



Article Reaction of Pyrrolobenzothiazines with Schiff Bases and Carbodiimides: Approach to Angular 6/5/5/5-Tetracyclic Spiroheterocycles

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Abstract: 1*H*-Pyrrole-2,3-diones, fused at [e]-side with a heterocycle, are suitable platforms for the synthesis of various angular polycyclic alkaloid-like spiroheterocycles. Recently discovered sulfur-containing [e]-fused 1*H*-pyrrole-2,3-diones (aroylpyrrolobenzothiazinetriones) tend to exhibit unusual reactivity. Based on these peculiar representatives of [e]-fused 1*H*-pyrrole-2,3-diones, we have developed an approach to an unprecedented 6/5/5/5-tetracyclic alkaloid-like spiroheterocyclic system of benzo[d]pyrrolo[3',4':2,3]pyrrolo[2,1-b]thiazole via their reaction with Schiff bases and carbodiimides. The experimental results have been supplemented with DFT computational studies. The synthesized alkaloid-like 6/5/5/5-tetracyclic compounds have been tested for their biotechnological potential as growth stimulants in the green algae *Chlorella vulgaris*.

Keywords: carbodiimide; *Chlorella*; DFT calculations; nitrogen heterocycle; 1*H*-pyrrole-2,3-dione; Schiff base; sulfur heterocycle

1. Introduction

To improve the clinical success, reduce the undesirable side effects caused by the binding promiscuity of drug candidates, and speed up the lead optimization process, it is necessary to look for ways to expand the medicinal chemistry synthetic toolbox to be able to target more complex three-dimensional (3D) chemical space [1–4]. The 3D shape of a molecule is the most important factor determining its biological activity [5–7]. Due to these, angular polycyclic alkaloid-like spirocycles are attractive objects for drug discovery and related studies [8].

[*e*]-Fused 1*H*-pyrrole-2,3-diones (FPDs) (Figure 1) are versatile starting materials for the synthesis of various heterocyclic systems [9–12], including angular polycyclic alkaloid-like spiroheterocycles (for example, 6/6/5/5- [13], 6/6/5/6- [14], 6/6/5/6/6- [15], 6/6/5/6/5- [15], 6/5/7/5- [16], 6/5/7/6- [17], 6/6/5/7/6- [18], 6/7/5/6- [19], 5/6/5/6- systems [20] and some others (Figure 1)).

Exploring the scope of the recently discovered by us nucleophile-induced ring contraction reaction in FPDs [21], we unexpectedly found an approach to an unprecedented angular 6/5/5/5-tetracyclic alkaloid-like spiroheterocyclic system of benzo[*d*]pyrrolo[3',4':2,3]



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). pyrrolo[2,1-*b*]thiazole (Figure 2). Such a 6/5/5/5-tetracyclic framework is a quite interesting one, since it is present in natural products (retigeranic acid, a sesterterpene from Himalayan lichens *Lobaria retigera* and *Lobaria subretigeria* [22]) and synthetic biologically active molecules [23,24] (Figure 2).



Figure 1. Selected examples of angular polycyclic alkaloid-like spiroheterocyclic systems, synthesized on the basis of FPDs.



Figure 2. 6/5/5/5-Tetracyclic alkaloid-like spiroheterocyclic system reported in this study and related compounds.

Thus, herein, we report the first synthetic approach to an unprecedented 6/5/5/5-tetracyclic alkaloid-like spiroheterocyclic system of benzo[*d*]pyrrolo[3',4':2,3]pyrrolo[2,1-*b*]thiazole (Figure 2) via the reaction of pyrrolobenzothiazines (FPDs incorporating 1,4-benzothiazine moiety) with Schiff bases and carbodiimides. The experimental results were supplemented with DFT computational studies to elucidate the mechanism and stereoselectivity of the reaction. The biotechnological potential of the reported benzo[*d*]pyrrolo[3',4':2,3] pyrrolo[2,1-*b*]thiazoles as growth stimulants and promoters of pigments accumulation in the green algae *Chlorella vulgaris* was demonstrated.

