

Comprehensive Analysis of CRISPR/Cas9-Mediated Mutagenesis in *Arabidopsis thaliana* by Genome-wide Sequencing

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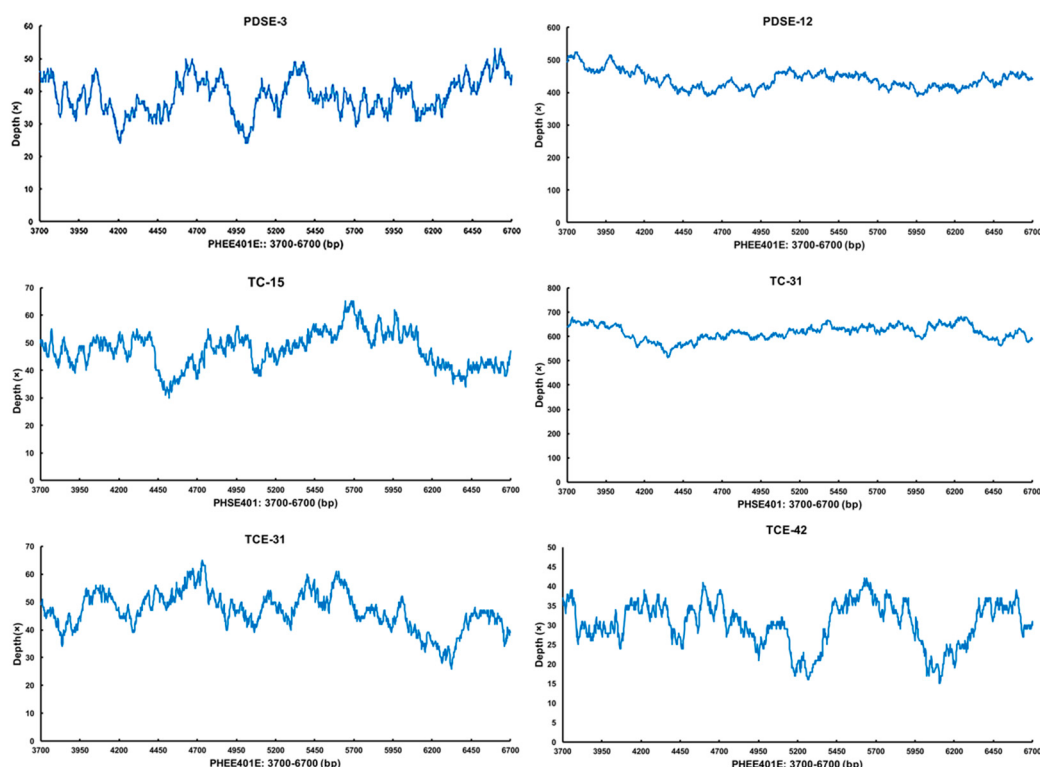


Figure S1. Sequencing depth distribution of inserted vector in the CRISPR/Cas9-edited mutants. The partially continuous specific sequences of two vectors were used for this plot.

Table S1. Sequencing depth and mapping ratio of samples for WGS.

Sample	Sequencing Depth(x)	Mapping Ratio(%)
WT1	94.43	92.61
WT2	84.67	92.86
WT3	87.94	92.74
PDSE-3	139.93	94.36
PDSE-12	132.61	94.43
TC-15	138.52	94.7
TC-31	127.79	94.06
TCE-31	126.64	93.78
TCE-42	95.85	93.67

Table S2. The number of structural variants of mutants.

Mutant	PDSE-12	PDSE-3	TC-15	TC-31	TCE-31	TCE-42
SV	63	72	77	78	80	19
SV (<10kb)	18	16	16	15	24	4

Table S3 Editing efficiency of targets for deep sequencing.

Name	WT	PDSE-3	PDSE-MIX	TC-15	TC-31	TC-MIX	TCE-MIX
<i>PDS-1_target</i>	3.30E-04	2.59E-04	2.21E-04	-	-	-	-
<i>PDS-1-1</i>	5.69E-05	9.10E-05	5.97E-05	-	-	-	-
<i>PDS-1-2</i>	1.74E-04	1.94E-04	1.80E-04	-	-	-	-
<i>PDS-1-3</i>	4.01E-04	4.43E-04	3.32E-04	-	-	-	-
<i>PDS-1-4</i>	1.08E-04	9.09E-05	1.06E-04	-	-	-	-
<i>PDS-2_target</i>	1.28E-02	4.95E-01	9.20E-01	-	-	-	-
<i>PDS-2-1</i>	2.35E-04	2.39E-04	2.08E-04	-	-	-	-
<i>PDS-2-2</i>	7.87E-05	4.94E-05	9.54E-05	-	-	-	-
<i>CPC_target</i>	1.41E-02	-	-	9.95E-01	9.95E-01	9.67E-01	8.88E-01
<i>CPC-1</i>	9.64E-05	-	-	1.40E-04	1.55E-04	1.13E-04	1.23E-04
<i>CPC-2</i>	1.08E-04	-	-	1.31E-04	1.15E-04	1.44E-04	1.58E-04
<i>CPC-3</i>	1.62E-04	-	-	1.54E-04	1.39E-04	2.19E-04	1.90E-04
<i>CPC-4</i>	4.66E-04	-	-	6.07E-04	5.92E-04	3.72E-04	3.46E-04
<i>CPC-5</i>	1.68E-04	-	-	1.23E-04	1.12E-04	1.54E-04	1.67E-04
<i>TRY_target</i>	2.16E-03	-	-	9.93E-01	9.96E-01	7.24E-01	9.30E-01
<i>TRY-1</i>	1.41E-04	-	-	5.38E-04	2.28E-04	1.85E-04	1.65E-04
<i>TRY-2</i>	2.73E-03	-	-	1.86E-01	9.81E-01	1.61E-02	9.24E-02
<i>TRY-3</i>	2.53E-04	-	-	1.41E-04	1.28E-04	1.71E-04	1.64E-04
<i>TRY-4</i>	4.94E-05	-	-	5.00E-05	3.93E-05	5.20E-05	4.62E-05
<i>TRY-5</i>	1.82E-04	-	-	-	-	-	-
<i>TRY-6</i>	1.40E-04	-	-	2.22E-04	1.61E-04	1.70E-04	1.66E-04
<i>TRY-7</i>	5.40E-04	-	-	5.16E-04	4.82E-04	6.01E-04	5.58E-04

“-” represents that the site in the sample was not detected.

