



Article

# MicroRNA-31 Emerges as a Predictive Biomarker of Pathological Response and Outcome in Locally Advanced Rectal Cancer

Cristina Caramés <sup>1</sup>, Ion Cristobal <sup>2,\*</sup>, Víctor Moreno <sup>1</sup>, Juan P. Marín <sup>3</sup>, Paula González-Alonso <sup>4</sup>, Blanca Torrejón <sup>2</sup>, Pablo Minguez <sup>5</sup>, Ana Leon <sup>1</sup>, José I. Martín <sup>1</sup>, Roberto Hernández <sup>1</sup>, Manuel Pedregal <sup>1</sup>, María J. Martín <sup>6</sup>, Delia Cortés <sup>7</sup>, Damian García-Olmo <sup>7</sup>, María J. Fernández <sup>4</sup>, Federico Rojo <sup>4</sup> and Jesús García-Foncillas <sup>1,2,\*</sup>

- Medical Oncology Department, Oncohealth Institute, Health Research Institute FJD-UAM, University Hospital Fundación Jiménez Díaz, Avda. Reyes Católicos-2, 28040 Madrid, Spain; ccarames@fjd.es (C.C.); victor.moreno@fjd.es (V.M.); Aleon@fjd.es (A.L.); JIMartin@fjd.es (J.I.M.); rhernandezl@quironsalud.es (R.H.); manuel.pedregal@quironsalud.es (M.P.)
- Traslational Oncology Division, Oncohealth Institute, Health Research Institute FJD-UAM, University Hospital Fundación Jiménez Díaz, Avda. Reyes Católicos-2, 28040 Madrid, Spain; blanca.torrejon@quironsalud.es
- Radiation Oncology Department, University Hospital Fundación Jiménez Díaz, Avda. Reyes Católicos-2, 28040 Madrid, Spain; marin5si@yahoo.com
- <sup>4</sup> Pathology Department, Health Research Institute FJD-UAM, University Hospital Fundación Jiménez Díaz, Avda. Reyes Católicos-2, 28040 Madrid, Spain; paula.galonso@fjd.es (P.G.-A.); mgg10167@gmail.com (M.J.F.); frojo@fjd.es (F.R.)
- Department of Genetics and Genomics, Health Research Institute FJD-UAM, University Hospital Fundación Jiménez Díaz, Avda. Reyes Católicos-2, 28040 Madrid, Spain; pablo.minguez@quironsalud.es
- Gastroenterology Department, University Hospital Fundación Jiménez Díaz, Avda. Reyes Católicos-2, 28040 Madrid, Spain; MJMartin@fjd.es
- Surgery Department, Oncohealth Institute, Health Research Institute FJD-UAM, University Hospital Fundación Jiménez Díaz, Avda. Reyes Católicos-2, 28040 Madrid, Spain; DCORTES@fjd.es (D.C.); Damian.garcia@uam.es (D.G.-O.)
- \* Correspondence: ion.cristobal@fjd.es (I.C.); jesus.garciafoncillas@oncohealth.eu (J.G.-F.); Tel.: +34-915-504-800 (I.C. & J.G.-F.)

Academic Editor: William Chi-shing Cho

Received: 22 April 2016; Accepted: 27 May 2016; Published: 3 June 2016

**Abstract:** Neoadjuvant chemoradiotherapy (CRT) followed by total mesorectal excision has emerged as the standard treatment for locally advanced rectal cancer (LARC) patients. However, many cases do not respond to neoadjuvant CRT, suffering unnecessary toxicities and surgery delays. Thus, identification of predictive biomarkers for neoadjuvant CRT is a current clinical need. In the present study, microRNA-31 expression was measured in formalin-fixed paraffin-embedded (FFPE) biopsies from 78 patients diagnosed with LARC who were treated with neoadjuvant CRT. Then, the obtained results were correlated with clinical and pathological characteristics and outcome. High microRNA-31 (miR-31) levels were found overexpressed in 34.2% of cases. Its overexpression significantly predicted poor pathological response (p = 0.018) and worse overall survival (OS) (p = 0.008). The odds ratio for no pathological response among patients with miR-31 overexpression was 0.18 (Confidence Interval = 0.06 to 0.57; p = 0.003). Multivariate analysis corroborated the clinical impact of miR-31 in determining pathological response to neoadjuvant CRT as well as OS. Altogether, miR-31 quantification emerges as a novel valuable clinical tool to predict both pathological response and outcome in LARC patients.

**Keywords:** miR-31; neoadjuvant chemoradiotherapy; rectal cancer; prognosis

#### 1. Introduction

Colorectal carcinoma (CRC) incidence, morbidity, and mortality rates vary markedly around the world, and rectal carcinoma represents approximately 28% of all CRC [1]. The standard of care for locally advanced rectal cancer (LARC) involves a multidisciplinary approach consisting of neoadjuvant chemoradiotherapy (CRT) followed by total mesorectal excision (TME) surgery. This fact was established after several studies showing that CRT before total mesorectal excision (neoadjuvant chemoradiotherapy) was associated with a lower percentage of patients with local recurrence than either with total mesorectal excision surgery followed by adjuvant CRT or total mesorectal excision surgery alone [2,3]. After the initiation of neoadjuvant CRT, a relevant observation is the range of pathological tumor response, from cases achieving a pathological complete response (pCR; ypT0N0) to others getting an absence of tumor regression at all or even tumor progression during therapy [4,5]. Of importance, pCR detected by pathological examination of the resected specimen is associated with significantly better prognosis as compared with patients with residual tumor, particularly residual nodal disease. Similarly, minimal residual disease is associated with better prognosis than gross residual disease [6,7]. Currently, the neoadjuvant CRT regimen most widely used is based on fluoropyrimidines concurrent with conventional fractionation radiotherapy [2]. Nevertheless, more than one-third of cases develop distant metastasis within 10 years from diagnosis and complete pathological response only occurs in 8% to 14% of patients. Thus, those patients who do not respond suffer undesired toxicities and delays in the resection of the primary tumor [8,9].

