

Supplementary Materials

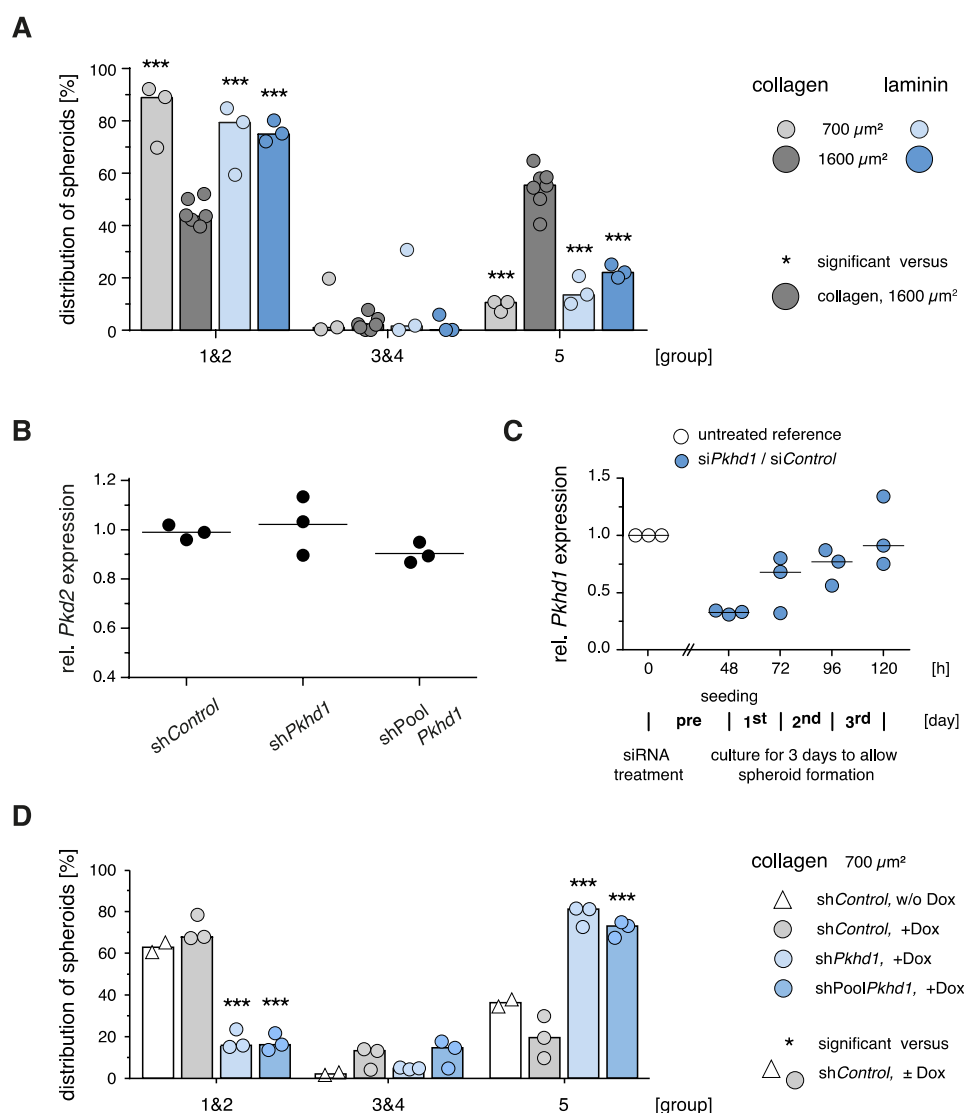


Figure S1. Sensitivity of spheroid formation to confinement, ECM coating and *Pkhd1* silencing by shRNA confirms function of morphogenesis assay. **(A)** Impact of confinement and ECM proteins on epithelial characteristics. Spheroid formation of MDCKII cells on 700 or 1600 μm^2 pattern coated with collagen or laminin. ($n = 3\text{--}7$ independent experiments, median bars; >150 spheroids per condition; two-way ANOVA/Tukey's, $p < 0.001$, ***). **(B)** *Pkd2* mRNA is not specifically reduced in *Pkhd1* silenced MDCK II cells as compared to control treated cells ($n = 3$ independent experiments). **(C)** Time course of the MDCKII spheroid assay is shown in its temporal relation to siRNA treatment of cells and si*Pkhd1* knockdown. Knockdown was determined as detailed in the legend of Figure 1C ($n = 3$ independent experiments). **(D)** Spheroid formation is defective in shRNA-silenced MDCKII TetON-cells [17] with sh*Pkhd1* and shPool*Pkhd1* as compared to sh*Control* expression. Three days after induction of shRNA with doxycycline, cells were seeded on 700 μm^2 collagen-coated disc-shaped micropattern. Non-induced sh*Control* (-Dox) was included as further control and spheroid formation analyzed for all conditions as described for siRNA, Figure 1E ($n = 2\text{--}3$ independent experiments, median bars; >130 spheroids per condition; two-way ANOVA/Tukey's, $p < 0.001$, ***).

