



Review

# Role of Hypoxia and Metabolism in the Development of Neointimal Hyperplasia in Arteriovenous Fistulas

Nirvana Sadaghianloo <sup>1,2,\*</sup>, Julie Contenti <sup>1,3</sup>, Alan Dardik <sup>4,5</sup> and Nathalie M. Mazure <sup>1,2</sup> 

<sup>1</sup> Centre de Méditerranéen de Médecine Moléculaire (C3M), Université Côte d'Azur, INSERM U1065, 151 Route de St Antoine de Ginestière, BP2 3194, 06204 Nice CEDEX 03, France; contenti.j@chu-nice.fr (J.C.); mazure@unice.fr (N.M.M.)

<sup>2</sup> Department of Vascular Surgery, Centre Hospitalier Universitaire de Nice, 06000 Nice, France

<sup>3</sup> Department of Emergency Medicine, Centre Hospitalier Universitaire de Nice, 06000 Nice, France

<sup>4</sup> Department of Surgery and the Vascular Biology and Therapeutics Program, Yale University, New Haven, CT 06520, USA; alan.dardik@yale.edu

<sup>5</sup> Department of Surgery, VA Connecticut Healthcare Systems, West Haven, CT 06516, USA

\* Correspondence: sadaghianloo.n@chu-nice.fr

Received: 4 October 2019; Accepted: 25 October 2019; Published: 29 October 2019



**Abstract:** For patients with end-stage renal disease requiring hemodialysis, their vascular access is both their lifeline and their Achilles heel. Despite being recommended as primary vascular access, the arteriovenous fistula (AVF) shows sub-optimal results, with about 50% of patients needing a revision during the year following creation. After the AVF is created, the venous wall must adapt to new environment. While hemodynamic changes are responsible for the adaptation of the extracellular matrix and activation of the endothelium, surgical dissection and mobilization of the vein disrupt the *vasa vasorum*, causing wall ischemia and oxidative stress. As a consequence, migration and proliferation of vascular cells participate in venous wall thickening by a mechanism of neointimal hyperplasia (NH). When aggressive, NH causes stenosis and AVF dysfunction. In this review we show how hypoxia, metabolism, and flow parameters are intricate mechanisms responsible for the development of NH and stenosis during AVF maturation.

**Keywords:** arteriovenous fistula; hypoxia; hypoxia-inducible factor; metabolism; neointimal hyperplasia

## 1. End-Stage Renal Disease and Arteriovenous Fistulas

### 1.1. Chronic Kidney Disease

Chronic kidney disease (CKD) describes the gradual loss of kidney function. At the stage of kidney failure, also called end-stage renal disease (ESRD) renal function must be replaced by either a new functioning kidney or an artificial kidney in order to maintain life. The main etiology of CKD is related to the diabetes-metabolic syndrome-obesity bundle and hypertension. As the prevalence of these diseases is growing exponentially worldwide, so is CKD [1]. In 2016, 124,675 new cases of ESRD were reported in the USA, with a total of nearly 500,000 patients receiving dialysis treatment and over 200,000 patients living with a kidney transplant [2]. Hemodialysis is the most commonly utilized therapeutic approach for the treatment of ESRD in the majority of countries [3]. Current hemodialysis devices run at a flow of 300 to 400 mL/min, meaning that the blood flow should be at least 500 to 600 mL/min [4]. As a comparison, the standard blood flow rate in a forearm radial artery is less than 40 mL/min [5]. Several types of vascular access have been designed and used for that purpose.

## 1.2. Arteriovenous fistulas (AVFs)

Hemodialysis arteriovenous fistulas (AVFs) are direct communications between a vein and an artery of the patient (Figure 1), first described in 1966, by Brescia et al. [6]. Bringing the arterial flow to the vein induces a large increase in the venous flow. With progressive maturation, which usually takes around six weeks, the venous wall becomes thicker, the venous diameter larger (>5–6 mm), and the venous flow reaches 600 to 1200 mL/min [5].

AVFs are considered the optimal vascular access for hemodialysis because of their potential for durability, lower risk of infection, and reduced need for intervention to ensure patency compared to grafts or catheters; however, literature and registries reveal a high prevalence of failure of newly placed fistulas never becoming functional (early failure) [4,7]. Based on the United States Renal Data System (USRDS), among AVFs placed between June 2014 and May 2016, 39% failed to mature sufficiently for use in dialysis. Of those that matured and were finally used, the median time to the first use was 108 days [2]. Late failure worsens the outcome of these AVFs, with a recent systematic review and meta-analysis reporting that primary patency rates of AVFs were 60% at 1 year and 51% at 2 years, meaning almost half of the patients had at least one reintervention on their AVF to maintain patency [8]. Secondary patency rates of 71% at 1 year and 64% at 2 years, meaning 30–40% of all AVFs were abandoned after a short time [8]. These reinterventions add to the burden of hemodialysis patients, already tired by the disease, the 3-times a week visit to the dialysis facility, the dialysis sessions themselves, and all the burden of vascular access monitoring [9]. Vascular access care is also responsible for a significant proportion of health care costs, in particular in the first year of hemodialysis. In the USRDS report, the cost of hemodialysis treatment was evaluated in the US at \$90,971 per patient per year [2].

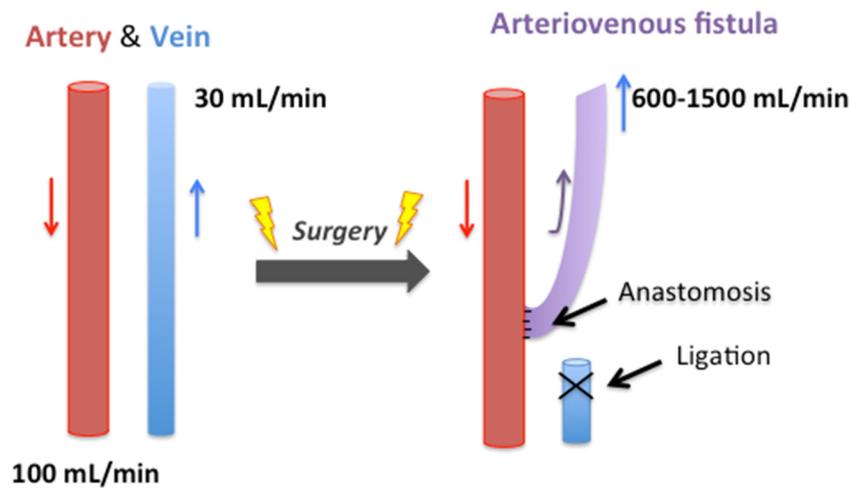
These data show that even the AVF, the best vascular access for hemodialysis, is clearly sub-optimal for a permanent treatment. The vascular access and dialysis community needs to better understand and correct the mechanisms leading to AVF failure.

## 2. Pathophysiology of AVFs

### 2.1. How AVFs Mature

During AVF creation, the surgeon directly connects the artery, a high-pressure pulsatile flow conduit, to a vein, which is ordinarily a low-pressure steady flow conduit (Figure 1). The pressure gradient results in an immediate increase in flow in both the artery and vein. The resulting hemodynamic change initiates a vascular remodeling response within both vessels. Pressure in the venous segment increases upon AVF creation and remains relatively constant during the time course of remodeling and thereafter. Outward remodeling is thought to be mediated by the venous endothelium and adventitia that sense these hemodynamic forces and integrates them to allow successful adaptation without loss of luminal area and vessel patency [10]. Venous diameter expansion is a critical element of outward remodeling and predicts the clinical success of AVF [11].

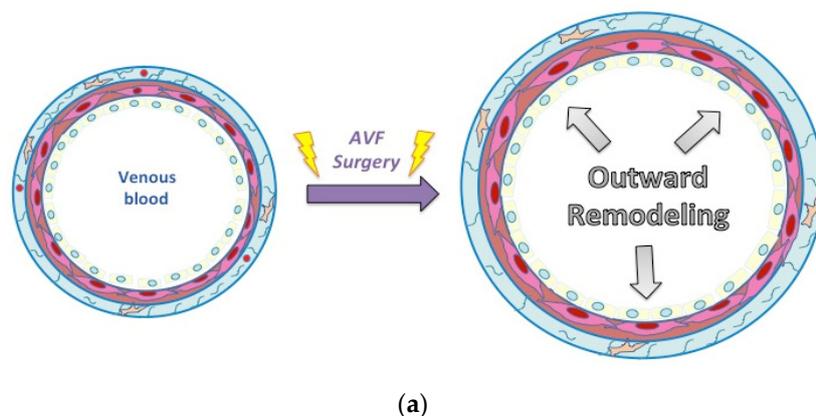
The venous wall is composed of three layers (Figure 2A) [12]. The external layer, called the adventitia, acts like a thin but resistant envelope to the vessel. It is composed of extracellular matrix (ECM), fibroblasts and immune cells. *Vasa vasorum*, small vessels, also run around the vein within the adventitia, to bring arterial blood flow to the venous wall [13]. The middle layer, called the media, is made of smooth muscle cells (SMC) and some extracellular fibers. The internal layer, also called intima, is composed of endothelial cells. During remodeling, hemodynamic changes and surgical injuries have an impact on all three layers of the venous wall. Endothelial and adventitial signaling induces, in turn, structural changes in the cells and the ECM, which affect the venous caliber and thickness (Figure 2B,C). Lack of outward remodeling and/or excess in wall thickening creates a disequilibrium that leads to AVF non-maturation and early failure (Figure 2D) [12].



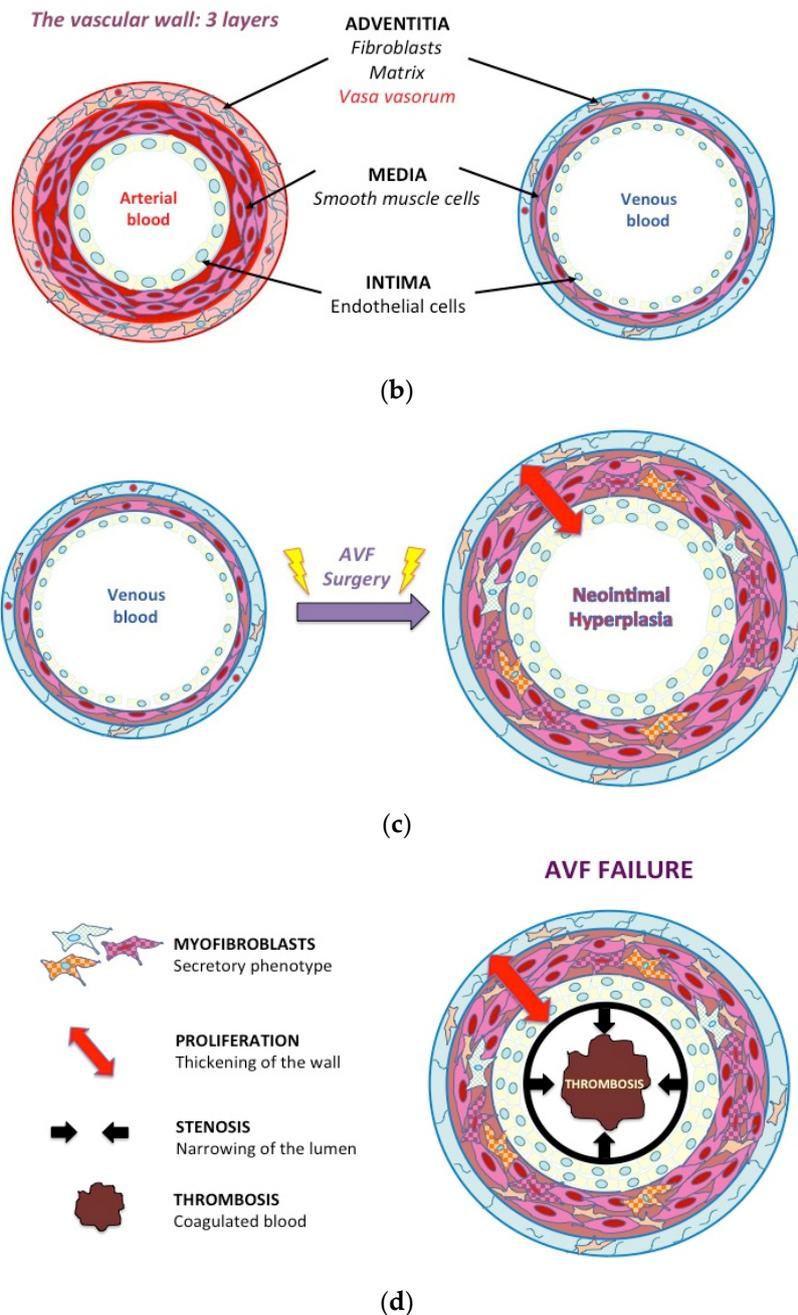
**Figure 1.** A fistula is direct communication between a vein and an artery, frequently on the upper limb of the patient. This communication is made by the surgeon with the help of a suture. The vein is dissected and ligated its extremity, and sewn to the artery. The arterial blood (red arrow) flows into the vein (blue arrow) through this anastomosis. The fistula matures, and the blood flow in the fistula (purple arrow) reaches around 1 L per minute.

## 2.2. Flow and Hemodynamic Parameters in AVF

The increased hemodynamics of arterial flow that increases vessel wall shear stress (WSS) are critical events after AVF creation that promotes AVF adaptation [14]. In a patient-specific end-to-side fistula, image-based computational fluid dynamics studies showed laminar flow within the arterial limb and a complex multi-directional and reciprocating flow field on the inner side of the swing segment in the proximal venous limb [15]. Wall thickening is predisposed to occur in the inner wall of the venous segment near the anastomosis and has a strong inverse correlation with magnitudes of shear stress, but is also related to flow patterns [16]. Although data is sparse in the venous vasculature and venous cells, studies on the arterial vasculature have shown that disturbed flow, with low and reciprocating WSS, is known to induce selective expression of atherogenic and thrombogenic genes that are pro-oxidant, proinflammatory, procoagulant, and proapoptotic in endothelial cells [17,18], and to stimulate vascular smooth muscle cell migration and proliferation, all of which may enhance wall thickening [19–21].



(a)  
**Figure 2.** Cont.



**Figure 2.** Mechanisms of venous wall remodeling. (a) Similar to the arterial wall, the venous wall is composed of three layers: the external layer, called the adventitia, composed of extracellular matrix (ECM), fibroblasts, immune cells, and *vasa vasorum*. The middle layer, called the media, is made of smooth muscle cells (SMC) and some extracellular fibers. The internal layer, also called intima, is composed of endothelial cells. (b) and (c), upon arteriovenous fistula (AVF) creation, in response to the altered hemodynamic environment, several structural changes occur in the vessel wall. Surgery induces endothelial denudation, matrix reorganization to accommodate outward expansion but also vascular SMC proliferation and migration, contributing to NH formation. (d) Aggressive intimal hyperplasia induces stenosis (narrowing) of the vessel that can provoke thrombosis (occlusion).

### 3. Neointimal Hyperplasia in AVF

#### 3.1. Definitions

Wall thickening is the adaptation of the vessel wall to increased pressure and abnormal WSS. This process involves the expansion of all the vessel layers via ECM deposition, cell proliferation and migration [22]. The thickening of the venous wall composed of these matrix proteins and proliferative and secretive cells is called neointimal hyperplasia (NH). Excessive neointimal hyperplasia causes stenosis. As a consequence of stenosis, the blood flow may be diminished and the AVF becomes unusable for dialysis. In extreme cases, stenosis may cause thrombosis, a complete occlusion of the vessel because of non-circulating blood (Figure 2D). Several types of cells are involved in NH, including ECs, SMCs, and adventitial fibroblasts [23].

#### 3.2. Role of Endothelial Cells

The release of chemotactic and inflammatory mediators from the endothelium during surgical manipulation and hemodynamic variation are substantial during the initial phase of adaptation [24]. Directly after AVF creation, high magnitudes of arterial flow result in both passive vascular distention and nitric oxide (NO) synthesis by endothelial cells with subsequent vascular SMC relaxation, resulting in acute vasodilation [25,26]. NO is produced by endothelial nitric oxide synthase (eNOS) and may contribute to adaptive vein wall remodeling through its anti-inflammatory, anti-thrombotic, and antiproliferative properties. Both eNOS and inducible nitric oxide synthase (iNOS) are upregulated in the AVF, and pharmacological inhibition of eNOS results in increased monocyte chemoattractant protein-1 and interleukin (IL) 8, leading to NH [27]. Endothelin-1 (ET-1) is an inflammatory mediator of vasoconstriction and endothelial proliferation. ET-1 expression is upregulated in the venous wall and within areas of NH in AVF as well as in the plasma of patients with chronic renal failure and hemodialysis. ET-1 may mediate wall thickening in response to localized hemodynamic forces [28,29]. Selectins facilitate leukocyte adhesion. P-selectin is present on ECs and platelets, and E-selectin is present on ECs. Intercellular adhesion molecule (ICAM) and vascular cell adhesion molecule (VCAM) facilitate additional binding and migration [30]. P-selectin and E-selectin expression are both upregulated early after AVF creation, followed by decreased P-selectin expression after 1 month [27]. VCAM-1, but not ICAM-1, is highly expressed in thrombosed and stenotic AVF [31].

#### 3.3. Role of Smooth Muscle Cells

The canonical origin of proliferating cells in NH is the vessel media that contains SMC. Endothelial and smooth muscle cell injury, consequences of hemodynamic stress and mechanical injury, results in the migration of SMCs from the media into the intima, where the SMC proliferate and differentiate into a secretory phenotype (myofibroblasts). This process of injury, followed by migration, proliferation and differentiation, is orchestrated by a large number of mediators, such as cell cycle regulators (p27-p16 [32], p38 mitogen-activated protein kinase [33]) and growth factors (vascular endothelial growth factor (VEGF) [34], platelet derived growth factor (PDGF) [27], basic fibroblast growth factor (bFGF) [35,36], insulin-like growth factor-1 (IGF-1) [37], etc.). Expression of adhesion molecule  $\beta$ -catenin and proto-oncogene c-Myc is increased one week after AVF creation, correlating with decreased adhesion molecule N-cadherin, associated with vascular SMC proliferation [38]. Vascular SMC may also become resistant to NO, decreasing vascular SMC relaxation and preventing AVF maturation by reducing outward remodeling [39]. As shown recently, mature SMCs are keys to the equilibrium between medial wall thickening (proliferation of differentiated SMCs that promotes maturation) and neointimal hyperplasia (dedifferentiated SMCs that cause NH and failure) [40]. Interestingly, vascular SMCs from the proximal artery may also contribute to venous NH: in a murine AVF model, increased Notch signaling could drive migration of these cells to the venous outflow tract [41].

### 3.4. Role of Fibroblasts

More recent studies have shown that the adventitial fibroblasts are critical during venous adaptation to arterial flow and help maintain venous wall integrity and hemostasis after surgical creation of the AVF [23,42]. Fibroblast precursors residing in the venous adventitia sense the abrupt changes in mechanical forces produced by the arterial flow to rapidly adjust their genomic expression program to help increase vascular resistance [43]. They differentiate into myofibroblasts, form matrix bundles of contractile microfilaments and extensive cell-to-matrix attachment sites, and secrete matrix metalloproteinases (MMPs), collagen, and ECM proteins that strengthen the fistula wall. Recent data suggest that NH in AVFs consists mainly of smooth muscle  $\alpha$ -actin-positive, vimentin-positive and desmin-negative myofibroblasts that have probably migrated from the adventitial layer [23,44]. Analysis of venous segments of failed AVF has shown increased adventitial fibrosis, myofibroblast activation, and capillary rarefaction [45].

