

Figure S1. Alignment of amino acid sequences of BAS2 of *C. gloeosporioides* from *H. brasiliensis*, *C. gloeosporioides* Nara gc5, *C. higginsianum*, *C. graminicola*, *M. oryzae*, and *P. anserina*.



Figure S2. Phylogenetic analysis of BAS2 proteins. The neighbor joining phylogenetic tree was constructed by MEGA 7.0 according to the evolutionary relationship between BAS2 proteins in *C. gloeosporioides* from *H. brasiliensis, C. gloeosporioides* Nara gc5, *C. higginsianum, C. graminicola, M. oryzae,* and *P. anserina*.



Figure S3. (A) The gene deletion strategy. Putative mutants were screened with diagnostic primers, indicated by black triangles. **(B)** Confirmation of the correct recombination of deletion cassettes with gene loci by Southern blot.



Figure S4. Strategy for generation of BAS2-GFP fusion overexpressing mutants. (**A**) Expression system using the gene loci of nitrate reductase (*niaD*). Promoter of ToxA and terminator nos were used for the gene expression; the Hygromycin phosphotransferase gene (*HPH*) was used for transformant selection. (**B**) The open reading frame of *BAS2* and the coding sequence of GFP with an N-terminal linker were linked together to construct the fusion expressing vector.



Figure S5. Phenotype assays of ΔniaD. **(A)** Growth rate assay of WT and ΔniaD cultured on complete medium (CM) and minimal medium (MM). **(B)** Mean lesion diameters after inoculation with conidia for 3 days. Bars represent standard deviation (SD). **(C)** Disease symptoms of rubber-tree leaves after inoculation with conidia suspension for 3 days.



Figure S6. Two-dimensional patterns of extracellular proteomes of WT and Δ BAS2. Arrows indicate protein spots that were upregulated in abundance more than 1.5-fold between WT and Δ BAS2.

Table S1. PCR	rimers used in this study. Lowercase letters indicate induced restriction sites.

Numbe	Primer	Sequence $(5' \rightarrow 3')$	Application
r			
1	BAS2-5F	cgagctcTCGCAAAAATGTTCCAG	BAS2 deletion
2	BAS2-5R	gctctagaTTTCGCGGTAGTTGAGTG	BAS2 deletion
3	BAS2-3F	ccatcgatGCTGGAAATGCAGAAACT	BAS2 deletion
4	BAS2-3R	ggggtaccAACATGGGGCAGGAGAC	BAS2 deletion
5	BAS2-JC5F	GCGCATTCTTTGAGGTTTCTTG	Δ BAS2 diagnosis
6	HYG-JCR	TGAGTTCAGGCTTTTTCATTTGG	Δ BAS2 diagnosis
7	HYG-JCF	ACAGCGGTCATTGACTGGAGCGA	Δ BAS2 diagnosis
8	BAS2- IC3R	AAGGGCGGCGACAGTGAAGAGG	ΔBAS2 diagnosis
9	niaD-5F	gagetcAAGGAGTCCCGTTTGTT	<i>niaD</i> deletion, expression
10	niaD-5R	gcggccgcACTGACGACTGGCTTGTC	<i>niaD</i> deletion, expression system
11	niaD-3F	gtcgacACGAGCTGCCGTTTTTAG	<i>niaD</i> deletion, expression system
12	niaD-3R	ggtaccCGGTCACGACGCTGTAA	<i>niaD</i> deletion, expression system
13	niaD-JC5F	TGCCAGTAGCGTGGTTTAGGTC	∆niaD diagnosis
14	niaD-JC5R	tctagaAATTTCCCCGATCGTTC	∆niaD diagnosis
15	niaD-JC3F	ACAGCGGTCATTGACTGGAGCGA	Δ niaD diagnosis
16	niaD-JC3R	AGTGTCCCAGATGTCGTGTTGC	Δ niaD diagnosis
17	Ptoxa-F	gtcgacTGGAATGCATGGAGGAG	Expression system of C. gloeosporioides
18	Ptoxa-R	atcgatGACCTATATTCATTCAT	Expression system of C. gloeosporioides
19	Tnos-F	tctagaAATTTCCCCGATCGTTC	Expression system of C. gloeosporioides
20	Tnos-R	gcggccgcCCGATCTAGTAACATAG	Expression system of C. gloeosporioides
21	HPH-F	gtcgacAACTGATATTGAAGGAG	Expression system of C.
22	HPH-R	gtcgacAACTGGTTCCCGGTCGG	Expression system of C.
23	cBAS2-F1	ctgcagATGGTCCGCATCACTCT	BAS2-GFP fusion expressing
24	cBAS2-R1	actagtGAAACCTTGCTTCTTGG	BAS2-GFP fusion expressing mutant
25	GFPlink-F	actagtGGAGCTGGTGCAGGCGCTGGA GC CGGTGCCATGGTGAGCAAGGGCGA	BAS2-GFP fusion expressing mutant

26	GFP-R	tctagaTTACTTGTACAGCTCGT	BAS2-GFP fusion expressing
			mutant
27	cBAS2-F2	tctagaATGGTCCGCATCACTCT	Transient expression in rubber-
			tree protoplasts
28	cBAS2-R2	gagctctGAAACCTTGCTTCTTGG	Transient expression in rubber-
			tree protoplasts