



*Hypothesis*

# Nanoparticles and Nanomaterials as Plant Biostimulants

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**Abstract:** Biostimulants are materials that when applied in small amounts are capable of promoting plant growth. Nanoparticles (NPs) and nanomaterials (NMs) can be considered as biostimulants since, in specific ranges of concentration, generally in small levels, they increase plant growth. Pristine NPs and NMs have a high density of surface charges capable of unspecific interactions with the surface charges of the cell walls and membranes of plant cells. In the same way, functionalized NPs and NMs, and the NPs and NMs with a corona formed after the exposition to natural fluids such as water, soil solution, or the interior of organisms, present a high density of surface charges that interact with specific charged groups in cell surfaces. The magnitude of the interaction will depend on the materials adhered to the corona, but high-density charges located in a small volume cause an intense interaction capable of disturbing the density of surface charges of cell walls and membranes. The electrostatic disturbance can have an impact on the electrical potentials of the outer and inner surfaces, as well as on the transmembrane electrical potential, modifying the activity of the integral proteins of the membranes. The extension of the cellular response can range from biostimulation to cell death and will depend on the concentration, size, and the characteristics of the corona.

**Keywords:** biostimulation; stress tolerance; elicitors; corona; hormesis; nutritional quality; growth promoters

## 1. Introduction

Biostimulation of an organism is a phenomenon of modification in metabolic processes that allow the more efficient use of environmental resources, more significant growth or yield, and more tolerance to adverse environmental factors. Biostimulation has been described as a general biological phenomena dependent of the interactions between cell molecular structures and external impulses or stimuli [1]. After the application of a biostimulating agent, a sequence of events of perception, transduction, signaling, effector action, and modification in gene expression, metabolism or cellular (and organismal) characters occurs. Environmental agents that cause biostimulation in living organisms can be physical,

chemical, or biological [2,3]. In cultivated plants, biostimulation has been defined as the result of applying substances or microorganisms that do not directly add essential elements, known plant hormones, or molecules with suppressive effect of diseases [4,5].

Nanoparticles of metals, semimetals, and non-metals (NPs) and nanomaterials (NMs) of C (such as carbon nanotubes), organic (such as chitosan) and other materials such as nanoclays are materials with at least one dimension less than 100 nm. This small size gives rise to properties different from those exhibited by the bulk material of the same composition. These new properties provide the material with an added value that has multiple applications in automotive, energy, pharmaceutical, medical, and agricultural industries, among others [6].

In the present manuscript it is proposed that due to the physicochemical characteristics of the NPs and NMs, as well as the responses induced in the plants, NPs and NMs could be considered in the group of compounds useful for crops known as biostimulants. The capacity of these materials, applied in generally small amounts and by foliar spray or in nutrient solutions, has been described to modify the composition and nutraceutical quality of food crops, as well as their tolerance to stress [7–9]. The purpose of the proposal is that, at a time when the legislation on agricultural biostimulants is being discussed [4,5], it is perhaps an appropriate time for the consideration of NPs and NMs to be included as a kind of biostimulant, taking into account the enormous potential utility for the agricultural sector. In this context, the effectiveness of NPs and NMs applied in minimal quantities should be highlighted. The use in low concentrations is recommended because it decreases the risk of a plant's toxicity [10,11] and, in the case of NPs of metals such as Fe, Mn, and Cu, the low amounts applied may decrease the transfer to the soils and food chain, in comparison with fertilizer bulk materials [12].

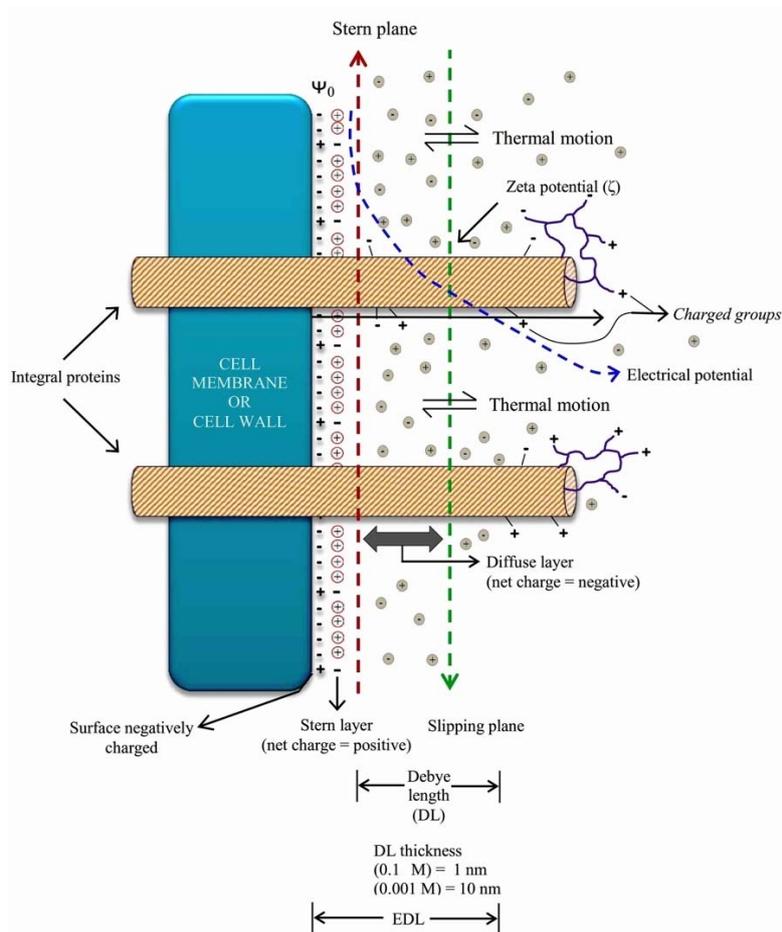
Apparently, biostimulation by NPs and NMs occurs in two phases. The first is of a physicochemical nature through the energy and surface charges of the NPs and NMs, which are associated with the surfaces of walls and cell membranes modifying the activity of receptors, transporters, and other proteins, also modifying the transport of metabolites and ions to cells [13]. The second phase occurs through biochemical processes that respond to the release of the chemical elements from the NP or NM (for example  $\text{Fe}^{3+}$  from Fe NPs or carbon compounds from NPs of chitosan) and that would be analogous to the responses to metal ions or organic molecules in their bulk form [14,15]. The first phase, the interaction of surfaces, in the case of a physicochemical process, can be independent from the composition, this is why it is possible that practically any type of inorganic (Cu, CuO, silica) or organic (polystyrene, chitosan) NP or NM in specific concentration ranges has this kind of biostimulating capacity. Indeed, it has been proposed that the surface area of the NPs and NMs, instead of the concentration, be used as a predictor of the potential biological impact [16].

In addition to the Introduction, the manuscript has three main sections. In Section 2, the subject of the superficial charges of the cells is included. Section 3 consists of the explanation about the surface charges of the NPs and NMs and the interaction of the surface charges of cells and NPs and NMs; explaining how the interaction process can cause biostimulation. Section 4 includes examples of the biostimulation process.

## 2. The Charges of the Cell's Surface

The walls and cell membranes, the latter through the plasmodesmata [17], are in contact with the apoplast, where there are abundant organic and inorganic chemical species with charge. Several chemical interactions occur at the wall-membrane-apoplast interface, such as acid/base dissociations, ionization of functional groups, ion adsorption or other species charged on surfaces, as well as the partial dissolution of some of the structural components of the membranes and cell walls. The intensity and final balance of these interactions depend on the pH, ionic strength, oxidation-reduction potential, and other dynamic properties of the extracellular medium. When reaching a point of dynamic equilibrium between the cell surfaces and the fluid medium, the interface acquires a net charge negative since the number of positive charges at the interface is less than that of negative charges [18]. It modifies, by attraction/repulsion, the random distribution created by the thermal movement of the

ions in the solution of the apoplast. The result is a dynamic electric double layer (EDL) in the region near the surface of the membrane [19] (Figure 1).



**Figure 1.** Schematic representation of the electric double layer (EDL) as a dynamic structure on the surface of a charged cell wall or membrane of a living cell, nanoparticle (NP) or a nanomaterial (NM) when it is exposed to a fluid. The term EDL refers to two parallel layers of charge on the surface. The Stern layer (with a positive net charge), consists of ions adsorbed onto the surface due to chemical interactions. The diffuse layer (with a negative net charge) is composed of ions attracted to the surface charges of the Stern layer via the Coulomb force. The diffuse layer is made of free ions that move in the fluid under the influence of electric attraction and thermal motion. The Debye length is the thickness of the mobile ions of the EDL and marks the distance under the influence of the electric potential of the surface. The zeta potential is the electrical potential at the slipping plane. The volume contained under the slipping plane shows tangential molecular motion with respect to the surface. In diluted solutions (0.001 M) the Debye length is about 10 nm but decreases with the ionic strength (1 nm with 0.1 M). In plants, the Debye length is within 1–2 nm, because in biological fluids the ionic strength is often about 0.15 M [20]. Since the transmembrane domains of integral proteins can protrude from 2 to 7 nm [21], groups with positive and negative charges of proteins and recognition sites or receptors are located outside the EDL. On the other hand, the peripheral proteins associated with integral proteins are located even further away from the EDL.

The net negative charge of cell surfaces depends on the sum of negative and positive charges at the interface with the external environment. The net negative charge occurs since the number of positive charges at the interface is less than that of negative charges. The net negative charge of cell surfaces determines the interaction with the ions in solution both in the EDL of the external side

(apoplast) and in the inner side of the cell membrane. The ions in solution in the apoplast are available at the cell interface through diffusion, and their concentration and activity depend on the final balance of charges at the interface [18]. On the other hand, the number of charges per unit area (charge density), determines the electrical potential of the surface of the membranes represented by  $\psi_0$  (Figure 1) as well as the transmembrane potential that maintains the structure and functionality of the integral proteins [18,19].

Additionally, to the primary surface charges of the membrane that fundamentally make up the EDL, there are negative and positive charges resulting from the presence of integral and peripheral proteins (receptors, channels, response proteins, among others), and of the lateral flow of protons associated with proton pumps [22]. The same occurs in the cell wall, where metabolism-related and response-related proteins determine the presence of negative and positive free charges, in addition to the surface charges of structural biomolecules that make up the EDL (Figure 1). These free charges of a membrane's or cell wall's proteins, can be found outside the main volume of influence of the EDL. They offer molecular interplay sites with a mixture of negative and positive charges to biomolecules, polymers, xenotoxics, NPs, and NMs. The interactions can be of the non-specific electrostatic type, like those that supposedly occur with NPs and NMs, or recognition interactions with native receptors of surface proteins [23], both interactions being able to induce plant biostimulation [24].