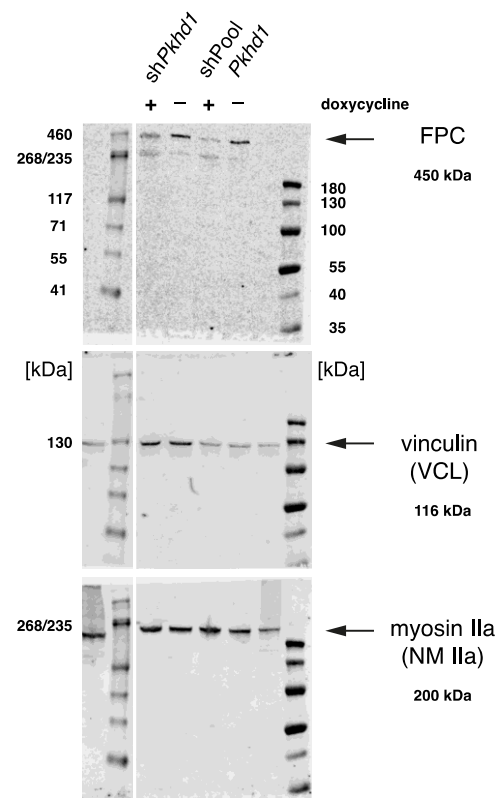


Figure S2. Full-length immunoblot membranes of Figure 1D, showing detection of fibrocystin (FPC), and load controls, vinculin (VCL) and non-muscle myosin IIa (NM IIa), alongside with molecular weight marker bands. Note: protein levels are compared within treatment groups, sh*Pkhd1* and sh*PoolPkd1*, with (+) and without (-) induction of shRNA by doxycycline. There is no impact of shRNA on VCL and NM IIa levels.

