

Article

MicroRNA-Regulated Protein-Protein Interaction Networks and Their Functions in Breast Cancer

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Abstract: MicroRNAs, which are small endogenous RNA regulators, have been associated with various types of cancer. Breast cancer is a major health threat for women worldwide. Many miRNAs were reported to be associated with the progression and carcinogenesis of breast cancer. In this study, we aimed to discover novel breast cancer-related miRNAs and to elucidate their functions. First, we identified confident miRNA-target pairs by combining data from miRNA target prediction databases and expression profiles of miRNA and mRNA. Then, miRNA-regulated protein interaction networks (PINs) were constructed with confident pairs and known interaction data in the human protein reference database (HPRD). Finally, the functions of miRNA-regulated PINs were elucidated by functional enrichment analysis. From the results, we identified some previously reported breast cancer-related miRNAs and functions of the PINs, e.g., miR-125b, miR-125a,

miR-21, and miR-497. Some novel miRNAs without known association to breast cancer were also found, and the putative functions of their PINs were also elucidated. These include miR-139 and miR-383. Furthermore, we validated our results by receiver operating characteristic (ROC) curve analysis using our miRNA expression profile data, gene expression-based outcome for breast cancer online (GOBO) survival analysis, and a literature search. Our results may provide new insights for research in breast cancer-associated miRNAs.

Keywords: miRNA; breast cancer; protein interaction network; functional analysis

1. Introduction

Breast cancer is a global health threat for women. According to a 2008 survey [1], breast cancer was the leading cause of cancer deaths in women. Our knowledge of possible risk factors has led to developments in diagnostic methods, drugs, and surgery procedures for treatment [2,3]; however, the details of breast carcinoma progression, and perhaps most importantly, how to cure breast cancer, remain elusive.

Previous research has identified a number of risk factors for breast cancer. Early menarche, late menopause, obesity, late first full pregnancy, and hormone replacement therapy were considered as high risk factors for breast cancer [2]. Breast cancer risk has also been reported to be related to fat intake in diets rich in red meats and high-fat dairy foods [3].

MicroRNAs (miRNAs) [4], are short endogenous non-coding RNAs which are able to regulate gene expression. After miRNA precursors are transcribed from the genome or generated from spliceosomes, they are exported to the cytoplasm and further processed by the Dicer complex [5]. The mature miRNA is then bound to Argonaute protein, forming a miRNA-protein complex known as the RNA-induced silencing complex (RISC) [6], miRNP, or RNAi (RNA interference) enzyme complex [7,8]. The RISC has been reported to down-regulate target genes by translational repression [9] or mRNA cleavage [10].

Like other protein-based regulators, miRNAs have been associated with cancer. Calin *et al.*, reported that miR-15 and miR-16 were deleted in leukemia [11], which was believed to be one of the earliest reports associating miRNAs with cancer [12]. After this report, many miRNAs were found to act as tumor suppressors or oncogenes (also known as oncomirs). For example, miR-21 was identified as an oncomir in hepatocellular cancer [13], breast cancer [14], and kidney cancer [15]. On the other hand, let-7c was found to be a tumor suppressor in prostate cancer [16], and miR-181a was reported as a tumor suppressor in glioma [17]. Further, miR-125b [18], and miR-145 [19] were identified as tumor suppressors in breast cancer, and miR-125a was found to repress tumor growth in breast cancer [20]. Thus, it is highly likely that miRNAs play an important role in breast cancer.

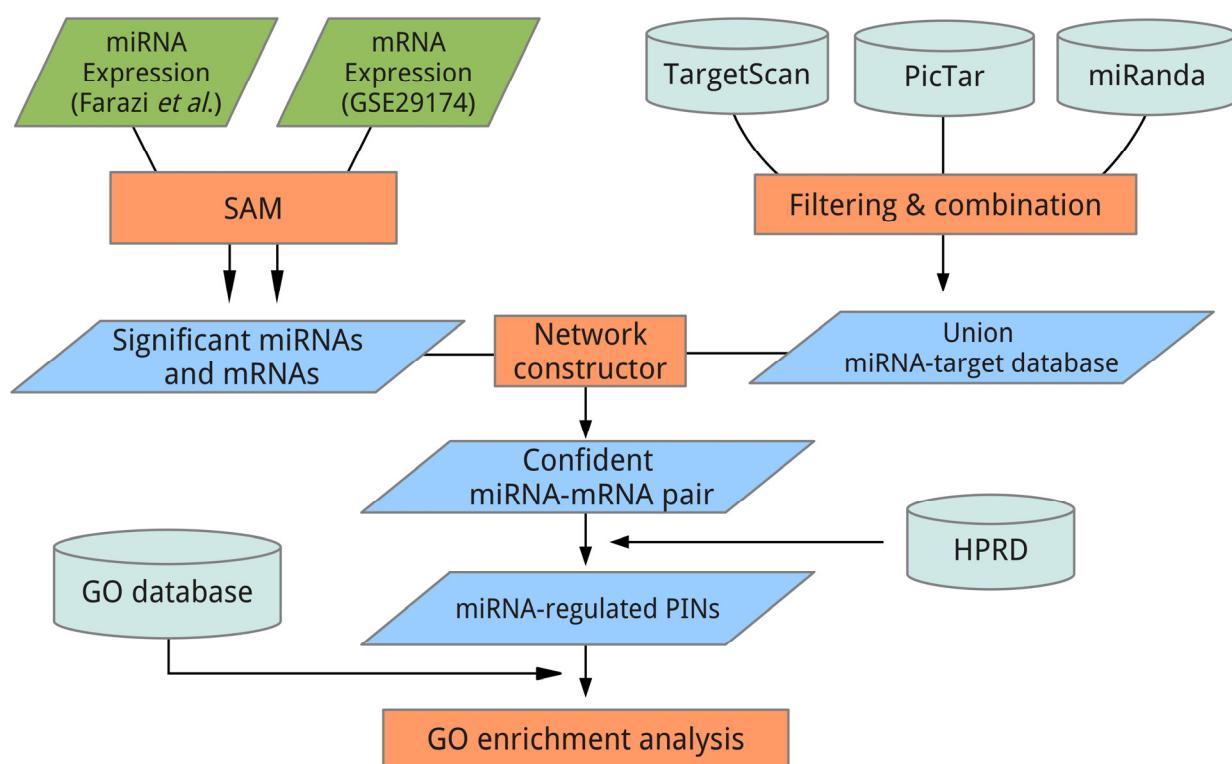
Since miRNA functions by regulating its target genes, we may deduce the effects of miRNAs by analyzing their regulated networks. To use such a method to elucidate miRNA functions, targets of miRNAs should be deduced. Currently, predictions in most target prediction database are based on sequence and statistical methods [21]. For example, in TargetScan, seed base pairing, target site context, conservation of target site and miRNA, and site accessibility are considered in the prediction process [22].

Another method to elucidate miRNA targets is to integrate expression profiles of miRNA and mRNA. In the work of Huang *et al.* [23], a Bayesian-based algorithm, GenMiR++, was developed to predict possible targets of 104 miRNAs in humans. They also verified their results by RT-PCR and microarray experiments. However, the power of other sequence-based target prediction algorithms was not utilized in their work.

It is also possible to combine sequence-based target prediction and expression-based target prediction methods. By integrating expression data into sequence-based predictions, possible false positives can be reduced. Previously, miRNA-mRNA interactions were explored with splitting-averaging Bayesian networks [24]. In that work, expression profiles of miRNA and mRNA from public databases, miRNA target prediction databases, and miRNA sequence information were integrated together to discover miRNA-mRNA interaction networks.

Here, we combined expression profiles of miRNA and mRNA, and three target prediction databases, TargetScan, PicTar and miRanda, to obtain confident miRNA-mRNA relationships and construct miRNA-regulated protein-protein networks for breast cancer. Furthermore, we explored the functions of the miRNAs by inspecting the underlying protein interaction networks (PINs) of the miRNAs with functional enrichment analysis. This method, as described in Figure 1, was used to elucidate the functions of gastric cancer-related miRNAs in our previous work [25]. In that study, a gastric cancer-associated miRNA, miR-148a, was identified and validated as being involved in tumor proliferation, invasion, migration, and the survival rate of the patients. By using a similar method, we aimed to elucidate breast cancer-related miRNA-regulated PINs and their functions.

Figure 1. Analysis flow chart used in this work. After expression profiles and target prediction databases were fetched and preprocessed, they were subjected to the analysis process described here and in the “Experimental Section”.



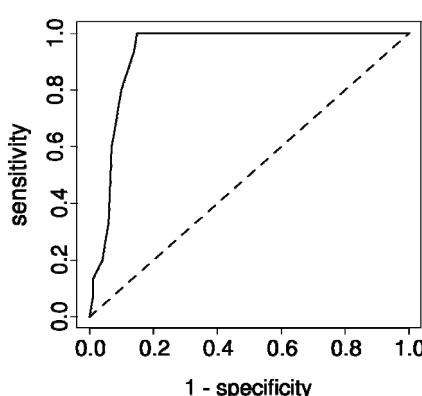
2. Results and Discussion

To construct miRNA-regulated PINs, differentially expressed miRNAs and genes from the dataset from Farazi *et al.* [26] were extracted following proper processing of the expression profiles. From our selected public miRNA dataset, we found 89 down-regulated miRNAs (93 prior to fold-change filtering) and only 1 up-regulated miRNA (Table S1). In gene expression dataset GSE29174, we found a total of 1268 down-regulated genes and 587 up-regulated genes before applying the fold change filter. There were 726 down-regulated genes (Table S2) and 437 up-regulated genes (Table S3) after significantly and differentially expressed genes were filtered by fold change (fold change >2).

From the results of SAM analysis, we identified some well-known breast cancer-related miRNAs (Table S1). For example, miR-214-3p [27] and miR-335-5p [28] have been previously reported to be down-regulated in breast cancer. Let-7c was found to be down-regulated in this work, while let-7a, another member of the let-7 family, was found to be down-regulated in another work [29]. MicroRNAs of the let-7 family were also reportedly down-regulated in several types of cancer [30]. We also found that miR-21-5p, the sole up-regulated miRNA in our list, was also previously found to be up-regulated [14,31]. However, changes in the expression of most of the miRNAs in our down-regulated list have not been reported in the literature. Therefore, we could not rule out the possibility that these miRNAs were novel breast cancer-related miRNAs. There are also some well-known miRNAs not presented in our list (for such a list, one may see [32–34]). The reason that some known miRNAs, for example, miR-19a, miR-155 and miR-205, did not show up in our result might be that we used a very stringent threshold (described in Experimental Section) when selecting differentially expressed miRNAs for PIN construction.

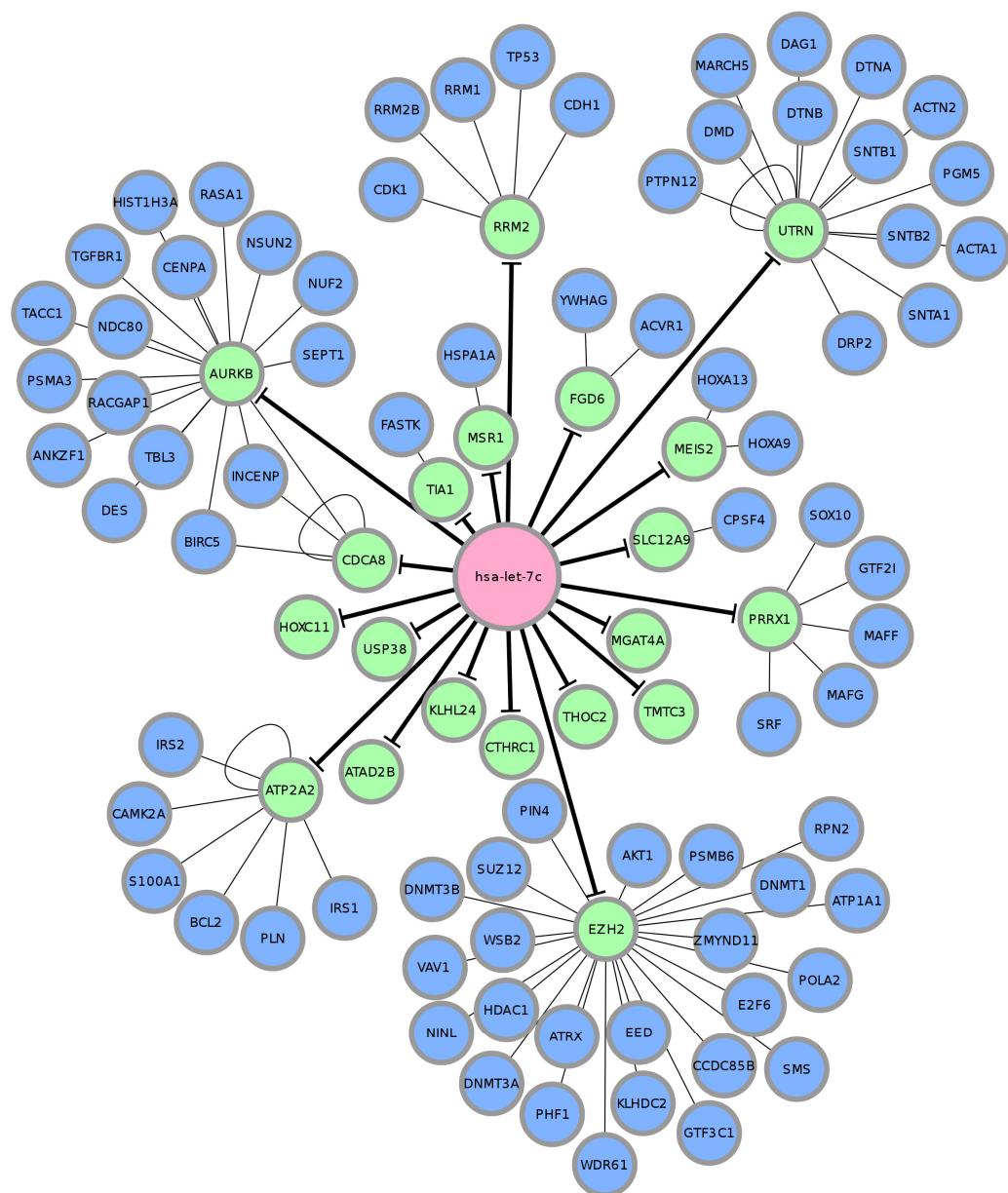
Since the miRNAs of the miRNA-regulated PINs were differentially expressed between normal and tumor tissues, and we identified some cancer-related functions in our functional enrichment analysis, the miRNAs may potentially be useful diagnostic markers for breast cancer. To verify this, we applied ROC curve analysis on the miRNA expression profile that was not used in constructing the miRNA-regulated PINs. Notably, our results (Figures 2 and S1 and Table S4) showed that let-7c (Figure 2), miR-497-5p, miR-125b-5p, and some other miRNAs of miRNA-regulated PINs, performed well when used as breast cancer diagnostic markers.

Figure 2. Receiver operating characteristic (ROC) curve of let-7c from our miRNA array dataset. For ROC curves of other miRNAs, see Figure S1.



Following elucidation of differentially expressed miRNAs and genes, miRNA-regulated PINs could then be constructed. We identified and constructed partial networks, containing the miRNA and its direct target, with the differentially expressed miRNAs and genes as described in the “Experimental Section”. We then extended the network by appending known interactions from the HPRD database. Finally, 18 miRNA-regulated PINs were constructed by the steps described above (Figure 3, Figures S2–S13, and Table S5).

Figure 3. The let-7c-regulated protein interaction network (PIN). This is one of the 18 miRNA-regulated PINs constructed in this work. Figures of all other miRNA-regulated PINs are displayed in Supplementary Figures S2–S13.



After construction of the 18 PINs was completed, we observed that the sizes of the PINs were not similar: some of the miRNAs seemed to regulate larger sized PINs, while other miRNAs affected only a small number of genes. Small miRNA-regulated PINs may be caused by the strict *q*-value threshold

set during SAM analysis, the processing steps performed on the target prediction databases discussed previously, and possibly by lack of protein-protein interaction data for some proteins in the HPRD. Although the HPRD may be considered the most comprehensive source of protein-protein interaction data [35], some proteins may not have been considered and researched by other investigators, and therefore, interaction data for those proteins would not be included in the HPRD. However, it may be true that some of the miRNA-regulated PINs were small in breast cancer, since the construction of the PINs were based on differentially expressed miRNAs and genes between normal tissues and tumor samples, and those miRNAs with small PINs may not be as important as others with larger PINs.

To elucidate the functions of a miRNA with a regulated PIN, GO enrichment analysis was applied to the miRNA-regulated PINs. We did not consider the miRNAs with ≤ 5 genes in their regulated PINs, and some of the miRNA-regulated PINs had no enriched functions using the defined threshold, $FDR < 0.0001$. To exclude GO terms that describe a broad range of concepts, we only included high level GO terms, *i.e.*, larger than 5.

The results of the GO enrichment analysis for let-7c related to cancer are listed in Table 1. (Results of all miRNA-regulated PINs were in Table S6). We defined a GO term as cancer-related if a GO term contained “cell proliferation”, “cell death”, “apoptosis”, “signaling”, “microtubule”, and “actin”. We noted that 7 miRNAs had enriched GO terms related to apoptosis, cell death, and cell proliferation, *i.e.*, miR-520d-3p, miR-497-5p, miR-125b-5p, miR-21-5p, miR-31-5p, let-7c, and miR-125-5p. Further, some miRNA-regulated PINs may have functions other than cell survival. For example, the nerve growth factor receptor pathway was enriched in miR-regulated PINs of miR-520d-3p, miR-497-5p, miR-125a-5p, miR-125b-5p, and miR-31-5p, and the epidermal growth factor receptor pathway was enriched in miR-regulated PINs of miR-520d-3p, miR-21-5p, and miR-497-5p. Most of the miRNAs had been previously described and were known to be implicated in breast cancer. Let-7c was not only likely to be down-regulated in breast cancer [29], but was also found to be a tumor suppressor in prostate cancer [16]. Another reported tumor suppressor was miR-125b-5p, which was found to be down-regulated in breast tumor tissue [18], and this finding was consistent with our functional enrichment results. The miR-125a-5p-regulated PIN was found to be able to inhibit apoptosis and regulate epithelial cell proliferation, and has been reported to repress cell growth [36].