There are currently no effective methods to predict which patients will respond to neoadjuvant CRT. Despite some prognostic factors of local recurrence, distant recurrence and outcome have been proposed after tumor mesorectal excision surgery [10–12]; neoadjuvant CRT predictive biomarkers with an impact in recurrence and outcome are not still established in clinical practice [12–14]. The identification of patients who have a higher possibility of responding to preoperative CRT could be important in improving survival and decreasing treatment morbidity and local control in LARC. Moreover, patients who are potential non-responders could be moved to alternative therapeutic strategies. Therefore, the identification of novel alterations with predictive value of response to neoadjuvant CRT would be of high relevance for an optimal multidisciplinary treatment approach in this LARC patient subgroup.

MicroRNAs (miRNAs) are short (18–27 nucleotides) non-coding single-stranded RNA molecules which negatively regulate the expression of specific target genes at the post-transcriptional level. Depending on their target genes, microRNA deregulation has been related to CRC development, progression, or therapy response. Moreover, they can be easily detected in rectal cancer tissue and blood, so they have been proposed as promising biomarkers for diagnosis, prognosis, and monitoring therapies in this disease [15–17]. MicroRNA-31 (miR-31) maps on 9p21.3 and is one of the most significantly deregulated miRNAs in rectal cancer. It has been reported to be overexpressed in CRC cell lines as well as in rectal tumor tissue compared with the normal paired rectal mucosa [18]. In functional studies, the inhibition of miR-31 is able to impair CRC cell proliferation, invasion, and promote apoptosis—which suggests its oncogenic role in this disease [19]. Furthermore, high miR-31 levels have been described to correlate with tumor stage [18,20], poor prognosis [21], and shorter progression-free survival in CRC patients treated with anti-epithelial growth factor receptor (anti-EGFR) therapies [21]. Of importance, miR-31 suppression increases sensitivity to 5-fluorouracil (5-FU) and affects cell migration and invasion in the CRC cell line HCT-116 [22]. However, miR-31 ability to predict neoadjuvant CRT pathological response and outcome in LARC is a relevant question that remains to be explored. Here, we report that miR-31 deregulation is a common event in LARC that determines poor outcome. Moreover, this alteration is of high therapeutic relevance because it defines a subgroup of LARC patients who will not respond to neoadjuvant CRT.

#### 2. Results

# 2.1. Prevalence of miR-31 Deregulation in Locally Advanced Rectal Cancer (LARC) and Its Relation with Pathological and Clinical Characteristics

Eighty-two patients with LARC treated with neoadjuvant CRT were selected. All of them had a minimal follow-up greater than three years. The cohort flowchart diagram is shown in Figure 1. Owing to the loss of clinical follow-up data, four patients were excluded. Complete response, moderate response, minimal response, and a poor response was observed in 12.3%, 37%, 18.3%, and 28.4% of patients, respectively.

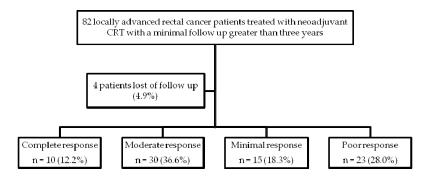


Figure 1. Patients flowchart through the study. CRT: chemoradiotherapy.

High miR-31 levels were observed in 34.2% of cases (27 out of 78). To explore the potential value of miR-31 predicting response to neoadjuvant CRT, patients with different grades of pathological response were compared. Interestingly, those patients who developed complete response tended to have low microRNA-31 levels, whereas the subgroup of patients with a poor pathological response showed high levels of miR-31. (p = 0.018). No statistical significance was observed in age, sex, Eastern Cooperative Oncology Group (ECOG) scale of performance status, histological grade, clinical stage, tumor size, and lymph node stage between patients with high and low miR-31 expression levels (Table 1). Similarly, no differences were found for pathological stage and downstaging.

**Table 1.** Baseline clinical and pathological characteristics of our locally advanced rectal cancer (LARC) cohort and its relation with miR-31 expression.

Clinical and pathological characteristics		n° Cases (%)	n° miR-31 High <sup>1</sup> (%)	n° miR-31 Low <sup>2</sup> (%)	р
miR-31 <sup>3 pre</sup> CRT		78 (100%)	27 (34.2)	51 (65.8)	
Age	<60 >60	24 (30.7) 54 (69.3)	10 (41.6) 17 (31.4)	14 (58,4) 37 (68.6)	0.267
Sex	Male Female	47 (60) 31 (40)	15 (31.9) 8 (68.1)	32 (68.1) 19 (31.9)	0.454
ECOG <sup>4</sup>	0 I	49 (62.8) 29 (37.2)	14 (28.5) 13 (71.5)	35 (71.5) 16 (28.5)	0.113
Clinical stage pre CRT <sup>5</sup>	II III	4 (5.2) 73 (94.8)	3 (75) 24 (30.7)	1 (25) 49 (69.3)	0.121
Neoadjuvant CRT	RT + 5-FU <sup>6</sup> based	78 (100%)			
Adjuvant therapy	5-FU FOLFOX <sup>7</sup> Other	55 (70.5) 6 (7.7) 17 (21.8)	20 (36.4) 3 (50) 4 (23.5)	35 (63.6) 3 (50) 13 (76.5)	0.407
Grade pre CRT	Low High ND <sup>8</sup>	20 (25.8) 50 (64) 8 (10.2)	5 (25) 20 (40) 2 (25)	15 (75) 30 (60) 6 (75)	0.410