Table S4 Re-analysis results for rice mutants.

Plant Name	NCBI SRA Accession	On Target	Off-target Site	Off-target Sequence
Cas9 Backbone-1	SRR6344862	-	-	-
Cas9 Backbone-2	SRR6344859	-	-	-
Cas9-E1	SRR6344819	Cas9-E	-	-
Cas9-E2	SRR6344879	Cas9-E	-	-
Cas9-J1	SRR6344848	sgRNA01	Chr9:17099504-17099516	GCAGCTCTGACATGTGGGCCCG G
			Chr5:19226195-19226217	ACAGCTCTGACATGTGGGCCCG G
			Chr2:23446231-23446253	GCAGCTCTGACATATGGGCCCG G
			Chr1:31035301-31035323	GCAGCTCTGACATGTGGGTCCG G
			Chr5:13975206-13975228	GGAGCTCTGACATGTGGGCCCG G
			Chr1:22043898-22043920	GCAGCTCTGACATGTGGGCCTG G
			Chr7:22958478-22958500	GCAGCACTGACATGTGGGCCCG G

Cas9-J2	SRR6344845	sgRNA02	Chr9:17967312-17967334	GCAGCTCTGACATGTAGGCCGG G
			Chr12:8340812-8340834	GCAGTTCTGACATGTGGGCCCA G
			Chr10:8570768-8570790	GTAGCTCTGACATATGGGCCGG G
			-	-
		sgRNA01	Chr9:17099504-17099516	GCAGCTCTGACATGTGGGCCCG G
		sgRNA02	Chr5:19226195-19226217	ACAGCTCTGACATGTGGGCCCG G
			Chr2:23446231-23446253	GCAGCTCTGACATATGGGCCCG G
			Chr1:31035301-31035323	GCAGCTCTGACATGTGGGTCCG G
			Chr5:13975206-13975228	GGAGCTCTGACATGTGGGCCCG G
			Chr1:22043898-22043920	GCAGCTCTGACATGTGGGCCTG G
			Chr6:24673722-24673744	GCAGCTCTGACATGTGGGCCTG G
			Chr1:19854951-19854973	ACAGCTCCGACATGTGGGCCCG G
			Chr9:17967312-17967334	GCAGCTCTGACATGTAGGCCGG G
			Chr3:2628500-2628522	GCAGCTCTGACATGTGGGTCTGG
			Chr1:4302680-4302702	ATAGCTCTGACATGTGGGCTCGG
			Chr10:8570768-8570790	GTAGCTCTGACATATGGGCCGG G
			-	-
			-	-
			-	-
			-	-
			-	-
			-	-

“-” represents that there is no off target in the mutant for the sgRNA. New off-target sites within 3 mismatches are highlighted in yellow.

Table S5 Target sequences for Digenome-seq.

Target	Sequence
<i>CPC</i>	ATCCCGGGACGGACGCCGAGG
<i>TRY</i>	GAAGTGAGCAGTATCGAATGGG
<i>PDS-1</i>	AGGGTGCTGGTAGGACATCTGG
<i>PDS-2</i>	GGATGGTAATCCTCCGAAAGG
<i>GLV1-1</i>	CTCTTTGGACGTCTTCAATGG
<i>GLV6-1</i>	GCTAGTGCACAAAGAAAGAGGG
<i>GLV7-2</i>	CTGAAACGAAGATGAAGAGAGG
<i>GLV8-2</i>	AAGACAAGCAATAAAGCTGAGG
<i>GLV10-1</i>	AAGGATCATTGAAGCAACATGG
<i>GLV10-2</i>	GATAATCTGCAAATAAGAGCGG

All target sites are listed in 5' to 3'.

Table S6 Primers used for PCR amplification and sequencing of target sites.

Target	Primer/F	Primer/R
<i>PDS-1</i>	TAAGCAGGATACCAGATGAA	CATGTCAAAGGCGCTAAA
<i>PDS-2</i>	TGTTCTTCCTGACGTCTAA	ACTCATACCCTCTCTGTTG
<i>CPC</i>	CGATGGAGGCTGGTAAA	CTGTCAATGACTGTGTTTCA
<i>TRY</i>	TCTGAAGGTACCTCTCTATTC	GCCAGCCAAGCATTAT

Table S7 Target sequences and primers used for target amplification deep sequencing.

