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All-Trans Retinoic Acid Treatment Is Associated with Prohibitin Expression in Renal Interstitial Fibrosis Rats

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Abstract: This study was performed to investigate the association of prohibitin with renal interstitial fibrosis (RIF) lesion and to explore the association of all-trans retinoic acid (ATRA) treatment with prohibitin expression in RIF rats. Rats were divided into three groups: the sham operation group (SHO), the model group subjected to unilateral ureteral obstruction (UUO), and the model group treated with ATRA (GA). Renal tissues were collected at 14 and 28 days after surgery, and the relevant indicators were detected. In comparison with the SHO group, the RIF index in the UUO group was markedly elevated ($p < 0.01$), and the RIF index in the GA group was alleviated compared with that in the UUO group ($p < 0.01$). Compared with the SHO group, the expression of prohibitin (protein or mRNA) in the UUO group was significantly reduced (each $p < 0.01$). Prohibitin expression in the GA group was markedly increased when compared with that in the UUO ($p < 0.01$). The expression of TGF- β 1 (protein and mRNA), protein expressions of Col-IV, fibronectin, α -SMA and cleaved Caspase-3, ROS generation and cell apoptosis index in the UUO group were markedly higher than those in the SHO group (all $p < 0.01$), and their expressions in the GA group were markedly down-regulated compared to those in the UUO group (all $p < 0.01$, respectively). The protein expression of prohibitin was negatively correlated with the RIF index, protein expression of TGF- β 1, Col-IV, fibronectin, α -SMA or cleaved Caspase-3, ROS generation and the cell apoptosis index (each $p < 0.01$). In conclusion, lower expression of prohibitin is associated with the RIF, and ATRA treatment is associated with increased prohibitin, which can prevent the progression of RIF.

Keywords: all-trans retinoic acid; prohibitin, extracellular matrix; cell apoptosis; renal interstitial fibrosis; unilateral ureteral obstruction

1. Introduction

Renal interstitial fibrosis (RIF) is the final common pathway leading to chronic kidney diseases and end-stage renal disease [1,2], and its effective therapeutic strategy is wanting at present. Unilateral ureteral obstruction (UUO) is a well-characterized model of experimental obstructive nephropathy, culminating in RIF [3,4]. Currently, investigations have found that the reactive oxidative species (ROS) can contribute to the progression of renal disease induced by UUO [5,6].

Prohibitin is a ubiquitous protein with a number of different molecular functions [7] and it is mainly located on the inner mitochondrial membrane and nuclei [8]. It plays a pivotal role in the processes of cell differentiation and apoptosis [9]. The over-expression of prohibitin could protect the mitochondria from ROS-induced injury [10]. The abnormal expression of prohibitin might be associated with RIF progression.

All-trans retinoic acid (ATRA), a natural derivative of vitamin A [11], has been shown to have a variety of functions, including a role as an antioxidant and regulator of cell differentiation and apoptosis [12,13]. Some investigations [14–16] reported that ATRA could play a protective role in some renal diseases, but the detailed mechanism was not clear. Interestingly, ATRA treatment can suppress the generation of ROS [12], and have an anti-oxidative effect [17]. As mentioned above, we drew up a hypothesis that ATRA treatment is associated with the expression of prohibitin. This investigation was performed to explore this hypothesis in RIF rats induced by UUO.

2. Results

2.1. Renal Morphology

Light microscopy showed more collagen deposition, fibroblast proliferation and diffused lymphocytes filtration in the renal tubulointerstitium of the UUO group when compared with those in the SHO group (blue arrowhead in Figure 1), but the lesion degree in the GA group was less serious than that in the UUO group (blue arrowhead in Figure 1). The RIF score in the UUO group was significantly increased over that in the SHO group, and the RIF index in the GA group was markedly alleviated compared with that in the UUO group (all $p < 0.01$; Figure 1).

Figure 1. (1) Statistical parameters in three groups. *: $p < 0.01$ compared with sham operation group (SHO), **: $p < 0.01$ compared with unilateral ureteral obstruction group (UUO); (2) Tissue characteristics in three groups. Masson staining for SHO group (A₁: 14-day; A₂: 28-day), UUO group (A₃: 14-day; A₄: 28-day) and model group treated with ATRA (GA) (A₅: 14-day; A₆: 28-day). Representative samples of immunohistochemical staining for TGF- β 1 (SHO: B₁: 14-day, B₂: 28-day; UUO: B₃: 14-day, B₄: 28-day; GA: B₅: 14-day; B₆: 28-day), Col-IV (SHO: C₁: 14-day, C₂: 28-day; UUO: C₃: 14-day, C₄: 28-day; GA: C₅: 14-day; C₆: 28-day), and fibronectin (SHO: D₁: 14-day, D₂: 28-day; UUO: D₃: 14-day; D₄: 28-day; GA: D₅: 14-day; D₆: 28-day) were observed in three groups. Magnification $\times 400$.