### 3. The Surface Charges of Nanoparticles (NPs) and Nanomaterials (NMs) and Its Interaction with Cell Surface Charges

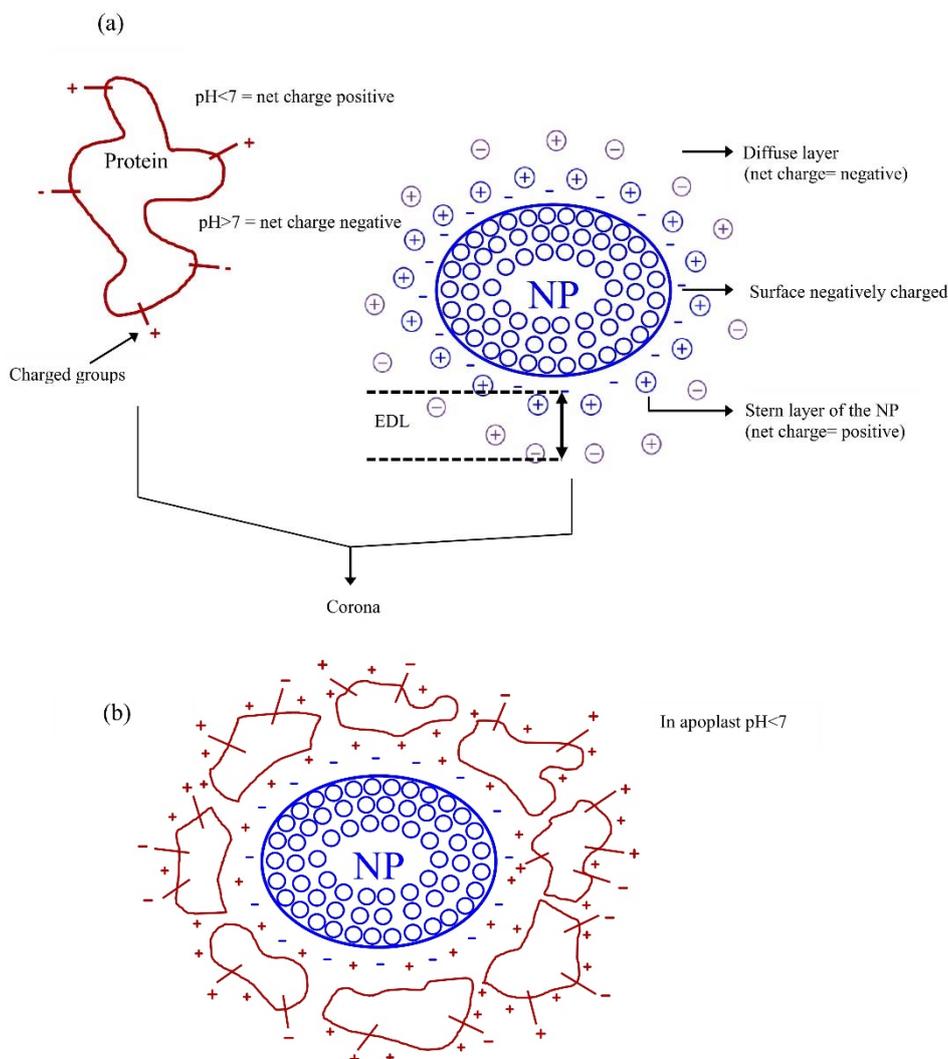
While several properties of NPs and NMs such as surface charge, surface energy, size, shape, roughness, porosity, hydrophobicity, and hydrophilicity have an impact on the interaction with cell surfaces [20,25], the emphasis in this manuscript will be focused on the charges and the charges-related energy surface of the materials. The reason is that the above enumerated set of properties of the NPs and NMs seem to modify the interactions with the cells increasing their internment in the cells [26], but it is the interaction between the surface charges of NPs–NMs–cells that initially trigger changes in metabolism and gene expression in cells [27].

Like any other material, the NPs and NMs have superficial free energy derived from the exposure of the electrons at the interface of the material and the surrounding medium. The difference with the bulk material is that the NPs and NMs, due to their high surface/volume ratio, have a much higher amount of surface free energy, which mainly explains the higher reactivity of the materials in the nanometric form [16,28,29]. This high level of superficial free energy supposes an entropy imbalance that tends to diminish spontaneously, through the interaction with other molecules or forming aggregates of NPs that precipitate. In both cases the surface exposed at the interface with the medium is decreased [30].

When the NPs or NMs are placed in a natural environment such as water, soil, epidermis, or the internal fluids of plants, they adsorb organic matter components or biomolecules (mainly proteins and peptides) from the environment, forming a corona. The composition of the corona depends initially on the most abundant biomolecules (soft corona), but over time adopts a particular composition that probably has the lowest free energy (hard corona), without necessarily including the most abundant biomolecules of the medium [31]. The corona formation is a mechanism for decreasing the surface free energy of NPs or NMs, so it happens spontaneously, regardless of whether the character of the pristine NP is cationic, anionic, or neutral [23].

The formation of the corona on a NP or NM occurs by the action of biomolecules or organic molecules that also have surface charges, forming molecular monolayers when their molecular weight is high (such as albumin) or multilayers of molecules when the molecular weight is low (such as lysozyme) [32]. The coronas of the NPs or NMs have a high density of surface charges and therefore high free energy, since it is concentrated in a small volume with large exposure area. A direct relationship has been reported between the number of proteins present in the corona and the surface charge density of the NP or NM [33,34].

The surface charge of pristine state of NPs and NMs in a natural environment is commonly negative, with a positive Stern layer and a negative diffuse layer, forming its EDL with a very high density of charges (Figure 2a) [28]. In the laboratory or during the industrial process, the original surface with negative net charge can be modified by functionalizing NPs or NMs, taking a positive, negative, or neutral character [35].



**Figure 2.** Graphical representation of formation of NP and NM corona in natural fluids. In (a) (left side) the net surface charge of the protein is positive at  $\text{pH} < 7$  (with a negatively charged Stern layer). In (a) (right side) the pristine nanoparticles (NPs) shows a surface negative charge and a positive charged Stern layer. In (b) as a result of the diffuse layers of proteins and NPs having opposite charges, the electrostatic interactions that give rise to the corona occur.

The formation of the corona depends on the chemical characteristics of the environment and on the surface charge of the chemicals used to functionalize the NPs and NMs [23]. In particular, when a corona is formed by interacting the NPs with natural fluids (with  $\text{pH}$  values around or less than 7), the cover formed by proteins or peptides will have a positive surface charge, with a negative Stern layer and a positive diffuse layer, resulting in a corona with an EDL of positive net surface charge [36] (Figure 2b), whose magnitude will change with the values of  $\text{pH}$  and ionic strength of the solution. The corona composition is modified as long as NPs are transferred from one medium to another, for example, from an abiotic to a biotic environment (and vice versa) or from an extracellular to an intracellular environment [37]. Even in structurally and chemically identical NMs and NPs,

the structure of the different proteins in the corona determine the cellular receptors and components with which the NPs interact [23] since the NPs express functional epitopes depending on the type of corona [38].

The positive surface charge of the proteins in the corona have a negative Stern layer (contrary to the positive Stern layer of the cell surfaces), which probably will dictate most of the interactions with both the ions and biomolecules of the apoplast and the initial attraction with the EDL of the cell surfaces. The initial interactions do not prohibit the interaction with anionic or cationic components of cell walls or membranes through positive and negative charge groups located on the surface of the corona [23] or in the core of the NP or NM that is not entirely covered by the corona [39].

The composition of the corona dictates to a large extent the capacity of binding with the receptor sites of the cell wall and cell membrane [40]. The interaction between the corona of NPs and NMs and cell surfaces is a result of the decrease in free surface energy of the corona when attaching to the components of the cell wall or membrane [20].

In the proteins that form the corona, the net surface charge that interacts with the EDL of NPs and NMs depends on the isoelectric point of each protein. In eukaryotes, the isoelectric point of proteins varies in a pH range of 6 to 8 [41], which partially explains the dynamic character of the corona and its different composition in different biological media, since the pH can vary with values from 5 to 8 in the different cell compartments [42]. The different composition of the coronas of NPs creates a specific chemical identity, constituting a factor that adds complexity to the impact of NPs in plants. According to the characteristics of the corona, the same type of NPs can produce different cellular responses, from biostimulation to toxicity [40].

In addition to the above, the surface charges of metallic NPs form a very reactive electronic plasma that, in addition to interacting with the biomolecules of the medium, also do so with photons, creating plasmons which under certain conditions trigger cellular responses [43].

Moving now to the topic of cell wall and membrane surface charges, the amount of electric charges per unit area is the density of charges ( $\sigma = q/A$ ;  $q$  = charge in Coulombs (C),  $A$  = area). The number of binding sites with a negative charge on the surface of a membrane was established with a value of  $0.3074 \mu\text{mol m}^{-2}$ , while the number of neutral sites (phospholipids) was determined with a value of  $2.4 \mu\text{mol m}^{-2}$ . The density of charges on the surface of a plant cell membrane was estimated with a value of  $\sigma_0 = -30 \text{ mC m}^{-2}$  [18].

The density of charges modifies the cellular behavior against ions by changing the electrical potential of the surface of the membranes ( $\psi_0$ ) as well as the transmembrane potential that keeps the ion transport channels and other integral proteins functional [18]. The activity of the  $M$  charged  $Z$  ions on the surface of the membrane and the cell wall  $\{M^Z\}_0$  is described regarding the activity of the ions in the surrounding solution  $\{M^Z\}$  and  $\psi_0$  using the equation of Nernst:

$$\{M^Z\}_0 = \{M^Z\} \exp\left[\frac{-Z_1 F \psi_0}{RT}\right] \quad (1)$$

For example, a value of  $\psi_0 = -59.2 \text{ mV}$  will increase (compared to the contiguous solution) the activities of monovalent cations on the cell surface by 10 times, while for divalent and trivalent cations it will increase by 100 and 1000 times, respectively. The anions will show the opposite behavior. Thus, plant responses to ions, including intracellular functions such as electron transport in chloroplasts and mitochondria, will show little correlation with  $\{M^Z\}$  and high correlation with  $\{M^Z\}_0$  [18].