Int. J. Mol. Sci. 2016, 17, 878 4 of 11

TO 1 '	1.	•	$\sim$	
Tab:	Ie -		เดท	11

Clinical and pathological characteristics		n° Cases (%)	n° miR-31 High <sup>1</sup> (%)	n° miR-31 Low <sup>2</sup> (%)	p	
	урТ0	10 (12.8)	1 (10)	9 (90)		
ypT <sup>9</sup>	урТ1-2 урТ3-4	33 (42.4) 32 (41)	13 (39.4) 12 (37.5)	20 (60.6) 20 (62.5)	0.372	
	ypTx	3 (3.8)	1 (33.3)	2 (66.7)		
ypN <sup>10</sup>	pN0	61 (78.2)	18 (29.5)	43 (70.5)	0.007	
ypin	pN+	17 (21.8)	9 (52.9)	8 (47.1)	0.086	
	ypT0N0	10 (12.8)	1 (10)	9 (90)		
Pathological stage	ypI	30 (38.5)	11 (36.6)	19 (63.4)	0.133	
i attiological stage	ypII	21 (27)	6 (28.6)	15 (71.4)	0.133	
	ypIII	17 (21.7)	9 (52.9)	8 (47.1)		
Downstaging	No	19 (24.3)	9 (47.4)	10 (52.6)	0.142	
Downstagnig	Yes	59 (75.7)	18 (52.6)	41 (47.4)	0.143	
	Complete response	10 (12.8)	1 (10)	9 (90)		
Pathology response	Moderate response	30 (38.5)	7 (23.3)	23 (76.7)	0.018	
r uniology response	Minimal response	15 (19.2)	5 (33.3)	10 (66.7)	0.016	
	Poor response	23 (29.5)	14 (60.8)	9 (39.2)		

<sup>&</sup>lt;sup>1</sup> miR-31 high =  $\Delta C_T$  miR-31 < 0.34 (cut-off established by a receiver operating characteristic (ROC) curve);

2.2. miR-31 Deregulation Predicts Pathological Response to Neoadjuvant Chemoradiotherapy (CRT) in LARC Patients

Receiver operating characteristic (ROC) curves were generated in order to investigate the potential utility of miR-31 as a predictive biomarker of response to neoadjuvant CRT. miR-31 expression levels provide an area under the curve (AUC) value of 0.71 (95% confidence interval (CI) = 0.57 to 0.84; p = 0.001) with 60.8% sensitivity and 76.3% specificity in distinguishing rectal cancer from poor responders to minimal, moderate, and complete responders. Negative predictive value (NPV) was 82.3% and positive predictive value (PPV) was 51.8% (Figure 2 and Table 2).

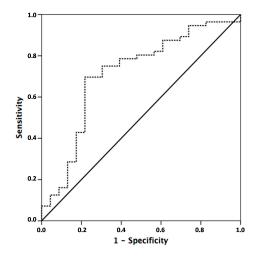


Figure 2. Receiver operating characteristic (ROC) Curve. Based on this ROC curve, the cutoff point that provides the best sensitivity and specificity to separate rectal cancer from any response to absence of response (Ryan 3) was selected. According with this criteria, miR-31 high (or overexpressed) was defined as  $\Delta C_T$  miR-31 < 0.34 and miR-31 low was defined as  $\Delta C_T$  miR-31 > 0.34. The dash line represents the coordinated points of the ROC curve. The solid line represents the ROC curve diagonal reference line.

 $<sup>^2</sup>$  miR-31 low =  $\Delta C_T$  miR-31 > 0.34 (cut-off established using a ROC curve);  $^3$  miR-31 = microRNA 31;  $^4$  ECOG = Eastern Cooperative Oncology Group;  $^5$  CRT = Chemoradiotherapy;  $^6$  5-FU = 5-fluorouracil;

FOLFOX = Fluorouracil, Oxaliplatin, Leucovorin; 8 ND = No data; 9 ypT = tumor size after CRT;  $^{\rm 10}$  ypN = pathological lymph node after CRT.

Int. J. Mol. Sci. 2016, 17, 878 5 of 11

Responders vs. Non-Responders				
Response	Response <sup>1</sup>	Non-Response <sup>2</sup>	Total	
miR-31 high <sup>3</sup>	13	14	27	
miR-31 low <sup>4</sup>	42	9	51	
Total	55	23	78	
	NPV <sup>5</sup> (%) 82.3 Specificity (%) 76 PPV <sup>6</sup> (%) 51.8 Sensitivity (%) 60			

Table 2. MiR-31 predicts pathological response to neoadjuvant CRT.

Moreover, multivariable logistic regression analyses revealed that miR-31 expression levels measured before the beginning of neoadjuvant CRT is a predictive pathological response marker. The odds ratio for non-responders was 0.18 (95% CI = 0.06 to 0.57; p = 0.003) (Table 3).

**Table 3.** Multivariable logistic analysis for miR-31 expression measured pre neoadjuvant CRT and the rest of clinic-pathological factors measured before neoadjuvant CRT in responder and non-responder patients.

Responders <sup>1</sup> vs. Non-Responders <sup>2</sup>	<sup>3</sup> OR (95% CI <sup>4</sup> )	p
Age, <60 vs. >60	0.97 (0.30 to 3.06)	0.962
Gender, Female vs. Male	1.25 (0.41 to 3.86)	0.452
ECOG <sup>5</sup> , O vs. I	0.79 (0.25 to 2.51)	0.799
Grade pre CRT <sup>6</sup> , high-moderate <i>vs.</i> low	1.25 (0.53 to 2.96)	0.600
Clinical stage, II vs. III	2.51 (0.20 to 30.37)	0.468
miR-31 high <sup>7</sup> vs. low <sup>8</sup>	0.18 (0.06 to 0.57)	0.003

 $<sup>^{1}</sup>$  "Response" = moderate, minimal or complete response;  $^{2}$  "Non-Response" = poor pathological response;  $^{3}$  OR = Odds ratio;  $^{4}$  CI = Confidence interval;  $^{5}$  ECOG = Eastern Cooperative Oncology Group;  $^{6}$  CRT = chemoradiotherapy;  $^{7}$  miR-31 high =  $\Delta C_{T}$  miR-31 < 0.34 (cutoff derived by receive operating characteristic curve);  $^{8}$  miR-31 low=  $\Delta C_{T}$  miR-31 > 0.34 (cutoff obtained by a receive operating characteristic curve).