### 3.5. Role of Inflammatory Cells

Inflammatory cells and cytokines, such as CD68-positive macrophages, CD3-positive lymphocytes, IL-6, IL-8, and monocyte chemo-attractant protein-1 are also known to be part of the NH process in AVFs [29,46–48]. This inflammatory state, exacerbated by CKD and dialysis, leads towards the up-regulation of numerous growth factors and cytokines, such as insulin-like growth factor-1, platelet-derived growth factor and basic fibroblast growth factor [27,37]. They all play a significant role in the stimulation of cell proliferation and migration. Even transforming growth factor- $\beta$ 1, usually known for its anti-inflammatory properties, is produced by ECs, vascular SMCs and inflammatory cells within the venous wall, leading to ECM deposition. These cells also participate in NH in AVFs [36,37,49].

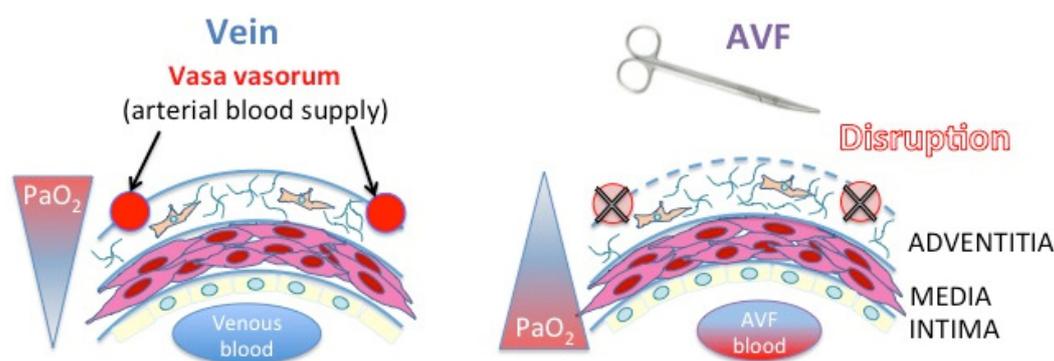
### 3.6. Role of Extracellular Matrix

The equilibrium between ECM degradation and accumulation is closely related to the maturation process of AVFs. In a murine AVF model, Hall et al. showed that initial ECM degradation occurs early after AVF creation, coincident with an early increase in expression of matrix metalloproteinases (MMP-2 and -9) and tissue inhibitor of metalloproteinase-1 (TIMP-1) [50]. Although the role of TIMP-1 is not clear, an imbalance in MMP and TIMP may contribute to AVF failure [51]. By day 7, there is an increase in collagen and elastin production and a change in patterns of MMP expression. By day 21, expression of MMPs is reduced and expression of larger structural and non-collagenous matrix proteins, such as fibronectin and perlecan, is increased. Other studies have shown that decreased expression of MMP-1, MMP-3, and MMP-9 is linked to increased AVF failure and stenosis [52]. Giachelli et al. have found that the expression of osteopontin (OPN) was up-regulated in the development of NH in a model of arterial balloon-injury. OPN was originally identified as matrix protein in bones, facilitating cell adhesion and migration, but this secreted protein has also been found in several experimental models of AVF maturation [50,53]. Its pro-inflammatory properties may favor vascular remodeling and development of NH [54].

### 3.7. Dissection Injury: Ischemia and Oxidative Stress

Hemodynamic changes and surgery-induced mechanical injury to the venous wall have been proposed as the etiology of juxta-anastomotic stenosis and maturation failure of AVF; however, a more recent theory proposes that hypoxia and oxidative stress may also play a role. Starting from the observation that during traditional AVF creation the vein is dissected free from the surrounding tissue, and that this same dissected segment is responsible for the majority of stenosis, researchers hypothesized that the rupture of the vasa vasorum during surgical dissection may induce hypoxia, low oxygen ( $O_2$ ) concentration, in the venous wall [12,34] (Figure 3). Additionally, the  $O_2$  partial pressure ( $pO_2$ ) in the venous blood is approximately 35–45 mmHg. However, after AVF creation,  $pO_2$  in the

fistula increases to the level of arterial  $pO_2$  (73–100 mmHg). Thus it is possible that interruption of the arterial blood supply followed by the sudden increase in luminal  $pO_2$  causes an ischemia-reperfusion phenomenon within the venous wall. Both of these phenomena provoke oxidative stress and stimulate numerous signaling pathways [36]. Interestingly, Joddar et al. have shown that arterial levels of oxygen stimulate NH in veins via a ROS-dependent mechanism [55]. In addition, two computational studies suggest that hypoxic segments of vessels may be linked to hemodynamic patterns and may correlate with stenosis [56,57]. Thus the two pathophysiological theories, flow and ischemia, may be related.



**Figure 3.** Changes in the venous wall after surgery: The *vasa vasorum* disruption creates a new oxygen gradient. The *vasa vasorum* in the adventitia provides oxygenated blood to the wall, whereas the intima is in contact with a venous blood low in oxygen. During surgery, the vein is dissected, disrupting the *vasa vasorum*, and potentially decreasing oxygen levels. On the other side, oxygenated arterial blood mixed to venous blood potentially creates an inversion of the oxygen gradient.

#### 4. The Hypoxic Pathway in AVF

##### 4.1. Hypoxia-Inducible Factors (HIFs), Keys to the Hypoxic Environment

Maintaining oxygen homeostasis is essential for the survival of all aerobic organisms. Therefore, a change in  $pO_2$  is a major physiological stimulus that elicits rapid cellular responses. HIFs are key transcription factors that regulate the cellular response to hypoxic injury by stimulating the transcription of genes that help restore  $O_2$  and maintain effective energy production [58]. HIFs belong to the large family of basic-helix-loop-helix transcription factors and bind DNA as a heterodimer composed of one oxygen-regulated  $\alpha$ -subunit (isoforms HIF-1 $\alpha$ , HIF-2 $\alpha$ , or HIF-3 $\alpha$ ) and two  $\beta$  stable subunits (HIF-1 $\beta$  or aryl hydrocarbon receptor nuclear translocator (ARNT), and HIF-2 $\beta$  or ARNT2) [59,60].

##### 4.2. Oxygen-Dependent Regulation of HIFs

The discovery of molecular mechanisms behind oxygen-dependent regulation of HIFs was rewarded in 2019 by the Nobel Prize in Physiology or Medicine. In well-oxygenated cells, the HIF- $\alpha$  subunits are constantly and rapidly degraded by hydroxylation of proline residues within the oxygen-dependent degradation domain of HIF- $\alpha$ , with one of the shortest half-lives (less than 5 min) among cellular proteins (Figure 4A) [59,61,62]. Prolyl hydroxylation targets the HIF- $\alpha$  proteins for proteasomal degradation by promoting their interaction with the protein von Hippel-Lindau (pVHL) (Figure 4A) [59,61,63–65]. This post-translational modification of HIF- $\alpha$  is carried out by a family of prolyl hydroxylase enzymes, the prolyl hydroxylase domain proteins (PHD1, PHD2, PHD3) (Figure 4A) [61,62,66]. Since these enzymes bind oxygen directly, they are critical oxygen sensors within the hypoxic response pathway [67]. In hypoxia, these enzymes being inactivated, HIF- $\alpha$  proteins are not degraded and accumulate in cells (Figure 4B) [68]. An additional hydroxylation of an asparaginyl moiety at the end of the COOH-terminal transactivation domain of HIF-1 $\alpha$  or HIF-2 $\alpha$  subunit is catalyzed by an enzyme called factor-inhibiting HIF-1 (FIH-1) [69,70]. In normoxia, FIH-1 abrogates HIF transcriptional activity by inhibiting the binding of coactivators such as p300 and

C-AMP Response Element-binding protein [71–73]. Under hypoxia, PHD inactivation causes HIF- $\alpha$  accumulation, and FIH-1 inactivation allows HIF transcriptional activity. While the HIF- $\beta$ /ARNT subunits are constitutively expressed in the nucleus, the HIF- $\alpha$  subunits found in the cytoplasm have sensitivity towards O<sub>2</sub>. When the HIF- $\alpha$  subunit is stabilized and active, it leaves the cytoplasm to enter the nucleus, binds its  $\beta$ -subunit and cofactors, and induces the transcription of a large number of genes (1–2% of all human genes) (Figure 4B) [61]. All these genes have in common a hypoxia-responsive element (HRE: 5' -RCGTG-3') as an enhancer, either in their promoter, in the 3' untranslated region, or in introns [58–76]. These genes have key biological roles such as cell survival (*ADM* [77], *cyclin D1* [78], *BNIP3* [79], *NIX* [80]), cell proliferation (*cyclin G2* [81], *IGF2* [82], *TGF $\alpha$*  [83], *NOS2* [84]), cell motility (*AMF/GPI* [83], *LRP1* [81]), cytoskeletal structure and ECM (*vimentin* [83], some keratins [83]), erythropoiesis (*EPO*) [85], pH regulation (*carbonic anhydrase 9 and 12*) [86], nucleotide metabolism (*adenylate kinase 4*) [87], amino acid metabolism (*transglutaminase 2*) [81], iron metabolism (*transferrin* [88], *ceruleoplasmin* [89]), and glucose and energy metabolism (*HK2* [90], *GLUT1* [91], *GAPDH* [92], *LDHA* [93], *leptin* [94]).

A number of HIF-target genes also control angiogenesis: (VEGF [95], VEGF receptor (VEGFR)-1 and -2 [96]), vascular remodeling (angiopoetin-2 and -4, TIE1, TIE2) [97] and vascular tone (HO1 [98], NOS2 [84], ET1 [81]). As such, HIF pathways have been of great scientific interest for more than two decades, in particular in the fields of development [99,100], cancer [101,102] and cardiovascular diseases [103].

#### 4.3. Oxygen-Independent Regulation of HIFs

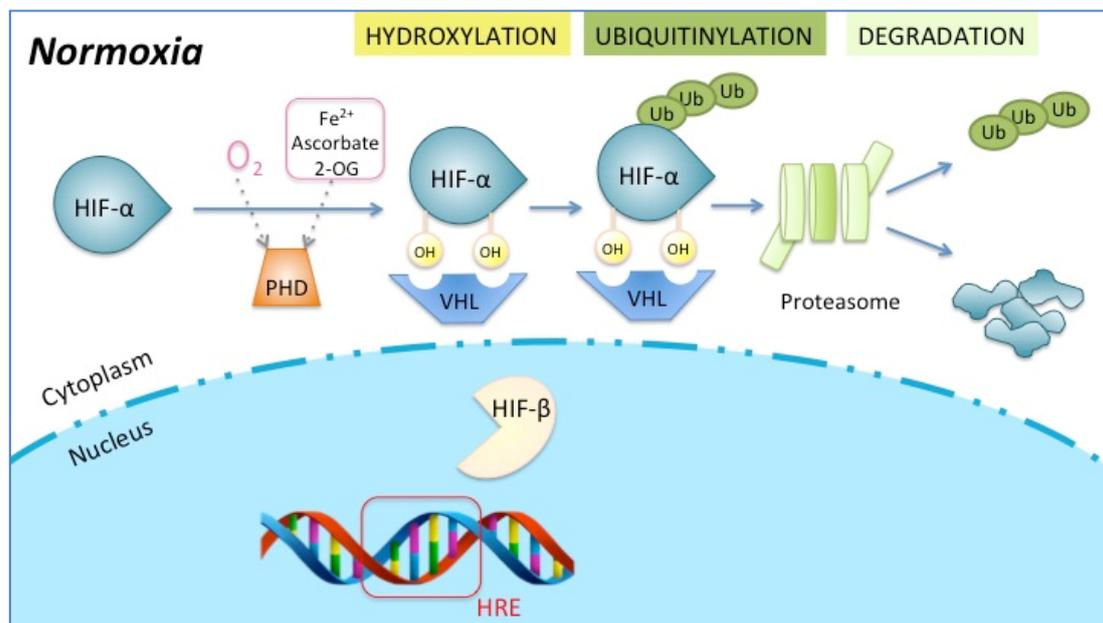
HIF- $\alpha$  can also be stabilized in normoxic conditions as a consequence of cell injury and oxidative stress. Several O<sub>2</sub>-independent mechanisms of HIF regulation have been identified such as receptor for activated C kinase 1 (RACK1) [104], carboxyl terminus of Hsc70-interaction protein (CHIP) and heat shock protein-70 (HSP70) [105], hypoxia-associated factor (HAF) [106], and Sirtuin-7 (SIRT7) [107]. They all mediate ubiquitination and proteasomal degradation. HIF-1 $\alpha$  responds also to non-hypoxic stimuli [68], including hormones like insulin [108,109], growth factors such as IGF-1 [110] or PDGF [111], vasoactive peptides such as nitric-oxide (NO) [112,113] or angiotensin 2 [111,114] and cytokines such as tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) [115]. It is assumed that these O<sub>2</sub>-independent stimuli contribute to an autocrine role in proliferation, survival or tissue repair, allowing early adaptation to variations in the cell environment.

Since many of these stimuli induce reactive oxygen species (ROS) production as part of their signaling cascade, it has been hypothesized that ROS participate directly in HIF stabilization [116]. Incubation of cells with H<sub>2</sub>O<sub>2</sub> leads to the stabilization of HIF- $\alpha$  proteins and activation of HIF target genes even when O<sub>2</sub> levels are high [117,118]. Furthermore, exposing keratinocytes to ultra-violet B irradiation activates HIF-1 $\alpha$  through generation of mitochondrial ROS [119]. Using junD-deficient cells that exhibit chronic oxidative stress, Gerald et al. discovered ROS-dependent iron-mediated regulation of PHD enzymes [120]. Since enhanced H<sub>2</sub>O<sub>2</sub> levels promote the oxidation of Fe(II) to Fe(III) through the Fenton reaction, it reduces the cellular pool of Fe(II) and increases even more drastically the proportion of active PHD. PHD and FIH-1 not only directly sense O<sub>2</sub> concentrations, as discussed above, but are also regulated by ROS (mostly H<sub>2</sub>O<sub>2</sub>) and metabolites through modulation of iron availability [120–122] and 2-OG accessibility [121], respectively. In addition to PHD inactivation, ROS has also been shown to act on HIF expression via activation of kinases [123]. ROS-induced HIF- $\alpha$  stabilization was reversed in various studies by iron supplementation, reducing agents or antioxidant compounds. Treating cells with anti-oxidant compounds, such as the free radical scavenger *N*-acetylcysteine [124–126], glutathione [126], or vitamins E and C [127,128], markedly attenuates HIF- $\alpha$  protein accumulation and expression of HIF target genes in various cell types, including hepatoma cells [124], vascular SMC [129], myocardial cells [127], gastric epithelial cells [125], renal tubuloepithelial cells [114], and macrophages [126]. Therefore, oxidative stress could trigger activation of HIF even in well-oxygenated conditions, thereby mimicking hypoxia signaling.

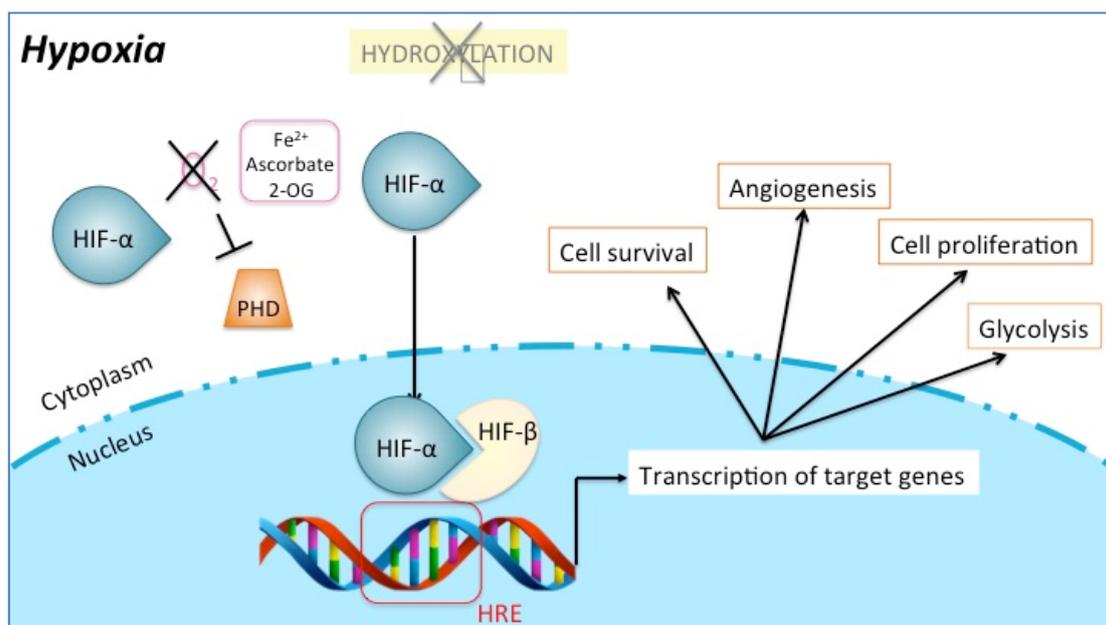
#### 4.4. HIF-1 vs. HIF-2

The expression of HIF-2 $\alpha$  was first believed to be restricted to blood vessels and in particular to vascular endothelial cells [85,130,131]. Since then, the expression of HIF-2 $\alpha$  has been found in several tissues, including hypoxic kidney, lung, colonic epithelia, hepatocytes, macrophages, muscle cells and astrocytes [132]. Some early studies showed that besides common target genes, each HIF- $\alpha$  isoform seems to have unique targets [133,134], and this transcriptional specificity may reside in the NH<sub>2</sub>-terminal transactivation domain, suggesting that HIF- $\alpha$  interactions with transcriptional co-factors are key to determine differential gene activation [135]. However, recent studies have shown that the mechanisms leading to selective gene transactivation are highly context specific. The individual HIFs have specific temporal and functional roles: HIF-1 drives the initial response to hypoxia (<24h) and HIF-2 drives the chronic response (>24 h) [136,137]. They can also exhibit antagonistic functions on the same final target. While HIF-1 $\alpha$  deletion in macrophages reduced the expression of iNOS and consequent production of NO [138], activation of HIF-2 $\alpha$  in the same cell type will reduce the pool of L-arginine (the source of NO) by inducing arginase 1 [139].