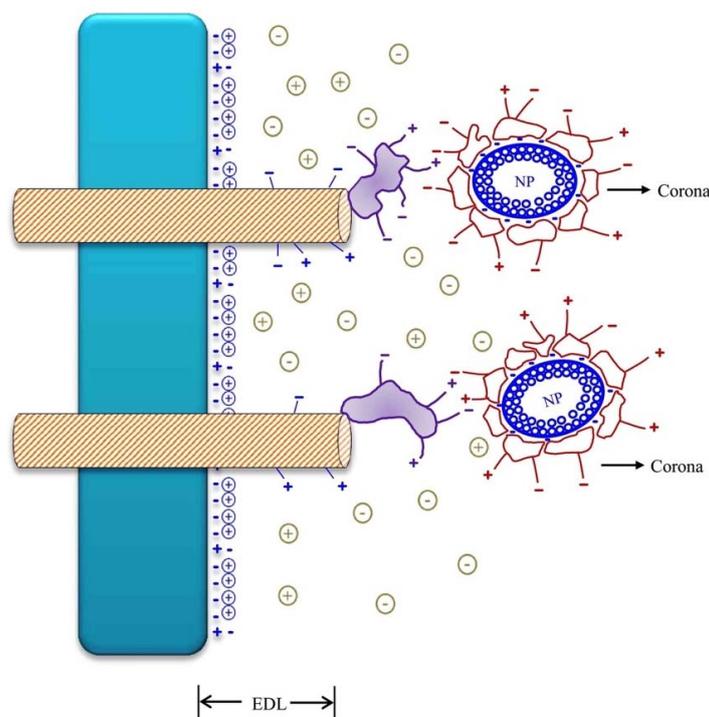
Changes in  $\psi_0$  are dynamic processes dependent on cellular metabolism and the composition of the cellular environment. The presence of cations reduces the negativity of  $\psi_0$ , reducing the absorption and impact of other cations, especially those of toxic heavy metals, the order of effectiveness being:  $\text{Al}^{3+} > \text{H}^+ > \text{Cu}^{2+} > \text{Ca}^{2+} \approx \text{Mg}^{2+} > \text{Na}^+ \approx \text{K}^+$ . The impact on anions is the opposite order. Although the protective effect of some cations such as  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$ , and  $\text{H}^+$  has been interpreted as a result of the competition for ligands in the membranes, an alternative interpretation is that the impact of protective cations also comes from effects associated with surface charges and changes in  $\psi_0$  [44].

The changes in  $\psi_0$  impacts on the activity of specific receptors, channels, transporters, or in the dynamics of exosomes that are involved in the import of hormones and other growth regulators, the ions of essential elements such as  $\text{NO}_3^-$ ,  $\text{K}^+$  and  $\text{SO}_4^{2-}$ , carbohydrates, lipids, lipoproteins, and the set of biomolecules necessary for growth and development [45,46]. Alterations in  $\psi_0$  will, therefore, have a significant effect on cellular metabolism.

The distribution of the charges on the surfaces of cell walls and cell membranes is heterogeneous at a scale of 10–50 nm due to the presence of proteins and other structures [20]. On the other hand, the distribution of the ions at the interface and in the solution with which the cell surface is in contact is considered as a product of thermal diffusion [18] (Figure 1). Therefore, the surrounding solution with ions in solution is a diffuse and homogeneous source of charges that interact with heterogeneous cell walls and membranes.

NPs and NMs with a positive net surface charge or with exposed positively or negatively charged groups are able to interact with the negative surface charges of the wall and cell membranes, as well as with the external negative and positive charges of integral and peripheral proteins, without necessarily intervening a specific cellular receptor [44] (Figure 3). In the absence of a corona, the net surface charge of a pristine NP or NM is commonly negative, as is that of cell surfaces. The negative surfaces mean that the repulsion between the EDL of the cell walls and membranes and those of the pristine NPs and NMs would make the joining process between the two surfaces more difficult. In the absence of a corona, as in pristine NPs, the process would possibly depend on the particle's hydrophobicity as an aggregation factor to decrease surface free energy in NPs and NMs [10,20].

Concerning the interaction with cell surfaces, NPs and NMs constitute a different case from that of ions [3]. It is assumed that the NPs and NMs are mobilized towards the cell surface by diffusion as it happens with the ions, but they do not seem to have specialized receptors. Therefore, the initial interaction with the cells seems to occur through attraction and complementation phenomena between the charged groups of the corona proteins and the peripheral and integral proteins that protrude from the EDL of the cell surfaces [21,47].



**Figure 3.** Graphical representation of the interaction of charges group of proteins of corona, cell wall or membrane.

The temporal magnitude of the interactions of the charges depends on the stability of the NPs and NMs, related to the composition of the corona. This later constitutes the source of surface charges of an NP or NM. A first step of the cell–surface interaction NP occurs through the union of the groups of integral or peripheral proteins with the proteins of the corona, a union that probably occurs between groups with opposite charges. A second step of this interaction apparently occurs by the action of the free energy of the surface of the corona and its hydrophobicity. High free energy and hydrophobic groups tend spontaneously to decrease the surface exposure to water molecules of the apoplast and bind with cell surfaces [20].

The second step results in the union of the corona of the nanoparticle with the cell surface, extensively disturbing (a) the electrical potential of the surfaces of the membranes, (b) the transmembrane potential and (c) the activity of the ions associated with the Stern layer and the diffuse layer.

The changes in the electrical properties of cell surfaces occur because, unlike the ions in the EDL, the NPs and NMs are not diffuse and homogeneous sources of charges. Due to their surface energy and density, surface charges are heterogeneous sources with a high concentration of charges in a small volume. It is probable that, due to this last characteristic, the impact of the NPs and NMs on cellular metabolism will be different and higher than that exerted by the diffuse charges of ions in solution, which could partially explain the observation that small amounts of NPs or NMs exert significant effects on the plants [48].

The final result of the changes in the electrical properties of cell surfaces is the modification of the activity of integral proteins like receptors or channels that unchains metabolic adjustments and changes in the gene expression as the consequence of the perceived signals, which will give rise to a new cellular phenotype and a biostimulation response [13].

NPs and NMs are likely to induce biostimulation due to having a very high surface/volume value and a significant surface charge density. As non-diffuse source of charges, it can modify the electric and transmembrane potentials by the interaction with the surface charges of the cell wall and cell membranes. The strong surface interactions could explain the cellular impact of carbon NMs [29] and, in the case of metallic elements such as Cu, Fe, Ce, Ti, and Ag, the cellular responses so different induced by the ionic forms in comparison with the nanometric structures [13,16].

As an additional point, the high surface free energy of the NPs and NMs can induce, in later stages of the interaction, cellular signaling through mechanisms other than the surface charges of membranes and cell walls. It has been proposed that NPs and NMs and their aggregated forms can block the transit of ions and metabolites at cell walls and membranes [13,29]. Under conditions of high concentration of NPs this blockade would partially explain the toxicity of NPs [13]. Other mechanisms that possibly induce significant changes in cellular behavior would be the conformational changes induced by interactions between NMs and enzymes [11], and the mechanical stimulation of NP or NM [10,49], that could lead to a mechanoreceptor-dependent response located in cell walls or membranes. Indeed, the corona proteins, due to the surface free energy of NPs or NMs, can adopt abnormal folding states [20], which can trigger defense responses of the damage associated molecular patterns (DAMPs) type [5]. An example would be the unfolded protein response (UPR) pathway, which occurs in the endoplasmic reticulum but is activated by receptors located in cell membranes and activated in the presence of proteins damaged by salinity or high temperatures [50].

The result of the processes mentioned above would be the extensive modification of the activity of the receptor, transporter and signaling proteins of the membranes, with impacts on metabolism and gene expression. This physicochemical biostimulation phenomenon resulting from surface interactions is expected to occur with any type of NP or NM, regardless of their chemical composition. It is assumed that, under a specific range of concentrations, which depends on the particular type of NPs or NMs and their properties, the initial stimulus derived from the interaction of surface charges, triggers a positive response in the cellular metabolism or biostimulation.

The organic NPs with surfaces modified to have a cationic character like poly(amidoamine) dendrimers, show more significant interaction with cell surfaces than the anionic NPs [51], disturbing with greater intensity the  $\psi_0$  and causing signaling events that modify metabolism and gene expression. However, it should be noted that, despite their negative net charge, anionic NPs like cyanoacrylic NPs [51] also interact with membranes and cell walls, possibly through the charged groups of the integral and peripheral proteins of the membrane and by the participation of other properties of NPs such as hydrophobicity [26]. An example is the NPs of polystyrene, that shows a different impact on the cells according to the surface groups and charges, observing greater toxicity in those with superficial amino groups (with cationic character) in comparison with those containing carboxyl groups (with anionic character) or had unmodified surface [52]. Although the protein corona of the polystyrene NPs adopts a negative net charge, the most effective interaction with cell surfaces occurs through scavenger receptors that bind a variety of ligands when the pristine NP was of a cationic nature, whereas with the anionic pristine NPs the interaction occurs through specific native receptors [23,53]. In the polymeric NPs of chitosan, it has been found that the absorption rate and fate in the cell are related to the surface charge, with the anionic and neutral NPs with less cellular uptake being located in the lysosomes, while the cationic ones showed more quickly internalization with a lysosome and perinuclear location. The differential locations of chitosan NPs causes different cell responses [54]. In other NPs like dextran-coated superparamagnetic iron oxide nanoparticles differential cellular locations or impacts occur because the corona does not entirely cover its nucleus, leaving some functional groups exposed and active to interact with cell surfaces [39].