Name	Target sequence	Primer/F	Primer/R
<i>PDS-1_target</i>	AGGGTGCTGGTAGGACATCTGG	GTTCAACCACTGTTTTCATCAGATAG	GATCAATGATCGGTTGCAGTG
<i>PDS-1-1</i>	GCGCCGCTGGTAGGACATCAG A	GTGCACGCTCAATAGAAACAGC	CAATTACGCTAGTGTGCCGAC
<i>PDS-1-2</i>	TTGATGCTGGTAGGACAACAGG	GCGAGTTGTTCTCGAGGAGC	CTCTCACAGTTGGGTGTGCG
<i>PDS-1-3</i>	CTCCTACTGGTTGGACATCTGG	GAGCCCAATGCCACAGGAC	GTGATAATCTTATTGGCTGATTTCG
<i>PDS-1-4</i>	GAGGAGCTGGTAGGACCTTCGG	CGTCACTGGCGCCAGTTC	CATGGCCAAGAAAAATCCAC
<i>PDS-2_target</i>	GGATGGTAATCTCCGGAAG G	GTGCCATCGTCATTGAGCTC	GTGTGTCATAAGCTGCAACAGATG
<i>PDS-2-1</i>	GGATGGTATATCTCTGAAAAGG	CGACGGATACTCAAGTATGGC	GTTTGGGCCGATATTCTTGCTAG
<i>PDS-2-2</i>	TGTTGGATATACTCCGAATGG	GTTAATTATCAATTTTAAGGTCACAC TG	GATTTGGCATTCAATGGAACG
<i>CPC_target</i>	ATCCCGGACGGACGCCGAG G	CTTGCTTGTGAATTAAGGAGAGG	GAATAAAAGGTATTTGGTTAAGTGTAAG
<i>CPC-1</i>	CTTCCGAGACAGACGCTCGGAG G	GACTGTGATGAATCACCGACAC	GACCAGAGTGATCACGCCAAG
<i>CPC-2</i>	ACTTCAGGACGGACGCTGGAG G	CAAGATCGAAGACGCAATCG	CTTCACCAAATAGTCAAAAGCCTG
<i>CPC-3</i>	ATGGAGGAACCAACGCCGAG G	GGAGTATTCTGGTCCGAATCAAG	CGCATATATCTCCAGAACAATCTAAG
<i>CPC-4</i>	AAACGGGAACGAACCCGGAG G	GCTGGGGGAAAGACTAAAAAAG	CATGTTTTTCTTATGCGGGGC
<i>CPC-5</i>	TTCCGGCCACGGACGCCGAGC G	GTTACGACCCGACGATATCG	GAGTCACCGGTGGAGGAGAAG
<i>TRY_target</i>	GAAGTGAGCAGTATCGAATGG G	GATTAGTTTCTAGTTTAAACAAAAAT TAATATC	CTTAGAAAAAGTAGAAGAAGAAGAAATT G
<i>TRY-1</i>	GAAGTGAGTAGCATCGAATGG G	GAATCGGACATGACAGAAGAAGAG	CAATTCATTACTCTACACTCATCCTC
<i>TRY-2</i>	GAGGTGAGTAGTATCGAATGG G	GATGGATCGAGAGAGAAAGAGTCTC	GCTTCTTGTCCGAAGGTCTG
<i>TRY-3</i>	GAAGTGAGCAGTCTTGAGTGGG	CTCCGCTATCTCATGATCTCCATC	GAGAAAACATGAAAAACCTTTTAC
<i>TRY-4</i>	GAAGTGAGTAGTATGAAATGG G	TATATAAACAGAAGAAGAAAAATGT TACG	GTCTACGCTCATTCAATTGACCTG
<i>TRY-5</i>	GAAGTAAGTAGTATCAAAAGG T	GATCACCTAGAAGATAACGGCTATA G	CTGTGGTAAGCTCTACATTCATTGC
<i>TRY-6</i>	GAGATGAAGAGTATCGAATTG G	CTTAATCCATCTTCTCTGATAATCG	GAAGGAGAGTAAAAAGAGACCGTAGAG
<i>TRY-7</i>	GAATACAGCAGTATCAAAGGG T	CGTACTGGCAGATAGAGCAGTTG	CTGATAGGTGCAGCTTTCAAATG

Table S8. Primers used for RT-qPCR amplification.

Gene	Primer/F	Primer/R
ACTIN2	CTGTGCCAATCTACGAGGGTT	CTGTGCCAATCTACGAGGGTT
zCas9	CTACCTGTACTACCTCCAGAAT	GAATGACTGTGGAACGATATGA

