



Review

# p63 at the Crossroads between Stemness and Metastasis in Breast Cancer

Veronica Gatti <sup>1,†</sup>, Lucilla Bongiorno-Borbone <sup>1,†</sup>, Claudia Fierro <sup>1</sup>,  
Margherita Annicchiarico-Petruzzelli <sup>2</sup>, Gerry Melino <sup>1,3</sup> and Angelo Peschiaroli <sup>4,\*</sup>

<sup>1</sup> Department of Experimental Medicine, TOR, University of Rome Tor Vergata, 00133 Rome, Italy; veronica\_gatti@yahoo.it (V.G.); lucilla.bongiorno@uniroma2.it (L.B.-B.); claudia\_fierro@libero.it (C.F.); melino@uniroma2.it (G.M.)

<sup>2</sup> Istituto Dermopatico dell'Immacolata, IDI-IRCCS, 00163 Rome, Italy; m.annicchiaricopetruzzelli@idi.it

<sup>3</sup> Medical Research Council, Toxicology Unit, University of Cambridge, Cambridge CB2 1PZ, UK

<sup>4</sup> National Research Council of Italy, Institute of Translational Pharmacology, 00133 Rome, Italy

\* Correspondence: angelo.peschiaroli@cnr.it

† These authors contributed equally to this work.

Received: 13 May 2019; Accepted: 29 May 2019; Published: 31 May 2019



**Abstract:** After lung cancer, breast cancer (BC) is the most frequent cause of cancer death among women, worldwide. Although advances in screening approaches and targeted therapeutic agents have decreased BC incidence and mortality, over the past five years, triple-negative breast cancer (TNBC) remains the breast cancer subtype that displays the worst prognosis, mainly due to the lack of clinically actionable targets. Genetic and molecular profiling has unveiled the high intrinsic heterogeneity of TNBC, with the basal-like molecular subtypes representing the most diffuse TNBC subtypes, characterized by the expression of basal epithelial markers, such as the transcription factor p63. In this review, we will provide a broad picture on the physiological role of p63, in maintaining the basal epithelial identity, as well as its involvement in breast cancer progression, emphasizing its relevance in tumor cell invasion and stemness.

**Keywords:** p53 family; epithelial tumors; TNBC; metastasis; cancer stem cell

## 1. Introduction

Although promising targeted therapeutic agents have been developed over the past two decades, breast cancers (BCs) remain the most frequent cause of cancer death among women worldwide, after lung cancer [1,2]. Based on the histological and molecular features, BCs are molecularly defined into four different subgroups—Luminal A and B, HER2 positive, and triple-negative breast cancers (TNBC). Luminal BCs represent the vast majority (60–80%) of BC cases in developing countries, and they are further classified as Luminal A and Luminal B [3–5].

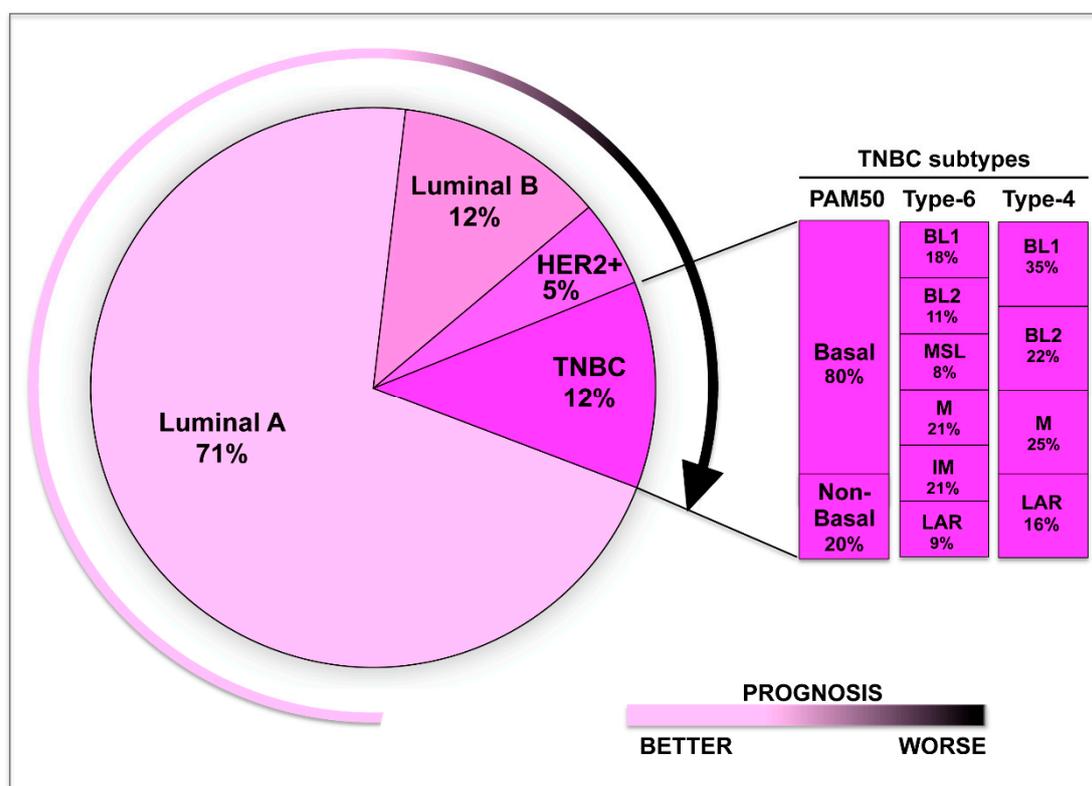
Luminal A are characterized by the expression of estrogen receptors (ER) and progesterone receptors (PR) and generally have the best prognosis among different BC types, responding well to the ER antagonist tamoxifen and aromatase inhibitors [6,7]. Luminal B tumors are ER-positive but PR-negative, and can be either positive or negative for the expression of human epidermal growth factor receptor 2 (HER2/ERBB2). In contrast to Luminal A, Luminal B tumors are more aggressive and do not respond well to endocrine treatment [8].

HER2-positive tumors display an overexpression or amplification of the HER2 oncogene and can be clinically treated with the anti-HER2 monoclonal antibody trastuzumab, in combination with conventional chemotherapy, improving the survival rate by 30% [9–12].

TNBC are generally defined on the basis of the lack of expression of hormone and the HER2 receptors [13]. The absence of these clinically actionable targets implies that TNBC therapy relies

merely on standard approaches, such as radiotherapy, and a combination of chemotherapeutic agents like anthracyclines, alkylating agents, taxanes, or platinum salts [13]. In addition, genetic and molecular profiling has unveiled the high heterogeneity of TNBC, a feature that likely limits the clinical management of these tumors and, as a consequence, TNBC prognosis [5,14–16]. TNBC is indeed the breast cancer subtype displaying the worst prognosis, and TNBC patients display low rates of disease-free survival, overall survival, and five-year survival.

Over the past years, genomic, transcriptomic, proteomic, and epigenomic profiling have allowed to classify TNBC into distinct clinical and molecular subtypes [15]. Lehmann and colleagues have subdivided TNBC into six subtypes—basal-like 1 (BL1), basal-like 2 (BL2), immune modulatory (IM), mesenchymal (M), mesenchymal stem-like (MSL), and luminal androgen receptor (LAR) [16] (see Figure 1). Subsequent studies of the same group have refined this classification on the basis of the contribution of the stromal and immune cells, to the gene expression profiles of the IM and MSL subtypes, respectively [17]. Therefore, TNBC might be simply categorized into four subtypes—BL1, BL2, mesenchymal, and LAR (Figure 1).



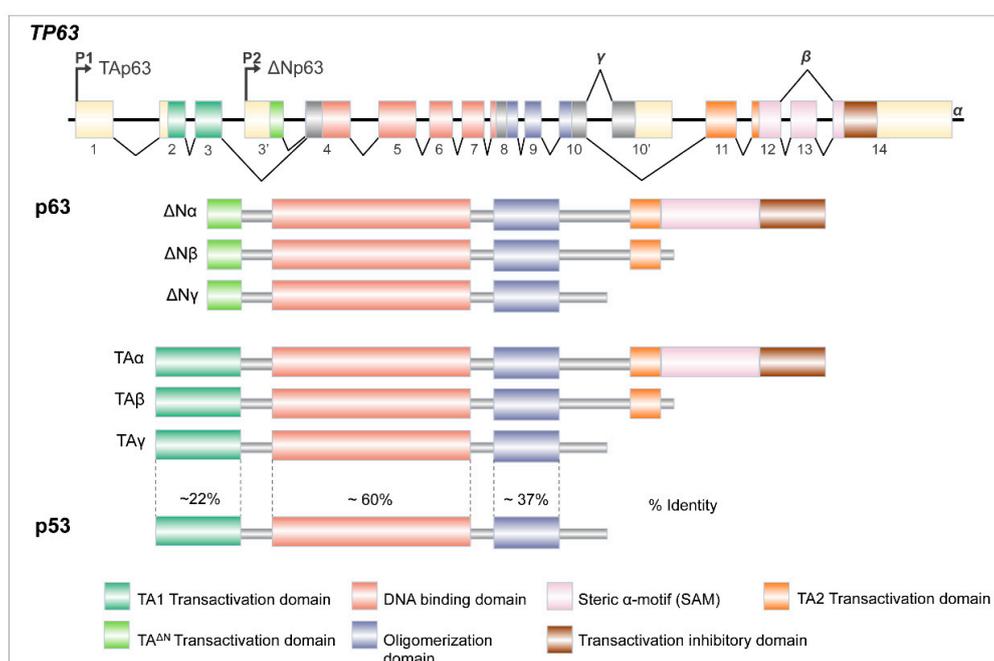
**Figure 1.** Schematic model showing the molecular heterogeneity of triple-negative breast cancer (TNBC). Based on the expression profile, TNBC can be classified into distinct subtypes—Basal-like 1 (BL1), basal-like 2 (BL2), immune modulatory (IM), mesenchymal (M), mesenchymal stem-like (MSL), and luminal androgen receptor (LAR). The percentage of each subtype is also indicated.

Each TNBC subtype displays distinct clinical features, response to neoadjuvant chemotherapy, and relapse-free survival rates, likely reflecting specific genetic and molecular features associated with each subgroup. Basal-like BCs are characterized by the expression of basal epithelial markers, including the transcription factor p63. The *TP63* gene belongs to the p53 family of transcription factors, which includes p63 itself, p73, and the canonical tumor suppressor gene p53 [18]. The activity of members of this family is intimately linked with the pathogenesis of different epithelial tumors, including BC. p53 is considered the guardian of the genome, since it preserves the genome integrity by inducing cell cycle arrest, apoptosis DNA repair, or metabolic adaptation, in response to different stresses [19–22]. In TNBC, *TP53* mutation is frequently observed with a mutational rate of 80% of cases, and is associated

with a more aggressive phenotype [23,24]. Besides losing their tumor suppressor functions, some p53 mutations, the so-called Gain-Of-Function (GOF) mutations, acquire oncogenic functions promoting metastatic dissemination and drug resistance [25–29]. Although the *TP63* gene is rarely mutated in TNBC, several studies have clearly demonstrated the relevance of p63 activity in controlling several tumor-associated events, especially in basal-like subgroups. In the next paragraphs, we will portray a general view of the physiological role of p63, in regulating mammary gland homeostasis, as well as the p63-activated pathways involved in breast cancer progression.

