Manuscript Kabilan et al.

Supplementary Table. A brief summary of radiation effects related to radiation hormesis observed in a variety of organisms/cell types.

Model organism	Range of dose/dose- rate	Effects	Reference
House Cricket (Acheta domesticus)	0.5-2 Gy	Survival fitness and reproductive maturity was observed at low (but not high) doses.	[1]
Rat (<i>Rattus norvegicus</i>)	0.75 Gy	γ - (but not X-ray) radiation promoted dermal wound healing; resistance to longitudinal stress.	[2]
Large Japanese Field Mouse (Apodemus speciosus)	0.05-21 mGy/day	Increased spermatogenesis in the mice treated with low doses of γ -radiation. The highest dose rate 21 mGy/day lead to an increase in neoplastic cell death which was not observed at lower dose rates.	[3]
Large Japanese Field Mouse (Apodemus speciosus)	4.1-13.9 μGy/hour	At low doses/dose rates of γ -radiation, there were no detrimental effects of spermatogenesis observed in the exposed mice population at Fukushima Daiichi Nuclear Power plant.	[4]
Fruit Fly (Drosophila Melanogaster)	400 mGy (irradiated at the rate of 0.014 mGy/hr for 12 days)	Chronic exposure to 400 mGy of γ-radiation upregulated DNA repair genes resulting in radioadaptive mechanisms.	[5]
Mouse (<i>Mus musculus</i>)	25-500 mGy	The frequency of aortic atherosclerotic lesion formation reduced in APOE ^{-/-} mice irradiated at 25-50 mGy while the control group (unexposed APOE ^{-/-} mice) showed a gradual increase in the lesion formation with time (8 months).	[6]
Fish (Danio rerio)	1.4-8.4 mGy	Low apoptotic signals observed at doses below 2.8 mGy resulting in radio adaptive responses. Bystander effect was also observed.	[7]
Fish (Danio rerio)	4.4 mGy	Active clearance of apoptotic cells was observed in response to low-dose radiation.	[8]
Worm C. elegans	0.5-1000 mGy	Increased activity of 20S proteasome in worms treated with highest dose (1 Gy) while this was not observed at lower doses.	[9]

Mouse	650 and 2000 mGy	At 650 mGy, photoreceptor apoptosis was reduced through upregulation	[10]
(Mus musculus)		of Prdx2 gene resulting in decreased neuro degradation in rd10 mice and	
		inducing radiation mediated neurohormesis.	
Mouse	500 mGy	Stimulation of osteoblast proliferation, enhanced wound healing.	[11]
(MC3T3-E1 cells)			
Human	0.01-2 Gy	A delay in cellular senescence with acute 30-50 mGy doses of γ -radiation.	[12]
(HELF-104 cells)			
Human	2-10 mGy	Decreased aneuploidy of chromosomes 1 and 2.	[13]
(CCD-986 sk)			
Fish – Medaka	0 - 5.88 Gy	Radioadaptive responses seen in fish pre-exposed to low doses.	[14]
(Oryzias latipes)		Communication of protective signals between irradiated and non-	
		irradiated individuals as an evidence of bystander effect.	
Bacteria	100 mGy	Treatment with 100 mGy of ionization radiation using tritium for 15 days	[15]
Photobacterium		did not induce any mutation in the bacterial genome. Bacterial	
Phosphoreum		luminescence was higher without tritium penetration into the bacterial	
		cells.	
Protozoan	0-50 mGy/year	The cellular proliferation is stimulated upon irradiation and this	[16]
Paramecium tetraurelia		stimulatory effect declines at doses below 50 mGy/year.	
Plant	0.1-10 Gy	No increase in homologous recombination frequency upon acute	[17]
Arabidopsis thaliana		irradiation at $0.1 - 0.5$ Gy. 5-6-fold increase in the the homologous	
		recombination frequency upon chronic irradiation at >200 mGy.	
Plant	50 mGy	50 mGy of acute γ -irradiation upregulated Calnexin-1 gene, membrane	[18]
Tradescantia BNL 4430		protein functions as a chaperone in endoplasmic reticulum.	
		Overexpression studies using <i>E. Coli</i> strain BL21 (DE3) increased the	
		cellular viability which suggest that IR upregulating Calnexin-1 in plants to	
		withstand the radiation associated stress.	
Plant	196 mGy	Induction of genes such as Superoxide Dismutase (SOD2), Glutathione	[19]
Stipa capillata		Reductase (GR), Peroxidases (POD), Catalase (CAT), Glucose-6-phosphate	
		Dehydrogenase (G6PDH) is observed upon irradiation.	
Protozoa	0.4 – 2.7 mGy	Stimulation of growth of the organism.	[20]
Tetrahymena pyriformis			
Bacteria	31 mGy/year	Cells irradiated with low doses showed stimulated growth rates and the	[21]
Synechococus lividus		stimulatory effect diminished as radiation dose increased.	