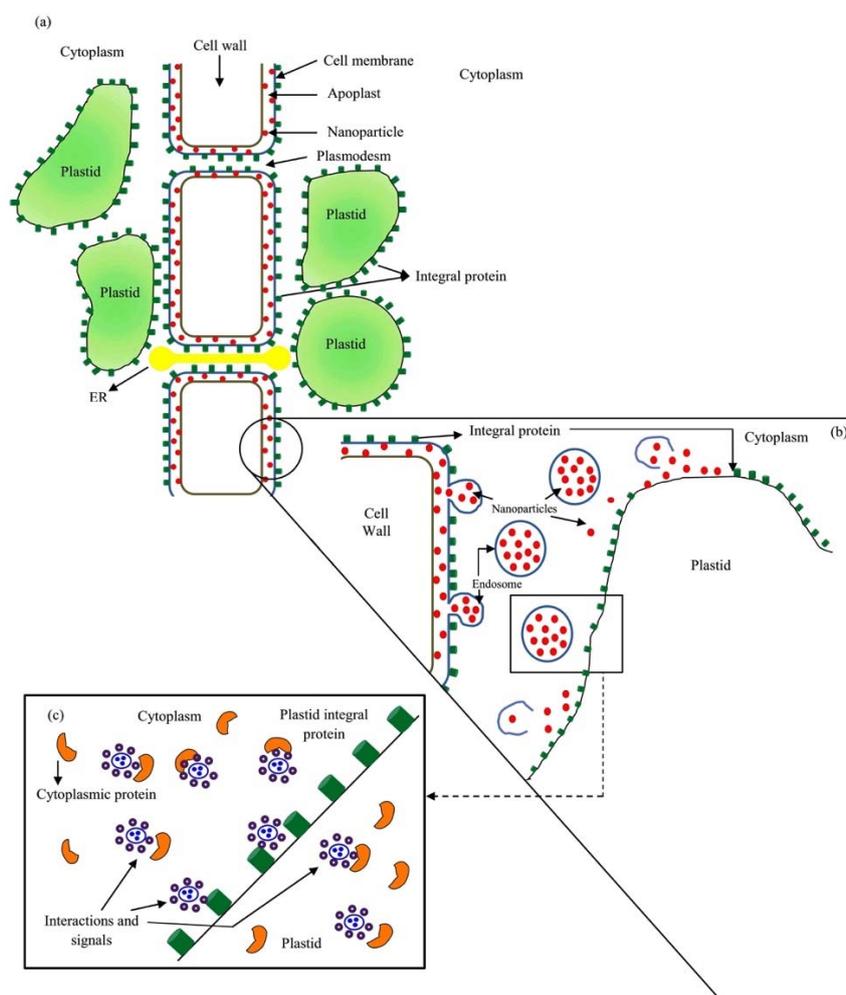
After the primary response to the changes in surface charges, a second series of biochemical stimuli are triggered by the entry of the NPs or by damage to the membranes or integral proteins that may occur in parallel with the internment. The primary and secondary responses will positively modify the cellular metabolism, causing biostimulation in the plant organism [14,55]. The process proposed above is schematized in the Figure 4.

In the metallic NPs, after the first surface interaction and cell signaling, the NPs internalized into the cell produces ionic forms of the element that are transported to the cytoplasm, impacting on metabolism according to concentration. This second-phase biochemical stimulation occurs by the presence of protein cofactors like  $Zn^{2+}$  or  $Cu^{2+}$  or by induction of oxidative stress [3,10,56,57].

Internalization occurs at the cell wall–apoplast or membrane–apoplast interface. Internalization occurs through pre-existing plasmodesmata pores up to 30 nm in diameter, modified pores with larger diameter, or by induction of new pores [58,59]. Intracellular mobility is achieved through the direct internalization or by endocytosis that occurs after the interactions between the surfaces of membranes and NPs; in the case of nanoporous materials, cell adhesion to the surface can occur [20,23,59–61]. Intracellular mobility has also been described for C nanotubes and other NMs [13]. The literature indicates that the NPs and NMs in the initial stages of internment do not maintain direct contact with the cytoplasm but remain in the endosomes. However, it has been described how endosome disruption events allow the release of NPs and NMs, thus reaching the cytoplasm, the internal membrane system and the organelle and nucleus membranes [62,63].

As with any other type of biostimulant, the definition of adequate concentration is specific for each type of NP or NM [16]. As expected, the increase in the concentration of the NPs or NMs beyond a certain threshold causes a negative response [7,13,63]. In the range of concentrations from very low to high, it is possible to observe positive responses followed by negative responses, and then followed by positive again. These alternate positive–negative responses may result in part from the cumulative dose over time, by phenomena of NPs aggregation, surface interactions, and cellular responses that do not follow a linear trend [3]. The absence of linear responses is explained by the presence of receptors that respond at different thresholds of the stimulus, as described for biostimulation with ultraviolet (UV) radiation [64]. The aggregation could be the explanation of the paradox that lower concentrations of NPs and NMs can induce more toxicity than higher concentrations [65]. Likewise, the non-linearity of the responses points to the need to distinguish the initial impact of the concentration of NPs and

NMs (which we assume causes the first biostimulation response) from the subsequent cumulative impact over time that depends on the dose.



**Figure 4.** The interaction between nanoparticles or nanomaterials with cell surfaces. (a) Binding occurs to the cell wall or the membrane through to the free energy or charged groups of the surface of the corona surrounding the NP or NM, as well as to hydrophobicity phenomena that causes aggregation between surfaces. (b) Subsequently to the binding to the cell wall or membrane, the NP or NM is admitted to the cytoplasm due to direct internalization, endocytosis phenomena, or pore formation. (c) Once the NP is internalized to the cytoplasm, interactions with cytoplasmic proteins, internal membranes or organelles occur, which causes a series of metabolic adjustments and changes in gene expression that are signaled to other cells located in another tissues or organs of the plant. In this phase, it can happen that the interaction of the NP and the cellular medium release components (such as  $\text{Cu}^{2+}$  from a copper NP) that in turn trigger other cellular responses.

#### 4. Responses of Plants to NPs and NMs

It is not the purpose of this manuscript to present a compilation of plant responses to NPs and NMs. The reader can refer to the works of [10,12,13,29,66–68], among others, to review the subject. Instead, the intention in this part of the manuscript is to present the information available in the literature about particular NPs or NMs. The purpose is to show that their use in specific concentrations induces, supposedly through interaction phenomena of charges between surfaces, changes in gene expression and positively impact on metabolism and growth.

As the first biostimulatory interaction between cells and NPs and NMs appears to be physicochemical, it is therefore independent of the chemical composition of the material and depends

more on its surface properties. That is, it occurs whether the NPs or NMs are made up of elements essential for plant nutrition or if they include other elements or even engineered structures such as carbon nanotubes or graphene. This complex phenomenon has been extensively studied in the case of NPs and NMs for medical use [3], although there is less information published in the case of plant cells, where the presence of a cell wall imposes a fundamental difference concerning stimulus-response dynamics.

In the case of NPs of essential elements, the initial response of biostimulation is dependent on the interaction of the surfaces that occurs with concentrations lower than those necessary for the element to cover or complement the nutritional needs of the plant [7,69]. According to [3] it is possible that responses to NPs and NMs can occur even in nanomolar concentration ranges, whereas [70,71] described favorable responses of the plants to micromolar concentrations of Cu NPs applied in chitosan hydrogels.

In a second phase of the biostimulatory process, the NPs of essential elements would provide ions that, when incorporated into the metabolism, originate the classical nutritional responses described for this category of elements. By increasing concentration of NPs, as with any biostimulant, the response goes from positive to neutral or negative. On the other hand, in the segment of concentrations below those that induce a positive effect the response is generally neutral in the first phase of the interaction between NPs and NMs, so that it could be classified as a hormetic response [3,14]. This type of response was described in tomato using NiO NPs [63] and in rice applying ZnO NPs [72].

Almost all the literature about the impacts of NPs and NMs on plants refers to toxicity effects. Many studies were designed using concentrations of NPs and NMs greater than those used, for example, for trace elements such as Fe, Zn, Mn, and Cu ( $<3 \text{ mg L}^{-1}$  or  $<0.05 \text{ mmol}$ ) in a nutrient solution. Another example would be graphene class materials, where concentrations of 500 to 2000  $\text{mg L}^{-1}$  are toxic to plants [73], at 50–100  $\text{mg L}^{-1}$  induces adaptation responses to stress as a higher concentration of the abscisic acid (ABA) hormone [74], whereas when applied to soil in low concentrations it promotes the absorption of water by the roots of plants [29,75]. However, the increasing agricultural use of NPs and NMs has added publications where the positive effects are explored using concentrations below those expected to be toxic [76]. This fact allowed to collect examples with NPs of  $\text{TiO}_2$  and Cu in which responses that can be described as biostimulation are indicated, possibly obtained through the action of mechanisms such as those described in the previous parts of this manuscript.

#### 4.1. Nanoparticles of Titanium Dioxide ( $\text{TiO}_2$ )

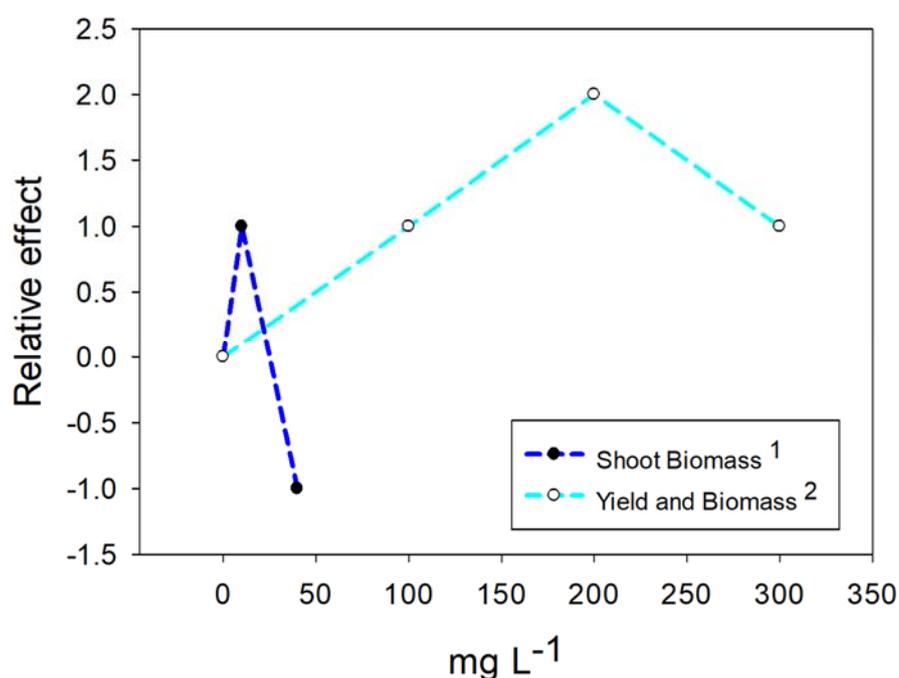
The nanomaterials of  $\text{TiO}_2$  have diverse uses such as solar cells, sensor devices, paints, and photo-catalysis.  $\text{TiO}_2$  exists in three forms: rutile (bulk), anatase and brookite. Most methods of synthesis and separation favor the more water-soluble and high surface-energy anatase structure over the lipophilic rutile structure [77]. The  $\text{TiO}_2$  NPs are among the most used in the industry, and it is estimated that 0.13 to 1200  $\mu\text{g}$  of  $\text{TiO}_2$  NPs per kg of soil are incorporated each year [78]. Experimental tests to verify toxicity, transfer to ecosystems and their use as promoters of plant metabolism have also increased in number in recent years [79].