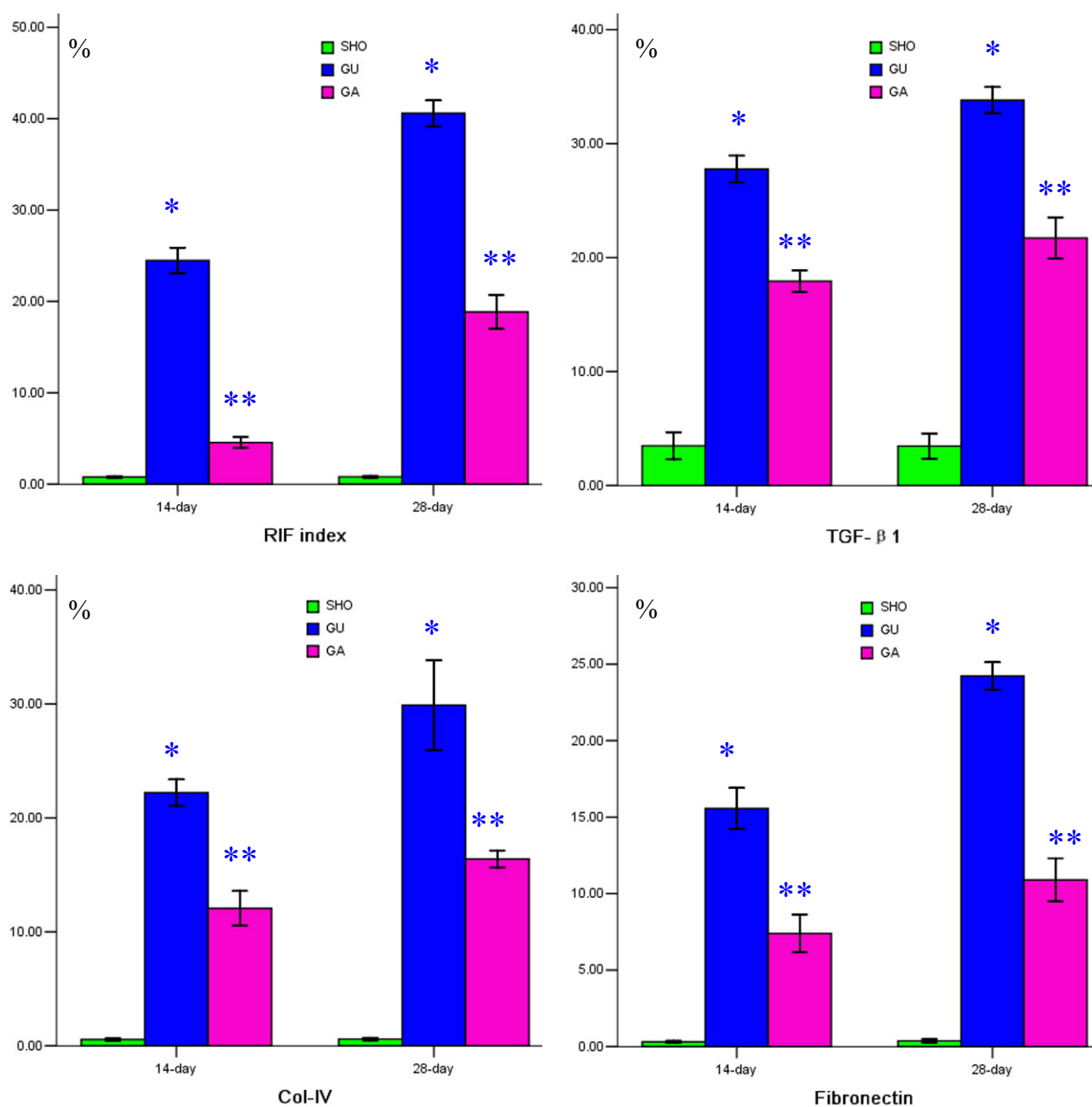
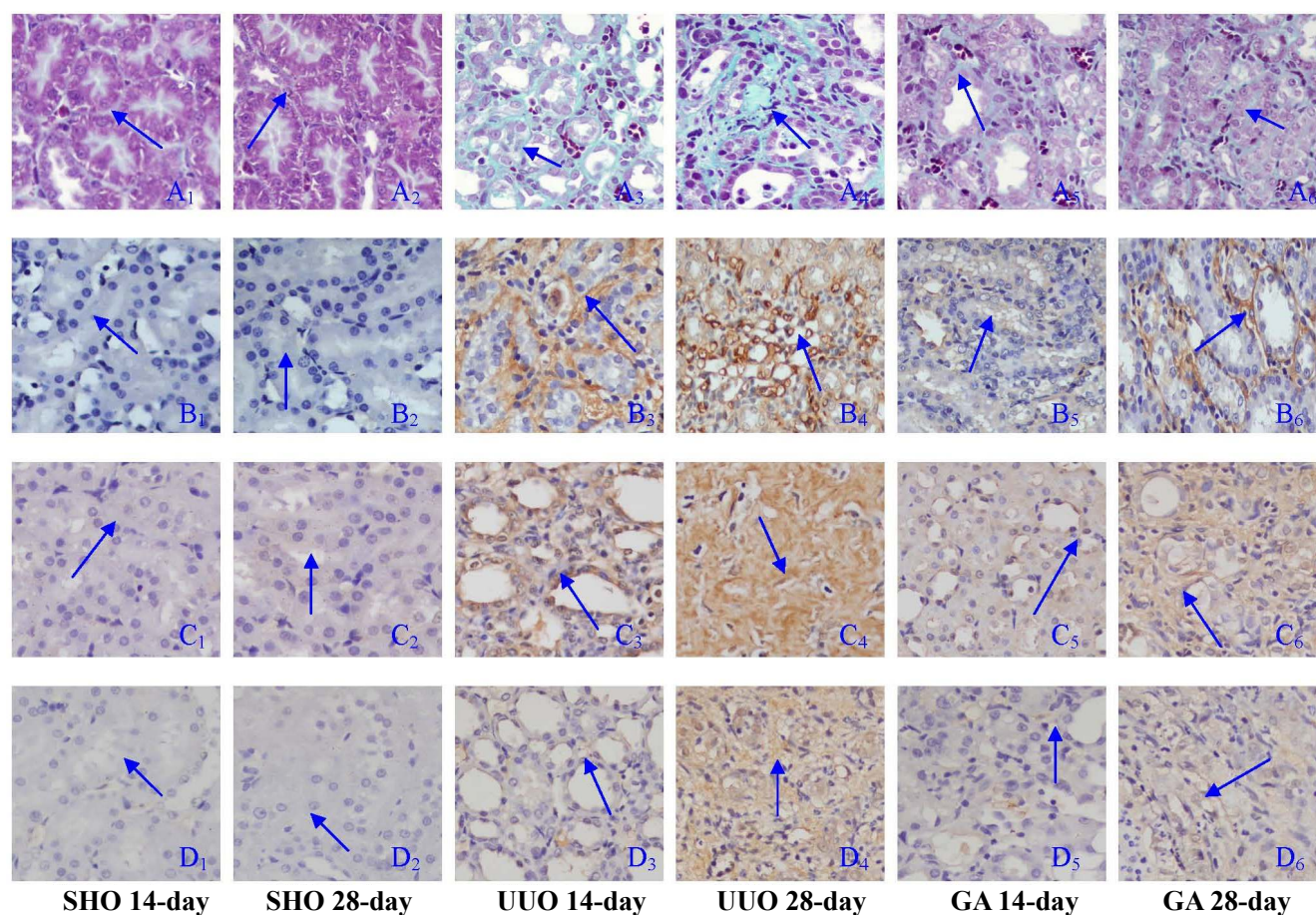


Figure 1. Cont.



2.2. Association of ATRA Treatment with TGF- β 1, Col-IV, Fibronectin, α -SMA, Prohibitin or Cleaved Caspase-3 Protein Expression

The staining of prohibitin in the SHO group was more significant than that in the UUO group or GA group (blue arrowhead in Figure 2). The stainings of TGF- β 1, Col-IV, fibronectin, α -SMA and cleaved Caspase-3 in the UUO group were much more significant compared with those in the SHO group / GA group (blue arrowhead in Figure 1 and 2). Comparison with that in the SHO group, the protein expression of prohibitin in the UUO group was significantly weakened, and protein expression of prohibitin in the ATRA treatment group was markedly increased over that in the UUO group ($p < 0.01$; Figure 2). The protein expressions of TGF- β 1, Col-IV, fibronectin, α -SMA and cleaved Caspase-3 in the UUO group were significantly increased when compared with those in the SHO group, and the TGF- β 1, Col-IV, fibronectin, α -SMA and cleaved Caspase-3 expressions in the GA group were lower than those in the UUO group (all $p < 0.01$; Figure 1 and 2). Protein expression of prohibitin was negatively correlated with the RIF index, protein expression of TGF- β 1, Col-IV, fibronectin, α -SMA or cleaved Caspase-3 ($r = -0.825, -0.786, -0.948, -0.817, -0.953, -0.863$; each $p < 0.01$, respectively).