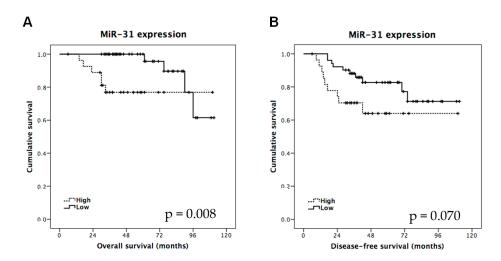
### 2.3. High miR-31 Levels Determines Poor Outcome in LARC Patients Treated with Neoadjuvant CRT

To further evaluate the clinical significance of miR-31 in LARC, we next investigated whether miR-31 levels could serve as a predictor of patient outcome. Interestingly, patients with higher miR-31 levels had statistically significantly worse overall survival (p = 0.008, Figure 3A). Despite high miR-31 expression also associated with shorter disease-free survival, significance was not achieved (p = 0.070)—probably due to the small number of cases included in this cohort (Figure 3B).

Additionally, Cox proportional hazard regression analyses showed that in the univariate analysis, poor outcome in LARC patients was related to high levels of miR-31 (Hazard Ratio (HR) = 5.077; 95% CI = 1.366 to 18.863; p = 0.015) and pathological stage (HR = 1.890; 95% CI = 1.027 to 3.478; p = 0.043). In the same way, multivariable analysis proved that high levels of miR-31 can be used as an independent prognostic biomarker for determining outcome in LARC patients treated with neoadjuvant CRT (HR = 0.206; 95% CI = 0.051 to 0.840; p = 0.028) (Table 4).

 $<sup>^1</sup>$  "Response" = moderate, minimal, or complete response;  $^2$  "Non-Response" = poor pathological response;  $^3$  miR-31 high =  $\Delta C_T$  miR-31 < 0.34 (cutoff derived by receive operating characteristic curve);  $^4$  miR-31 low=  $\Delta C_T$  miR-31 > 0.34 (cutoff derived by receive operating characteristic curve);  $^5$  NPV = negative predictive value;  $^6$  PPV = positive predictive value.

Int. J. Mol. Sci. 2016, 17, 878 6 of 11



**Figure 3.** Kaplan-Meier analysis of **(A)** overall survival (OS) and **(B)** disease-free survival (DFS) in LARC patients based upon miR-31 expression in primary tumors measured prior to the neoadjuvant treatment.

Table 4. LARC patients factors predictive of poor overall survival: Univariate and multivariable analyses.

Variables	Univariate		Multivariate	
valiables	HR <sup>1</sup> (95% CI <sup>2</sup> )	р	HR (95% CI)	p
Age, >60 vs. <60	0.915 (0.246 to 3.407)	0.895		
Sex, female vs. male	1.024 (0.56 to 1.86)	0.928		
Clinical stage, II vs. III	2.229 (0.274 to 18.132)	0.453		
Pathological ypT <sup>3</sup> , ypT0 <i>vs.</i> ypT1/T2 <i>vs.</i> ypT3/T4	2.064 (0.214 to 19.902)	0.531		
Pathological ypN <sup>4</sup> , N+ vs. N-	2.907 (0.918 to 9.211)	0.070		
Pathological stage	1.890 (1.027 to 3.478)	0.043	2.411 (1.136 to 5.114)	0.022
Pathological response, poor <i>vs.</i> complete <i>vs.</i> moderate <i>vs.</i> minimal	1.346 (0.70 to 2.56)	0.366		
MiR-31 expression, high <sup>5</sup> ( $\Delta C_T < 0.34$ ) vs. low <sup>6</sup> ( $\Delta C_T > 0.34$ )	5.077 (1.366 to 18.863)	0.015	0.206 (0.051 to 0.840)	0.028

<sup>&</sup>lt;sup>1</sup> HR = Hazard ratio; <sup>2</sup> CI = Confidence Interval; <sup>3</sup> ypT = tumor size after CRT; <sup>4</sup> ypN = pathological lymph node after CRT; <sup>5</sup> miR-31 high =  $\Delta C_T$  miR-31 < 0.34 (cutoff derived by receive operating characteristic curve); <sup>6</sup> miR-31 low =  $\Delta C_T$  miR-31 > 0.34 (cutoff derived by receive operating characteristic curve).

# 3. Discussion

Personalized treatment selection for patients with LARC relies on clinical variables measured prior to neoadjuvant CRT. Taken into account that, depending on the series, between 45% and 20% of the patients do not respond to neoadjuvant CRT therapy, and the lack of any clinical and pathological factors to guide the initial treatment, novel predictive biomarkers are needed to avoid toxicity and surgery delays. In this context, prior studies have reported the potential usefulness of some gene signatures and microRNA expression profiles to predict neoadjuvant CRT response [14,17]. However, those biomarkers are not well validated for use in daily clinical practice. Here, we show that miR-31 expression levels measured before starting any treatment have the potential ability to predict pathological response, overall survival, and progression-free survival in patients with LARC treated with 5-FU based neoadjuvant CRT.

