



F-box gene D5RF is regulated by *Agrobacterium* virulence protein VirD5 and essential for *Agrobacterium*-mediated plant transformation

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Supporting information

Supplementary materials can be found at <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE155112>.

Supplement Table 2 align_pct and align_region;

Supplement Table 3 *d5rf* vs col_0_deg_all;

Supplement Table 4 *d5rf* vs col_0_Goerich;

Supplement Table 5 *d5rf* vs col_0_KEGGenrich.

The RNA-Seq raw data are deposited in NCBI'S Sequence Read Archive (SRA) with accession code PRJNA647639.

Table S1. The putative genes directly regulated by VirD5.

Gene	Protein	No. of D5RE	Annotated functions
<i>At2g04230</i>	--	2	F-box and LRR protein
<i>AT5G16690</i>	<i>ATORC3</i>	2	Involved in double-strand break repair
<i>At3g19210</i>	<i>RAD54</i>	2	a member of the SWI2/SNF2 family of DNA-stimulated ATPases
<i>At5g49160</i>	<i>DDM2/MET1</i>	2	cytosine methyltransferase
<i>At3g49480.1</i>	<i>D5RF</i>	2	F-box/LRR-repeat protein
<i>At1g68910</i>	<i>WIT2</i>	2	WPP domain-interacting protein 2

Note: D5RE indicates VirD5 response element (CCGCNG).

Table S2. List of oligonucleotides.

Gene name	Primer name	Sequence	Restriction site
<i>AT3G49480.1</i>	p49480-F2	5'-GGgtaccTTACATAGAAACCACTAGTGACG-3'	<i>KpnI</i>
	P49480-R2	5'-CCGctcgagTCTGATCGGCTCTCCTTTC-3'	<i>XhoI</i>
	49480-LUC-F3	5'-GGgtaccTACCGGATGCGATATGGCAG-3'	<i>KpnI</i>
	49480-LUC-R3	5'-CGggtaccTCTGATCGGCTCTCCTTTCCA-3'	<i>BamHI</i>
	49480-F4	5'-GGgtaccATGCCTAAGAGCTTTTACTT-3'	<i>KpnI</i>
	49480-R4	5'-GGgtaccATATCTAAATGACATTCGAGC-3'	<i>KpnI</i>
	49480-QF1	5'-GTTGGAATCACTTTTGGTTTGTCTC-3'	-
	49480-QR1	5'-ACATCTTGAATAGAAGCTCGGTTGA-3'	-
<i>AT3G49480.2</i>	L49480-F1	5'-GGgtaccATGACAGAACTCGTCGAACATAG-3'	<i>KpnI</i>
	L49480-R1	5'-CATGcattggATATCTAAATGACATTCGAGCAG-3'	<i>NcoI</i>
<i>AT5G16690</i>	16690-QF1	5'-TGATAAGGTTGCCACAGTCATAGAA-3'	-
	16690-QR1	5'-GGTCTCCAAGTAAAGCAGATTGAAG-3'	-