All Publications Cited for Section 4.1

1. Amancio, S.; Nakajima, I.; Ban, Y.; Azuma, A.; Onoue, N.; Moriguchi, T.; Yamamoto, T.; Toki, S.; Endo, M. CRISPR/Cas9-mediated targeted mutagenesis in grape. *Plos ONE* **2017**, *12*, e0177966.
2. Andersson, M.; Turesson, H.; Nicolai, A.; Fält, A.S.; Samuelsson, M.; Hofvander, P. Efficient targeted multiallelic mutagenesis in tetraploid potato (*Solanum tuberosum*) by transient CRISPR-Cas9 expression in protoplasts. *Plant Cell Rep.* **2016**, *36*, 117–128.
3. Andersson, M.; Turesson, H.; Olsson, N.; Fält, A.S.; Ohlsson, P.; Gonzalez, M.N.; Samuelsson, M.; Hofvander, P. Genome editing in potato via CRISPR-Cas9 ribonucleoprotein delivery. *Physiol. Plant* **2018**, *164*, 378–384.
4. Bertier, L.D.; Ron, M.; Huo, H.; Bradford, K.J.; Britt, A.B.; Michelmore, R.W. High-Resolution Analysis of the Efficiency, Heritability, and Editing Outcomes of CRISPR/Cas9-Induced Modifications of NCED4 in Lettuce (*Lactuca sativa*). *G3 (Bethesda)* **2018**, *8*, 1513–1521.
5. Brooks, C.; Nekrasov, V.; Lippman, Z.B.; Van Eck, J. Efficient Gene Editing in Tomato in the First Generation Using the Clustered Regularly Interspaced Short Palindromic Repeats/CRISPR-Associated9 System. *Plant Physiol.* **2014**, *166*, 1292–1297.
6. Cai, Y.; Chen, L.; Liu, X.; Guo, C.; Sun, S.; Wu, C.; Jiang, B.; Han, T.; Hou, W. CRISPR/Cas9-mediated targeted mutagenesis of GmFT2a delays flowering time in soya bean. *Plant Biotechnol. J.* **2018**, *16*, 176–185.
7. Cai, Y.; Chen, L.; Liu, X.; Sun, S.; Wu, C.; Jiang, B.; Han, T.; Hou, W. CRISPR/Cas9-Mediated Genome Editing in Soybean Hairy Roots. *Plos ONE* **2015**, *10*, e0136064.
8. Chandrasekaran, J.; Brumin, M.; Wolf, D.; Leibman, D.; Klap, C.; Pearlsman, M.; Sherman, A.; Arazi, T.; Gal-On, A. Development of broad virus resistance in non-transgenic cucumber using CRISPR/Cas9 technology. *Mol. Plant. Pathol.* **2016**, *17*, 1140–1153.
9. Chen, X.; Lu, X.; Shu, N.; Wang, S.; Wang, J.; Wang, D.; Guo, L.; Ye, W. Targeted mutagenesis in cotton (*Gossypium hirsutum* L.) using the CRISPR/Cas9 system. *Sci. Rep.* **2017**, *7*, 44304.
10. Curtin, S.J.; Xiong, Y.; Michno, J.M.; Campbell, B.W.; Stec, A.O.; Cermak, T.; Starker, C.; Voytas, D.F.; Eamens, A.L.; Stupar, R.M. CRISPR/Cas9 and TALENs generate heritable mutations for genes involved in small RNA processing of Glycine max and Medicago truncatula. *Plant Biotechnol. J.* **2018**, *16*, 1125–1137.
11. D'Ambrosio, C.; Stigliani, A.L.; Giorio, G. CRISPR/Cas9 editing of carotenoid genes in tomato. *Transgenic Res.* **2018**, *27*, 367–378.
12. Deng, L.; Wang, H.; Sun, C.; Li, Q.; Jiang, H.; Du, M.; Li, C.B.; Li, C. Efficient generation of pink-fruited tomatoes using CRISPR/Cas9 system. *J. Genet. Genom.* **2018**, *45*, 51–54.
13. Du, H.; Zeng, X.; Zhao, M.; Cui, X.; Wang, Q.; Yang, H.; Cheng, H.; Yu, D. Efficient targeted mutagenesis in soybean by TALENs and CRISPR/Cas9. *J. Biotechnol.* **2016**, *217*, 90–97.
14. Feng, C.; Su, H.; Bai, H.; Wang, R.; Liu, Y.; Guo, X.; Liu, C.; Zhang, J.; Yuan, J.; Birchler, J.A.; et al. High-efficiency genome editing using a dmc1 promoter-controlled CRISPR/Cas9 system in maize. *Plant Biotechnol. J.* **2018**, *16*, 1848–1857.
15. Feng, C.; Yuan, J.; Wang, R.; Liu, Y.; Birchler, J. A.; Han, F. Efficient Targeted Genome Modification in Maize Using CRISPR/Cas9 System. *J. Genet. Genomics* **2015**, *43*, 37–43.
16. Feng, Z.; Zhang, B.; Ding, W.; Liu, X.; Yang, D.L.; Wei, P.; Cao, F.; Zhu, S.; Zhang, F.; Mao, Y.; et al. Efficient genome editing in plants using a CRISPR/Cas system. *Cell Res.* **2013**, *23*, 1229–1232.
17. Gao, R.; Feyissa, B.A.; Croft, M.; Hannoufa, A. Gene editing by CRISPR/Cas9 in the obligatory outcrossing Medicago sativa. *Planta* **2018**, *247*, 1043–1050.
18. Gao, W.; Long, L.; Tian, X.; Xu, F.; Liu, J.; Singh, P.K.; Botella, J.R.; Song, C. Genome Editing in Cotton with the CRISPR/Cas9 System. *Front. Plant Sci.* **2017**, *8*, 1364.
19. Gil-Humanes, J.; Wang, Y.; Liang, Z.; Shan, Q.; Ozuna, C.V.; Sánchez-León, S.; Baltes, N.J.; Starker, C.; Barro, F.; Gao, C.; et al. High-efficiency gene targeting in hexaploid wheat using DNA replicons and CRISPR/Cas9. *Plant J.* **2017**, *89*, 1251–1262.
20. He, Y.; Zhu, M.; Wang, L.; Wu, J.; Wang, Q.; Wang, R.; Zhao, Y. Programmed Self-Elimination of the CRISPR/Cas9 Construct Greatly Accelerates the Isolation of Edited and Transgene-Free Rice Plants. *Mol. Plant* **2018**, *11*, 1210–1213.
21. Ito, Y.; Nishizawa-Yokoi, A.; Endo, M.; Mikami, M.; Toki, S. CRISPR/Cas9-mediated mutagenesis of the RIN locus that regulates tomato fruit ripening. *Biochem. Biophys. Res. Commun.* **2015**, *467*, 76–82.
22. Janga, M.R.; Campbell, L.M.; Rathore, K.S. CRISPR/Cas9-mediated targeted mutagenesis in upland cotton (*Gossypium hirsutum* L.). *Plant Mol. Biol.* **2017**, *94*, 349–360.