## 2. *Tp63* Gene Architecture

p63 is the most ancient member of the p53 family of transcription factors, which also includes p73 [18,30]. Human *TP63* gene is located on chromosome 3q27–29, which consists of 15 exons spread over 220 kilobases, and can generate multiple isoforms, thanks to different promoter usage and alternative splicing at the C-terminus (Figure 2). p63 protein functions as tetramer and each monomer is arranged in structurally and functionally defined protein domains, closely related to p53's well-known homolog p53. All p63 isoforms encode a core DNA-binding domain (DBD), which is about 60% identical at the DBD of p53 and, as a consequence, can bind to the canonical p53 response element, as well as p63-specific DNA-binding sites. Next to the DBD and common to all p63 isoforms, there is an oligomerization domain (OD) that shares approximately 37% of identity with p53. This allows homo- and hetero-oligomerization between different isoforms of p63 itself and other p53-family members [30–32].



**Figure 2.** The *TP63* gene architecture. *TP63* gene encodes several proteins, thanks to two distinct promoters (P1 and P2) and differential splicing events at the 3' end of its RNA (see text for details).

p63 can be transcribed, starting from two distinct promoters, one (P1) arranged immediately before the first exon and the other (P2) located in the third intron. The choice between P1 and P2 yields two different proteins that might include (TA) or not ( $\Delta N$ ), the N-terminal p53-homologous TransActivation domain (TA1). In addition, the 3' end of both TA and  $\Delta N$  transcripts can undergo alternative splicing events, generating three different C-terminal isoforms ( $\alpha$ ,  $\beta$ , and  $\gamma$ ). The full-length  $\alpha$  isoforms contain two distinctive domains: a putative protein–protein interaction domain found in other transcription factors and signaling proteins called Sterile alpha-motif (SAM) and a transcription inhibition domain (TI). Importantly, the TI domain promotes a closed conformation, through the binding with TA1,

restraining the transcriptional activity of the TAp63 $\alpha$  isoform in the physiological condition [33,34]. Conversely, upon genotoxic stress, specific post-translational modifications release the T1–TA binding, activating TAp63 $\alpha$  transcriptional activity. The physiological relevance of this regulation has been clearly demonstrated in the oocytes, where DNA damage induces TAp63 $\alpha$  transcriptional activity toward pro-apoptotic genes, thus preserving, the female germline [35,36].

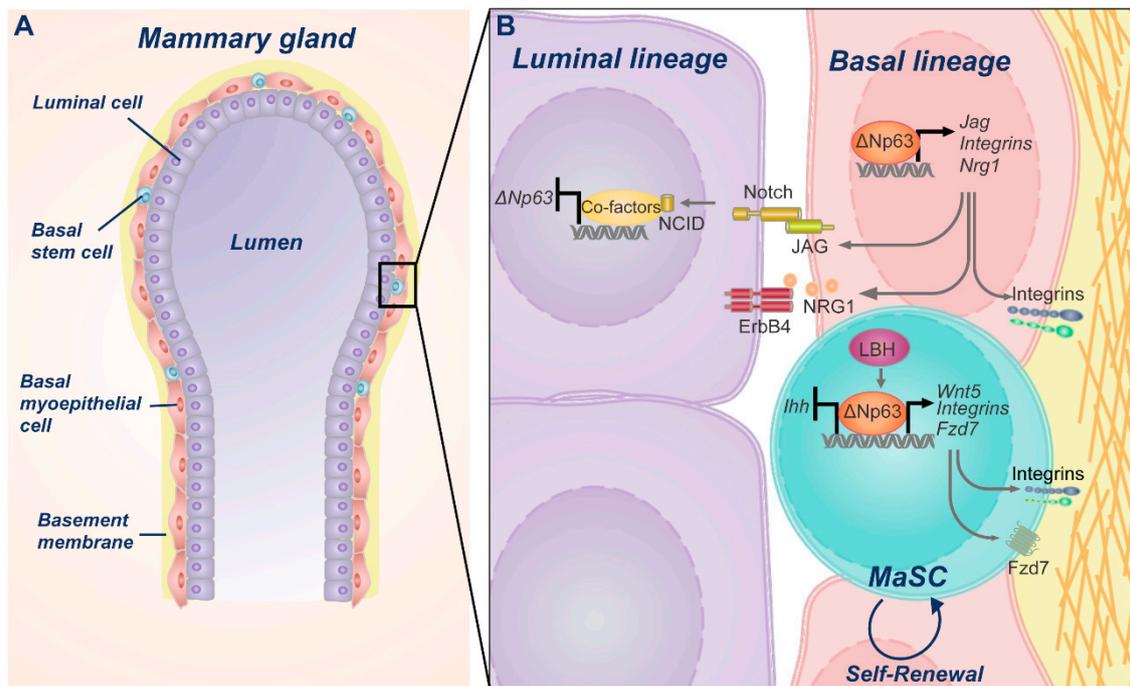
Due to the absence of a canonical TA1 domain, the  $\Delta$ Np63 isoforms have been initially described as dominant negative factors counteracting the TAp63 and p53 functions [32]. However, subsequent studies have clearly demonstrated that, in addition to this function,  $\Delta$ Np63 can act as a sequence-specific transcriptional activator or repressor. In detail, the  $\Delta$ Np63 $\alpha$  isoform can transactivate specific target genes, due to a second transcription activation domain (TA2), arranged within exons 11 and 12, and a residual transactivation activity in the N-terminal truncated region (TA $\Delta$ N) [37–40]. In addition,  $\Delta$ Np63 acts as a potent transcriptional repressor interacting with diverse epigenetic factors, such as histone deacetylases HDAC1 and HDAC2, the chromatin remodeling protein complex SRCAP, the SWI/SNF subunit ACTL6A, and the histone lysine methyl transferase SETDB1, which have all been implicated in the  $\Delta$ Np63-mediated transcriptional repression [41–44].

Multiple studies have analyzed the role of p63 isoforms in regulating physiological and pathological processes, such as tumor development. The general view emerging from these studies is that TAp63 acts as a tumor suppressor, while  $\Delta$ N isoforms promote cancer cell survival and proliferation. In the next paragraphs, we will describe the physiological function of p63 isoforms in maintaining mammary gland homeostasis, and decipher the p63-related pathways involved in breast tumor initiation and progression. If not specified, we used TAp63 and  $\Delta$ Np63 to indicate the  $\alpha$  isoforms, which are the most expressed p63 isoforms in epithelial tissues.

### 3. Physiological Role of p63 during Mammary Gland Development

Mammary gland consists of a bilayered epithelium, made of two distinct cell compartments, luminal and basal. The luminal compartment consists of a single layer of apically oriented, polarized epithelium surrounding the lumen of the gland ducts, which includes structural ductal cells and milk-producing alveolar cells. The basal compartment includes rare stem cells and myoepithelial cells that are arranged in a network around luminal cells and have a contractile function in milk ejection (see Figure 3A) [45,46]. Mammary gland is a plastic organ that undergoes several important changes during the lifetime. It reaches full development during puberty, goes through massive side branching of the ducts during pregnancy and undergoes involution, upon breastfeeding suspension. The impressive regenerative potential of the mammary gland epithelium implies the existence of a mammary epithelial cell population endowed with self-renewing properties. Many studies have indeed proven that mammary stem cells permanently residing within the basal compartment of the mammary gland can generate lineage-committed progenitors (luminal and myoepithelial progenitors), which in turn differentiate into myoepithelial, ductal, or alveolar cells [47–49].

Several lines of evidence have clearly indicated that p63 activity is critically involved in sustaining the proliferative potential and self-renewing capacity of mammary epithelial stem cells (see Figure 3B) [50–53]. The initial observation impinging on a fundamental role of p63, in mammary epithelial homeostasis, arises from the analysis of mice that are genetically devoid of all p63 isoforms. *TP63* null mice display a lack of all squamous epithelia and their derivatives, including mammary glands [52,54]. These defects have been ascribed, at least in part, to a loss of the proliferative potential of the epithelial stem cells and suggest a role for p63 in stemness maintenance [50,52,53]. Remarkably, the genetic selective deletion of  $\Delta$ Np63 isoforms recapitulates the mammary gland defects observed in the p63 global knock-out mice, strongly indicating that  $\Delta$ Np63 is the major p63 isoform governing mammary glands morphogenesis [50,55]. This notion has also been confirmed by additional data, showing that  $\Delta$ Np63 is the major p63 isoform expressed in the epithelial stem cells, where it drives organ reconstitution and ductal branching potential, after mammary epithelial cells transplantation into a cleared fat pad [50].



**Figure 3.** Schematic model of  $\Delta$ Np63 physiological role in mammary gland. (A) Representation of mature mammary gland duct showing an inner layer of luminal cells surrounded by a network of basal myoepithelial cells and scattered stem cells. (B) Magnification of the inter- and intra-cellular signaling pathways orchestrated by  $\Delta$ Np63, aimed at preserving the stem cell self-renewal potential, as well as determining the luminal or basal fate.

A relatively recent study suggests that TAp63 might also participate in controlling mammary stem cell activity. Indeed, it has been shown that mammary glands deficient for TAp63 have an increased regenerative potential and display an enrichment of mammary epithelial cells harboring stem-like properties [56]. These data suggest that TAp63 activity might restrain the self-renewing capacity of mammary stem cells, and imply that the p63 isoforms expression (TA and  $\Delta$ N) needs to be finely regulated in distinct cell compartments. Accordingly, it has been shown that hedgehog (Hh) signaling controls the beginning and the progression of the mammary regenerative cycle by differentially inducing the expression of  $\Delta$ Np63 and TAp63 isoforms in stem and progenitor cells, respectively [57]. The segregation of p63 isoforms is responsible for the TAp63-dependent induction of the Indian hedgehog (Ihh) in progenitor cells and Ihh repression by  $\Delta$ Np63 in stem cells. This circuit ensures a tightly regulated balance between a quiescent state maintenance and progenitors maturation.

Although these data suggest that TAp63 isoforms might also contribute to regulate the self-renewing capacity of mammary cells, diverse studies have clearly highlighted the relevance of  $\Delta$ Np63, in controlling molecular pathways involved in the regenerative potential of mammary epithelium, such as the WNT- $\beta$ /catenin pathway (see Figure 3B).  $\Delta$ Np63 activity intersects WNT signaling at multiple levels.  $\Delta$ Np63 and WNT pathway cooperates in the promotion of stemness, through a direct regulation of the FZD7 receptor and WNT5B ligand expression by  $\Delta$ Np63 [50]. The WNT/ $\beta$ -catenin target gene *LBH* (Limb, Bud and Hearth) is able to drive the expression of  $\Delta$ Np63 and to repress that of TAp63. *LBH*'s capability for maintaining a basal mammary stem cell state and to repress luminal differentiation, requires  $\Delta$ Np63 expression, suggesting that  $\Delta$ Np63 activity is pivotal to control cell fate determination and maintain the basal cell phenotype [58]. Accordingly,  $\Delta$ Np63 knockdown in primary human breast epithelial cells, with basal characteristics, induces a decrease of the basal markers, such as cytokeratin 14 (CK14) and integrin- $\alpha$ 6 (ITGA6) together with the appearance of luminal markers [51]. Reciprocally, ectopic expression of  $\Delta$ Np63 in isolated human luminal cells or in adult murine luminal cells, in vivo, is sufficient to induce a transition towards a basal cell fate [51,59].

The importance of  $\Delta$ Np63 in sustaining epithelial basal identity is also supported by the finding that the Bromodomain-containing protein 4 (BRD4) maintains a basal epithelial phenotype by regulating the expression of epithelial-specific genes, including *TP63* [60].

