



Figure S1. MPD-GFP expression and purification strategy. **A)** MPD1 or MPD2 expression vector scheme. MPD1 or MPD2 genes were cloned *in frame* using *EcoRI* and *PstI* restriction enzymes into the *E. coli* pTTQ18-C3-GFP-His vector. The resulting fusion protein would contain a C3 PreScission cleavage site followed by the superfolder Green Fluorescent Protein (sf-GFP) and a 10x Histidine C-terminal tag. **B)** Size Exclusion Chromatography (SEC) of purified MPD1/2-GFP-His proteins. The purified MPD1/2-GFP-Hisproteins by Ni⁺⁺-NTA affinity chromatography column were run into a Superdex 200 10/300 SEC column and both proteins show a main elution peak centered at a 15.1 mL volume (blue dashed line corresponds to MPD1-GFP-His and red line to MPD2-GFP). This result is compatible with a ca. 50 kDa molecular size protein that would correspond to a monomer of the fusion protein. **C and D)** The SEC fractions 1 to 4 were collected and run into an SDS-PAGE electrophoresis using a 10 % polyacrylamide gel. **C)** In Gel Fluorescence of the fractions 1 to 4 corresponding to the main peak of the SEC of both purified proteins MPD1 or MPD2 fusion proteins. The “*” shows the fluorescent protein that fits to the molecular weight of a dimer of MPD1/2-GFP fusion proteins. Non-boiled samples run as a dimer and boiled ones run as a monomer in SDS-PAGE electrophoresis (see figure 3). **D)** Coomassie staining of the same SDS-PAGE gel showed the purity of the protein sample (*) and small amounts of free GFP and some impurities that could be removed with a second purification step.