23. Jia, H.; Wang, N. Targeted genome editing of sweet orange using Cas9/sgRNA. *Plos ONE* **2014**, *9*, e93806.
24. Jia, H.; Zhang, Y.; Orbović, V.; Xu, J.; White, F.F.; Jones, J.B.; Wang, N. Genome editing of the disease susceptibility gene CsLOB1 in citrus confers resistance to citrus canker. *Plant Biotechnol. J.* **2017**, *15*, 817–823.
25. Li, J.F.; Norville, J.E.; Aach, J.; McCormack, M.; Zhang, D.; Bush, J.; Church, G.M.; Sheen, J. Multiplex and homologous recombination-mediated genome editing in Arabidopsis and Nicotiana benthamiana using guide RNA and Cas9. *Nat. Biotechnol.* **2013**, *31*, 688–691.
26. Jiang, W.; Zhou, H.; Bi, H.; Fromm, M.; Yang, B.; Weeks, D.P. Demonstration of CRISPR/Cas9/sgRNA-mediated targeted gene modification in Arabidopsis, tobacco, sorghum and rice. *Nucleic Acids Res.* **2013**, *41*, e188.
27. Jiang, W.Z.; Henry, I.M.; Lynagh, P.G.; Comai, L.; Cahoon, E.B.; Weeks, D.P. Significant enhancement of fatty acid composition in seeds of the allohexaploid, Camelina sativa, using CRISPR/Cas9 gene editing. *Plant Biotechnol. J.* **2017**, *15*, 648–657.
28. Kapusi, E.; Corcuera-Gomez, M.; Melnik, S.; Stoger, E. Heritable Genomic Fragment Deletions and Small Indels in the Putative ENGase Gene Induced by CRISPR/Cas9 in Barley. *Front. Plant Sci.* **2017**, *8*, 540.
29. Kaur, N.; Alok, A.; Shivani; Kaur, N.; Pandey, P.; Awasthi, P.; Tiwari, S. CRISPR/Cas9-mediated efficient editing in phytoene desaturase (PDS) demonstrates precise manipulation in banana cv. Rasthali genome. *Funct. Integr. Genom.* **2017**, *18*, 89–99.
30. Kim, D.; Alptekin, B.; Budak, H. CRISPR/Cas9 genome editing in wheat. *Funct. Integr. Genom.* **2017**, *18*, 31–41.
31. Klimek-Chodacka, M.; Oleszkiewicz, T.; Lowder, L.G.; Qi, Y.; Baranski, R. Efficient CRISPR/Cas9-based genome editing in carrot cells. *Plant Cell Rep.* **2018**, *37*, 575–586.
32. LeBlanc, C.; Zhang, F.; Mendez, J.; Lozano, Y.; Chatpar, K.; Irish, V.F.; Jacob, Y. Increased efficiency of targeted mutagenesis by CRISPR/Cas9 in plants using heat stress. *Plant J.* **2018**, *93*, 377–386.
33. Lee, K.; Zhang, Y.; Kleinstiver, B.P.; Guo, J.A.; Aryee, M.J.; Miller, J.; Malzahn, A.; Zarecor, S.; Lawrence-Dill, C.J.; Joung, J.K.; et al. Activities and specificities of CRISPR/Cas9 and Cas12a nucleases for targeted mutagenesis in maize. *Plant Biotechnol. J.* **2019**, *17*, 362–372.
34. Li, C.; Chen, C.; Chen, H.; Wang, S.; Chen, X.; Cui, Y. Verification of DNA motifs in Arabidopsis using CRISPR/Cas9-mediated mutagenesis. *Plant Biotechnol. J.* **2018**, *16*, 1446–1451.
35. Li, C.; Hao, M.; Wang, W.; Wang, H.; Chen, F.; Chu, W.; Zhang, B.; Mei, D.; Cheng, H.; Hu, Q. An Efficient CRISPR/Cas9 Platform for Rapidly Generating Simultaneous Mutagenesis of Multiple Gene Homoeologs in Allotetraploid Oilseed Rape. *Front. Plant Sci.* **2018**, *9*, 442.
36. Li, C.; Unver, T.; Zhang, B. A high-efficiency CRISPR/Cas9 system for targeted mutagenesis in Cotton (*Gossypium hirsutum* L.). *Sci. Rep.* **2017**, *7*, 43902.
37. Li, J.; Manghwar, H.; Sun, L.; Wang, P.; Wang, G.; Sheng, H.; Zhang, J.; Liu, H.; Qin, L.; Rui, H.; et al. Whole genome sequencing reveals rare off-target mutations and considerable inherent genetic or/and somaclonal variations in CRISPR. Cas9-edited cotton plants. *Plant Biotechnol. J.* **2018**, *17*, 858–868.
38. Li, J.; Zhang, H.; Si, X.; Tian, Y.; Chen, K.; Liu, J.; Chen, H.; Gao, C. Generation of thermosensitive male-sterile maize by targeted knockout of the ZmTMS5 gene. *J. Genet. Genom.* **2017**, *44*, 465–468.
39. Li, M.; Li, X.; Zhou, Z.; Wu, P.; Fang, M.; Pan, X.; Lin, Q.; Luo, W.; Wu, G.; Li, H. Reassessment of the Four Yield-related Genes Gn1a, DEP1, GS3, and IPA1 in Rice Using a CRISPR/Cas9 System. *Front. Plant Sci.* **2016**, *7*.
40. Li, R.; Fu, D.; Zhu, B.; Luo, Y.; Zhu, H. CRISPR/Cas9-mediated mutagenesis of lncRNA1459 alters tomato fruit ripening. *Plant J.* **2018**, *94*, 513–524.
41. Li, R.; Li, R.; Li, X.; Fu, D.; Zhu, B.; Tian, H.; Luo, Y.; Zhu, H. Multiplexed CRISPR/Cas9-mediated metabolic engineering of γ -aminobutyric acid levels in Solanum lycopersicum. *Plant Biotechnol. J.* **2018**, *16*, 415–427.
42. Li, R.; Zhang, L.; Wang, L.; Chen, L.; Zhao, R.; Sheng, J.; Shen, L. Reduction of Tomato-Plant Chilling Tolerance by CRISPR-Cas9-Mediated SICBF1 Mutagenesis. *J. Agric. Food Chem.* **2018**, *66*, 9042–9051.
43. Li, S.; Songmei, L.; Yanhua, L.; Haiping, L.; Yuanyuan, T.; Jianzhong, H.; Pengcheng, W.; Yao, S.Q. HRM-facilitated rapid identification and genotyping of mutations induced by CRISPR/Cas9 mutagenesis in rice. *Crop Breed. Appl. Biotechnol.* **2018**, *18*, 184–191.
44. Li, Z.; Liu, Z.B.; Xing, A.; Moon, B.P.; Koellhoffer, J.P.; Huang, L.; Ward, R.T.; Clifton, E.; Falco, S.C.; Cigan, A.M. Cas9-Guide RNA Directed Genome Editing in Soybean. *Plant Physiol.* **2015**, *169*, 960–970.
45. Liang, Z.; Chen, K.; Li, T.; Zhang, Y.; Wang, Y.; Zhao, Q.; Liu, J.; Zhang, H.; Liu, C.; Ran, Y.; et al. Efficient DNA-free genome editing of bread wheat using CRISPR/Cas9 ribonucleoprotein complexes. *Nat. Commun.* **2017**, *8*, 14261.