Furthermore, *BRCA1*, a DNA repair gene involved in mammary gland differentiation and implicated in the pathogenesis of basal-like breast cancer [61,62], is able to drive the expression of  $\Delta$ Np63 in basal cells and sustain NOTCH signaling in luminal cells [63]. Importantly, NOTCH signaling is fundamental to commit progenitor epithelial cells towards a luminal phenotype [51]. Mouse mammary epithelial cells bearing inactive NOTCH signaling, showed an impaired luminal lineage differentiation. Notably, NOTCH activity in determining luminal fate specification requires the NOTCH-dependent inhibition of  $\Delta$ Np63 expression.  $\Delta$ Np63 expression inhibits luminal cell differentiation and prevents overall reconstruction in mammary gland transplantation experiments. NOTCH- $\Delta$ Np63 crosstalk is further complicated by the evidence that  $\Delta$ Np63 promotes mammary stem cell quiescence, by increasing the expression and activity of NOTCH 3 [64], suggesting that specific regulation of different NOTCH receptors might regulate the fine balance between quiescence and commitment. Collectively, these data indicate that *BRCA1*,  $\Delta$ Np63, and NOTCH signaling takes part in a highly regulated crosstalk, ensuring the proper balance between stemness, cell fate specification, and differentiation.

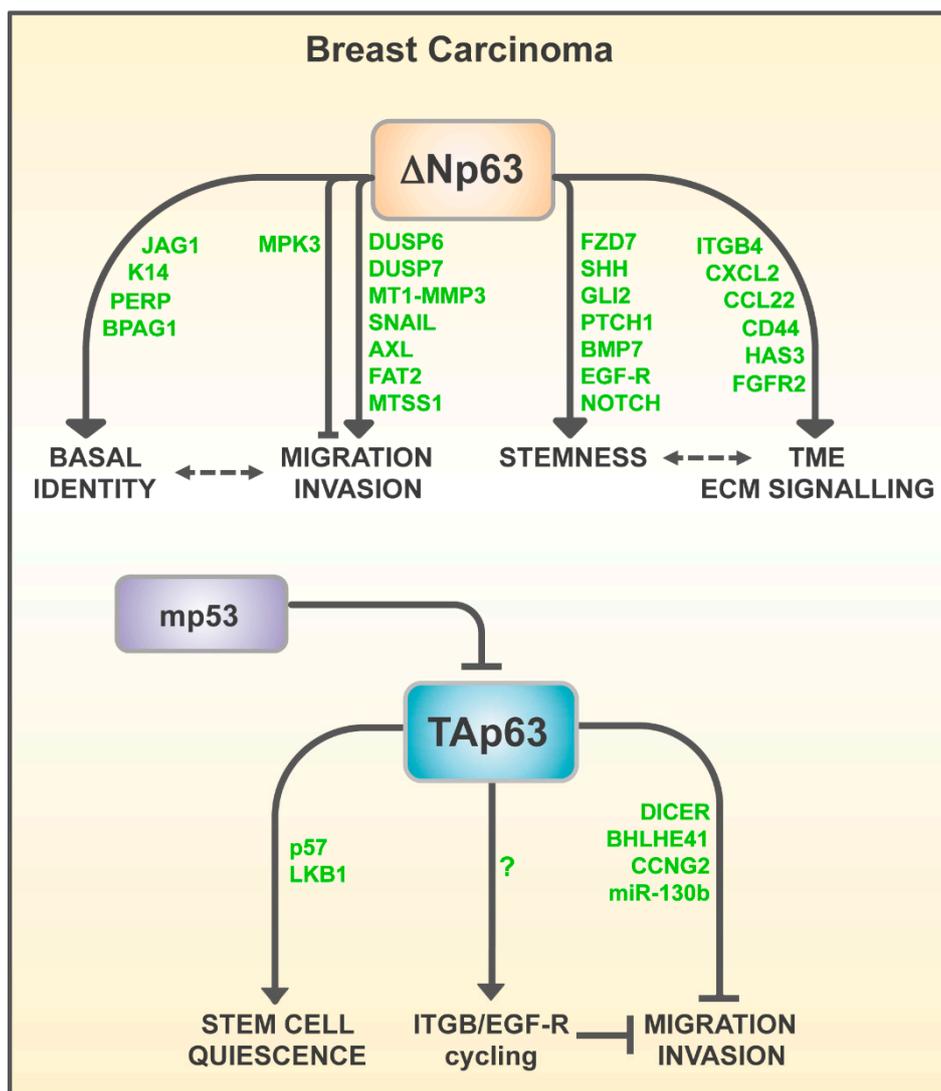
$\Delta$ Np63 activity as regulator of stemness and progenitor commitment is not restricted to basal-specific roles, but also modulates the homeostasis of specialized mammary cells, during lactation and post-lactational involution. Pregnancy-specific selective deletion of *TP63* in the basal cells is associated with a complete lactation failure, due to the impaired proliferation and differentiation of luminal cells. At the molecular level,  $\Delta$ Np63 induces the expression of the EGF family member Neuregulin 1 (*NRG1*), which triggers a paracrine signaling involving luminal *ERBB4/STAT5A* activation, leading to luminal progenitor proliferation, alveolar differentiation, and milk production [65].

During post-lactational involution,  $\Delta$ Np63 mediates the survival of Parity-Induced Mammary Epithelial Cells (PI-MECs), a heterogenous population that escapes apoptotic cell death during post-lactational involution survival. In detail,  $\Delta$ Np63 represses the expression of the STAT3-positive regulator oncostatin M and promotes the expression of *NRG1* and *NRG2* that, activating the pro-survival homolog *STAT5*, opposes the STAT3-induced apoptotic pathway [66,67].

In conclusion, these data clearly indicate a remarkable role of the  $\Delta$ Np63 in controlling molecular circuits involved in epithelial stemness and cell fate specification, ensuring the regenerative potential and plasticity of the mammary gland epithelium. In the next paragraphs, we will describe how p63 isoforms exploit several interrelated molecular pathways, to finely regulate breast tumor progression.

#### 4. $\Delta$ Np63 Oncogenic Pathways in Breast Cancer

In the past years, various transcriptional effectors of  $\Delta$ Np63-driven tumorigenesis in epithelial tumors have been identified and characterized. These  $\Delta$ Np63 transcriptional targets have been deeply investigated in squamous cell carcinoma, where  $\Delta$ Np63 acts as an oncogene by regulating expression of diverse tumor-related proteins, critically involved in the extracellular matrix (ECM) remodeling, growth factors-mediated signaling, and tumor microenvironment (TME) remodeling [68]. It is reasonable that these oncogenic routes might also be involved in the pathogenesis of breast carcinoma, contributing to enhancement of proliferation, stemness, and survival of breast tumors. In the following sections, we will focus our attention only on those  $\Delta$ Np63 transcriptional targets, which have been formally proved to play a critical role in breast carcinogenesis, emphasizing their role in controlling tumor metastasis and cancer cell stemness (see Figure 4).



**Figure 4.** Schematic model of p63-dependent routes involved in breast tumor progression (see text for details).

#### 4.1. ΔNp63 in Metastatic Dissemination

Metastasis represents the end-product of a multistep cell-biological process in which cancer cells acquire the ability to invade organs that are anatomically distant from the primary site. The metastasis cascade is modulated by many factors and epithelial-mesenchymal transition (EMT) has been identified as an important process for enhancing the invasive and migratory capabilities of tumor cells [69,70]. By maintaining epithelia identity, ΔNp63 should exert an anti-metastatic action that is able to repress the mesenchymal traits, thus inhibiting the EMT process. Accordingly, in squamous cell carcinoma, which predominantly expresses the ΔNp63 isoforms, p63 depletion led to the upregulation of mesenchymal markers associated with an increase of tumor invasion and metastasis [71]. However, a mesenchymal phenotype can be incompatible with growth in distant tissues [72,73] and basal-like breast cancers, which are intrinsically motile, can collectively invade the surrounding stroma, while maintaining epithelial features [74]. Furthermore, in spontaneous breast-to-lung metastasis models, the EMT process is not required for tumor cell dissemination but rather is implicated in modulating tumor chemoresistance [75,76]. Therefore, it is not surprising that conflicting evidence on the role ΔNp63 in regulating cell motility and invasiveness, has been reported. Here, we will describe some evidence arguing for the pro- or anti-metastatic action of ΔNp63 in breast tumors.

The idea that  $\Delta$ Np63 acts as a pro-migratory factor in breast cancer cells has been substantiated by two relatively recent papers which showed that  $\Delta$ Np63 is targeted by different oncogenic signaling, in order to enhance cell migration and invasion [77]. Yoh and colleagues reported that the expression of H-RasV12 oncogene in two non-transformed mammary epithelial cell lines (MCF10A and MCF12A) downmodulates the  $\Delta$ Np63 expression. This effect is associated with the induction of EMT, increased cell motility, and increased invasive behavior. Importantly, silencing of  $\Delta$ Np63, recapitulates the H-RasV12-mediated pro-migratory action, arguing that  $\Delta$ Np63 is a critical effector of the oncogenic activity of H-Ras. Although through a different mechanism, a further study demonstrated that activation of PI3Kinase or HER2 overexpression—two genetic events commonly observed in breast cancer [78,79]—positively regulate cell motility and tumor metastasis by targeting  $\Delta$ Np63 [80]. At the molecular level, HER2 and PI3Kinase activates AKT, which in turn phosphorylates and inhibits nuclear translocation, as well as the transactivation activity of FOXO3a, a positive regulator of  $\Delta$ Np63 transcription.

In contrast to these evidence, Vasilaki and colleagues reported that in p53 mutant breast cancer activation of Ras or TFG $\beta$  signaling induces the degradation of mutant p53, releasing  $\Delta$ Np63 transcriptional activity towards two target genes, dual specificity phosphatase 6 and 7 (DUSP6 and DUSP7) [81]. This Ras- $\Delta$ Np63-DUSPs circuit, promotes EGF-R-induced or TFG $\beta$ -induced breast carcinoma migration and invasion. Although this pathway unveils a potential molecular link between oncogenic events and an increased  $\Delta$ Np63 activity, the proposed molecular mechanism is barely reconcilable with the oncogenic function exerted by GOF p53 mutations in TNBC. Furthermore, it has been reported that in MDA-MB-231, a highly invasive TNBC cell line expressing mutant p53, the ectopic expression of  $\Delta$ Np63 is able to negatively modulate invasive behavior through the transcriptional regulation of MPK3, a regulator of ERK1/2 activity [82].

In addition, in prostate and breast cancer cell lines,  $\Delta$ Np63 restrains the migratory and invasive abilities of cancer cells, by driving the expression of miR-205, an important regulator of EMT [83–85]. On the other hand, miR-205 is also able to target p63 mRNA, creating a regulatory feedback loop, which contributes to the regulation of anti-Her2/EGFR therapy response [86]. These findings suggest that in some cellular context, and in specific genetic background,  $\Delta$ Np63 might restrain cell motility and invasion of breast cancer cells.

In contrast with these data, numerous reports unveiled a pro-migratory and pro-invasive action of  $\Delta$ Np63 in breast tumors, and different  $\Delta$ Np63 transcriptional effectors underlying these effects have been identified.

Lodillinsky and colleagues identified the membrane-type 1 membrane-anchored matrix metalloproteinase (MT1-MMP), an important protease involved in tumor invasion [87,88], as a  $\Delta$ Np63 target gene regulating the invasive properties of basal-like breast cancer cells [89]. During the transition between in situ to invasive carcinoma, MT1-MMP- $\Delta$ Np63 circuit is specifically activated in invasive cell clusters. In agreement with these data,  $\Delta$ Np63 expression has been also found at the edge of microinvasive and invasive tumor breast xenografts, suggesting that  $\Delta$ Np63 activity might be locally upregulated in invading cells. In addition to MT1-MMP,  $\Delta$ Np63 directly regulates the expression of another metalloproteinase, MMP13, although the relevance of this pathway in breast tumor has not been demonstrated [90].