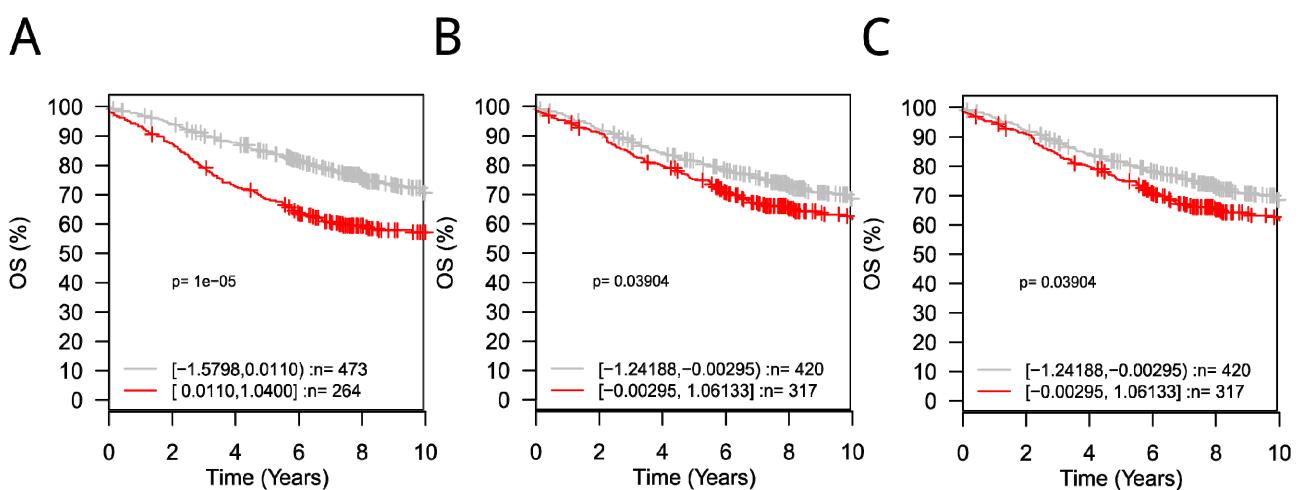
Table 1. Selected enriched functions of let-7c. Member genes of the let-7c-regulated network annotated with corresponding enriched functions are listed.

MIMAT0000064(hsa-let-7c)		
GO term	Genes	Adj. p-value
GO:0043067, Regulation of programmed cell death	RRM2B, ACVR1, TP53, RASA1, TGFBR1, PSMA3, BIRC5, ACTN2, HOXA13, IRS2, FASTK, VAV1, PSMB6, BCL2, CDK1, HDAC1, SOX10, TIA1, AKT1, AURKB	3.43×10^{-8}
GO:0043069, Negative regulation of programmed cell death	RRM2B, ACVR1, TP53, RASA1, TGFBR1, BIRC5, IRS2, BCL2, CDK1, HDAC1, SOX10, AKT1, AURKB	6.58×10^{-6}
GO:0060548, Negative regulation of cell death	RRM2B, ACVR1, TP53, RASA1, TGFBR1, BIRC5, IRS2, BCL2, CDK1, HDAC1, SOX10, AKT1, AURKB	8.92×10^{-6}
GO:0015630, Microtubule cytoskeleton	INCENP, SNTB2, SEPT1, TACC1, BIRC5, RACGAP1, PIN4, CDCA8, CDK1, PHF1, AKT1, AURKB, NINL, CCDC85B	5.15×10^{-5}

We also found that some breast cancer-specific functions were enriched in our results. For example, in the miR-497-5p-regulated PIN, the term “androgen receptor signaling pathway” was enriched. Although it is not clear whether androgens are related to breast cancer, androgen receptors are known to be up-regulated in breast cancer and related to node invasiveness [37].

To further verify the results of the enriched cancer-related functions, we used GOBO for survival analysis. Our hypothesis is that expression of genes annotated with enriched cancer-related terms may be related to survival outcome of patients. With exception to some cell death/proliferation-related terms, it is already known that some pathways or functions are also related to clinical outcomes. For example, cell proliferation-related GO terms have a high probability of affecting survival of cancer tissues, and patient outcome may worsen if cancer tissue survives. In addition, some signaling pathways were known to enhance invasiveness, migration abilities, or were associated with reduced patient survival. For example, the BMP signaling pathway is known to confer various tumor cells with enhanced migration and invasion abilities [38], and nerve growth factor receptor (NGFR) was found to be associated with overall survival of breast cancer [38]. Furthermore, Toll-like receptor 4 has been reported to promote adhesion and invasive migration in breast cancer [39]. Finally, the cytoskeleton plays an important role in regulating cell motility in all cells. Actin filaments are known to participate in the invasive migration of cancer cells [40]. Since some of these functions were present in our enriched terms, we wished to test if the expression of a gene set annotated to the cancer-related enriched terms in the PIN would be related to clinical outcome of patients.

Figure 4. Validation of let-7c result. Gene expression-based outcome for breast cancer online (GOBO) survival analysis of let-7c-regulated PIN members marked with the following functions: (A) Microtubule cytoskeleton; (B) Negative regulation of programmed cell death; and (C) negative regulation of cell death. **Red:** samples with high expression of selected gene set (PIN members); **grey:** samples with low expression of selected gene set (PIN members).



As shown in Figure 4, Figure S14 and Table S7, only some of the enriched terms were significantly associated with clinical outcome. This may be because the changes in these key genes occurred at the protein level, such as in protein expression or even post-translational modification; therefore, mRNA expression-centric tools like GOBO cannot explore association of such genes to clinical outcomes.

Alternatively, it is possible the miRNA did not regulate the whole pathway, or the miRNA did not target the key part of the pathway directly, and thus the clinical outcome of gene sets of the enriched GO terms in this condition cannot be determined. However, some functions associated with clinical outcomes were observed. For example, proteins annotated with the terms “microtubule cytoskeleton”, “negative regulation of programmed cell death”, and “negative regulation of cell death” in the let-7c-regulated PIN were related to 10 year survival rate of patients, as reported by GOBO (Figure 4). Also, the enriched term “regulation of epithelial cell proliferation” for both miR-125a-5p and miR-125b-5p were found to be associated with the 10-year survival rate of patients. Therefore, these results further supported the GO enrichment analysis discussed previously.

3. Experimental Section

3.1. miRNA Microarray Experiments

We performed a miRNA microarray to obtain the expression profiles for receiver operating characteristic (ROC) curve analysis. This dataset was deposited in Gene Expression Omnibus (GEO, <http://www.ncbi.nlm.nih.gov/geo/>, Series Accession: GSE45666). In total, there were 15 normal samples and 101 tumor samples in the expression profile. Detailed pathophysiological characteristics of these samples were in Table S8 and GSE45666. All human tissue samples collected from breast cancer patients were approved and human subject confidentiality was protected by the Institute Review Board of National Taiwan University Hospital (IRB, 20071211R).

Total RNA was extracted from tissues collected from the patients using Trizol® Reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer’s protocol. Purified RNA was quantified at OD260nm by using a ND-1000 spectrophotometer (Nanodrop Technology, Wilmington, DE, USA) and qualitated by using a Bioanalyzer 2100 with the RNA 6000 Nano LabChip kit (Agilent Technologies, Santa Clara, CA, USA).

After RNA extraction, 100 ng of total RNA was dephosphorylated and labeled with pCp-Cy3 using the Agilent miRNA Complete Labeling and Hyb Kit in conjunction with the microRNA Spike-In kit (Agilent Technologies, Santa Clara, CA, USA). Briefly, 2X hybridization buffer (Agilent Technologies) was added to the labeled mixture to a final volume of 45 µL. The mixture was heated for 5 min at 100 °C and immediately cooled to 0 °C. Each 45 µL sample was hybridized onto an Agilent human miRNA Microarray Release 12.0, 8 × 15 K (Agilent Technologies) at 55 °C for 20 h. After hybridization, slides were washed for 5 min in Gene Expression Wash Buffer 1 at room temperature, then for 5 min in Gene Expression Wash Buffer 2 at 37 °C. Slides were scanned on an Agilent microarray scanner (Agilent Technologies, model G2505C) at 100% and 5% sensitivity settings. Feature Extraction (Agilent Technologies) software version 10.7.3.1 (Agilent Technologies, Santa Clara, CA, USA) was used for image analysis.

3.2. mRNA Expression Profiles

For miRNA-regulated protein-protein interaction network construction, the mRNA expression profile was fetched from GEO (Series Accession: GSE29174). This dataset was produced by Farazi *et al.* [26], which was the only dataset publicly available with large size of tumor samples and

reasonably sized normal samples. In total, 161 clinical samples were collected from breast cancer patients by biopsy: 110 invasive ductal carcinoma (IDC), 11 normal, 17 ductal carcinoma *in situ* (DCIS), 1 mucinous A, 8 atypical medullary, 4 apocrine, 8 metaplastic, and 2 adenoid, as classified by Farazi *et al.* The 110 IDC samples were classified as the tumor group and the 11 normal samples were classified as the normal group in this study.

3.3. miRNA Expression Profiles

The miRNA expression profile used in this work was fetched from Table S4A of the work of Farazi *et al.* [26]. There were 189 samples in this dataset, with 6 of them from cell lines, and another 183 samples from patient tissues. In the 183 clinical samples collected from breast cancer patients by biopsy in this dataset, there were 128 IDC, 11 normal, 18 DCIS, 1 mucinous A, 8 atypical medullary, 4 apocrine, 9 metaplastic, and 2 adenoid cases, as classified by Farazi *et al.* The 128 IDC samples were classified as the tumor group and the 11 normal samples as were classified as the normal group.

3.4. Data Analysis

The overall workflow design was similar to the previous work of Tseng *et al.* [25] (see Figure 1). However, we applied the workflow on breast cancer expression profiles instead of gastric cancer as in the work of Tseng *et al.* Additionally, we used 3 miRNA target prediction databases, TargetScan (v6.0) [22,41,42], PicTar [43,44], and miRanda (release August 2010) [45] here, while only TargetScan was used in the previous study.

To construct the networks, we first elucidated differentially expressed miRNAs and mRNAs from published datasets. Both of miRNA (generated with miRNA-Seq technique) and mRNA array expression data were produced by Farazi *et al.* [26] from the same batch of clinical tumor samples. To obtain a list of differentially expressed genes and miRNAs between normal and tumor groups, we used significance analysis of microarrays (SAM) [46] implemented in R package samr (version 2.0; Stanford University, Stanford, CA, USA). We set the false discovery rate (FDR) as $\leq 0.0001\%$, fold change as ≥ 2.5 for miRNA, and fold change as ≥ 1.9 for genes as thresholds to reduce false positives. If a gene/miRNA was expressed higher in the normal group compared to the tumor group, we defined that gene/miRNA as a down-regulated gene/miRNA, and *vice versa*.

Following this, we paired the miRNAs and mRNAs with different expression trends. For example, an up-regulated miRNA would be paired with a down-regulated mRNA. If such pairs could be found in 2 (or more) of the 3 miRNA target prediction databases, they were then added to their corresponding miRNA-regulated network. To further extend the coverage of our network, we incorporated the human protein reference database (HPRD) [47], which contains experimentally verified interaction data, into our miRNA-regulated PINs.

Finally, we used gene ontology (GO) enrichment analysis to explore the function of the miRNA-regulated PINs. Hypergeometric tests were used to determine if a GO term was enriched in a PIN. In this section, we excluded PINs with less than 5 proteins. We also excluded GO terms with levels of less than 5 to avoid non-specific GO terms. Since we tested multiple GO terms on each miRNA-regulated PIN, we adjusted the significance of the test with the FDR method developed by Benjamini *et al.* [48]. We used the adjusted p -value < 0.0001 as our threshold. A GO term would

be excluded if its *p*-value was larger than 0.0001. Therefore, for each network, we ran the GO enrichment analysis, collected the calculated *p*-values, and adjusted these values using the methods described above.

3.5. ROC and GOBO Survival Analysis

After the PINs were constructed, we attempted to verify our results by literature search, ROC, and GOBO survival analysis [49]. To determine if the miRNAs we found could serve as classification markers for discriminating between normal and tumor samples, we applied ROC analysis on our miRNA array data (described in Section 2.1). ROC analysis is usually used to evaluate the efficiency of a classifier or a biological marker. R package ROCR [50] (version 1.0.4) was used to plot the ROC curve and calculate the area under curve (AUC). The standard error of AUC was then calculated as described in the work of Hanley and McNeil [51]. The *p*-value of AUC was thus calculated with standard error obtained in the previous step. To further validate if the PINs we found were related to cancer, we used survival analysis implemented in GOBO [49] (available at <http://co.bmc.lu.se/gobo>), which provides a large amount of breast cancer gene expression profiles collected from public databases with clinical outcome data. In both ROC analysis and GOBO survival analysis, we considered our results significant when the *p*-value was smaller than 0.05.

4. Conclusions

Using integrative analysis of miRNA and mRNA expression profiles, we have identified not only breast cancer-related miRNAs and genes, but also putative roles for miRNAs in cancer as elucidated from miRNA-regulated PINs constructed in this work. Here, some previously known functions of miRNAs were again presented in our results, e.g., the relationship between the miRNAs, let-7c, miR-125a-5p, miR-125b-5p, and miR-21-5p, and breast cancer were demonstrated in this research. Furthermore, we have identified additional miRNAs and their related functions that have not been previously reported or discussed, providing valuable resources for further research in breast cancer.

Acknowledgments

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Conflict of Interest

The authors declare no conflict of interest.

Supplementary Information

Table S1. Significantly differentially expressed miRNAs found in miRNA dataset in Farazi *et al.* [26]. There are 89 down-regulated miRNAs and 1 up-regulated miRNA in this list. *Q*-values reported by SAM were 0 for all miRNAs in this list.

miRBase Accession	miRNA Name	Fold Change	miRBase Accession	miRNA Name	Fold Change
MIMAT0004761	hsa-miR-483-5p	0.01	MIMAT0000077	hsa-miR-22-3p	0.21
MIMAT0004552	hsa-miR-139-3p	0.01	MIMAT0000089	hsa-miR-31-5p	0.21
MIMAT0000738	hsa-miR-383	0.02	MIMAT0004612	hsa-miR-186-3p	0.21
MIMAT0002856	hsa-miR-520d-3p	0.02	MIMAT0004592	hsa-miR-125b-1-3p	0.22
MIMAT0002811	hsa-miR-202-3p	0.03	MIMAT0001639	hsa-miR-409-3p	0.22
MIMAT0002177	hsa-miR-486-5p	0.04	MIMAT0015032	hsa-miR-3158-3p	0.22
MIMAT0022721	hsa-miR-1247-3p	0.05	MIMAT0004496	hsa-miR-23a-5p	0.22
MIMAT0002175	hsa-miR-485-5p	0.06	MIMAT0000690	hsa-miR-296-5p	0.22
MIMAT0000265	hsa-miR-204-5p	0.07	MIMAT0000731	hsa-miR-378a-5p	0.23
MIMAT0000752	hsa-miR-328	0.07	MIMAT0000448	hsa-miR-136-5p	0.23
MIMAT0000421	hsa-miR-122-5p	0.07	MIMAT0004796	hsa-miR-576-3p	0.23
MIMAT0000447	hsa-miR-134	0.08	MIMAT0010133	hsa-miR-2110	0.23
MIMAT0000722	hsa-miR-370	0.09	MIMAT0004951	hsa-miR-887	0.23
MIMAT0004513	hsa-miR-101-5p	0.09	MIMAT0003239	hsa-miR-574-3p	0.25
MIMAT0000446	hsa-miR-127-3p	0.10	MIMAT0005901	hsa-miR-1249	0.25
MIMAT0000097	hsa-miR-99a-5p	0.10	MIMAT0000510	hsa-miR-320a	0.26
MIMAT0004566	hsa-miR-218-2-3p	0.10	MIMAT0002172	hsa-miR-376b	0.26
MIMAT0000729	hsa-miR-376a-3p	0.11	MIMAT0000250	hsa-miR-139-5p	0.27
MIMAT0009197	hsa-miR-205-3p	0.11	MIMAT0005825	hsa-miR-1180	0.27
MIMAT0004615	hsa-miR-195-3p	0.11	MIMAT0000437	hsa-miR-145-5p	0.28
MIMAT0005899	hsa-miR-1247-5p	0.11	MIMAT0004601	hsa-miR-145-3p	0.28
MIMAT0000720	hsa-miR-376c	0.12	MIMAT0003322	hsa-miR-652-3p	0.28
MIMAT0000762	hsa-miR-324-3p	0.12	MIMAT0000756	hsa-miR-326	0.28
MIMAT0004679	hsa-miR-296-3p	0.12	MIMAT0000098	hsa-miR-100-5p	0.29
MIMAT0004614	hsa-miR-193a-5p	0.12	MIMAT0003296	hsa-miR-627	0.29
MIMAT0003880	hsa-miR-671-5p	0.12	MIMAT0002820	hsa-miR-497-5p	0.31
MIMAT0004795	hsa-miR-574-5p	0.12	MIMAT0004507	hsa-miR-92a-1-5p	0.31
MIMAT0004599	hsa-miR-143-5p	0.13	MIMAT0000271	hsa-miR-214-3p	0.32
MIMAT0000423	hsa-miR-125b-5p	0.13	MIMAT0004702	hsa-miR-339-3p	0.33
MIMAT0004957	hsa-miR-760	0.13	MIMAT0004611	hsa-miR-185-3p	0.33
MIMAT0004911	hsa-miR-874	0.14	MIMAT0000064	hsa-let-7c	0.34
MIMAT0004603	hsa-miR-125b-2-3p	0.15	MIMAT0004673	hsa-miR-29c-5p	0.35
MIMAT0004952	hsa-miR-665	0.15	MIMAT0000733	hsa-miR-379-5p	0.35
MIMAT0018205	hsa-miR-3928	0.15	MIMAT0004594	hsa-miR-132-5p	0.35
MIMAT0004767	hsa-miR-193b-5p	0.15	MIMAT0000765	hsa-miR-335-5p	0.35
MIMAT0002861	hsa-miR-518e-3p	0.15	MIMAT0002819	hsa-miR-193b-3p	0.36
MIMAT0004604	hsa-miR-127-5p	0.16	MIMAT0000088	hsa-miR-30a-3p	0.36
MIMAT0002807	hsa-miR-491-5p	0.16	MIMAT0005951	hsa-miR-1307-3p	0.36
MIMAT0004689	hsa-miR-377-5p	0.16	MIMAT0004597	hsa-miR-140-3p	0.37
MIMAT0004762	hsa-miR-486-3p	0.16	MIMAT0004556	hsa-miR-10b-3p	0.37
MIMAT0000732	hsa-miR-378a-3p	0.17	MIMAT0000272	hsa-miR-215	0.37
MIMAT0017981	hsa-miR-3605-5p	0.18	MIMAT0004511	hsa-miR-99a-3p	0.37
MIMAT0004605	hsa-miR-129-2-3p	0.19	MIMAT0000443	hsa-miR-125a-5p	0.38
MIMAT0006789	hsa-miR-1468	0.20	MIMAT0004482	hsa-let-7b-3p	0.38
MIMAT0000737	hsa-miR-382-5p	0.21	MIMAT0000076	hsa-miR-21-5p	6.58