46. Liang, Z.; Zhang, K.; Chen, K.; Gao, C. Targeted mutagenesis in *Zea mays* using TALENs and the CRISPR/Cas system. *J. Genet. Genom.* **2014**, *41*, 63–68.
47. Lin, C.S.; Hsu, C.T.; Yang, L.H.; Lee, L.Y.; Fu, J.Y.; Cheng, Q.W.; Wu, F.H.; Hsiao, H.C.; Zhang, Y.; Zhang, R.; et al. Application of protoplast technology to CRISPR/Cas9 mutagenesis: From single-cell mutation detection to mutant plant regeneration. *Plant Biotechnol. J.* **2018**, *16*, 1295–1310.
48. Liu, Y.; Merrick, P.; Zhang, Z.; Ji, C.; Yang, B.; Fei, S.Z. Targeted mutagenesis in tetraploid switchgrass (*Panicum virgatum* L.) using CRISPR/Cas9. *Plant Biotechnol. J.* **2018**, *16*, 381–393.
49. Lowder, L.G.; Zhang, D.; Baltes, N.J.; Paul, J.W. 3rd; Tang, X.; Zheng, X.; Voytas, D.F.; Hsieh, T.F.; Zhang, Y.; Qi, Y. A CRISPR/Cas9 Toolbox for Multiplexed Plant Genome Editing and Transcriptional Regulation. *Plant Physiol.* **2015**, *169*, 971–985.
50. Ma, X.; Zhang, Q.; Zhu, Q.; Liu, W.; Chen, Y.; Qiu, R.; Wang, B.; Yang, Z.; Li, H.; Lin, Y.; et al. A Robust CRISPR/Cas9 System for Convenient, High-Efficiency Multiplex Genome Editing in Monocot and Dicot Plants. *Mol. Plant* **2015**, *8*, 1274–1284.
51. Macovei, A.; Sevilla, N.R.; Cantos, C.; Jonson, G.B.; Slamet-Loedin, I.; Cermak, T.; Voytas, D.F.; Choi, I.R.; Chadha-Mohanty, P. Novel alleles of rice eIF4G generated by CRISPR/Cas9-targeted mutagenesis confer resistance to Rice tungro spherical virus. *Plant Biotechnol. J.* **2018**, *16*, 1918–1927.
52. Meng, Y.; Hou, Y.; Wang, H.; Ji, R.; Liu, B.; Wen, J.; Niu, L.; Lin, H. Targeted mutagenesis by CRISPR/Cas9 system in the model legume *Medicago truncatula*. *Plant. Cell Rep.* **2016**, *36*, 371–374.
53. Nakayasu, M.; Akiyama, R.; Lee, H.J.; Osakabe, K.; Osakabe, Y.; Watanabe, B.; Sugimoto, Y.; Umemoto, N.; Saito, K.; Muranaka, T.; et al. Generation of alpha-solanine-free hairy roots of potato by CRISPR/Cas9 mediated genome editing of the St16DOX gene. *Plant Physiol. Biochem.* **2018**, *131*, 70–77.
54. Nieves-Cordones, M.; Mohamed, S.; Tanoi, K.; Kobayashi, N.I.; Takagi, K.; Vernet, A.; Guiderdoni, E.; Perin, C.; Sentenac, H.; Very, A.A. Production of low-Cs(+) rice plants by inactivation of the K(+) transporter OsHAK1 with the CRISPR-Cas system. *Plant J.* **2017**, *92*, 43–56.
55. Nishitani, C.; Hirai, N.; Komori, S.; Wada, M.; Okada, K.; Osakabe, K.; Yamamoto, T.; Osakabe, Y. Efficient Genome Editing in Apple Using a CRISPR/Cas9 system. *Sci. Rep.* **2016**, *6*, 31481.
56. Odipio, J.; Alicai, T.; Ingelbrecht, I.; Nusinow, D.A.; Bart, R.; Taylor, N.J. Efficient CRISPR/Cas9 Genome Editing of Phytoene desaturase in Cassava. *Front. Plant Sci.* **2017**, *8*, 1780.
57. Osakabe, Y.; Watanabe, T.; Sugano, S.S.; Ueta, R.; Ishihara, R.; Shinozaki, K.; Osakabe, K. Optimization of CRISPR/Cas9 genome editing to modify abiotic stress responses in plants. *Sci. Rep.* **2016**, *6*, 26685.
58. Pan, C.; Ye, L.; Qin, L.; Liu, X.; He, Y.; Wang, J.; Chen, L.; Lu, G. CRISPR/Cas9-mediated efficient and heritable targeted mutagenesis in tomato plants in the first and later generations. *Sci. Rep.* **2016**, *6*, 24765.
59. Ren, C.; Liu, X.; Zhang, Z.; Wang, Y.; Duan, W.; Li, S.; Liang, Z. CRISPR/Cas9-mediated efficient targeted mutagenesis in Chardonnay (*Vitis vinifera* L.). *Sci. Rep.* **2016**, *6*, 32289.
60. Ron, M.; Kajala, K.; Pauluzzi, G.; Wang, D.; Reynoso, M.A.; Zumstein, K.; Garcha, J.; Winte, S.; Masson, H.; Inagaki, S.; et al. Hairy Root Transformation Using *Agrobacterium rhizogenes* as a Tool for Exploring Cell Type-Specific Gene Expression and Function Using Tomato as a Model. *Plant. Physiol.* **2014**, *166*, 455–469.
61. Sanchez-Leon, S.; Gil-Humanes, J.; Ozuna, C.V.; Gimenez, M.J.; Sousa, C.; Voytas, D.F.; Barro, F. Low-gluten, nontransgenic wheat engineered with CRISPR/Cas9. *Plant Biotechnol. J.* **2018**, *16*, 902–910.
62. Shan, Q.; Wang, Y.; Li, J.; Zhang, Y.; Chen, K.; Liang, Z.; Zhang, K.; Liu, J.; Xi, J.J.; Qiu, J.L. Targeted genome modification of crop plants using a CRISPR-Cas system. *Nat. Biotechnol.* **2013**, *31*, 686–688.
63. Shen, C.; Que, Z.; Xia, Y.; Tang, N.; Li, D.; He, R.; Cao, M. Knock out of the annexin gene OsAnn3 via CRISPR/Cas9-mediated genome editing decreased cold tolerance in rice. *J. Plant. Biol.* **2017**, *60*, 539–547.
64. Shen, L.; Hua, Y.; Fu, Y.; Li, J.; Liu, Q.; Jiao, X.; Xin, G.; Wang, J.; Wang, X.; Yan, C.; et al. Rapid generation of genetic diversity by multiplex CRISPR/Cas9 genome editing in rice. *Sci. China Life Sci.* **2017**, *60*, 506–515.
65. Shen, L.; Wang, C.; Fu, Y.; Wang, J.; Liu, Q.; Zhang, X.; Yan, C.; Qian, Q.; Wang, K. QTL editing confers opposing yield performance in different rice varieties. *J. Integr. Plant Biol.* **2018**, *60*, 89–93.
66. Singh, M.; Kumar, M.; Albertsen, M.C.; Young, J.K.; Cigan, A.M. Concurrent modifications in the three homeologs of Ms45 gene with CRISPR-Cas9 lead to rapid generation of male sterile bread wheat (*Triticum aestivum* L.). *Plant Mol. Biol.* **2018**, *97*, 371–383.
67. Soyk, S.; Lemmon, Z.H.; Oved, M.; Fisher, J.; Liberatore, K.L.; Park, S.J.; Goren, A.; Jiang, K.; Ramos, A.; van der Knaap, E.; et al. Bypassing Negative Epistasis on Yield in Tomato Imposed by a Domestication Gene. *Cell* **2017**, *169*, 1142–1155.
68. Soyk, S.; Muller, N.A.; Park, S.J.; Schmalenbach, I.; Jiang, K.; Hayama, R.; Zhang, L.; Van Eck, J.; Jimenez-Gomez, J.M.; Lippman, Z.B. Variation in the flowering gene SELF PRUNING 5G promotes day-neutrality