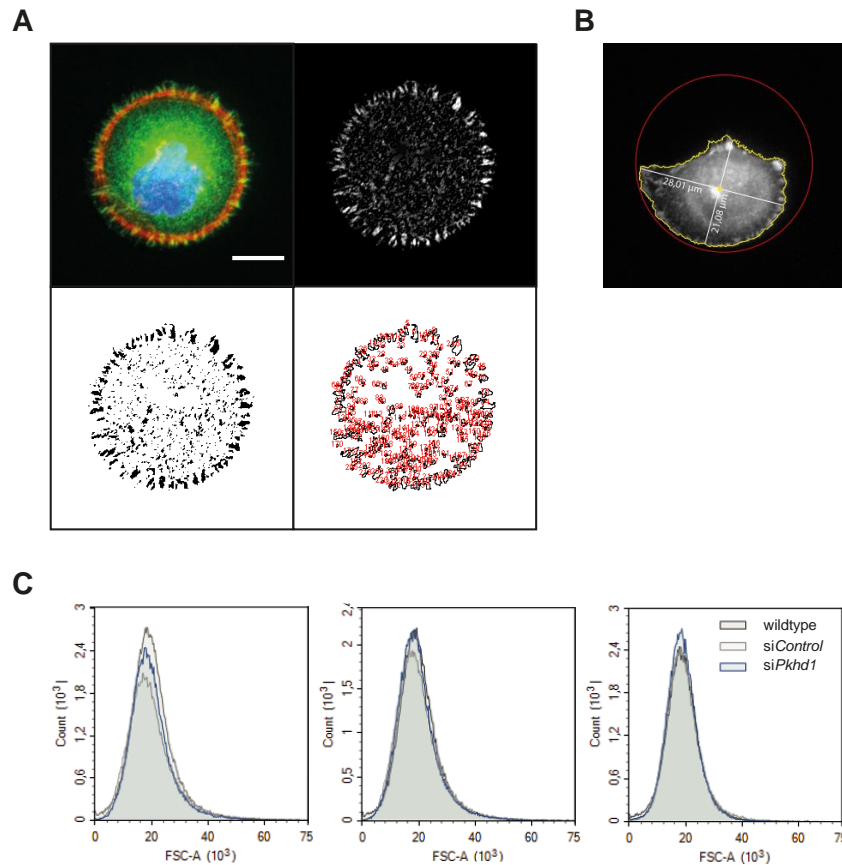


Figure S3. Parameter determination of adherent cells and size in solution. **(A)** Steps illustrate extraction of cell-ECM adhesion sites; (top left) 3-color fluorescence image for vinculin (green), f-actin (red) and nucleus (blue), (top right) background subtracted image (vinculin), (bottom left) binary mask, (bottom right) outlines of extracted adhesion sites (>0.1 μm^2). Outlines (without numbers) are routinely used for visual inspection of adhesion site assignment. Parameters are adjusted to exclude discrepancies between personal observation and automated detection. **(B)** Graph shows measurement of cell length and perpendicular width. **(C)** After siRNA treatment, cell size in suspension is not changed. FACS analysis of MDCKII wildtype, siControl and siPkh1-treated cells showing equal distribution of forward scatter (FSC) parameters for three independent knockdown experiments. Profiles and statistics reveal identical cell size/volume with no detectable shift to higher/lower values of FSC-A.

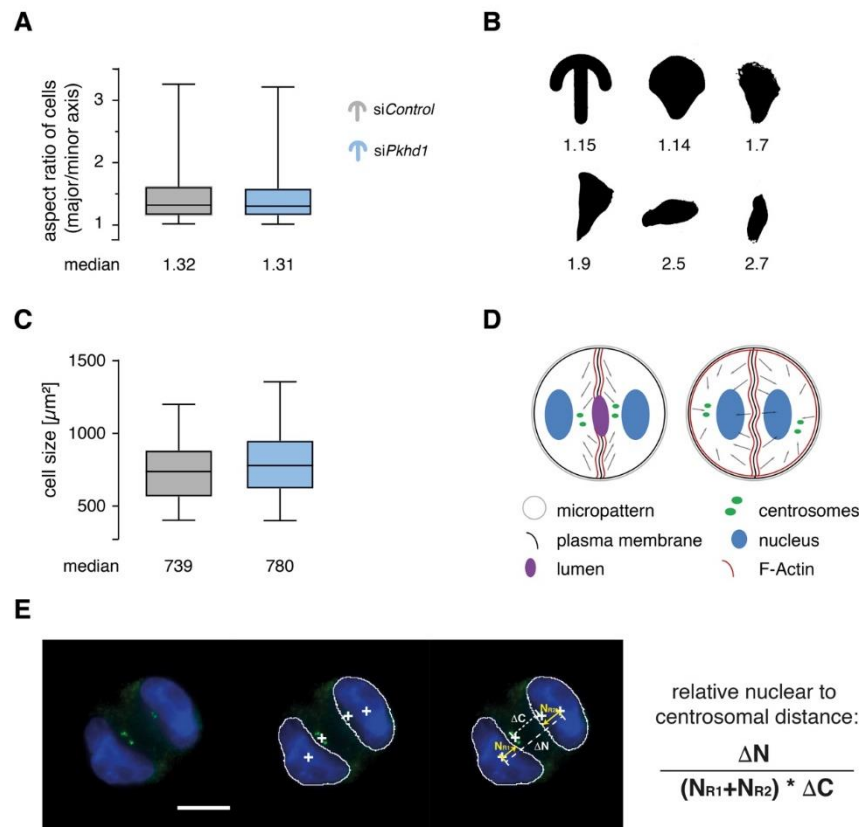


Figure S4. Shape and size of cells on crossbow and detection of centrosomes positions in 2-cell stages. Four hours after seeding, single cells were analyzed on crossbow-shaped micropatterns (1100 μm^2 , CW-M-A, CYTOO) for (A) cell geometry, determined as relation of major to minor cell axis, with (B) exemplary shapes to illustrate aspect ratios, and (C) spreading area, box plot and whiskers 5/95%. ($n = 250\text{--}310$ cells, $n = 4$ independent experiments; non-parametric t -test/Mann Whitney, not significant) (D) Model of centrosome and nuclear positioning in 2-cell states. Under confinement conditions, junctional tension is mostly restricted to cell-cell adhesions favoring induction of apical surface and lumen formation (left). When cell-matrix adhesion is high, peripheral tension pushes nuclei towards the cell-cell junction and inhibits polarization and initiation of lumen (right). Arrows indicate distribution of tension (E) Determination of relative nuclear to centrosomal distance; marks indicate centers of mass for nuclei and centrosomes. Size bar, 10 μm .