Figure 2. (1) Statistical parameters for α -SMA, prohibitin, cleaved Caspase-3 and cell apoptosis index in three groups. *: $p < 0.01$ compared with SHO, **: $p < 0.01$ compared with UUO; **(2)** Tissue characteristics in three groups. Representative samples of immunohistochemical staining for α -SMA (SHO: E₁: 14-day, E₂: 28-day; UUO: E₃: 14-day, E₄: 28-day; GA: E₅: 14-day; E₆: 28-day), prohibitin (SHO: F₁: 14-day, F₂: 28-day; UUO: F₃: 14-day, F₄: 28-day; GA: F₅: 14-day; F₆: 28-day), and cleaved Caspase-3 (SHO: G₁: 14-day, G₂: 28-day; UUO: G₃: 14-day, G₄: 28-day; GA: G₅: 14-day; G₆: 28-day) were observed in three groups. Cell apoptosis also detected in three groups (SHO: H₁: 14-day, H₂: 28-day; UUO: H₃: 14-day, H₄: 28-day; GA: H₅: 14-day; H₆: 28-day). Magnification $\times 400$.

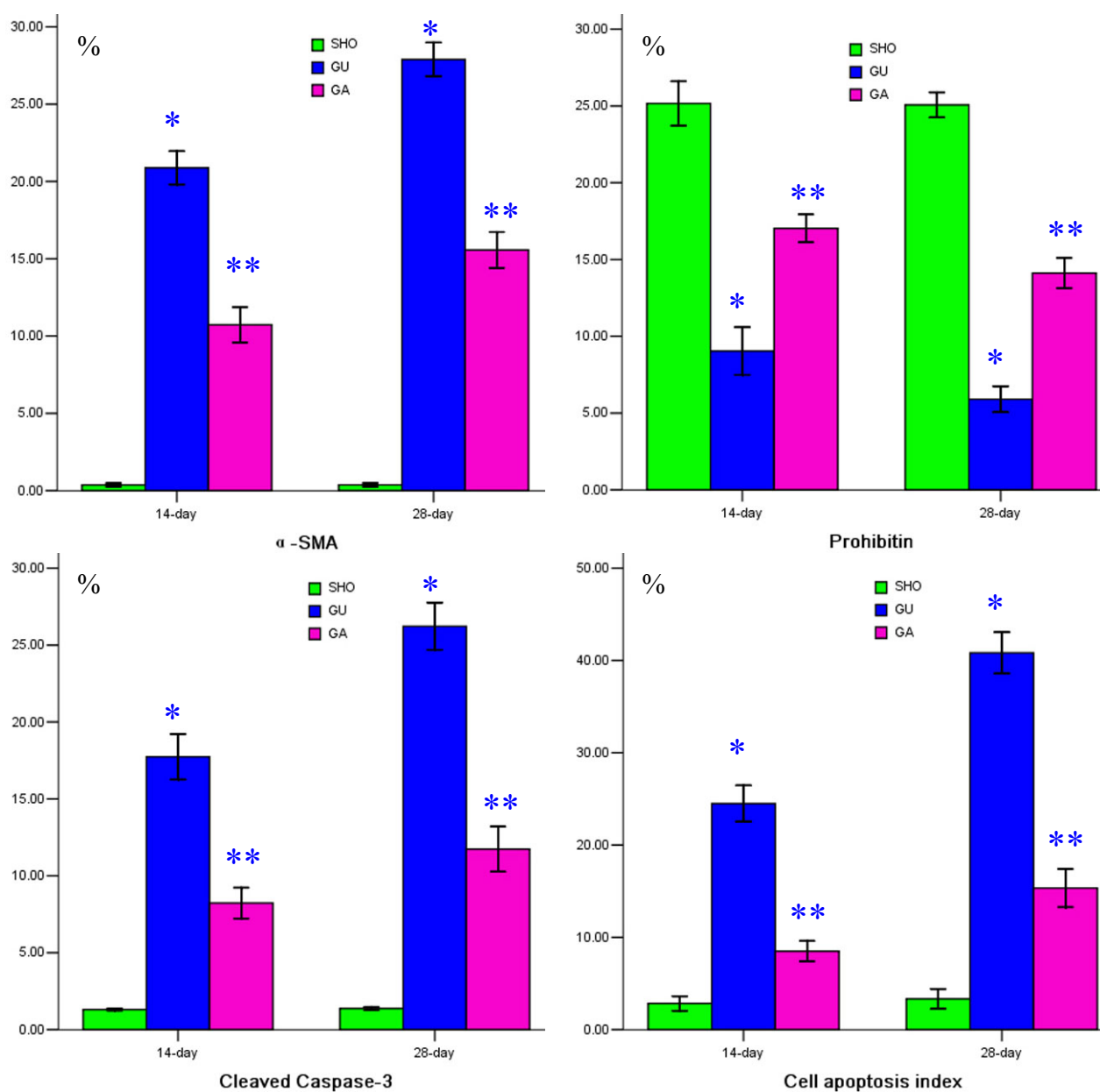
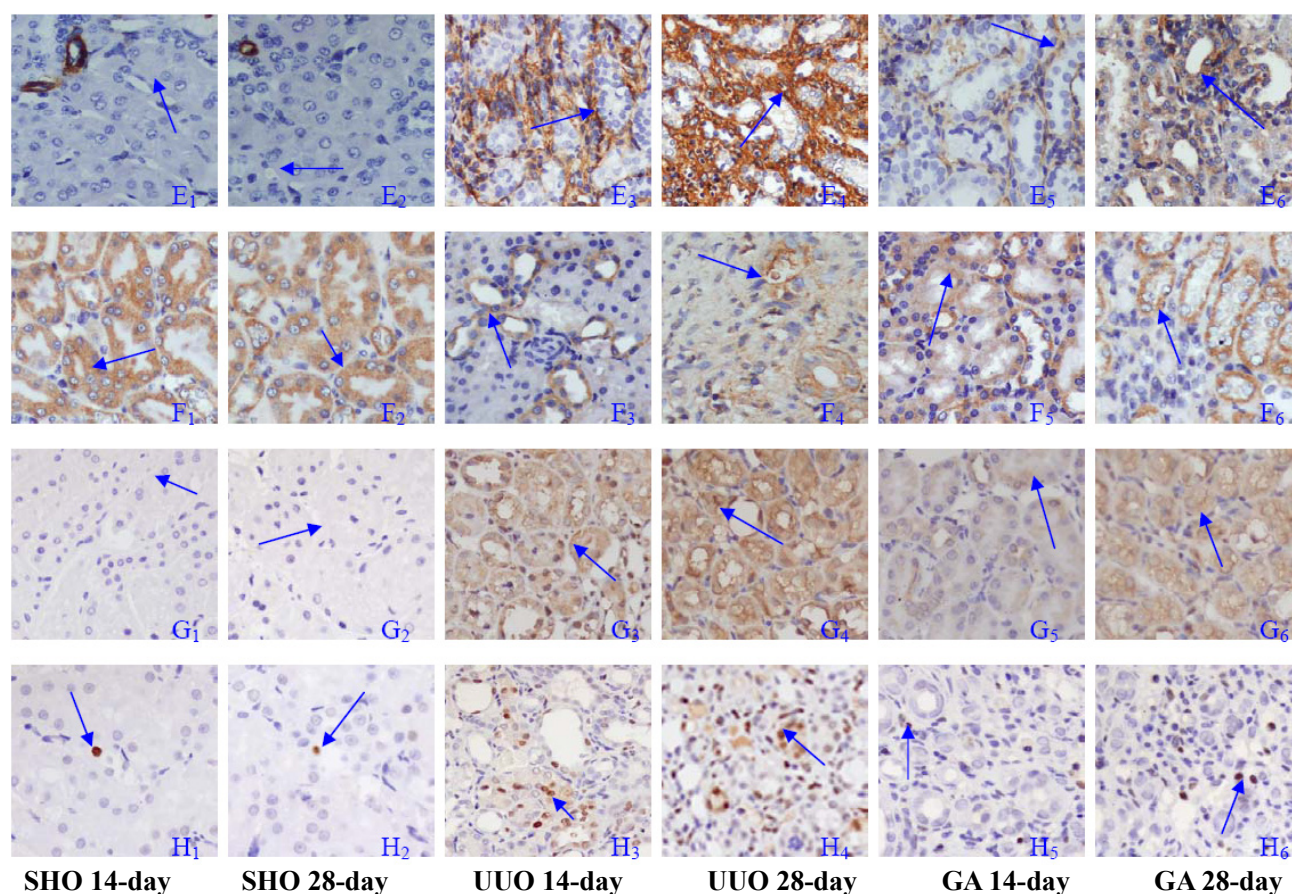