2. Results and Discussion

2.1. Chemistry

Recently, we reported [25] a new class of FPDs, aroylpyrrolobenzothiazinetriones (APBTTs) **1** (Scheme 1). APBTTs **1** were found to react as oxadienes in a hetero-Diels–Alder reaction with electron-rich dienophiles (alkoxyolefins, styrene) (Scheme 1) [25], which is a quite common reactivity for other known types of FPDs and monocyclic 1*H*-pyrrole-2,3-diones [9,10,14,15]. However, under the action of mononucleophiles (amines and alcohols), APBTTs **1** were found to undergo a ring-contraction reaction (Scheme 1) [21], which greatly distinguished their reactivity from the reactivity of their 5-oxa- and 5-aza-analogues [10].



Scheme 1. Reactivity APBTTs 1 in the reactions with electron-rich dienophiles and mononucleophiles.

5-Oxa- and 5-aza-analogues of APBTTs **1** seem not to react with Schiff bases and carbodiimides (C=N reagents) without thermal decomposition (for such reactions under thermal decomposition conditions, see the work presented in [12]). At the same time, monocyclic 1*H*-pyrrole-2,3-diones are known to react with carbodiimides as oxadienes in formal hetero-Diels–Alder reactions to produce the corresponding cycloadducts (Scheme 2) [26].



Scheme 2. Reaction of monocyclic 1H-pyrrole-2,3-diones with carbodiimides.

Considering the tendency for the unusual reactivity of APBTTs **1**, we studied their reaction with Schiff bases and carbodiimides under conditions without thermal decomposition of APBTTs **1** (for thermal decomposition of APBTTs **1**, see the work presented in [27]).

To start with, we tested the reaction of APBTT **1a** with *N*-benzylideneaniline **2a** (Scheme 3). As a result, 6/5/5/5-tetracyclic product **3a** was isolated by a simple crystallization from the reaction mixture in the yield of 52% (Scheme 3). Compound **3a** was obtained as a single ($3R^*$, $3aS^*$, $11aR^*$)-diastereomer, and its structure was unequivocally determined by a single crystal X-ray analysis (CCDC 2341688).



of APBTT **1a** with Schiff base **2a** (Table **1**).

Scheme 3. Reaction of APBTT 1a with N-benzylideneaniline 2a under test conditions.

Obviously, our hypothesis of the unusual reactivity of APBTTs 1 in reactions with C=N reagents was confirmed, which justified a more in-depth study of this transformation. Next, we carried out a series of experiments to optimize the conditions of the reaction

Table 1. Optimization of the reaction of APBTT **1a** with Schiff base **2a** under various conditions ¹.

Entry	Solvent	Time, ² min	Temperature, °C	HPLC Yield ³ of 3a, %
1	acetone	15	56	traces
2	acetonitrile	30	85	25
3	benzene	180	85	80
4	butyl acetate	90	126	41
5	chloroform	300	65	90
6	1,4-dioxane	180	105	66
7	DMAA	5	120	traces
8	DMF	5	120	traces
9	DMSO	5	120	traces
10	NMP	5	120	traces
11	THF	30	70	42
12	toluene	30	115	69
13	o-xylene	30	120	36

¹ Reaction scale: a mixture of APBTT **1a** (29.8 μ mol, 10 mg), Schiff base **2a** (29.8 μ mol, 5.4 mg), and an anhydrous solvent (500 μ L) was stirred in an oven-dried closed microreaction V-vial. ² The reaction progress was monitored visually by the disappearance of the dark violet color characteristic of APBTT **1a**. ³ Biphenyl was used as an internal standard; each entry was carried out in duplicate, and the yields are given as mean values.

According to Table 1, APBTT **1a** and Schiff base **2a** reacted most quickly in polar solvents (acetone, DMAA, DMSO, DMF, NMP (entries 1, 7–10, Table 1)), but the reaction proceeded unselectively (a difficult to identify mixture of products was observed), and only trace amounts of the target product **3a** were formed. Interestingly, in acetonitrile (entry 2, Table 1), which is a polar solvent too, the reaction yield was much higher, which could indicate that the low yields of the product **3a** in polar solvents were caused not only by the polarity of these solvents, but also by their specific solvation effects and their ability to react with the reaction intermediates. In nonpolar solvents (entries 3–6, 12, 13 Table 1), the reaction proceeded much slower and was more selective towards the product **3a**. The best yield of the product **3a** (HPLC yield of 90%) was observed in chloroform when heated for 5 h (entry 5, Table 1).