Table S2. Down-regulated genes found in dataset GSE29174. There are 726 down-regulated genes in this list. *Q*-values reported by SAM were 0 for all genes in this list.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
2949	GSTM5	0.06	619373	MBOAT4	0.17
10894	LYVE1	0.06	130399	ACVR1C	0.17
5950	RBP4	0.07	1646	AKR1C2	0.17
762	CA4	0.09	80763	C12orf39	0.17
54997	TESC	0.09	2159	F10	0.18
3489	IGFBP6	0.09	84889	SLC7A3	0.18
3952	LEP	0.09	1308	COL17A1	0.18
213	ALB	0.09	83699	SH3BGRL2	0.18
3131	HLF	0.10	84417	C2orf40	0.18
4023	LPL	0.10	4081	MAB21L1	0.18
10633	RASL10A	0.11	3484	IGFBP1	0.18
364	AQP7	0.11	5239	PGM5	0.19
1908	EDN3	0.11	4969	OGN	0.19
1811	SLC26A3	0.11	2719	GPC3	0.19
91851	CHRDL1	0.11	116362	RBP7	0.19
729359	PLIN4	0.13	948	CD36	0.19
1149	CIDEA	0.13	5764	PTN	0.19
5959	RDH5	0.13	3043	HBB	0.19
5348	FXYD1	0.14	56920	SEMA3G	0.20
5346	PLIN1	0.14	94274	PPP1R14A	0.20
10249	GLYAT	0.14	57447	NDRG2	0.20
158800	RHOXF1	0.14	84795	PYROXD2	0.20
221476	PI16	0.14	84649	DGAT2	0.20
3040	HBA2	0.14	2690	GHR	0.20
6939	TCF15	0.14	22802	CLCA4	0.20
79645	EFCAB1	0.14	5179	PENK	0.20
80343	SEL1L2	0.14	6663	SOX10	0.20
9413	FAM189A2	0.15	6649	SOD3	0.21
26289	AK5	0.15	54922	RASIP1	0.21
25891	PAMR1	0.15	8406	SRPX	0.21
3679	ITGA7	0.15	1446	CSN1S1	0.21
1264	CNN1	0.15	7123	CLEC3B	0.22
92304	SCGB3A1	0.15	9647	PPM1F	0.22
2167	FABP4	0.15	1842	ECM2	0.22
23285	KIAA1107	0.15	3909	LAMA3	0.22
7145	TNS1	0.16	8639	AOC3	0.23
4881	NPR1	0.16	2934	GSN	0.23
1028	CDKN1C	0.16	9370	ADIPOQ	0.23
1036	CDO1	0.16	3202	HOXA5	0.23
130271	PLEKHH2	0.16	9452	ITM2A	0.23
8736	MYOM1	0.16	6290	SAA3P	0.23
8908	GYG2	0.16	4604	MYBPC1	0.23

Table S2. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
79785	RERGL	0.16	1128	CHRM1	0.23
221091	LRRN4CL	0.17	83878	USHBP1	0.24
3991	LIPE	0.17	63970	TP53AIP1	0.24
27175	TUBG2	0.24	79192	IRX1	0.28
1346	COX7A1	0.24	3400	ID4	0.28
6376	CX3CL1	0.24	57519	STARD9	0.29
50486	G0S2	0.24	57666	FBRSL1	0.29
6285	S100B	0.24	3590	IL11RA	0.29
443	ASPA	0.24	57664	PLEKHA4	0.29
947	CD34	0.25	197257	LDHD	0.29
84632	AFAP1L2	0.25	66036	MTMR9	0.29
3866	KRT15	0.25	2321	FLT1	0.29
147463	ANKRD29	0.25	126	ADH1C	0.29
2878	GPX3	0.25	1363	CPE	0.29
7079	TIMP4	0.25	56131	PCDHB4	0.29
54345	SOX18	0.25	22915	MMRN1	0.29
51277	DNAJC27	0.25	7069	THRSP	0.29
84870	RSPO3	0.25	57161	PELI2	0.30
55323	LARP6	0.25	770	CA11	0.30
6387	CXCL12	0.25	53342	IL17D	0.30
137835	TMEM71	0.25	79987	SVEP1	0.30
5212	VIT	0.25	857	CAV1	0.30
26577	PCOLCE2	0.25	222166	C7orf41	0.30
845	CASQ2	0.25	27190	IL17B	0.30
6422	SFRP1	0.25	116159	CYYR1	0.30
10351	ABCA8	0.26	4487	MSX1	0.30
10840	ALDH1L1	0.26	9068	ANGPTL1	0.30
65983	GRAMD3	0.26	10411	RAPGEF3	0.30
84327	ZBED3	0.26	3199	HOXA2	0.30
57124	CD248	0.26	2944	GSTM1	0.30
3235	HOXD9	0.26	2920	CXCL2	0.30
2192	FBLN1	0.26	201134	CEP112	0.31
91653	BOC	0.26	220001	VWCE	0.31
4147	MATN2	0.26	83888	FGFBP2	0.31
126669	SHE	0.27	6366	CCL21	0.31
2788	GNG7	0.27	6711	SPTBN1	0.31
129804	FBLN7	0.27	85378	TUBGCP6	0.31
270	AMPD1	0.27	26040	SETBP1	0.31
79656	BEND5	0.27	4692	NDN	0.31
58503	PROL1	0.27	25890	ABI3BP	0.31
3316	HSPB2	0.27	23531	MMD	0.31
729440	CCDC61	0.27	30846	EHD2	0.31
54438	GFOD1	0.27	6196	RPS6KA2	0.31
5243	ABCB1	0.27	2009	EML1	0.31

Table S2. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
810	CALML3	0.27	6289	SAA2	0.31
6898	TAT	0.27	345275	HSD17B13	0.31
5648	MASP1	0.28	2701	GJA4	0.32
25999	CLIP3	0.28	112609	MRAP2	0.32
125875	CLDND2	0.28	727	C5	0.32
7102	TSPAN7	0.28	477	ATP1A2	0.32
1879	EBF1	0.28	9627	SNCAIP	0.32
23252	OTUD3	0.28	4435	CITED1	0.32
5493	PPL	0.28	10974	C10orf116	0.32
83987	CCDC8	0.28	11005	SPINK5	0.32
9073	CLDN8	0.28	80325	ABTB1	0.33
221981	THSD7A	0.28	221395	GPR116	0.33
64102	TNMD	0.28	10014	HDAC5	0.33
137872	ADHFE1	0.33	1489	CTF1	0.37
27151	CPAMD8	0.33	35	ACADS	0.37
387923	SERP2	0.33	3749	KCNC4	0.37
145581	LRFN5	0.33	140738	TMEM37	0.37
6263	RYR3	0.33	2791	GNG11	0.37
2354	FOSB	0.33	23604	DAPK2	0.37
51302	CYP39A1	0.33	10217	CTDSPL	0.37
4128	MAOA	0.34	23550	PSD4	0.37
117248	GALNTL2	0.34	4306	NR3C2	0.37
10268	RAMP3	0.34	119587	CPXM2	0.37
7730	ZNF177	0.34	7942	TFEB	0.37
10873	ME3	0.34	3815	KIT	0.37
7461	CLIP2	0.34	1805	DPT	0.37
7049	TGFBR3	0.34	23242	COBL	0.37
79901	CYBRD1	0.34	4313	MMP2	0.37
5152	PDE9A	0.34	4139	MARK1	0.37
50805	IRX4	0.34	9104	RGN	0.37
8644	AKR1C3	0.34	2329	FMO4	0.37
5915	RARB	0.34	25802	LMOD1	0.38
2770	GNAI1	0.34	4239	MFAP4	0.38
54996	2-Mar	0.35	10392	NOD1	0.38
79791	FBXO31	0.35	6794	STK11	0.38
54776	PPP1R12C	0.35	85458	DIXDC1	0.38
9079	LDB2	0.35	4123	MAN2C1	0.38
57104	PNPLA2	0.35	54476	RNF216	0.38
30008	EFEMP2	0.35	9920	KBTBD11	0.38
91461	PKDCC	0.35	6329	SCN4A	0.38
23368	PPP1R13B	0.35	10253	SPRY2	0.38
23461	ABCA5	0.35	1910	EDNRB	0.38
9572	NR1D1	0.35	9249	DHRS3	0.38
23338	PHF15	0.35	22869	ZNF510	0.38

Table S2. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
114800	CCDC85A	0.35	3384	ICAM2	0.38
2550	GABBR1	0.35	8613	PPAP2B	0.38
4638	MYLK	0.35	1950	EGF	0.38
2327	FMO2	0.35	55273	TMEM100	0.38
139411	PTCHD1	0.35	6297	SALL2	0.38
10391	CORO2B	0.35	9365	KL	0.38
25854	FAM149A	0.35	8863	PER3	0.38
55701	ARHGEF40	0.36	8404	SPARCL1	0.38
1759	DNM1	0.36	2202	EFEMP1	0.38
22849	CPEB3	0.36	8369	HIST1H4G	0.38
57716	PRX	0.36	5187	PER1	0.39
1628	DBP	0.36	30815	ST6GALNAC6	0.39
80031	SEMA6D	0.36	256364	EML3	0.39
259217	HSPA12A	0.36	57381	RHOJ	0.39
6909	TBX2	0.36	761	CA3	0.39
1511	CTSG	0.36	83989	FAM172A	0.39
79971	WLS	0.36	1408	CRY2	0.39
90865	IL33	0.36	2281	FKBP1B	0.39
11343	MGLL	0.36	51222	ZNF219	0.39
55800	SCN3B	0.36	54540	FAM193B	0.39
1949	EFNB3	0.36	4053	LTBP2	0.39
284217	LAMA1	0.36	55184	DZANK1	0.39
22927	HABP4	0.37	5740	PTGIS	0.39
23645	PPP1R15A	0.39	84814	PPAPDC3	0.42
342574	KRT27	0.39	79365	BHLHE41	0.42
83543	AIF1L	0.39	316	AOX1	0.42
624	BDKRB2	0.39	23380	SRGAP2	0.42
347	APOD	0.39	84033	OBSCN	0.42
84935	C13orf33	0.39	90353	CTU1	0.42
858	CAV2	0.39	9013	TAF1C	0.42
5138	PDE2A	0.40	474344	GIMAP6	0.42
114928	GPRASP2	0.40	84883	AIFM2	0.42
58190	CTDSP1	0.40	58480	RHOU	0.42
513	ATP5D	0.40	65982	ZSCAN18	0.42
57684	ZBTB26	0.40	666	BOK	0.42
7041	TGFB1I1	0.40	79762	C1orf115	0.42
5787	PTPRB	0.40	525	ATP6V1B1	0.42
7294	TXK	0.40	4675	NAP1L3	0.42
56301	SLC7A10	0.40	3257	HPS1	0.43
55937	APOM	0.40	55781	RIOK2	0.43
6368	CCL23	0.40	63947	DMRTC1	0.43
55020	TTC38	0.40	1969	EPHA2	0.43
134265	AFAP1L1	0.40	25927	CNRIP1	0.43
4485	MST1	0.40	57685	CACHD1	0.43

Table S2. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
51559	NT5DC3	0.40	29997	GLTSCR2	0.43
7169	TPM2	0.40	26051	PPP1R16B	0.43
51705	EMCN	0.40	83604	TMEM47	0.43
8938	BAIAP3	0.40	2308	FOXO1	0.43
10365	KLF2	0.40	55225	RAVER2	0.43
59	ACTA2	0.40	54839	LRRC49	0.43
80309	SPHKAP	0.40	122953	JDP2	0.43
3779	KCNMB1	0.41	29775	CARD10	0.43
10826	C5orf4	0.41	166	AES	0.43
219654	ZCCHC24	0.41	25924	MYRIP	0.43
92162	TMEM88	0.41	2852	GPER	0.43
7450	VWF	0.41	51421	AMOTL2	0.43
10266	RAMP2	0.41	124936	CYB5D2	0.43
25875	LETMD1	0.41	1294	COL7A1	0.43
1938	EEF2	0.41	127435	PODN	0.43
121551	BTBD11	0.41	84952	CGNL1	0.43
2119	ETV5	0.41	83483	PLVAP	0.43
9696	CROCC	0.41	1958	EGR1	0.43
1031	CDKN2C	0.41	230	ALDOC	0.43
9037	SEMA5A	0.41	65987	KCTD14	0.43
3397	ID1	0.41	4804	NGFR	0.44
84707	BEX2	0.41	64852	TUT1	0.44
57616	TSHZ3	0.41	84253	GARNL3	0.44
1471	CST3	0.41	5866	RAB3IL1	0.44
55214	LEPREL1	0.41	10608	MXD4	0.44
3914	LAMB3	0.41	4211	MEIS1	0.44
57478	USP31	0.41	83547	RILP	0.44
3783	KCNN4	0.41	9172	MYOM2	0.44
8839	WISP2	0.41	57192	MCOLN1	0.44
1583	CYP11A1	0.42	255877	BCL6B	0.44
10124	ARL4A	0.42	56904	SH3GLB2	0.44
738	C11orf2	0.42	51285	RASL12	0.44
29800	ZDHHC1	0.42	3425	IDUA	0.44
23135	KDM6B	0.44	402117	VWC2L	0.46
171024	SYNPO2	0.44	81490	PTDSS2	0.46
10350	ABCA9	0.44	283748	PLA2G4D	0.46
3691	ITGB4	0.44	23523	CABIN1	0.46
2348	FOLR1	0.44	6146	RPL22	0.46
11145	PLA2G16	0.44	85360	SYDE1	0.46
554	AVPR2	0.45	60468	BACH2	0.46
64072	CDH23	0.45	57451	ODZ2	0.46
80177	MYCT1	0.45	4013	VWA5A	0.46
5957	RCVRN	0.45	339768	ESPNL	0.46
408	ARRB1	0.45	3860	KRT13	0.46

Table S2. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
144699	FBXL14	0.45	7094	TLN1	0.46
83719	YPEL3	0.45	4232	MEST	0.46
22841	RAB11FIP2	0.45	1410	CRYAB	0.46
283927	NUDT7	0.45	57452	GALNTL1	0.47
293	SLC25A6	0.45	63935	PCIF1	0.47
90507	SCRN2	0.45	25873	RPL36	0.47
37	ACADVL	0.45	9812	KIAA0141	0.47
112744	IL17F	0.45	51665	ASB1	0.47
6709	SPTAN1	0.45	64123	ELTD1	0.47
8086	AAAS	0.45	6122	RPL3	0.47
7423	VEGFB	0.45	222962	SLC29A4	0.47
64221	ROBO3	0.45	23102	TBC1D2B	0.47
7273	TTN	0.45	3476	IGBP1	0.47
2657	GDF1	0.45	93408	MYL10	0.47
59271	C21orf63	0.45	5310	PKD1	0.47
132160	PPM1M	0.45	4628	MYH10	0.47
27244	SESN1	0.45	221935	SDK1	0.47
51310	SLC22A17	0.45	23328	SASH1	0.47
4828	NMB	0.45	8522	GAS7	0.47
54360	CYTL1	0.45	10023	FRAT1	0.47
203245	NAIF1	0.45	7301	TYRO3	0.47
23166	STAB1	0.45	2767	GNA11	0.47
2121	EVC	0.45	9457	FHL5	0.47
116496	FAM129A	0.45	4094	MAF	0.47
23239	PHLPP1	0.45	65268	WNK2	0.47
51673	TPPP3	0.45	54585	LZTFL1	0.47
64094	SMOC2	0.45	375449	MAST4	0.47
6383	SDC2	0.45	138311	FAM69B	0.47
2180	ACSL1	0.45	160622	GRASP	0.47
23770	FKBP8	0.45	22837	COBLL1	0.47
55901	THSD1	0.46	51435	SCARA3	0.47
25895	METTL21B	0.46	217	ALDH2	0.47
23731	C9orf5	0.46	6236	RRAD	0.47
126393	HSPB6	0.46	8322	FZD4	0.47
4056	LTC4S	0.46	653275	CFC1B	0.47
79825	CCDC48	0.46	10908	PNPLA6	0.47
10810	WASF3	0.46	57526	PCDH19	0.47
29911	HOOK2	0.46	8424	BBOX1	0.47
583	BBS2	0.46	9905	SGSM2	0.48
28984	C13orf15	0.46	10435	CDC42EP2	0.48
1465	CSRP1	0.46	23087	TRIM35	0.48
55258	THNSL2	0.46	60314	C12orf10	0.48
161198	CLEC14A	0.46	1073	CFL2	0.48
3699	ITIH3	0.48	5256	PHKA2	0.49

