



1 Article

2 **Aspalathin-Enriched Green Rooibos Extract Reduces** 3 **Hepatic Insulin Resistance by Modulating PI3K/AKT** 4 **and AMPK Pathways**

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24 **Supplementary material**

25 ***In vitro* data**

26 **Effect of GRE on normal C3A liver cells without palmitate**

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28 The effect of the GRE on glucose uptake by C3A liver cells was used to determine the optimal
29 effective concentration *in vitro*. The extracts GRE was freshly made up in cell culture tested sterile
30 water at a stock concentration of 0.1 mg/μl, the stock solution was further diluted to working
31 solutions of (100, 10, 1, 0.1, 0.01, 0.001, 0.0001 μg/mL) thereafter glucose uptake was performed,
32 insulin was used as positive control.
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35 **Results**

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38 We performed an *in vitro* glucose uptake study to determine the most active concentration for GRE
39 on normal C3A liver cells. Glucose uptake was enhanced in a dose-dependent manner at all
40 concentrations tested by all GRE concentration. This increase was without any cytotoxicity (Data not
41 shown). Therefore, a concentration of 10μM was selected for further assays.

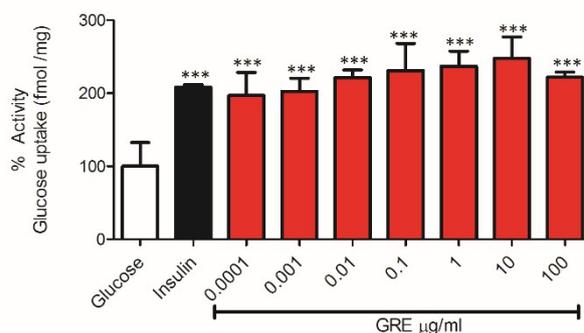


Figure 1S. The effect green rooibos extract (GRE) on glucose uptake in C3A liver cells (B). Cells were cultured in 8 mM glucose in DMEM without phenol red and pyruvate containing log dilutions of GRE for 3 h. Glucose uptake was measured. GRE increased glucose uptake similar insulin at all concentrations. Results are expressed as the mean of three independent experiments expressed relative to control (8 mM glucose) at 100% ± SD; ***p < 0.001.

***In vivo* models and animal housing**

This study was approved by the Ethics Committee for Research on Animals (ECRA) of the South African Medical Research Council (SAMRC) (ECRA 11/03/H). The study was executed in accordance with the principles and guidelines of the SAMRC as outlined in Guidelines on Ethics for Medical Research: Use of Animals in Research and Training, 2004 (<http://www.mrc.ac.za/ethics/ethicsbook3.pdf>). Three-week-old weaning male Wistar rats were obtained and housed at the Primate Unit of the South African Medical Research Council (Tygerberg, South Africa). The rats were housed individually in wired top and bottom polycarbonate cages, fitted with Perspex houses and maintained in a temperature-controlled room of 24-26°C, humidity of 45-55% with 15-20 air changes per hour and on a 12 h light/dark cycle.