Pearson's lab also confirmed a pro-metastatic role of  $\Delta$ Np63 in breast cancer [91,92]. In MCFDCIS and HCC1806 cells,  $\Delta$ Np63 silencing markedly reduces tumor cell motility by transcriptionally regulating the expression of the transcription factor Snail, the EMT-related tyrosine kinase Axl, and the transmembrane protein FAT2. The relevance of these  $\Delta$ Np63-mediated transcriptional pathways in breast tumors have been confirmed by the worse prognosis of basal-like breast cancer patients displaying high expression of p63 and Slug, or p63 and FAT2. Accordingly, in basal-like breast tumors, high  $\Delta$ Np63 $\alpha$  expression in a p53-mutated genetic background is associated with a shorter overall survival [81].

Another  $\Delta$ Np63 target gene potentially involved in breast tumor progression is the Metastasis Suppressor 1 (*MTSS1*), also known as Missing in Metastasis, or *MIM* or *BEG4*. *MTSS1* is a gene

involved in the regulation of actin-based cytoskeleton organization, whose tumor-related function is controversial, since it might act as pro-or anti-metastatic factor, depending on the tumor context [93–95]. In some breast cancer cell lines,  $\Delta$ Np63 directly drives the expression of MTSS1, enhancing the cellular migration and cytoskeleton rearrangements [81]. Notably, in three human breast tumors datasets, the MTSS1/p63 co-expression is a negative prognostic factor on patient survival [96]. Although these data are far from being conclusive, they potentially suggest a molecular effector of the pro-migratory function of  $\Delta$ Np63, at least in some human breast tumors.

As mentioned above,  $\Delta$ Np63 expression is restricted to the tumor periphery, in coincidence with the invading cell cluster. In agreement with these data, an elegant study of Ewald's lab has identified p63 as a critical basal epithelial gene controlling the collective invasion process of several breast cancer subtypes [74]. Collective invasion is a process in which tumor cells invade the ECM, cohesively, as a multicellular unit. By exploiting mouse models of the luminal-type and basal-type breast carcinogenesis, Cheung and colleagues found that cells leading collective invasion, display a conserved, basal epithelial gene expression program that is necessary to exploit the invasive process. In detail, invading tumor cells activate the expression of CK14 and p63 (likely the  $\Delta$ Np63 isoform), which are required for breast cancer cells to invade the surrounding tissues [74]. It is likely that during the collective invasion of p63 transcriptional activity, it is necessary to sustain a basal epithelial program, in order to maintain a hybrid epithelial/mesenchymal state, which is more permissive to invasion, with respect to a fully mesenchymal state. In addition to these data, this study also suggests that distinct ECM rearrangements might be critical to induce p63 expression during the collective invasion process, highlighting the importance of ECM-tumor crosstalk, during breast tumor progression. Although not fully explored, it is reasonable that ECM remodeling, cellular stroma or specific signals of the TME could sustain  $\Delta$ Np63 expression and activity, during the invasive process. For instance, breast cancer cells expressing  $\Delta$ Np63 are reliant upon the presence of stromal cells or specific compositions of the ECM, to initiate ECM remodeling that permits a collective invasion [74,91]. In TNBC and in osteosarcoma cells, a TGF $\beta$ -rich environment modulates the pro-metastatic function of  $\Delta$ Np63 [81,97]. In MCF-7 cells, an estrogen-receptor-positive breast cancer cell line, estrogen enhances cell viability and motility by inducing the  $\Delta$ Np63-ITGB4 pathway [98]. In diverse basal/triple-negative breast cancer cells p63 is able to regulate the expression of milk fat globule-EGF8 (MFG-E8), which functions in this context as a pro-survival and pro-tumorigenic factor. Conversely, in ER and erbB2-positive breast cancer cells, MFG-E8 exhibits tumor suppressive functions and is not regulated by p63 [99]. Collectively, these data indicate that TME could be determinant in modulating the pro-metastatic activity of  $\Delta$ Np63. On the other hand,  $\Delta$ Np63 is also able to model TME, in order to create a favorable niche to sustain the metastatic capabilities of breast cancer cells. In TNBC cells,  $\Delta$ Np63 directly regulates the transcription of two chemokines, CXCL2 and CCL22, which drives the recruitment of myeloid-derived immunosuppressor cells (MDSCs) [100]. MDSCs secrete pro-metastatic factors, such as MMP9 and chitinase 3-like 1, which in turn promote TNBC tumor progression and metastasis.

All together, these data suggest that  $\Delta$ Np63 exploits multiple pathways, including the induction of EMT-related factors, metallopeptidases, and activation of basal epithelial program, each likely contributing to increase the invasive capabilities of breast tumors cells. Of course, genetic and molecular features or distinct TME might critically impact and influence the biological activity of  $\Delta$ Np63, potentially explaining the conflicting evidence on the role that  $\Delta$ Np63 plays in regulating cell motility and invasiveness in breast cancer cells.

#### 4.2. $\Delta$ Np63 and Stemness of Breast Cancer Cells

As discussed in the previous paragraphs,  $\Delta$ Np63 is required to maintain the self-renewing capacity of stem cells, in diverse epithelial structures, including mammary glands [101]. Based on its role in regulating mammary stem cells homeostasis, it is not surprising that the deregulated activity of  $\Delta$ Np63 has been implicated in controlling the stemness properties of the breast cancer cells. By exploiting a mouse model of basal-type breast cancer tumorigenesis, Chakrabarti and colleagues identified a

$\Delta$ Np63-driven pathway, which is capable of governing the tumor-initiating activity of breast cancer cells [50].  $\Delta$ Np63 depletion significantly reduces tumorsphere formation, in vitro, and decreases tumor volume in vivo. As discussed before,  $\Delta$ Np63 enhances the WNT signaling, a critical regulator of epithelial stem cell homeostasis, by directly driving the expression of FZD7, a receptor for the WNT ligands. Importantly, FZD7 or  $\Delta$ Np63 depletion, exerts a similar effect on tumorspheres formation, suggesting that activation of the WNT signaling is a critical effector of the  $\Delta$ Np63-dependent control of breast cancer stemness, at least in basal-type breast cancer.

The prominent role of  $\Delta$ Np63 in controlling the self-renewal potential and expansion of mammary cancer stem cells, has also been confirmed in a mouse model of luminal type breast carcinogenesis. Downregulation of p63 in MMTV-ErbB2-derived mammospheres, which almost exclusively express the  $\Delta$ Np63 isoform, significantly limits the self-renewal capacity of cancer stem cells in vitro, and delays tumor growth, in vivo [102]. At the molecular level,  $\Delta$ Np63 enhances the Hedgehog signaling, a relevant pathway in stemness regulation, by directly controlling the expression of Sonic Hedgehog (*SHH*), GLI family zinc finger 2 (*GLI2*), and Patched1 (*PTCH1*) genes.

In addition to the WNT and Hh pathway, other transcriptional targets of  $\Delta$ Np63 involved in stemness regulation of breast cancer has been identified and characterized. One of these targets is the Notch1 receptor. As discussed above, in mammary glands, NOTCH signaling and  $\Delta$ Np63 activity are functionally interconnected in a finely regulated cross-talk, ensuring a proper balance between the luminal commitment and maintenance of the basal cell fates. In the MCF7 breast carcinoma cell line, overexpression of  $\Delta$ Np63 enhances the cancer stem cell-like features and leads to an increase of cancer cell proliferation, clonogenicity, and incidence of xenograft tumor growth, in vivo [103]. These effects have been associated with a transcriptional upregulation of NOTCH1 by  $\Delta$ Np63.

Another  $\Delta$ Np63 transcriptional target involved in breast cancer stemness is BMP7, a member of the bone morphogenetic proteins of the TGF $\beta$  superfamily of cytokines [104]. By using an in vitro cell system and a mouse model of breast cancer, Balboni and colleagues identified BMP7 as a bona fide  $\Delta$ Np63 target gene. Notably, activation of BMP7 signaling is a common event observed in human breast cancers, mainly in the basal molecular subtype, where it can regulate EMT, epithelial cell plasticity, and tumorigenicity.

One well-established biological outcome of  $\Delta$ Np63 activity in mammary gland is the modulation of ECM-mediated signaling. In normal mammary epithelial cells and in several epithelial tumors, p63 directly regulates the expression of integrin receptors (e.g., ITGB1, ITGB4, ITGA6, ITGA3), growth factor receptors (e.g., EGF-R, FGFR2), ECM components (e.g., laminin, collagen), and adhesion molecules (e.g., PERP), which ultimately might impact cancer stem cell homeostasis [68,101,105–108]. For instance, in basal-type breast tumors,  $\Delta$ Np63 regulates the expression of the hyaluronic acid (HA) synthase gene *HAS3* and the HA receptor *CD44* sustaining, thus, the synthesis and the signaling of HA [109]. Inhibition of HA synthesis or, at higher extent, depletion of CD44, decreases the stem-like properties of basal-type breast cancer cells. The relevance of the HA receptor CD44 in regulating breast cancer stemness has also been confirmed by Di Franco and colleagues, which showed that in breast cancer stem cells derived from human primary tumors,  $\Delta$ Np63 exerts a pro-metastatic action and enhances the stem-like features via upregulation of the *CD44v6* isoform expression [110]. These data suggest that HA metabolism and signaling is an important effector of the  $\Delta$ Np63-mediated stemness control.

In addition to sustain HA-mediated signaling,  $\Delta$ Np63 might also favor the activation of several tyrosine kinase receptors, such as the EGF-R, whose signaling is important to sustain the proliferation and survival of cancer stem cells [111,112].

Collectively, these results clearly indicate that the pro-tumorigenic action of  $\Delta$ Np63 in breast cancer is intimately linked to its ability to orchestrate several pathways, such as ECM remodeling, cytokine- or growth factors-mediated signaling, each of them contributing to maintain the stem-like features of breast cancer cells.

## 5. Role of TAp63 in Breast Carcinogenesis

In contrast to  $\Delta$ Np63, several lines of evidence have indicated that TAp63 activity might exert a tumor suppressor function in different human neoplasia, including breast carcinoma (see Figure 4). The initial observation that TAp63 might behave as tumor suppressor, emerged from the gain of function studies, which demonstrated that, in different cancer cell lines, TAp63 overexpression induces cell cycle arrest and cell death [113]. Although rare mutations of the *TP63* gene have been reported in human tumors, multiple studies have reported undetectable or very low expression of this isoform, in different human neoplasia, including invasive and metastatic lesions of mammary carcinoma [114]. Interestingly, the rare *TP63* mutations reported in squamous cell carcinoma are located in the TA domain, suggesting that inhibition of TAp63 isoform expression might favor tumor development [115]. The most compelling evidence impinging for a tumor suppressor role of TAp63, arises from the analysis of mice with genetically deleted TAp63 isoforms. TAp63 KO and, to a higher extent, TAp63 heterozygote mice, develop metastatic mammary and lung adenocarcinoma [116]. The anti-metastatic action of TAp63 has been further corroborated by the findings that p53 GOF mutations enhance the metastatic capabilities of tumor cells by, at least in part, inhibiting the TAp63 activity. Adorno and colleagues reported that in TNBC cells, TGF $\beta$ -mediated cell invasion is mediated by the formation of ternary complex (SMAD2-mutant p53-p63), which results in the inhibition of the transactivating action of p63, toward two p63-specific target genes, *BHLHE41* (which encodes SHARP1) and *CCNG2* (cyclin G2) [117]. Importantly, TNBC patients with a low expression of *BHLHE41* and *CCNG2*, show a reduced survival and are at a higher risk of metastasis. The anti-metastatic action of TAp63 has been also confirmed in another study, which demonstrated that in H1299 cells, which exclusively express the TAp63 isoform, p53 mutants directly interact with TAp63 in a TGF $\beta$ -independent manner, resulting in the inhibition of integrin- $\alpha$ 5 $\beta$ 1 and EGF-R endocytosis cycling [118]. A further validation of the tumor suppressor function of TAp63 in breast carcinogenesis, has emerged by the finding that in TNBC cells, the prolyl isomerase Pin1 promotes mutant p53-dependent inhibition of p63, favoring the migration and invasion of tumor cells [119].