<i>AT5G49610</i>	9160-QF1	5'-GAGTTTGCAGGGAACATAAATCACA-3'	-
	9160-QR1	5'-TTCTTGAGATGTAGGGCTTCTTTGA-3'	-
<i>AT3G19210</i>	19210-QF1	5'-ACATGGGTTTAGGGAAGACATTACA-3'	-
	19210-QR1	5'-TCCTGTCTCCAACCCATTTCTTAAT-3'	-
<i>AT1G68910</i>	8910-QF1	5'-GAAATGGAAAATGCAAATGAGTCGG-3'	-
	8910-QR1	5'-ACTTCTAATTCCCTGACTTGCTTCT-3'	-
<i>AT2G04230</i>	4230-QF1	5'-AGAGTTAGAGGAGGCAAAAATCAGT-3'	
	4230-QR1	5'-TTTAACCTCAAGGGGTGATAAGTGT-3'	
<i>VirD5</i>	D-F03	5'-ggatccATGACAGGAAAGTCGAAAGTT-3'	<i>Bam</i> HI
	D-R03	5'-gtcgacTCAGCGTTTAAACGCTTTGTC-3'	<i>Sal</i> II
<i>AT1G70880</i>	70880-QF	5'-GGCTCACCCCTGAAACTCTCC-3'	
	70880-QR-	5'-ACGTGCGGCTCAGTGTTTAT-3'	
<i>AT1G70890</i>	70890-QF	5'-GGGAGTCAGGGAGTATTGCG-3'	
	70890-QR-	5'-CTCGGCCAAGAGATGTTTCGT-3'	
<i>AT2G01520</i>	1520-QF	5'-ACACAAGGAAGAGAGATGGCG-3'	
	1520-QR	5'-GGGGAAGAGGTGTTCTCAC-3'	
<i>AT2G01530</i>	1530-QF	5'-AAGAAGCGCACACACACAAC-3'	
	1530-QR	5'-GTCGTGGACAGTGACACCTT-3'	
<i>AT2G02120</i>	2120-QF	5'-TCAGCCGTTCTTTTCTTGGTG-3'	
	2120-QR	5'-GCTCACGCATTTACCCTTGA-3'	
<i>AT3G26460</i>	26460-QF	5'-CAACATCACCATCATCCCTATATGT-3'	
	26460-QR	5'-AGAAAGTAACTCCACAAGACCCA-3'	
<i>AT5G36910</i>	36910-QF	5'-GCATGCTTTCCGTGTCATCC-3'	
	36910-QR	5'-TTGGAGAGTGGTCAAGGCAC-3'	
<i>AT5G44430</i>	4430-QF	5'-TGGTGCTTGATCGTGTGTGTA-3'	
	4430-QR	5'-AACAGTCCAACGTCGTACATAA-3'	

-500 TTACATAGAAACCCTAGTGACGTAAAAAGCAATACTGTTTGGTGGTTTCTGAAGAGTCATTCCGTTT
-431 TCACAAGGCACCTTTGTTAGAAGCGTTTGTGGATCGACCTCGGTCCACAATGTCCTATTAATGTTAATCC
-362 TGTAAAGTGGGTGCAAAAAGTACTTGATACCAAGGGCGGATCTAGAATGTAATTTAATGTGGGGCACCA

VirD5 response element

-293 TATATATAAATAGTACTAAGTATAATTTAATTACCTAAATTGTATTGTATTATAAAATTATTTTTACAA
-224 AAATAACA GTGCGGAGACTTGAAGTTCGGGTAGTAGGCACTTTTTGCTGGAAAACATTGAGTTA

VirD5 response element

-155 CCTGTATTACATTTGTTTAGTAGTAGCTTATAAAAATTTATAGAATTAGTATCAGGCACGTGCCCCAC
-86 CTGCCTCCAACGTGGGTCCGCACTGCTTGATACTCGTGTGCGTTGGCTAACTTTCAGGCTCCTTTGGA
-17 AAGGAGAGCCGATCAGA

Figure S1. The promoter sequence of *D5RF.1* gene (500 bp upstream of the translational start site), On the chromosome 3: 18342043-18342543. The red sequences are the possible binding sites of VirD5 protein (D5RE).