Ti is not an element considered essential for plants, but different reports point to it as a beneficial element for plants both when it is in soil and applied in foliar sprays [80]. In the soil, the Ti is relatively abundant and reaches 30  $\text{mg L}^{-1}$  in the soil solution [78], so that the toxicity in the plants is reached with relatively high concentrations in the tissues, around  $10^{-4} \text{ M}$  [81].

Taking into account the high toxicity threshold of Ti on plants, the effects of  $\text{TiO}_2$  NPs on plants are perhaps mainly the result of surface interaction phenomena. The release of  $\text{Ti}^{2+}$ , which could be beneficial for the plants, seems to be a mechanism that contributes less to the response considering that  $\text{TiO}_2$  is a poorly soluble compound in water [82]. The NPs of  $\text{TiO}_2$  have been applied as promoters of antioxidant activity and increased tolerance to different stresses: cold [83], heat [84], drought [80,85–87], NaCl [88], and ultraviolet (UV) light [89], in concentrations ranging from 5 to 2500  $\text{mg L}^{-1}$  in plants of different species. Senescent seeds were treated with concentrations from 2500 to even 40,000  $\text{mg L}^{-1}$ ,

with the best results at  $2500 \text{ mg L}^{-1}$ . In the latter case it is very likely that the  $\text{TiO}_2$  NPs have improved the germinative response through their capacity to increase the metabolism of the seed reserves and the transport of water to the internal tissues, as reported for the NPs of Ag [90] and graphene [91].

In experiments without application of stress,  $0.01$  to  $100 \text{ mg L}^{-1}$  NPs of  $\text{TiO}_2$  were used, finding toxicity on flax plants with all concentrations [65]. Higher concentrations (from  $300$  to  $2500 \text{ mg L}^{-1}$ ) have been applied in spinach, benefiting the plants through the photocatalytic and photoprotective capacity of the  $\text{TiO}_2$  NPs [78,92,93]. In all the cases described, factors such as plant species, characteristics of the NPs and the conditions of the growth environment were decisive in the responses. However, a constant in all the works was that the positive responses were achieved with the lowest concentrations of  $\text{TiO}_2$ . Figure 5 represents the results obtained on dragonhead and wheat plants by [85] and [80], respectively. It shows the response to low concentrations for the biomass variables of the plant, which constitute the physiological summary of the plant facing the stimulation of the NPs. It is possible to see that in both studies there is an intermediate concentration with the most significant positive impact, while the higher concentration resulted in lower biomass and yield.



**Figure 5.** The relative effects of  $\text{TiO}_2$  NPs application on plants. On the Y axis the zero number represents the control treatment, a positive value represents a positive effect of the NPs, and a negative value represents an adverse effect on the specified variable. The superscript indicates the reference: dragonhead plants 1 [85], and wheat plants 2 [80].

Responses such as those presented in Figure 5, when not referring to essential elements, are considered as hormesis [14] and are characteristic of biostimulants that, according to their agricultural definition [4] induce greater growth without providing nutrients to the plants. Taking into account that the  $\text{TiO}_2$  NPs do not provide any nutrient, in addition to causing responses in plants from levels as low as  $0.01 \text{ mg L}^{-1}$ , it can be assumed that they are acting in accordance with the accepted definition of biostimulant. The action would possibly occur through interaction between the surfaces of NPs and cells in a first phase, and later through aggregation phenomena that modify the response and decrease the toxicity, especially for high concentrations [3,65].

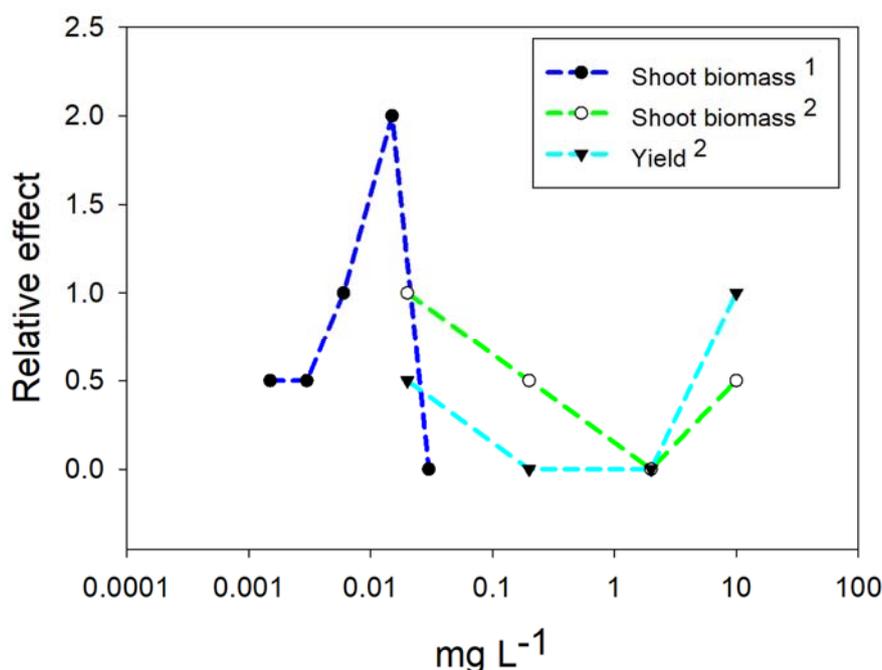
#### 4.2. Nanoparticles of Copper (Cu)

Cu nanoparticles are used in industry for their catalytic, optical, electrical, and antimicrobial characteristics. In particular, this latter antimicrobial capacity makes Cu NPs, together with those of

Ag, an attractive agent for use in clothing and footwear and textiles, food and water packaging and, in general, as a broad spectrum antimicrobial agent [94]. Unlike Ti, the toxicity threshold of Cu is reached at low concentrations. In a nutrient solution, the recommended amount of copper is  $0.026 \text{ mg L}^{-1}$ , reaching toxic effects in a range of  $0.06\text{--}0.32 \text{ mg L}^{-1}$  depending on the plant species [95]. The Cu and CuO NPs in suspension release tiny amounts of  $\text{Cu}^{2+}$  ions (approximately 0.5%, equivalent to  $5 \text{ mg L}^{-1}$  of  $\text{Cu}^{2+}$  per  $1000 \text{ mg L}^{-1}$  of NPs of CuO) that are supposed to act in conjunction with the interactive effect of the surfaces of NPs and cells. The concentration threshold for adverse impact on plant metabolism appears located at  $12\text{--}64 \text{ mg L}^{-1}$  NPs of CuO. However, it was observed that even  $10 \text{ mg L}^{-1}$  of CuO NPs substantially decreased plant growth, showing significant DNA damage caused by the production of a significant amount of free radicals [96].

Foliar spraying is a practice of NPs and NMs application that substantially decreases the toxicity on plants thanks to the chemical interactions between the components of the leaf cuticle and the applied materials [97]. The NPs of Cu or CuO have been applied by foliar application with two localized applications using a suspension with  $250 \text{ mg L}^{-1}$  of Cu NPs to decrease the risk of toxicity [98]. This concentration of Cu NPs possibly would provide  $1.25 \text{ mg L}^{-1}$  of  $\text{Cu}^{2+}$ . On the other hand, by applying CuO NPs ( $50, 100, \text{ and } 200 \text{ mg L}^{-1}$ ) by leaf spraying, mixed results were obtained, increasing the fruit production per plant, but decreasing some quality characteristics [99]. Another alternative is to apply to the substrate the Cu NPs in the form of complexes with biocompatible polymers, with positive effects at a concentration of  $0.0015 \text{ mg L}^{-1}$  up to  $1 \text{ mg L}^{-1}$  in tomato and *Vigna radiata* [71,100], equivalent to  $0.02$  to  $10 \text{ mg}$  of Cu NPs  $\text{plant}^{-1}$  in tomato plants [48,70]. Figure 6 shows the results obtained on the biomass and yield of the tomato plants when applying the substrate Cu NPs encapsulated in biocompatible polymers.

The definition of biostimulant does not include fertilizers with salts that directly contribute  $\text{Cu}^{2+}$  or  $\text{Cu}^+$  or other essential elements [4]. However, Cu or CuO NPs are not materials that directly contribute Cu and their application in the form of suspension releases approximately 0.5% of ionic Cu [96].



**Figure 6.** Relative effects of Cu NPs application on tomato plants. On the Y axis, zero represents the control treatment, a positive value represents a positive effect on characteristic specified. 1 [71]; 2 [70].

On the other hand, in Figure 6 the concentrations of Cu NPs added to the medium are very low, with positive responses of the plants being observed at levels, for example  $0.01 \text{ mg L}^{-1}$  of Cu NPs,

with what would expect nanomolar concentrations of  $\text{Cu}^{2+}$ , or approximately 500 times less ionic copper than the recommended for a nutrient solution. It is very likely that the biostimulation response observed is primarily due to the surface interaction phenomena of the NPs [3] followed by a possible effect of hormesis caused by the presence of  $\text{Cu}^{2+}$  in minimal amounts [14].

## 5. Conclusions

The surface charges of NPs and NMs interact with the surface charges of plant cells, inducing plant responses from biostimulation to toxicity.

When applied in small amounts by foliar spray or in nutrient solutions, NPs and NMs positively change the metabolism, composition and nutraceutical quality of food crops, as well as their tolerance to stress.