The selective expression of HIF-1 $\alpha$  vs. HIF-2 $\alpha$  is the result of a complex regulation process rather than the sole consequence of cell type or O<sub>2</sub>-dependant stabilization of HIF- $\alpha$  subunits. HIF-1 $\alpha$  vs. HIF-2 $\alpha$  expression can be regulated selectively at the level of transcription, translation, or protein stability [133]. Lin et al. have shown that chromatin remodeling via differential acetylation of core histones in the promoter of HIF-1 $\alpha$  or -2 $\alpha$  may regulate their transcription [140]. Another well-known regulator of HIF-1 $\alpha$  gene transcription is nuclear factor-KB (NF-KB), but it has not been shown to control HIF-2 $\alpha$  [141]. As above, the phosphatidylinositol-3-kinases/protein kinase B/mechanistic target of rapamycin (PI3K/AKT/mTOR) pathway regulates HIF $\alpha$  gene translation. However, Toschi et al. have shown that, in the same renal clear cell carcinoma cell line, HIF-1 $\alpha$  expression was both mTOR complex 1 and 2 (mTORC1, mTORC2)-dependent, whereas HIF-2 $\alpha$  was only mTORC2 dependent [142]. As for the stability of HIF- $\alpha$  subunits, HAF binds and leads HIF-1 $\alpha$  towards proteasomal degradation under normoxic and hypoxic conditions in a pVHL-independent manner, but does not decrease HIF-2 $\alpha$  levels [106]. In contrast, HAF binds HIF2 $\alpha$  at a distinct C-terminal region and promotes HIF-2 $\alpha$  transcriptional activity: cells switch from a HIF-1 $\alpha$  to a HIF-2 $\alpha$  transcriptional program [59,137]. In addition, HSP70 and CHIP bind and degrade HIF-1 $\alpha$  (but not HIF-2 $\alpha$ ) under conditions of prolonged hypoxia in cultured cells, whereas rapid reoxygenation destabilizes both HIF-1 $\alpha$  and HIF-2 $\alpha$  proteins via the PHD-pVHL-proteasome pathway (Figure 4) [59,105]. This is one mechanism by which HIF-2 is considered to be the effector of prolonged hypoxia, whereas HIF-1 drives the initial response to hypoxia [59]. Other mechanisms of HIF differential regulation include sirtuins (SIRT1–7), a family of redox-sensitive, NAD<sup>+</sup>-dependent deacetylases and/or adenosine diphosphate (ADP)-ribosyltransferases, which are known to regulate complex changes in gene expression, metabolism, and redox status in mammalian cells [143]. SIRT1 acts at a post-translational level: forming a complex with HIF-2 $\alpha$ , SIRT1 deacetylates lysine residues in the N-TAD, which enhances HIF-2 $\alpha$  transcriptional activity [59,144]. However, deacetylation of lysine residues in HIF-1 $\alpha$  results in the repression of the HIF-1 $\alpha$  transcriptional activity [145]; these opposing effects of SIRT1 may result in a switch towards either HIF-1 $\alpha$  or -2 $\alpha$  transcriptional programs in response to a metabolic switch in hypoxic tumors. The authors propose a mechanism of positive feedback in which under hypoxia HIF-1 $\alpha$  promotes glycolysis, reducing NAD<sup>+</sup>/NADH (nicotinamide adenine dinucleotide, oxidized and reduced forms, respectively) ratios and inhibiting SIRT1, thereby further increasing the HIF-1 $\alpha$  activity and presumably decreasing the HIF-2 $\alpha$  activity [145]. Given the disparate actions and differential regulation of HIF-1 and -2, targeting the hypoxic pathway needs to determine whether broad or selective inhibition is required, depending on the cell type and the expected effect.



(a)

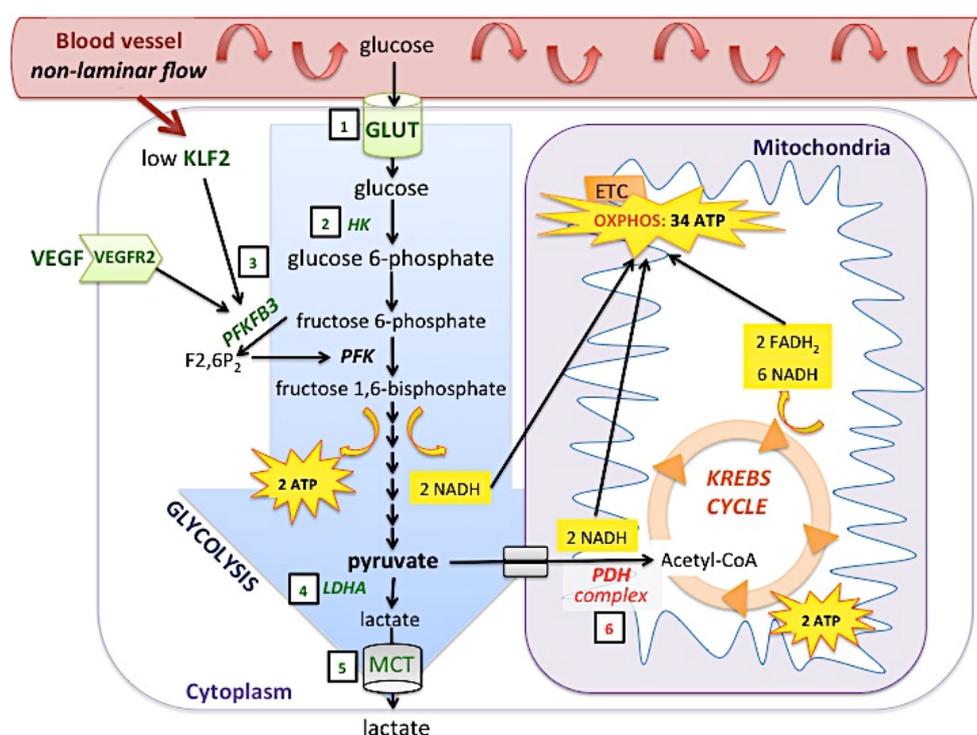


(b)

**Figure 4.** Molecular mechanisms of oxygen sensing and signaling. (a) In normoxia, hypoxia-inducible factor- $\alpha$  subunit (HIF- $\alpha$ ) is hydroxylated through a combination of  $\alpha$ -ketoglutarate, molecular oxygen ( $O_2$ ), one or more of the prolyl hydroxylase (PHD) isoenzymes and the asparaginyl hydroxylase factor inhibiting HIF (grey dotted arrow). The von Hippel-Lindau protein (VHL)-containing E3 ubiquitin ligase targets the  $\alpha$ -subunit for polyubiquitylation and degradation in the proteasome. (b) In hypoxia, low oxygen (black cross line) causes inactivation of PHD (black T arrow). The  $\alpha$ -subunit is no longer hydroxylated (grey cross line) and becomes stabilized. In the nucleus, HIF- $\alpha$  binds to the HIF- $\beta$  subunit and with the help of cofactors, the complex becomes transcriptionally active upon binding to the hypoxia-responsive element (HRE, red box) consensus sequence on the promoter of target genes. HIF-target genes are numerous, with key roles in cell survival, angiogenesis, proliferation, and glycolysis.

## 5. HIF and Cell Metabolism

Metabolism comprises a series of interconnected pathways that can function in the presence or absence of O<sub>2</sub>. Two of them, glycolysis and oxidative phosphorylation (OXPHOS) are represented in Figure 5. During glycolytic metabolism 1 molecule of glucose is converted into 2 molecules of pyruvate and generates 2 molecules of ATP and NADH. Lactate dehydrogenase A (LDHA) further reduces pyruvate into lactate [146]. During OXPHOS, 1 molecule of glucose is converted into 2 molecules of pyruvate, which are transported into the mitochondria, converted to acetyl coenzyme A and oxidized to CO<sub>2</sub> in the Krebs cycle. The NADH and FADH<sub>2</sub> generated in this process provide electrons to the electron transport chain in the inner mitochondrial membrane (respiratory cytochromes) [147]. Electrons are added to O<sub>2</sub> at complex IV (cytochrome c oxidase or COX), finally generating 38 molecules of ATP per molecule of glucose (Figure 5).



**Figure 5.** Glucose metabolism is regulated by hypoxia-inducible factors (HIFs). During glycolysis, glucose enters the cytoplasm through the glucose transporter (GLUT) and is transformed into glucose 6-phosphate by hexokinase (HK). Eight successive enzymatic reactions then occur to produce 2 molecules of pyruvate, 2 ATP and 2 NADH. At the end, cytosolic pyruvate has two possibilities: transformation into lactate by the enzyme lactate dehydrogenase A (LDHA)—lactate being then exported through the monocarboxylate transporter (MCT), or transferred into the mitochondria and transformed into Acetyl-CoA via the pyruvate dehydrogenase complex (PDH complex). In the second option, Acetyl-coA goes into the Krebs cycle to produce 2 ATP, 6 NADH, and 2 FADH<sub>2</sub>. NADH and FADH<sub>2</sub> will further be used in redox reactions in the electron transport chain (ETC) to produce more ATP via oxidative phosphorylation (OXPHOS). HIF intervenes in several steps during glucose metabolism: by inducing GLUT it allows more entry of glucose (1) and by inducing hexokinase (2) and PFKFB3 (3), it enhances glycolysis. Note that non-laminar blood flow decreases the expression of Kruppel-like factor 2 (KLF2), which has otherwise an inhibitory action on HIF-1 and PFKFB3 expression. VEGF and its receptor VEGFR2 are also induced by HIF-1 and enhance PFKFB3 expression. Further, pyruvate is pushed towards lactate transformation by up-regulation of LDHA (4) and some isoforms of MCT (5). At the same time, HIF prevents the PDH complex from transforming pyruvate into Acetyl-coA (6). Green: activation by HIF, Red: inhibition by HIF.

### 5.1. Metabolic Adaptation to Hypoxia

Metabolic adaptation to hypoxia involves mainly a switch from oxidative (aerobic) to glycolytic (anaerobic) metabolism by cells. With this energy compensation, the cell continues to generate ATP and, despite the hypoxia, try to meet the metabolic demands and maintain the expenditure of energy of normoxic conditions. During hypoxia, HIF-1 stimulates glycolysis through the up-regulation of key glycolytic genes, including *GLUT1*, *GLUT3*, *HK1*, *HK2*, and *LDHA* [148]. HIFs also reduce mitochondrial O<sub>2</sub> consumption by attenuating glucose oxidation: they up-regulate the expression of pyruvate dehydrogenase kinase, which inhibits the activity of the pyruvate dehydrogenase complex (PDH complex), a key enzyme regulating the entry of pyruvate into the Krebs cycle through conversion to acetyl CoA [149,150]. Finally, the subunit composition of COX is altered in hypoxic cells by increased expression of the COX4–2 subunit, which optimizes the COX activity under hypoxic conditions, and increases degradation of the COX4–1 subunit by the LON protease, which optimizes the COX activity under aerobic conditions [146].

### 5.2. Metabolism of Endothelial Cells

The metabolism of ECs has been largely studied in the domain of cancer and atherosclerosis but is also of great interest in the problematic AVF. All ECs (arterial, venous, microvascular) are preferentially glycolytic, rather than OXPHOS since 85% of their ATP is produced by converting glucose into lactate [151,152]. One hypothesis is that downregulation of OXPHOS metabolism protects them from excessive ROS and oxidative stress [152]. At the same time, they save O<sub>2</sub> to enhance the diffusion of oxygen to perivascular cells [152]. In addition, glycolysis produces less ATP but with faster kinetics, which is necessary for the rapid revascularization of hypoxic tissues when needed [153]. A hypoxic environment stimulates EC proliferation, and hypoxic proliferation of ECs increases glycolysis because of HIF-1 upregulation (both hypoxia- and growth factor- induced) [151].

### 5.3. Shear Stress, Hypoxia, and Metabolism in Endothelial Cells

Interestingly, hemodynamics and shear stress also play a role in EC metabolism. Laminar shear stress induces Kruppel-like factor-2 (KLF2) in ECs [154], whereas non-laminar (disturbed or turbulent) flow inhibits KLF2 (Figure 5). KLF2 is a zinc-finger transcription factor that maintains anti-proliferative and anti-inflammatory phenotypes of quiescent ECs. By inactivating p65 (subunit of NF-KB), KLF2 down-regulates cell adhesion (E-selectin) and prothrombotic molecules (thrombomodulin), inhibits VEGFR2, and up-regulates eNOS [155–159]. Doddaballapur et al. showed that laminar shear-stress induced KLF2 reduces glucose uptake and represses the key glycolytic enzyme 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase 3 (PFKFB3) [160]. This enzyme produces fructose-2,6-bisphosphate (F2,6P2), a strong allosteric activator of phosphofructokinase-1 (PFK1), a rate-limiting enzyme of glycolysis (Figure 5) [161]. Thereby, glycolytic metabolism is limited and blood flow keeps ECs in a resting state [162]. Kawanami et al. showed that KLF2 expression is also induced in hypoxic ECs, and over-expression of KLF2 inhibits HIF-1 $\alpha$  (but not HIF-2 $\alpha$ ) stabilization and downstream expression of ANG2, IL-8 and VEGF [163]. Conversely, inhibition of KLF2 increases HIF-1 $\alpha$  and its target-gene expression [163]. As such, KLF2 links hemodynamics, hypoxia and metabolism in ECs.

## 6. HIF and Vascular Diseases

In vascular pathology, hypoxia is typically the consequence of altered, destroyed or absent blood vessels leading to tissue ischemia. The mechanisms of the HIF response have therefore been studied in ischemic vascular diseases, such as myocardial infarction (consequence of obstruction of coronary arteries) or peripheral vascular disease (PAD, consequence of obstruction of arteries going to the limbs) [164,165].

Besides tissue ischemia, mechanisms of hypoxia and ischemia can occur within the vascular wall itself, with implications for diseases such as atherosclerosis [166], arterial aneurysm [167], chronic venous insufficiency [168] and thromboembolism [169,170]. There is growing evidence that the HIF pathway is implicated—for better or for worse—in the response of the vascular wall to inadequate oxygenation and/or increased cellular oxygen, a demand secondary to various stresses or injuries [171,172].

### 6.1. HIF and Atherosclerosis

Atherosclerosis is a condition in which fatty deposits (plaques) develop in the arterial wall, leading to stenosis and ultimately to thrombosis of the vessel [173]. Atherosclerotic plaques are mainly composed of cholesterol, other lipids, calcium, fibrin and inflammatory cells [174]. Oxygen diffusion through space-occupying atherosclerotic plaques could be limited due to wall thickening. Hypoxia could also originate from the high oxygen demand of metabolically active inflammatory cells, such as macrophage foam cells that play a pivotal role in this disease [175]. Detection of HIF-1 $\alpha$  and activation of its target genes occurs in atherosclerotic plaques [166,176]. During atherogenesis (plaque build-up), hypoxia increases angiogenesis [166], releases pro-inflammatory mediators [177], activates MMPs [178], forms reactive oxygen species [179], and oxidizes low-density lipoprotein [180,181]. Necrotic cores within atherosclerotic plaques can lead to plaque instability and thromboembolic complications. The HIF-1 $\alpha$  protein is found in tissue surrounding atherosclerotic arteries, inflammatory macrophages [166] and arterial SMCs around these necrotic cores [182,183]. Proangiogenic HIF-target genes, such as VEGF-A, are responsible for intra-plaque neoangiogenesis, contributing to plaque instability [184]. HIF-1 activity also alters the lipid metabolism of macrophages and vascular SMC [172,176,185]. Finally, besides hypoxia, several non-hypoxic stimuli of HIF have also been found in plaques and plaque macrophages, such as inflammation, ROS, vasoactive and thrombotic factors [129,186,187].

### 6.2. HIF and NH in Vascular Grafts

The role of the HIF pathway in the development of NH has also been studied. When atherosclerotic lesions occupy a long segment of the artery, surgeons frequently use a graft to bypass the lesion and provide blood to distal tissues. Either venous or prosthetic, these grafts are prone to NH. Although vein grafts and prosthetic grafts are not identical, because of different hemodynamics and oxygen tension, the characteristics of NH in vein grafts are similar to those of NH that form in AVFs [188]. In studies with animal models, grafts showed increased hypoxia within the vessel wall in regions of neointimal hyperplasia [189,190]. A study using a porcine model showed an association of HIF-1 $\alpha$  expression with the degree of neointimal hyperplasia in grafts, and suggested that reducing hypoxia might inhibit venous NH formation, possibly by reducing the phenotypic switch of fibroblasts to myofibroblasts [191]. Paradoxically, increased activation of the HIF pathway may also protect against NH formation. Nakao et al. studied the role of carbon monoxide, a physiologically important vasodilator that acts via cyclic guanosine monophosphate, in the formation of NH [192]. They showed that a vein graft placed immediately after harvest led to less NH than if the graft was rinsed with lactated Ringer for 2 h prior to surgical placement. However, if the lactated Ringer solution was saturated with carbon monoxide, it significantly inhibited NH compared to the untreated control. The effects of carbon monoxide in inhibiting NH formation seemed to be associated with increased HIF-1 $\alpha$  and VEGF expression a few hours after grafting. This effect, including the induction of VEGF, was reversed by treatment with YC-1, a HIF-1 $\alpha$  inhibitor, supporting the conclusion that these effects were mediated through the activation of the HIF pathway. These apparently contradictory studies suggest that, at least in the setting of vein graft NH, transient activation of the HIF pathway may protect the wall from hypoxia, whereas chronic activation may maintain the ongoing NH process. Carbon monoxide may also display vasoconstrictor effects depending on oxygen and metabolic homeostasis, including ROS production, highlighting the important role of metabolism in the vascular wall [193].

### 6.3. HIF and NH in Vascular Access

Both prosthetic arteriovenous graft and AVF failure may be associated with increased activation of the HIF pathway. Misra et al. investigated HIF-1 $\alpha$  and several target genes in stenotic segments from prosthetic grafts compared with AVFs [194]. The authors compared the expression of HIF-1 $\alpha$ , several MMPs and TIMPs, and macrophage migration inhibitory factor (MIF), a proinflammatory factor that regulates proliferation and migration of vascular SMC, which is affected by HIF-1 $\alpha$  in human vascular SMCs exposed to hypoxia [195]. Specimens from patients with prosthetic grafts expressed significantly more HIF-1 $\alpha$ , MIF, TIMP-1, pro-MMP-2, and pro-MMP-9 compared with control veins. Pro-MMP-9 was also up-regulated in AVFs compared with control samples. The expression of MIF and TIMP-1 was significantly increased in prosthetic graft specimens compared with AVFs. Expression of HIF-1 $\alpha$  in AVFs seemed to be higher than in controls although the differences reported in this study were not statistically significant. The authors concluded that there were major differences in the expression HIF-1 $\alpha$ , and proteins regulated by HIF-1 in specimens removed from patients with prosthetic grafts and AVFs. They consequently developed porcine and mouse models of vascular access failure and found that HIF-1 $\alpha$  and HIF-1-associated protein expression were increased in failed conduits compared with controls [196–198].

### 6.4. HIF Target Genes in Vascular Access

We previously studied oxidative stress and HIF-1 expression in a murine model of AVF maturation. Our team used a microarray analysis to show downregulation of 27 of the 83 genes involved in the OXPHOS pathway, downregulation of 12 of the 18 genes involved in mitochondrial long-chain fatty acid beta-oxidation, and 10 of the 13 genes involved in mitochondrial unsaturated fatty acid beta-oxidation, suggesting the presence of oxidative stress during early AVF maturation [199], and which was consistent with previous research [36]. We then showed increased excreted hydrogen peroxide, nitrotyrosine staining, and NOX-2 (**nicotinamide adenine dinucleotide phosphate oxidase - 2**) staining in the venous limb of the AVF during early maturation compared to control veins, confirming the presence of oxidative stress during AVF maturation. We also showed up-regulation of HIF-1 $\alpha$  mRNA and protein expression in AVFs. Immunofluorescence of the AVF showed increased immunoreactivity of HIF-1-associated heme oxygenase-1 (HO-1) and VEGF-A, at both days 3 and 7 compared with control veins [199].

Interestingly, recent research has shown that expression of the HIF-1-target *HO1* is related to AVF function [200–203]. HO-1 is a cytoprotective and rate-limiting enzyme responsible for heme degradation, generating free iron, biliverdin, and carbon monoxide. Biliverdin is subsequently converted to bilirubin by biliverdin reductase, and free iron is rapidly sequestered by ferritin [204]. Bilirubin is a free radical scavenger that blocks lipid peroxidation [205]. HO has antioxidant, anti-inflammatory, anti-apoptotic, and angiogenic functions through its reactive products [98,204]. HO-1 is an inducible isoform, whereas HO-2 is expressed constitutively. Patients with *HO-1* gene polymorphisms characterized by long GT repeats, resulting in less HO-1 production, were more likely to have worse AVF patency [200,201]. In a murine model of AVF, Junco et al. showed that *HO-1* gene expression was markedly induced in the vascular SMCs. *HO-1* knockout mice had reduced patency and increased expression of pro-inflammatory and pro-oxidant mediators, such as MCP-1, MMP-2, and MMP-9 [206]. Subsequently, Kang et al., using the same model, showed that the adventitial delivery of an adeno-associated viral gene *HO-1* improved AVF blood flow and decreased venous wall thickness [203]. A functional AVF also requires HO-2 [202]. Shear stress also regulates HO-1 activity, with high flow inducing HO-1 to generate NO and mitochondria-derived hydrogen peroxide. Low flow induces lower levels of HO-1 that lead to macrophage infiltration and superoxide production within the vessel wall. However, the authors did not report whether these results were HIF-dependent. While they suggest an important role for HO in promoting outward remodeling and in preventing NH [207], we may think that HIF expression in this setting would be beneficial.