Figure 2. Cont.



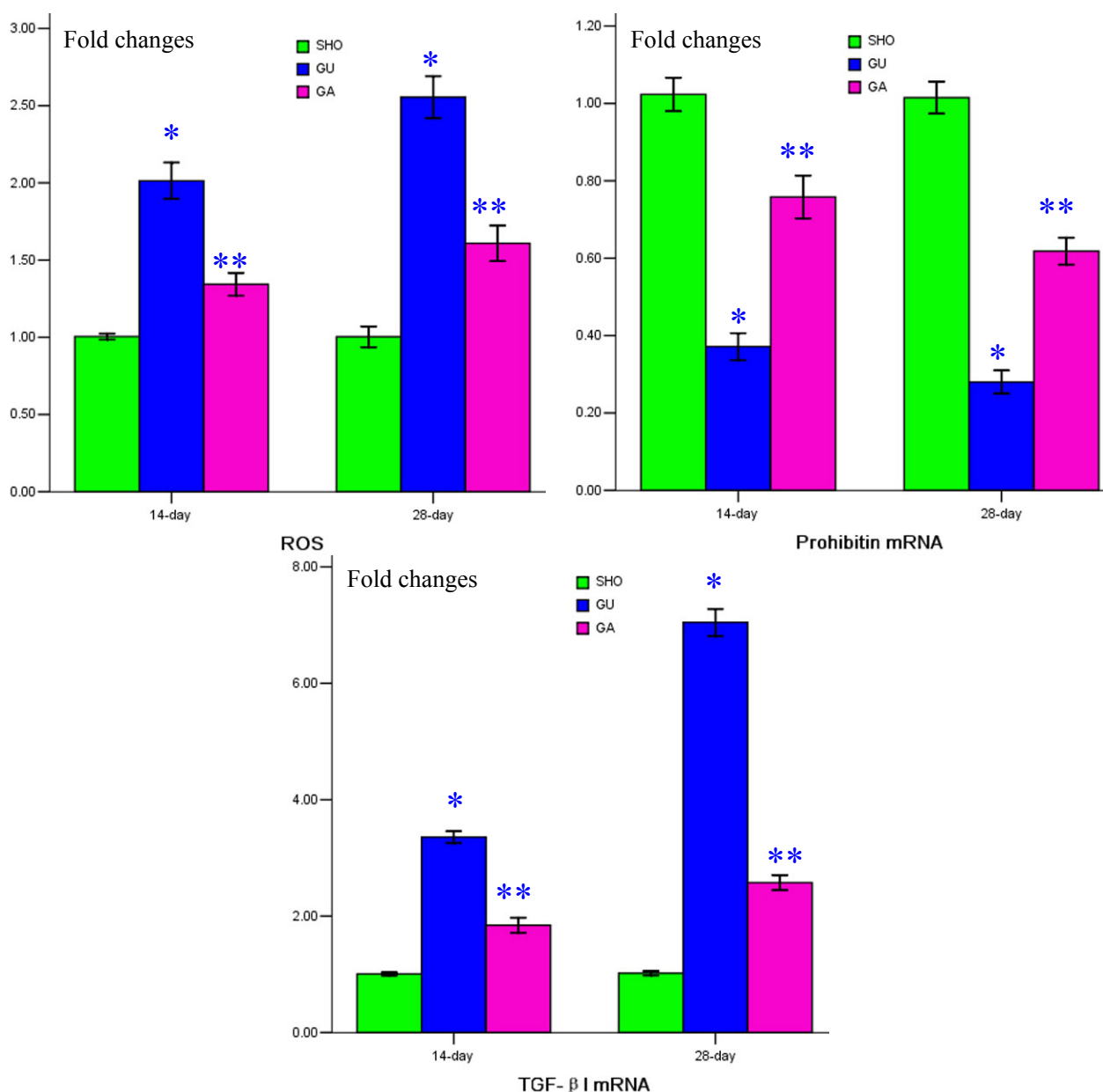
2.3. Association of ATRA Treatment with Cell Apoptosis

The staining of cell apoptosis of the renal tubulointerstitium was much more significant in the UUU group than in the SHO group (blue arrowhead in Figure 2), and the cell apoptosis index in the UUU group was significantly increased when compared with that in SHO ($p < 0.01$; Figure 2). Cell apoptosis in the renal tubulointerstitium in the ATRA treatment group was attenuated (blue arrowhead in Figure 2), and the apoptosis index in the GA group was significantly reduced over that in the UUU group ($p < 0.01$; Figure 2). Protein expression of prohibitin was negatively correlated with the cell apoptosis index ($r = -0.886, p < 0.01$).