- and early yield in tomato. *Nat. Genet.* **2017**, *49*, 162–168.
69. Sun, Q.; Lin, L.; Liu, D.; Wu, D.; Fang, Y.; Wu, J.; Wang, Y. CRISPR/Cas9-Mediated Multiplex Genome Editing of the BnWRKY11 and BnWRKY70 Genes in Brassica napus L. *Int J. Mol. Sci.* **2018**, *19*, E2716.
 70. Sun, Y.; Jiao, G.; Liu, Z.; Zhang, X.; Li, J.; Guo, X.; Du, W.; Du, J.; Francis, F.; Zhao, Y.; et al. Generation of High-Amylose Rice through CRISPR/Cas9-Mediated Targeted Mutagenesis of Starch Branching Enzymes. *Front. Plant Sci.* **2017**, *8*, 298.
 71. Tang, F.; Yang, S.; Liu, J.; Zhu, H. Rj4, a Gene Controlling Nodulation Specificity in Soybeans, Encodes a Thaumatin-Like Protein But Not the One Previously Reported. *Plant Physiol.* **2016**, *170*, 26–32.
 72. Tang, L.; Mao, B.; Li, Y.; Lv, Q.; Zhang, L.; Chen, C.; He, H.; Wang, W.; Zeng, X.; Shao, Y.; et al. Knockout of OsNramp5 using the CRISPR/Cas9 system produces low Cd-accumulating indica rice without compromising yield. *Sci. Rep.* **2017**, *7*, 14438.
 73. Tang, X.; Zheng, X.; Qi, Y.; Zhang, D.; Cheng, Y.; Tang, A.; Voytas, D.F.; Zhang, Y. A Single Transcript CRISPR-Cas9 System for Efficient Genome Editing in Plants. *Mol. Plant* **2016**, *9*, 1088–1091.
 74. Tashkandi, M.; Ali, Z.; Aljedaani, F.; Shami, A.; Mahfouz, M.M. Engineering resistance against Tomato yellow leaf curl virus via the CRISPR/Cas9 system in tomato. *Plant Signal. Behav.* **2018**, *13*, e1525996.
 75. Tian, S.; Jiang, L.; Gao, Q.; Zhang, J.; Zong, M.; Zhang, H.; Ren, Y.; Guo, S.; Gong, G.; Liu, F.; et al. Efficient CRISPR/Cas9-based gene knockout in watermelon. *Plant Cell Rep.* **2017**, *36*, 399–406.
 76. Ueta, R.; Abe, C.; Watanabe, T.; Sugano, S.S.; Ishihara, R.; Ezura, H.; Osakabe, Y.; Osakabe, K. Rapid breeding of parthenocarpic tomato plants using CRISPR/Cas9. *Sci. Rep.* **2017**, *7*, 507.
 77. Wang, D.; Samsulrizal, N.H.; Yan, C.; Allcock, N.S.; Craigon, J.; Blanco-Ulate, B.; Ortega-Salazar, I.; Marcus, S.E.; Bagheri, H.M.; Perez Fons, L.; et al. Characterization of CRISPR Mutants Targeting Genes Modulating Pectin Degradation in Ripening Tomato. *Plant Physiol.* **2019**, *179*, 544–557.
 78. Wang, F.; Wang, C.; Liu, P.; Lei, C.; Hao, W.; Gao, Y.; Liu, Y.G.; Zhao, K. Enhanced Rice Blast Resistance by CRISPR/Cas9-Targeted Mutagenesis of the ERF Transcription Factor Gene OsERF922. *Plos ONE* **2016**, *11*, e0154027.
 79. Wang, L.; Chen, L.; Li, R.; Zhao, R.; Yang, M.; Sheng, J.; Shen, L. Reduced Drought Tolerance by CRISPR/Cas9-Mediated SIMAPK3 Mutagenesis in Tomato Plants. *J. Agric. Food Chem.* **2017**, *65*, 8674–8682.
 80. Wang, L.; Wang, L.; Tan, Q.; Fan, Q.; Zhu, H.; Hong, Z.; Zhang, Z.; Duanmu, D. Efficient Inactivation of Symbiotic Nitrogen Fixation Related Genes in Lotus japonicus Using CRISPR-Cas9. *Front. Plant Sci.* **2016**, *7*.
 81. Wang, P.; Zhang, J.; Sun, L.; Ma, Y.; Xu, J.; Liang, S.; Deng, J.; Tan, J.; Zhang, Q.; Tu, L.; et al. High efficient multisites genome editing in allotetraploid cotton (*Gossypium hirsutum*) using CRISPR/Cas9 system. *Plant Biotechnol. J.* **2018**, *16*, 137–150.
 82. Wang, W.; Pan, Q.; He, F.; Akhunova, A.; Chao, S.; Trick, H.; Akhunov, E. Transgenerational CRISPR-Cas9 Activity Facilitates Multiplex Gene Editing in Allopolyploid Wheat. *Cris. J.* **2018**, *1*, 65–74.
 83. Wang, X.; Tu, M.; Wang, D.; Liu, J.; Li, Y.; Li, Z.; Wang, Y.; Wang, X. CRISPR/Cas9-mediated efficient targeted mutagenesis in grape in the first generation. *Plant Biotechnol. J.* **2018**, *16*, 844–855.
 84. Wang, Y.; Cheng, X.; Shan, Q.; Zhang, Y.; Liu, J.; Gao, C.; Qiu, J.L. Simultaneous editing of three homoeoalleles in hexaploid bread wheat confers heritable resistance to powdery mildew. *Nat. Biotechnol.* **2014**, *32*, 947–951.
 85. Wang, Z.; Wang, S.; Li, D.; Zhang, Q.; Li, L.; Zhong, C.; Liu, Y.; Huang, H. Optimized paired-sgRNA/Cas9 cloning and expression cassette triggers high-efficiency multiplex genome editing in kiwifruit. *Plant Biotechnol. J.* **2018**, *16*, 1424–1433.
 86. Woo, J.W.; Kim, J.; Kwon, S.I.; Corvalan, C.; Cho, S.W.; Kim, H.; Kim, S.G.; Kim, S.T.; Choe, S.; Kim, J.S. DNA-free genome editing in plants with preassembled CRISPR-Cas9 ribonucleoproteins. *Nat. Biotechnol.* **2015**, *33*, 1162–1164.
 87. Xie, K.; Yang, Y. RNA-guided genome editing in plants using a CRISPR-Cas system. *Mol. Plant* **2013**, *6*, 1975–1983.
 88. Yang, Y.; Zhu, G.; Li, R.; Yan, S.; Fu, D.; Zhu, B.; Tian, H.; Luo, Y.; Zhu, H. The RNA Editing Factor SlORRM4 Is Required for Normal Fruit Ripening in Tomato. *Plant Physiol.* **2017**, *175*, 1690–1702.
 89. Yin, X.; Biswal, A.K.; Dionora, J.; Perdigon, K.M.; Balahadia, C.P.; Mazumdar, S.; Chater, C.; Lin, H.C.; Coe, R.A.; Kretschmar, T.; et al. CRISPR-Cas9 and CRISPR-Cpf1 mediated targeting of a stomatal developmental gene EPFL9 in rice. *Plant Cell Rep.* **2017**, *36*, 745–757.
 90. Yu, Q.H.; Wang, B.; Li, N.; Tang, Y.; Yang, S.; Yang, T.; Xu, J.; Guo, C.; Yan, P.; Wang, Q.; et al. CRISPR/Cas9-induced Targeted Mutagenesis and Gene Replacement to Generate Long-shelf Life Tomato Lines. *Sci. Rep.*