VEGF-A is necessary at low concentrations to promote endothelial cell health, NO and prostacyclin production, vasodilation, antithrombosis, and suppression of SMC proliferation but promotes angiogenesis and vasculogenesis at high concentrations [13]. Increased VEGF expression has been associated with early AVF thrombosis in human patients [208], and the VEGF-936C/C gene polymorphism has been associated with a 5.54-fold increase in risk of late AVF thrombosis [209]. Adventitial delivery of a lentivirus inhibiting VEGF-A expression decreases cell proliferation, inhibits NH and increases patency in a mouse model of AVF [34]. Systemic VEGF receptor gene transfer in rats decreases carotid artery restenosis, suggesting the utility of targeting this pathway to inhibit NH [210].

## 7. HIFs as Therapeutic Targets

Several experimental studies have shown that inhibiting the HIF-1 target gene VEGF decreases NH [34]. Further studies are needed to understand if the HIF pathway should be inhibited or activated in this setting. These studies can evaluate the broad range of therapeutic agents tested in other fields, in particular in cancer, since pharmacological targeting of HIF has been a major field of research these past two decades [101,211].

### 7.1. O<sub>2</sub> Treatment

In a study using a rabbit iliac AVF model, Lata et al. demonstrated that NH is inhibited by the administration of 30% supplemental oxygen [212]. Based on the pattern of SMC proliferation, they suggested that 21 days of oxygen supplementation would be enough to control the initial burst of proliferation and subsequent NH. Further experiments by the same team demonstrated that HIF-1 $\alpha$  was stabilized, VEGFR-2 up-regulated, and plasma VEGF-A was increased in rabbits with AVFs placed in normoxia [213]. In addition, these markers were significantly decreased when rabbits with AVFs were placed in 30% oxygen chambers for 21 days after surgery. Plasma collected in the hyperoxic group induced significantly less SMC and EC proliferation in vitro [213]. These results support the role of hypoxia and HIF-1 stabilization in NH. However, translation of this treatment to humans would mean wearing a nasal cannula all day. It is likely that it would be very difficult to convince patients who already carry a heavy medical burden to use supplemental oxygen therapy. Paradoxically, several studies conducted in different non-vascular experimental settings and cell types have shown that absolute hyperoxia induces HIF stabilization and activation of HIF targets [214–218]. The putative mechanisms may involve oxidative stress and ROS production, consequences of disturbed mitochondrial metabolism. In the AVF, both hypoxia (induced by the rupture of the vasa vasorum) and relative hyperoxia (induced by an increase in blood PaO<sub>2</sub>) may cause HIF stabilization, with a balance between the two mechanisms over the maturation period potentially leading towards NH and AVF dysfunction. Future studies may use ROS scavengers and/or regulate other metabolic targets to discriminate between the two mechanisms.

### 7.2. Pharmacological Treatment

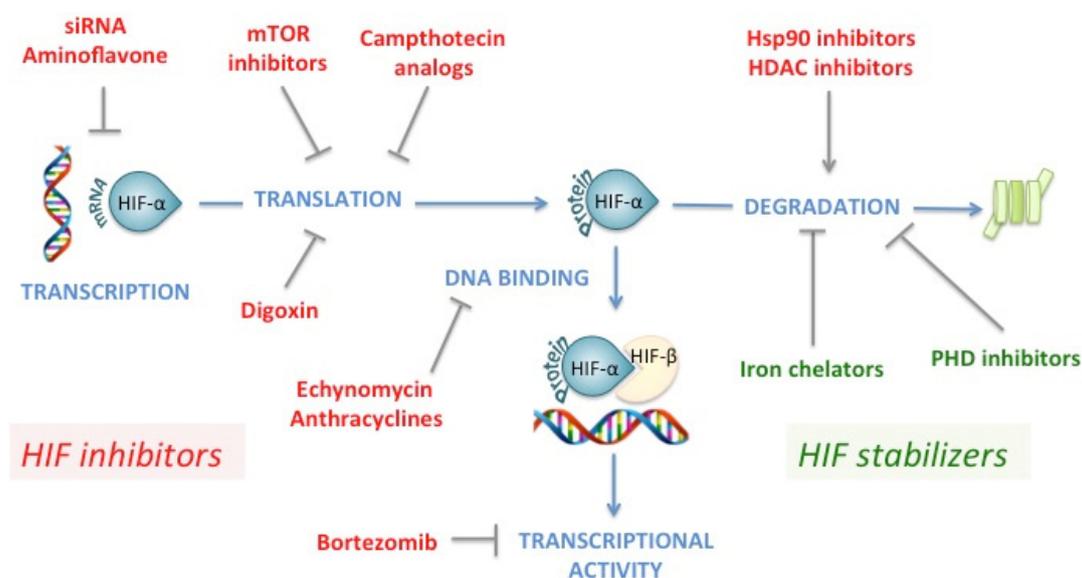
Many existing drugs or drugs used in clinical trials can specifically, but mostly non-specifically, target HIF at different points of its regulation [219,220].

#### 7.2.1. HIF Inhibition

According to their putative mechanism of action, HIF inhibitors can be divided into agents that modulate (Figure 6):

- HIF transcriptional activity, such as the FDA-approved chemotherapeutic drug PD-341 (Bortezomib, impairs binding of p300 to HIF1- $\alpha$  by enhancing the binding of FIH-1 to HIF1- $\alpha$ ) [221],
- HIF- $\alpha$  mRNA expression, such as the experimental compounds EZN-2968 (antisense oligonucleotide) [222] and aminoflavone (ligand of aryl hydrocarbon receptor) [223],

- HIF- $\alpha$  protein translation, such as FDA-approved chemotherapeutic drug topotecan (camptothecin analog, inhibitor of topoisomerase I) [211,224], experimental compound EZN-2208 (derived from irinotecan, another camptothecin analog) [225], FDA-approved cardiac glycoside digoxin [226], and FDA-approved chemotherapeutic drugs temsirolimus and everolimus (mTOR inhibitors) [227–229],
- HIF- $\alpha$  protein degradation, such as Hsp90 inhibitors [230,231] or histone deacetylase (HDAC) inhibitors [232],
- HIF DNA binding, such as echinomycin (a cyclic peptide of the family of quinoxaline antibiotics) [233] and anthracyclines (such as the FDA-approved chemotherapeutic drug doxorubicin) [234].



**Figure 6.** Pharmacological treatments targeting the HIF pathway. *Green: stabilization of HIF- $\alpha$ , red: destabilization of HIF- $\alpha$ . Blue arrows: physiological process, grey arrows: activation, grey T arrows: inhibition.*

### 7.2.2. HIF Activation

One of the easiest ways to stabilize HIF- $\alpha$  is to act on the repressing enzymes PHDs and FIH-1. These enzymes being dependent on Fe(II) and 2-OG, iron chelators such as desferioxamine [34] and 2-OG analogs such as dimethylxalylglycine (DMOG) [63] have been used in many experimental studies to stabilize HIF-1 $\alpha$ . Importantly, these therapeutics are not isoform specific as the compounds inhibit all PHDs and therefore, all HIF- $\alpha$ . In the setting of atherosclerosis and treatment of PAD, experimental studies evaluated intramuscular injection of AdCA5 and intravenous administration of bone marrow-derived angiogenic cells that were cultured in the presence of the PHD inhibitor DMOG. The combined therapy increased perfusion, motor function and limb salvage in aged mice subjected to femoral artery ligation [165,235]. Based on these promising findings, a Phase I dose-escalation human clinical trial was performed using an adenoviral delivery system to deliver the active form of HIF1- $\alpha$  into the lower extremity of patients with critical limb ischemia. In addition, the therapy was well tolerated [236]. However, in a trial evaluating the efficacy of intramuscular administration of three different doses of Ad2/HIF1A/VP16 on walking time in patients with PAD and intermittent claudication, there was no significant clinical improvement when comparing the placebo and each HIF1- $\alpha$  dosage group [237]. This led to the conclusion that gene therapy with intramuscular administration of Ad2/HIF1A/VP16 was not an effective treatment for patients with intermittent claudication [237]. Although the reasons underlying failure remain unclear, concerns include the efficacy of gene transfer, including whether the gene transfer was increasing HIF to sufficient levels in the critical tissue

compartment. For the treatment of CKD-induced anemia, promising compounds targeting PHD (Roxadustat, Vadadustat, Daprodustat, Molidustat) have been tested in patients with significant improvement of their hemoglobin level (mainly as a consequence of HIF-1-induced expression of EPO) [238–240]. These inhibitors are likely to become an important tool for the management of anemia in patients with CKD, where the oral route is a significant advantage. However, long-term safety of HIF activation must be determined, in particular with regards to HIF pathway tumorigenicity. Further studies are ongoing [238].

## 8. Conclusions

Increasing evidence shows that in AVFs, NH induced by upstream hemodynamic changes is linked to downstream metabolic disruption, involving the regulation of the response to hypoxia. Unfortunately, the common denominator of most HIF inhibitors and activators described so far is the lack of specificity, which indicates that they inhibit multiple targets and that HIF inhibition cannot be easily separated from other activities exerted by these agents [241]. Moreover, the majority of drugs primarily target HIF-1, although they may also act on HIF-2. One of the challenges of pharmacologically targeting the HIF pathway is to identify which isoforms are responsible for the disease and to find an isoform-specific drug. One of the most striking advances in the field is the successful targeting of HIF-2 in renal cell carcinoma, using a small molecule called PT2399, specifically designed to inhibit the binding of HIF-2 $\alpha$  to the HIF- $\beta$  subunit [242–245].

Besides finding HIF specific, isoform specific molecules, one major challenge will be to target delivery of these drugs to specific organs, to limit off-target and secondary effects.

**Author Contributions:** N.S., J.C., A.D., and N.M.M. conceptualized, wrote, and reviewed the article.

**Funding:** N.S.: J.C. and N.M.M. were supported by grants from the Fondation Flavien, the Fondation ARC, La Ligue contre le Cancer, Fondation de France, and the Société Française de Chirurgie Vasculaire et Endovasculaire. A.D. was supported by NIH grants R01HL128406 and R01HL1444476.

**Conflicts of Interest:** The authors declare no conflict of interest.

## Abbreviations

ADM	Adrenomedullin
ADP	Adenosine DiPhosphate
AKT	Protein kinase B
AMF/GPI	Autocrine Motility Factor/Glucose-6-Phosphate Isomerase
AMP	Adenosine MonoPhosphate
ANG	ANGiopoietin
ARNT	Aryl hydrocarbon Receptor Nuclear Translocator
ATP	Adenosine TriPhosphate
AVF	ArterioVenous Fistula
bFGF	basic Fibroblast Growth Factor
BNIP3	BCL2/adenovirus E1B 19 kDa protein-Interacting Protein 3
CHIP	Carboxyl terminus of Hsp70-interacting protein
CKD	Chronic Kidney Disease
COX	Cytochrome C Oxidase
CREB	C-AMP Response Element-binding protein
DMOG	DiMethylOxalylGlycine
DNA	Deoxyribonucleic acid
EC	Endothelial Cells
ECM	Extra-Cellular Matrix
eNOS	endothelial Nitric Oxide Synthase
EPO	ErythroPOetin
ESRD	End-Stage Renal Disease

ET-1	Endothelin-1
FAD	Flavin Adenine Dinucleotide
FDA	Food & Drug Administration
FIH	Factor Inhibiting HIF
GLUT	GLUcose Transporter
HAF	Hypoxia-Associated Factor
HDAC	Histone DeACetylase
HIF	Hypoxia-Inducible Factor
HK	HexoKinase
HO-1	Heme Oxygenase-1
HRE	Hypoxia-Responsive Element
Hsp	Heat-shock protein
ICAM	InterCellular Adhesion Molecule
IGF	Insulin-like Growth Factor
IL	InterLeukine
iNOS	inducible Nitric Oxide Synthase
KLF2	Kruppel-Like Factor 2
LDHA	Lactate DeHydrogenase A
LRP1	Low density lipoprotein recpetor-related protein 1
MCT	MonoCarboxylate Transporter
MMP	Matrix MetalloProteinase
mRNA	messenger RiboNucleic Acid
mTOR	mechanistic Target Of Rapamycin
mTORC	mTOR Complex
NAD	Nicotinamide Adenine Dinucleotide
NADH	Reduced form of NAD
NF-KB	Nuclear Factor-KB
NH	Neointimal Hyperplasia
NO	Nitric Oxide
NOX	<b>Nicotinamide adenine dinucleotide phosphate oxidase</b>
OPN	Osteopontin
PAD	Peripheral Arterial Disease
PDGF	Platelet Derived Growth Factor
PDH	Pyruvate DeHydrogenase
PFK	PhospoFructoKinase
PFKFB3	6-PhospoFructo-2-Kinase/Fructose-2,6-Bisphospatase 3
PHD	Prolyl Hydroxylase Domain
PI3K	Phosphatidylinositol-3-kinases
RACK	Receptor for Activated Protein Kinase C
ROS	Reactive Oxygen Species
SIRT	Sirtuin
SMC	Smooth Muscle Cells
TGF	Transforming-Growth Factor
TIE	Tyrosine kinase with Immunoglobulin-like and EGF-like domains
TIMP	Tissue Inhibitor of MetalloProteinase
TNF	Tumor Necrosis Factor
USRDS	United States Renal Data System
VCAM	Vascular Cell Adhesion Molecule
VEGF	Vascular Endothelial Growth Factor
VHL	von Hippel-Lindau
WSS	Wall Shear Stress

## References

1. Jha, V.; Garcia-Garcia, G.; Iseki, K.; Li, Z.; Naicker, S.; Plattner, B.; Saran, R.; Wang, A.Y.; Yang, C.W. Chronic kidney disease: Global dimension and perspectives. *Lancet* **2013**, *382*, 260–272. [CrossRef]
2. United States Renal Data System. *2018 USRDS Annual Data Report: Epidemiology of Kidney Disease in the United States*; National Institutes of Health, National Institute of Diabetes and Digestive and Kidney Diseases: Bethesda, MD, USA, 2018; Available online: <https://www.usrds.org/adr.aspx> (accessed on 3 October 2019).
3. Czyzewski, L.; Sanko-Resmer, J.; Wyzgal, J.; Kurowski, A. Assessment of health-related quality of life of patients after kidney transplantation in comparison with hemodialysis and peritoneal dialysis. *Ann. Transpl.* **2014**, *19*, 576–585.
4. Vascular Access Work, G. Clinical practice guidelines for vascular access. *Am. J. Kidney Dis.* **2006**, *48* (Suppl. 1), S176–S247.
5. Konner, K.; Nonnast-Daniel, B.; Ritz, E. The arteriovenous fistula. *J. Am. Soc. Nephrol.* **2003**, *14*, 1669–1680. [CrossRef]
6. Brescia, M.J.; Cimino, J.E.; Appel, K.; Hurwich, B.J. Chronic hemodialysis using venipuncture and a surgically created arteriovenous fistula. *N. Engl. J. Med.* **1966**, *275*, 1089–1092. [CrossRef]
7. Dember, L.M.; Beck, G.J.; Allon, M.; Delmez, J.A.; Dixon, B.S.; Greenberg, A.; Himmelfarb, J.; Vazquez, M.A.; Gassman, J.J.; Greene, T.; et al. Effect of clopidogrel on early failure of arteriovenous fistulas for hemodialysis: A randomized controlled trial. *JAMA* **2008**, *299*, 2164–2171. [CrossRef]
8. Al-Jaishi, A.A.; Oliver, M.J.; Thomas, S.M.; Lok, C.E.; Zhang, J.C.; Garg, A.X.; Kosa, S.D.; Quinn, R.R.; Moist, L.M. Patency rates of the arteriovenous fistula for hemodialysis: A systematic review and meta-analysis. *Am. J. Kidney Dis.* **2014**, *63*, 464–478. [CrossRef]
9. Hedin, U. Vascular access: A never-ending story. *J. Cardiovasc. Surg. (Torino)* **2014**, *55*, 793–801.
10. Roy-Chaudhury, P.; Spergel, L.M.; Besarab, A.; Asif, A.; Ravani, P. Biology of arteriovenous fistula failure. *J. Nephrol.* **2007**, *20*, 150–163.
11. Feldman, H.I.; Joffe, M.; Rosas, S.E.; Burns, J.E.; Knauss, J.; Brayman, K. Predictors of successful arteriovenous fistula maturation. *Am. J. Kidney Dis.* **2003**, *42*, 1000–1012. [CrossRef]
12. Rothuizen, T.C.; Wong, C.; Quax, P.H.A.; van Zonneveld, A.J.; Rabelink, T.J.; Rotmans, J.I. Arteriovenous access failure: More than just intimal hyperplasia? *Nephrol. Dial. Transplant.* **2013**, *28*, 1085–1092. [CrossRef] [PubMed]
13. Hu, H.; Patel, S.; Hanisch, J.J.; Santana, J.M.; Hashimoto, T.; Bai, H.; Kudze, T.; Foster, T.R.; Guo, J.; Yatsula, B.; et al. Future research directions to improve fistula maturation and reduce access failure. *Semin. Vasc. Surg.* **2016**, *29*, 153–171. [CrossRef] [PubMed]
14. Remuzzi, A.; Ene-Iordache, B. Novel paradigms for dialysis vascular access: Upstream hemodynamics and vascular remodeling in dialysis access stenosis. *Clin. J. Am. Soc. Nephrol.* **2013**, *8*, 2186–2193. [CrossRef] [PubMed]
15. Browne, L.D.; Bashar, K.; Griffin, P.; Kavanagh, E.G.; Walsh, S.R.; Walsh, M.T. The Role of shear stress in arteriovenous fistula maturation and failure: A systematic review. *PLoS ONE* **2015**, *10*, e0145795. [CrossRef]
16. Jia, L.; Wang, L.; Wei, F.; Yu, H.; Dong, H.; Wang, B.; Lu, Z.; Sun, G.; Chen, H.; Meng, J.; et al. Effects of wall shear stress in venous neointimal hyperplasia of arteriovenous fistulae. *Nephrology (Carlton)* **2015**, *20*, 335–342. [CrossRef]
17. Chiu, J.-J.; Chien, S. Effects of disturbed flow on vascular endothelium: Pathophysiological basis and clinical perspectives. *Physiol. Rev.* **2011**, *91*, 327–387. [CrossRef]
18. Dai, G.; Kaazempur-Mofrad, M.R.; Natarajan, S.; Zhang, Y.; Vaughn, S.; Blackman, B.R.; Kamm, R.D.; Garcia-Cardena, G.; Gimbrone, M.A., Jr. Distinct endothelial phenotypes evoked by arterial waveforms derived from atherosclerosis-susceptible and -resistant regions of human vasculature. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 14871–14876. [CrossRef]
19. Fan, L.; Karino, T. Effect of a disturbed flow on proliferation of the cells of a hybrid vascular graft. *Biorheology* **2010**, *47*, 31–38.
20. Fan, L.; Sakai, J.; Bessho, S.; Wada, S.; Karino, T. Effect of a disturbed flow on adhesion of monocytes to a model of an arterial wall. *Biorheology* **2010**, *47*, 15–29.