It should be mentioned that, during the optimization of the reaction of APBTT **1a** with the Schiff base **2a** at room temperature in anhydrous butyl acetate, acetonitrile, and acetone, we observed the formation of product **4a** in significant amounts (Scheme 4). Under these conditions, the reaction proceeded very slowly (about a month), and obviously, side reactions took place. Since the reaction vials were not sealed, it could be assumed that atmospheric moisture affected the reaction (Scheme 4). Moreover, the reaction of APBTT **1a** with the Schiff base **2a** at room temperature in acetic acid, containing traces of water, produced compound **4a** in 24 h in a very good isolated yield (86%) (Scheme 4).



Scheme 4. Reaction of APBTT **1a** with *N*-benzylideneaniline **2a** in the presence of atmospheric moisture at room temperature.

To prove our hypothesis of the formation of compound **4a**, we carried out a reaction of APBTTs **1a**,**b** with aniline **5a** and benzylamine **5b** (Scheme 5). As a result, we isolated target products **4a**,**b** in yields of 95% and 46%, respectively. The structure of compound **4b** was unequivocally confirmed by a single crystal X-ray analysis (CCDC 2341690).



Scheme 5. Reaction of APBTTs 1a,b with amines 5a,b.

Then, we performed reactions of APBTTs **1a–f** with Schiff bases **2a–g** and azine **2h** to determine the reactant scope (Table 2).

As a result, we found that the reaction of APBTTs **1** with Schiff bases **2** performed under optimized conditions (chloroform as the solvent) produced target products **3** in poor to very good HPLC yields (Conditions A, Table 2). However, our attempts to isolate products **3** from such reaction mixtures (in scale of 298 µmol) were unsuccessful. Moreover, we observed that compounds **3** underwent unfavorable transformations during our attempts to isolate and purify them by column chromatography. For these reasons, we replaced chloroform with benzene in these reactions. As a result, we noticed a decreasing tendency in the HPLC yields of products **3** (Conditions B, Table 2), which correlated with the optimization data for the test reaction of APBTT **1a** with Schiff base **2a** (Table 1). However, products **3** were easily isolated and purified by simple crystallization directly from the reaction mixtures (in scale of 298 µmol, benzene).

We also observed that the nature of the aryl substituents in the examined (Table 2) APBTTs **1** and Schiff bases **2** did not significantly affect the yields of the corresponding products **3**. However, the reactions of APBTT **1a** with 1-phenyl-*N*-(pyridin-2-yl)methanimine **2i**, 1-((phenylimino)methyl)naphthalen-2-ol **2j**, *N*-(4-nitrophenyl)-1-phenylmethanimine, *N*-mesityl-1-(4-nitrophenyl)methanimine **2k**, *N*-(2-chlorophenyl)-1-phenylmethanimine **2l**, and *N*,*N*-dimethyl-4-((phenylimino)methyl)aniline **2m** did not produce the desired products **3**, which was possibly caused by the presence of additional nucleophilic centers, *o*-substituents, or strong electron-withdrawing groups in the molecules of these Schiff bases. Moreover, the reaction with *N*-benzylmethanimine **2n** did not give the corresponding

product **3** too, possibly due to the absence of aromatic substituent at the CH part of this Schiff base, which could stabilize reaction intermediates.

Table 2. The reaction of APBTTs 1a–f with Schiff bases 2a–g and azine 2h.

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Enter		- 1	~ 2	HPLC Yie	eld of 3, %	T 1 (1)(113 (0 0)
Entry	Entry Ar	R ¹	R ² -	Conditions A ¹	Conditions B ²	Isolated Yield ^o of 3, %
3a	Ph	Ph	Ph	90	80	52
3b	C ₆ H ₄ Me-4	Ph	Ph	81	48	18
3c	C_6H_4F-4	Ph	Ph	n/d ⁴	29	25
3d	C ₆ H ₄ Br-4	Ph	Ph	n/d	18	17
3e	2-furyl	Ph	Ph	n/d	72	47
3f	2-thienyl	Ph	Ph	n/d	47	45
3g	Ph	$C_6H_4NO_2-3$	trans-CH=CHPh	22	14	12
3h	Ph	Ph	C_6H_4I-4	33	36	28
3i	Ph	C ₆ H ₄ OMe-4	C_6H_4Br-4	53	63	28
3j	Ph	C_6H_4Cl-4	C ₆ H ₃ (OMe) ₂ -3,4	n/d	42	32 ⁵
3k	Ph	Ph	C ₆ H ₃ (OMe) ₂ -3,4	43	44	23 ⁵
31	Ph	Bn	C ₆ H ₄ Br-4	37	34	33
3m	Ph	N=CHPh	Ph	n/d	n/d	26