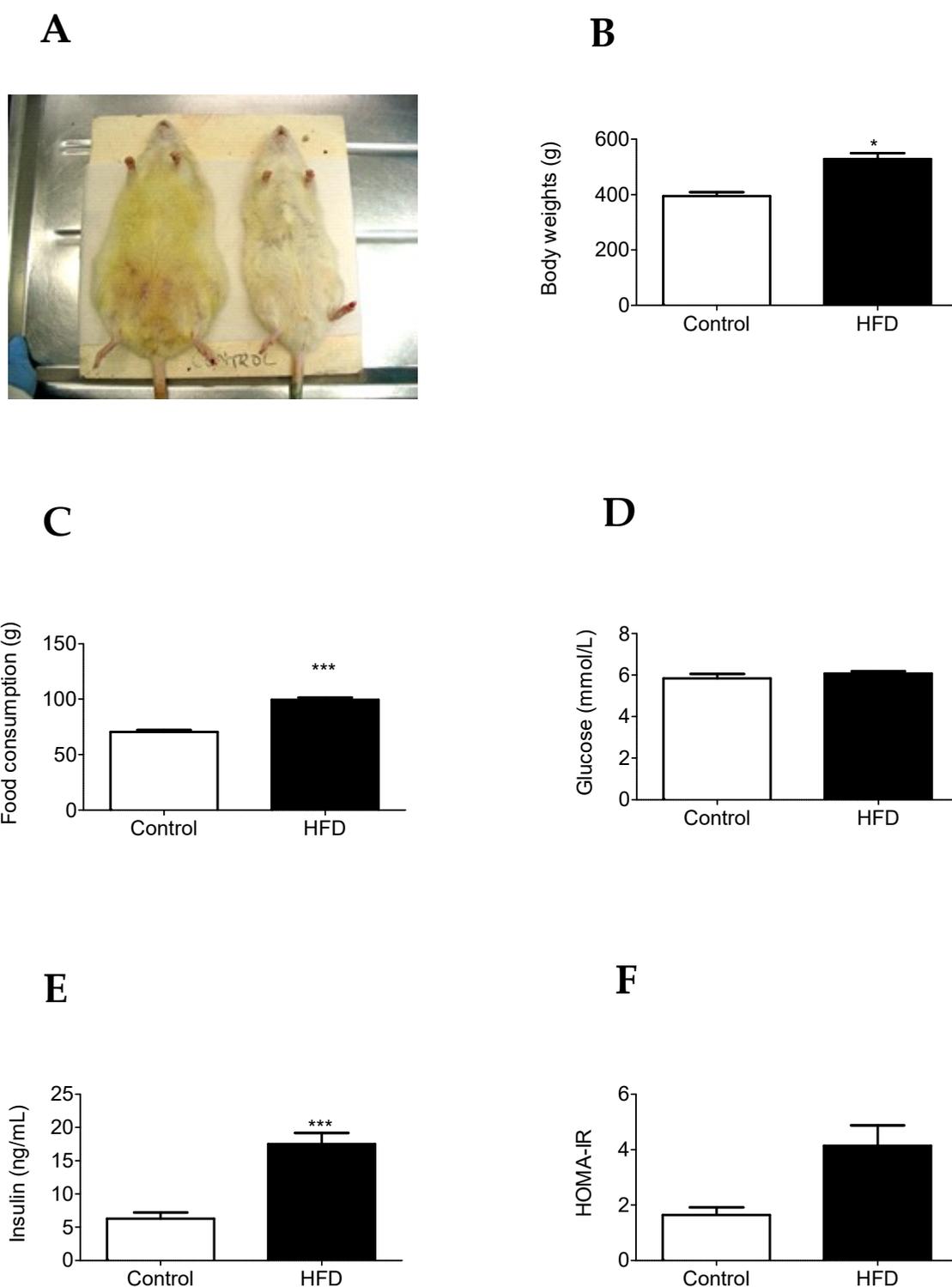
Establishment of OBIR rat model

Rats were divided into 2 groups, control diet group received standard maintenance rat chow (Atlas Animal Feed, Cape Town) and water *ad libitum* (n= 10). High-fat diet (HFD) and 15% sucrose/fructose in drinking water *ad libitum* (n= 10). Diets were administered for three months. Metabolic parameters were measured before termination.

Results

After 3 months of feeding with high fat diet, rats displayed increased body weight and food consumption (Figure 4S A, B, C), while there was no significant change in blood glucose levels between control and high fed diet. However, even though these rats demonstrated ability to

85 maintain normoglycemia, serum insulin concentrations at three months were increased in high fat
86 diet (Figure 2S D and E). The Homeostatic model assessment-insulin resistance (HOMA-IR) value of
87 the in the high fat diet rats were non-significantly increased when compared to control rats (Figure
88 2S, F).
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Figure 2S. High fat diet (HFD) induced obese and insulin resistance (OBIR) in male Wistar rats. Graphs shows body weight (A-B), food intake (C), glucose levels (D), insulin (E) and homeostatic model assessment-insulin resistance (HOMA-IR) (F). Wistar rats were treated with or without high fat diet for 12 weeks. Metabolic parameters were measured after 12 weeks. Results are presented as mean \pm SEM of Wistar rats (n = 10). * $p < 0.05$; *** $p < 0.001$ versus normal control.