21. Ene-Iordache, B.; Cattaneo, L.; Dubini, G.; Remuzzi, A. Effect of anastomosis angle on the localization of disturbed flow in side-to-end fistulae for haemodialysis access. *Nephrol. Dial. Transplant.* **2013**, *28*, 997–1005. [[CrossRef](#)]
22. Muto, A.; Model, L.; Ziegler, K.; Eghbalieh, S.D.; Dardik, A. Mechanisms of vein graft adaptation to the arterial circulation: Insights into the neointimal algorithm and management strategies. *Circ. J.* **2010**, *74*, 1501–1512. [[CrossRef](#)] [[PubMed](#)]
23. Roy-Chaudhury, P.; Wang, Y.; Krishnamoorthy, M.; Zhang, J.; Banerjee, R.; Munda, R.; Heffelfinger, S.; Arend, L. Cellular phenotypes in human stenotic lesions from haemodialysis vascular access. *Nephrol. Dial. Transplant.* **2009**, *24*, 2786–2791. [[CrossRef](#)] [[PubMed](#)]
24. Tseng, C.N.; Karlof, E.; Chang, Y.T.; Lengquist, M.; Rotzius, P.; Berggren, P.O.; Hedin, U.; Eriksson, E.E. Contribution of endothelial injury and inflammation in early phase to vein graft failure: The causal factors impact on the development of intimal hyperplasia in murine models. *PLoS ONE* **2014**, *9*, e98904. [[CrossRef](#)] [[PubMed](#)]
25. Kang, H.Y.; Fan, Y.B.; Deng, X.Y. Vascular smooth muscle cell glycocalyx modulates shear-induced proliferation, migration, and NO production responses. *Am. J. Physiol. Heart C* **2011**, *300*, H76–H83. [[CrossRef](#)]
26. Pike, D.; Shiu, Y.T.; Cho, Y.F.; Le, H.; Somarathna, M.; Isayeva, T.; Guo, L.; Symons, J.D.; Kevil, C.G.; Totenhagen, J.; et al. The effect of endothelial nitric oxide synthase on the hemodynamics and wall mechanics in murine arteriovenous fistulas. *Sci. Rep.* **2019**, *9*, 4299. [[CrossRef](#)]
27. Croatt, A.J.; Grande, J.P.; Hernandez, M.C.; Ackerman, A.W.; Katusic, Z.S.; Nath, K.A. Characterization of a model of an arteriovenous fistula in the rat: The effect of L-NAME. *Am. J. Pathol.* **2010**, *176*, 2530–2541. [[CrossRef](#)]
28. Jones, G.T.; van Rij, A.M.; Packer, S.G.; Walker, R.J.; Stehbens, W.E. Venous endothelial changes in therapeutic arteriovenous fistulae. *Atherosclerosis* **1998**, *137*, 149–156. [[CrossRef](#)]
29. Nath, K.A.; Kanakiriya, S.K.; Grande, J.P.; Croatt, A.J.; Katusic, Z.S. Increased venous proinflammatory gene expression and intimal hyperplasia in an aorto-caval fistula model in the rat. *Am. J. Pathol.* **2003**, *162*, 2079–2090. [[CrossRef](#)]
30. Schepers, A.; Eefting, D.; Bonta, P.I.; Grimbergen, J.M.; de Vries, M.R.; van Weel, V.; de Vries, C.J.; Egashira, K.; van Bockel, J.H.; Quax, P.H. Anti-MCP-1 gene therapy inhibits vascular smooth muscle cells proliferation and attenuates vein graft thickening both in vitro and in vivo. *Arterioscler. Thromb. Vasc. Biol.* **2006**, *26*, 2063–2069. [[CrossRef](#)]
31. Chang, C.J.; Ko, Y.S.; Ko, P.J.; Hsu, L.A.; Chen, C.F.; Yang, C.W.; Hsu, T.S.; Pang, J.H. Thrombosed arteriovenous fistula for hemodialysis access is characterized by a marked inflammatory activity. *Kidney Int.* **2005**, *68*, 1312–1319. [[CrossRef](#)]
32. Tsui, L.V.; Camrud, A.; Mondesire, J.; Carlson, P.; Zayek, N.; Camrud, L.; Donahue, B.; Bauer, S.; Lin, A.; Frey, D.; et al. p27-p16 fusion gene inhibits angioplasty-induced neointimal hyperplasia and coronary artery occlusion. *Circ. Res.* **2001**, *89*, 323–328. [[CrossRef](#)] [[PubMed](#)]
33. Ohashi, N.; Matsumori, A.; Furukawa, Y.; Ono, K.; Okada, M.; Iwasaki, A.; Miyamoto, T.; Nakano, A.; Sasayama, S. Role of p38 mitogen-activated protein kinase in neointimal hyperplasia after vascular injury. *Arterioscler. Thromb. Vasc. Biol.* **2000**, *20*, 2521–2526. [[CrossRef](#)] [[PubMed](#)]
34. Yang, B.; Janardhanan, R.; Vohra, P.; Greene, E.L.; Bhattacharya, S.; Withers, S.; Roy, B.; Nieves Torres, E.C.; Mandrekar, J.; Leof, E.B.; et al. Adventitial transduction of lentivirus-shRNA-VEGF-A in arteriovenous fistula reduces venous stenosis formation. *Kidney Int.* **2014**, *85*, 289–306. [[CrossRef](#)] [[PubMed](#)]
35. Lindner, V.; Reidy, M.A. Proliferation of smooth muscle cells after vascular injury is inhibited by an antibody against basic fibroblast growth factor. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 3739–3743. [[CrossRef](#)]
36. Weiss, M.F.; Scivittaro, V.; Anderson, J.M. Oxidative stress and increased expression of growth factors in lesions of failed hemodialysis access. *Am. J. Kidney Dis.* **2001**, *37*, 970–980. [[CrossRef](#)]
37. Stracke, S.; Konner, K.; Kostlin, I.; Friedl, R.; Jehle, P.M.; Hombach, V.; Keller, F.; Waltenberger, J. Increased expression of TGF-beta1 and IGF-I in inflammatory stenotic lesions of hemodialysis fistulas. *Kidney Int.* **2002**, *61*, 1011–1019. [[CrossRef](#)]
38. Nath, K.A.; Grande, J.P.; Kang, L.; Juncos, J.P.; Ackerman, A.W.; Croatt, A.J.; Katusic, Z.S. ss-Catenin is markedly induced in a murine model of an arteriovenous fistula: The effect of metalloproteinase inhibition. *Am. J. Physiol. Renal Physiol.* **2010**, *299*, F1270–F1277. [[CrossRef](#)]

39. Geenen, I.L.; Kolk, F.F.; Molin, D.G.; Wagenaar, A.; Compeer, M.G.; Tordoir, J.H.; Schurink, G.W.; De Mey, J.G.; Post, M.J. Nitric oxide resistance reduces arteriovenous fistula maturation in chronic kidney disease in rats. *PLoS ONE* **2016**, *11*, e0146212. [[CrossRef](#)]
40. Zhao, J.; Jour'd'heuil, F.L.; Xue, M.; Conti, D.; Lopez-Soler, R.I.; Ginnan, R.; Asif, A.; Singer, H.A.; Jour'd'heuil, D.; Long, X. Dual function for mature vascular smooth muscle cells during arteriovenous fistula remodeling. *J. Am. Heart Assoc.* **2017**, *6*, e004891. [[CrossRef](#)]
41. Liang, M.; Wang, Y.; Liang, A.; Mitch, W.E.; Roy-Chaudhury, P.; Han, G.; Cheng, J. Migration of smooth muscle cells from the arterial anastomosis of arteriovenous fistulas requires Notch activation to form neointima. *Kidney Int.* **2015**, *88*, 490–502. [[CrossRef](#)]
42. Duque, J.C.; Vazquez-Padron, R.I. Myofibroblasts: The ideal target to prevent arteriovenous fistula failure? *Kidney Int.* **2014**, *85*, 234–236. [[CrossRef](#)] [[PubMed](#)]
43. Coen, M.; Gabbiani, G.; Bochaton-Piallat, M.L. Myofibroblast-mediated adventitial remodeling: An underestimated player in arterial pathology. *Arterioscler. Thromb. Vasc. Biol.* **2011**, *31*, 2391–2396. [[CrossRef](#)] [[PubMed](#)]
44. Wang, Y.; Krishnamoorthy, M.; Banerjee, R.; Zhang, J.; Rudich, S.; Holland, C.; Arend, L. Roy-Chaudhury, P. Venous stenosis in a pig arteriovenous fistula model—anatomy, mechanisms and cellular phenotypes. *Nephrol. Dial. Transplant.* **2008**, *23*, 525–533. [[CrossRef](#)] [[PubMed](#)]
45. Simone, S.; Loverre, A.; Cariello, M.; Divella, C.; Castellano, G.; Gesualdo, L.; Pertosa, G.; Grandaliano, G. Arteriovenous fistula stenosis in hemodialysis patients is characterized by an increased adventitial fibrosis. *J. Nephrol.* **2014**, *27*, 555–562. [[CrossRef](#)] [[PubMed](#)]
46. Liang, A.; Wang, Y.; Han, G.; Truong, L.; Cheng, J. Chronic kidney disease accelerates endothelial barrier dysfunction in a mouse model of an arteriovenous fistula. *Am. J. Physiol. Renal Physiol.* **2013**, *304*, F1413–F1420. [[CrossRef](#)]
47. Wasse, H.; Huang, R.; Naqvi, N.; Smith, E.; Wang, D.; Husain, A. Inflammation, oxidation and venous neointimal hyperplasia precede vascular injury from AVF creation in CKD patients. *J. Vasc. Access.* **2012**, *13*, 168–174. [[CrossRef](#)]
48. Juncos, J.P.; Grande, J.P.; Kang, L.; Ackerman, A.W.; Croatt, A.J.; Katusic, Z.S.; Nath, K.A. MCP-1 Contributes to arteriovenous fistula failure. *J. Am. Soc. Nephrol.* **2011**, *22*, 43–48. [[CrossRef](#)]
49. Ikegaya, N.; Yamamoto, T.; Takeshita, A.; Watanabe, T.; Yonemura, K.; Miyaji, T.; Ohishi, K.; Furuhashi, M.; Maruyama, Y.; Hishida, A. Elevated erythropoietin receptor and transforming growth factor-beta1 expression in stenotic arteriovenous fistulae used for hemodialysis. *J. Am. Soc. Nephrol.* **2000**, *11*, 928–935.
50. Hall, M.R.; Yamamoto, K.; Protack, C.D.; Tsuneki, M.; Kuwahara, G.; Assi, R.; Brownson, K.E.; Bai, H.; Madri, J.A.; Dardik, A. Temporal regulation of venous extracellular matrix components during arteriovenous fistula maturation. *J. Vasc. Access.* **2015**, *16*, 93–106. [[CrossRef](#)]
51. Lee, E.S.; Shen, Q.; Pitts, R.L.; Guo, M.; Wu, M.H.; Sun, S.C.; Yuan, S.Y. Serum metalloproteinases MMP-2, MMP-9, and metalloproteinase tissue inhibitors in patients are associated with arteriovenous fistula maturation. *J. Vasc. Surg.* **2011**, *54*, 454–459. [[CrossRef](#)]
52. Lin, C.C.; Yang, W.C.; Chung, M.Y.; Lee, P.C. Functional polymorphisms in matrix metalloproteinases-1, -3, -9 are associated with arteriovenous fistula patency in hemodialysis patients. *Clin. J. Am. Soc. Nephrol.* **2010**, *5*, 1805–1814. [[CrossRef](#)] [[PubMed](#)]
53. Kokubo, T.; Ishikawa, N.; Uchida, H.; Chasnoff, S.E.; Xie, X.; Mathew, S.; Hruska, K.A.; Choi, E.T. CKD Accelerates development of neointimal hyperplasia in arteriovenous fistulas. *J. Am. Soc. Nephrol.* **2009**, *20*, 1236–1245. [[CrossRef](#)] [[PubMed](#)]
54. Scatena, M.; Liaw, L.; Giachelli, C.M. Osteopontin: A multifunctional molecule regulating chronic inflammation and vascular disease. *Arterioscler. Thromb. Vasc. Biol.* **2007**, *27*, 2302–2309. [[CrossRef](#)] [[PubMed](#)]
55. Joddar, B.; Firstenberg, M.S.; Reen, R.K.; Varadharaj, S.; Khan, M.; Childers, R.C.; Zweier, J.L.; Gooch, K.J. Arterial levels of oxygen stimulate intimal hyperplasia in human saphenous veins via a ROS-dependent mechanism. *PLoS ONE* **2015**, *10*, e0120301. [[CrossRef](#)] [[PubMed](#)]
56. Sun, N.; Wood, N.B.; Hughes, A.D.; Thom, S.A.; Xu, X.Y. Fluid-wall modelling of mass transfer in an axisymmetric stenosis: Effects of shear-dependent transport properties. *Ann. Biomed. Eng.* **2006**, *34*, 1119–1128. [[CrossRef](#)] [[PubMed](#)]

57. Iori, F.; Grechy, L.; Corbett, R.W.; Gedroyc, W.; Duncan, N.; Caro, C.G.; Vincent, P.E. The effect of in-plane arterial curvature on blood flow and oxygen transport in arterio-venous fistulae. *Phys. Fluids* (1994) **2015**, 27, 031903. [[CrossRef](#)] [[PubMed](#)]
58. Semenza, G.L. Hypoxia-inducible factor 1 (HIF-1) pathway. *Sci. STKE* **2007**, 2007, cm8. [[CrossRef](#)]
59. Koh, M.Y.; Powis, G. Passing the baton: The HIF switch. *Trends Biochem. Sci.* **2012**, 37, 364–372. [[CrossRef](#)]
60. Wenger, R.H. Cellular adaptation to hypoxia: O<sub>2</sub>-sensing protein hydroxylases, hypoxia-inducible transcription factors, and O<sub>2</sub>-regulated gene expression. *FASEB J.* **2002**, 16, 1151–1162. [[CrossRef](#)]
61. Eltzschig, H.K.; Bratton, D.L.; Colgan, S.P. Targeting hypoxia signalling for the treatment of ischaemic and inflammatory diseases. *Nat. Rev. Drug Discov.* **2014**, 13, 852–869. [[CrossRef](#)]
62. Epstein, A.C.; Gleadle, J.M.; McNeill, L.A.; Hewitson, K.S.; O'Rourke, J.; Mole, D.R.; Mukherji, M.; Metzen, E.; Wilson, M.I.; Dhanda, A.; et al. Elegans EGL-9 and mammalian homologs define a family of dioxygenases that regulate HIF by prolyl hydroxylation. *Cell* **2001**, 107, 43–54. [[CrossRef](#)]
63. Jaakkola, P.; Mole, D.R.; Tian, Y.M.; Wilson, M.I.; Gielbert, J.; Gaskell, S.J.; von Kriegsheim, A.; Hebestreit, H.F.; Mukherji, M.; Schofield, C.J.; et al. Targeting of HIF- $\alpha$  to the von Hippel-Lindau ubiquitylation complex by O<sub>2</sub>-regulated prolyl hydroxylation. *Science* **2001**, 292, 468–472. [[CrossRef](#)] [[PubMed](#)]
64. Lisztwan, J.; Imbert, G.; Wirbelauer, C.; Gstaiger, M.; Krek, W. The von Hippel-Lindau tumor suppressor protein is a component of an E3 ubiquitin-protein ligase activity. *Genes Dev.* **1999**, 13, 1822–1833. [[CrossRef](#)] [[PubMed](#)]
65. Ivan, M.; Kondo, K.; Yang, H.; Kim, W.; Valiando, J.; Ohh, M.; Salic, A.; Asara, J.M.; Lane, W.S.; Kaelin, W.G., Jr. HIF $\alpha$  targeted for VHL-mediated destruction by proline hydroxylation: Implications for O<sub>2</sub> sensing. *Science* **2001**, 292, 464–468. [[CrossRef](#)] [[PubMed](#)]
66. Bruick, R.K.; McKnight, S.L. A conserved family of prolyl-4-hydroxylases that modify HIF. *Science* **2001**, 294, 1337–1340. [[CrossRef](#)] [[PubMed](#)]
67. Mazure, N.M.; Brahimi-Horn, M.C.; Berta, M.A.; Benizri, E.; Bilton, R.L.; Dayan, F.; Ginouves, A.; Berra, E.; Pouyssegur, J. HIF-1: Master and commander of the hypoxic world. A pharmacological approach to its regulation by siRNAs. *Biochem. Pharmacol.* **2004**, 68, 971–980. [[CrossRef](#)] [[PubMed](#)]
68. Brahimi-Horn, M.C.; Pouyssegur, J. HIF at a glance. *J. Cell Sci.* **2009**, 122, 1055–1057. [[CrossRef](#)]
69. Lando, D.; Peet, D.J.; Gorman, J.J.; Whelan, D.A.; Whitelaw, M.L.; Bruick, R.K. FIH-1 is an asparaginyl hydroxylase enzyme that regulates the transcriptional activity of hypoxia-inducible factor. *Genes Dev.* **2002**, 16, 1466–1471. [[CrossRef](#)]
70. Mahon, P.C.; Hirota, K.; Semenza, G.L. FIH-1: A novel protein that interacts with HIF-1 $\alpha$  and VHL to mediate repression of HIF-1 transcriptional activity. *Genes Dev.* **2001**, 15, 2675–2686. [[CrossRef](#)]
71. Lando, D.; Peet, D.J.; Whelan, D.A.; Gorman, J.J.; Whitelaw, M.L. Asparagine hydroxylation of the HIF transactivation domain a hypoxic switch. *Science* **2002**, 295, 858–861. [[CrossRef](#)]
72. Dayan, F.; Roux, D.; Brahimi-Horn, M.C.; Pouyssegur, J.; Mazure, N.M. The oxygen sensor factor-inhibiting hypoxia-inducible factor-1 controls expression of distinct genes through the bifunctional transcriptional character of hypoxia-inducible factor-1 $\alpha$ . *Cancer Res.* **2006**, 66, 3688–3698. [[CrossRef](#)] [[PubMed](#)]
73. Kung, A.L.; Wang, S.; Klco, J.M.; Kaelin, W.G.; Livingston, D.M. Suppression of tumor growth through disruption of hypoxia-inducible transcription. *Nat. Med.* **2000**, 6, 1335–1340. [[CrossRef](#)]
74. Tazuke, S.I.; Mazure, N.M.; Sugawara, J.; Carland, G.; Faessen, G.H.; Suen, L.F.; Irwin, J.C.; Powell, D.R.; Giaccia, A.J.; Giudice, L.C. Hypoxia stimulates insulin-like growth factor binding protein 1 (IGFBP-1) gene expression in HepG2 cells: A possible model for IGFBP-1 expression in fetal hypoxia. *Proc. Natl. Acad. Sci. USA* **1998**, 95, 10188–10193. [[CrossRef](#)] [[PubMed](#)]
75. Pugh, C.W.; Tan, C.C.; Jones, R.W.; Ratcliffe, P.J. Functional analysis of an oxygen-regulated transcriptional enhancer lying 3' to the mouse erythropoietin gene. *Proc. Natl. Acad. Sci. USA* **1991**, 88, 10553–10557. [[CrossRef](#)] [[PubMed](#)]
76. Wang, G.L.; Semenza, G.L. Purification and characterization of hypoxia-inducible factor 1. *J. Biol. Chem.* **1995**, 270, 1230–1237. [[CrossRef](#)]
77. Frede, S.; Freitag, P.; Otto, T.; Heilmaier, C.; Fandrey, J. The proinflammatory cytokine interleukin 1 $\beta$  and hypoxia cooperatively induce the expression of adrenomedullin in ovarian carcinoma cells through hypoxia inducible factor 1 activation. *Cancer Res.* **2005**, 65, 4690–4697. [[CrossRef](#)]