Yeast	7 mGy	Radiation induced higher survivability rate in melanin containing yeast	[22]
Wangiella dermatitidis		compared to melanin lacking yeast. This effect was due to enhanced DNA	
		repair mechanisms and genome stability.	

SUPPLEMENTARY REFERENCES:

- 1. Shephard, A.M.; Aksenov, V.; Tran, J.; Nelson, C.J.; Boreham, D.R.; Rollo, C.D. Hormetic effects of early juvenile radiation exposure on adult reproduction and offspring performance in the cricket (Acheta domesticus). *Dose-Response* **2018**, *16*, doi:10.1177/1559325818797499.
- 2. Jabbari, N.; Farjah, G.H.; Ghadimi, B.; Zanjani, H.; Heshmatian, B. Acceleration of skin wound healing by low-dose indirect ionizing radiation in male rats. *Kaohsiung J. Med. Sci.* **2017**, *33*, 385–393, doi:10.1016/j.kjms.2017.05.013.
- 3. Takino, S.; Yamashiro, H.; Sugano, Y.; Fujishima, Y.; Nakata, A.; Kasai, K.; Hayashi, G.; Urushihara, Y.; Suzuki, M.; Shinoda, H.; et al. Analysis of the Effect of Chronic and Low-Dose Radiation Exposure on Spermatogenic Cells of Male Large Japanese Field Mice (Apodemus speciosus) after the Fukushima Daiichi Nuclear Power Plant Accident. *Radiat. Res.* **2017**, *187*, 161, doi:10.1667/rr14234.1.
- 4. Okano, T.; Ishiniwa, H.; Onuma, M.; Shindo, J.; Yokohata, Y.; Tamaoki, M. Effects of environmental radiation on testes and spermatogenesis in wild large Japanese field mice (Apodemus speciosus) from Fukushima. *Sci. Rep.* **2016**, *6*, 1–8, doi:10.1038/srep23601.
- 5. Koval, L.; Proshkina, E.; Shaposhnikov, M.; Moskalev, A. The role of DNA repair genes in radiation-induced adaptive response in Drosophila melanogaster is differential and conditional. *Biogerontology* **2020**, *21*, 45–56, doi:10.1007/s10522-019-09842-1.
- 6. Mitchel, R.E.J.; Hasu, M.; Bugden, M.; Wyatt, H.; Little, M.P.; Gola, A.; Hildebrandt, G.; Priest, N.D.; Whitman, S.C. Low-Dose Radiation Exposure and Atherosclerosis in ApoE –/– Mice . *Radiat. Res.* **2011**, *175*, 665–676, doi:10.1667/rr2176.1.
- 7. Choi, V.W.Y.; Cheung, A.L.Y.; Cheng, S.H.; Yu, K.N. Hormetic effect induced by alpha-particle-induced stress communicated in vivo between zebrafish embryos. *Environ. Sci. Technol.* **2012**, *46*, 11678–11683, doi:10.1021/es301838s.
- 8. Yuen Ping Ng, C.; Han Cheng, S.; Ngok Yu, K. Effect of Photon Hormesis on Dose Responses to Alpha Particles in Zebrafish Embryos. *Int. J. Mol. Sci. Artic.* **2017**, doi:10.3390/ijms18020385.
- 9. Dubois, C.; Lecomte, C.; Ruys, S.P. dit; Kuzmic, M.; Della-Vedova, C.; Dubourg, N.; Galas, S.; Frelon, S. Precoce and opposite response of proteasome activity after acute or chronic exposure of C. elegans to γ-radiation. *Sci. Rep.* **2018**, *8*, doi:10.1038/s41598-018-29033-1.
- 10. Otani, A.; Kojima, H.; Guo, C.; Oishi, A.; Yoshimura, N. Low-Dose-Rate, Low-Dose Irradiation Delays Neurodegeneration in a Model of Retinitis Pigmentosa. *Am. J. Pathol.* **2012**, *180*, 328–336, doi:10.1016/j.ajpath.2011.09.025.