In addition to SHARP1 and cyclin G2, another critical effector of the anti-metastatic activity of TAp63 is DICER, an endoribonuclease able to generate mature miRs, by processing the pre-miRs. TAp63 KO mice displaying reduced levels of DICER and murine TAp63 is able to bind to and transactivate the *Dicer* promoter [116]. The relevance of TAp63-DICER pathway has been revealed by the inhibitory effect of mutant p53 on the TAp63-dependent induction of DICER expression, and as a consequence, the reduction of the expression of certain pro-metastatic miRs, such as miR-130b and miR-200 [120].

Collectively these results indicate that during breast tumor initiation and progression, the decreased expression of TAp63 or its functional inhibition by mutant p53, might alter the expression of several TAp63 anti-metastatic target genes, thus increasing the metastatic potential of cancer cells.

As discussed before, TAp63 activity has been implicated in restraining the self-renewing ability of breast cancer cells [56]. In TAp63<sup>-/-</sup> mammary adenocarcinomas or in human xenograft mammary tumors depleted of TAp63, TAp63 deficiency significantly unleashes the stemness potential of human mammary cancer cells. Although these studies unveil a potential functional link between the TAp63 tumor suppressor activity and stem cells homeostasis in breast tumors, they hardly align with previous studies showing that  $\Delta$ Np63 is the only p63 isoform expressed in mammary stem cells, capable of regulating mammary stem cells activity and promoting breast cancer initiation in basal-like or luminal-like breast cancers. Likely, the analysis of tumor incidence and breast cancer stem cell activity in a mouse model of breast carcinogenesis selectively deficient for TAp63, could help clarify these controversial results.

## 6. Future Directions

TNBC is a highly malignant cancer characterized by high molecular heterogeneity and poor response to conventional therapeutic options. These clinical and molecular features negatively affect the survival rate of TNBC patients, urgently prompting to identify novel clinically actionable oncogenic

routes. The transcription factor p63 has emerged as an important regulator of two highly interconnected oncogenic pathways in breast cancer, tumor cell dissemination, and stemness. The role of the *TP63* gene in breast cancer has been historically complicated by the fact that its gene codifies for two main isoforms, TAp63 and  $\Delta$ Np63, having opposite functions. Although its involvement in breast cancer progression need to be further validated in mouse models of breast carcinogenesis, it seems that TAp63 might counteract  $\Delta$ Np63 in controlling cancer cell stemness and tumor invasion, implying that the expression of these two isoforms need to be finely regulated during breast cancer progression. In this context, future studies should be directed to investigate the molecular pathways controlling the expression of p63 isoforms, as well as their epigenetic and transcriptome landscape in breast cancer subtypes. For instance, our knowledge about the role of p63 isoforms in modulating the expression of non-coding RNAs, is very limited. Furthermore, since TAp63 $\alpha$  isoform needs post-transcriptional modifications to be transcriptionally active, it would be interesting to decipher the intracellular pathways controlling its anti-metastatic activity, as well as the role of other transcriptionally active TAp63 isoforms, such as TAp63 $\gamma$ , during breast cancer progression. On the same line,  $\Delta$ Np63 expression and activity needs to be tightly regulated during breast cancer progression, especially during the collective invasion process. Although some data have suggested the importance of ECM- $\Delta$ Np63 crosstalk in tumor progression, our knowledge on the molecular pathways regulating this circuit, as well as the  $\Delta$ Np63-dependent transcriptome and epigenetic landscape during collective invasion, is quite limited and represents a challenging question for future studies. Another fascinating aspect of the functional relationship between  $\Delta$ Np63 and TME, regard its potential involvement in modulating the immunolandscape of TNBC and as a consequence, the potential clinical use of immunotherapy. Indeed, immunotherapy might be a promising treatment modality in TNBC, mainly in basal-like subtypes, due to their high levels of neo-antigens [15,121]. Although several evidence have indicated that  $\Delta$ Np63 might transcriptionally regulate some cytokines and interleukins, the impact of  $\Delta$ Np63 on modelling the immunolandscape of TNBC still needs to be elucidated. In conclusion, we believe that further investigations on the role of p63 isoforms and their connection with TME remodeling would expand our knowledge on the pathogenesis of breast tumors, potentially unveiling novel therapeutically actionable pathways.

**Author Contributions:** A.P., G.M., V.G., and L.B.-B. wrote the paper. V.G., C.F., M.A.P., and L.B.-B. prepared the figures.

**Acknowledgments:** This work has been supported by AIRC #20473 grant, Regione Lazio through LazioInnova Progetto Gruppo di Ricerca n 85-2017-14986 to G.M. Partial support has been also provided by the MAECI Italy–China cooperation grant (#PGR00774).

**Conflicts of Interest:** The authors have no conflicts of interest to declare.

## References

1. DeSantis, C.E.; Ma, J.; Goding Sauer, A.; Newman, L.A.; Jemal, A. Breast cancer statistics, 2017, racial disparity in mortality by state. *CA A Cancer J. Clin.* **2017**, *67*, 439–448. [[CrossRef](#)] [[PubMed](#)]
2. Abubakar, I.I.; Tillmann, T.; Banerjee, A. Global, regional, and national age-sex specific all-cause and cause-specific mortality for 240 causes of death, 1990–2013: A systematic analysis for the Global Burden of Disease Study 2013. *Lancet* **2015**, *385*, 117–171.
3. Anderson, W.F.; Katki, H.A.; Rosenberg, P.S. Incidence of breast cancer in the United States: Current and future trends. *J. Natl. Cancer Inst.* **2011**, *103*, 1397–1402. [[CrossRef](#)] [[PubMed](#)]
4. Sledge, G.W.; Mamounas, E.P.; Hortobagyi, G.N.; Burstein, H.J.; Goodwin, P.J.; Wolff, A.C. Past, present, and future challenges in breast cancer treatment. *J. Clin. Oncol.* **2014**, *32*, 1979–1986. [[CrossRef](#)]
5. The Cancer Genome Atlas Network. Comprehensive molecular portraits of human breast tumours. *Nature* **2012**, *490*, 61–70. [[CrossRef](#)] [[PubMed](#)]
6. Chia, S.K.; Bramwell, V.H.; Tu, D.; Shepherd, L.E.; Jiang, S.; Vickery, T.; Mardis, E.; Leung, S.; Ung, K.; Pritchard, K.I.; et al. A 50-gene intrinsic subtype classifier for prognosis and prediction of benefit from adjuvant tamoxifen. *Clin. Cancer Res.* **2012**, *18*, 4465–4472. [[CrossRef](#)]

7. Dunbier, A.K.; Anderson, H.; Ghazoui, Z.; Salter, J.; Parker, J.S.; Perou, C.M.; Smith, I.E.; Dowsett, M. Association between breast cancer subtypes and response to neoadjuvant anastrozole. *Steroids* **2011**, *76*, 736–740. [[CrossRef](#)] [[PubMed](#)]
8. Sorlie, T.; Perou, C.M.; Tibshirani, R.; Aas, T.; Geisler, S.; Johnsen, H.; Hastie, T.; Eisen, M.B.; van de Rijn, M.; Jeffrey, S.S.; et al. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 10869–10874. [[CrossRef](#)] [[PubMed](#)]
9. Romond, E.H.; Perez, E.A.; Bryant, J.; Suman, V.J.; Geyer, C.E., Jr.; Davidson, N.E.; Tan-Chiu, E.; Martino, S.; Paik, S.; Kaufman, P.A.; et al. Trastuzumab plus adjuvant chemotherapy for operable HER2-positive breast cancer. *N. Engl. J. Med.* **2005**, *353*, 1673–1684. [[CrossRef](#)]
10. Piccart-Gebhart, M.J.; Procter, M.; Leyland-Jones, B.; Goldhirsch, A.; Untch, M.; Smith, I.; Gianni, L.; Baselga, J.; Bell, R.; Jackisch, C.; et al. Trastuzumab after adjuvant chemotherapy in HER2-positive breast cancer. *N. Engl. J. Med.* **2005**, *353*, 1659–1672. [[CrossRef](#)]
11. Hosonaga, M.; Arima, Y.; Sampetean, O.; Komura, D.; Koya, I.; Sasaki, T.; Sato, E.; Okano, H.; Kudoh, J.; Ishikawa, S.; et al. HER2 heterogeneity is associated with poor survival in HER2-positive breast cancer. *Int. J. Mol. Sci.* **2018**, *19*, 2158. [[CrossRef](#)]
12. de Angelo, R.M.; Almeida, M.O.; de Paula, H.; Honorio, K.M. Studies on the Dual Activity of EGFR and HER-2 Inhibitors Using Structure-Based Drug Design Techniques. *Int. J. Mol. Sci.* **2018**, *19*, 3728. [[CrossRef](#)]
13. Foulkes, W.D.; Smith, I.E.; Reis-Filho, J.S. Triple-negative breast cancer. *N. Engl. J. Med.* **2010**, *363*, 1938–1948. [[CrossRef](#)]
14. Shaheen, S.; Fawaz, F.; Shah, S.; Busselberg, D. Differential expression and pathway analysis in drug-resistant triple-negative breast cancer cell lines using RNASeq analysis. *Int. J. Mol. Sci.* **2018**, *19*, 1810. [[CrossRef](#)]
15. Kalimutho, M.; Parsons, K.; Mittal, D.; Lopez, J.A.; Srihari, S.; Khanna, K.K. Targeted therapies for triple-negative breast cancer: Combating a stubborn disease. *Trends Pharmacol. Sci.* **2015**, *36*, 822–846. [[CrossRef](#)]
16. Lehmann, B.D.; Bauer, J.A.; Chen, X.; Sanders, M.E.; Chakravarthy, A.B.; Shyr, Y.; Pietenpol, J.A. Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. *J. Clin. Investig.* **2011**, *121*, 2750–2767. [[CrossRef](#)]
17. Lehmann, B.D.; Jovanovic, B.; Chen, X.; Estrada, M.V.; Johnson, K.N.; Shyr, Y.; Moses, H.L.; Sanders, M.E.; Pietenpol, J.A. Refinement of triple-negative breast cancer molecular subtypes: Implications for neoadjuvant chemotherapy selection. *PLoS ONE* **2016**, *11*, e0157368. [[CrossRef](#)]
18. Yang, A.; McKeon, F. P63 and P73: P53 mimics, menaces and more. *Nat. Rev. Mol. Cell Biol.* **2000**, *1*, 199–207. [[CrossRef](#)]
19. Aubrey, B.J.; Kelly, G.L.; Janic, A.; Herold, M.J.; Strasser, A. How does p53 induce apoptosis and how does this relate to p53-mediated tumour suppression? *Cell Death Differ.* **2018**, *25*, 104–113. [[CrossRef](#)]
20. Charni, M.; Aloni-Grinstein, R.; Molchadsky, A.; Rotter, V. P53 on the crossroad between regeneration and cancer. *Cell Death Differ.* **2017**, *24*, 8–14. [[CrossRef](#)]
21. Kaiser, A.M.; Attardi, L.D. Deconstructing networks of p53-mediated tumor suppression in vivo. *Cell Death Differ.* **2018**, *25*, 93–103. [[CrossRef](#)]
22. Mello, S.S.; Attardi, L.D. Deciphering p53 signaling in tumor suppression. *Curr. Opin. Cell Biol.* **2018**, *51*, 65–72. [[CrossRef](#)]
23. Baugh, E.H.; Ke, H.; Levine, A.J.; Bonneau, R.A.; Chan, C.S. Why are there hotspot mutations in the TP53 gene in human cancers? *Cell Death Differ.* **2018**, *25*, 154–160. [[CrossRef](#)]
24. Muller, P.A.; Vousden, K.H. Mutant p53 in cancer: New functions and therapeutic opportunities. *Cancer Cell* **2014**, *25*, 304–317. [[CrossRef](#)]
25. Alexandrova, E.M.; Mirza, S.A.; Xu, S.; Schulz-Heddergott, R.; Marchenko, N.D.; Moll, U.M. P53 loss-of-heterozygosity is a necessary prerequisite for mutant p53 stabilization and gain-of-function in vivo. *Cell Death Dis.* **2017**, *8*, e2661. [[CrossRef](#)]
26. Amelio, I.; Mancini, M.; Petrova, V.; Cairns, R.A.; Vikhрева, P.; Nicolai, S.; Marini, A.; Antonov, A.A.; Le Quesne, J.; Baena Acevedo, J.D.; et al. P53 mutants cooperate with HIF-1 in transcriptional regulation of extracellular matrix components to promote tumor progression. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, E10869–E10878. [[CrossRef](#)]
27. Kim, M.P.; Lozano, G. Mutant p53 partners in crime. *Cell Death Differ.* **2018**, *25*, 161–168. [[CrossRef](#)]