78. Zatyka, M.; da Silva, N.F.; Clifford, S.C.; Morris, M.R.; Wiesener, M.S.; Eckardt, K.U.; Houlston, R.S.; Richards, F.M.; Latif, F.; Maher, E.R. Identification of cyclin D1 and other novel targets for the von Hippel-Lindau tumor suppressor gene by expression array analysis and investigation of cyclin D1 genotype as a modifier in von Hippel-Lindau disease. *Cancer Res.* **2002**, *62*, 3803–3811.
79. Mazure, N.M.; Pouyssegur, J. Atypical BH3-domains of BNIP3 and BNIP3L lead to autophagy in hypoxia. *Autophagy* **2009**, *5*, 868–869. [[CrossRef](#)]
80. Sowter, H.M.; Ratcliffe, P.J.; Watson, P.; Greenberg, A.H.; Harris, A.L. HIF-1-dependent regulation of hypoxic induction of the cell death factors BNIP3 and NIX in human tumors. *Cancer Res.* **2001**, *61*, 6669–6673.
81. Wykoff, C.C.; Pugh, C.W.; Maxwell, P.H.; Harris, A.L.; Ratcliffe, P.J. Identification of novel hypoxia dependent and independent target genes of the von Hippel-Lindau (VHL) tumour suppressor by mRNA differential expression profiling. *Oncogene* **2000**, *19*, 6297–6305. [[CrossRef](#)]
82. Feldser, D.; Agani, F.; Iyer, N.V.; Pak, B.; Ferreira, G.; Semenza, G.L. Reciprocal positive regulation of hypoxia-inducible factor 1alpha and insulin-like growth factor 2. *Cancer Res.* **1999**, *59*, 3915–3918. [[PubMed](#)]
83. Krishnamachary, B.; Berg-Dixon, S.; Kelly, B.; Agani, F.; Feldser, D.; Ferreira, G.; Iyer, N.; LaRusch, J.; Pak, B.; Taghavi, P.; et al. Regulation of colon carcinoma cell invasion by hypoxia-inducible factor 1. *Cancer Res.* **2003**, *63*, 1138–1143. [[PubMed](#)]
84. Jung, F.; Palmer, L.A.; Zhou, N.; Johns, R.A. Hypoxic regulation of inducible nitric oxide synthase via hypoxia inducible factor-1 in cardiac myocytes. *Circ. Res.* **2000**, *86*, 319–325. [[CrossRef](#)] [[PubMed](#)]
85. Wang, G.L.; Semenza, G.L. General involvement of hypoxia-inducible factor 1 in transcriptional response to hypoxia. *Proc. Natl. Acad. Sci. USA* **1993**, *90*, 4304–4308. [[CrossRef](#)]
86. Wykoff, C.C.; Beasley, N.J.; Watson, P.H.; Turner, K.J.; Pastorek, J.; Sibtain, A.; Wilson, G.D.; Turley, H.; Talks, K.L.; Maxwell, P.H.; et al. Hypoxia-inducible expression of tumor-associated carbonic anhydrases. *Cancer Res.* **2000**, *60*, 7075–7083.
87. Greijer, A.E.; van der Groep, P.; Kemming, D.; Shvarts, A.; Semenza, G.L.; Meijer, G.A.; van de Wiel, M.A.; Belien, J.A.; Van Diest, P.J.; van der Wall, E. Up-regulation of gene expression by hypoxia is mediated predominantly by hypoxia-inducible factor 1 (HIF-1). *J. Pathol.* **2005**, *206*, 291–304. [[CrossRef](#)]
88. Rolfs, A.; Kvietikova, I.; Gassmann, M.; Wenger, R.H. Oxygen-regulated transferrin expression is mediated by hypoxia-inducible factor-1. *J. Biol. Chem.* **1997**, *272*, 20055–20062. [[CrossRef](#)]
89. Mukhopadhyay, C.K.; Mazumder, B.; Fox, P.L. Role of hypoxia-inducible factor-1 in transcriptional activation of ceruloplasmin by iron deficiency. *J. Biol. Chem.* **2000**, *275*, 21048–21054. [[CrossRef](#)]
90. Menendez, M.T.; Teygong, C.; Wade, K.; Florimond, C.; Blader, I.J. siRNA Screening Identifies the Host Hexokinase 2 (HK2) Gene as an Important Hypoxia-Inducible Transcription Factor 1 (HIF-1) Target Gene in Toxoplasma gondii-Infected Cells. *mBio* **2015**, *6*, e00462. [[CrossRef](#)]
91. Ebert, B.L.; Firth, J.D.; Ratcliffe, P.J. Hypoxia and mitochondrial inhibitors regulate expression of glucose transporter-1 via distinct Cis-acting sequences. *J. Biol. Chem.* **1995**, *270*, 29083–29089. [[CrossRef](#)]
92. Graven, K.K.; Bellur, D.; Klahn, B.D.; Lowrey, S.L.; Amberger, E. HIF-2alpha regulates glyceraldehyde-3-phosphate dehydrogenase expression in endothelial cells. *Biochim. Biophys. Acta* **2003**, *1626*, 10–18. [[CrossRef](#)]
93. Firth, J.D.; Ebert, B.L.; Pugh, C.W.; Ratcliffe, P.J. Oxygen-regulated control elements in the phosphoglycerate kinase 1 and lactate dehydrogenase A genes: Similarities with the erythropoietin 3' enhancer. *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 6496–6500. [[CrossRef](#)] [[PubMed](#)]
94. Grosfeld, A.; Andre, J.; Hauguel-De Mouzon, S.; Berra, E.; Pouyssegur, J.; Guerre-Millo, M. Hypoxia-inducible factor 1 transactivates the human leptin gene promoter. *J. Biol. Chem.* **2002**, *277*, 42953–42957. [[CrossRef](#)] [[PubMed](#)]
95. Tsuzuki, Y.; Fukumura, D.; Oosthuysen, B.; Koike, C.; Carmeliet, P.; Jain, R.K. Vascular endothelial growth factor (VEGF) modulation by targeting hypoxia-inducible factor-1alpha-> hypoxia response element-> VEGF cascade differentially regulates vascular response and growth rate in tumors. *Cancer Res.* **2000**, *60*, 6248–6252.
96. Das, B.; Yeger, H.; Tsuchida, R.; Torkin, R.; Gee, M.F.; Thorner, P.S.; Shibuya, M.; Malkin, D.; Baruchel, S. A hypoxia-driven vascular endothelial growth factor/Flt1 autocrine loop interacts with hypoxia-inducible factor-1alpha through mitogen-activated protein kinase/extracellular signal-regulated kinase 1/2 pathway in neuroblastoma. *Cancer Res.* **2005**, *65*, 7267–7275. [[CrossRef](#)]

97. Yamakawa, M.; Liu, L.X.; Date, T.; Belanger, A.J.; Vincent, K.A.; Akita, G.Y.; Kuriyama, T.; Cheng, S.H.; Gregory, R.J.; Jiang, C. Hypoxia-inducible factor-1 mediates activation of cultured vascular endothelial cells by inducing multiple angiogenic factors. *Circ. Res.* **2003**, *93*, 664–673. [[CrossRef](#)]
98. Kim, Y.M.; Pae, H.O.; Park, J.E.; Lee, Y.C.; Woo, J.M.; Kim, N.H.; Choi, Y.K.; Lee, B.S.; Kim, S.R.; Chung, H.T. Heme oxygenase in the regulation of vascular biology: From molecular mechanisms to therapeutic opportunities. *Antioxid. Redox Signal.* **2011**, *14*, 137–167. [[CrossRef](#)]
99. Shimoda, L.A.; Semenza, G.L. HIF and the lung: Role of hypoxia-inducible factors in pulmonary development and disease. *Am. J. Respir. Crit. Care Med.* **2011**, *183*, 152–156. [[CrossRef](#)]
100. Semenza, G.L.; Agani, F.; Iyer, N.; Kotch, L.; Laughner, E.; Leung, S.; Yu, A. Regulation of cardiovascular development and physiology by hypoxia-inducible factor 1. *Ann. NY Acad. Sci.* **1999**, *874*, 262–268. [[CrossRef](#)]
101. Semenza, G.L. Targeting HIF-1 for cancer therapy. *Nat. Rev. Cancer* **2003**, *3*, 721–732. [[CrossRef](#)]
102. Schito, L.; Semenza, G.L. Hypoxia-Inducible Factors: Master Regulators of Cancer Progression. *Trends Cancer* **2016**, *2*, 758–770. [[CrossRef](#)] [[PubMed](#)]
103. Semenza, G.L. Hypoxia-inducible factor 1 and cardiovascular disease. *Annu. Rev. Physiol.* **2014**, *76*, 39–56. [[CrossRef](#)] [[PubMed](#)]
104. Baek, J.H.; Liu, Y.V.; McDonald, K.R.; Wesley, J.B.; Hubbi, M.E.; Byun, H.; Semenza, G.L. Spermidine/spermine-N1-acetyltransferase 2 is an essential component of the ubiquitin ligase complex that regulates hypoxia-inducible factor 1alpha. *J. Biol. Chem.* **2007**, *282*, 23572–23580. [[CrossRef](#)] [[PubMed](#)]
105. Luo, W.; Zhong, J.; Chang, R.; Hu, H.; Pandey, A.; Semenza, G.L. Hsp70 and CHIP selectively mediate ubiquitination and degradation of hypoxia-inducible factor (HIF)-1alpha but Not HIF-2alpha. *J. Biol. Chem.* **2010**, *285*, 3651–3663. [[CrossRef](#)]
106. Koh, M.Y.; Darnay, B.G.; Powis, G. Hypoxia-associated factor, a novel E3-ubiquitin ligase, binds and ubiquitinates hypoxia-inducible factor 1alpha, leading to its oxygen-independent degradation. *Mol. Cell. Biol.* **2008**, *28*, 7081–7095. [[CrossRef](#)]
107. Hubbi, M.E.; Hu, H.; Kshitiz; Gilkes, D.M.; Semenza, G.L. Sirtuin-7 inhibits the activity of hypoxia-inducible factors. *J. Biol. Chem.* **2013**, *288*, 20768–20775. [[CrossRef](#)]
108. Zelzer, E.; Levy, Y.; Kahana, C.; Shilo, B.Z.; Rubinstein, M.; Cohen, B. Insulin induces transcription of target genes through the hypoxia-inducible factor HIF-1alpha/ARNT. *EMBO J.* **1998**, *17*, 5085–5094. [[CrossRef](#)]
109. Treins, C.; Giorgetti-Peraldi, S.; Murdaca, J.; Semenza, G.L.; Van Obberghen, E. Insulin stimulates hypoxia-inducible factor 1 through a phosphatidylinositol 3-kinase/target of rapamycin-dependent signaling pathway. *J. Biol. Chem.* **2002**, *277*, 27975–27981. [[CrossRef](#)]
110. Fukuda, R.; Hirota, K.; Fan, F.; Jung, Y.D.; Ellis, L.M.; Semenza, G.L. Insulin-like growth factor 1 induces hypoxia-inducible factor 1-mediated vascular endothelial growth factor expression, which is dependent on MAP kinase and phosphatidylinositol 3-kinase signaling in colon cancer cells. *J. Biol. Chem.* **2002**, *277*, 38205–38211. [[CrossRef](#)]
111. Richard, D.E.; Berra, E.; Pouyssegur, J. Nonhypoxic pathway mediates the induction of hypoxia-inducible factor 1alpha in vascular smooth muscle cells. *J. Biol. Chem.* **2000**, *275*, 26765–26771.
112. Kohl, R.; Zhou, J.; Brune, B. Reactive oxygen species attenuate nitric-oxide-mediated hypoxia-inducible factor-1alpha stabilization. *Free Radic. Biol. Med.* **2006**, *40*, 1430–1442. [[CrossRef](#)] [[PubMed](#)]
113. Sandau, K.B.; Fandrey, J.; Brune, B. Accumulation of HIF-1alpha under the influence of nitric oxide. *Blood* **2001**, *97*, 1009–1015. [[CrossRef](#)] [[PubMed](#)]
114. Sanchez-Lopez, E.; Lopez, A.F.; Esteban, V.; Yague, S.; Egido, J.; Ruiz-Ortega, M.; Alvarez-Arroyo, M.V. Angiotensin II regulates vascular endothelial growth factor via hypoxia-inducible factor-1alpha induction and redox mechanisms in the kidney. *Antioxid. Redox Signal.* **2005**, *7*, 1275–1284. [[CrossRef](#)] [[PubMed](#)]
115. Hellwig-Burgel, T.; Rutkowski, K.; Metzen, E.; Fandrey, J.; Jelkmann, W. Interleukin-1beta and tumor necrosis factor-alpha stimulate DNA binding of hypoxia-inducible factor-1. *Blood* **1999**, *94*, 1561–1567. [[CrossRef](#)]
116. Pouyssegur, J.; Mechta-Grigoriou, F. Redox regulation of the hypoxia-inducible factor. *Biol. Chem.* **2006**, *387*, 1337–1346. [[CrossRef](#)]
117. Chandel, N.S.; McClintock, D.S.; Feliciano, C.E.; Wood, T.M.; Melendez, J.A.; Rodriguez, A.M.; Schumacker, P.T. Reactive oxygen species generated at mitochondrial complex III stabilize hypoxia-inducible factor-1alpha during hypoxia: A mechanism of O2 sensing. *J. Biol. Chem.* **2000**, *275*, 25130–25138. [[CrossRef](#)]

118. Brauchle, M.; Funk, J.O.; Kind, P.; Werner, S. Ultraviolet B and H<sub>2</sub>O<sub>2</sub> are potent inducers of vascular endothelial growth factor expression in cultured keratinocytes. *J. Biol. Chem.* **1996**, *271*, 21793–21797. [[CrossRef](#)]
119. Rezvani, H.R.; Dedieu, S.; North, S.; Belloc, F.; Rossignol, R.; Letellier, T.; de Verneuil, H.; Taieb, A.; Mazurier, F. Hypoxia-inducible factor-1alpha, a key factor in the keratinocyte response to UVB exposure. *J. Biol. Chem.* **2007**, *282*, 16413–16422. [[CrossRef](#)]
120. Gerald, D.; Berra, E.; Frapart, Y.M.; Chan, D.A.; Giaccia, A.J.; Mansuy, D.; Pouyssegur, J.; Yaniv, M.; Mechta-Grigoriou, F. JunD reduces tumor angiogenesis by protecting cells from oxidative stress. *Cell* **2004**, *118*, 781–794. [[CrossRef](#)]
121. Lu, H.; Dalgard, C.L.; Mohyeldin, A.; McFate, T.; Tait, A.S.; Verma, A. Reversible inactivation of HIF-1 prolyl hydroxylases allows cell metabolism to control basal HIF-1. *J. Biol. Chem.* **2005**, *280*, 41928–41939. [[CrossRef](#)]
122. Knowles, H.J.; Mole, D.R.; Ratcliffe, P.J.; Harris, A.L. Normoxic stabilization of hypoxia-inducible factor-1alpha by modulation of the labile iron pool in differentiating U937 macrophages: Effect of natural resistance-associated macrophage protein 1. *Cancer Res.* **2006**, *66*, 2600–2607. [[CrossRef](#)] [[PubMed](#)]
123. Wang, G.L.; Jiang, B.H.; Semenza, G.L. Effect of protein kinase and phosphatase inhibitors on expression of hypoxia-inducible factor 1. *Biochem. Biophys. Res. Commun.* **1995**, *216*, 669–675. [[CrossRef](#)] [[PubMed](#)]
124. Tacchini, L.; Dansi, P.; Matteucci, E.; Desiderio, M.A. Hepatocyte growth factor signalling stimulates hypoxia inducible factor-1 (HIF-1) activity in HepG2 hepatoma cells. *Carcinogenesis* **2001**, *22*, 1363–1371. [[CrossRef](#)] [[PubMed](#)]
125. Park, J.H.; Kim, T.Y.; Jong, H.S.; Kim, T.Y.; Chun, Y.S.; Park, J.W.; Lee, C.T.; Jung, H.C.; Kim, N.K.; Bang, Y.J. Gastric epithelial reactive oxygen species prevent normoxic degradation of hypoxia-inducible factor-1alpha in gastric cancer cells. *Clin. Cancer Res.* **2003**, *9*, 433–440.
126. Shatrov, V.A.; Sumbayev, V.V.; Zhou, J.; Brune, B. Oxidized low-density lipoprotein (oxLDL) triggers hypoxia-inducible factor-1alpha (HIF-1alpha) accumulation via redox-dependent mechanisms. *Blood* **2003**, *101*, 4847–4849. [[CrossRef](#)] [[PubMed](#)]
127. Zhu, X.Y.; Rodriguez-Porcel, M.; Bentley, M.D.; Chade, A.R.; Sica, V.; Napoli, C.; Caplice, N.; Ritman, E.L.; Lerman, A.; Lerman, L.O. Antioxidant intervention attenuates myocardial neovascularization in hypercholesterolemia. *Circulation* **2004**, *109*, 2109–2115. [[CrossRef](#)] [[PubMed](#)]
128. Rodriguez, J.A.; Nespereira, B.; Perez-Illzarbe, M.; Eguinoa, E.; Paramo, J.A. Vitamins C and E prevent endothelial VEGF and VEGFR-2 overexpression induced by porcine hypercholesterolemic LDL. *Cardiovasc. Res.* **2005**, *65*, 665–673. [[CrossRef](#)]
129. Gorchach, A.; Diebold, I.; Schini-Kerth, V.B.; Berchner-Pfannschmidt, U.; Roth, U.; Brandes, R.P.; Kietzmann, T.; Busse, R. Thrombin activates the hypoxia-inducible factor-1 signaling pathway in vascular smooth muscle cells: Role of the p22(phox)-containing NADPH oxidase. *Circ. Res.* **2001**, *89*, 47–54. [[CrossRef](#)]
130. Semenza, G.L.; Wang, G.L. A nuclear factor induced by hypoxia via de novo protein synthesis binds to the human erythropoietin gene enhancer at a site required for transcriptional activation. *Mol. Cell. Biol.* **1992**, *12*, 5447–5454. [[CrossRef](#)]
131. Tian, H.; McKnight, S.L.; Russell, D.W. Endothelial PAS domain protein 1 (EPAS1), a transcription factor selectively expressed in endothelial cells. *Genes Dev.* **1997**, *11*, 72–82. [[CrossRef](#)]
132. Wiesener, M.S.; Jurgensen, J.S.; Rosenberger, C.; Scholze, C.K.; Horstrup, J.H.; Warnecke, C.; Mandriota, S.; Bechmann, I.; Frei, U.A.; Pugh, C.W.; et al. Widespread hypoxia-inducible expression of HIF-2alpha in distinct cell populations of different organs. *FASEB J.* **2003**, *17*, 271–273. [[CrossRef](#)] [[PubMed](#)]
133. Keith, B.; Johnson, R.S.; Simon, M.C. HIF1alpha and HIF2alpha: Sibling rivalry in hypoxic tumour growth and progression. *Nat. Rev. Cancer* **2011**, *12*, 9–22. [[CrossRef](#)] [[PubMed](#)]
134. Hu, C.J.; Wang, L.Y.; Chodosh, L.A.; Keith, B.; Simon, M.C. Differential roles of hypoxia-inducible factor 1alpha (HIF-1alpha) and HIF-2alpha in hypoxic gene regulation. *Mol. Cell. Biol.* **2003**, *23*, 9361–9374. [[CrossRef](#)] [[PubMed](#)]
135. Hu, C.J.; Sataur, A.; Wang, L.; Chen, H.; Simon, M.C. The N-terminal transactivation domain confers target gene specificity of hypoxia-inducible factors HIF-1alpha and HIF-2alpha. *Mol. Biol. Cell* **2007**, *18*, 4528–4542. [[CrossRef](#)] [[PubMed](#)]