Table S2. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
92922	CCDC102A	0.48	6237	RRAS	0.49
65057	ACD	0.48	5288	PIK3C2G	0.49
9095	TBX19	0.48	10252	SPRY1	0.49
6441	SFTPД	0.48	79026	AHNAK	0.49
22846	VASH1	0.48	9693	RAPGEF2	0.49
51066	C3orf32	0.48	51226	COPZ2	0.49
23179	RGL1	0.48	158326	FREM1	0.49
4664	NAB1	0.48	1956	EGFR	0.49
50511	SYCP3	0.48	5360	PLTP	0.49
6430	SRSF5	0.48	290	ANPEP	0.49
11078	TRIOBP	0.48	1756	DMD	0.49
78991	PCYOX1L	0.48	5118	PCOLCE	0.49
6623	SNCG	0.48	56654	NPDC1	0.49
23384	SPECC1L	0.48	9254	CACNA2D2	0.49
53826	FXYD6	0.48	55536	CDCA7L	0.49
9397	NMT2	0.48	124975	GGT6	0.49
6041	RNASEL	0.48	1906	EDN1	0.49
113510	HELQ	0.48	81029	WNT5B	0.49
64788	LMF1	0.48	2646	GCKR	0.49
2217	FCGRT	0.48	9811	CTIF	0.50
79720	VPS37B	0.48	145376	PPP1R36	0.50
6764	ST5	0.48	222865	TMEM130	0.50
252969	NEIL2	0.48	92999	ZBTB47	0.50
8987	STBD1	0.48	168002	DACT2	0.50
41	ACCN2	0.48	6829	SUPT5H	0.50
7905	REEP5	0.48	9992	KCNE2	0.50
5919	RARRES2	0.48	58509	C19orf29	0.50
10544	PROCR	0.48	79706	PRKRIP1	0.50
6876	TAGLN	0.48	1153	CIRBP	0.50
8436	SDPR	0.49	9639	ARHGEF10	0.50
23500	DAAM2	0.49	4054	LTBP3	0.50
130132	RFTN2	0.49	1120	CHKB	0.50
80310	PDGFD	0.49	286046	XKR6	0.50
4215	MAP3K3	0.49	9590	AKAP12	0.50
282775	OR5J2	0.49	64115	C10orf54	0.50
51161	C3orf18	0.49	2067	ERCC1	0.50
29098	RANGRF	0.49	7507	XPA	0.50
53336	CPXCR1	0.49	22897	CEP164	0.50
9081	PRY	0.49	652	BMP4	0.50
9459	ARHGEF6	0.49	55702	CCDC94	0.50
2995	GYPC	0.49	57613	KIAA1467	0.50
23057	NMNAT2	0.49	28514	DLL1	0.50
4669	NAGLU	0.49	169270	ZNF596	0.50
6452	SH3BP2	0.49	83982	IFI27L2	0.50

Table S2. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
51458	RHCG	0.49	2247	FGF2	0.50
1112	FOXN3	0.49	26248	OR2K2	0.50
29954	POMT2	0.49	84303	CHCHD6	0.50
9612	NCOR2	0.49	3615	IMPDH2	0.50
3198	HOXA1	0.49	1813	DRD2	0.50
5311	PKD2	0.49	80148	PQLC1	0.50
2946	GSTM2	0.49	390081	OR52E4	0.50
2109	ETFB	0.49	352954	GATS	0.50
56062	KLHL4	0.49	90871	C9orf123	0.50
6915	TBXA2R	0.50	50945	TBX22	0.52
64288	ZNF323	0.50	5204	PFDN5	0.52
5195	PEX14	0.50	5338	PLD2	0.52
84557	MAP1LC3A	0.50	94	ACVRL1	0.52
6164	RPL34	0.50	54039	PCBP3	0.52
8835	SOCS2	0.50	7691	ZNF132	0.52
2735	GLI1	0.50	338	APOB	0.52
26022	TMEM98	0.50	84658	EMR3	0.52
3908	LAMA2	0.50	283232	TMEM80	0.52
1825	DSC3	0.50	5430	POLR2A	0.52
5730	PTGDS	0.50	54623	PAF1	0.52
162515	SLC16A11	0.51	11070	TMEM115	0.52
274	BIN1	0.51	10395	DLC1	0.52
79654	HECTD3	0.51	57140	RNPEPL1	0.52
22863	ATG14	0.51	79781	IQCA1	0.52
25949	SYF2	0.51	1838	DTNB	0.52
84872	ZC3H10	0.51	51386	EIF3L	0.52
23187	PHLDB1	0.51	56919	DHX33	0.52
5434	POLR2E	0.51	57542	KLHDC5	0.52
6181	RPLP2	0.51	3628	INPP1	0.52
6141	RPL18	0.51	4520	MTF1	0.52
84747	UNC119B	0.51	8547	FCN3	0.52
23399	CTDNEP1	0.51	60401	EDA2R	0.52
599	BCL2L2	0.51	8082	SSPN	0.52
197258	FUK	0.51	80755	AARSD1	0.52
5207	PFKFB1	0.51	710	SERPING1	0.52
8131	NPRL3	0.51	56246	MRAP	0.52
25839	COG4	0.51	10555	AGPAT2	0.52
10816	SPINT3	0.51	949	SCARB1	0.52
60485	SAV1	0.51	23743	BHMT2	0.52
5681	PSKH1	0.51	3910	LAMA4	0.52
80318	GKAP1	0.51	60370	AVPI1	0.52
57088	PLSCR4	0.51	5021	OXTR	0.52
93129	ORA13	0.51	55997	CFC1	0.52
5829	PXN	0.51	23144	ZC3H3	0.52

Table S2. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
56776	FMN2	0.51	150709	ANKAR	0.52
85456	TNKS1BP1	0.51	6591	SNAI2	0.52
283	ANG	0.51	10129	FRY	0.52
7035	TFPI	0.51	5166	PDK4	0.52
51232	CRIM1	0.51	146433	IL34	0.52
112616	CMTM7	0.51	118812	MORN4	0.53
22981	NINL	0.51	10516	FBLN5	0.53
8727	CTNNAL1	0.51	9463	PICK1	0.53
9902	MRC2	0.51	127495	LRRC39	0.53
10900	RUNDYC3A	0.51	7753	ZNF202	0.53
51299	NRN1	0.51	79827	CLMP	0.53
79632	FAM184A	0.52	203260	CCDC107	0.53
80820	EEPDL1	0.52	83657	DYNLRB2	0.53

Table S1. Up-regulated genes found in dataset GSE29174. There are 437 up-regulated genes in this list. *Q*-values reported by SAM were 0 for all genes in this list.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
1300	COL10A1	42.74	54443	ANLN	5.79
3007	HIST1H1D	29.72	6710	SPTB	5.71
8366	HIST1H4B	25.58	7272	TTK	5.64
6286	S100P	25.19	10635	RAD51AP1	5.49
1301	COL11A1	24.72	4069	LYZ	5.37
3627	CXCL10	17.83	55183	RIF1	5.34
4283	CXCL9	15.88	891	CCNB1	5.34
1387	CREBBP	12.83	91543	RSAD2	5.31
27299	ADAMDEC1	12.78	81610	FAM83D	5.24
54986	ULK4	12.46	64581	CLEC7A	5.10
55771	PRR11	12.02	10051	SMC4	5.02
54790	TET2	11.25	4085	MAD2L1	4.96
6241	RRM2	10.60	55872	PBK	4.83
3433	IFIT2	10.49	991	CDC20	4.82
6999	TDO2	9.73	9221	NOLC1	4.74
1656	DDX6	9.72	2124	EVI2B	4.66
55088	C10orf118	9.37	375248	ANKRD36	4.66
9648	GCC2	9.24	1164	CKS2	4.64
6696	SPP1	8.92	1230	CCR1	4.62
2803	GOLGA4	8.57	890	CCNA2	4.56
83540	NUF2	7.73	127933	UHMK1	4.49
10112	KIF20A	7.66	10274	STAG1	4.45
9833	MELK	7.59	597	BCL2A1	4.43
55165	CEP55	7.50	55355	HJURP	4.41
10142	AKAP9	7.44	54210	TREM1	4.36
9447	AIM2	7.42	253558	LCLAT1	4.26

Table S3. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
2706	GJB2	7.33	1033	CDKN3	4.24
6498	SKIL	7.13	79801	SHCBP1	4.23
219285	SAMD9L	7.06	126731	C1orf96	4.21
10261	IGSF6	7.01	6772	STAT1	4.20
2335	FN1	6.95	55729	ATF7IP	4.14
699	BUB1	6.75	6713	SQLE	4.14
1058	CENPA	6.75	157570	ESCO2	4.10
332	BIRC5	6.73	79871	RPAP2	4.09
51203	NUSAP1	6.59	9493	KIF23	4.09
259266	ASPM	6.54	4751	NEK2	4.05
1063	CENPF	6.49	10631	POSTN	4.03
165918	RNF168	6.44	23515	MORC3	4.02
9232	PTTG1	6.34	7153	TOP2A	4.02
5996	RGS1	6.07	10403	NDC80	4.00
29089	UBE2T	5.96	10915	TCERG1	3.99
22974	TPX2	5.94	57650	KIAA1524	3.99
4321	MMP12	5.91	23049	SMG1	3.93
983	CDK1	5.89	80231	CXorf21	3.87
85444	LRRCC1	5.87	5111	PCNA	3.86
29121	CLEC2D	3.83	79682	MLF1IP	3.11
4090	SMAD5	3.80	29123	ANKRD11	3.09
2123	EVI2A	3.80	5429	POLH	3.09
57695	USP37	3.79	701	BUB1B	3.07
133418	EMB	3.76	200030	NBPF11	3.06
4131	MAP1B	3.76	55677	IWS1	3.06
9787	DLGAP5	3.75	160418	TMT3	3.04
9768	KIAA0101	3.74	9147	NEMF	3.04
54625	PARP14	3.73	11320	MGAT4A	3.04
2215	FCGR3B	3.71	5238	PGM3	3.03
9134	CCNE2	3.70	2820	GPD2	3.02
3117	HLA-DQA1	3.68	388886	FAM211B	3.01
10380	BPNT1	3.67	7852	CXCR4	3.00
79056	PRRG4	3.63	57082	CASC5	2.99
10673	TNFSF13B	3.63	22926	ATF6	2.98
8467	SMARCA5	3.61	7594	ZNF43	2.98
115908	CTHRC1	3.61	968	CD68	2.97
3428	IFI16	3.61	7171	TPM4	2.96
1520	CTSS	3.61	11004	KIF2C	2.96
10797	MTHFD2	3.57	10808	HSPH1	2.95
55681	SCYL2	3.57	84909	C9orf3	2.94
9749	PHACTR2	3.57	1894	ECT2	2.93
94240	EPSTI1	3.56	1629	DBT	2.92
64151	NCAPG	3.51	116969	ART5	2.90
25879	DCAF13	3.51	3227	HOXC11	2.88

Table S3. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
116064	LRRC58	3.47	3149	HMGB3	2.87
29899	GPSM2	3.47	10437	IFI30	2.87
135114	HINT3	3.45	57489	ODF2L	2.87
27333	GOLIM4	3.43	2151	F2RL2	2.86
55839	CENPN	3.43	23215	PRRC2C	2.85
23213	SULF1	3.41	128710	C20orf94	2.85
81671	VMP1	3.39	23594	ORC6	2.84
9889	ZBED4	3.36	5205	ATP8B1	2.83
3092	HIP1	3.34	51430	C1orf9	2.80
51512	GTSE1	3.34	57405	SPC25	2.80
92797	HELB	3.34	112401	BIRC8	2.80
51426	POLK	3.30	3606	IL18	2.80
5611	DNAJC3	3.30	115362	GBP5	2.80
6596	HLTF	3.28	50515	CHST11	2.79
9910	RABGAP1L	3.25	83461	CDCA3	2.79
528	ATP6V1C1	3.23	10744	PTTG2	2.78
3833	KIFC1	3.23	51765	MST4	2.77
197131	UBR1	3.20	10926	DBF4	2.76
29923	HILPDA	3.20	27125	AFF4	2.75
28998	MRPL13	3.19	10615	SPAG5	2.75
58527	C6orf115	3.19	55143	CDCA8	2.74
79000	C1orf135	3.19	51602	NOP58	2.74
9857	CEP350	3.18	51478	HSD17B7	2.73
84296	GINS4	3.18	2209	FCGR1A	2.73
81034	SLC25A32	3.15	9958	USP15	2.72
55723	ASF1B	3.14	5469	MED1	2.72
7110	TMF1	3.14	8813	DPM1	2.70
84081	NSRP1	3.14	6731	SRP72	2.70
23075	SWAP70	3.12	9991	PTBP3	2.70
6726	SRP9	2.69	79866	BORA	2.41
55215	FANCI	2.68	7072	TIA1	2.40
57590	WDFY1	2.67	55632	G2E3	2.40
55142	HAUS2	2.66	2213	FCGR2B	2.40
23047	PDS5B	2.66	3987	LIMS1	2.39
5373	PMM2	2.66	829	CAPZA1	2.39
11065	UBE2C	2.66	26973	CHORDC1	2.38
23085	ERC1	2.66	435	ASL	2.38
389197	C4orf50	2.65	29979	UBQLN1	2.38
11260	XPOT	2.65	8548	BLZF1	2.37
29980	DONSON	2.65	9694	TTC35	2.37
64399	HHIP	2.64	55055	ZWILCH	2.36
6453	ITSN1	2.63	4481	MSR1	2.36
29108	PYCARD	2.63	10213	PSMD14	2.35
9877	ZC3H11A	2.62	9966	TNFSF15	2.35

Table S3. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
81624	DIAPH3	2.62	51582	AZIN1	2.35
79723	SUV39H2	2.61	54843	SYTL2	2.34
55789	DEPDC1B	2.61	9039	UBA3	2.33
10097	ACTR2	2.59	933	CD22	2.33
23036	ZNF292	2.58	5685	PSMA4	2.33
22936	ELL2	2.57	9885	OSBPL2	2.33
8477	GPR65	2.57	9262	STK17B	2.33
23397	NCAPH	2.57	56942	C16orf61	2.32
3015	H2AFZ	2.54	10767	HBS1L	2.32
55749	CCAR1	2.53	87178	PNPT1	2.32
25937	WWTR1	2.52	6303	SAT1	2.32
360023	ZBTB41	2.51	7316	UBC	2.32
5080	PAX6	2.51	4205	MEF2A	2.32
4193	MDM2	2.51	85465	EPT1	2.31
24137	KIF4A	2.51	84640	USP38	2.31
9212	AURKB	2.51	5810	RAD1	2.30
168850	ZNF800	2.50	64397	ZFP106	2.29
55109	AGGF1	2.49	5706	PSMC6	2.29
23185	LARP4B	2.49	22948	CCT5	2.29
51571	FAM49B	2.49	10672	GNA13	2.29
51077	FCF1	2.49	339344	MYPOP	2.28
23167	EFR3A	2.49	7292	TNFSF4	2.28
23468	CBX5	2.48	57103	C12orf5	2.28
5396	PRRX1	2.48	388403	YPEL2	2.28
10096	ACTR3	2.47	54876	DCAF16	2.27
10308	ZNF267	2.47	113235	SLC46A1	2.27
6782	HSPA13	2.47	11177	BAZ1A	2.27
3832	KIF11	2.47	339175	METTL2A	2.26
917	CD3G	2.47	26586	CKAP2	2.26
80821	DDHD1	2.46	55785	FGD6	2.26
52	ACP1	2.46	24145	PANX1	2.25
4179	CD46	2.46	253461	ZBTB38	2.25
10499	NCOA2	2.44	23232	TBC1D12	2.25
60558	GUF1	2.44	995	CDC25C	2.25
55676	SLC30A6	2.43	55974	SLC50A1	2.25
6646	SOAT1	2.43	472	ATM	2.25
5440	POLR2K	2.43	23008	KLHDC10	2.24
84955	NUDCD1	2.42	10024	TROAP	2.24
54739	XAF1	2.42	9521	EEF1E1	2.24
84295	PHF6	2.23	7402	UTRN	2.09
7295	TXN	2.23	55589	BMP2K	2.08
2710	GK	2.23	158747	MOSPD2	2.08
10905	MAN1A2	2.22	56886	UGGT1	2.07
6780	STAU1	2.22	203100	HTRA4	2.07

Table S3. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
10282	BET1	2.22	55279	ZNF654	2.07
134430	WDR36	2.21	54499	TMCO1	2.07
4299	AFF1	2.21	81930	KIF18A	2.07
6747	SSR3	2.21	142686	ASB14	2.06
7334	UBE2N	2.21	55209	SETD5	2.06
5965	RECQL	2.21	9736	USP34	2.04
4605	MYBL2	2.2	116285	ACSM1	2.04
6093	ROCK1	2.19	2201	FBN2	2.04
161725	OTUD7A	2.19	963	CD53	2.04
23518	R3HDM1	2.18	55159	RFWD3	2.03
2239	GPC4	2.18	9871	SEC24D	2.03
28977	MRPL42	2.18	9887	SMG7	2.02
64859	OBFC2A	2.18	23376	UFL1	2.02
3845	KRAS	2.18	79646	PANK3	2.01
51388	NIP7	2.18	50613	UBQLN3	2.00
7586	ZKSCAN1	2.18	201595	STT3B	2.00
10762	NUP50	2.17	59345	GNB4	1.99
7328	UBE2H	2.17	5876	RABGGTB	1.99
10730	YME1L1	2.17	79820	CATSPERB	1.99
23093	TTLL5	2.17	6637	SNRPG	1.99
6790	AURKA	2.17	51330	TNFRSF12A	1.99
22889	KIAA0907	2.17	9928	KIF14	1.99
10875	FGL2	2.17	286097	EFHA2	1.98
23161	SNX13	2.17	9131	AIFM1	1.98
9169	SCAF11	2.16	488	ATP2A2	1.98
1788	DNMT3A	2.15	23042	PDXDC1	1.98
9088	PKMYT1	2.15	7114	TMSB4X	1.98
23033	DOPEY1	2.13	9123	SLC16A3	1.98
89882	TPD52L3	2.13	54454	ATAD2B	1.97
6556	SLC11A1	2.13	23143	LRCH1	1.97
64216	TFB2M	2.13	4212	MEIS2	1.97
3071	NCKAP1L	2.13	1457	CSNK2A1	1.97
51068	NMD3	2.13	80012	PHC3	1.97
509	ATP5C1	2.13	128497	SPATA25	1.96
953	ENTPD1	2.13	186	AGTR2	1.96
51105	PHF20L1	2.13	53981	CPSF2	1.96
5062	PAK2	2.13	56996	SLC12A9	1.96
9205	ZMYM5	2.12	1584	CYP11B1	1.96
55157	DARS2	2.12	133619	PRRC1	1.96
8520	HAT1	2.11	4288	MKI67	1.96
79739	TTLL7	2.11	9014	TAF1B	1.96
9495	AKAP5	2.10	55858	TMEM165	1.96
3181	HNRNPA2B1	2.10	2212	FCGR2A	1.96

Table S3. Cont.