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98 *Treatment of OBIR rat model*

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100 Since the OBIR model was partially established, in the current study we used only OBIR rats' vs
101 OBIR rats treated with different concentrations of GRE.

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103 **Food and water intake in OBIR rats**

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105 Twenty-four hour water and food intake were determined, in metabolic cages, prior to
106 commencement of treatment with Rooibos extract as well as 12 weeks after treatment with GRE.
107 Oral glucose tolerance test (OGTT) was performed after 12 weeks of treatment with GRE. After 16 h
108 of fast, rats were gavaged with GRE treatments 1 h (t = -60 min) before administration of a glucose
109 bolus (t = 0 min) at 2 g/kg glucose (50% Dextrose-Fresenius 50%). Plasma glucose concentrations
110 were determined at -60 and at 0, 15, 30, 60, 120 and 240 min, respectively, relative to the glucose
111 bolus (t = 0)

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118 **Results**

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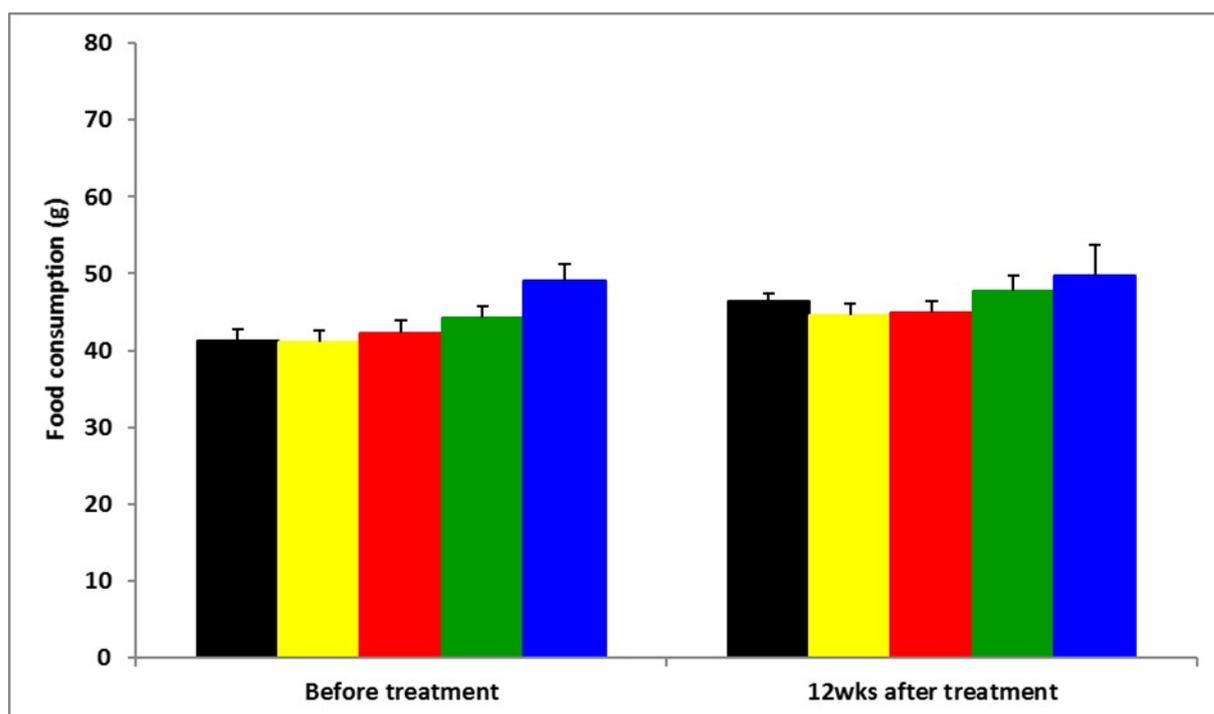
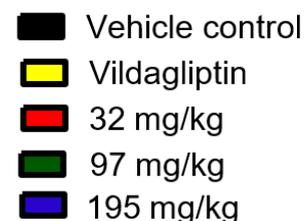
120 **Food consumption**

