



1 Supplementary Materials

2 Peptides Derived from (RRWQWRMKKLG)₂-K-Ahx 3 Induce Selective Cellular Death in Breast Cancer Cell 4 Lines through Apoptotic Pathway

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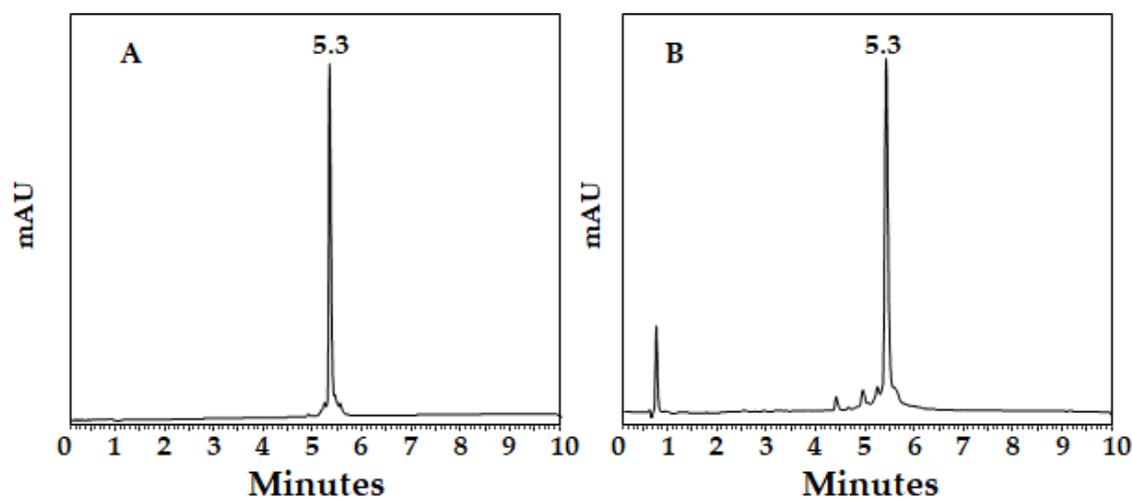
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11 Analysis by RP-HPLC of peptide 26[M] after four months of storage.

12 Dimer ²⁶[M]: (RRWQWRMKKLG)₂-K-Ahx, (10 μL, 1 mg/mL) was analyzed, at different times of storage,
13 by RP-HPLC using an Agilent Series 1260 chromatograph; Chromolith® C-18 monolithic column (50 × 4.6 mm).
14 Solvent A: 0.05% TFA in water, Solvent B: 0.05% TFA in ACN, for the analysis an elution gradient of 5 to 50% of
15 solvent B was used, gradient time was 8 minutes at a flow rate of 2.0 mL/min, and 210 nm for detection.

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18 **Figure S1.** Analysis by RP-HPLC of purified peptide ²⁶[M]: (RRWQWRMKKLG)₂-K-Ahx at zero time (A) and
19 after four months of storage (B).
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21 Synthesis of the dimeric peptides by SPPS-Fmoc/tBu

22 The dimeric peptides (RRWQWRMKKLG)₂-K-Ahx (²⁶[M]) and its analogs were synthesized
23 using the solid phase peptide synthesis (SPPS) methodology by the Fmoc/tBu strategy. The Rink
24 Amide resin (0.46 meq/g, Figure S2a) was used as solid support, this resin (200 mg) was conditioned
25 with DMF for 12 h. Figure S2 shows the general scheme for obtaining the dimeric peptides, please
26 note that in red is shown the 26th residue that was Methionine for the original peptide, ²⁶[M], and it
27 was replaced by Lys, Asp, Ala, Phe or Leu for the dimers ²⁶[K], ²⁶[D], ²⁶[A], ²⁶[F] and ²⁶[L], respectively.

28 Briefly, SPPS-Fmoc/tBu involved three steps [48,49], Figure S2. (Step 1) The Fmoc group removal:
 29 this reaction was carried out by treating the protected resin with 4-methylpiperidine (2.5% v/v) in
 30 DMF, 10 min at room temperature (RT) ($\times 2$). (Step 2) The Fmoc-amino acid coupling: for attaching
 31 each amino acid, the corresponding Fmoc-amino acid (0.46 mmol) was first treated with DCC/6-
 32 HOBT (0.45/0.46 mmol) in DMF, the reaction was stirred vigorously for 15 min at RT. This mixture
 33 was then added to the unprotected resin and allowed to react for 1 h, with stirring. The Kaiser test
 34 was performed to monitor both the Fmoc removal and coupling reaction. For obtaining each dimer,
 35 the amino acid Fmoc-6-aminohexanoic (*Ahx*) was bound as the C-terminal residue (Figure S2c-d),
 36 then Fmoc-Lys(Fmoc)-OH amino acid was attached (Figure S2e), and by Fmoc removal reaction it
 37 was generated two free amino group, at α and ϵ position, and over them the target sequence was
 38 constructed, generating a protected dimeric structure (Figure S2f). (Step 3) Cleavage reaction: the side
 39 chains protecting groups removal and the peptide cleavage from the resin was carried out by treating
 40 the dried resin-peptide with a solution containing TFA/H₂O/TIS/EDT (9.5/2.5/2.5/2.5%, v/v), the
 41 reaction was stirred for 4 h at RT. Then, the resin was filtered and the solution was treated with cold
 42 ethyl ether precipitating the dimer. Finally, crude products were purified by SPE [50] and analyzed
 43 by RP-HPLC and MALDI-TOF MS.

