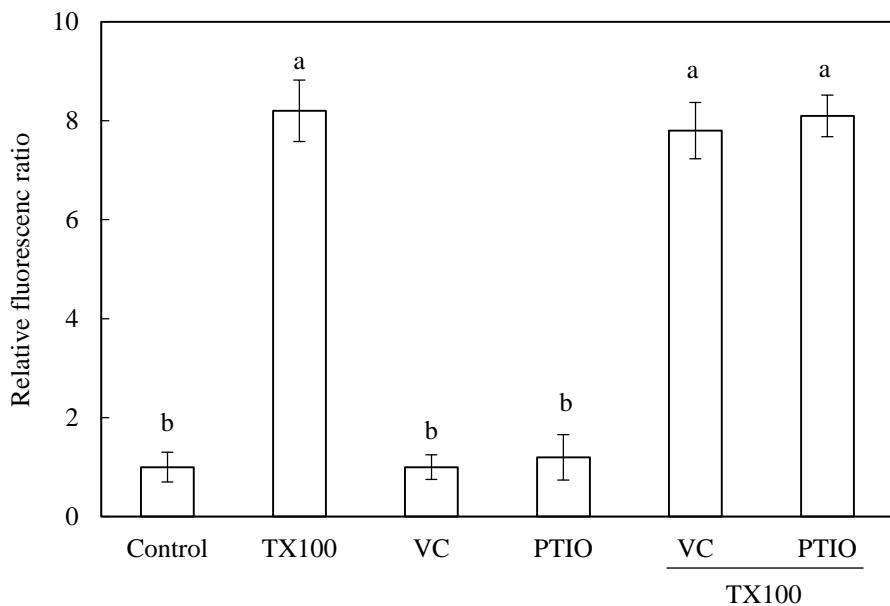


Figure S1. The effect of TX100, VC and PTTIO on membrane permeability of hyphal cells in the extractive fermentation. TX100 was added at 25 g/L at 36 h of the fermentation. The membrane integrity of hyphal cells was detected by a nucleic acid stain fluorescent dye SYTOX Green at 0.5 μ mol/L. The dosage and procedure for VC and PTIO addition was specified in Figure 3 and 4. The relative fluorescence ratio is defined to the ratio of fluorescence intensity at a given time to that of the control. Values are mean \pm SD from three independent experiments. Different letters above the bars mean significant differences ($p < 0.05$).