NCBI gene ID	Gene Symbol	Fold Change	NCBI gene ID	Gene Symbol	Fold Change
389898	UBE2NL	2.10	10075	HUWE1	1.96
29850	TRPM5	2.10	220988	HNRNPA3	1.96
3070	HELLS	2.10	80146	UXS1	1.95
331	XIAP	2.09	122011	CSNK1A1L	1.95
55751	TMEM184C	2.09	150468	CKAP2L	1.95
2146	EZH2	2.09	84624	FNDC1	1.95
26057	ANKRD17	1.95	7332	UBE2L3	1.92
128061	C1orf131	1.95	3336	HSPE1	1.92
64090	GAL3ST2	1.94	54800	KLHL24	1.92
130507	UBR3	1.93	2290	FOXG1	1.91
2298	FOXD4	1.93	50848	F11R	1.91
123169	LEO1	1.93	10627	MYL12A	1.91
57187	THOC2	1.93	5074	PAWR	1.91
148789	B3GALNT2	1.93	6476	SI	1.91
58508	MLL3	1.92	1009	CDH11	1.90
5701	PSMC2	1.92	29066	ZC3H7A	1.90
148066	ZNRF4	1.92	51319	RSRC1	1.90
6670	SP3	1.92			

Table S4. Result of ROC curve analysis on our miRNA array data. ROC analysis was done to validate the diagnostic value of the miRNA in the miRNA-regulated PINs. AUC: area under (ROC) curve; * *p*-value < 0.05; *** *p*-value < 0.001.

miRBase Accession	miRNA name	AUC	<i>p</i> -value
MIMAT0002856	hsa-miR-520d-3p	0.49	0.549112
MIMAT0000265	hsa-miR-204-5p	0.98	6.47×10^{-10} ***
MIMAT0000272	hsa-miR-215	0.21	0.999782
MIMAT0000271	hsa-miR-214-3p	0.68	0.010387 *
MIMAT0002820	hsa-miR-497-5p	0.99	2.75×10^{-10} ***
MIMAT0000076	hsa-miR-21-5p	0.78	0.000184 ***
MIMAT0000738	hsa-miR-383	0.60	0.106284
MIMAT0000423	hsa-miR-125b-5p	0.99	2.48×10^{-10} ***
MIMAT0000064	hsa-let-7c	0.93	3.79×10^{-8} ***
MIMAT0000089	hsa-miR-31-5p	0.80	8.63×10^{-5} ***
MIMAT0000077	hsa-miR-22-3p	0.27	0.99749
MIMAT0000098	hsa-miR-100-5p	0.98	5.55×10^{-10} ***
MIMAT0000097	hsa-miR-99a-5p	0.99	2.55×10^{-10} ***
MIMAT0000443	hsa-miR-125a-5p	0.31	0.990694
MIMAT0002819	hsa-miR-193b-3p	0.41	0.86128
MIMAT0000250	hsa-miR-139-5p	0.99	2.42×10^{-10} ***
MIMAT0000437	hsa-miR-145-5p	0.96	3.14×10^{-9} ***
MIMAT0000421	hsa-miR-122-5p	0.48	0.597483

Table S5. Summary of constructed miRNA-regulated networks. L0 gene: genes connected directly to the miRNA (*i.e.*, direct target of miRNA); L1 gene: genes not connected directly to the miRNA.

miRBase Accession	miR name	Total gene count	L0 count	L1 count
MIMAT0002819	hsa-miR-193b-3p	16	1	15
MIMAT0000250	hsa-miR-139-5p	28	10	18
MIMAT0000437	hsa-miR-145-5p	86	22	64
MIMAT0000423	hsa-miR-125b-5p	211	16	195
MIMAT0000443	hsa-miR-125a-5p	206	14	192
MIMAT0000097	hsa-miR-99a-5p	14	1	13
MIMAT0000265	hsa-miR-204-5p	64	18	46
MIMAT0000076	hsa-miR-21-5p	91	16	75
MIMAT0000064	hsa-let-7c	96	20	76
MIMAT0000421	hsa-miR-122-5p	5	3	2
MIMAT0000098	hsa-miR-100-5p	14	1	13
MIMAT0000272	hsa-miR-215	3	3	0
MIMAT0000271	hsa-miR-214-3p	14	8	6
MIMAT0000738	hsa-miR-383	34	3	31
MIMAT0002856	hsa-miR-520d-3p	146	23	123
MIMAT0000077	hsa-miR-22-3p	46	11	35
MIMAT0002820	hsa-miR-497-5p	267	32	235
MIMAT0000089	hsa-miR-31-5p	34	3	31

Table S6. Specific enriched GO terms of each miRNA-regulated PINs. Genes annotated with the specific GO term in the PIN were also listed in this table. Adj. *p*-value: multiple-test adjusted *p*-value calculated by the method described in the work of Benjamini and Yekutieli [48].

MIMAT0002856(hsa-miR-520d-3p)		
GO term	Genes	Adj. <i>p</i> -value
GO:0007169, Transmembrane receptor protein tyrosine kinase signaling pathway	SH3KBP1, HDAC2, RET, ABI1, LYN, GRB2, SORBS1, CLTC, CLTA, CDC42, CASP9, RAF1, SRC, AP2A1, AP2B1, MAPK3, ARHGEF7, PRKCA, RPS6, PRKAR2B, MAPK1, ARHGEF6, CDK1, SH3GL2, EIF4G1, HDAC1, ECT2, MKNK1, CASP3, PRKACA, ADRB2, PRKAR2A, EIF4B, SHC1, RAC1	2.77×10^{-29}
GO:0048011, Nerve growth factor receptor signaling pathway	HDAC2, GRB2, CLTC, CLTA, CASP9, RAF1, SRC, AP2A1, AP2B1, MAPK3, ARHGEF7, PRKCA, PRKAR2B, MAPK1, ARHGEF6, CDK1, SH3GL2, HDAC1, ECT2, CASP3, PRKACA, PRKAR2A, SHC1, RAC1	5.09×10^{-24}
GO:0007173, Epidermal growth factor receptor signaling pathway	SH3KBP1, GRB2, CLTC, CLTA, CDC42, CASP9, RAF1, SRC, AP2A1, AP2B1, MAPK3, ARHGEF7, PRKCA, PRKAR2B, MAPK1, CDK1, SH3GL2, PRKACA, PRKAR2A, SHC1	2.96×10^{-22}
GO:0043067, Regulation of programmed cell death	HDAC2, STK17B, ESR1, ABL1, LYN, TP53, GABRB3, PAK2, LCK, CASP9, RAF1, PLK1, ARHGEF7, PRKCA, RPS6, SH3RF1, MAPK1, IFT57, ARHGAP10, ARHGEF6, CDK1, APAF1, HDAC1, ECT2, CASP3, SOX10, EP300, ARAF, TFAP2A, ADRB2, HCK, KLHL20, CASP8, HIP1, RAC1	4.76×10^{-19}
GO:0042058, Regulation of epidermal growth factor receptor signaling pathway	SH3KBP1, ESR1, GRB2, CLTC, CLTA, CDC42, AP2A1, AP2B1, ARHGEF7, SH3GL2, SHC1	2.36×10^{-12}
GO:0008543, Fibroblast growth factor receptor signaling pathway	GRB2, CASP9, RAF1, SRC, MAPK3, PRKCA, PRKAR2B, MAPK1, CDK1, MKNK1, PRKACA, PRKAR2A, SHC1	2.39×10^{-12}
GO:0043068, Positive regulation of programmed cell death	STK17B, ABL1, LYN, TP53, LCK, CASP9, ARHGEF7, PRKCA, RPS6, SH3RF1, MAPK1, ARHGEF6, APAF1, ECT2, CASP3, EP300, TFAP2A, ADRB2, CASP8, HIP1, RAC1	3.09×10^{-12}
GO:0010942, Positive regulation of cell death	STK17B, ABL1, LYN, TP53, LCK, CASP9, ARHGEF7, PRKCA, RPS6, SH3RF1, MAPK1, ARHGEF6, APAF1, ECT2, CASP3, EP300, TFAP2A, ADRB2, CASP8, HIP1, RAC1	4.49×10^{-12}
GO:0006917, Induction of apoptosis	STK17B, ABL1, TP53, LCK, CASP9, ARHGEF7, PRKCA, SH3RF1, MAPK1, ARHGEF6, APAF1, ECT2, CASP3, EP300, CASP8, HIP1, RAC1	6.91×10^{-11}
GO:0012502, Induction of programmed cell death	STK17B, ABL1, TP53, LCK, CASP9, ARHGEF7, PRKCA, SH3RF1, MAPK1, ARHGEF6, APAF1, ECT2, CASP3, EP300, CASP8, HIP1, RAC1	7.42×10^{-11}

Table S6. Cont.

MIMAT0002856(hsa-miR-520d-3p)		
GO term	Genes	Adj. p-value
GO:0042059, Negative regulation of epidermal growth factor receptor signaling pathway	SH3KBP1, GRB2, CLTC, CLTA, CDC42, AP2A1, AP2B1, ARHGEF7, SH3GL2	8.63×10^{-11}
GO:0015630, Microtubule cytoskeleton	STMN1, SORBS1, SMAD4, CLTC, CDC42, LCK, RACGAP1, PLK1, PRKAR2B, YES1, MAPK1, IFT57, CDK1, ECT2, PRKACA, RB1, EP300, CCNB1, CHAF1B, TFAP2A, CASP8, PRKAR2A	4.75×10^{-10}
GO:0060548, Negative regulation of cell death	HDAC2, ESR1, TP53, SMAD4, RAF1, PLK1, PRKCA, RPS6, SH3RF1, CDK1, HDAC1, CASP3, SOX10, ARAF, TFAP2A, HCK, KLHL20	6.27×10^{-8}
GO:0008286, Insulin receptor signaling pathway	GRB2, SORBS1, RAF1, MAPK3, RPS6, MAPK1, CDK1, EIF4G1, EIF4B, SHC1	2.15×10^{-7}
GO:0043069, Negative regulation of programmed cell death	HDAC2, ESR1, TP53, RAF1, PLK1, PRKCA, RPS6, SH3RF1, CDK1, HDAC1, CASP3, SOX10, ARAF, TFAP2A, HCK, KLHL20	3.13×10^{-7}
GO:0008284, Positive regulation of cell proliferation	HDAC2, ESR1, LYN, CDC42, E2F1, PRKCA, MAPK1, CDK1, RHOG, HDAC1, NCK1, SOX10, CCNB1, ADRB2, HCK, SHC1	8.34×10^{-7}
GO:0051988, Regulation of attachment of spindle microtubules to kinetochore	CDC42, RACGAP1, ECT2, CCNB1	3.27×10^{-5}
GO:0008629, Induction of apoptosis by intracellular signals	ABL1, TP53, CASP9, APAF1, CASP3, EP300, CASP8	5.83×10^{-5}
MIMAT0002820(hsa-miR-497-5p)		
GO term	Genes	Adj. p-value
GO:0043067, Regulation of programmed cell death	ESR1, MEN1, ABL1, HIPK3, PPARGC1A, SIAH1, SH3RF1, PAK2, LCK, MED1, PPARG, CBX4, ARHGEF7, YWHAB, RXRA, ACVR1, MAPK1, CASP3, CASP6, AR, PTPRF, MDM2, BRCA1, MLH1, RAB27A, PIAS4, FAF1, RAC1, VHL, SKI, NR4A1, LYN, TP53, PSMC2, GATA1, GATA6, GATA3, RAF1, CDKN1B, PLK1, PSMD11, HOXA13, RPS6, ESR2, ARHGAP10, ARHGEF6, SMAD3, SKIL, RYR2, PSEN1, HCK, TRAF2	2.67×10^{-25}
GO:0043068, Positive regulation of programmed cell death	MEN1, ABL1, SIAH1, SH3RF1, LCK, PPARG, ARHGEF7, YWHAB, RXRA, MAPK1, CASP3, CASP6, PTPRF, BRCA1, MLH1, RAB27A, PIAS4, FAF1, RAC1, NR4A1, LYN, TP53, GATA6, CDKN1B, HOXA13, RPS6, ESR2, ARHGEF6, SMAD3, RYR2, PSEN1, TRAF2	1.74×10^{-17}
GO:0010942, Positive regulation of cell death	MEN1, ABL1, SIAH1, SH3RF1, LCK, PPARG, ARHGEF7, YWHAB, RXRA, MAPK1, CASP3, CASP6, PTPRF, BRCA1, MLH1, RAB27A, PIAS4, FAF1, RAC1, NR4A1, LYN, TP53, GATA6, CDKN1B, HOXA13, RPS6, ESR2, ARHGEF6, SMAD3, RYR2, PSEN1, TRAF2	3.08×10^{-17}

Table S6. Cont.

MIMAT0002820(hsa-miR-497-5p)		
GO term	Genes	Adj. p-value
GO:0008285, Negative regulation of cell proliferation	MEN1, MED1, PPARG, RXRA, CASP3, AR, PTPRF, VDR, VHL, SKI, LYN, TP53, TOB1, GATA1, GATA3, RAF1, HNF4A, CDKN1B, BRD7, MED25, ESR2, ABI1, SMAD1, SMAD2, SMAD3, SMAD4, SOX7	3.85×10^{-14}
GO:0015629, Actin cytoskeleton	ABL1, SORBS1, FLNA, SEPT7, ANLN, MACF1, HAP1, SH3PXD2A, IQGAP2, BRCA1, ACTC1, ACTA1, MYL2, MYLK, SORBS2, ARPC4, ARPC5, ACTR2, ACTR3, ARPC1B, WASF1, WASF2, HCK	2.52×10^{-13}
GO:0006917, Induction of apoptosis	ABL1, SH3RF1, LCK, PPARG, ARHGEF7, YWHAB, MAPK1, CASP3, CASP6, BRCA1, MLH1, RAB27A, RAC1, NR4A1, TP53, CDKN1B, ARHGEF6, SMAD3, RYR2, PSEN1, TRAF2	9.69×10^{-11}
GO:0012502, Induction of programmed cell death	ABL1, SH3RF1, LCK, PPARG, ARHGEF7, YWHAB, MAPK1, CASP3, CASP6, BRCA1, MLH1, RAB27A, RAC1, NR4A1, TP53, CDKN1B, ARHGEF6, SMAD3, RYR2, PSEN1, TRAF2	1.06×10^{-10}
GO:0007178, Transmembrane receptor protein serine/threonine kinase signaling pathway	ACVR1, SMURF2, SKI, GDF6, BMP6, ZNF8, GATA4, HNF4A, SMAD1, SMAD2, SMAD3, SMAD4, SMAD5, RYR2	1.22×10^{-10}
GO:0007169, Transmembrane receptor protein tyrosine kinase signaling pathway	SORBS1, CDC42, SRC, MAPK3, ARHGEF7, YWHAB, MAPK1, SH3GL2, CASP3, MDM2, EIF4G1, RAC1, SH3KBP1, NR4A1, LYN, GRB2, RAF1, CDKN1B, RPS6, ABI1, ARHGEF6, MKNK1, PSEN1, EIF4B	1.67×10^{-10}
GO:0090092, Regulation of transmembrane receptor protein serine/threonine kinase signaling pathway	MEN1, ACVR1, SMURF2, SKI, GDF6, TP53, BMP6, GATA4, GATA6, HOXA13, SMAD2, SMAD3, SMAD4, SKIL	7.51×10^{-10}
GO:0030509, BMP signaling pathway	ACVR1, SMURF2, SKI, GDF6, BMP6, ZNF8, SMAD1, SMAD4, SMAD5, RYR2	3.25×10^{-9}
GO:0060548, Negative regulation of cell death	ESR1, HIPK3, PPARGC1A, SH3RF1, MED1, CBX4, ACVR1, CASP3, AR, MDM2, VHL, TP53, GATA1, GATA6, GATA3, RAF1, CDKN1B, PLK1, RPS6, SMAD3, SMAD4, PSEN1, HCK	7.88×10^{-9}
GO:0007173, Epidermal growth factor receptor signaling pathway	CDC42, SRC, MAPK3, ARHGEF7, YWHAB, MAPK1, SH3GL2, MDM2, SH3KBP1, NR4A1, GRB2, RAF1, CDKN1B	9.22×10^{-9}
GO:0030521, Androgen receptor signaling pathway	PPARGC1A, MED14, MED1, AR, BRCA1, MED12, PIAS1, RAN, NR1I3	1.42×10^{-8}
GO:0043069, Negative regulation of programmed cell death	ESR1, HIPK3, PPARGC1A, SH3RF1, MED1, CBX4, ACVR1, CASP3, AR, MDM2, VHL, TP53, GATA1, GATA6, GATA3, RAF1, CDKN1B, PLK1, RPS6, SMAD3, PSEN1, HCK	2.44×10^{-8}
GO:0048011, Nerve growth factor receptor signaling pathway	SRC, MAPK3, ARHGEF7, YWHAB, MAPK1, SH3GL2, CASP3, MDM2, RAC1, NR4A1, GRB2, RAF1, CDKN1B, ARHGEF6, PSEN1	2.64×10^{-8}
GO:0032956, Regulation of actin cytoskeleton organization	ABL1, LRP1, ARPC4, ARPC5, ACTR3, ARPC1B, SMAD3, NCK1, SORBS3, HCK, LIMK1	6.42×10^{-6}

Table S6. Cont.