We decided to evaluate the potential clinical value of miR-31 in predicting CRT response due to the previously-reported oncogenic role of this microRNA in CRC [23]. We found a similar prevalence (around 34%) of miR-31 overexpression to what has been previously reported in the literature (between

10% and 38% depending on the cohort and the stage of the disease) [23,24]. In concordance with our findings, it has been reported that miR-31 suppression increments sensitivity to 5-FU at the initial stage, and also affects invasion and cell migration in HCT-116 cells [22]. Moreover, several studies have reported the potential value of microRNAs to predict neoadjuvant CRT response; however, these works did not evaluate miR-31 expression levels [25,26].

In our study, high miR-31 levels were significantly associated with the lack of pathological response and patients with low miR-31 expression tended to have better response. As indicated above, the pathological response can be predicted with a specificity of 76.3%. Prior works pointed out that the degree of tumor regression might be a very important clinical tool, as it could be used as a prognostic marker; however, the clinical significance of tumor pathological response is still under investigation [7,27]. In our cohort, pathological response is not associated with overall survival (OS) and disease-free survival (DFS). However, we showed that miR-31 deregulation is able to predict pathological response, DFS, and OS. In this way, 78% of the patients with high miR-31 expression were alive at 6 years follow up compared with 96% of those with low miR-31. In addition, 65% of the patients with high miR-31 had a recurrence within the first three years of follow up, compared with 86% of the patients with low miR-31. Those results suggest the strength of miR-31 as a predictive biomarker to guide multidisciplinary treatment in patients with LARC, since cases with high miR-31 had no appreciable clinical benefit from neoadjuvant CRT. Considering these results, high miR-31 patients could benefit from an alternative therapeutic approach, different than 5-FU-based neoadjuvant CRT.

Measuring miR-31 expression levels as a predictive biomarker for neoadjuvant CRT in LARC has the advantage that is easily detected from small amounts of routinely-prepared formalin-fixed paraffin-embedded (FFPE) endoscopic samples by using a RT-PCR. Furthermore, microRNAs have the ability to continue stable, even when subjected to extreme conditions such as very low or high pH levels, boiling, longer storage time, and multiple freeze-thaw cycles [28]. Limitations of our study include the low number of cases included, the lack of validation in an independent larger cohort, and that it was a retrospective study instead of a prospective one. Therefore, the conclusions provided by this study have to be interpreted with caution and the potential clinical usefulness of miR-31 has to be further confirmed in forthcoming studies and controlled randomized clinical trials before a potential inclusion in clinical protocols. However, it is also true that this study is larger than other studies evaluating neoadjuvant CRT treatments, and the treatment regimen was very homogeneous.

Finally, prior works have described the interaction of miR-31 with important tumor suppressor genes, such as the enhancer of zeste homolog 2 (EZH2) [29] and the hypoxia-inducible transcription factor  $1\alpha$  (HIF- $1\alpha$ ) [30]. In the same way, in other tumor models, miR-31 represses the regulatory subunit B alpha isoform of the tumor suppressor PP2A (PPP2R2A) [31]. We previously reported that this subunit downregulation is a common event in colorectal cancer patients and its relation with the resistance to 5-FU [32]. According to this and despite there are likely to be multiple explanations for primary or acquired resistance to neoadjuvant CRT—exploring the role and interaction of miR-31 and PPP2R2A to predict neoadjuvant CRT response in LARC patients could be of great interest for future research.

#### 4. Materials and Methods

#### 4.1. Patient Selection

Eighty-one patients diagnosed with LARC and treated with neoadjuvant CRT between the beginning of 2007 and the end of 2012 (6 years) at the University Hospital Fundación Jiménez Díaz, (Madrid, Spain) were retrospectively selected and included in this study. They were followed-up until February 2016.

All patients had a preoperative staging based on transrectal ultrasound (TRUS) and/or magnetic resonance image (MRI) of the pelvis. In addition, a full body computed tomography scan (FBCT) was performed to exclude stage IV disease. The patients were treated with rule-based chemoradiotherapy

regimens based on 5-FU and underwent surgery after one and a half to two months after neoadjuvant CRT completion. Every patient gave written informed consent for tissue storage and analysis at Fundación Jiménez Díaz biobank, Madrid (Spain). Fundación Jiménez Díaz University Hospital, Madrid (Spain) institutional review board approved the study.

#### 4.2. Pathologic Response and Tumor Samples

The Fundación Jimenez Diaz biobank provided the tumor samples. All tissue derived from the surgical resection were classified in concordance with the College of American Pathologist guidelines for invasive carcinomas (TNM, 7th ed.). Two independent pathologists blinded to all patients' clinical data evaluated tumor regression grade according to the modified Ryan classification that categorizes tumors in four levels of response: complete, moderate, minimal, and poor response. Complete response score 0 indicates no viable cancer cells; moderate score 1 indicates single cells or little groups of cancer cells; minimal score 2 indicates residual cancer outgrown by fibrosis; and poor response scores 3 indicates minimal or no tumor kill with extensive residual cancer. Every regression grade was compared with the primary tumor, in concordance with clinical guidelines [33].

#### 4.3. RNA Isolation

We isolated total RNA from formalin-fixed paraffin-embedded (FFPE) tumor biopsies, applying RecoverAll Total Nucleic Acid Isolation kit Ambion (Thermo Fisher Scientific, USA in accordance with manufacturer's instructions. We eluted total RNA and quantified it with a NanoDrop Spectrophotometer (Thermo scientific, Waltham, MA, USA)

# 4.4. Quantification of MicroRNA Expression Levels

Samples were reverse transcribed using the TaqManHMicroRNA Reverse Transcription Kit (Applied Biosystems Waltham, MA, USA). Working with TaqMan MicroRNA Assays (Applied Biosystems, USA) specific for miR-31 (reference number: 002279) and U6B as internal control, mature miRNAs were quantified by quantitative real-time reverse transcription polymerase chain reaction (RT-PCR). TaqMan MicroRNA assays have an ultra-high sensitivity and can detect as few as ten copies of the target transcript in a sample and also have a dynamic range greater than seven logs. Reactions were carried out using an Applied Biosystems 7500 Sequence Detection System. Conditions: 95 °C for 10 min, followed by 45 cycles of 95 °C for 15 s, and 60 °C for 1 min. The  $\Delta C_T$  method was performed to analyze relative gene expression data.