2.4. Association of ATRA Treatment with ROS Generation, mRNA Expressions of Prohibitin and TGF- β 1

When compared with that in the SHO group, the generation of ROS in the UUU group was markedly increased, especially in 28-days ($p < 0.01$; Figure 3). ROS generation in the GA group was alleviated when compared with that of the UUU group ($p < 0.01$; Figure 3). Protein expression of prohibitin was negatively correlated with ROS generation ($r = -0.859, p < 0.01$). Consistently lower prohibitin mRNA expression and higher TGF- β 1 mRNA expression were shown in the renal tissue of the UUU group when compared with that in the SHO group (all $p < 0.01$; Figure 3). Prohibitin mRNA expression was significantly up-regulated and TGF- β 1 mRNA expression was markedly reduced in the GA group when compared with the UUU group (all $p < 0.01$; Figure 3).

Figure 3. Statistical parameters for reactive oxidative species (ROS), prohibitin mRNA, and TGF- β 1 mRNA. *: $p < 0.01$ compared with SHO, **: $p < 0.01$ compared with UUO.



3. Discussion

Of all the cytokines and growth factors involved, TGF- β 1 plays a pivotal role in the processes of renal interstitial fibrosis (RIF) [18]. TGF- β 1 is known as one of the major mediators that leads to RIF by inducing the production of α -SMA and ECM (Col-IV and fibronectin) in the renal tubulointerstitium [19–21]. Those indicators were chosen and evaluated in this investigation.

Prohibitin is an ubiquitous protein and might play a pivotal role in the process of cell apoptosis, and ATRA treatment can have an antioxidant effect and regulate the cell apoptosis. In this investigation, the ROS generation, the cleaved Caspase-3 and the cell apoptosis index were also evaluated.

In our study, we found that the expressions of TGF- β 1, α -SMA, Col-IV and fibronectin in the UUO group were markedly increased when compared to those in the SHO group, especially in 28-days.

However, the expression of prohibitin in the UUO group was markedly reduced over that in SHO. Protein expression of prohibitin was negatively correlated with the RIF index, protein expression of TGF- β 1, α -SMA, Col-IV, fibronectin or cleaved Caspase-3, ROS generation or cell apoptosis index. The number of apoptotic cells in the UUO group was much more than in the normal control group. The lower expression of prohibitin was associated with increased cell apoptosis and RIF lesion. Interestingly, the prohibitin mainly located in renal tubular epithelial cells (RTEC) and apoptotic cells, observed by us, was mainly contributed by RTEC in our study. We speculated that the mechanism might be as follows: The expression of prohibitin in RTEC in the UUO model was reduced. The generation of TGF- β 1 and ROS was increased, which induced the expressions of α -SMA, Col-IV and fibronectin in the renal tubulointerstitium and increased the production of cleaved Caspase-3 in RTEC. The cell apoptosis index was also up-regulated. The less expression of prohibitin was associated with increased expressions of ROS, TGF- β 1, α -SMA, Col-IV, fibronectin, cleaved Caspase-3 and the index of cell apoptosis.

Prohibitin is a highly conserved, ubiquitously expressed protein that participates in diverse processes including mitochondrial chaperone, growth and apoptosis [22] and is regarded as an apoptosis-regulating protein [23]. Prohibitin has played a protective role in some studies. Theiss *et al.* [24] reported that the elevation of prohibitin in the surface epithelial cells of the colon could reduce the severity of colitis in mice, suggesting that prohibitin might represent a novel therapeutic target in inflammatory bowel disease. Ko *et al.* [22] found that hepatocyte-specific prohibitin deficiency resulted in marked liver injury, oxidative stress, and fibrosis with development of hepatocellular carcinoma, suggesting that the prohibitin was a tumor suppressor in hepatocytes. Muraguchi *et al.* [25] performed an investigation into H9C2 cardiomyocytes, and found that prohibitin might function as a survival factor against hypoxia-induced cell death. Liu *et al.* [10] conducted a study in cardiomyocytes, and their data indicated that prohibitin could protect the cardiomyocytes from oxidative stress-induced damage. The results from these studies mentioned above drew a consistent conclusion that prohibitin was associated with injury of cells or tissues. The results were similar to ours. So, how to regulate the abnormal expression of prohibitin is very important; prohibitin might be a potential therapeutic target for regulation of cell apoptosis and prevention of cell or tissue injury. However, cell culture using RTEC, *etc.*, and the inhibition of the signalling pathway of prohibitin need to be conducted to explore further its detailed mechanism in the progression of RIF.

In this investigation, we found that ATRA treatment was associated with increased expression of prohibitin, attenuation of the lesion of the renal tubulointerstitium and reduction in the expressions of TGF- β 1, α -SMA, Col-IV, fibronectin, ROS and cleaved Caspase-3. Kishimoto *et al.* [26] also performed an investigation on the association of ATRA treatment with RIF progression in UUO model mice, and found that a significant improvement was observed in histological and immunological findings in the ATRA treatment group. Their results were similar to ours.

There was no report investigating whether ATRA treatment was associated with the expression of prohibitin *in vivo* or *in vitro*. Li *et al.* [9] showed that prohibitin was localized in the nuclear matrix in human neuroblastoma SK-N-SH cells and its distribution was altered due to retinoic acid treatment, but they did not explore whether the retinoic acid treatment was associated with prohibitin expression.

4. Materials and Methods

4.1. Animal Model

Wistar male rats (6-week-old) were purchased from the Experimental Animal center of Guangxi Medicinal University, Nanning, China. The rats were divided into 3 groups: sham operation group (SHO), model group subjected to unilateral ureteral obstruction (UUO), model group treated with ATRA (GA); $n = 40$, respectively. The ureter was ligated at approximately 1 cm below the renal hilum with a 3-0 silk suture. The abdominal wound was closed, and the rats were returned to the cages. Control rats underwent abdominal incision and approximation with no ligation of the ureter [27,28]. The rats in the GA group were treated with ATRA (Sigma Co., USA; 15 mg/kg·d) in corn oil once daily by oral gavage, from the first day before operation. Rats in the UUO and SHO groups were administered with saline. Twenty rats of the three groups were sacrificed 14 and 28 days after surgery respectively, and the renal tissue was collected for the determination of histological and molecular biology.

The Animal Care and Use Committee of Guangxi Medical University approved all protocols.