136. Holmquist-Mengelbier, L.; Fredlund, E.; Lofstedt, T.; Noguera, R.; Navarro, S.; Nilsson, H.; Pietras, A.; Vallon-Christersson, J.; Borg, A.; Gradin, K.; et al. Recruitment of HIF-1alpha and HIF-2alpha to common target genes is differentially regulated in neuroblastoma: HIF-2alpha promotes an aggressive phenotype. *Cancer Cell* **2006**, *10*, 413–423. [[CrossRef](#)]
137. Koh, M.Y.; Lemos, R., Jr.; Liu, X.; Powis, G. The hypoxia-associated factor switches cells from HIF-1alpha- to HIF-2alpha-dependent signaling promoting stem cell characteristics, aggressive tumor growth and invasion. *Cancer Res.* **2011**, *71*, 4015–4027. [[CrossRef](#)]
138. Doedens, A.L.; Stockmann, C.; Rubinstein, M.P.; Liao, D.; Zhang, N.; DeNardo, D.G.; Coussens, L.M.; Karin, M.; Goldrath, A.W.; Johnson, R.S. Macrophage expression of hypoxia-inducible factor-1 alpha suppresses T-cell function and promotes tumor progression. *Cancer Res.* **2010**, *70*, 7465–7475. [[CrossRef](#)]
139. Takeda, N.; O’Dea, E.L.; Doedens, A.; Kim, J.W.; Weidemann, A.; Stockmann, C.; Asagiri, M.; Simon, M.C.; Hoffmann, A.; Johnson, R.S. Differential activation and antagonistic function of HIF-1alpha isoforms in macrophages are essential for NO homeostasis. *Genes Dev.* **2010**, *24*, 491–501. [[CrossRef](#)]
140. Lin, Q.; Cong, X.; Yun, Z. Differential hypoxic regulation of hypoxia-inducible factors 1alpha and 2alpha. *Mol. Cancer Res.* **2011**, *9*, 757–765. [[CrossRef](#)]
141. Rius, J.; Guma, M.; Schachtrup, C.; Akassoglou, K.; Zinkernagel, A.S.; Nizet, V.; Johnson, R.S.; Haddad, G.G.; Karin, M. NF-kappaB links innate immunity to the hypoxic response through transcriptional regulation of HIF-1alpha. *Nature* **2008**, *453*, 807–811. [[CrossRef](#)]
142. Toschi, A.; Lee, E.; Gadir, N.; Ohh, M.; Foster, D.A. Differential dependence of hypoxia-inducible factors 1 alpha and 2 alpha on mTORC1 and mTORC2. *J. Biol. Chem.* **2008**, *283*, 34495–34499. [[CrossRef](#)] [[PubMed](#)]
143. Finkel, T.; Deng, C.X.; Mostoslavsky, R. Recent progress in the biology and physiology of sirtuins. *Nature* **2009**, *460*, 587–591. [[CrossRef](#)] [[PubMed](#)]
144. Dioum, E.M.; Chen, R.; Alexander, M.S.; Zhang, Q.; Hogg, R.T.; Gerard, R.D.; Garcia, J.A. Regulation of hypoxia-inducible factor 2alpha signaling by the stress-responsive deacetylase sirtuin 1. *Science* **2009**, *324*, 1289–1293. [[CrossRef](#)] [[PubMed](#)]
145. Lim, J.H.; Lee, Y.M.; Chun, Y.S.; Chen, J.; Kim, J.E.; Park, J.W. Sirtuin 1 modulates cellular responses to hypoxia by deacetylating hypoxia-inducible factor 1alpha. *Mol. Cell* **2010**, *38*, 864–878. [[CrossRef](#)] [[PubMed](#)]
146. Semenza, G.L. Oxygen-dependent regulation of mitochondrial respiration by hypoxia-inducible factor 1. *Biochem. J.* **2007**, *405*, 1–9. [[CrossRef](#)]
147. Bhargava, P.; Schnellmann, R.G. Mitochondrial energetics in the kidney. *Nat. Rev. Nephrol.* **2017**, *13*, 629–646. [[CrossRef](#)]
148. Iyer, N.V.; Kotch, L.E.; Agani, F.; Leung, S.W.; Laughner, E.; Wenger, R.H.; Gassmann, M.; Gearhart, J.D.; Lawler, A.M.; Yu, A.Y.; et al. Cellular and developmental control of O2 homeostasis by hypoxia-inducible factor 1 alpha. *Genes Dev.* **1998**, *12*, 149–162. [[CrossRef](#)]
149. Kim, J.W.; Tchernyshyov, I.; Semenza, G.L.; Dang, C.V. HIF-1-mediated expression of pyruvate dehydrogenase kinase: A metabolic switch required for cellular adaptation to hypoxia. *Cell Metab.* **2006**, *3*, 177–185. [[CrossRef](#)]
150. Papandreou, I.; Cairns, R.A.; Fontana, L.; Lim, A.L.; Denko, N.C. HIF-1 mediates adaptation to hypoxia by actively downregulating mitochondrial oxygen consumption. *Cell Metab.* **2006**, *3*, 187–197. [[CrossRef](#)]
151. Rohlenova, K.; Veys, K.; Miranda-Santos, I.; De Bock, K.; Carmeliet, P. Endothelial cell metabolism in health and disease. *Trends Cell Biol.* **2018**, *28*, 224–236. [[CrossRef](#)]
152. De Bock, K.; Georgiadou, M.; Carmeliet, P. Role of endothelial cell metabolism in vessel sprouting. *Cell Metab.* **2013**, *18*, 634–647. [[CrossRef](#)] [[PubMed](#)]
153. Helmlinger, G.; Endo, M.; Ferrara, N.; Hlatky, L.; Jain, R.K. Formation of endothelial cell networks. *Nature* **2000**, *405*, 139–141. [[CrossRef](#)] [[PubMed](#)]
154. Dekker, R.J.; van Soest, S.; Fontijn, R.D.; Salamanca, S.; de Groot, P.G.; VanBavel, E.; Pannekoek, H.; Horrevoets, A.J. Prolonged fluid shear stress induces a distinct set of endothelial cell genes, most specifically lung Kruppel-like factor (KLF2). *Blood* **2002**, *100*, 1689–1698. [[CrossRef](#)] [[PubMed](#)]
155. Lin, Z.; Kumar, A.; SenBanerjee, S.; Staniszewski, K.; Parmar, K.; Vaughan, D.E.; Gimbrone, M.A., Jr.; Balasubramanian, V.; Garcia-Cardena, G.; Jain, M.K. Kruppel-like factor 2 (KLF2) regulates endothelial thrombotic function. *Circ. Res.* **2005**, *96*, e48–e57. [[CrossRef](#)] [[PubMed](#)]
156. Chiplunkar, A.R.; Curtis, B.C.; Eades, G.L.; Kane, M.S.; Fox, S.J.; Haar, J.L.; Lloyd, J.A. The Kruppel-like factor 2 and Kruppel-like factor 4 genes interact to maintain endothelial integrity in mouse embryonic vasculogenesis. *BMC Dev. Biol.* **2013**, *13*, 40. [[CrossRef](#)]

157. Tuomisto, T.T.; Lumivuori, H.; Kansanen, E.; Hakkinen, S.K.; Turunen, M.P.; van Thienen, J.V.; Horrevoets, A.J.; Levonen, A.L.; Yla-Herttuala, S. Simvastatin has an anti-inflammatory effect on macrophages via upregulation of an atheroprotective transcription factor, Kruppel-like factor 2. *Cardiovasc. Res.* **2008**, *78*, 175–184. [[CrossRef](#)]
158. Sweet, D.R.; Fan, L.; Hsieh, P.N.; Jain, M.K. Kruppel-Like Factors in Vascular Inflammation: Mechanistic Insights and Therapeutic Potential. *Front. Cardiovasc. Med.* **2018**, *5*, 6. [[CrossRef](#)]
159. Bhattacharya, R.; Senbanerjee, S.; Lin, Z.; Mir, S.; Hamik, A.; Wang, P.; Mukherjee, P.; Mukhopadhyay, D.; Jain, M.K. Inhibition of vascular permeability factor/vascular endothelial growth factor-mediated angiogenesis by the Kruppel-like factor KLF2. *J. Biol. Chem.* **2005**, *280*, 28848–28851. [[CrossRef](#)]
160. Doddaballapur, A.; Michalik, K.M.; Manavski, Y.; Lucas, T.; Houtkooper, R.H.; You, X.; Chen, W.; Zeiher, A.M.; Potente, M.; Dimmeler, S.; et al. Laminar shear stress inhibits endothelial cell metabolism via KLF2-mediated repression of PFKFB3. *Arterioscler. Thromb. Vasc. Biol.* **2015**, *35*, 137–145. [[CrossRef](#)]
161. De Bock, K.; Georgiadou, M.; Schoors, S.; Kuchnio, A.; Wong, B.W.; Cantelmo, A.R.; Quaegebeur, A.; Ghesquiere, B.; Cauwenberghs, S.; Eelen, G.; et al. Role of PFKFB3-driven glycolysis in vessel sprouting. *Cell* **2013**, *154*, 651–663. [[CrossRef](#)]
162. Schoors, S.; De Bock, K.; Cantelmo, A.R.; Georgiadou, M.; Ghesquiere, B.; Cauwenberghs, S.; Kuchnio, A.; Wong, B.W.; Quaegebeur, A.; Goveia, J.; et al. Partial and transient reduction of glycolysis by PFKFB3 blockade reduces pathological angiogenesis. *Cell Metab.* **2014**, *19*, 37–48. [[CrossRef](#)] [[PubMed](#)]
163. Kawanami, D.; Mahabeleshwar, G.H.; Lin, Z.; Atkins, G.B.; Hamik, A.; Haldar, S.M.; Maemura, K.; Lamanna, J.C.; Jain, M.K. Kruppel-like factor 2 inhibits hypoxia-inducible factor 1alpha expression and function in the endothelium. *J. Biol. Chem.* **2009**, *284*, 20522–20530. [[CrossRef](#)] [[PubMed](#)]
164. Resar, J.R.; Roguin, A.; Voner, J.; Nasir, K.; Hennebry, T.A.; Miller, J.M.; Ingersoll, R.; Kasch, L.M.; Semenza, G.L. Hypoxia-inducible factor 1alpha polymorphism and coronary collaterals in patients with ischemic heart disease. *Chest* **2005**, *128*, 787–791. [[CrossRef](#)] [[PubMed](#)]
165. Bosch-Marce, M.; Okuyama, H.; Wesley, J.B.; Sarkar, K.; Kimura, H.; Liu, Y.V.; Zhang, H.; Strazza, M.; Rey, S.; Savino, L.; et al. Effects of aging and hypoxia-inducible factor-1 activity on angiogenic cell mobilization and recovery of perfusion after limb ischemia. *Circ. Res.* **2007**, *101*, 1310–1318. [[CrossRef](#)] [[PubMed](#)]
166. Sluimer, J.C.; Gasc, J.M.; van Wanroij, J.L.; Kisters, N.; Groeneweg, M.; Sollewijn Gelpke, M.D.; Cleutjens, J.P.; van den Akker, L.H.; Corvol, P.; Wouters, B.G.; et al. Hypoxia, hypoxia-inducible transcription factor, and macrophages in human atherosclerotic plaques are correlated with intraplaque angiogenesis. *J. Am. Coll. Cardiol.* **2008**, *51*, 1258–1265. [[CrossRef](#)]
167. Vorp, D.A.; Lee, P.C.; Wang, D.H.; Makaroun, M.S.; Nemoto, E.M.; Ogawa, S.; Webster, M.W. Association of intraluminal thrombus in abdominal aortic aneurysm with local hypoxia and wall weakening. *J. Vasc. Surg.* **2001**, *34*, 291–299. [[CrossRef](#)]
168. Lee, J.D.; Lai, C.H.; Yang, W.K.; Lee, T.H. Increased expression of hypoxia-inducible factor-1alpha and metallothionein in varicocele and varicose veins. *Phlebology* **2012**, *27*, 409–415. [[CrossRef](#)]
169. Evans, C.E.; Humphries, J.; Mattock, K.; Waltham, M.; Wadoodi, A.; Saha, P.; Modarai, B.; Maxwell, P.H.; Smith, A. Hypoxia and upregulation of hypoxia-inducible factor 1{alpha} stimulate venous thrombus recanalization. *Arterioscler. Thromb. Vasc. Biol.* **2010**, *30*, 2443–2451. [[CrossRef](#)]
170. Evans, C.E.; Humphries, J.; Waltham, M.; Saha, P.; Mattock, K.; Patel, A.; Ahmad, A.; Wadoodi, A.; Modarai, B.; Burnand, K.; et al. Upregulation of hypoxia-inducible factor 1 alpha in local vein wall is associated with enhanced venous thrombus resolution. *Thromb. Res.* **2011**, *128*, 346–351. [[CrossRef](#)]
171. Semenza, G.L. Hypoxia-inducible factors in physiology and medicine. *Cell* **2012**, *148*, 399–408. [[CrossRef](#)]
172. Lim, C.S.; Kiriakidis, S.; Sandison, A.; Paleolog, E.M.; Davies, A.H. Hypoxia-inducible factor pathway and diseases of the vascular wall. *J. Vasc. Surg.* **2013**, *58*, 219–230. [[CrossRef](#)] [[PubMed](#)]
173. Hansson, G.K. Inflammation, atherosclerosis, and coronary artery disease. *N. Engl. J. Med.* **2005**, *352*, 1685–1695. [[CrossRef](#)] [[PubMed](#)]
174. Lusis, A.J. Atherosclerosis. *Nature* **2000**, *407*, 233–241. [[CrossRef](#)] [[PubMed](#)]
175. Murdoch, C.; Muthana, M.; Lewis, C.E. Hypoxia regulates macrophage functions in inflammation. *J. Immunol.* **2005**, *175*, 6257–6263. [[CrossRef](#)]
176. Parathath, S.; Mick, S.L.; Feig, J.E.; Joaquin, V.; Grauer, L.; Habel, D.M.; Gassmann, M.; Gardner, L.B.; Fisher, E.A. Hypoxia is present in murine atherosclerotic plaques and has multiple adverse effects on macrophage lipid metabolism. *Circ. Res.* **2011**, *109*, 1141–1152. [[CrossRef](#)]

177. Cramer, T.; Yamanishi, Y.; Clausen, B.E.; Forster, I.; Pawlinski, R.; Mackman, N.; Haase, V.H.; Jaenisch, R.; Corr, M.; Nizet, V.; et al. HIF-1alpha is essential for myeloid cell-mediated inflammation. *Cell* **2003**, *112*, 645–657. [[CrossRef](#)]
178. Burke, B.; Giannoudis, A.; Corke, K.P.; Gill, D.; Wells, M.; Ziegler-Heitbrock, L.; Lewis, C.E. Hypoxia-induced gene expression in human macrophages: Implications for ischemic tissues and hypoxia-regulated gene therapy. *Am. J. Pathol.* **2003**, *163*, 1233–1243. [[CrossRef](#)]
179. Lobanova, E.M.; Tahanovich, A.D. Hydrogen peroxide metabolism in alveolar macrophages after exposure to hypoxia and heat. *Physiol. Res.* **2006**, *55*, 569–575.
180. Rydberg, E.K.; Krettek, A.; Ullstrom, C.; Ekstrom, K.; Svensson, P.A.; Carlsson, L.M.; Jonsson-Rylander, A.C.; Hansson, G.I.; McPheat, W.; Wiklund, O.; et al. Hypoxia increases LDL oxidation and expression of 15-lipoxygenase-2 in human macrophages. *Arterioscler. Thromb. Vasc. Biol.* **2004**, *24*, 2040–2045. [[CrossRef](#)]
181. Lattimore, J.D.; Wilcox, I.; Nakhla, S.; Langenfeld, M.; Jessup, W.; Celermajer, D.S. Repetitive hypoxia increases lipid loading in human macrophages—a potentially atherogenic effect. *Atherosclerosis* **2005**, *179*, 255–259. [[CrossRef](#)]
182. Luque, A.; Turu, M.; Juan-Babot, O.; Cardona, P.; Font, A.; Carvajal, A.; Slevin, M.; Iborra, E.; Rubio, F.; Badimon, L.; et al. Overexpression of hypoxia/inflammatory markers in atherosclerotic carotid plaques. *Front. Biosci.* **2008**, *13*, 6483–6490. [[CrossRef](#)] [[PubMed](#)]
183. Vink, A.; Schoneveld, A.H.; Lamers, D.; Houben, A.J.; van der Groep, P.; van Diest, P.J.; Pasterkamp, G. HIF-1 alpha expression is associated with an atheromatous inflammatory plaque phenotype and upregulated in activated macrophages. *Atherosclerosis* **2007**, *195*, e69–e75. [[CrossRef](#)] [[PubMed](#)]
184. Higashida, T.; Kanno, H.; Nakano, M.; Funakoshi, K.; Yamamoto, I. Expression of hypoxia-inducible angiogenic proteins (hypoxia-inducible factor-1alpha, vascular endothelial growth factor, and E26 transformation-specific-1) and plaque hemorrhage in human carotid atherosclerosis. *J. Neurosurg.* **2008**, *109*, 83–91. [[CrossRef](#)] [[PubMed](#)]
185. Jiang, G.; Li, T.; Qiu, Y.; Rui, Y.; Chen, W.; Lou, Y. RNA interference for HIF-1alpha inhibits foam cells formation in vitro. *Eur. J. Pharmacol.* **2007**, *562*, 183–190. [[CrossRef](#)] [[PubMed](#)]
186. Blouin, C.C.; Page, E.L.; Soucy, G.M.; Richard, D.E. Hypoxic gene activation by lipopolysaccharide in macrophages: Implication of hypoxia-inducible factor 1alpha. *Blood* **2004**, *103*, 1124–1130. [[CrossRef](#)] [[PubMed](#)]
187. Cramer, T.; Johnson, R.S. A novel role for the hypoxia inducible transcription factor HIF-1alpha: Critical regulation of inflammatory cell function. *Cell Cycle* **2003**, *2*, 192–193. [[CrossRef](#)]
188. Lu, D.Y.; Chen, E.Y.; Wong, D.J.; Yamamoto, K.; Protack, C.D.; Williams, W.T.; Assi, R.; Hall, M.R.; Sadaghianloo, N.; Dardik, A. Vein graft adaptation and fistula maturation in the arterial environment. *J. Surg. Res.* **2014**, *188*, 162–173. [[CrossRef](#)]
189. Lee, E.S.; Bauer, G.E.; Caldwell, M.P.; Santilli, S.M. Association of artery wall hypoxia and cellular proliferation at a vascular anastomosis. *J. Surg. Res.* **2000**, *91*, 32–37. [[CrossRef](#)]
190. Santilli, S.M.; Wernsing, S.E.; Lee, E.S. Transarterial wall oxygen gradients at a prosthetic vascular graft to artery anastomosis in the rabbit. *J. Vasc. Surg.* **2000**, *31*, 1229–1239. [[CrossRef](#)]
191. Hughes, D.; Fu, A.A.; Puggioni, A.; Glockner, J.F.; Anwer, B.; McGuire, A.M.; Mukhopadhyay, D.; Misra, S. Adventitial transplantation of blood outgrowth endothelial cells in porcine haemodialysis grafts alleviates hypoxia and decreases neointimal proliferation through a matrix metalloproteinase-9-mediated pathway—A pilot study. *Nephrol. Dial. Transplant.* **2009**, *24*, 85–96. [[CrossRef](#)]
192. Nakao, A.; Huang, C.S.; Stolz, D.B.; Wang, Y.; Franks, J.M.; Tochigi, N.; Billiar, T.R.; Toyoda, Y.; Tzeng, E.; McCurry, K.R. Ex vivo carbon monoxide delivery inhibits intimal hyperplasia in arterialized vein grafts. *Cardiovasc. Res.* **2011**, *89*, 457–463. [[CrossRef](#)] [[PubMed](#)]
193. Rochette, L.; Cottin, Y.; Zeller, M.; Vergely, C. Carbon monoxide: Mechanisms of action and potential clinical implications. *Pharmacol. Ther.* **2013**, *137*, 133–152. [[CrossRef](#)] [[PubMed](#)]
194. Misra, S.; Fu, A.A.; Rajan, D.K.; Juncos, L.A.; McKusick, M.A.; Bjarnason, H.; Mukhopadhyay, D. Expression of hypoxia inducible factor-1 $\alpha$ , macrophage migration inhibition factor, matrix metalloproteinase-2 and -9, and their inhibitors in hemodialysis grafts and arteriovenous fistulas. *J. Vasc. Interv. Radiol.* **2008**, *19*, 252–259. [[CrossRef](#)] [[PubMed](#)]