Biostimulation by NPs and NMs occurs in a specific range of concentrations that depends on the particular type of NPs or NMs and their properties. Biostimulation seems to occur in two stages. The initial phase, of a physicochemical nature, is the result of the interaction of surface charges; and the second phase results from the series of biochemical stimuli triggered by the entry of NPs and NMs or by the alterations in the membranes or integral proteins that can occur in parallel with the cellular internment.

Taking into account that the first phase of biostimulation seems to be physicochemical, it is concluded that virtually any NP or NM is capable of inducing biostimulation as long as it is applied in plants with the appropriate concentrations and physicochemical characteristics.

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## References

1. Vygovsky, Y.N.; Malov, A.N. *The Physics of Laser Biostimulation*; JSC MILTA-PKP GIT: Moscow, Russia, 2003.
2. Cabrera-De la Fuente, M.; González-Morales, S.; Juárez-Maldonado, A.; Leija-Martínez, P.; Benavides-Mendoza, A. Plant nutrition and agronomic management to obtain crops with better nutritional and nutraceutical quality. In *Therapeutic Foods*; Holban, A.M., Grumezescu, A.M., Eds.; Academic Press: London, UK, 2018; pp. 99–140. ISBN 978-0-12-811517-6.
3. Bell, I.R.; Ives, J.A.; Jonas, W.B. Nonlinear effects of nanoparticles: Biological variability from hormetic doses, small particle sizes, and dynamic adaptive interactions. *Dose Resp.* **2014**, *12*, 202–232. [[CrossRef](#)] [[PubMed](#)]
4. Du Jardin, P. Plant biostimulants: Definition, concept, main categories and regulation. *Sci. Hortic.* **2015**, *196*, 3–14. [[CrossRef](#)]
5. Brown, P.; Saa, S. Biostimulants in agriculture. *Front. Plant Sci.* **2015**, *6*, 671. [[CrossRef](#)] [[PubMed](#)]
6. Judy, J.D.; Bertsch, P.M. Bioavailability, toxicity, and fate of manufactured nanomaterials in terrestrial ecosystems. *Adv. Agron.* **2014**, *123*, 1–64. [[CrossRef](#)]
7. Juárez-Maldonado, A.; González-Morales, S.; Cabrera-De la Fuente, M.; Medrano-Macías, J.; Benavides-Mendoza, A. Nanometals as promoters of nutraceutical quality in crop plants. In *Impact of Nanoscience in Food Industry*; Grumezescu, A.M., Holban, A.M., Eds.; Academic Press: London, UK, 2018; pp. 277–310. ISBN 978-0-12-811441-4.
8. Byczyńska, A. Nano-silver as a potential biostimulant for plant—A review. *World Sci. News* **2017**, *86*, 180–192.
9. Campos, E.V.R.; de Oliveira, J.L.; Fernandes Fraceto, L.; Grillo, R. Global market of nanomaterials and colloidal formulations for agriculture: An overview. In *Emerging Trends in Agri-nanotechnology: Fundamental and Applied Aspects*; Singh, H.B., Mishra, S., Fernandes Fraceto, L., de Lima, R., Eds.; CAB International: Wallingford, UK, 2018; p. 302. ISBN 9781786391445.
10. Seabra, A.B.; Paula, A.J.; de Lima, R.; Alves, O.L.; Durán, N. Nanotoxicity of graphene and graphene oxide. *Chem. Res. Toxicol.* **2014**, *27*, 159–168. [[CrossRef](#)] [[PubMed](#)]

11. Chen, M.; Zeng, G.; Xu, P.; Yan, M.; Xiong, W.; Zhou, S. Interaction of carbon nanotubes with microbial enzymes: Conformational transitions and potential toxicity. *Environ. Sci. Nano* **2017**, *4*, 1954–1960. [[CrossRef](#)]
12. Morales-Díaz, A.B.A.B.; Ortega-Ortíz, H.; Juárez-Maldonado, A.; Cadenas-Pliego, G.; González-Morales, S.; Benavides-Mendoza, A. Application of nanoelements in plant nutrition and its impact in ecosystems. *Adv. Nat. Sci. Nanosci. Nanotechnol.* **2017**, *8*, 013001. [[CrossRef](#)]
13. Zuverza-Mena, N.; Martínez-Fernández, D.; Du, W.; Hernandez-Viezcás, J.A.; Bonilla-Bird, N.; López-Moreno, M.L.; Komárek, M.; Peralta-Video, J.R.; Gardea-Torresdey, J.L. Exposure of engineered nanomaterials to plants: Insights into the physiological and biochemical responses-A review. *Plant Physiol. Biochem.* **2017**, *110*, 236–264. [[CrossRef](#)]
14. Poschenrieder, C.; Cabot, C.; Martos, S.; Gallego, B.; Barceló, J. Do toxic ions induce hormesis in plants? *Plant Sci.* **2013**, *212*, 15–25. [[CrossRef](#)]
15. Poschenrieder, C.; Tolrà, R.; Barceló, J. Can metals defend plants against biotic stress? *Trends Plant Sci.* **2006**, *11*, 288–295. [[CrossRef](#)] [[PubMed](#)]
16. Hillegass, J.M.; Shukla, A.; Lathrop, S.A.; MacPherson, M.B.; Fukagawa, N.K.; Mossman, B.T. Assessing nanotoxicity in cells in vitro. *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.* **2010**, *2*, 219–231. [[CrossRef](#)] [[PubMed](#)]
17. Knox, J.P.; Benitez-Alfonso, Y. Roles and regulation of plant cell walls surrounding plasmodesmata. *Curr. Opin. Plant Biol.* **2014**, *22*, 93–100. [[CrossRef](#)]
18. Kinraide, T.B.; Wang, P. The surface charge density of plant cell membranes (sigma): An attempt to resolve conflicting values for intrinsic sigma. *J. Exp. Bot.* **2010**, *61*, 2507–2518. [[CrossRef](#)] [[PubMed](#)]
19. Wiley, D.; Weihs, G.F. Surface charge density. In *Encyclopedia of Membranes*; Springer: Berlin/Heidelberg, Germany, 2016; pp. 1864–1866.
20. Nel, A.E.; Mädler, L.; Velegol, D.; Xia, T.; Hoek, E.M.V.; Somasundaran, P.; Klaessig, F.; Castranova, V.; Thompson, M. Understanding biophysicochemical interactions at the nano–bio interface. *Nat. Mater.* **2009**, *8*, 543–557. [[CrossRef](#)] [[PubMed](#)]
21. Milo, R.; Phillips, R. *Cell Biology by the Numbers*; Garland Science: New York, NY, USA, 2015; ISBN 978-0-8153-4163-5.
22. Yoshinaga, M.Y.; Kellermann, M.Y.; Valentine, D.L.; Valentine, R.C. Phospholipids and glycolipids mediate proton containment and circulation along the surface of energy-transducing membranes. *Prog. Lipid Res.* **2016**, *64*, 1–15. [[CrossRef](#)]
23. Fleischer, C.C.; Payne, C.K. Nanoparticle-cell interactions: Molecular structure of the protein corona and cellular outcomes. *Acc. Chem. Res.* **2014**, *47*, 2651–2659. [[CrossRef](#)]
24. Canellas, L.P.; Olivares, F.L.; Aguiar, N.O.; Jones, D.L.; Nebbioso, A.; Mazzei, P.; Piccolo, A. Humic and fulvic acids as biostimulants in horticulture. *Sci. Hortic.* **2015**, *196*, 15–27. [[CrossRef](#)]
25. Zhao, J.; Stenzel, M.H. Entry of nanoparticles into cells: The importance of nanoparticle properties. *Polym. Chem.* **2018**, *9*, 259–272. [[CrossRef](#)]
26. Abou Matar, T.; Karam, P. The role of hydrophobicity in the cellular uptake of negatively charged macromolecules. *Macromol. Biosci.* **2018**, *18*, 1700309. [[CrossRef](#)]
27. Chang, Y.-N.; Zhang, M.; Xia, L.; Zhang, J.; Xing, G. The toxic effects and mechanisms of CuO and ZnO nanoparticles. *Materials* **2012**, *5*, 2850–2871. [[CrossRef](#)]
28. Fischer, F.D.; Waitz, T.; Vollath, D.; Simha, N.K. On the role of surface energy and surface stress in phase-transforming nanoparticles. *Prog. Mater. Sci.* **2008**, *53*, 481–527. [[CrossRef](#)]
29. Chen, M.; Zhou, S.; Zhu, Y.; Sun, Y.; Zeng, G.; Yang, C.; Xu, P.; Yan, M.; Liu, Z.; Zhang, W. Toxicity of carbon nanomaterials to plants, animals and microbes: Recent progress from 2015-present. *Chemosphere* **2018**, *206*, 255–264. [[CrossRef](#)] [[PubMed](#)]
30. Loosli, F.; Vitorazi, L.; Berret, J.-F.; Stoll, S. Towards a better understanding on agglomeration mechanisms and thermodynamic properties of TiO<sub>2</sub> nanoparticles interacting with natural organic matter. *Water Res.* **2015**, *80*, 139–148. [[CrossRef](#)] [[PubMed](#)]
31. Docter, D.; Westmeier, D.; Markiewicz, M.; Stolte, S.; Knauer, S.K.; Stauber, R.H. The nanoparticle biomolecule corona: Lessons learned—Challenge accepted? *Chem. Soc. Rev.* **2015**, *44*, 6094–6121. [[CrossRef](#)] [[PubMed](#)]
32. Aramesh, M.; Shimoni, O.; Ostrikov, K.; Prawer, S.; Cervenka, J. Surface charge effects in protein adsorption on nanodiamonds. *Nanoscale* **2015**, *7*, 5726–5736. [[CrossRef](#)]