¹ Conditions A: a mixture of APBTT **1** (29.8 µmol), Schiff base **2** (29.8 µmol), and anhydrous chloroform (500 µL) was stirred in an oven-dried closed microreaction V-vial at 65 °C, until a clear orange or yellow solution formed. ² Conditions B: a mixture of APBTT **1** (29.8 µmol), Schiff base **2** (29.8 µmol) and anhydrous benzene (2.5 mL) was stirred in an oven-dried closed microreaction V-vial at 85 °C, until a clear orange or yellow solution formed. ³ Reaction conditions are given in Section 3.1.2. ⁴ n/d = this was not done. ⁵ Data only for (3*R**,3a*S**,11a*R**)-diastereomer is given.

Noteworthy, when synthesizing product **3d**, we succeeded in isolating by-product **6d** from the reaction mixture in the isolated yield of 2% (Scheme 6). The structure of compound **6d** was unequivocally confirmed by a single crystal X-ray analysis (CCDC 2341691). Apparently, the formation of product **6d** proceeded through an intermediate **A** (conditions for formation of compounds **A** were discussed in the work presented in [21]) and the hydrolysis of Schiff base **2a** (Scheme 6).



Scheme 6. Formation of by-product 6e in the reaction of APBTT 1e with *N*-benzylideneaniline 2a.

Moreover, in cases of compounds 3j and 3k, we succeeded in isolating their stereoisomers, compounds 3'j and 3'k (isolated yields of 7 and 28%, respectively). Indeed, for structures of compounds 3, there are four possible diastereomers with ($3R^*$, $3aS^*$, $11aR^*$), ($3S^*$, $3aS^*$, $11aR^*$), ($3S^*$, $3aR^*$, $11aR^*$), and ($3R^*$, $3aR^*$, $11aR^*$) relative configurations (Figure 3).

In our scope studies (Table 2), we isolated exclusively (3*R**,3a*S**,11a*R**) diastereomers for compounds 3a–i,l,m. In our scope studies (Table 2), we were unable to detect and determine any other diastereomers by HPLC due to the absence of their reference samples; NMR spectra of crude reaction mixtures were also not suitable for these purposes. But, in cases of compounds 3j and 3k, we isolated both (3*R**,3a*S**,11a*R**) diastereomers 3j,k and their stereoisomers 3'j,k with unknown relative configurations. Diastereomers 3'j,k were found to be unstable in HPLC studies, long NMR experiments in solutions and during our attempts to grow a crystal suitable for a single crystal X-ray analysis, which was possibly caused by their reaction with water.



Figure 3. Possible diastereomers of compounds 3.

The ¹H NMR and IR spectra of compounds **3***j*,**k** and compounds **3**'*j*,**k** are very similar and cannot be used for their identification relative to each other. But their ¹³C NMR spectra are quite different in the region of 62–82 ppm (where sp³ atoms—C³, C^{3a}, C^{11a}—appear), which is enough to distinguish them from each other. Fragments of their ¹³C NMR spectra in these regions look as follows:

> ¹³C NMR of **3j** (100 MHz, DMSO-*d*₆): δ = 79.3, **66.0**, 65.2 ppm; ¹³C NMR of **3'j** (100 MHz, DMSO-*d*₆): δ = 79.2, **70.6**, 64.2 ppm; ¹³C NMR of **3k** (100 MHz, DMSO-*d*₆): δ = 79.6, **65.7**, 65.5 ppm; ¹³C NMR of **3'k** (100 MHz, DMSO-*d*₆): δ = 79.3, **70.8**, 64.5 ppm.

However, this information is not enough to determine the relative configuration of compounds 3'.