4.2. Renal Morphology

After 10% neutral formaldehyde fixation, the renal tissue was dehydrated through a graded ethanol series and embedded in paraffin. Sections were prepared on a microtome and stained with Masson's trichrome staining. Renal morphology was observed by light microscopy, the severity of the renal lesion was presented by the RIF index. Blue granular and linear deposits were interpreted as positive areas for collagen staining. Semi-quantitative evaluation was performed by computer-assisted image analysis (DMR + Q550, Leica Co., Germany). 400-fold original magnifications in ten fields (ignoring the fields containing glomerular parts) were measured, selected from coded sections for each rat at random. The extent of interstitial fibrosis was scored as: absent (0), involving less than 25% of the area (1), involving 26 to 50% of the area (2), and involving greater than 50% of the area (3) [29,30]. RIF index was obtained by the formula as follow: $\text{RIF index} = (0 \times n_0 + 1 \times n_1 + 2 \times n_2 + 3 \times n_3) / (n_0 + n_1 + n_2 + n_3) = (0 \times n_0 + 1 \times n_1 + 2 \times n_2 + 3 \times n_3) / 10$; where n_0 , n_1 , n_2 and n_3 were the number of extent of interstitial fibrosis as absent, less than 25% of the area, 26 to 50% of the area, and greater than 50% of the area, respectively. All the fields were selected from coded sections for each rat at random and the scores obtained by two investigators were averaged.

4.3. Immunohistochemical Analysis of the Protein Expressions of Prohibitin, Transforming Growth Factor- β 1 (TGF- β 1), Collagen-IV (Col-IV), Fibronectin, α -Smooth Muscle Actin (α -SMA) and Cleaved Caspase-3

Kidney tissue fixed with 4% buffered paraformaldehyde was embedded in paraffin, and 4 μ m thick sections were stained. The positive area was measured using a computer-aided manipulator (DMR + Q550, Leica Co., Germany). For immunohistochemical analysis of prohibitin, TGF- β 1, Col-IV, fibronectin, α -SMA and cleaved Caspase-3, the sections were deparaffinized and washed with PBS. In order to retrieve the antigenicity from formalin fixation, the sections were incubated for 10 min in 10 mmol/L sodium citrate buffer using a microwave oven. After the incubation, all the sections were treated with 3%

H₂O₂ in methanol for 10 min. All sections were then incubated with anti-prohibitin antibody (1:300) (Neomarker Lab, Co., USA), anti-TGF- β 1 antibody (1:100) (Zhongshan, Co., Beijing, China), anti-Col-IV antibody (ready-to-use kit; Bo Shide, Co., Wuhan, China), anti-fibronectin antibody (1:50) (Zhongshan, Co., Beijing, China), anti- α -SMA antibody (ready-to-use kit; Shanghai Changdao, Co., Inc., China) and anti-cleaved-Caspase-3 antibody (1:200) (Thermo Fisher Scientific, Co., Runcorn, UK), respectively. After incubation with second antibody immunoglobulin (Shanghai Changdao, Co., Shanghai, China), the sections were stained with diaminobenzidine (Maixin Bio, Co., Fuzhou, China). Prohibitin, TGF- β 1, Col-IV, fibronectin, α -SMA or cleaved Caspase-3 in renal tissue was measured by semi-quantitative evaluation. During evaluation of the interstitial areas, fields containing glomerular parts were ignored. All the evaluations were performed by two of the authors blind to the experimental code.

4.4. Apoptosis Assay

Cell apoptosis was examined by the TdT mediated dUTP nick end labelling (TUNEL) assay (Roche Inc., Basel, Switzerland) as described previously [31,32]. Six slides from each kidney were evaluated for percentage of apoptotic cells by using the TUNEL assay. Then 10 watch fields, which did not include the glomerular parts, were chosen at random under the microscope for each section. Brown staining of cell nuclei was on considered apoptotic cells. Positive brown cells and total cells were counted. The formula for the apoptosis index as the indicator of apoptosis was as follows [33,34]: cell apoptosis index = positive cells/total cells \times 100%. The scores obtained by two investigators were averaged.

4.5. ROS Measurement

The cortical material of freshly dissected renal tissues was extracted, washed, weighed and homogenized in ice cold 0.9% normal saline solution. The homogenate was centrifuged at 4000 \times g for 15 min at 25 °C and the supernatant was used for the determination [35]. The 500 μ L of homogenates were used for detection of the ROS generation by the ROS sensitive dye, 2,7-dichlorodihydro-fluorescein diacetate (DCF-DA; Invitrogen, Co., USA), as an indicator [36,37]. Sample (50 μ g protein) was incubated with 10 μ L of DCF-DA (10 μ M) for 3 h at 37 °C. The fluorescent product formed was measured by spectrofluorometer at 485/525 nm. Changes in fluorescence were expressed as an arbitrary unit [36].

4.6. Real Time Reverse Transcription Polymerase Chain Reaction to Detect Prohibitin/TGF- β 1 mRNA Expression in Renal Tissue

Renal tissue was homogenized, and total RNA was extracted with TRIzol (Beijing Tiangen, Co., China). Primers were designed according to primer design principles by Primer Premier 5.0. The primers for prohibitin, TGF- β 1 and internal control β -actin were as follows: F 5'-TGGCGTTAGCGGTTACAGGAG-3' and R 5'-GAGGATGCGTAGTGTGATGTTGAC-3' for prohibitin; F 5'-TGAGCACTGAAGCGAAAGCC-3' and R 5'-GATTCAAGTCAACTGTGGAGCAAC-3' for TGF- β 1; F 5'-GCCCCTGAGGAGCACCTGT-3' and R 5'-ACGCTCGGTCAGGATCTTCA-3' for β -actin. One microgram total RNA from the renal tissue of each rat was reverse transcribed into cDNA with an ExScript RT reagent kit (Takara Biotechnology, Co., Dalian, China). Prohibitin, TGF- β 1 and β -actin were amplified with SYBR Premix Ex Taq (Beijing Tiangen, Co., China). Gene expression of β -actin was also

measured in each sample, and was used as an internal control for loading and reverse transcription efficiency. The analysis for each sample was performed in triplicate. The average threshold cycle (Ct, the cycles of template amplification to the threshold) was worked out as the value of each sample. The data of fold of changes was analyzed using $2^{-\Delta\Delta Ct}$ [38,39]. For example, the $\Delta\Delta Ct$ of prohibitin mRNA expression in UUO group in 14-day was as follow: $\Delta\Delta Ct_{\text{prohibitin, 14-day, UUO group}} = (Ct_{\text{prohibitin, 14-day, UUO group}} - Ct_{\beta\text{-actin, 14-day, UUO group}}) - (Ct_{\text{prohibitin, 14-day, SHO group}} - Ct_{\beta\text{-actin, 14-day, SHO group}})$, and the fold of changes of prohibitin mRNA expression in the UUO group in 14-days was $2^{-\Delta\Delta Ct_{\text{prohibitin, 14-day, UUO group}}}$.

4.7. Statistical Analysis

The data are shown as mean \pm standard deviation. Analysis of covariance and Student-Newman Keuls post-tests was performed to determine the differences among groups, and Pearson's correlation coefficient was used to determine the relationships between the indicators. A value of $p < 0.05$ was considered as a significant difference. Statistical analysis was performed using the statistical package for social studies SPSS version 13.0 (SPSS, Chicago, IL, USA).