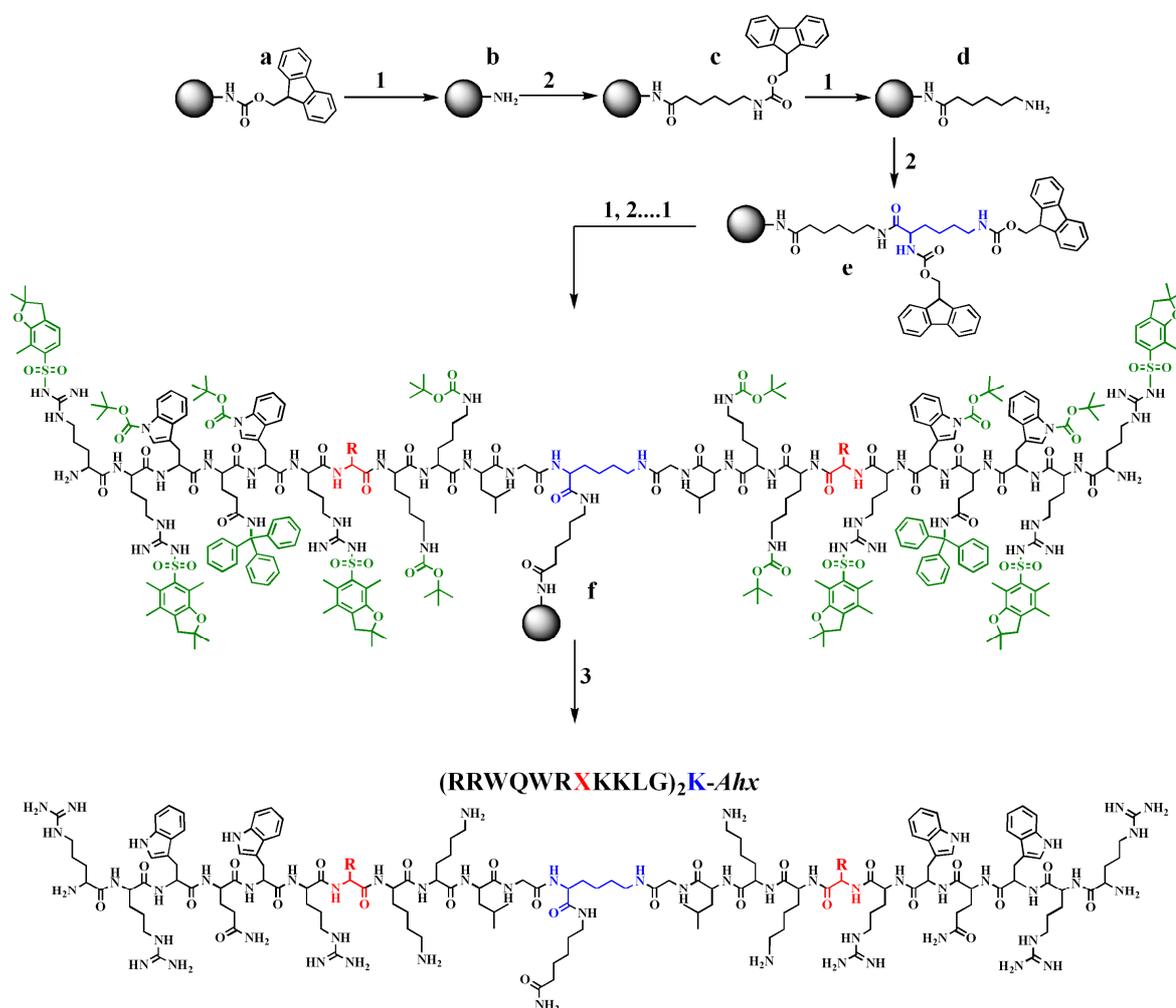
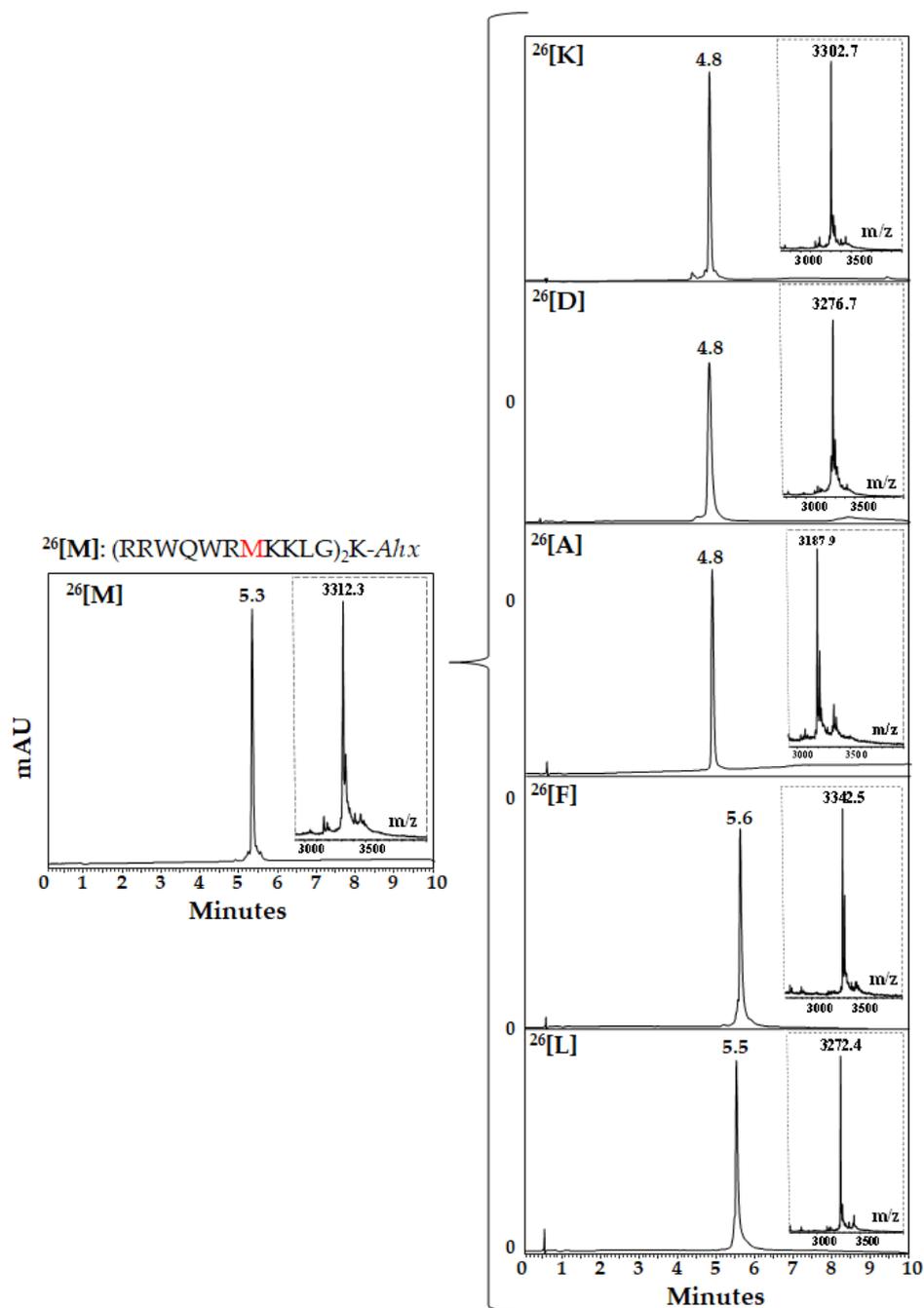


Figure S2. Synthesis of dimeric peptides by SPPS-Fmoc/tBu. Lysine residue used for building the two branches, in blue. The amino acid at position 26, in red. The protecting groups of the side chain from the amino acids, in green. 1: The Fmoc group removal. 2: The Fmoc-amino acid coupling. 3: Cleavage reaction.

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Purified Dimer Characterization by RP-HPLC and MALDI-TOF MS



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57 **Figure S3.** Characterization of purified (RRWQWRMKKLG)₂-K-Ahx (26 [M]) and its analogs. Chromatographic
 58 profiles and MALDI-TOF MS spectra (in dashed square). The residue at position 26 corresponds to Lys, Asp,
 59 Ala, Phe or Leu for 26 [K], 26 [D], 26 [A], 26 [F] and 26 [L], respectively. Peptides were analyzed as described above and
 60 in Materials and Methods



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