#### 4.5. Statistical Analysis

We used SPSS v14.0 (SPSS Inc, Chicago, IL, USA) for statistical analyses. We applied the  $\chi 2$  test (Fisher exact test) based on bimodal distribution of data to evaluate the correlation between miR-31 overexpression and the clinical and pathological variables. All reported p values are two-sided. We made a receiver operating characteristic (ROC) curve to establish a cutoff for miR-31 expression, calculated its 95% confidence interval, and selected the cutoff point that provides us with the best specificity and sensitivity to differentiate rectal cancer pathological response, in order to assess the potential usefulness of miR-31 as a predictive biomarker. According to this criteria, we considered up-regulation when miR-31 expression levels ( $\Delta C_{\rm T}$ ) were lower than 0.34. We defined DFS as the time from surgery until recurrence, appearance of a secondary tumor, or death, and OS as the time from the date of diagnosis to the date of last follow-up or death, and applied the Kaplan-Meier method and survival comparisons with the log-rank test and Breslow. Then, we adjusted the Cox proportional hazards model by taking into consideration significant parameters in univariate analysis and considered a p value <0.05 statistically significant. We followed the Reporting Recommendations for Tumor Marker Prognostic Studies (REMARK) guideline [34].

#### 5. Conclusions

In conclusion, our work shows evidence for the potential utility of miR-31 as a predictive biomarker for pathological response and outcome in patients with LARC treated with neoadjuvant CRT—a concept that can be very interesting to better select treatment options in patients with LARC and that can be incorporated into daily clinical practice. However, more studies are needed to better define the biological and clinical significance of miR-31 in rectal cancer. Furthermore, a validation of these results in a larger independent LARC patient cohort is necessary to validate our results.

**Acknowledgments:** This work was supported by PT13/0010/0012, PI13/02609 and PI15/00934 grants from "Instituto de Salud Carlos III FEDER, Madrid (Spain)".

**Author Contributions:** Jesús García-Foncillas, Cristina Caramés and Ion Cristobal conceived and designed the experiments; Víctor Moreno, Cristina Caramés, Pablo Minguez and Ion Cristobal performed the experiments and analyzed the data; Juan P. Marín, Paula González-Alonso, Blanca Torrejón, Ana Leon, José I. Martín, María J. Martín, María J. Fernández, Damian García-Olmo, Delia Cortés, Manuel Pedregal, Roberto Hernández, and Federico Rojo contributed reagents/materials/analysis tools and helped to draft the manuscript; Cristina Caramés wrote the paper.

**Conflicts of Interest:** The authors declare no conflict of interest. The founding sponsors had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

#### Abbreviations

LARC Locally advanced rectal cancer

CRT Chemoradiotherapy

MiR MicroRNA MiR-31 MicroRNA-31

FFPE formalin-fixed paraffin-embedded

5-FU 5-fluorouracil

pCR pathological complete response; ypT0N0

CRC colorectal cancer

anti-EGFR anti-epithelial growth factor receptor

RT-PCR quantitative real-time polymerase chain reaction

PPP2R2A regulatory subunit B alpha isoform of the tumor suppressor PP2A

MRI Magnetic resonance image TRUS Transrectal ultrasound

FBCT full body computed tomography scan

REMARK Reporting Recommendations for Tumor Marker Prognostic Studies

#### References

- 1. Brenner, H.; Kloor, M.; Pox, C.P. Colorectal cancer. Lancet 2014, 383, 1490–1502. [CrossRef]
- 2. Sauer, R.; Liersch, T.; Rodel, C. Preoperative *versus* postoperative chemoradiotherapy for locally advanced rectal cancer: Results of the German CAO/ARO/AIO-94 randomized phase III trial after a median follow-up of 11 years. *J. Clin. Oncol.* **2012**, *30*, 1926–1933. [CrossRef] [PubMed]
- 3. Van Gijn, W.; Marijnen, C.A.; van de Velde, C.J. Preoperative radiotherapy combined with total mesorectal excision for resectable rectal cancer: 12 years follow-up of the multicentre, randomised controlled TME trial. *Lancet Oncol.* **2011**, *12*, 575–582. [CrossRef]
- 4. Chetty, R.; Gill, P.; Govender, D.; Bateman, A.; Chang, H.J.; Deshpande, V.; Driman, D.; Gomez, M.; Greywoode, G.; Jaynes, E.; *et al.* International study group on rectal cancer regression grading: Interobserver variability with commonly used regression grading systems. *Hum. Pathol.* **2012**, *43*, 1917–1923. [CrossRef] [PubMed]
- 5. MacGregor, T.P.; Maughan, T.S.; Sharma, R.A. Pathological grading of regression following neoadjuvant chemoradiation therapy: The clinical need is now. *J. Clin. Pathol.* **2012**, *65*, 867–871. [CrossRef] [PubMed]
- Fokas, E.; Liersch, T.; Fietkau, R.; Hohenberger, W.; Beissbarth, T.; Hess, C.; Becker, H.; Ghadimi, M.; Mrak, K.; Merkel, S.; et al. Tumor regression grading after preoperative chemoradiotherapy for locally advanced rectal carcinoma revisited: Updated results of the CAO/ARO/AIO-94 trial. J. Clin. Oncol. 2014, 32, 1554–1562.
  [CrossRef] [PubMed]

7. Rödel, C.; Martus, P.; Papadoupolos, T.; Füzesi, L.; Klimpfinger, M.; Fietkau, R.; Liersch, T.; Hohenberger, W.; Raab, R.; Sauer, R.; *et al.* Prognostic significance of tumor regression after preoperative chemoradiotherapy for rectal cancer. *J. Clin. Oncol.* **2005**, *23*, 8688–8696. [CrossRef] [PubMed]