Then, to elucidate the possible mechanism of the reaction and compare the thermodynamics and kinetics of possible stereoisomers formation, we performed computational DFT studies of a model reaction between the APBTT **1a** and Schiff base **2a** (Scheme 7). We proposed several reaction pathways for **1a** \rightarrow **3a** transformation (Scheme 7), but the results of DFT calculations revealed only one, very energetically unprofitable, intermediate **I2** on the potential energy surface and indicated that the formation of product **3a** occurred directly from the orientation complex **OC**. The hypothetical transformation **OC** \rightarrow **3a** (via transition state **TS** (Figure 4)) is less thermodynamically profitable (by 4.7 kcal/mol in terms of Gibbs-free energies of reaction, Table 3, Figure 5) compared to the alternative hypothetical transformation **OC** \rightarrow **3'a** (via transition state **TS'** (Figure 4)) but more kinetically favorable (by 1.6 kcal/mol in terms of Gibbs-free energies of activation, Table 3, Figure 5). Thus, diastereomer **3a** is a kinetically controlled product, and diastereomer **3'a** is a thermodynamically controlled one (Figure 5).



Scheme 7. Proposed reaction pathways for $1a \rightarrow 3a$ (or 3'a) transformation.



Figure 4. Transition states structures for reactions leading from **OC** to adducts **3a** and **3'a**, according to the DFT calculations (M06-2X/6-31G* level of theory).

Elementary Stage	ΔΕ	ΔΗ	ΔG
1a + 2a ightarrow OC	-7.2	-5.6	6.7
$\mathbf{OC} ightarrow \mathbf{I2}$	12.1	12.4	16.4
$\mathbf{OC} ightarrow \mathbf{TS}$	4.3	4.2	9.8
OC ightarrow TS'	5.3	4.7	11.4
$\mathbf{OC} ightarrow \mathbf{3a}$	-32.1	-30.5	-24.7
OC ightarrow 3'a	-37.1	-35.4	-29.4

Table 3. Calculated values of the total electronic energies, enthalpies, and Gibbs-free energies of reaction (ΔE , ΔH , and ΔG in kcal/mol) for elementary stages of **1a** \rightarrow **3a** (or **3'a**) transformation ¹.

^{$\overline{1}$} The DFT calculations were carried out at the M06-2X/6-31G^{*} level of theory.



Figure 5. Energy profile for the elementary stages of different pathways for $1a \rightarrow 3a$ (or 3'a) transformation.

Since our computational studies revealed only intermediate-**I2**-like transition states **TS** and **TS'** (their 3D structures are available in Supplementary Materials) in the reaction, and these transition states could afford only diastereomers $(3R^*, 3aS^*, 11aR^*)$ and $(3S^*, 3aS^*, 11aR^*)$ (Scheme 7), we suggest that compounds **3'j**,**k** had $(3S^*, 3aS^*, 11aR^*)$ a relative configuration (Figure 3).

Then, in order to expand the reagent scope of the reaction, we studied the reactions of APBTTs 1 with carbodiimides 7.

First, we tested the reaction of APBTT **1a** with *N*,*N*'-dicyclohexylcarbodiimide (DCC) **7a** (Scheme 8). As we expected, 6/5/5/5-tetracyclic product **8a** was isolated by a simple crystallization from the reaction mixture in the yield of 44% (Scheme 8). Compound **8a** was obtained as a single diastereomer, and its structure was unequivocally determined by a single crystal X-ray analysis (CCDC 2341689).



Scheme 8. Reaction of APBTT 1a with DCC 7a under test conditions.

Next, we performed the optimization of the reaction conditions (Table 4). As a result, we did not observe any correlation between the polarity of the solvent and the yield of the product **8a**. The best yield of the product **8a** (HPLC yield of 96%) was observed in 1,4-dioxane when heated for 40 min (entry 6, Table 4). Our optimization studies at room temperature, as expected, revealed that reactions took place over 14 days, during which time both APBTT **1a** and DCC **7a** underwent side reactions with water.

Entry	Solvent	Time, ² min	Temperature, $^{\circ}C$	HPLC Yield ³ of 8a, $\%$
1	acetone	60	56	31
2	acetonitrile	30	85	74
3	benzene	60	85	50
4	butyl acetate	10	126	44
5	chloroform	180	65	35
6	1,4-dioxane	40	105	96
7	DMAA	5	120	16
8	DMF	5	120	0
9	NMP	5	120	0
10	THF	180	70	49
11	toluene	20	115	56
12	<i>p</i> -xylene	40	120	36

Table 4. Optimization of the reaction of APBTT 1a with DCC 7a under various conditions ¹.