5. Conclusion

In conclusion, lower expression of prohibitin is associated with RIF progression, and ATRA treatment is associated with increased prohibitin in UUO rats, although the detailed mechanism is not fully understood. However, cell culture and further investigations should be conducted to explore the detailed mechanism.

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Conflict of Interest

The authors have declared that no competing interests exist.

References

1. Bielez, B.; Sirin, Y.; Si, H.; Niranjana, T.; Gruenwald, A.; Ahn, S.; Kato, H.; Pullman, J.; Gessler, M.; Haase, V.H.; *et al.* Epithelial Notch signaling regulates interstitial fibrosis development in the kidneys of mice and humans. *J. Clin. Invest.* **2010**, *120*, 4040–4054.

2. Kushiyama, T.; Oda, T.; Yamada, M.; Higashi, K.; Yamamoto, K.; Sakurai, Y.; Miura, S.; Kumagai, H. Alteration in the phenotype of macrophages in the repair of renal interstitial fibrosis in mice. *Nephrology (Carlton)* **2011**, *16*, 522–535.
3. Morisada, N.; Nomura, M.; Nishii, H.; Furuno, Y.; Sakanashi, M.; Sabanai, K.; Toyohira, Y.; Ueno, S.; Watanabe, S.; Tamura, M.; *et al.* Complete disruption of all nitric oxide synthase genes causes markedly accelerated renal lesion formation following unilateral ureteral obstruction in mice *in vivo*. *J. Pharmacol. Sci.* **2010**, *114*, 379–389.
4. Correa-Costa, M.; Semedo, P.; Monteiro, A.P.; Silva, R.C.; Pereira, R.L.; Goncalves, G.M.; Marques, G.D.; Cenedeze, M.A.; Faleiros, A.C.; Keller, A.C.; *et al.* Induction of heme oxygenase-1 can halt and even reverse renal tubule-interstitial fibrosis. *PLoS One* **2010**, *5*, e14298.
5. Manucha, W. Biochemical-molecular markers in unilateral ureteral obstruction. *Biocell* **2007**, *31*, 1–12.
6. Manucha, W.; Carrizo, L.; Ruete, C.; Molina, H.; Valles, P. Angiotensin II type I antagonist on oxidative stress and heat shock protein 70 (HSP 70) expression in obstructive nephropathy. *Cell Mol. Biol. (Noisy-le-Grand)* **2005**, *51*, 547–555.
7. Puppin, C.; Passon, N.; Franzoni, A.; Russo, D.; Damante, G. Histone deacetylase inhibitors control the transcription and alternative splicing of prohibitin in thyroid tumor cells. *Oncol. Rep.* **2011**, *25*, 393–397.
8. Zhu, B.; Zhai, J.; Zhu, H.; Kyprianou, N. Prohibitin regulates TGF-beta induced apoptosis as a downstream effector of Smad-dependent and -independent signaling. *Prostate* **2010**, *70*, 17–26.
9. Li, Q.F.; Liang, Y.; Shi, S.L.; Liu, Q.R.; Xu, D.H.; Jing, G.J.; Wang, S.Y.; Kong, H.Y. Localization of prohibitin in the nuclear matrix and alteration of its expression during differentiation of human neuroblastoma SK-N-SH cells induced by retinoic acid. *Cell. Mol. Neurobiol.* **2011**, *31*, 203–211.
10. Liu, X.; Ren, Z.; Zhan, R.; Wang, X.; Wang, X.; Zhang, Z.; Leng, X.; Yang, Z.; Qian, L. Prohibitin protects against oxidative stress-induced cell injury in cultured neonatal cardiomyocyte. *Cell Stress Chaperones* **2009**, *14*, 311–319.
11. Zhou, T.B.; Qin, Y.H.; Ou, C.; Lei, F.Y.; Su, L.N.; Huang, W.F.; Zhao, Y.J. All-trans retinoic acid can regulate the expressions of gelatinases and apolipoprotein E in glomerulosclerosis rats. *Vascul. Pharmacol.* **2011**, *55*, 169–177.
12. de Rosa, S.; Cirillo, P.; Paglia, A.; Sasso, L.; di Palma, V.; Chiariello, M. Reactive oxygen species and antioxidants in the pathophysiology of cardiovascular disease: Does the actual knowledge justify a clinical approach? *Curr. Vasc. Pharmacol.* **2010**, *8*, 259–275.
13. Lee, H.P.; Casadesus, G.; Zhu, X.; Lee, H.G.; Perry, G.; Smith, M.A.; Gustaw-Rothenberg, K.; Lerner, A. All-trans retinoic acid as a novel therapeutic strategy for Alzheimer's disease. *Expert Rev. Neurother.* **2009**, *9*, 1615–1621.
14. Liu, X.; Lu, L.; Tao, B.B.; Zhou, A.L.; Zhu, Y.C. Amelioration of glomerulosclerosis with all-trans retinoic acid is linked to decreased plasminogen activator inhibitor-1 and alpha-smooth muscle actin. *Acta Pharmacol. Sin.* **2011**, *32*, 70–78.
15. Ratnam, K.K.; Feng, X.; Chuang, P.Y.; Verma, V.; Lu, T.C.; Wang, J.; Jin, Y.; Farias, E.F.; Napoli, J.L.; Chen, N.; *et al.* Role of the retinoic acid receptor-alpha in HIV-associated nephropathy. *Kidney Int.* **2011**, *79*, 624–634.