28. Morrison, C.D.; Chang, J.C.; Keri, R.A.; Schiemann, W.P. Mutant p53 dictates the oncogenic activity of c-Abl in triple-negative breast cancers. *Cell Death Dis.* **2017**, *8*, e2899. [[CrossRef](#)]
29. Vaughan, C.A.; Singh, S.; Grossman, S.R.; Windle, B.; Deb, S.P.; Deb, S. Gain-of-function p53 activates multiple signaling pathways to induce oncogenicity in lung cancer cells. *Mol. Oncol.* **2017**, *11*, 696–711. [[CrossRef](#)]
30. Dotsch, V.; Bernassola, F.; Coutandin, D.; Candi, E.; Melino, G. P63 and p73, the ancestors of p53. *Cold Spring Harb. Perspect. Biol.* **2010**, *2*, a004887. [[CrossRef](#)]
31. Vanbokhoven, H.; Melino, G.; Candi, E.; Declercq, W. P63, a story of mice and men. *J. Investig. Dermatol.* **2011**, *131*, 1196–1207. [[CrossRef](#)]
32. Yang, A.; Kaghad, M.; Wang, Y.; Gillett, E.; Fleming, M.D.; Dotsch, V.; Andrews, N.C.; Caput, D.; McKeon, F. P63, a p53 homolog at 3q27–29, encodes multiple products with transactivating, death-inducing, and dominant-negative activities. *Mol. Cell* **1998**, *2*, 305–316. [[CrossRef](#)]
33. Serber, Z.; Lai, H.C.; Yang, A.; Ou, H.D.; Sigal, M.S.; Kelly, A.E.; Darimont, B.D.; Duijf, P.H.; Van Bokhoven, H.; McKeon, F.; et al. A C-terminal inhibitory domain controls the activity of p63 by an intramolecular mechanism. *Mol. Cell. Biol.* **2002**, *22*, 8601–8611. [[CrossRef](#)]
34. Straub, W.E.; Weber, T.A.; Schafer, B.; Candi, E.; Durst, F.; Ou, H.D.; Rajalingam, K.; Melino, G.; Dotsch, V. The C-terminus of p63 contains multiple regulatory elements with different functions. *Cell Death Dis.* **2010**, *1*, e5. [[CrossRef](#)] [[PubMed](#)]
35. Suh, E.K.; Yang, A.; Kettenbach, A.; Bamberger, C.; Michaelis, A.H.; Zhu, Z.; Elvin, J.A.; Bronson, R.T.; Crum, C.P.; McKeon, F. P63 protects the female germ line during meiotic arrest. *Nature* **2006**, *444*, 624–628. [[CrossRef](#)] [[PubMed](#)]
36. Gonfloni, S.; Di Tella, L.; Caldarola, S.; Cannata, S.M.; Klinger, F.G.; Di Bartolomeo, C.; Mattei, M.; Candi, E.; De Felici, M.; Melino, G.; et al. Inhibition of the c-Abl-TAP63 pathway protects mouse oocytes from chemotherapy-induced death. *Nat. Med.* **2009**, *15*, 1179–1185. [[CrossRef](#)] [[PubMed](#)]
37. Dohn, M.; Zhang, S.; Chen, X. P63alpha and DeltaNp63alpha can induce cell cycle arrest and apoptosis and differentially regulate p53 target genes. *Oncogene* **2001**, *20*, 3193–3205. [[CrossRef](#)]
38. Ghioni, P.; Bolognese, F.; Duijf, P.H.; Van Bokhoven, H.; Mantovani, R.; Guerrini, L. Complex transcriptional effects of p63 isoforms: Identification of novel activation and repression domains. *Mol. Cell. Biol.* **2002**, *22*, 8659–8668. [[CrossRef](#)]
39. Helton, E.S.; Zhu, J.; Chen, X. The unique NH2-terminally deleted (DeltaN) residues, the PXXP motif, and the PPXY motif are required for the transcriptional activity of the DeltaN variant of p63. *J. Biol. Chem.* **2006**, *281*, 2533–2542. [[CrossRef](#)]
40. Lena, A.M.; Duca, S.; Novelli, F.; Melino, S.; Annicchiarico-Petruzzelli, M.; Melino, G.; Candi, E. Amino-terminal residues of DeltaNp63, mutated in ectodermal dysplasia, are required for its transcriptional activity. *Biochem. Biophys. Res. Commun.* **2015**, *467*, 434–440. [[CrossRef](#)] [[PubMed](#)]
41. Ramsey, M.R.; He, L.; Forster, N.; Ory, B.; Ellisen, L.W. Physical association of HDAC1 and HDAC2 with p63 mediates transcriptional repression and tumor maintenance in squamous cell carcinoma. *Cancer Res.* **2011**, *71*, 4373–4379. [[CrossRef](#)]
42. Gallant-Behm, C.L.; Ramsey, M.R.; Bensard, C.L.; Nojek, I.; Tran, J.; Liu, M.; Ellisen, L.W.; Espinosa, J.M. DeltaNp63alpha represses anti-proliferative genes via H2A.Z deposition. *Genes Dev.* **2012**, *26*, 2325–2336. [[CrossRef](#)]
43. Regina, C.; Compagnone, M.; Peschiaroli, A.; Lena, A.; Annicchiarico-Petruzzelli, M.; Piro, M.C.; Melino, G.; Candi, E. Setdb1, a novel interactor of DeltaNp63, is involved in breast tumorigenesis. *Oncotarget* **2016**, *7*, 28836–28848. [[CrossRef](#)]
44. Saladi, S.V.; Ross, K.; Karaayvaz, M.; Tata, P.R.; Mou, H.; Rajagopal, J.; Ramaswamy, S.; Ellisen, L.W. ACTL6A Is Co-Amplified with p63 in Squamous Cell Carcinoma to Drive YAP Activation, Regenerative Proliferation, and Poor Prognosis. *Cancer Cell* **2017**, *31*, 35–49. [[CrossRef](#)]
45. Inman, J.L.; Robertson, C.; Mott, J.D.; Bissell, M.J. Mammary gland development: Cell fate specification, stem cells and the microenvironment. *Development* **2015**, *142*, 1028–1042. [[CrossRef](#)] [[PubMed](#)]
46. McNally, S.; Stein, T. Overview of Mammary Gland Development: A Comparison of Mouse and Human. *Methods Mol. Biol.* **2017**, *1501*, 1–17. [[PubMed](#)]
47. Stingl, J.; Raouf, A.; Eirew, P.; Eaves, C.J. Deciphering the mammary epithelial cell hierarchy. *Cell Cycle* **2006**, *5*, 1519–1522. [[CrossRef](#)]