MIMAT0002820(hsa-miR-497-5p)		
GO term	Genes	Adj. p-value
GO:0008543, Fibroblast growth factor receptor signaling pathway	SRC, MAPK3, YWHAB, MAPK1, MDM2, NR4A1, GRB2, RAF1, CDKN1B, MKNK1	9.06×10^{-6}
GO:0042059, Negative regulation of epidermal growth factor receptor signaling pathway	CDC42, ARHGEF7, SH3GL2, PTPRF, SH3KBP1, GRB2, PSEN1	1.11×10^{-5}
GO:0042058, Regulation of epidermal growth factor receptor signaling pathway	ESR1, CDC42, ARHGEF7, SH3GL2, PTPRF, SH3KBP1, GRB2, PSEN1	1.17×10^{-5}
GO:0007179, Transforming growth factor beta receptor signaling pathway	ACVR1, SMURF2, SKI, SMAD1, SMAD2, SMAD3, SMAD4, SMAD5	1.67×10^{-5}
GO:0015630, Microtubule cytoskeleton	STMN1, RIF1, SORBS1, CDC42, LCK, RACGAP1, YES1, YWHAB, MAPK1, SEPT7, KIF23, CDC16, MACF1, BRCA1, FEZ1, NCOR1, PLK1, CHD3, SMAD4, CEP350, CDC27, PSEN1	2.14×10^{-5}
GO:0017015, Regulation of transforming growth factor beta receptor signaling pathway	MEN1, SMURF2, SKI, TP53, SMAD2, SMAD3, SMAD4, SKIL	2.30×10^{-5}
GO:0070302, Regulation of stress-activated protein kinase signaling cascade	MEN1, ZEB2, HIPK3, SH3RF1, CDC42, MAPK3, MAPK1, LYN, NCOR1, TRAF2	2.74×10^{-5}
GO:0001959, Regulation of cytokine-mediated signaling pathway	HSP90AB1, MED1, PPARG, PTPRF, NR1H2, PIAS1, IL36RN, HIPK1	6.35×10^{-5}
GO:0008284, Positive regulation of cell proliferation	ESR1, CDC42, MED1, RARA, MAPK1, AR, MDM2, NR4A1, LYN, FZR1, BMP6, GATA1, GATA4, GATA6, CDKN1B, NCK1, HCLS1, HCK	7.29×10^{-5}

Table S6. Cont.

MIMAT0000423(hsa-miR-125b-5p)		
GO term	Genes	Adj. p-value
GO:0043067, Regulation of programmed cell death	HMGA2, PML, PRNP, FGF2, XRCC4, BRCA1, IGFBP3, HDAC3, CTNNB1, CD5, CDK1, NKX2-5, MEF2C, PRKCI, CASP2, PSMA4, PSMA3, CFDP1, CAV1, FAF1, YWHAB, HIF1A, RELA, TCF7L2, TNFSF12, PSEN2, TP53, TOP2A, TNFRSF4, BID, MYC, JUN, OGT, CDKN1A, HOXA13, RNF7, PPP2R4, HDAC2, HDAC1, SNCA, PTEN, NFKBIA, IFI16, NOL3, TRAF2, HSP90B1	4.56×10^{-24}
GO:0060548, Negative regulation of cell death	HMGA2, PRNP, FGF2, XRCC4, HDAC3, CTNNB1, CDK1, NKX2-5, MEF2C, PRKCI, CFDP1, HIF1A, RELA, TCF7L2, PSEN2, TP53, TNFRSF4, MYC, JUN, CDKN1A, RNF7, HDAC2, HDAC1, SNCA, PTEN, MGMT, NFKBIA, NOL3, HSP90B1	8.04×10^{-17}
GO:0008284, Positive regulation of cell proliferation	HMGA2, FGF2, XRCC4, CDC25B, CTNNB1, EGR1, AGGF1, CDK1, NKX2-5, MEF2C, PRKCI, IRS1, HIF1A, RELA, HCLS1, TNFSF12, ARNT, PTPRC, TNFSF4, TNFRSF4, MYC, JUN, FGF1, CDKN1A, HDAC2, HDAC1, NOLC1, PTEN	2.20×10^{-15}
GO:0043069, Negative regulation of programmed cell death	HMGA2, PRNP, XRCC4, HDAC3, CTNNB1, CDK1, NKX2-5, MEF2C, PRKCI, CFDP1, HIF1A, RELA, TCF7L2, PSEN2, TP53, TNFRSF4, MYC, JUN, CDKN1A, RNF7, HDAC2, HDAC1, SNCA, PTEN, NFKBIA, NOL3, HSP90B1	3.62×10^{-15}
GO:0043068, Positive regulation of programmed cell death	HMGA2, PML, BRCA1, IGFBP3, CTNNB1, CD5, MEF2C, PRKCI, CASP2, CAV1, FAF1, YWHAB, TNFSF12, PSEN2, TP53, TOP2A, BID, JUN, OGT, CDKN1A, HOXA13, RNF7, PPP2R4, PTEN, IFI16, TRAF2	4.51×10^{-14}
GO:0010942, Positive regulation of cell death	HMGA2, PML, BRCA1, IGFBP3, CTNNB1, CD5, MEF2C, PRKCI, CASP2, CAV1, FAF1, YWHAB, TNFSF12, PSEN2, TP53, TOP2A, BID, JUN, OGT, CDKN1A, HOXA13, RNF7, PPP2R4, PTEN, IFI16, TRAF2	7.15×10^{-14}
GO:0006916, Anti-apoptosis	PRNP, HDAC3, CDK1, NKX2-5, MEF2C, PRKCI, CFDP1, RELA, TCF7L2, PSEN2, RNF7, HDAC1, SNCA, NFKBIA, NOL3, HSP90B1	1.25×10^{-10}
GO:0008285, Negative regulation of cell proliferation	SERPINF1, SRF, PML, PRNP, FGF2, CSNK2B, IGFBP3, CTNNB1, CAV1, HMGA1, VDR, CDH5, HSF1, COL18A1, TP53, MYC, JUN, CDKN1A, PAK1, PTEN	2.08×10^{-9}
GO:0015630, Microtubule cytoskeleton	STMN1, KIF1C, RANGAP1, CDC25B, BRCA1, HDAC3, CTNNB1, PIN4, HSPH1, RANBP9, CDK1, SPTAN1, YWHAQ, DVL1, FKBP4, YWHAB, CCDC85B, MAPT, PSEN2, TOP2A, SPIB, MYC, OGT, APEX1, PAFAH1B1	2.24×10^{-9}
GO:0048011, Nerve growth factor receptor signaling pathway	HDAC3, CDK1, MEF2C, PRKCI, CASP2, IRS1, YWHAB, RELA, PSEN2, HDAC2, HDAC1, PTEN, ATF1, NFKBIA	1.99×10^{-8}
GO:0050678, Regulation of epithelial cell proliferation	SERPINF1, PGR, FGF2, CTNNB1, AGGF1, CAV1, HIF1A, TNFSF12, ARNT, MYC, JUN, FGF1, PTEN	3.41×10^{-8}
GO:0007169, Transmembrane receptor protein tyrosine kinase signaling pathway	FGF2, HDAC3, CDK1, MEF2C, PRKCI, CASP2, IRS1, FIBP, PTPN1, YWHAB, RELA, PSEN2, FGF1, HDAC2, HDAC1, PTEN, ATF1, NFKBIA, EIF4EBP1	6.39×10^{-8}

Table S6. Cont.

MIMAT0000423(hsa-miR-125b-5p)		
GO term	Genes	Adj. p-value
GO:0006917, Induction of apoptosis	PML, BRCA1, CD5, CASP2, CAV1, YWHAB, TNFSF12, PSEN2, TP53, BID, OGT, CDKN1A, RNF7, PTEN, IFI16, TRAF2	1.68×10^{-7}
GO:0012502, Induction of programmed cell death	PML, BRCA1, CD5, CASP2, CAV1, YWHAB, TNFSF12, PSEN2, TP53, BID, OGT, CDKN1A, RNF7, PTEN, IFI16, TRAF2	1.81×10^{-7}
GO:0035666, TRIF-dependent toll-like receptor signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	2.40×10^{-6}
GO:0034138, Toll-like receptor 3 signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	2.71×10^{-6}
GO:0051693, Actin filament capping	SPTB, SPTBN1, SPTAN1, SPTA1, ADD1, EPB49	3.43×10^{-6}
GO:0002756, MyD88-independent toll-like receptor signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	3.82×10^{-6}
GO:0034134, Toll-like receptor 2 signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	5.33×10^{-6}
GO:0034130, Toll-like receptor 1 signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	5.33×10^{-6}
GO:0030835, Negative regulation of actin filament depolymerization	SPTB, SPTBN1, SPTAN1, SPTA1, ADD1, EPB49	5.58×10^{-6}
GO:0015629, Actin cytoskeleton	WAS, CDH1, BRCA1, SPTB, SPTBN1, SPTAN1, CTDP1, STX1A, SPTA1, PAK1, SNCA, ADD1, EPB41, EPB49	5.58×10^{-6}
GO:0002755, MyD88-dependent toll-like receptor signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	7.66×10^{-6}
GO:0034142, Toll-like receptor 4 signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	1.14×10^{-5}
GO:0050679, Positive regulation of epithelial cell proliferation	FGF2, CTNNB1, AGGF1, HIF1A, TNFSF12, ARNT, MYC, JUN, FGF1	1.14×10^{-5}

Table S6. Cont.

MIMAT0000076(hsa-miR-21-5p)		
GO term	Genes	Adj. p-value
GO:0030834, Regulation of actin filament depolymerization	SPTB, SPTBN1, SPTAN1, SPTA1, ADD1, EPB49	1.27×10^{-5}
GO:0030837, Negative regulation of actin filament polymerization	SPTB, SPTBN1, SPTAN1, SPTA1, ADD1, EPB49	2.71×10^{-5}
GO:0002224, Toll-like receptor signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	2.96×10^{-5}
GO:0008629, Induction of apoptosis by intracellular signals	PML, BRCA1, YWHAB, TP53, BID, CDKN1A, RNF7, IFI16	3.52×10^{-5}
GO:0043067, Regulation of programmed cell death	SPRY2, TP53, ADAMTSL4, ETS1, TDGF1, RAF1, HOXA5, HOXA13, MSX1, MSX2, NKX2-5, CBL, INHBB, COL4A3, ACVR1C, TRAF2	1.92×10^{-5}
GO:0007173, Epidermal growth factor receptor signaling pathway	SPRY2, SPRY1, GRB2, PTPN11, TDGF1, RAF1, CBL	9.44×10^{-5}
MIMAT0000250(hsa-miR-139-5p)		
GO term	Genes	Adj. p-value
GO:0035583, Negative regulation of transforming growth factor beta receptor signaling pathway by extracellular sequestering of TGFbeta	LTBP1, FBN1, FBN2	2.61×10^{-5}
MIMAT0000089(hsa-miR-31-5p)		
GO term	Genes	Adj. p-value
GO:0007187, G-protein signaling, coupled to cyclic nucleotide second messenger	GNA12, GNA13, DRD5, MTNR1A, S1PR3, TSHR, S1PR4	8.74×10^{-7}
GO:0019935, Cyclic-nucleotide-mediated signaling	GNA12, GNA13, DRD5, MTNR1A, S1PR3, TSHR, S1PR4	1.60×10^{-6}
GO:0007188, G-protein signaling, coupled to cAMP nucleotide second messenger	GNA12, GNA13, DRD5, S1PR3, TSHR, S1PR4	6.82×10^{-6}

Table S6. Cont.

MIMAT0000089(hsa-miR-31-5p)		
GO term	Genes	Adj. p-value
GO:0048011, Nerve growth factor receptor signaling pathway	ARHGEF1, PRKCD, ARHGEF12, PRKACA, PRKCE, MCF2, ARHGEF11	6.93×10^{-6}
GO:0019933, CAMP-mediated signaling	GNA12, GNA13, DRD5, S1PR3, TSHR, S1PR4	1.05×10^{-5}
GO:0043067, Regulation of programmed cell death	PTK2B, PRKCD, TGFBR1, ARHGEF12, CTNNB1, PRKCE, TIA1, MCF2, FASTK, ARHGEF11, F2R	1.25×10^{-5}
GO:0003376, Sphingosine-1-phosphate signaling pathway	S1PR3, S1PR2, S1PR4	3.22×10^{-5}
GO:0007169, Transmembrane receptor protein tyrosine kinase signaling pathway	PTK2B, ARHGEF1, PRKCD, ARHGEF12, PRKACA, PRKCE, MCF2, ARHGEF11	5.20×10^{-5}
GO:0043068, Positive regulation of programmed cell death	TGFBR1, ARHGEF12, CTNNB1, PRKCE, TIA1, MCF2, FASTK, ARHGEF11	7.67×10^{-5}
GO:0010942, Positive regulation of cell death	TGFBR1, ARHGEF12, CTNNB1, PRKCE, TIA1, MCF2, FASTK, ARHGEF11	8.78×10^{-5}
GO:0006917, Induction of apoptosis	TGFBR1, ARHGEF12, PRKCE, TIA1, MCF2, FASTK, ARHGEF11	9.32×10^{-5}
GO:0012502, Induction of programmed cell death	TGFBR1, ARHGEF12, PRKCE, TIA1, MCF2, FASTK, ARHGEF11	9.51×10^{-5}
MIMAT0000437(hsa-miR-145-5p)		
GO term	Genes	Adj. p-value
GO:0030509, BMP signaling pathway	BMP6, ZNF8, ACVR1, SMAD1, SMAD4, RYR2, SMAD5, SMURF2, GDF6	1.37×10^{-11}
GO:0007178, Transmembrane receptor protein serine/threonine kinase signaling pathway	BMP6, ZNF8, ACVR1, SMAD1, SMAD4, RYR2, SMAD5, SMURF2, GDF6	2.56×10^{-8}
GO:0090092, Regulation of transmembrane receptor protein serine/threonine kinase signaling pathway	MEN1, TP53, BMP6, HOXA13, ACVR1, SMAD4, SULF1, SMURF2, GDF6	8.22×10^{-8}

Table S6. Cont.

MIMAT0000064(hsa-let-7c)		
GO term	Genes	Adj. p-value
GO:0043067, Regulation of programmed cell death	RRM2B, ACVR1, TP53, RASA1, TGFBR1, PSMA3, BIRC5, ACTN2, HOXA13, IRS2, FASTK, VAV1, PSMB6, BCL2, CDK1, HDAC1, SOX10, TIA1, AKT1, AURKB	3.43×10^{-8}
GO:0043069, Negative regulation of programmed cell death	RRM2B, ACVR1, TP53, RASA1, TGFBR1, BIRC5, IRS2, BCL2, CDK1, HDAC1, SOX10, AKT1, AURKB	6.58×10^{-6}
GO:0060548, Negative regulation of cell death	RRM2B, ACVR1, TP53, RASA1, TGFBR1, BIRC5, IRS2, BCL2, CDK1, HDAC1, SOX10, AKT1, AURKB	8.92×10^{-6}
GO:0015630, Microtubule cytoskeleton	INCENP, SNTB2, SEPT1, TACC1, BIRC5, RACGAP1, PIN4, CDCA8, CDK1, PHF1, AKT1, AURKB, NINL, CCDC85B	5.15×10^{-5}
GO:0043067, Regulation of programmed cell death	HMGA2, PML, PRNP, FGF2, XRCC4, BRCA1, IGFBP3, HDAC3, CTNNB1, CD5, CDK1, NKX2-5, MEF2C, PRKCI, CASP2, PSMA4, PSMA3, CFDP1, CAV1, FAF1, YWHAB, HIF1A, RELA, TCF7L2, TNFSF12, PSEN2, TP53, TOP2A, TNFRSF4, BID, MYC, JUN, OGT, CDKN1A, RNF7, PPP2R4, HDAC2, HDAC1, SNCA, PTEN, NFKBIA, IFI16, NOL3, TRAF2, HSP90B1	1.62×10^{-23}
GO:0060548, Negative regulation of cell death	HMGA2, PRNP, FGF2, XRCC4, HDAC3, CTNNB1, CDK1, NKX2-5, MEF2C, PRKCI, CFDP1, HIF1A, RELA, TCF7L2, PSEN2, TP53, TNFRSF4, MYC, JUN, CDKN1A, RNF7, HDAC2, HDAC1, SNCA, PTEN, MGMT, NFKBIA, NOL3, HSP90B1	4.53×10^{-17}
GO:0008284, Positive regulation of cell proliferation	HMGA2, FGF2, XRCC4, CDC25B, CTNNB1, EGR1, AGGF1, CDK1, NKX2-5, MEF2C, PRKCI, IRS1, HIF1A, RELA, HCLS1, TNFSF12, ARNT, PTPRC, TNFSF4, TNFRSF4, MYC, JUN, FGF1, CDKN1A, HDAC2, HDAC1, NOLC1, PTEN	1.23×10^{-15}
MIMAT0000443(hsa-miR-125a-5p)		
GO term	Genes	Adj. p-value
GO:0043069, Negative regulation of programmed cell death	HMGA2, PRNP, XRCC4, HDAC3, CTNNB1, CDK1, NKX2-5, MEF2C, PRKCI, CFDP1, HIF1A, RELA, TCF7L2, PSEN2, TP53, TNFRSF4, MYC, JUN, CDKN1A, RNF7, HDAC2, HDAC1, SNCA, PTEN, NFKBIA, NOL3, HSP90B1	2.02×10^{-15}
GO:0043068, Positive regulation of programmed cell death	HMGA2, PML, BRCA1, IGFBP3, CTNNB1, CD5, MEF2C, PRKCI, CASP2, CAV1, FAF1, YWHAB, TNFSF12, PSEN2, TP53, TOP2A, BID, JUN, OGT, CDKN1A, RNF7, PPP2R4, PTEN, IFI16, TRAF2	2.80×10^{-13}
GO:0010942, Positive regulation of cell death	HMGA2, PML, BRCA1, IGFBP3, CTNNB1, CD5, MEF2C, PRKCI, CASP2, CAV1, FAF1, YWHAB, TNFSF12, PSEN2, TP53, TOP2A, BID, JUN, OGT, CDKN1A, RNF7, PPP2R4, PTEN, IFI16, TRAF2	4.35×10^{-13}
GO:0006916, Anti-apoptosis	PRNP, HDAC3, CDK1, NKX2-5, MEF2C, PRKCI, CFDP1, RELA, TCF7L2, PSEN2, RNF7, HDAC1, SNCA, NFKBIA, NOL3, HSP90B1	9.04×10^{-11}
GO:0008285, Negative regulation of cell proliferation	SERPINF1, SRF, PML, PRNP, FGF2, CSNK2B, IGFBP3, CTNNB1, CAV1, HMGA1, VDR, CDH5, HSF1, COL18A1, TP53, MYC, JUN, CDKN1A, PAK1, PTEN	1.32×10^{-9}

Table S6. Cont.