16. Zhou, T.B.; Qin, Y.H.; Lei, F.Y.; Su, L.N.; Zhao, Y.J.; Huang, W.F. All-trans retinoic acid regulates the expression of apolipoprotein E in rats with glomerulosclerosis induced by Adriamycin. *Exp. Mol. Pathol.* **2011**, *90*, 287–294.
17. Rao, J.; Zhang, C.; Wang, P.; Lu, L.; Zhang, F. All-trans retinoic acid alleviates hepatic ischemia/reperfusion injury by enhancing manganese superoxide dismutase in rats. *Biol. Pharm. Bull.* **2010**, *33*, 869–875.
18. Chen, C.L.; Chou, K.J.; Lee, P.T.; Chen, Y.S.; Chang, T.Y.; Hsu, C.Y.; Huang, W.C.; Chung, H.M.; Fang, H.C. Erythropoietin suppresses epithelial to mesenchymal transition and intercepts Smad signal transduction through a MEK-dependent mechanism in pig kidney (LLC-PK1) cell lines. *Exp. Cell Res.* **2010**, *316*, 1109–1118.
19. Djamali, A.; Vidyasagar, A.; Yagci, G.; Huang, L.J.; Reese, S. Mycophenolic acid may delay allograft fibrosis by inhibiting transforming growth factor-beta1-induced activation of Nox-2 through the nuclear factor- κ B pathway. *Transplantation* **2010**, *90*, 387–393.
20. Zhou, G.; Li, C.; Cai, L. Advanced glycation end-products induce connective tissue growth factor-mediated renal fibrosis predominantly through transforming growth factor beta-independent pathway. *Am. J. Pathol.* **2004**, *165*, 2033–2043.
21. Kelley, R.; Werdin, E.S.; Bruce, A.T.; Choudhury, S.; Wallace, S.M.; Ilagan, R.M.; Cox, B.R.; Tatsumi-Ficht, P.; Rivera, E.A.; Spencer, T.; *et al.* Tubular cell-enriched subpopulation of primary renal cells improves survival and augments kidney function in rodent model of chronic kidney disease. *Am. J. Physiol. Renal. Physiol.* **2010**, *299*, 1026–1039.
22. Ko, K.S.; Tomasi, M.L.; Iglesias-Ara, A.; French, B.A.; French, S.W.; Ramani, K.; Lozano, J.J.; Oh, P.; He, L.; Stiles, B.L.; *et al.* Liver-specific deletion of prohibitin 1 results in spontaneous liver injury, fibrosis, and hepatocellular carcinoma in mice. *Hepatology* **2010**, *52*, 2096–2108.
23. Sato, S.; Murata, A.; Orihara, T.; Shirakawa, T.; Suenaga, K.; Kigoshi, H.; Uesugi, M. Marine natural product aurilide activates the OPA1-mediated apoptosis by binding to prohibitin. *Chem. Biol.* **2011**, *18*, 131–139.
24. Theiss, A.L.; Laroui, H.; Obertone, T.S.; Chowdhury, I.; Thompson, W.E.; Merlin, D.; Sitaraman, S.V. Nanoparticle-based therapeutic delivery of prohibitin to the colonic epithelial cells ameliorates acute murine colitis. *Inflamm. Bowel Dis.* **2011**, *17*, 1163–1176.
25. Muraguchi, T.; Kawawa, A.; Kubota, S. Prohibitin protects against hypoxia-induced H9c2 cardiomyocyte cell death. *Biomed. Res.* **2010**, *31*, 113–122.
26. Kishimoto, K.; Kinoshita, K.; Hino, S.; Yano, T.; Nagare, Y.; Shimazu, H.; Nozaki, Y.; Sugiyama, M.; Ikoma, S.; Funauchi, M. Therapeutic effect of retinoic acid on unilateral ureteral obstruction model. *Nephron Exp. Nephrol.* **2011**, *118*, 69–78.
27. Xie, P.; Sun, L.; Nayak, B.; Haruna, Y.; Liu, F.Y.; Kashihara, N.; Kanwar, Y.S. C/EBP- β modulates transcription of tubulointerstitial nephritis antigen in obstructive uropathy. *J. Am. Soc. Nephrol.* **2009**, *20*, 807–819.
28. Zhang, D.; Sun, L.; Xian, W.; Liu, F.; Ling, G.; Xiao, L.; Liu, Y.; Peng, Y.; Haruna, Y.; Kanwar, Y.S. Low-dose paclitaxel ameliorates renal fibrosis in rat UUO model by inhibition of TGF- β /Smad activity. *Lab. Invest.* **2010**, *90*, 436–447.

29. Radford, M.G., Jr.; Donadio, J.V., Jr.; Bergstralh, E.J.; Grande, J.P. Predicting renal outcome in IgA nephropathy. *J. Am. Soc. Nephrol.* **1997**, *8*, 199–207.
30. Zhou, T.B.; Qin, Y.H.; Zhou, C.; Lei, F.Y.; Zhao, Y.J.; Chen, J.; Su, L.N.; Huang, W.F. Less expression of prohibitin is associated with increased Caspase-3 expression and cell apoptosis in renal interstitial fibrosis rats. *Nephrology (Carlton)* **2012**, *17*, 189–196.
31. Hu, X.; Zhou, X.; He, B.; Xu, C.; Wu, L.; Cui, B.; Wen, H.; Lu, Z.; Jiang, H. Minocycline protects against myocardial ischemia and reperfusion injury by inhibiting high mobility group box 1 protein in rats. *Eur. J. Pharmacol.* **2010**, *638*, 84–89.
32. Ha, T.; Hua, F.; Liu, X.; Ma, J.; McMullen, J.R.; Shioi, T.; Izumo, S.; Kelley, J.; Gao, X.; Browder, W.; *et al.* Lipopolysaccharide-induced myocardial protection against ischaemia/reperfusion injury is mediated through a PI3K/Akt-dependent mechanism. *Cardiovasc. Res.* **2008**, *78*, 546–553.
33. Li, W.; Wang, J.; Jiang, H.R.; Xu, X.L.; Zhang, J.; Liu, M.L.; Zhai, L.Y. Combined effects of cyclooxygenase-1 and cyclooxygenase-2 selective inhibitors on ovarian carcinoma *in vivo*. *Int. J. Mol. Sci.* **2011**, *12*, 668–681.
34. Lai, J.Y.; Li, Y.T.; Wang, T.P. *In vitro* response of retinal pigment epithelial cells exposed to chitosan materials prepared with different cross-linkers. *Int. J. Mol. Sci.* **2010**, *11*, 5256–5272.
35. Olalekan, L.A.; Lawal, A.F.; Ologundudu, A.; Adeniran, O.Y.; Omonkhua, A.; Obi, F. Antioxidant effects of heated garlic juice on cadmium-induced liver damage in rats as compared to ascorbic acid. *J. Toxicol. Sci.* **2011**, *36*, 549–557.
36. Zhang, Y.; Peng, T.; Zhu, H.; Zheng, X.; Zhang, X.; Jiang, N.; Cheng, X.; Lai, X.; Shunnar, A.; Singh, M.; *et al.* Prevention of hyperglycemia-induced myocardial apoptosis by gene silencing of Toll-like receptor-4. *J. Transl. Med.* **2010**, *8*, doi:10.1186/1479-5876-8-133.
37. Chen, N.; Ko, M. Schisandrin B-induced glutathione antioxidant response and cardioprotection are mediated by reactive oxidant species production in rat hearts. *Biol. Pharm. Bull.* **2010**, *33*, 825–829.
38. Zhou, T.B.; Qin, Y.H.; Lei, F.Y.; Su, L.N.; Zhao, Y.J.; Huang, W.F. Less gelatinases is associated with apolipoprotein E accumulation in glomerulosclerosis rats. *Histol. Histopathol.* **2012**, *27*, 249–256.
39. Zhou, T.B.; Qin, Y.H.; Lei, F.Y.; Zhao, Y.J.; Huang, W.F. Association of PAX2 with cell apoptosis in unilateral ureteral obstruction rats. *Ren. Fail.* **2012**, *34*, 194–202.