¹ Reaction scale: a mixture of APBTT **1a** (29.8 μmol, 10 mg), DCC **7a** (31.3 μmol, 6.5 mg) and anhydrous solvent (500 μL) was stirred in an oven-dried closed microreaction V-vial. ² The reaction progress was monitored visually by the disappearance of the dark violet color characteristic of APBTT **1a**. ³ Biphenyl was used as an internal standard, each entry was carried out in duplicate, and the yields are given as mean values.

Then, we performed reactions of APBTTs 1a-g with carbodiimides 7a,b to determine the reactant scope (Table 5). Initially, we tried to perform the scope examination under optimal conditions in 1,4-dioxane (entry 6, Table 4). But under these conditions, our attempts to isolate products 8 from such reaction mixtures (in scale of 298 µmol) were unsuccessful, and we observed that compounds 8 underwent unfavorable transformations during our attempts to isolate and purify them by column chromatography. A similar situation was observed when we tried to apply acetonitrile as the reaction solvent (entry 2, Table 4). Thus, we had to carry out these reactions in toluene (entry 11, Table 4). In toluene, products 8 were easily isolated by simple crystallization from the reaction mixtures.

Table 5. The reaction of APBTTs 1a-g with carbodiimides 7a,b.



Entry	Ar	Alk	HPLC Yield ¹ of 8, %	Isolated Yield ² of 8, %
8a	Ph	Су	56	44
8b	C_6H_4Me-4	Cy	65	15
8c	C_6H_4F-4	Су	68	11
8d	C_6H_4Br-4	Cy	52	31
8e	2-furyl	Су	62	45
8f	2-thienyl	Су	59	59
8g	C ₆ H ₄ Cl-4	Су	58	25
8h	Ph	Pr-i	71	65
8i	C ₆ H ₄ Me-4	Pr-i	75	75
8j	C ₆ H ₄ Cl-4	Pr-i	73	72

¹ Reaction scale: a mixture of APBTT **1** (29.8 μmol), carbodiimide **7** (31.3 μmol), and anhydrous toluene (500 μL) was stirred in an oven-dried closed microreaction V-vial. ² Reaction conditions are given in Section 3.1.3.

As a result, we found that the reaction of APBTTs **1** with carbodiimides **7** performed in toluene produced target products **8** in fair to good HPLC yields (Table 5). We observed that the nature of the aryl substituents in examined (Table 5) APBTTs **1** did not significantly affect the yields of the corresponding products **8**.

2.2. Biology

The green unicellular algae *Chlorella vulgaris* serves as a source of valuable metabolites for the food industry [28–30], agriculture [22–34], cosmetics [35–37], and biodiesel production [38–42]. There is a significant demand for chemical stimulants that promote the growth of this algae, as well as the accumulation of lipids [43–45], proteins [46,47], carbohydrates [48–50], and pigments [51] such as chlorophylls [52,53] and carotenoids [54–58]. We selected several synthesized compounds with favorable characteristics, such as high synthesis yield and improved solubility in polar solvents. These compounds were added in varying concentrations to *C. vulgaris* cultures, which were then cultivated, followed by the measurement of algae growth (cell concentration) and the accumulation of pigments.

The bioactivity study was conducted in two stages. Initially, a screening experiment in small volumes of algal cultures (96-well plates) was performed (Table 6). Two compounds (**3a** and **8j**) that promoted the growth of *C. vulgaris* were further analyzed in a subsequent experiment. Algae were grown in 50-mL Erlenmeyer flasks in the presence of the tested compounds at concentrations ranging from 1×10^{-7} mol/L to 1×10^{-4} mol/L. Glucose served as a positive control due to its ability to enhance algae growth, while the negative control was culture fluid containing 1% DMSO, which was used for the dilution of the tested compounds.

_	Difference ¹ in Algae Cell Concentration between Cultures Containing the Compounds under Study and Control Cultures					
Entry	Concentration of Compounds in Culture Medium					
	$1 imes 10^{-5}$ mol/L	$1 imes 10^{-6}$ mol/L	$1 imes 10^{-7}$ mol/L			
3a	13.7 ²	-2.7	-9.1			
8a	-21.5	-18.7	3.2			
8h	-5.8	-0.5	-5.7			
8i	-7.4	-8.3	-3.3			
8j	-12.7	6.3	15.1 ²			
Glucose $(2 g/L)$	102.6					

Table 6. The difference in algae cell concentration between cultures containing the compounds under study and the control cultures.