- 8. Gérard, J.P.; Conroy, T.; Bonnetain, F.; Bouché, O.; Chapet, O.; Closon-Dejardin, M.T.; Untereiner, M.; Leduc, B.; Francois, É.; Maurel, J.; *et al.* Preoperative radiotherapy with or without concurrent fluorouracil and leucovorin in T3-4 rectal cancers: Results of FFCD 9203. *J. Clin. Oncol.* 2006, 24, 4620–4625. [CrossRef] [PubMed]
- 9. Bosset, J.F.; Collette, L.; Calais, G.; Mineur, L.; Maingon, P.; Radosevic-Jelic, L.; Daban, A.; Bardet, E.; Beny, A.; Ollier, J.C.; *et al.* Chemotherapy with preoperative radiotherapy in rectal cancer. *N. Engl. J. Med.* **2006**, 355, 1114–1123. [CrossRef] [PubMed]
- 10. Nagtegaal, I.D.; Gosens, M.J.; Marijnen, C.A.M.; Rutten, H.J.; van de Velde, C.J.; van Krieken, J.H. Combinations of tumor and treatment parameters are more discriminative for prognosis than the present TNM system in rectal cancer. *J. Clin. Oncol.* **2007**, *25*, 1647–1650. [CrossRef] [PubMed]
- 11. Kim, N.K.; Baik, S.H.; Seong, J.S.; Kim, H.; Roh, J.K.; Lee, K.Y.; Sohn, S.K.; Cho, C.H. Oncologic outcomes after neoadjuvant chemoradiation followed by curative resection with tumor-specific mesorectal excision for fixed locally advanced rectal cancer. *Ann. Surg.* **2006**, 244, 1024–1030. [CrossRef] [PubMed]
- 12. Bertolini, F.; Bengala, C.; Losi, L.; Pagano, M.; Iachetta, F.; Dealis, C.; Jovic, G.; Depenni, R.; Zironi, S.; Falchi, A.M.; *et al.* Prognostic and predictive value of baseline and posttreatment molecular marker expression in locally advanced rectal cancer treated with neoadjuvant chemoradiotherapy. *Int. J. Radiat. Oncol. Biol. Phys.* **2007**, *68*, 1455–1461. [CrossRef] [PubMed]
- 13. Kuremsky, J.G.; Tepper, J.E.; McLeod, H.L. Biomarkers for response to neoadjuvant chemoradiation for rectal cancer. *Int. J. Radiat. Oncol. Biol. Phys.* **2009**, *74*, 673–688. [CrossRef] [PubMed]
- 14. Casado, E.; Garcia, V.M.; Sanchez, J.J.; Blanco, M.; Maurel, J.; Feliu, J.; Fernandez-Martos, C.; de Castro, J.; Castelo, B.; Belda-Iniesta, C.; *et al.* A combined strategy of SAGE and quantitative PCR provides a 13-gene signature that predicts preoperative chemoradiotherapy response and outcome in rectal cancer. *Clin. Cancer Res.* **2011**, *17*, 4145–4154. [CrossRef] [PubMed]
- 15. Kong, Y.W.; Ferland-McCollough, D.; Jackson, T.J.; Bushell, M. MicroRNAs in cancer management. *Lancet Oncol.* **2012**, 13, e249–e258. [CrossRef]
- 16. Toiyama, Y.; Takahashi, M.; Hur, K.; Nagasaka, T.; Tanaka, K.; Inoue, Y.; Kusunoki, M.; Boland, C.R.; Goel, A. Serum miR-21 as a diagnostic and prognostic biomarker in colorectal cancer. *J. Natl. Cancer Inst.* **2013**, *105*, 849–859. [CrossRef] [PubMed]
- 17. Caramés, C.; Cristóbal, I.; Moreno, V.; del Puerto, L.; Moreno, I.; Rodriguez, M.; Marín, J.P.; Correa, A.V.; Hernández, R.; Zenzola, V.; *et al.* MicroRNA-21 predicts response to preoperative chemoradiotherapy in locally advanced rectal cancer. *Int. J. Colorectal Dis.* **2015**, *30*, 899–906. [CrossRef] [PubMed]
- 18. Bandrés, E.; Cubedo, E.; Agirre, X.; Malumbres, R.; Zárate, R.; Ramirez, N.; Abajo, A.; Navarro, A.; Moreno, I.; Monzó, M.; *et al.* Identification by real-time PCR of 13 mature microRNAs differentially expressed in colorectal cancer and non-tumoral tissues. *Mol. Cancer* **2006**, *5*, 29. [CrossRef] [PubMed]
- 19. Sun, D.; Yu, F.; Ma, Y.; Zhao, R.; Chen, X.; Zhu, J.; Zhang, C.Y.; Chen, J.; Zhang, J. MicroRNA-31 activates the RAS pathway and functions as an oncogenic MicroRNA in human colorectal cancer by repressing RAS p21 GTPase activating protein 1 (RASA1). *J. Biol. Chem.* **2013**, *288*, 9508–9518. [CrossRef] [PubMed]
- 20. Slattery, M.L.; Herrick, J.S.; Mullany, L.E.; Valeri, N.; Stevens, J.; Caan, B.J.; Samowitz, W.; Wolff, R.K. An evaluation and replication of miRNAs with disease stage and colorectal cancer-specific mortality. *Int. J. Cancer* 2015, 137, 428–438. [CrossRef] [PubMed]
- 21. Nosho, K.; Igarashi, H.; Nojima, M.; Ito, M.; Maruyama, R.; Yoshii, S.; Naito, T.; Sukawa, Y.; Mikami, M.; Sumioka, W.; *et al.* Association of microRNA-31 with BRAF mutation, colorectal cancer survival and serrated pathway. *Carcinogenesis* **2014**, *35*, 776–783. [CrossRef] [PubMed]
- 22. Wang, C.J.; Stratmann, J.; Zhou, Z.G.; Sun, X.F. Suppression of microRNA-31 increases sensitivity to 5-FU at an early stage, and affects cell migration and invasion in HCT-116 colon cancer cells. *BMC Cancer* **2010**, *10*, 616. [CrossRef] [PubMed]
- 23. Manceau, G.; Imbeaud, S.; Thiébaut, R.; Liébaert, F.; Fontaine, K.; Rousseau, F.; Génin, B.; Le Corre, D.; Didelot, A.; Vincent, M.; *et al.* Hsa-miR-31-3p expression is linked to progression-free survival in patients with KRAS wild-type metastatic colorectal cancer treated with anti-EGFR therapy. *Clin. Cancer Res.* **2014**, *20*, 3338–3347. [CrossRef] [PubMed]