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127 **Figure 3S. Obese insulin resistant (OBIR) Wistar rats treated with an aspalathin-enriched green**
128 **rooibos extract (GRE). Graphs depict food intake before and after 12-week treatment period.**

129 The effect of different doses of GRE (32, 97 and 195 mg/kg BW) were assessed for their effect on food
130 intake. No significant results were observed within groups before and after treatment. Results are
131 presented as mean \pm SEM of OBIR rats (n = 12).

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136 **Water consumption**

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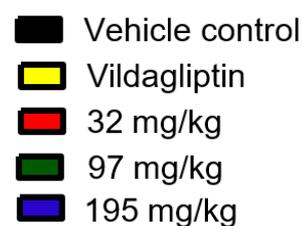
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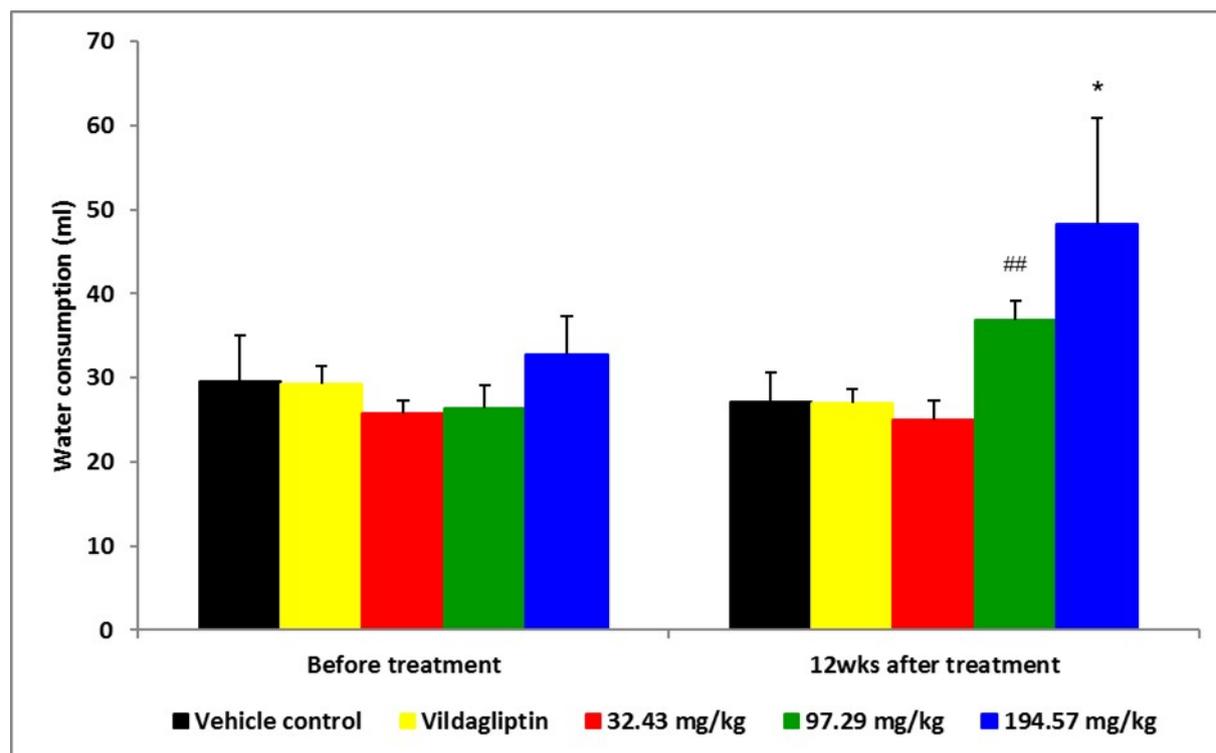
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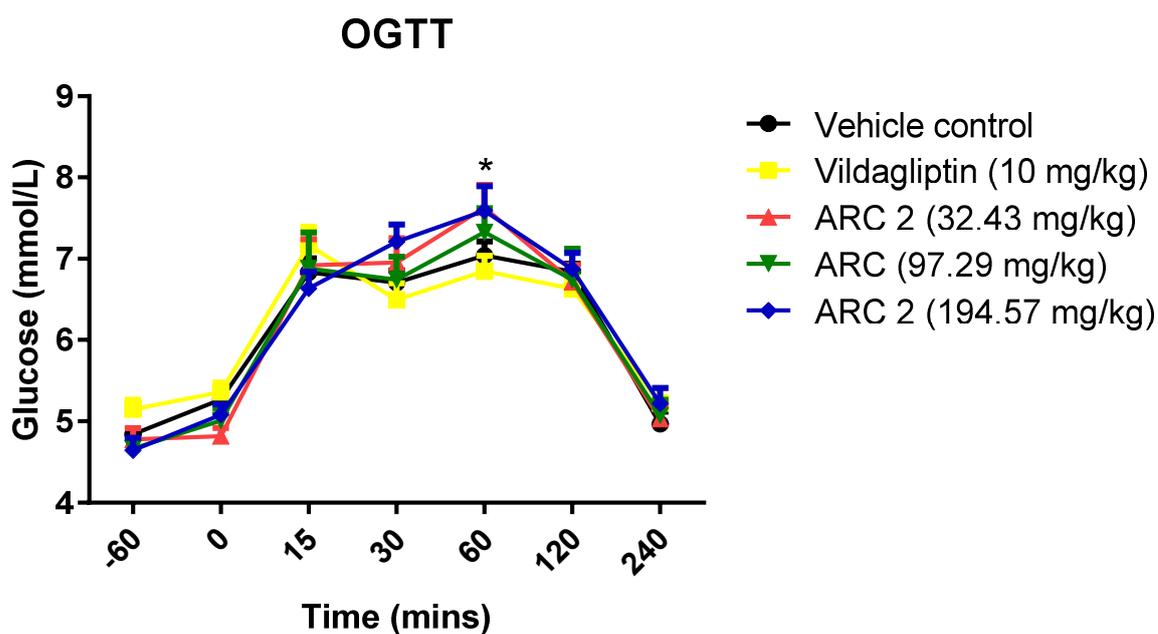