¹ Expressed as a percentage of the negative control. ² Bold indicates conditions that result in cell concentrations exceeding the established threshold (mean of control plus three standard deviations).

Both **3a** and **8j** increased the chlorophyll content in cells and/or algae growth at specific concentrations (Tables 7 and 8). The most notable effect was the nearly 30% increase in chlorophyll content in cells in the presence of 1×10^{-4} mol/L **3a**, although this was accompanied by a 17.8% decrease in cell growth. Chlorophylls are utilized as natural colorants in the food, cosmetics, and textile industries [59]. As *C. vulgaris* cells are enriched with chlorophyll (up to 4.5% of dry weight), it can be considered one of the most prominent natural sources of these pigments with substantial commercial potential [60]. Therefore, the synthesized compounds can be utilized to enhance the efficiency of algal chlorophyll production for industrial applications.

Table 7. The impact of adding compound **3a** on the concentration of *Chlorella* cells and the cellular content of pigments.

Parameter	Negative Control ¹	Positive Control ²	$1 imes 10^{-4}$ mol/L	$1 imes 10^{-5}$ mol/L	$1 imes 10^{-6}$ mol/L	$1 imes 10^{-7}$ mol/L
Concentration of cells, 10 ⁶ cell/mL Chlorophyll a, µg/10 ⁷ cells	$\begin{array}{c} 20.73 \pm 1.05 \ ^{3} \\ 1.713 \pm 0.064 \end{array}$	$\begin{array}{c} 75.00 \pm 1.08 \\ 1.873 \pm 0.002 \end{array}$	$\begin{array}{c} 17.01 \pm 0.32 \\ 2.310 \pm 0.094 \end{array}$	$\begin{array}{c} 20.56 \pm 0.91 \\ 1.842 \pm 0.004 \end{array}$	$\begin{array}{c} 23.93 \pm 0.71 \\ 1.546 \pm 0.066 \end{array}$	$\begin{array}{c} 22.86 \pm 0.85 \\ 1.520 \pm 0.150 \end{array}$

Concentration of cells, 106 cell/mL

Chlorophyll a, $\mu g/10^7$ cells

Chlorophyll b, $\mu g/10^7$ cells

Carotenoids, $\mu g/10^7$ cells

 18.93 ± 1.83

 1.807 ± 0.041

 1.729 ± 0.047

0.215 + 0.025

Parameter	Negative Control ¹	Positive Control ²	$1 imes 10^{-4} \text{ mol/L}$	$1 imes 10^{-5}$ mol/L	$1 imes 10^{-6}$ mol/L	$1 imes 10^{-7} \text{ mol/L}$
Chlorophyll b, μg/10 ⁷ cells Carotenoids, μg/10 ⁷ cells	$\begin{array}{c} 1.723 \pm 0.046 \\ 0.203 \pm 0.015 \end{array}$	$\frac{1.994 \pm 0.015}{n/d^{4}}$	$\begin{array}{c} 2.223 \pm 0.089 \\ 0.235 \pm 0.010 \end{array}$	$\begin{array}{c} 1.797 \pm 0.029 \\ 0.210 \pm 0.018 \end{array}$	$\begin{array}{c} 1.512 \pm 0.047 \\ 0.173 \pm 0.016 \end{array}$	$\begin{array}{c} 1.540 \pm 0.130 \\ 0.169 \pm 0.007 \end{array}$
	1 Culture medium flasks \pm standar	m with 1% of DMSO d deviation. $4 n/d =$. ² Culture medium this was not done.	n with 1% of DMSO	and 2 g/L of gluce	ose. ³ Mean of three
	Table 8.The ircontent of pigr	npact of adding co nents.	ompound 8j on t	the concentratior	n of <i>Chlorella</i> cells	and the cellular
Parameter	Negative Control ¹	Positive Control ²	$1 imes 10^{-4}$ mol/L	$1 imes 10^{-5}$ mol/L	$1 imes 10^{-6}$ mol/L	$1 imes 10^{-7}$ mol/L

Ta	b	le	7.	Cont.