195. Fu, H.; Luo, F.; Yang, L.; Wu, W.; Liu, X. Hypoxia stimulates the expression of macrophage migration inhibitory factor in human vascular smooth muscle cells via HIF-1 $\alpha$  dependent pathway. *BMC Cell Biol.* **2010**, *11*, 66. [[CrossRef](#)]
196. Misra, S.; Fu, A.A.; Puggioni, A.; Glockner, J.F.; McKusick, M.A.; Bjarnason, H.; Mukhopadhyay, D. Proteomic Profiling in Early Venous Stenosis Formation in a Porcine Model of Hemodialysis Graft. *J. Vasc. Interv. Radiol.* **2009**, *20*, 241–251. [[CrossRef](#)] [[PubMed](#)]
197. Misra, S.; Fu, A.A.; Puggioni, A.; Glockner, J.F.; Rajan, D.K.; McKusick, M.A.; Bjarnason, H.; Mukhopadhyay, D. Increased Expression of Hypoxia-inducible Factor-1 $\alpha$  in venous stenosis of arteriovenous polytetrafluoroethylene grafts in a chronic renal insufficiency porcine model. *J. Vasc. Interv. Radiol.* **2008**, *19*, 260–265. [[CrossRef](#)] [[PubMed](#)]
198. Misra, S.; Shergill, U.; Yang, B.; Janardhanan, R.; Misra, K.D. Increased Expression of HIF-1 $\alpha$ , VEGF-A and Its Receptors, MMP-2, TIMP-1, and ADAMTS-1 at the venous stenosis of arteriovenous fistula in a mouse model with renal insufficiency. *J. Vasc. Interv. Radiol.* **2010**, *21*, 1255–1261. [[CrossRef](#)] [[PubMed](#)]
199. Sadaghianloo, N.; Yamamoto, K.; Bai, H.; Tsuneki, M.; Protack, C.D.; Hall, M.R.; Declémy, S.; Hassen-Khodja, R.; Madri, J.; Dardik, A. Increased oxidative stress and hypoxia inducible factor-1 expression during arteriovenous fistula maturation. *Ann. Vasc. Surg.* **2017**, *41*, 225–234. [[CrossRef](#)]
200. Lin, C.C.; Chung, M.Y.; Yang, W.C.; Lin, S.J.; Lee, P.C. Length polymorphisms of heme oxygenase-1 determine the effect of far-infrared therapy on the function of arteriovenous fistula in hemodialysis patients: A novel physicogenomic study. *Nephrol. Dial. Transplant.* **2013**, *28*, 1284–1293. [[CrossRef](#)]
201. Lin, C.C.; Yang, W.C.; Lin, S.J.; Chen, T.W.; Lee, W.S.; Chang, C.F.; Lee, P.C.; Lee, S.D.; Su, T.S.; Fann, C.S.; et al. Length polymorphism in heme oxygenase-1 is associated with arteriovenous fistula patency in hemodialysis patients. *Kidney Int.* **2006**, *69*, 165–172. [[CrossRef](#)]
202. Kang, L.; Grande, J.P.; Farrugia, G.; Croatt, A.J.; Katusic, Z.S.; Nath, K.A. Functioning of an arteriovenous fistula requires heme oxygenase-2. *Am. J. Physiol. Renal Physiol.* **2013**, *305*, F545–F552. [[CrossRef](#)] [[PubMed](#)]
203. Kang, L.; Grande, J.P.; Hillestad, M.L.; Croatt, A.J.; Barry, M.A.; Katusic, Z.S.; Nath, K.A. A new model of an arteriovenous fistula in chronic kidney disease in the mouse: Beneficial effects of upregulated heme oxygenase-1. *Am. J. Physiol. Renal Physiol.* **2016**, *310*, F466–F476. [[CrossRef](#)] [[PubMed](#)]
204. Lever, J.M.; Boddu, R.; George, J.F.; Agarwal, A. Heme oxygenase-1 in kidney health and disease. *Antioxid. Redox Signal.* **2016**, *25*, 165–183. [[CrossRef](#)] [[PubMed](#)]
205. Adin, C.A.; Croker, B.P.; Agarwal, A. Protective effects of exogenous bilirubin on ischemia-reperfusion injury in the isolated, perfused rat kidney. *Am. J. Physiol. Renal Physiol.* **2005**, *288*, F778–F784. [[CrossRef](#)]
206. Juncos, J.P.; Tracz, M.J.; Croatt, A.J.; Grande, J.P.; Ackerman, A.W.; Katusic, Z.S.; Nath, K.A. Genetic deficiency of heme oxygenase-1 impairs functionality and form of an arteriovenous fistula in the mouse. *Kidney Int.* **2008**, *74*, 47–51. [[CrossRef](#)]
207. Freidja, M.L.; Toutain, B.; Caillon, A.; Desquret, V.; Lambert, D.; Loufrani, L.; Procaccio, V.; Henrion, D. Heme oxygenase 1 is differentially involved in blood flow-dependent arterial remodeling: Role of inflammation, oxidative stress, and nitric oxide. *Hypertension* **2011**, *58*, 225–231. [[CrossRef](#)]
208. Zohny, S.F.; Abd el-Fattah, M. Evaluation of circulating vascular endothelial growth factor and soluble adhesion molecules as reliable predictors of native arteriovenous fistula thrombosis in chronic hemodialysis patients. *Clin. Biochem.* **2008**, *41*, 1175–1180. [[CrossRef](#)]
209. Candan, F.; Yildiz, G.; Kayatas, M. Role of the VEGF 936 gene polymorphism and VEGF-A levels in the late-term arteriovenous fistula thrombosis in patients undergoing hemodialysis. *Int. Urol. Nephrol.* **2014**, *46*, 1815–1823. [[CrossRef](#)]
210. Ohtani, K.; Egashira, K.; Hiasa, K.; Zhao, Q.; Kitamoto, S.; Ishibashi, M.; Usui, M.; Inoue, S.; Yonemitsu, Y.; Sueishi, K.; et al. Blockade of vascular endothelial growth factor suppresses experimental restenosis after intraluminal injury by inhibiting recruitment of monocyte lineage cells. *Circulation* **2004**, *110*, 2444–2452. [[CrossRef](#)]
211. Rapisarda, A.; Uranchimeg, B.; Scudiero, D.A.; Selby, M.; Sausville, E.A.; Shoemaker, R.H.; Melillo, G. Identification of small molecule inhibitors of hypoxia-inducible factor 1 transcriptional activation pathway. *Cancer Res.* **2002**, *62*, 4316–4324.
212. Lata, C.; Green, D.; Wan, J.; Roy, S.; Santilli, S.M. The role of short-term oxygen administration in the prevention of intimal hyperplasia. *J. Vasc. Surg.* **2013**, *58*, 452–459. [[CrossRef](#)] [[PubMed](#)]

213. Wan, J.; Lata, C.; Santilli, A.; Green, D.; Roy, S.; Santilli, S. Supplemental oxygen reverses hypoxia-induced smooth muscle cell proliferation by modulating HIF- $\alpha$  and VEGF levels in a rabbit arteriovenous fistula model. *Ann. Vasc. Surg.* **2014**, *28*, 725–736. [[CrossRef](#)] [[PubMed](#)]
214. Terraneo, L.; Paroni, R.; Bianciardi, P.; Giallongo, T.; Carelli, S.; Gorio, A.; Samaja, M. Brain adaptation to hypoxia and hyperoxia in mice. *Redox Biol.* **2017**, *11*, 12–20. [[CrossRef](#)] [[PubMed](#)]
215. Terraneo, L.; Virgili, E.; Caretti, A.; Bianciardi, P.; Samaja, M. In vivo hyperoxia induces hypoxia-inducible factor-1 $\alpha$  overexpression in LNCaP tumors without affecting the tumor growth rate. *Int. J. Biochem. Cell Biol.* **2014**, *51C*, 65–74. [[CrossRef](#)]
216. Marconi, G.D.; Zara, S.; De Colli, M.; Di Valerio, V.; Rapino, M.; Zaramella, P.; Dedja, A.; Macchi, V.; De Caro, R.; Porzionato, A. Postnatal hyperoxia exposure differentially affects hepatocytes and liver haemopoietic cells in newborn rats. *PLoS ONE* **2014**, *9*, e105005. [[CrossRef](#)]
217. Zara, S.; Macchi, V.; De Caro, R.; Rapino, M.; Cataldi, A.; Porzionato, A. pPKC $\alpha$  mediated-HIF-1 $\alpha$  activation related to the morphological modifications occurring in neonatal myocardial tissue in response to severe and mild hyperoxia. *Eur. J. Histochem.* **2012**, *56*, e2. [[CrossRef](#)]
218. Vandegriff, K.D.; Malavalli, A.; Lohman, J.; Young, M.A.; Terraneo, L.; Virgili, E.; Bianciardi, P.; Caretti, A.; Samaja, M. Impact of acellular hemoglobin-based oxygen carriers on brain apoptosis in rats. *Transfusion* **2014**, *54*, 45–54. [[CrossRef](#)]
219. Wigerup, C.; Pahlman, S.; Bexell, D. Therapeutic targeting of hypoxia and hypoxia-inducible factors in cancer. *Pharmacol. Ther.* **2016**, *164*, 152–169. [[CrossRef](#)]
220. Melillo, G. Targeting hypoxia cell signaling for cancer therapy. *Cancer Metastasis Rev.* **2007**, *26*, 341–352. [[CrossRef](#)]
221. Shin, D.H.; Chun, Y.S.; Lee, D.S.; Huang, L.E.; Park, J.W. Bortezomib inhibits tumor adaptation to hypoxia by stimulating the FIH-mediated repression of hypoxia-inducible factor-1. *Blood* **2008**, *111*, 3131–3136. [[CrossRef](#)]
222. Greenberger, L.M.; Horak, I.D.; Filpula, D.; Sapra, P.; Westergaard, M.; Frydenlund, H.F.; Albaek, C.; Schroder, H.; Orum, H. A RNA antagonist of hypoxia-inducible factor-1 $\alpha$ , EZN-2968, inhibits tumor cell growth. *Mol. Cancer Ther.* **2008**, *7*, 3598–3608. [[CrossRef](#)] [[PubMed](#)]
223. Terzuoli, E.; Puppo, M.; Rapisarda, A.; Uranchimeg, B.; Cao, L.; Burger, A.M.; Ziche, M.; Melillo, G. Aminoflavone, a ligand of the aryl hydrocarbon receptor, inhibits HIF-1 $\alpha$  expression in an AhR-independent fashion. *Cancer Res.* **2010**, *70*, 6837–6848. [[CrossRef](#)] [[PubMed](#)]
224. Rapisarda, A.; Uranchimeg, B.; Sordet, O.; Pommier, Y.; Shoemaker, R.H.; Melillo, G. Topoisomerase I-mediated inhibition of hypoxia-inducible factor 1: Mechanism and therapeutic implications. *Cancer Res.* **2004**, *64*, 1475–1482. [[CrossRef](#)] [[PubMed](#)]
225. Sapra, P.; Zhao, H.; Mehlig, M.; Malaby, J.; Kraft, P.; Longley, C.; Greenberger, L.M.; Horak, I.D. Novel delivery of SN38 markedly inhibits tumor growth in xenografts, including a camptothecin-11-refractory model. *Clin. Cancer Res.* **2008**, *14*, 1888–1896. [[CrossRef](#)] [[PubMed](#)]
226. Zhang, H.; Qian, D.Z.; Tan, Y.S.; Lee, K.; Gao, P.; Ren, Y.R.; Rey, S.; Hammers, H.; Chang, D.; Pili, R.; et al. Digoxin and other cardiac glycosides inhibit HIF-1 $\alpha$  synthesis and block tumor growth. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 19579–19586. [[CrossRef](#)] [[PubMed](#)]
227. Del Bufalo, D.; Ciuffreda, L.; Trisciuglio, D.; Desideri, M.; Cognetti, F.; Zupi, G.; Milella, M. Antiangiogenic potential of the Mammalian target of rapamycin inhibitor temsirolimus. *Cancer Res.* **2006**, *66*, 5549–5554. [[CrossRef](#)] [[PubMed](#)]
228. Manegold, P.C.; Paringer, C.; Kulka, U.; Krimmel, K.; Eichhorn, M.E.; Wilkowski, R.; Jauch, K.W.; Guba, M.; Bruns, C.J. Antiangiogenic therapy with mammalian target of rapamycin inhibitor RAD001 (everolimus) increases radiosensitivity in solid cancer. *Clin. Cancer Res.* **2008**, *14*, 892–900. [[CrossRef](#)]
229. Majumder, P.K.; Febbo, P.G.; Bikoff, R.; Berger, R.; Xue, Q.; McMahon, L.M.; Manola, J.; Brugarolas, J.; McDonnell, T.J.; Golub, T.R.; et al. mTOR inhibition reverses Akt-dependent prostate intraepithelial neoplasia through regulation of apoptotic and HIF-1-dependent pathways. *Nat. Med.* **2004**, *10*, 594–601. [[CrossRef](#)]
230. Isaacs, J.S.; Jung, Y.J.; Mimnaugh, E.G.; Martinez, A.; Cuttitta, F.; Neckers, L.M. Hsp90 regulates a von Hippel Lindau-independent hypoxia-inducible factor-1  $\alpha$ -degradative pathway. *J. Biol. Chem.* **2002**, *277*, 29936–29944. [[CrossRef](#)]

231. Hur, E.; Kim, H.H.; Choi, S.M.; Kim, J.H.; Yim, S.; Kwon, H.J.; Choi, Y.; Kim, D.K.; Lee, M.O.; Park, H. Reduction of hypoxia-induced transcription through the repression of hypoxia-inducible factor-1alpha/aryl hydrocarbon receptor nuclear translocator DNA binding by the 90-kDa heat-shock protein inhibitor radicicol. *Mol. Pharmacol.* **2002**, *62*, 975–982. [[CrossRef](#)]
232. Ellis, L.; Hammers, H.; Pili, R. Targeting tumor angiogenesis with histone deacetylase inhibitors. *Cancer Lett.* **2009**, *280*, 145–153. [[CrossRef](#)] [[PubMed](#)]
233. Kong, D.; Park, E.J.; Stephen, A.G.; Calvani, M.; Cardellina, J.H.; Monks, A.; Fisher, R.J.; Shoemaker, R.H.; Melillo, G. Echinomycin, a small-molecule inhibitor of hypoxia-inducible factor-1 DNA-binding activity. *Cancer Res.* **2005**, *65*, 9047–9055. [[CrossRef](#)] [[PubMed](#)]
234. Lee, K.; Qian, D.Z.; Rey, S.; Wei, H.; Liu, J.O.; Semenza, G.L. Anthracycline chemotherapy inhibits HIF-1 transcriptional activity and tumor-induced mobilization of circulating angiogenic cells. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 2353–2358. [[CrossRef](#)] [[PubMed](#)]
235. Rey, S.; Lee, K.; Wang, C.J.; Gupta, K.; Chen, S.; McMillan, A.; Bhise, N.; Levchenko, A.; Semenza, G.L. Synergistic effect of HIF-1alpha gene therapy and HIF-1-activated bone marrow-derived angiogenic cells in a mouse model of limb ischemia. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 20399–20404. [[CrossRef](#)] [[PubMed](#)]
236. Rajagopalan, S.; Olin, J.; Deitcher, S.; Pieczek, A.; Laird, J.; Grossman, P.M.; Goldman, C.K.; McEllin, K.; Kelly, R.; Chronos, N. Use of a constitutively active hypoxia-inducible factor-1alpha transgene as a therapeutic strategy in no-option critical limb ischemia patients: Phase I dose-escalation experience. *Circulation* **2007**, *115*, 1234–1243. [[CrossRef](#)]
237. Creager, M.A.; Olin, J.W.; Belch, J.J.; Moneta, G.L.; Henry, T.D.; Rajagopalan, S.; Annex, B.H.; Hiatt, W.R. Effect of hypoxia-inducible factor-1alpha gene therapy on walking performance in patients with intermittent claudication. *Circulation* **2011**, *124*, 1765–1773. [[CrossRef](#)]
238. Gupta, N.; Wish, J.B. Hypoxia-inducible factor prolyl hydroxylase inhibitors: A potential new treatment for anemia in patients with CKD. *Am. J. Kidney Dis.* **2017**, *69*, 815–826. [[CrossRef](#)]
239. Brigandi, R.A.; Johnson, B.; Oei, C.; Westerman, M.; Olbina, G.; de Zoysa, J.; Roger, S.D.; Sahay, M.; Cross, N.; McMahon, L.; et al. A novel hypoxia-inducible factor-prolyl hydroxylase inhibitor (GSK1278863) for anemia in CKD: A 28-day, phase 2A randomized trial. *Am. J. Kidney Dis.* **2016**, *67*, 861–871. [[CrossRef](#)]
240. Provenzano, R.; Besarab, A.; Wright, S.; Dua, S.; Zeig, S.; Nguyen, P.; Poole, L.; Saikali, K.G.; Saha, G.; Hemmerich, S.; et al. Roxadustat (FG-4592) versus epoetin alfa for anemia in patients receiving maintenance hemodialysis: A phase 2, randomized, 6- to 19-week, open-label, active-comparator, dose-ranging, safety and exploratory efficacy study. *Am. J. Kidney Dis.* **2016**, *67*, 912–924. [[CrossRef](#)]
241. Onnis, B.; Rapisarda, A.; Melillo, G. Development of HIF-1 inhibitors for cancer therapy. *J. Cell. Mol. Med.* **2009**, *13*, 2780–2786. [[CrossRef](#)]
242. Chen, W.; Hill, H.; Christie, A.; Kim, M.S.; Holloman, E.; Pavia-Jimenez, A.; Homayoun, F.; Ma, Y.; Patel, N.; Yell, P.; et al. Targeting renal cell carcinoma with a HIF-2 antagonist. *Nature* **2016**, *539*, 112–117. [[CrossRef](#)] [[PubMed](#)]
243. Scheuermann, T.H.; Li, Q.; Ma, H.W.; Key, J.; Zhang, L.; Chen, R.; Garcia, J.A.; Naidoo, J.; Longgood, J.; Frantz, D.E.; et al. Allosteric inhibition of hypoxia inducible factor-2 with small molecules. *Nat. Chem. Biol.* **2013**, *9*, 271–276. [[CrossRef](#)] [[PubMed](#)]
244. Cho, H.; Du, X.; Rizzi, J.P.; Liberzon, E.; Chakraborty, A.A.; Gao, W.; Carvo, I.; Signoretti, S.; Bruick, R.K.; Josey, J.A.; et al. On-target efficacy of a HIF-2alpha antagonist in preclinical kidney cancer models. *Nature* **2016**, *539*, 107–111. [[CrossRef](#)] [[PubMed](#)]
245. Wallace, E.M.; Rizzi, J.P.; Han, G.; Wehn, P.M.; Cao, Z.; Du, X.; Cheng, T.; Czerwinski, R.M.; Dixon, D.D.; Goggin, B.S.; et al. A small-molecule antagonist of HIF2alpha Is efficacious in preclinical models of renal cell carcinoma. *Cancer Res.* **2016**, *76*, 5491–5500. [[CrossRef](#)] [[PubMed](#)]