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145 **Figure 4S. Obese insulin resistant (OBIR) Wistar rats treated with an aspalathin-enriched green**
146 **rooibos extract (GRE). Graphs depict water intake before and after 12-week treatment period.**

147 The effect of different doses of GRE (32, 97 and 195 mg/kg BW) were assessed for their effect on
148 water intake. The highest dose (194.57 mg/kg/day) significantly increased water intake compared to
149 the baseline controls. Treatment group with 97.29 mg/kg/day displayed significantly increased
150 water intake compared to its baseline values. Results are presented as mean \pm SEM of OBIR rats (n =
151 12). * $p \leq 0.05$ versus respective baseline values; ## $p < 0.01$ versus its baseline values at 12 weeks after
152 treatment. Baseline indicates measurements taken before treatment.

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156 Figure 5S. Obese insulin resistant (OBIR) Wistar rats treated with an aspalathin-enriched green
 157 rooibos extract (GRE). Graphs shows oral glucose tolerance test (OGTT) after 12 weeks of
 158 treatment with GRE and after 16 h of fast. The effect of different doses of GRE (32, 97 to 195 mg/kg
 159 BW) were assessed for their effect on OGTT after 16 hours of fast) * $p \leq 0.05$ significance between
 160 Vildagliptin and GRE (194.57).

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