48. Shackleton, M.; Vaillant, F.; Simpson, K.J.; Stingl, J.; Smyth, G.K.; Asselin-Labat, M.L.; Wu, L.; Lindeman, G.J.; Visvader, J.E. Generation of a functional mammary gland from a single stem cell. *Nature* **2006**, *439*, 84–88. [[CrossRef](#)] [[PubMed](#)]
49. Van Keymeulen, A.; Fioramonti, M.; Centonze, A.; Bouvencourt, G.; Achouri, Y.; Blanpain, C. Lineage-Restricted Mammary Stem Cells Sustain the Development, Homeostasis, and Regeneration of the Estrogen Receptor Positive Lineage. *Cell Rep.* **2017**, *20*, 1525–1532. [[CrossRef](#)]
50. Chakrabarti, R.; Wei, Y.; Hwang, J.; Hang, X.; Andres Blanco, M.; Choudhury, A.; Tiede, B.; Romano, R.A.; DeCoste, C.; Mercatali, L.; et al. DeltaNp63 promotes stem cell activity in mammary gland development and basal-like breast cancer by enhancing Fzd7 expression and Wnt signalling. *Nat. Cell Biol.* **2014**, *16*, 1004–1015. [[CrossRef](#)]
51. Yalcin-Ozuysal, O.; Fiche, M.; Guitierrez, M.; Wagner, K.U.; Raffoul, W.; Briskin, C. Antagonistic roles of Notch and p63 in controlling mammary epithelial cell fates. *Cell Death Diff.* **2010**, *17*, 1600–1612. [[CrossRef](#)]
52. Yang, A.; Schweitzer, R.; Sun, D.; Kaghad, M.; Walker, N.; Bronson, R.T.; Tabin, C.; Sharpe, A.; Caput, D.; Crum, C.; et al. P63 is essential for regenerative proliferation in limb, craniofacial and epithelial development. *Nature* **1999**, *398*, 714–718. [[CrossRef](#)]
53. Senoo, M.; Pinto, F.; Crum, C.P.; McKeon, F. P63 Is essential for the proliferative potential of stem cells in stratified epithelia. *Cell* **2007**, *129*, 523–536. [[CrossRef](#)]
54. Mills, A.A.; Zheng, B.; Wang, X.J.; Vogel, H.; Roop, D.R.; Bradley, A. P63 is a p53 homologue required for limb and epidermal morphogenesis. *Nature* **1999**, *398*, 708–713. [[CrossRef](#)]
55. Koster, M.I.; Dai, D.; Marinari, B.; Sano, Y.; Costanzo, A.; Karin, M.; Roop, D.R. P63 induces key target genes required for epidermal morphogenesis. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 3255–3260. [[CrossRef](#)]
56. Su, X.; Napoli, M.; Abbas, H.A.; Venkatanarayan, A.; Bui, N.H.B.; Coarfa, C.; Gi, Y.J.; Kittrell, F.; Gunaratne, P.H.; Medina, D.; et al. TAp63 suppresses mammary tumorigenesis through regulation of the Hippo pathway. *Oncogene* **2017**, *36*, 2377–2393. [[CrossRef](#)]
57. Li, N.; Singh, S.; Cherukuri, P.; Li, H.; Yuan, Z.; Ellisen, L.W.; Wang, B.; Robbins, D.; DiRenzo, J. Reciprocal intraepithelial interactions between TP63 and hedgehog signaling regulate quiescence and activation of progenitor elaboration by mammary stem cells. *Stem Cells* **2008**, *26*, 1253–1264. [[CrossRef](#)]
58. Lindley, L.E.; Curtis, K.M.; Sanchez-Mejias, A.; Rieger, M.E.; Robbins, D.J.; Briegel, K.J. The WNT-controlled transcriptional regulator LBH is required for mammary stem cell expansion and maintenance of the basal lineage. *Development* **2015**, *142*, 893–904. [[CrossRef](#)]
59. Wuidart, A.; Sifrim, A.; Fioramonti, M.; Matsumura, S.; Brisebarre, A.; Brown, D.; Centonze, A.; Dannau, A.; Dubois, C.; Van Keymeulen, A.; et al. Early lineage segregation of multipotent embryonic mammary gland progenitors. *Nat. Cell Biol.* **2018**, *20*, 666–676. [[CrossRef](#)]
60. Nagarajan, S.; Bedi, U.; Budida, A.; Hamdan, F.H.; Mishra, V.K.; Najafova, Z.; Xie, W.; Alawi, M.; Indenbirken, D.; Knapp, S.; et al. BRD4 promotes p63 and GRHL3 expression downstream of FOXO in mammary epithelial cells. *Nucleic Acids Res.* **2017**, *45*, 3130–3145. [[CrossRef](#)]
61. Foulkes, W.D. BRCA1 functions as a breast stem cell regulator. *J. Med. Genet.* **2004**, *41*, 1–5. [[CrossRef](#)]
62. Molyneux, G.; Geyer, F.C.; Magnay, F.A.; McCarthy, A.; Kendrick, H.; Natrajan, R.; Mackay, A.; Grigoriadis, A.; Tutt, A.; Ashworth, A.; et al. BRCA1 basal-like breast cancers originate from luminal epithelial progenitors and not from basal stem cells. *Cell Stem Cell* **2010**, *7*, 403–417. [[CrossRef](#)]
63. Buckley, N.E.; Conlon, S.J.; Jirstrom, K.; Kay, E.W.; Crawford, N.T.; O’Grady, A.; Sheehan, K.; Mc Dade, S.S.; Wang, C.W.; McCance, D.J.; et al. The DeltaNp63 proteins are key allies of BRCA1 in the prevention of basal-like breast cancer. *Cancer Res.* **2011**, *71*, 1933–1944. [[CrossRef](#)]
64. Kent, S.; Hutchinson, J.; Balboni, A.; Decastro, A.; Cherukuri, P.; Drenzo, J. DeltaNp63alpha promotes cellular quiescence via induction and activation of Notch3. *Cell Cycle* **2011**, *10*, 3111–3118. [[CrossRef](#)]
65. Forster, N.; Saladi, S.V.; van Bragt, M.; Sfondouris, M.E.; Jones, F.E.; Li, Z.; Ellisen, L.W. Basal cell signaling by p63 controls luminal progenitor function and lactation via NRG1. *Dev. Cell* **2014**, *28*, 147–160. [[CrossRef](#)]
66. Yallowitz, A.R.; Alexandrova, E.M.; Talos, F.; Xu, S.; Marchenko, N.D.; Moll, U.M. P63 is a prosurvival factor in the adult mammary gland during post-lactational involution, affecting PI-MECs and ErbB2 tumorigenesis. *Cell Death Diff.* **2014**, *21*, 645–654. [[CrossRef](#)]
67. Hughes, K.; Watson, C.J. The Multifaceted Role of STAT3 in Mammary Gland Involution and Breast Cancer. *Int. J. Mol. Sci.* **2018**, *19*, 1695. [[CrossRef](#)]

68. Gatti, V.; Fierro, C.; Annicchiarico-Petruzzelli, M.; Melino, G.; Peschiaroli, A. DeltaNp63 in squamous cell carcinoma: Defining the oncogenic routes affecting epigenetic landscape and tumour microenvironment. *Mol. Oncol.* **2019**, *13*, 981–1001.
69. Tsubakihara, Y.; Moustakas, A. Epithelial-Mesenchymal Transition and Metastasis under the Control of Transforming Growth Factor beta. *Int. J. Mol. Sci.* **2018**, *19*, 3672. [[CrossRef](#)]
70. Brabletz, T.; Kalluri, R.; Nieto, M.A.; Weinberg, R.A. EMT in cancer. *Nat. Rev. Cancer* **2018**, *18*, 128–134. [[CrossRef](#)]
71. Barbieri, C.E.; Tang, L.J.; Brown, K.A.; Pietenpol, J.A. Loss of p63 leads to increased cell migration and up-regulation of genes involved in invasion and metastasis. *Cancer Res.* **2006**, *66*, 7589–7597. [[CrossRef](#)]
72. Ocana, O.H.; Corcoles, R.; Fabra, A.; Moreno-Bueno, G.; Acloque, H.; Vega, S.; Barrallo-Gimeno, A.; Cano, A.; Nieto, M.A. Metastatic colonization requires the repression of the epithelial-mesenchymal transition inducer Prrx1. *Cancer Cell* **2012**, *22*, 709–724. [[CrossRef](#)] [[PubMed](#)]
73. Tsai, J.H.; Donaher, J.L.; Murphy, D.A.; Chau, S.; Yang, J. Spatiotemporal regulation of epithelial-mesenchymal transition is essential for squamous cell carcinoma metastasis. *Cancer Cell* **2012**, *22*, 725–736. [[CrossRef](#)] [[PubMed](#)]
74. Cheung, K.J.; Gabrielson, E.; Werb, Z.; Ewald, A.J. Collective invasion in breast cancer requires a conserved basal epithelial program. *Cell* **2013**, *155*, 1639–1651. [[CrossRef](#)] [[PubMed](#)]
75. Fischer, K.R.; Durrans, A.; Lee, S.; Sheng, J.; Li, F.; Wong, S.T.; Choi, H.; El Rayes, T.; Ryu, S.; Troeger, J.; et al. Epithelial-to-mesenchymal transition is not required for lung metastasis but contributes to chemoresistance. *Nature* **2015**, *527*, 472–476. [[CrossRef](#)]
76. Zheng, X.; Carstens, J.L.; Kim, J.; Scheible, M.; Kaye, J.; Sugimoto, H.; Wu, C.C.; LeBleu, V.S.; Kalluri, R. Epithelial-to-mesenchymal transition is dispensable for metastasis but induces chemoresistance in pancreatic cancer. *Nature* **2015**, *527*, 525–530. [[CrossRef](#)]
77. Yoh, K.E.; Regunath, K.; Guzman, A.; Lee, S.M.; Pfister, N.T.; Akanni, O.; Kaufman, L.J.; Prives, C.; Prywes, R. Repression of p63 and induction of EMT by mutant Ras in mammary epithelial cells. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, E6107–E6116. [[CrossRef](#)]
78. Pereira, B.; Chin, S.F.; Rueda, O.M.; Vollan, H.K.; Provenzano, E.; Bardwell, H.A.; Pugh, M.; Jones, L.; Russell, R.; Sammut, S.J.; et al. The somatic mutation profiles of 2,433 breast cancers refines their genomic and transcriptomic landscapes. *Nat. Commun.* **2016**, *7*, 11479. [[CrossRef](#)]
79. Koepfen, H.K.; Wright, B.D.; Burt, A.D.; Quirke, P.; McNicol, A.M.; Dybdal, N.O.; Sliwkowski, M.X.; Hillan, K.J. Overexpression of HER2/neu in solid tumours: An immunohistochemical survey. *Histopathology* **2001**, *38*, 96–104. [[CrossRef](#)]
80. Hu, L.; Liang, S.; Chen, H.; Lv, T.; Wu, J.; Chen, D.; Wu, M.; Sun, S.; Zhang, H.; You, H.; et al. DeltaNp63alpha is a common inhibitory target in oncogenic PI3K/Ras/Her2-induced cell motility and tumor metastasis. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, E3964–E3973. [[CrossRef](#)]
81. Vasilaki, E.; Morikawa, M.; Koinuma, D.; Mizutani, A.; Hirano, Y.; Ehata, S.; Sundqvist, A.; Kawasaki, N.; Cedervall, J.; Olsson, A.K.; et al. Ras and TGF-beta signaling enhance cancer progression by promoting the DeltaNp63 transcriptional program. *Sci. Signal.* **2016**, *9*, ra84. [[CrossRef](#)]
82. Bergholz, J.; Zhang, Y.; Wu, J.; Meng, L.; Walsh, E.M.; Rai, A.; Sherman, M.Y.; Xiao, Z.X. DeltaNp63alpha regulates Erk signaling via MKP3 to inhibit cancer metastasis. *Oncogene* **2014**, *33*, 212–224. [[CrossRef](#)]
83. Tucci, P.; Agostini, M.; Grespi, F.; Markert, E.K.; Terrinoni, A.; Vousden, K.H.; Muller, P.A.; Dotsch, V.; Kehrlöesser, S.; Sayan, B.S.; et al. Loss of p63 and its microRNA-205 target results in enhanced cell migration and metastasis in prostate cancer. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 15312–15317. [[CrossRef](#)]
84. Rhodes, L.V.; Martin, E.C.; Segar, H.C.; Miller, D.F.; Buechlein, A.; Rusch, D.B.; Nephew, K.P.; Burow, M.E.; Collins-Burow, B.M. Dual regulation by microRNA-200b-3p and microRNA-200b-5p in the inhibition of epithelial-to-mesenchymal transition in triple-negative breast cancer. *Oncotarget* **2015**, *6*, 16638–16652. [[CrossRef](#)]
85. De Cola, A.; Lamolinara, A.; Lanuti, P.; Rossi, C.; Iezzi, M.; Marchisio, M.; Todaro, M.; De Laurenzi, V. MiR-205-5p inhibition by locked nucleic acids impairs metastatic potential of breast cancer cells. *Cell Death Dis.* **2018**, *9*, 821. [[CrossRef](#)]
86. De Cola, A.; Volpe, S.; Budani, M.C.; Ferracin, M.; Lattanzio, R.; Turdo, A.; D’Agostino, D.; Capone, E.; Stassi, G.; Todaro, M.; et al. MiR-205-5p-mediated downregulation of ErbB/HER receptors in breast cancer stem cells results in targeted therapy resistance. *Cell Death Dis.* **2015**, *6*, e1823. [[CrossRef](#)]