33. Gessner, A.; Lieske, A.; Paulke, B.R.; Müller, R.H. Influence of surface charge density on protein adsorption on polymeric nanoparticles: Analysis by two-dimensional electrophoresis. *Eur. J. Pharm. Biopharm.* **2002**, *54*, 165–170. [[CrossRef](#)]
34. Gunawan, C.; Lim, M.; Marquis, C.P.; Amal, R. Nanoparticle–protein corona complexes govern the biological fates and functions of nanoparticles. *J. Mater. Chem. B* **2014**, *2*, 2060. [[CrossRef](#)]
35. Mout, R.; Moyano, D.F.; Rana, S.; Rotello, V.M. Surface functionalization of nanoparticles for nanomedicine. *Chem. Soc. Rev.* **2012**, *41*, 2539. [[CrossRef](#)]
36. Gitlin, I.; Carbeck, J.D.; Whitesides, G.M. Why are proteins charged? Networks of charge–charge interactions in proteins measured by charge ladders and capillary electrophoresis. *Angew. Chem. Int. Ed.* **2006**, *45*, 3022–3060. [[CrossRef](#)]
37. Lundqvist, M.; Stigler, J.; Cedervall, T.; Berggård, T.; Flanagan, M.B.; Lynch, I.; Elia, G.; Dawson, K. The evolution of the protein corona around nanoparticles: A test study. *ACS Nano* **2011**, *5*, 7503–7509. [[CrossRef](#)]
38. Monopoli, M.P.; Walczyk, D.; Campbell, A.; Elia, G.; Lynch, I.; Bombelli, F.B.; Dawson, K.A. Physical-chemical aspects of protein corona: Relevance to in vitro and in vivo biological impacts of nanoparticles. *J. Am. Chem. Soc.* **2011**, *133*, 2525–2534. [[CrossRef](#)] [[PubMed](#)]
39. Rahman, M.; Laurent, S.; Tawil, N.; Yahia, L.; Mahmoudi, M. Nanoparticle and protein corona. In *Protein–Nanoparticle Interactions. The Bio–Nano Interface*; Rahman, M., Laurent, S., Tawil, N., Yahia, L., Mahmoud, M., Eds.; Springer: Berlin/Heidelberg, Germany, 2013; pp. 21–44. ISBN 978-3-642-37555-2.
40. Jackson, T.C.; Patani, B.O.; Israel, M.B. Nanomaterials and cell interactions: A review. *J. Biomater. Nanobiotechnol.* **2017**, *8*, 1–9. [[CrossRef](#)]
41. Kozłowski, L.P. Proteome-pI: Proteome isoelectric point database. *Nucleic Acids Res.* **2017**, *45*, D1112–D1116. [[CrossRef](#)] [[PubMed](#)]
42. Shen, J.; Zeng, Y.; Zhuang, X.; Sun, L.; Yao, X.; Pimpl, P.; Jiang, L. Organelle pH in the Arabidopsis endomembrane system. *Mol. Plant* **2013**, *6*, 1419–1437. [[CrossRef](#)] [[PubMed](#)]
43. Rejiya, C.S.; Kumar, J.; Raji, V.; Vibin, M.; Abraham, A. Laser immunotherapy with gold nanorods causes selective killing of tumour cells. *Pharmacol. Res.* **2012**, *65*, 261–269. [[CrossRef](#)]
44. Kinraide, T.B. Plasma membrane surface potential ( $\psi_{pm}$ ) as a determinant of ion bioavailability: A critical analysis of new and published toxicological studies and a simplified method for the computation of plant  $\psi_{pm}$ . *Environ. Toxicol. Chem.* **2006**, *25*, 3188–3198. [[CrossRef](#)] [[PubMed](#)]
45. Haak, D.C.; Fukao, T.; Grene, R.; Hua, Z.; Ivanov, R.; Perrella, G.; Li, S.; Verslues, P.E.; Nakashima, K.; Batelli, G.; et al. Multilevel regulation of abiotic stress responses in plants. *Front. Plant Sci.* **2017**, *8*, 1564. [[CrossRef](#)]
46. Stanley, S. Biological nanoparticles and their influence on organisms. *Curr. Opin. Biotechnol.* **2014**, *28*, 69–74. [[CrossRef](#)]
47. Alberts, B.; Johnson, A.; Lewis, J.; Raff, M.; Roberts, K.; Walter, P. *Molecular Biology of the Cell*, 4th ed.; Garland Science: New York, NY, USA, 2002; ISBN 978-0815332183.
48. Hernández-Hernández, H.; González-Morales, S.; Benavides-Mendoza, A.; Ortega-Ortiz, H.; Cadenas-Pliego, G.; Juárez-Maldonado, A. Effects of chitosan–PVA and Cu nanoparticles on the growth and antioxidant capacity of tomato under saline stress. *Molecules* **2018**, *23*, 178. [[CrossRef](#)]
49. Septiadi, D.; Crippa, F.; Moore, T.L.; Rothen-Rutishauser, B.; Petri-Fink, A. Nanoparticle–cell interaction: A cell mechanics perspective. *Adv. Mater.* **2018**, *30*, 1704463. [[CrossRef](#)] [[PubMed](#)]
50. Liu, J.-X.; Srivastava, R.; Che, P.; Howell, S.H. Salt stress responses in Arabidopsis utilize a signal transduction pathway related to endoplasmic reticulum stress signaling. *Plant J.* **2007**, *51*, 897–909. [[CrossRef](#)] [[PubMed](#)]
51. Fröhlich, E. The role of surface charge in cellular uptake and cytotoxicity of medical nanoparticles. *Int. J. Nanomed.* **2012**, *7*, 5577–5591. [[CrossRef](#)]
52. Ruenraroengsak, P.; Tetley, T.D. Differential bioreactivity of neutral, cationic and anionic polystyrene nanoparticles with cells from the human alveolar compartment: Robust response of alveolar type 1 epithelial cells. *Part. Fibre Toxicol.* **2015**, *12*, 19. [[CrossRef](#)] [[PubMed](#)]
53. Fleischer, C.C.; Payne, C.K. Nanoparticle surface charge mediates the cellular receptors used by protein–nanoparticle complexes. *J. Phys. Chem. B* **2012**, *116*, 8901–8907. [[CrossRef](#)]
54. Yue, Z.-G.; Wei, W.; Lv, P.-P.; Yue, H.; Wang, L.-Y.; Su, Z.-G.; Ma, G.-H. Surface charge affects cellular uptake and intracellular trafficking of chitosan-based nanoparticles. *Biomacromolecules* **2011**, *12*, 2440–2446. [[CrossRef](#)]

55. Wang, J. Engineered Nanomaterials and Plant Interactions: Uptake, Translocation, Transformation and Physiological Effects. Ph.D. Thesis, Rice University, Houston, TX, USA, 2014.
56. Nair, R.; Mohamed, M.S.; Gao, W.; Maekawa, T.; Yoshida, Y.; Ajayan, P.M.; Kumar, D.S. Effect of carbon nanomaterials on the germination and growth of rice plants. *J. Nanosci. Nanotechnol.* **2012**, *12*, 2212–2220. [[CrossRef](#)]
57. Wang, D.; Lin, Z.; Wang, T.; Yao, Z.; Qin, M.; Zheng, S.; Lu, W. Where does the toxicity of metal oxide nanoparticles come from: The nanoparticles, the ions, or a combination of both? *J. Hazard. Mater.* **2016**, *308*, 328–334. [[CrossRef](#)]
58. Nair, R.; Varghese, S.H.; Nair, B.G.; Maekawa, T.; Yoshida, Y.; Kumar, D.S. Nanoparticulate material delivery to plants. *Plant Sci.* **2010**, *179*, 154–163. [[CrossRef](#)]
59. Wang, P.; Lombi, E.; Zhao, F.-J.; Kopittke, P.M. Nanotechnology: A new opportunity in plant sciences. *Trends Plant Sci.* **2016**, *21*, 699–712. [[CrossRef](#)] [[PubMed](#)]
60. Verma, A.; Uzun, O.; Hu, Y.; Han, H.-S.; Watson, N.; Chen, S.; Irvine, D.J.; Stellacci, F. Surface-structure-regulated cell-membrane penetration by monolayer-protected nanoparticles. *Nat. Mater.* **2008**, *7*, 588–595. [[CrossRef](#)] [[PubMed](#)]
61. Lesniak, A.; Salvati, A.; Santos-Martinez, M.J.; Radomski, M.W.; Dawson, K.A.; Åberg, C. Nanoparticle adhesion to the cell membrane and Its effect on nanoparticle uptake efficiency. *J. Am. Chem. Soc.* **2013**, *135*, 1438–1444. [[CrossRef](#)]
62. Verma, A.; Stellacci, F. Effect of surface properties on nanoparticle-cell interactions. *Small* **2010**, *6*, 12–21. [[CrossRef](#)] [[PubMed](#)]
63. Faisal, M.; Saquib, Q.; Alatar, A.A.; Al-Khedhairi, A.A.; Hegazy, A.K.; Musarrat, J. Phytotoxic hazards of NiO-nanoparticles in tomato: A study on mechanism of cell death. *J. Hazard. Mater.* **2013**, *250–251*, 318–332. [[CrossRef](#)] [[PubMed](#)]
64. Díaz-Leyva, C.E.; Bacópulos-Mejía, E.; Ruiz-Torres, N.A.; Ibarra-Jiménez, L.; González-Morales, S.; Benavides-Mendoza, A. Irradiation of tomato seeds with Uv-B and UV-C: Impact on germination, vigor and growth. *Rev. Mex. Ciencias Agrícolas* **2017**, *8*, 103–116.
65. Clément, L.; Hurel, C.; Marmier, N. Toxicity of TiO<sub>2</sub> nanoparticles to cladocerans, algae, rotifers and plants—Effects of size and crystalline structure. *Chemosphere* **2013**, *90*, 1083–1090. [[CrossRef](#)] [[PubMed](#)]
66. Mukherjee, K.; Acharya, K. Toxicological effect of metal oxide nanoparticles on soil and aquatic habitats. *Arch. Environ. Contam. Toxicol.* **2018**, *75*, 1–12. [[CrossRef](#)] [[PubMed](#)]
67. Buzea, C.; Pacheco, I.I.; Robbie, K. Nanomaterials and nanoparticles: Sources and toxicity. *Biointerphases* **2007**, *2*, MR17. [[CrossRef](#)]
68. Capaldi Arruda, S.C.; Diniz Silva, A.L.; Moretto Galazzi, R.; Antunes Azevedo, R.; Zezzi Arruda, M.A.; Arruda, S.C.C.; Silva, A.L.D.; Galazzi, R.M.; Azevedo, R.A.; Arruda, M.A.Z.; et al. Nanoparticles applied to plant science: A review. *Talanta* **2015**, *131*, 693–705. [[CrossRef](#)] [[PubMed](#)]
69. Faizan, M.; Faraz, A.; Yusuf, M.; Khan, S.T.; Hayat, S. Zinc oxide nanoparticle-mediated changes in photosynthetic efficiency and antioxidant system of tomato plants. *Photosynthetica* **2018**, *56*, 678–686. [[CrossRef](#)]
70. Hernández-Hernández, H.; Benavides-Mendoza, A.; Ortega-Ortiz, H.; Hernández-Fuentes, A.D.; Juárez-Maldonado, A. Cu Nanoparticles in chitosan-PVA hydrogels as promoters of growth, productivity and fruit quality in tomato. *Emir. J. Food Agric.* **2017**, *29*. [[CrossRef](#)]
71. Juárez-Maldonado, A.; Ortega-Ortiz, H.; Pérez-Labrada, F.; Cadenas-Pliego, G.; Benavides-Mendoza, A. Cu Nanoparticles absorbed on chitosan hydrogels positively alter morphological, production, and quality characteristics of tomato. *J. Appl. Bot. Food Qual.* **2016**, *89*, 183–189. [[CrossRef](#)]
72. Boonyanitipong, P.; Kositsup, B.; Kumar, P.; Baruah, S.; Dutta, J. Toxicity of ZnO and TiO<sub>2</sub> Nanoparticles on germinating Rice seed *Oryza sativa* L. *Int. J. Biosci. Biochem. Bioinform.* **2011**, *1*, 4–7. [[CrossRef](#)]
73. Begum, P.; Ikhtiar, R.; Fugetsu, B. Graphene phytotoxicity in the seedling stage of cabbage, tomato, red spinach, and lettuce. *Carbon N. Y.* **2011**, *49*, 3907–3919. [[CrossRef](#)]
74. Cheng, F.; Liu, Y.-F.; Lu, G.-Y.; Zhang, X.-K.; Xie, L.-L.; Yuan, C.-F.; Xu, B.-B. Graphene oxide modulates root growth of *Brassica napus* L. and regulates ABA and IAA concentration. *J. Plant Physiol.* **2016**, *193*, 57–63. [[CrossRef](#)] [[PubMed](#)]
75. He, Y.; Hu, R.; Zhong, Y.; Zhao, X.; Chen, Q.; Zhu, H. Graphene oxide as a water transporter promoting germination of plants in soil. *Nano Res.* **2018**, *11*, 1928–1937. [[CrossRef](#)]