MIMAT0000443(hsa-miR-125a-5p)		
GO term	Genes	Adj. p-value
GO:0015630, Microtubule cytoskeleton	STMN1, KIF1C, RANGAP1, CDC25B, BRCA1, HDAC3, CTNNB1, PIN4, HSPH1, RANBP9, CDK1, SPTAN1, YWHAQ, DVL1, FKBP4, YWHAB, CCDC85B, MAPT, PSEN2, TOP2A, SPIB, MYC, OGT, APEX1, PAFAH1B1	1.32×10^{-9}
GO:0048011, Nerve growth factor receptor signaling pathway	HDAC3, CDK1, MEF2C, PRKCI, CASP2, IRS1, YWHAB, RELA, PSEN2, HDAC2, HDAC1, PTEN, ATF1, NFKBIA	1.50×10^{-8}
GO:0050678, Regulation of epithelial cell proliferation	SERPINF1, PGR, FGF2, CTNNB1, AGGF1, CAV1, HIF1A, TNFSF12, ARNT, MYC, JUN, FGF1, PTEN	2.66×10^{-8}
GO:0007169, Transmembrane receptor protein tyrosine kinase signaling pathway	FGF2, HDAC3, CDK1, MEF2C, PRKCI, CASP2, IRS1, FIBP, PTPN1, YWHAB, RELA, PSEN2, FGF1, HDAC2, HDAC1, PTEN, ATF1, NFKBIA, EIF4EBP1	4.37×10^{-8}
GO:0006917, Induction of apoptosis	PML, BRCA1, CD5, CASP2, CAV1, YWHAB, TNFSF12, PSEN2, TP53, BID, OGT, CDKN1A, RNF7, PTEN, IFI16, TRAF2	1.22×10^{-7}
GO:0012502, Induction of programmed cell death	PML, BRCA1, CD5, CASP2, CAV1, YWHAB, TNFSF12, PSEN2, TP53, BID, OGT, CDKN1A, RNF7, PTEN, IFI16, TRAF2	1.31×10^{-7}
GO:0035666, TRIF-dependent toll-like receptor signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	2.01×10^{-6}
GO:0034138, Toll-like receptor 3 signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	2.27×10^{-6}
GO:0051693, Actin filament capping	SPTB, SPTBN1, SPTAN1, SPTA1, ADD1, EPB49	2.97×10^{-6}
GO:0002756, MyD88-independent toll-like receptor signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	3.20×10^{-6}
GO:0015629, Actin cytoskeleton	WAS, CDH1, BRCA1, SPTB, SPTBN1, SPTAN1, CTDP1, STX1A, SPTA1, PAK1, SNCA, ADD1, EPB41, EPB49	4.25×10^{-6}
GO:0034134, Toll-like receptor 2 signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	4.43×10^{-6}
GO:0034130, Toll-like receptor 1 signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	4.43×10^{-6}
GO:0030835, Negative regulation of actin filament depolymerization	SPTB, SPTBN1, SPTAN1, SPTA1, ADD1, EPB49	4.72×10^{-6}

Table S6. Cont.

MIMAT0000443(hsa-miR-125a-5p)		
GO term	Genes	Adj. p-value
GO:0002755, MyD88-dependent toll-like receptor signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	6.41×10^{-6}
GO:0050679, Positive regulation of epithelial cell proliferation	FGF2, CTNNB1, AGGF1, HIF1A, TNFSF12, ARNT, MYC, JUN, FGF1	9.31×10^{-6}
GO:0034142, Toll-like receptor 4 signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	9.31×10^{-6}
GO:0030834, Regulation of actin filament depolymerization	SPTB, SPTBN1, SPTAN1, SPTA1, ADD1, EPB49	1.09×10^{-5}
GO:0030837, Negative regulation of actin filament polymerization	SPTB, SPTBN1, SPTAN1, SPTA1, ADD1, EPB49	2.37×10^{-5}
GO:0002224, Toll-like receptor signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	2.48×10^{-5}
GO:0008629, Induction of apoptosis by intracellular signals	PML, BRCA1, YWHAB, TP53, BID, CDKN1A, RNF7, IFI16	2.92×10^{-5}
GO:0050851, Antigen receptor-mediated signaling pathway	WAS, MEF2C, RELA, PSEN2, PTPRC, PAK1, PTEN, NFKBIA	8.55×10^{-5}
GO:0002221, Pattern recognition receptor signaling pathway	ATF2, CDK1, MEF2C, FOS, RELA, JUN, ATF1, NFKBIA	9.15×10^{-5}
GO:0001936, Regulation of endothelial cell proliferation	FGF2, AGGF1, CAV1, HIF1A, TNFSF12, ARNT, JUN	9.47×10^{-5}
MIMAT000077(hsa-miR-22-3p)		
GO term	Genes	Adj. p-value
GO:0035583, Negative regulation of transforming growth factor beta receptor signaling pathway by extracellular sequestering of TGFbeta	FBN2, FBN1, LTBP1	1.45×10^{-5}
MIMAT0000265(hsa-miR-204-5p)		
GO term	Genes	Adj. p-value
GO:0035583, Negative regulation of transforming growth factor beta receptor signaling pathway by extracellular sequestering of TGFbeta	FBN1, FBN2, LTBP1	6.31×10^{-5}

Table S7. GOBO survival analysis results. Genes which was annotated with the specified GO term of proteins in the PIN would be used as input gene set for GOBO analysis.
 * $p < 0.05$, ** $p < 0.01$; *** $p < 0.001$.

miRNA	GO term	<i>p</i> -value
MIMAT0000064(hsa-let-7c)	GO:0015630, Microtubule cytoskeleton	9.97×10^{-6} ***
	GO:0043067, Regulation of programmed cell death	0.268067
	GO:0043069, Negative regulation of programmed cell death	0.0390439 *
	GO:0060548, Negative regulation of cell death	0.0390439 *
MIMAT0000076(hsa-miR-21-5p)	GO:0007173, Epidermal growth factor receptor signaling pathway	0.721139
	GO:0043067, Regulation of programmed cell death	0.266312
MIMAT0000077(hsa-miR-22-3p)	GO:0035583, Negative regulation of transforming growth factor beta receptor signaling pathway by extracellular sequestering of TGFbeta	0.940727
	GO:0003376, Sphingosine-1-phosphate signaling pathway	0.062202
	GO:0006917, Induction of apoptosis	0.048584 *
	GO:0007169, Transmembrane receptor protein tyrosine kinase signaling pathway	0.050408
	GO:0007187, G-protein signaling, coupled to cyclic nucleotide second messenger	0.289466
	GO:0007188, G-protein signaling, coupled to cAMP nucleotide second messenger	0.687572
	GO:0010942, Positive regulation of cell death	0.356228
	GO:0012502, Induction of programmed cell death	0.048584 *
	GO:0019933, CAMP-mediated signaling	0.687572
MIMAT0000089(hsa-miR-31-5p)	GO:0019935, Cyclic-nucleotide-mediated signaling	0.289466
	GO:0043067, Regulation of programmed cell death	0.694486
	GO:0043068, Positive regulation of programmed cell death	0.356228
	GO:0048011, Nerve growth factor receptor signaling pathway	0.154543
	GO:0035583, Negative regulation of transforming growth factor beta receptor signaling pathway by extracellular sequestering of TGFbeta	0.940727
	GO:0035583, Negative regulation of transforming growth factor beta receptor signaling pathway by extracellular sequestering of TGFbeta	0.940727
	GO:0002224, Toll-like receptor signaling pathway	0.380928
MIMAT0000265(hsa-miR-204-5p)	GO:0002755, MyD88-dependent toll-like receptor signaling pathway	0.380928
	GO:0002756, MyD88-independent toll-like receptor signaling pathway	0.380928
	GO:0006916, Anti-apoptosis	0.0593
	GO:0006917, Induction of apoptosis	0.618064
	GO:0007169, Transmembrane receptor protein tyrosine kinase signaling pathway	0.776269
	GO:0008284, Positive regulation of cell proliferation	0.882324
MIMAT0000423(hsa-miR-125b-5p)	GO:0008285, Negative regulation of cell proliferation	0.883393
	GO:0008629, Induction of apoptosis by intracellular signals	0.073118
	GO:0010942, Positive regulation of cell death	0.972892
	GO:0012502, Induction of programmed cell death	0.618064
	GO:0015629, Actin cytoskeleton	0.596528

Table S7. Cont.

miRNA	GO term	p-value
MIMAT0000423(hsa-miR-125b-5p)	GO:0015630, Microtubule cytoskeleton	0.028245 *
	GO:0030834, Regulation of actin filament depolymerization	0.654383
	GO:0030835, Negative regulation of actin filament depolymerization	0.654383
	GO:0030837, Negative regulation of actin filament polymerization	0.654383
	GO:0034130, Toll-like receptor 1 signaling pathway	0.380928
	GO:0034134, Toll-like receptor 2 signaling pathway	0.380928
	GO:0034138, Toll-like receptor 3 signaling pathway	0.380928
	GO:0034142, Toll-like receptor 4 signaling pathway	0.380928
	GO:0035666, TRIF-dependent toll-like receptor signaling pathway	0.380928
	GO:0043067, Regulation of programmed cell death	0.643418
	GO:0043068, Positive regulation of programmed cell death	0.972892
	GO:0043069, Negative regulation of programmed cell death	0.492576
	GO:0048011, Nerve growth factor receptor signaling pathway	0.171634
	GO:0050678, Regulation of epithelial cell proliferation	0.002205 **
	GO:0050679, Positive regulation of epithelial cell proliferation	0.205483
	GO:0051693, Actin filament capping	0.654383
	GO:0060548, Negative regulation of cell death	0.413665
MIMAT0000437(hsa-miR-145-5p)	GO:0007178, Transmembrane receptor protein serine/threonine kinase signaling pathway	0.196953
	GO:0030509, BMP signaling pathway	0.196953
	GO:0090092, Regulation of transmembrane receptor protein serine/threonine kinase signaling pathway	0.843529
	GO:0001936, Regulation of endothelial cell proliferation	0.115146
	GO:0002221, Pattern recognition receptor signaling pathway	0.380928
	GO:0002224, Toll-like receptor signaling pathway	0.380928
	GO:0002755, MyD88-dependent toll-like receptor signaling pathway	0.380928
	GO:0002756, MyD88-independent toll-like receptor signaling pathway	0.380928
MIMAT0000443(hsa-miR-125a-5p)	GO:0006916, Anti-apoptosis	0.0593
	GO:0006917, Induction of apoptosis	0.618064
	GO:0007169, Transmembrane receptor protein tyrosine kinase signaling pathway	0.776269
	GO:0008284, Positive regulation of cell proliferation	0.882324
	GO:0008285, Negative regulation of cell proliferation	0.883393
	GO:0008629, Induction of apoptosis by intracellular signals	0.073118
	GO:0010942, Positive regulation of cell death	0.972892
	GO:0012502, Induction of programmed cell death	0.618064

Table S7. Cont.

miRNA	GO term	p-value
MIMAT0000443(hsa-miR-125a-5p)	GO:0015629, Actin cytoskeleton	0.596528
	GO:0015630, Microtubule cytoskeleton	0.028245 *
	GO:0030834, Regulation of actin filament depolymerization	0.654383
	GO:0030835, Negative regulation of actin filament depolymerization	0.654383
	GO:0030837, Negative regulation of actin filament polymerization	0.654383
	GO:0034130, Toll-like receptor 1 signaling pathway	0.380928
	GO:0034134, Toll-like receptor 2 signaling pathway	0.380928
	GO:0034138, Toll-like receptor 3 signaling pathway	0.380928
	GO:0034142, Toll-like receptor 4 signaling pathway	0.380928
	GO:0035666, TRIF-dependent toll-like receptor signaling pathway	0.380928
	GO:0043067, Regulation of programmed cell death	0.643418
	GO:0043068, Positive regulation of programmed cell death	0.972892
	GO:0043069, Negative regulation of programmed cell death	0.492576
	GO:0048011, Nerve growth factor receptor signaling pathway	0.171634
	GO:0050678, Regulation of epithelial cell proliferation	0.002205 **
	GO:0050679, Positive regulation of epithelial cell proliferation	0.205483
	GO:0050851, Antigen receptor-mediated signaling pathway	0.103325
	GO:0051693, Actin filament capping	0.654383
	GO:0060548, Negative regulation of cell death	0.413665
MIMAT0002820(hsa-miR-497-5p)	GO:0001959, Regulation of cytokine-mediated signaling pathway	0.06699
	GO:0006917, Induction of apoptosis	0.142401
	GO:0007169, Transmembrane receptor protein tyrosine kinase signaling pathway	0.837635
	GO:0007173, Epidermal growth factor receptor signaling pathway	0.387447
	GO:0007178, Transmembrane receptor protein serine/threonine kinase signaling pathway	0.240167
	GO:0007179, Transforming growth factor beta receptor signaling pathway	0.017876 *
	GO:0008284, Positive regulation of cell proliferation	0.430255
	GO:0008285, Negative regulation of cell proliferation	0.149994
	GO:0008543, Fibroblast growth factor receptor signaling pathway	0.521978
	GO:0010942, Positive regulation of cell death	0.237692
	GO:0012502, Induction of programmed cell death	0.142401
	GO:0015629, Actin cytoskeleton	0.228804
	GO:0015630, Microtubule cytoskeleton	0.18331
	GO:0017015, Regulation of transforming growth factor beta receptor signaling pathway	0.128773

Table S7. Cont.

miRNA	GO term	p-value
MIMAT0002820(hsa-miR-497-5p)	GO:0030509, BMP signaling pathway	0.39837
	GO:0030521, Androgen receptor signaling pathway	0.383811
	GO:0032956, Regulation of actin cytoskeleton organization	0.91762
	GO:0042058, Regulation of epidermal growth factor receptor signaling pathway	0.934045
	GO:0042059, Negative regulation of epidermal growth factor receptor signaling pathway	0.789492
	GO:0043067, Regulation of programmed cell death	0.111544
	GO:0043068, Positive regulation of programmed cell death	0.237692
	GO:0043069, Negative regulation of programmed cell death	0.856892
	GO:0048011, Nerve growth factor receptor signaling pathway	0.471986
	GO:0060548, Negative regulation of cell death	0.667437
	GO:0070302, Regulation of stress-activated protein kinase signaling cascade	0.561032
	GO:0090092, Regulation of transmembrane receptor protein serine/threonine kinase signaling pathway	0.182314
	GO:0006917, Induction of apoptosis	0.489781
	GO:0007169, Transmembrane receptor protein tyrosine kinase signaling pathway	0.171689
	GO:0007173, Epidermal growth factor receptor signaling pathway	0.449696
	GO:0008284, Positive regulation of cell proliferation	0.05916
	GO:0008286, Insulin receptor signaling pathway	0.237933
MIMAT0002856(hsa-miR-520d-3p)	GO:0008543, Fibroblast growth factor receptor signaling pathway	0.159318
	GO:0008629, Induction of apoptosis by intracellular signals	0.502822
	GO:0010942, Positive regulation of cell death	0.076906
	GO:0012502, Induction of programmed cell death	0.489781
	GO:0015630, Microtubule cytoskeleton	0.000292 ***
	GO:0042058, Regulation of epidermal growth factor receptor signaling pathway	0.762633
	GO:0042059, Negative regulation of epidermal growth factor receptor signaling pathway	0.826854
	GO:0043067, Regulation of programmed cell death	0.740092
	GO:0043068, Positive regulation of programmed cell death	0.076906
	GO:0043069, Negative regulation of programmed cell death	0.067499
GO:0048011, Nerve growth factor receptor signaling pathway	GO:0048011, Nerve growth factor receptor signaling pathway	0.014926 *
	GO:0051988, Regulation of attachment of spindle microtubules to kinetochore	$7.48 \times 10^{-6} ***$
	GO:0060548, Negative regulation of cell death	0.213035

Table S8. Pathophysiological characteristics of miRNA array data used in ROC curve analysis.