87. Hotary, K.B.; Allen, E.D.; Brooks, P.C.; Datta, N.S.; Long, M.W.; Weiss, S.J. Membrane type I matrix metalloproteinase usurps tumor growth control imposed by the three-dimensional extracellular matrix. *Cell* **2003**, *114*, 33–45. [[CrossRef](#)]
88. Perentes, J.Y.; Kirkpatrick, N.D.; Nagano, S.; Smith, E.Y.; Shaver, C.M.; Sgroi, D.; Garkavtsev, I.; Munn, L.L.; Jain, R.K.; Boucher, Y. Cancer cell-associated MT1-MMP promotes blood vessel invasion and distant metastasis in triple-negative mammary tumors. *Cancer Res.* **2011**, *71*, 4527–4538. [[CrossRef](#)]
89. Lodillinsky, C.; Infante, E.; Guichard, A.; Chaligne, R.; Fuhrmann, L.; Cyrta, J.; Irondelle, M.; Lagoutte, E.; Vacher, S.; Bonsang-Kitzis, H.; et al. P63/MT1-MMP axis is required for in situ to invasive transition in basal-like breast cancer. *Oncogene* **2016**, *35*, 344–357. [[CrossRef](#)]
90. Celardo, I.; Antonov, A.; Amelio, I.; Annicchiarico-Petruzzelli, M.; Melino, G. P63 transcriptionally regulates the expression of matrix metalloproteinase 13. *Oncotarget* **2014**, *5*, 1279–1289. [[CrossRef](#)]
91. Dang, T.T.; Esparza, M.A.; Maine, E.A.; Westcott, J.M.; Pearson, G.W. DeltaNp63alpha promotes breast cancer cell motility through the selective activation of components of the epithelial-to-mesenchymal transition program. *Cancer Res.* **2015**, *75*, 3925–3935. [[CrossRef](#)]
92. Dang, T.T.; Westcott, J.M.; Maine, E.A.; Kanchwala, M.; Xing, C.; Pearson, G.W. DeltaNp63alpha induces the expression of FAT2 and Slug to promote tumor invasion. *Oncotarget* **2016**, *7*, 28592–28611. [[CrossRef](#)] [[PubMed](#)]
93. Fan, H.; Chen, L.; Zhang, F.; Quan, Y.; Su, X.; Qiu, X.; Zhao, Z.; Kong, K.L.; Dong, S.; Song, Y.; et al. MTSS1, a novel target of DNA methyltransferase 3B, functions as a tumor suppressor in hepatocellular carcinoma. *Oncogene* **2012**, *31*, 2298–2308. [[CrossRef](#)]
94. Mertz, K.D.; Pathria, G.; Wagner, C.; Saarikangas, J.; Sboner, A.; Romanov, J.; Gschaidner, M.; Lenz, F.; Neumann, F.; Schreiner, W.; et al. MTSS1 is a metastasis driver in a subset of human melanomas. *Nat. Commun.* **2014**, *5*, 3465. [[CrossRef](#)] [[PubMed](#)]
95. Machesky, L.M.; Johnston, S.A. MIM: A multifunctional scaffold protein. *J. Mol. Med.* **2007**, *85*, 569–576. [[CrossRef](#)]
96. Giacobbe, A.; Compagnone, M.; Bongiorno-Borbone, L.; Antonov, A.; Markert, E.K.; Zhou, J.H.; Annicchiarico-Petruzzelli, M.; Melino, G.; Peschiaroli, A. P63 controls cell migration and invasion by transcriptional regulation of MTSS1. *Oncogene* **2016**, *35*, 1602–1608. [[CrossRef](#)]
97. Rodriguez Calleja, L.; Jacques, C.; Lamoureux, F.; Baud'huin, M.; Tellez Gabriel, M.; Quillard, T.; Sahay, D.; Perrot, P.; Amiaud, J.; Charrier, C.; et al. DeltaNp63alpha Silences a miRNA Program to Aberrantly Initiate a Wound-Healing Program That Promotes TGFbeta-Induced Metastasis. *Cancer Res.* **2016**, *76*, 3236–3251. [[CrossRef](#)]
98. Ho, J.Y.; Chang, F.W.; Huang, F.S.; Liu, J.M.; Liu, Y.P.; Chen, S.P.; Liu, Y.L.; Cheng, K.C.; Yu, C.P.; Hsu, R.J. Estrogen Enhances the Cell Viability and Motility of Breast Cancer Cells through the ERalpha-DeltaNp63-Integrin beta4 Signaling Pathway. *PLoS ONE* **2016**, *11*, e0148301. [[CrossRef](#)]
99. Yang, C.; Hayashida, T.; Forster, N.; Li, C.; Shen, D.; Maheswaran, S.; Chen, L.; Anderson, K.S.; Ellisen, L.W.; Sgroi, D.; et al. The integrin alpha(v)beta(3-5) ligand MFG-E8 is a p63/p73 target gene in triple-negative breast cancers but exhibits suppressive functions in ER(+) and erbB2(+) breast cancers. *Cancer Res.* **2011**, *71*, 937–945. [[CrossRef](#)]
100. Kumar, S.; Wilkes, D.W.; Samuel, N.; Blanco, M.A.; Nayak, A.; Alicea-Torres, K.; Gluck, C.; Sinha, S.; Gabrilovich, D.; Chakrabarti, R. DeltaNp63-driven recruitment of myeloid-derived suppressor cells promotes metastasis in triple-negative breast cancer. *J. Clin. Investig.* **2018**, *128*, 5095–5109. [[CrossRef](#)] [[PubMed](#)]
101. Melino, G.; Memmi, E.M.; Pelicci, P.G.; Bernassola, F. Maintaining epithelial stemness with p63. *Sci. Signal.* **2015**, *8*, re9. [[CrossRef](#)]
102. Memmi, E.M.; Sanarico, A.G.; Giacobbe, A.; Peschiaroli, A.; Frezza, V.; Cicalese, A.; Pisati, F.; Tosoni, D.; Zhou, H.; Tonon, G.; et al. P63 Sustains self-renewal of mammary cancer stem cells through regulation of Sonic Hedgehog signaling. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 3499–3504. [[CrossRef](#)]
103. Du, Z.; Li, J.; Wang, L.; Bian, C.; Wang, Q.; Liao, L.; Dou, X.; Bian, X.; Zhao, R.C. Overexpression of DeltaNp63alpha induces a stem cell phenotype in MCF7 breast carcinoma cell line through the Notch pathway. *Cancer Sci.* **2010**, *101*, 2417–2424. [[CrossRef](#)]
104. Balboni, A.L.; Hutchinson, J.A.; DeCastro, A.J.; Cherukuri, P.; Liby, K.; Sporn, M.B.; Schwartz, G.N.; Wells, W.A.; Sempere, L.F.; Yu, P.B.; et al. DeltaNp63alpha-mediated activation of bone morphogenetic protein signaling governs stem cell activity and plasticity in normal and malignant mammary epithelial cells. *Cancer Res.* **2013**, *73*, 1020–1030. [[CrossRef](#)]

105. Carroll, D.K.; Carroll, J.S.; Leong, C.O.; Cheng, F.; Brown, M.; Mills, A.A.; Brugge, J.S.; Ellisen, L.W. P63 regulates an adhesion programme and cell survival in epithelial cells. *Nat. Cell Biol.* **2006**, *8*, 551–561. [[CrossRef](#)]
106. Ihrie, R.A.; Marques, M.R.; Nguyen, B.T.; Horner, J.S.; Papazoglu, C.; Bronson, R.T.; Mills, A.A.; Attardi, L.D. Perp is a p63-regulated gene essential for epithelial integrity. *Cell* **2005**, *120*, 843–856. [[CrossRef](#)]
107. Kurata, S.; Okuyama, T.; Osada, M.; Watanabe, T.; Tomimori, Y.; Sato, S.; Iwai, A.; Tsuji, T.; Ikawa, Y.; Katoh, I. P51/p63 Controls subunit alpha3 of the major epidermis integrin anchoring the stem cells to the niche. *J. Biol. Chem.* **2004**, *279*, 50069–50077. [[CrossRef](#)]
108. Compagnone, M.; Gatti, V.; Presutti, D.; Ruberti, G.; Fierro, C.; Markert, E.K.; Vousden, K.H.; Zhou, H.; Mauriello, A.; Anemone, L.; et al. DeltaNp63-mediated regulation of hyaluronic acid metabolism and signaling supports HNSCC tumorigenesis. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 13254–13259. [[CrossRef](#)]
109. Gatti, V.; Fierro, C.; Compagnone, M.; Giangrazi, F.; Markert, E.K.; Bongiorno-Borbone, L.; Melino, G.; Peschiaroli, A. DeltaNp63 regulates the expression of hyaluronic acid-related genes in breast cancer cells. *Oncogenesis* **2018**, *7*, 65. [[CrossRef](#)]
110. Di Franco, S.; Turdo, A.; Benfante, A.; Colorito, M.L.; Gaggianesi, M.; Apuzzo, T.; Kandimalla, R.; Chinnici, A.; Barcaroli, D.; Mangiapane, L.R.; et al. DeltaNp63 drives metastasis in breast cancer cells via PI3K/CD44v6 axis. *Oncotarget* **2016**, *7*, 54157–54173. [[CrossRef](#)]
111. Holcakova, J.; Nekulova, M.; Orzol, P.; Nenutil, R.; Podhorec, J.; Svoboda, M.; Dvorakova, P.; Pjehova, M.; Hernychova, L.; Vojtesek, B.; et al. DeltaNp63 activates EGFR signaling to induce loss of adhesion in triple-negative basal-like breast cancer cells. *Breast Cancer Res. Treat.* **2017**, *163*, 475–484. [[CrossRef](#)] [[PubMed](#)]
112. Sigismund, S.; Avanzato, D.; Lanzetti, L. Emerging functions of the EGFR in cancer. *Mol. Oncol.* **2018**, *12*, 3–20. [[CrossRef](#)]
113. Gressner, O.; Schilling, T.; Lorenz, K.; Schulze Schleithoff, E.; Koch, A.; Schulze-Bergkamen, H.; Lena, A.M.; Candi, E.; Terrinoni, A.; Catani, M.V.; et al. TAp63alpha induces apoptosis by activating signaling via death receptors and mitochondria. *EMBO J.* **2005**, *24*, 2458–2471. [[CrossRef](#)] [[PubMed](#)]
114. Melino, G. p63 is a suppressor of tumorigenesis and metastasis interacting with mutant p53. *Cell Death Diff.* **2011**, *18*, 1487–1499. [[CrossRef](#)] [[PubMed](#)]
115. Stransky, N.; Egloff, A.M.; Tward, A.D.; Kostic, A.D.; Cibulskis, K.; Sivachenko, A.; Kryukov, G.V.; Lawrence, M.S.; Sougnez, C.; McKenna, A.; et al. The mutational landscape of head and neck squamous cell carcinoma. *Science* **2011**, *333*, 1157–1160. [[CrossRef](#)] [[PubMed](#)]
116. Su, X.; Chakravarti, D.; Cho, M.S.; Liu, L.; Gi, Y.J.; Lin, Y.L.; Leung, M.L.; El-Naggar, A.; Creighton, C.J.; Suraokar, M.B.; et al. TAp63 suppresses metastasis through coordinate regulation of Dicer and miRNAs. *Nature* **2010**, *467*, 986–990. [[CrossRef](#)]
117. Adorno, M.; Cordenonsi, M.; Montagner, M.; Dupont, S.; Wong, C.; Hann, B.; Solari, A.; Bobisse, S.; Rondina, M.B.; Guzzardo, V.; et al. A Mutant-p53/Smad complex opposes p63 to empower TGFbeta-induced metastasis. *Cell* **2009**, *137*, 87–98. [[CrossRef](#)] [[PubMed](#)]
118. Muller, P.A.; Caswell, P.T.; Doyle, B.; Iwanicki, M.P.; Tan, E.H.; Karim, S.; Lukashchuk, N.; Gillespie, D.A.; Ludwig, R.L.; Gosselin, P.; et al. Mutant p53 drives invasion by promoting integrin recycling. *Cell* **2009**, *139*, 1327–1341. [[CrossRef](#)] [[PubMed](#)]
119. Girardini, J.E.; Napoli, M.; Piazza, S.; Rustighi, A.; Marotta, C.; Radaelli, E.; Capaci, V.; Jordan, L.; Quinlan, P.; Thompson, A.; et al. A Pin1/mutant p53 axis promotes aggressiveness in breast cancer. *Cancer Cell* **2011**, *20*, 79–91. [[CrossRef](#)]
120. Muller, P.A.; Trinidad, A.G.; Caswell, P.T.; Norman, J.C.; Vousden, K.H. Mutant p53 regulates Dicer through p63-dependent and -independent mechanisms to promote an invasive phenotype. *J. Biol. Chem.* **2014**, *289*, 122–132. [[CrossRef](#)]
121. Emens, L.A. Breast Cancer Immunotherapy: Facts and Hopes. *Clin. Cancer Res.* **2018**, *24*, 511–520. [[CrossRef](#)]