- 11. Chen, M.; Huang, Q.; Xu, W.; She, C.; Xie, Z.G.; Mao, Y.T.; Dong, Q.R.; Ling, M. Low-dose X-ray irradiation promotes osteoblast proliferation, differentiation and fracture healing. *PLoS One* **2014**, *9*, doi:10.1371/journal.pone.0104016.
- 12. Velegzhaninov, I.O.; Ermakova, A. V.; Klokov, D.Y. Low dose ionizing irradiation suppresses cellular senescence in normal human fibroblasts. *Int. J. Radiat. Biol.* **2018**, *94*, 825–828, doi:10.1080/09553002.2018.1492167.
- 13. Cho, Y.H.; Kim, S.Y.; Woo, H.D.; Kim, Y.J.; Ha, S.W.; Chung, H.W. Delayed numerical chromosome aberrations in human fibroblasts by low dose of radiation. *Int. J. Environ. Res. Public Health* **2015**, *12*, 15162–15172, doi:10.3390/ijerph121214979.
- 14. Smith, R.W.; Mothersill, C.; Hinton, T.; Seymour, C.B. Exposure to low level chronic radiation leads to adaptation to a subsequent acute X-ray dose and communication of modified acute X-ray induced bystander signals in medaka (Japanese rice fish, Oryzias latipes). *Int. J. Radiat. Biol.* **2011**, *87*, 1011–1022, doi:10.3109/09553002.2011.587861.
- 15. Rozhko, T. V.; Badun, G.A.; Razzhivina, I.A.; Guseynov, O.A.; Guseynova, V.E.; Kudryasheva, N.S. On the mechanism of biological activation by tritium. *J. Environ. Radioact.* **2016**, *157*, 131–135, doi:10.1016/j.jenvrad.2016.03.017.
- 16. Planel, H.; Soleilhavoup, J.P.; Tixador, R.; Richoilley, G.; Conter, A.; Croute, F.; Caratero, C.; Gaubin, Y. Influence on cell proliferation of background radiation or exposure to very low, chronic γ radiation. *Health Phys.* **1987**, *52*, 571–578, doi:10.1097/00004032-198705000-00007.
- 17. Kovalchuk, O.; Arkhipov, A.; Barylyak, I.; Karachov, I.; Titov, V.; Hohn, B.; Kovalchuk, I. Plants experiencing chronic internal exposure to ionizing radiation exhibit higher frequency of homologous recombination than acutely irradiated plants. *Mutat. Res. Fundam. Mol. Mech. Mutagen.* **2000**, *449*, 47–56, doi:10.1016/S0027-5107(00)00029-4.
- 18. Ha, H.J.; Subburaj, S.; Kim, Y.S.; Kim, J.B.; Kang, S.Y.; Lee, G.J. Molecular characterization and identification of calnexin 1 as a radiation biomarker from tradescantia BNL4430. *Plants* **2020**, *9*, doi:10.3390/plants9030387.
- 19. Zaka, R.; Vandecasteele, C.M.; Misset, M.T. Effects of low chronic doses of ionizing radiation on antioxidant enzymes and G 6 PDH activities in Stipa capillata (Poaceae)., doi:10.1093/jxb/erf041.
- Luckey, T. Ionizing Radiation Promotes Protozoan Luckey, T. (1986). Ionizing Radiation Promotes Protozoan Reproduction PubMed. Retrieved March 26, 2020, from Radition Research website: https://pubmed.ncbi.nlm.nih.gov/3097750/Reproduction - PubMed Available online: https://pubmed.ncbi.nlm.nih.gov/3097750/ (accessed on Mar 26, 2020).
- Conter, A.; Dupouy, D.; Planel, H. Effects of Dose Rate on Response of Synechococcus lividus to Very Low Doses of Chronic γ Radiation: Influence of Enzymatic Equipment of Starting CellsEffects of Dose Rate on Response of Synechococcus lividus to Very Low Doses of Chronic g Radiation: Influence of Enzymatic Equipment of Starting Cells. *Radiat. Res.* **1986**, *105*, 379, doi:10.2307/3576693.
- 22. Robertson, K.L.; Mostaghim, A.; Cuomo, C.A.; Soto, C.M.; Lebedev, N.; Bailey, R.F.; Wang, Z. Adaptation of the Black Yeast Wangiella

dermatitidis to Ionizing Radiation: Molecular and Cellular Mechanisms. *PLoS One* **2012**, *7*, e48674, doi:10.1371/journal.pone.0048674.