24. Mlcochova, J.; Faltejskova-Vychytilova, P.; Ferracin, M.; Zagatti, B.; Radova, L.; Svoboda, M.; Nemecek, R.; John, S.; Kiss, I.; Vyzula, R.; *et al.* MicroRNA expression profiling identifies miR-31-5p/3p as associated with time to progression in wild-type RAS metastatic colorectal cancer treated with cetuximab. *Oncotarget* **2015**, *6*, 38695–38704. [PubMed]

- 25. Della Vittoria Scarpati, G.; Falcetta, F.; Carlomagno, C.; Ubezio, P.; Marchini, S.; de Stefano, A.; Singh, V.K.; D'Incalci, M.; de Placido, S.; Pepe, S. A specific miRNA signature correlates with complete pathological response to neoadjuvant chemoradiotherapy in locally advanced rectal cancer. *Int. J. Radiat. Oncol. Biol. Phys.* **2012**, *83*, 1113–1119. [CrossRef] [PubMed]
- 26. Svoboda, M.; Sana, J.; Fabian, P.; Kocakova, I.; Gombosova, J.; Nekvindova, J.; Radova, L.; Vyzula, R.; Slaby, O. MicroRNA expression profile associated with response to neoadjuvant chemoradiotherapy in locally advanced rectal cancer patients. *Radiat. Oncol.* **2012**, *7*, 195. [CrossRef] [PubMed]
- 27. Park, I.J.; You, Y.N.; Agarwal, A.; Skibber, J.M.; Rodriguez-Bigas, M.A.; Eng, C.; Feig, B.W.; Das, P.; Krishnan, S.; Crane, C.H.; *et al.* Neoadjuvant treatment response as an early response indicator for patients with rectal cancer. *J. Clin. Oncol.* **2012**, *30*, 1770–1776. [CrossRef] [PubMed]
- 28. Kurihara, H.; Maruyama, R.; Ishiguro, K.; Kanno, S.; Yamamoto, I.; Ishigami, K.; Mitsuhashi, K.; Igarashi, H.; Ito, M.; Tanuma, T.; *et al.* The relationship between EZH2 expression and microRNA-31 in colorectal cancer and the role in evolution of the serrated pathway. *Oncotarget* **2016**, *7*, 122704–122717.
- 29. Toiyama, Y.; Okugawa, Y.; Goel, A. DNA methylation and MicroRNA biomarkers for noninvasive detection of gastric and colorectal cancer. *Biochem. Biophys. Res. Commun.* **2014**, 455, 43–57. [CrossRef] [PubMed]
- 30. Chen, T.; Yao, L.Q.; Shi, Q.; Ren, Z.; Ye, L.C.; Xu, J.M.; Zhou, P.H.; Zhong, Y.S. MicroRNA-31 contributes to colorectal cancer development by targeting factor inhibiting HIF-1α (FIH-1α). *Cancer Biol. Ther.* **2014**, *15*, 516–523. [CrossRef] [PubMed]
- 31. Liu, X.; Sempere, L.F.; Ouyang, H.; Memoli, V.A.; Andrew, A.S.; Luo, Y.; Demidenko, E.; Korc, M.; Shi, W.; Preis, M.; *et al.* MicroRNA-31 functions as an oncogenic microRNA in mouse and human lung cancer cells by repressing specific tumor suppressors. *J. Clin. Investig.* **2010**, 120, 1298–1320. [CrossRef] [PubMed]
- 32. Cristobal, I.; Manso, R.; Rincon, R.; Carames, C.; Senin, C.; Borrero, A.; Martinez-Useros, J.; Rodriguez, M.; Zazo, S.; Martinez-Aguilera, O.; *et al.* PP2A inhibition is a common event in colorectal cancer and its restoration using FTY720 shows promising therapeutic potential. *Mol. Cancer Ther.* **2014**, *13*, 938–947. [CrossRef] [PubMed]
- 33. Greenson, J.K.; Bonner, J.D.; Ben-Yzhak, O.; Cohen, H.I.; Miselevich, I.; Resnick, M.B.; Trougouboff, P.; Tomsho, L.D.; Kim, E.; Low, M.; *et al.* Phenotype of microsatellite unstable colorectal carcinomas: Well-differentiated and focally mucinous tumors and the absence of dirty necrosis correlate with microsatellite instability. *Am. J. Surg. Pathol.* **2003**, *27*, 563–570. [CrossRef] [PubMed]
- 34. McShane, L.M.; Altman, D.G.; Sauerbrei, W.; Taube, S.E.; Gion, M.; Clark, G.M. Reporting recommendations for tumor MARKer prognostic studies (REMARK). *Breast Cancer Res. Treat.* **2006**, *100*, 229–235. [CrossRef] [PubMed]



© 2016 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC-BY) license (http://creativecommons.org/licenses/by/4.0/).