76. de la Rosa, G.; García-Castañeda, C.; Vázquez-Núñez, E.; Alonso-Castro, Á.J.; Basurto-Islas, G.; Mendoza, Á.; Cruz-Jiménez, G.; Molina, C. Physiological and biochemical response of plants to engineered NMs: Implications on future design. *Plant Physiol. Biochem.* **2017**, *110*, 226–235. [[CrossRef](#)]
77. Reyes-Coronado, D.; Rodríguez-Gattorno, G.; Espinosa-Pesqueira, M.E.; Cab, C.; de Coss, R.; Oskam, G. Phase-pure TiO<sub>2</sub> nanoparticles: Anatase, brookite and rutile. *Nanotechnology* **2008**, *19*, 145605. [[CrossRef](#)]
78. Lyu, S.; Wei, X.; Chen, J.; Wang, C.; Wang, X.; Pan, D. Titanium as a beneficial element for crop production. *Front. Plant Sci.* **2017**, *8*, 597. [[CrossRef](#)]
79. Mattiello, A.; Marchiol, L. Application of nanotechnology in agriculture: Assessment of TiO<sub>2</sub> nanoparticle effects on barley. In *Application of Titanium Dioxide*; Janus, M., Ed.; InTech: London, UK, 2017; pp. 23–39.
80. Jaberzadeh, A.; Moaveni, P.; Tohidi Moghadam, H.R.; Zahedi, H. Influence of bulk and nanoparticles titanium foliar application on some agronomic traits, seed gluten and starch contents of wheat subjected to water deficit stress. *Not. Bot. Horti Agrobot. Cluj-Napoca* **2013**, *41*, 201–207. [[CrossRef](#)]
81. Wallace, A.; Alexander, G.V.; Chaudhry, F.M. Phytotoxicity of cobalt, vanadium, titanium, silver, and chromium. *Commun. Soil Sci. Plant Anal.* **1977**, *8*, 751–756. [[CrossRef](#)]
82. Tlustoš, P.; Cígler, P.; Hrubý, M.; Kužel, S.; Száková, J.; Balík, J. The role of titanium in biomass production and its influence on essential elements' contents in field growing crops. *Plant Soil Environ.* **2005**, *51*, 19–25. [[CrossRef](#)]
83. Mohammadi, R.; Maali-Amiri, R.; Mantri, N.L. Effect of TiO<sub>2</sub> nanoparticles on oxidative damage and antioxidant defense systems in chickpea seedlings during cold stress. *Russ. J. Plant Physiol.* **2014**, *61*, 768–775. [[CrossRef](#)]
84. Qi, M.; Liu, Y.; Li, T. Nano- TiO<sub>2</sub> improve the photosynthesis of tomato leaves under mild heat stress. *Biol. Trace Elem. Res.* **2013**, *156*, 323–328. [[CrossRef](#)] [[PubMed](#)]
85. Mohammadi, H.; Esmailpour, M.; Gheranpaye, A. Effects of TiO<sub>2</sub> nanoparticles and water-deficit stress on morpho-physiological characteristics of dragonhead (*Dracocephalum moldavica* L.) plants. *Acta Agric. Slov.* **2016**, *107*, 385–396. [[CrossRef](#)]
86. Aghdam, M.T.B.; Mohammadi, H.; Ghorbanpour, M. Effects of nanoparticulate anatase titanium dioxide on physiological and biochemical performance of *Linum usitatissimum* (Linaceae) under well-watered and drought stress conditions. *Braz. J. Bot.* **2016**, *39*, 139–146. [[CrossRef](#)]
87. Kiapour, H.; Moaveni, P.; Habibi, D.; Sani, B. Evaluation of the application of gibberellic acid and titanium dioxide nanoparticles under drought stress on some traits of basil (*Ocimum basilicum* L.). *Int. J. Agron. Agric. Res.* **2015**, *6*, 138–150.
88. Khan, M.N. Nano-titanium Dioxide (Nano-TiO<sub>2</sub>) mitigates NaCl stress by enhancing antioxidative enzymes and accumulation of compatible solutes in tomato (*Lycopersicon esculentum* Mill.). *J. Plant Sci.* **2016**, *11*, 1–11. [[CrossRef](#)]
89. Zheng, L.; Mingyu, S.; Xiao, W.; Chao, L.; Chunxiang, Q.; Liang, C.; Hao, H.; Xiaoqing, L.; Fashui, H. Antioxidant stress is promoted by nano-anatase in spinach chloroplasts under UV-B radiation. *Biol. Trace Elem. Res.* **2008**, *121*, 69–79. [[CrossRef](#)]
90. Mahakham, W.; Sarmah, A.K.; Maensiri, S.; Theerakulpisut, P. Nanoprimering technology for enhancing germination and starch metabolism of aged rice seeds using phytosynthesized silver nanoparticles. *Sci. Rep.* **2017**, *7*, 8263. [[CrossRef](#)]
91. Zhang, M.; Gao, B.; Chen, J.; Li, Y. Effects of graphene on seed germination and seedling growth. *J. Nanoparticle Res.* **2015**, *17*, 78. [[CrossRef](#)]
92. Gao, F.; Liu, C.; Qu, C.; Zheng, L.; Yang, F.; Su, M.; Hong, F. Was improvement of spinach growth by nano-TiO<sub>2</sub> treatment related to the changes of Rubisco activase? *Biometals* **2008**, *21*, 211–217. [[CrossRef](#)] [[PubMed](#)]
93. Yang, F.; Hong, F.; You, W.; Liu, C.; Gao, F.; Wu, C.; Yang, P. Influences of nano anatase TiO<sub>2</sub> on nitrogen metabolism of growing spinach. *Biol. Trace Elem. Res.* **2006**, *110*, 179–190. [[CrossRef](#)]
94. Ingle, A.P.; Duran, N.; Rai, M. Bioactivity, mechanism of action, and cytotoxicity of copper-based nanoparticles: A review. *Appl. Microbiol. Biotechnol.* **2014**, *98*, 1001–1009. [[CrossRef](#)] [[PubMed](#)]
95. Ebbs, S.D.; Kochian, L.V. Toxicity of Zinc and Copper to Brassica species: Implications for phytoremediation. *J. Environ. Qual.* **1997**, *26*, 776–781. [[CrossRef](#)]
96. Atha, D.H.; Wang, H.; Petersen, E.J.; Cleveland, D.; Holbrook, R.D.; Jaruga, P.; Dizdaroglu, M.; Xing, B.; Nelson, B.C. Copper oxide nanoparticle mediated DNA damage in terrestrial plant models. *Environ. Sci. Technol.* **2012**, *46*, 1819–1827. [[CrossRef](#)] [[PubMed](#)]

97. Hu, J.; Guo, H.; Li, J.; Wang, Y.; Xiao, L.; Xing, B. Interaction of  $\gamma$ -Fe<sub>2</sub>O<sub>3</sub> nanoparticles with *Citrus maxima* leaves and the corresponding physiological effects via foliar application. *J. Nanobiotechnol.* **2017**, *15*, 51. [[CrossRef](#)] [[PubMed](#)]
98. Hernández-Fuentes, A.; López-Vargas, E.; Pinedo-Espinoza, J.; Campos-Montiel, R.; Valdés-Reyna, J.; Juárez-Maldonado, A. Postharvest behavior of bioactive compounds in tomato fruits treated with Cu nanoparticles and NaCl stress. *Appl. Sci.* **2017**, *7*, 980. [[CrossRef](#)]
99. Hong, J.; Wang, L.; Sun, Y.; Zhao, L.; Niu, G.; Tan, W.; Rico, C.M.; Peralta-Videa, J.R.; Gardea-Torresdey, J.L. Foliar applied nanoscale and microscale CeO<sub>2</sub> and CuO alter cucumber (*Cucumis sativus*) fruit quality. *Sci. Total Environ.* **2016**, *563*, 904–911. [[CrossRef](#)] [[PubMed](#)]
100. Pradhan, S.; Patra, P.; Mitra, S.; Dey, K.K.; Basu, S.; Chandra, S.; Palit, P.; Goswami, A. Copper nanoparticle (CuNP) nanochain arrays with a reduced toxicity response: A biophysical and biochemical outlook on *Vigna radiata*. *J. Agric. Food Chem.* **2015**, *63*, 2606–2617. [[CrossRef](#)]



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