2017, 7, 11874.

91. Zhang, F.; LeBlanc, C.; Irish, V.F.; Jacob, Y. Rapid and efficient CRISPR/Cas9 gene editing in Citrus using the YAO promoter. *Plant Cell Rep.* **2017**, 36, 1883–1887.
92. Zhang, J.; Zhang, H.; Botella, J.R.; Zhu, J.K. Generation of new glutinous rice by CRISPR/Cas9-targeted mutagenesis of the Waxy gene in elite rice varieties. *J. Integr. Plant Biol.* **2018**, 60, 369–375.
93. Zhang, Y.; Li, D.; Zhang, D.; Zhao, X.; Cao, X.; Dong, L.; Liu, J.; Chen, K.; Zhang, H.; Gao, C.; et al. Analysis of the functions of TaGW2 homoeologs in wheat grain weight and protein content traits. *Plant J.* **2018**, 94, 857–866.
94. Zhang, Y.; Liang, Z.; Zong, Y.; Wang, Y.; Liu, J.; Chen, K.; Qiu, J.-L.; Gao, C. Efficient and transgene-free genome editing in wheat through transient expression of CRISPR/Cas9 DNA or RNA. *Nat. Commun.* **2016**, 7, 12617.
95. Zhang, Z.; Ge, X.; Luo, X.; Wang, P.; Fan, Q.; Hu, G.; Xiao, J.; Li, F.; Wu, J. Simultaneous Editing of Two Copies of Gh14-3-3d Confers Enhanced Transgene-Clean Plant Defense Against *Verticillium dahliae* in Allotetraploid Upland Cotton. *Front. Plant Sci.* **2018**, 9, doi: 10.3389/fpls.2018.00842.