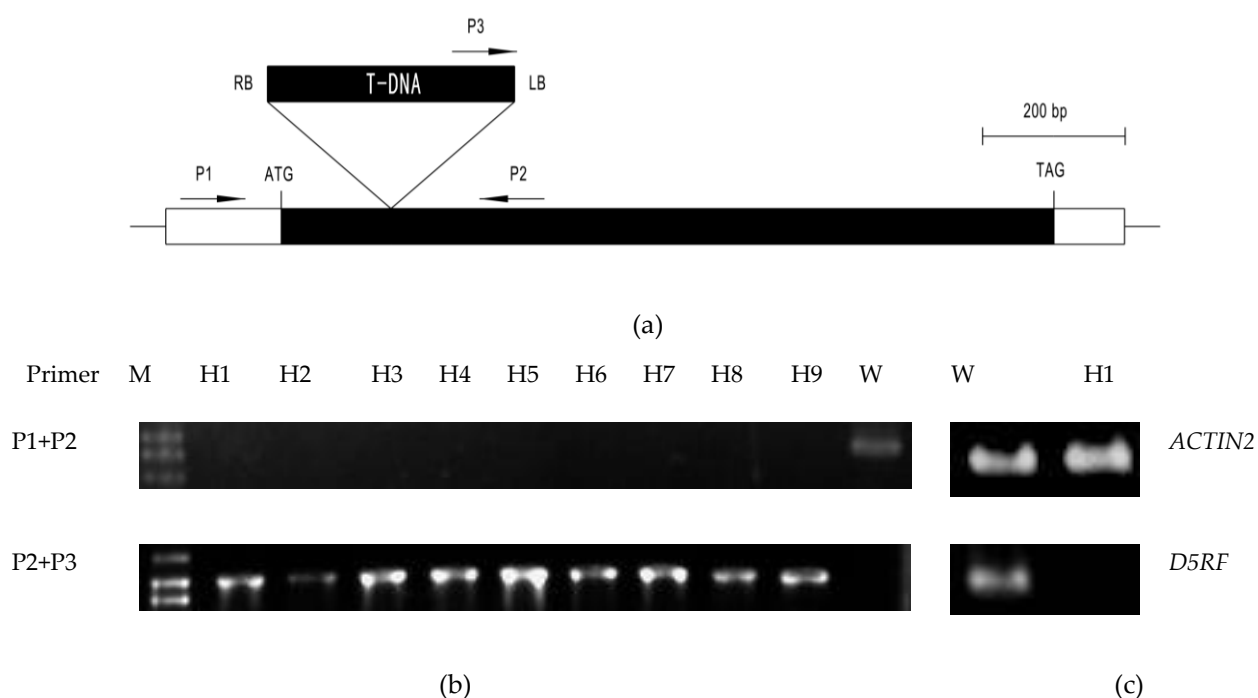


Figure S2. Identification of the *d5rf* mutant. (a) *D5RF.1* structure. The T-DNA was inserted into the exon approximately 200 bp downstream of the translational start site. P1, P2 and P3 indicate primers used for genotyping; RB and LB indicate the T-DNA right border and left border, respectively. (b) PCR genotyping of *d5rf* mutants. M indicates Marker. W and H indicate the wild-type and homozygous *d5rf* mutant plants, respectively. (c) RT-PCR analysis of *d5rf* and wild-type plants. RT-PCR showed that the expression level of *D5RF* was depressed in the *d5rf* mutant. Actin was used as the control; W, wild-type; H, homozygous *d5rf* mutant.

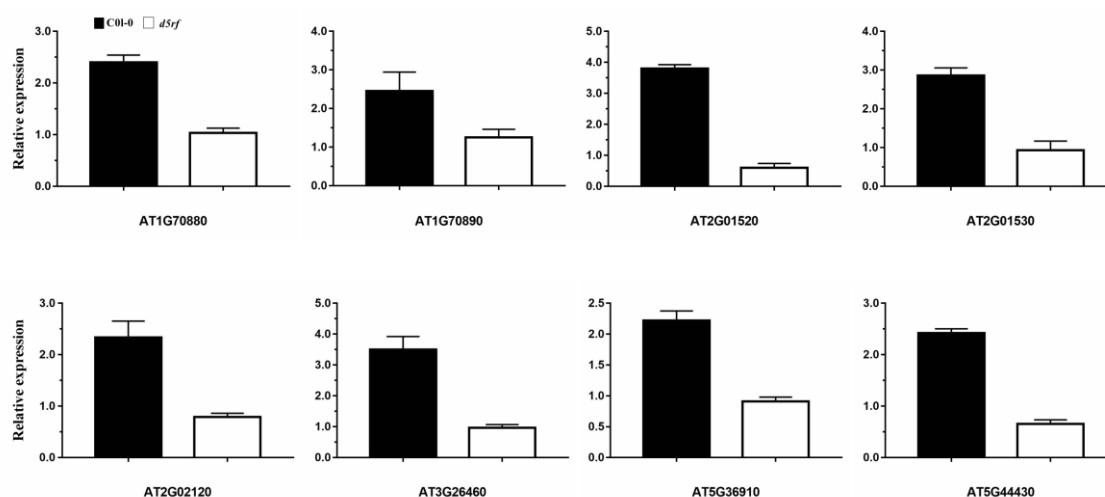


Figure S3. Validation of the RNA-Seq Data by Real-Time qRT-PCR. Eight different pathogen-responsive Genes genes that had less transcript abundance in *d5rf* compared with Col-0, based on RNA-Seq experiments, were selected for validation. Total RNA was extracted from leaves of wild-type Col-0 and *d5rf*. The first-strand cDNA was synthesized and used for qRT-PCR using gene-specific primers. The data represent the average of three biological replicates. each biological replicate includes three technical replicates. SE values were shown as error bars.