Sample Name	ER	PR	HER	TNM	Stage	Grade
S621T1	0	0	0	pT1N0M0	I	2
S434T1	1	0	1	T2N1M0	IIB	3
S403T1	0	1	0	T2N1M0	IIB	2
S459T1	1	0	0	T4N0M1	IV	1
S455N1	1	0	0	T3N3M1	IV	3
S545T1	0	0	0	pT2N0M0	IIA	3
S173N1	0	0	1	T2N3M0	IIIC	3
S363T1	1	1	0	T2N1M0	IIB	1
S909T1	1	1	1	pT3N3aM0	IIIC	(Unknown)
S645T1	0	1	1	pT1bN0M0	I	3
S898T1	1	0	0	pT2N0(i-)M0	IIA	(Unknown)
S201T1	1	0	0	T1N1M0	IIA	2
S631T1	1	1	1	T2N3aM1	IV	2
S303T1	0	0	1	T2N0M0	IIA	2
S502T1	0	0	0	pT3N0M0	IIB	3
S498N1	1	1	1	pT1cN1aM0	IIA	2
S536T1	1	0	1	T1cN1miM0	IIA	2
S660T1	0	0	1	T1N0M0	I	3
S358N1	0	0	1	T2pN2M0	IIIA	2
S665T1	0	0	0	T2N3M0	IIIC	3
S475T1	0	0	1	pT1cN0M0	I	3
S423T1	1	1	0	T2N3M0	IIIC	1
S507T1	0	0	0	pT1cN1aM0	IIA	3
S891T1	1	0	0	pT2NxM0	IIA	2
S422T1	1	1	0	T2N0M0	IIA	1
S961T1	0	0	1	T2N2aM0	IIIA	2
S622T1	0	0	0	pT2N0M0	IIA	2
S454T1	0	0	0	T1cN0M0	I	2
S433T1	1	0	0	T2N0M0	IIA	1
S673T1	0	0	0	pT2N0M0	IIA	2
S450T1	0	0	1	T2N1M0	IIB	3
S430T1	1	1	0	T2N3M0	IIIC	2
S437T1	1	0	0	T3N1M0	IIIA	3
S574T1	0	0	0	T2N0M0	IIA	2
S401T1	1	1	0	T1N0M0	I	1
S427N1	0	0	0	T3N1M0	IIIA	2
S894T1	0	0	0	pT1cN0M0	I	2
S929T1	1	0	0	pT2N0M0	IIA	2
S173T1	0	0	1	T2N3M0	IIIC	3
S622N1	0	0	0	pT2N0M0	IIA	2
S562T1	1	1	1	pT1aN0M0	I	3
S602T1	0	0	0	pT1cN0M0	I	3
S490T1	1	1	1	pT2N0M0	IIA	2
S677T1	0	0	0	pT2N1M0	IIB	3
S881T1	0	0	0	pT2N1aM0	IIB	3
S619T1	0	0	0	pT2N0M0	IIA	3
S446T1	0	0	0	T2N2M0	IIIA	3
S446T2	0	0	0	T2N2M0	IIIA	3
S453T1	1	1	1	T2N1M0	IIB	2
S562N1	1	1	1	pT1aN0M0	I	3
S557T1	0	0	0	pT2N0M0	IIA	3

Table S8. *Cont.*

Sample Name	ER	PR	HER	TNM	Stage	Grade
S594T1	1	1	1	pT2N3aM0	IIIC	3
S582T1	0	0	0	pT2N0M0	IIA	3
S358T1	0	0	1	T2pN2M0	IIIA	2
S368T1	0	0	1	T3N3M0	IIIC	2
S175T1	1	1	0	T3N3Mx	IIIC	3
S357T1	0	0	0	T2pN1M0	IIIC	3
S653T1	0	0	0	pT3N3aM0	IIIC	2
S722T1	1	1	1	pT1N0M0	I	3
S593T1	0	0	0	pT1N0M0	I	3
S543T1	1	1	1	pT2N1M0	IIB	2
S498T1	1	1	1	pT1cN1aM0	IIA	2
S389T1	1	0	0	T2N1M0	IIB	3
S614T1	0	1	1	TxN0M1	IIIA	(Unknown)
S536N1	1	0	1	T1cN1miM0	IIA	2
S462T1	1	1	0	T3N3M0	IIIC	2
S477T1	0	0	0	pT1cN0M0	I	3
S917T1	0	0	0	T2N0M0	IIA	(Unknown)
S213T1	0	0	1	T1cN1M0	IIA	3
S628T1	0	0	0	T1N0M0	I	2
S291T1	0	0	0	T3pN2M0	IIIA	3
S593N1	0	0	0	pT1N0M0	I	3
S418T1	1	1	0	T2N3M0	IIIC	2
S420T1	1	1	0	T1N0M0	I	2
S363N1	1	1	0	T2N1M0	IIB	1
S629T1	1	0	1	pT1N0M0	I	2
S586T1	1	1	1	pT2N3aM0	IIIC	2
S415T1	0	1	0	T2N2M0	IIIA	2
S439T1	1	1	0	T2N2M0	IIIA	2
S941N1	0	0	0	pT1cN1aM0	IIA	3
S893T1	0	0	0	pT2N1micM0	IIB	3
S380T1	1	0	0	T2N1M0	IIB	2
S906N1	1	1	1	pT3N3aM0	IIIC	2
S918T1	0	0	0	pT2N0M0	IIA	3
S400T1	0	0	1	T2N0M0	IIA	2
S328T1	0	0	0	T1cpN0M0	I	3
S367T1	1	0	0	T1N1M0	IIIA	1
S420N1	1	1	0	T1N0M0	I	2
S922T1	0	0	0	pT1cN0(i-)M0	I	3
S896T1	0	0	1	pT2N1aM0	IIB	2
S410T1	0	1	1	T1N0M0	I	3
S572T1	0	0	1	T4N1aM0	IIIC	3
S448T1	0	0	0	T1N0M0	I	3
S207T1	0	0	1	T2N0M0	IIA	3
S604T1	0	0	1	pT2N1M0	IIB	3
S379T1	0	0	1	T2N2M0	IIIA	2
S906T1	1	1	1	pT3N3aM0	IIIC	2
S941T2	0	0	0	pT1cN1aM0	IIA	3
S941T1	0	0	0	pT1cN1aM0	IIA	3
S375T1	1	1	0	T1N0M0	I	2
S427T1	0	0	0	T3N1M0	IIIA	2
S417T1	0	0	0	pT2N0M0	IIA	3

Table S8. Cont.

Sample Name	ER	PR	HER	TNM	Stage
S180T1	1	0	0	T4NxM0	IIIC
S455T1	1	0	0	T3N3M1	IV
S887T1	0	0	0	pT2N0M0	IIA
S445T1	1	0	1	T2N1M0	IIB
S469T1	1	1	0	T2N0M0	IIA
S469T2	1	1	0	T2N0M0	IIA
S483T1	1	1	0	T2N3M0	IIIC
S444T1	1	1	0	T1NxM0	(Unknown)
S909N1	1	1	1	pT3N3aM0	IIIC
S464T1	1	1	0	T1N0M0	I
S894N1	0	0	0	pT1cN0M0	I
S698T1	1	1	1	pT1cN0M0	I
S452T1	0	0	0	T1N0M0	I
S474T1	(Unknown)	(Unknown)	(Unknown)	(Unknown)	(Unknown)

References

1. Jemal, A.; Center, M.M.; DeSantis, C.; Ward, E.M. Global patterns of cancer incidence and mortality rates and trends. *Cancer Epidemiol. Biomark. Prev.* **2010**, *19*, 1893–1907.
2. Veronesi, U.; Boyle, P.; Goldhirsch, A.; Orecchia, R.; Viale, G. Breast cancer. *Lancet* **2005**, *365*, 1727–1741.
3. Cho, E.; Spiegelman, D.; Hunter, D.J.; Chen, W.Y.; Stampfer, M.J.; Colditz, G.A.; Willett, W.C. Premenopausal fat intake and risk of breast cancer. *J. Natl. Cancer Inst.* **2003**, *95*, 1079–1085.
4. Huttenhofer, A.; Schattner, P.; Polacek, N. Non-coding RNAs: Hope or hype? *Trends Genet.* **2005**, *21*, 289–297.
5. Kim, V.N.; Han, J.; Siomi, M.C. Biogenesis of small RNAs in animals. *Nat. Rev. Mol. Cell Biol.* **2009**, *10*, 126–139.
6. Kwak, P.B.; Iwasaki, S.; Tomari, Y. The microRNA pathway and cancer. *Cancer Sci.* **2010**, *101*, 2309–2315.
7. Hutvagner, G.; Zamore, P.D. A microRNA in a multiple-turnover RNAi enzyme complex. *Science* **2002**, *297*, 2056–2060.
8. Mourelatos, Z.; Dostie, J.; Paushkin, S.; Sharma, A.; Charroux, B.; Abel, L.; Rappaport, J.; Mann, M.; Dreyfuss, G. miRNPs: A novel class of ribonucleoproteins containing numerous microRNAs. *Genes Dev.* **2002**, *16*, 720–728.
9. Nelson, P.T.; Hatzigeorgiou, A.G.; Mourelatos, Z. miRNP:mRNA association in polyribosomes in a human neuronal cell line. *RNA* **2004**, *10*, 387–394.
10. Meister, G.; Landthaler, M.; Patkaniowska, A.; Dorsett, Y.; Teng, G.; Tuschl, T. Human Argonaute2 mediates RNA cleavage targeted by miRNAs and siRNAs. *Mol. Cell* **2004**, *15*, 185–197.
11. Calin, G.A.; Dumitru, C.D.; Shimizu, M.; Bichi, R.; Zupo, S.; Noch, E.; Aldler, H.; Rattan, S.; Keating, M.; Rai, K.; et al. Frequent deletions and down-regulation of micro-RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 15524–15529.

12. Zhang, B.; Pan, X.; Cobb, G.P.; Anderson, T.A. microRNAs as oncogenes and tumor suppressors. *Dev. Biol.* **2007**, *302*, 1–12.
13. Meng, F.; Henson, R.; Wehbe-Janek, H.; Ghoshal, K.; Jacob, S.T.; Patel, T. MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer. *Gastroenterology* **2007**, *133*, 647–658.
14. Rask, L.; Balslev, E.; Jorgensen, S.; Eriksen, J.; Flyger, H.; Moller, S.; Hogdall, E.; Litman, T.; Nielsen, B.S. High expression of miR-21 in tumor stroma correlates with increased cancer cell proliferation in human breast cancer. *APMIS* **2011**, *119*, 663–673.
15. Zaman, M.S.; Shahryari, V.; Deng, G.; Thamminana, S.; Saini, S.; Majid, S.; Chang, I.; Hirata, H.; Ueno, K.; Yamamura, S.; et al. Up-regulation of microRNA-21 correlates with lower kidney cancer survival. *PLoS One* **2012**, *7*, e31060.
16. Nadiminty, N.; Tummala, R.; Lou, W.; Zhu, Y.; Shi, X.B.; Zou, J.X.; Chen, H.; Zhang, J.; Chen, X.; Luo, J.; et al. MicroRNA let-7c is downregulated in prostate cancer and suppresses prostate cancer growth. *PLoS One* **2012**, *7*, e32832.
17. Wang, X.F.; Shi, Z.M.; Wang, X.R.; Cao, L.; Wang, Y.Y.; Zhang, J.X.; Yin, Y.; Luo, H.; Kang, C.S.; Liu, N.; et al. MiR-181d acts as a tumor suppressor in glioma by targeting K-ras and Bcl-2. *J. Cancer Res. Clin. Oncol.* **2012**, *138*, 573–584.
18. Zhang, Y.; Yan, L.X.; Wu, Q.N.; Du, Z.M.; Chen, J.; Liao, D.Z.; Huang, M.Y.; Hou, J.H.; Wu, Q.L.; Zeng, M.S.; et al. miR-125b is methylated and functions as a tumor suppressor by regulating the ETS1 proto-oncogene in human invasive breast cancer. *Cancer Res.* **2011**, *71*, 3552–3562.
19. Sachdeva, M.; Mo, Y.Y. MicroRNA-145 suppresses cell invasion and metastasis by directly targeting mucin 1. *Cancer Res.* **2010**, *70*, 378–387.
20. Guo, X.; Wu, Y.; Hartley, R.S. MicroRNA-125a represses cell growth by targeting HuR in breast cancer. *RNA Biol.* **2009**, *6*, 575–583.
21. Jin, H.; Tuo, W.; Lian, H.; Liu, Q.; Zhu, X.Q.; Gao, H. Strategies to identify microRNA targets: New advances. *N. Biotechnol.* **2010**, *27*, 734–738.
22. Friedman, R.C.; Farh, K.K.; Burge, C.B.; Bartel, D.P. Most mammalian mRNAs are conserved targets of microRNAs. *Genome Res.* **2009**, *19*, 92–105.
23. Huang, J.C.; Babak, T.; Corson, T.W.; Chua, G.; Khan, S.; Gallie, B.L.; Hughes, T.R.; Blencowe, B.J.; Frey, B.J.; Morris, Q.D. Using expression profiling data to identify human microRNA targets. *Nat. Methods* **2007**, *4*, 1045–1049.
24. Liu, B.; Li, J.; Tsykin, A.; Liu, L.; Gaur, A.B.; Goodall, G.J. Exploring complex miRNA-mRNA interactions with Bayesian networks by splitting-averaging strategy. *BMC Bioinforma.* **2009**, *10*, 408.
25. Tseng, C.W.; Lin, C.C.; Chen, C.N.; Huang, H.C.; Juan, H.F. Integrative network analysis reveals active microRNAs and their functions in gastric cancer. *BMC Syst. Biol.* **2011**, *5*, 99.
26. Farazi, T.A.; Horlings, H.M.; Ten Hoeve, J.J.; Mihailovic, A.; Halfwerk, H.; Morozov, P.; Brown, M.; Hafner, M.; Reyal, F.; van Kouwenhove, M.; et al. MicroRNA sequence and expression analysis in breast tumors by deep sequencing. *Cancer Res.* **2011**, *71*, 4443–4453.

27. Derfoul, A.; Juan, A.H.; Difflippantonio, M.J.; Palanisamy, N.; Ried, T.; Sartorelli, V. Decreased microRNA-214 levels in breast cancer cells coincides with increased cell proliferation, invasion and accumulation of the Polycomb Ezh2 methyltransferase. *Carcinogenesis* **2011**, *32*, 1607–1614.
28. Heyn, H.; Engelmann, M.; Schreek, S.; Ahrens, P.; Lehmann, U.; Kreipe, H.; Schlegelberger, B.; Beger, C. MicroRNA miR-335 is crucial for the BRCA1 regulatory cascade in breast cancer development. *Int. J. Cancer* **2011**, *129*, 2797–2806.
29. Sempere, L.F.; Christensen, M.; Silahtaroglu, A.; Bak, M.; Heath, C.V.; Schwartz, G.; Wells, W.; Kauppinen, S.; Cole, C.N. Altered MicroRNA expression confined to specific epithelial cell subpopulations in breast cancer. *Cancer Res.* **2007**, *67*, 11612–11620.
30. Zhu, X.M.; Wu, L.J.; Xu, J.; Yang, R.; Wu, F.S. Let-7c microRNA expression and clinical significance in hepatocellular carcinoma. *J. Int. Med. Res.* **2011**, *39*, 2323–2329.
31. Iyevleva, A.G.; Kuligina, E.; Mitiushkina, N.V.; Togo, A.V.; Miki, Y.; Imyanitov, E.N. High level of miR-21, miR-10b, and miR-31 expression in bilateral vs. unilateral breast carcinomas. *Breast Cancer Res. Treat.* **2012**, *131*, 1049–1059.
32. Rutnam, Z.J.; Yang, B.B. The involvement of microRNAs in malignant transformation. *Histol. Histopathol.* **2012**, *27*, 1263–1270.
33. Liu, H. MicroRNAs in breast cancer initiation and progression. *Cell Mol. Life Sci.* **2012**, *69*, 3587–3599.
34. Zhang, Z.J.; Ma, S.L. miRNAs in breast cancer tumorigenesis (Review). *Oncol. Rep.* **2012**, *27*, 903–910.
35. Mathivanan, S.; Periaswamy, B.; Gandhi, T.K.; Kandasamy, K.; Suresh, S.; Mohmood, R.; Ramachandra, Y.L.; Pandey, A. An evaluation of human protein-protein interaction data in the public domain. *BMC Bioinforma.* **2006**, *7*, S19.
36. Halbert, C.H.; Kumanyika, S.; Bowman, M.; Bellamy, S.L.; Briggs, V.; Brown, S.; Bryant, B.; Delmoor, E.; Johnson, J.C.; Purnell, J.; et al. Participation rates and representativeness of African Americans recruited to a health promotion program. *Health Educ. Res.* **2010**, *25*, 6–13.
37. Nicolas Diaz-Chico, B.; German Rodriguez, F.; Gonzalez, A.; Ramirez, R.; Bilbao, C.; Cabrera de Leon, A.; Aguirre Jaime, A.; Chirino, R.; Navarro, D.; Diaz-Chico, J.C. Androgens and androgen receptors in breast cancer. *J. Steroid Biochem. Mol. Biol.* **2007**, *105*, 1–15.
38. Sethi, N.; Kang, Y. Dysregulation of developmental pathways in bone metastasis. *Bone* **2011**, *48*, 16–22.
39. Liao, S.J.; Zhou, Y.H.; Yuan, Y.; Li, D.; Wu, F.H.; Wang, Q.; Zhu, J.H.; Yan, B.; Wei, J.J.; Zhang, G.M.; et al. Triggering of Toll-like receptor 4 on metastatic breast cancer cells promotes alphavbeta3-mediated adhesion and invasive migration. *Breast Cancer Res. Treat.* **2011**, *133*, 853–863.
40. Jiang, P.; Enomoto, A.; Takahashi, M. Cell biology of the movement of breast cancer cells: Intracellular signalling and the actin cytoskeleton. *Cancer Lett.* **2009**, *284*, 122–130.
41. Garcia, D.M.; Baek, D.; Shin, C.; Bell, G.W.; Grimson, A.; Bartel, D.P. Weak seed-pairing stability and high target-site abundance decrease the proficiency of lsy-6 and other microRNAs. *Nat. Struct. Mol. Biol.* **2011**, *18*, 1139–1146.
42. Lewis, B.P.; Burge, C.B.; Bartel, D.P. Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell* **2005**, *120*, 15–20.

43. Lall, S.; Grun, D.; Krek, A.; Chen, K.; Wang, Y.L.; Dewey, C.N.; Sood, P.; Colombo, T.; Bray, N.; Macmenamin, P.; *et al.* A genome-wide map of conserved microRNA targets in *C. elegans*. *Curr. Biol.* **2006**, *16*, 460–471.
44. Krek, A.; Grun, D.; Poy, M.N.; Wolf, R.; Rosenberg, L.; Epstein, E.J.; MacMenamin, P.; da Piedade, I.; Gunsalus, K.C.; Stoffel, M.; *et al.* Combinatorial microRNA target predictions. *Nat. Genet.* **2005**, *37*, 495–500.
45. Betel, D.; Wilson, M.; Gabow, A.; Marks, D.S.; Sander, C. The microRNA.org resource: Targets and expression. *Nucleic Acids Res.* **2008**, *36*, D149–D153.
46. Tusher, V.G.; Tibshirani, R.; Chu, G. Significance analysis of microarrays applied to the ionizing radiation response. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 5116–5121.
47. Keshava Prasad, T.S.; Goel, R.; Kandasamy, K.; Keerthikumar, S.; Kumar, S.; Mathivanan, S.; Telikicherla, D.; Raju, R.; Shafreen, B.; Venugopal, A.; *et al.* Human Protein Reference Database—2009 update. *Nucleic Acids Res.* **2009**, *37*, D767–D772.
48. Benjamini, Y.; Yekutieli, D. The control of the false discovery rate in multiple testing under dependency. *Ann. Stat.* **2001**, *29*, 1165–1188.
49. Ringner, M.; Fredlund, E.; Hakkinen, J.; Borg, A.; Staaf, J. GOBO: Gene expression-based outcome for breast cancer online. *PLoS One* **2011**, *6*, e17911.
50. Sing, T.; Sander, O.; Beerenwinkel, N.; Lengauer, T. ROCR: Visualizing classifier performance in R. *Bioinformatics* **2005**, *21*, 3940–3941.
51. Hanley, J.A.; McNeil, B.J. The meaning and use of the area under a receiver operating characteristic (ROC) curve. *Radiology* **1982**, *143*, 29–36.

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