 19.10 ± 2.00^{-3}

 1.795 ± 0.041

 1.738 ± 0.060

 0.207 ± 0.021

¹ Culture medium with 1% of DMSO. ² Culture medium with 1% of DMSO and 2 g/L of glucose. ³ Mean of three flasks \pm standard deviation. ⁴ n/d = this was not done.

 1840 ± 267

 1.904 ± 0.106

 1.859 ± 0.116

 0.213 ± 0.029

 17.60 ± 0.36

 2.033 ± 0.053

 2.043 ± 0.096

0.222 + 0.014

 20.50 ± 0.62

 1.631 ± 0.066

 1.609 ± 0.097

 0.187 ± 0.011

3. Materials and Methods

3.1. Synthetic Methods and Analytic Data of Compounds

 70.13 ± 0.50

 1.818 ± 0.082

 1.787 ± 0.077

n/d⁴

3.1.1. General Information

¹H and ¹³C NMR spectra (Supplementary Materials) were acquired on a Bruker Avance III 400 HD spectrometer (Bruker BioSpin AG, Faellanden, Switzerland) (at 400 and 100 MHz, respectively) in CDCl₃ or DMSO- d_6 , using solvent residual signals (in ¹³C NMR, 77.00 for CDCl₃, 39.52 for DMSO-*d*₆; in ¹H NMR, 7.26 for CDCl₃, 2.50 for DMSO-*d*₆) as internal standards. ¹⁹F NMR spectra (Supplementary Materials) were acquired on a Bruker Avance III 400 HD spectrometer (Bruker BioSpin AG, Faellanden, Switzerland) (at 376 MHz) in CDCl₃ or DMSO- d_6 using no internal standard. IR spectra were recorded on a Perkin-Elmer Spectrum Two spectrometer (PerkinElmer Inc., Waltham, MA, USA) from mulls in mineral oil. Melting points were measured on a Mettler Toledo MP70 apparatus (Mettler-Toledo (MTADA), Schwerzenbach, Switzerland). Elemental analyses were carried out on a Vario MICRO Cube analyzer (Elementar Analysensysteme GmbH, Langenselbold, Germany). The reaction conditions were optimized using HPLC-UV on Hitachi Chromaster (Hitachi High-Tech, Tokyo, Japan) [NUCLEODUR C18 Gravity column (particle size 3 μm; eluent acetonitrile-water, flow rate 1.5 mL/min); Hitachi Chromaster 5430 diode array detector (λ 210–750 nm)]. The single crystal X-ray analyses of compounds **3a**, **4b**, **6d**, 8a were performed on an Xcalibur Ruby diffractometer (Agilent Technologies, Wroclaw, Poland). The empirical absorption correction was introduced by a multi-scan method using the SCALE3 ABSPACK algorithm [61]. Using OLEX2 [62], the structures were solved with the SHELXT [63] or SUPERFLIP [64] program and refined by the full-matrix leastsquares minimization in the anisotropic approximation for all non-hydrogen atoms with the SHELXL [65] program. Hydrogen atoms bound to carbon were positioned geometrically and refined using a riding model. The hydrogen atoms of NH and OH groups were refined independently with isotropic displacement parameters. The APBTTs 1a-g were obtained according to reported procedures [21,25]. The compounds 2a-n were obtained according to the reported procedures [66–80]. Benzene, toluene, o-xylene, p-xylene, 1,4-dioxane, and THF for procedures with compounds 1 were distilled over Na before use. Acetone, butyl acetate, and chloroform for procedures with compounds 1 were distilled over P_2O_5 before the use. DMAA, DMF, DMSO, NMP, and acetonitrile for procedures with compounds 1 were dried over molecular sieves 4Å before the use. All procedures with APBTTs 1 were performed in an oven-dried glassware. All other solvents and reagents were purchased from commercial vendors and were used as received. Thin-layer chromatography (TLC) was performed on ALUGRAM Xtra SIL G/UV254 silica gel 60 plates (Macherey-Nagel, Düren, Germany) using EtOAC/toluene, 1:5 v/v, EtOAc, toluene as eluents; spots were visualized with iodine vapor and/or UV light (254, 365 nm) in the light of a TLC viewing cabinet Petrolaser TLC-254/365 Thin Layer Chromatography Dark Room (Petrolaser, St. Petersburg, Russia). In ¹³C NMR spectra of compounds **3a**,**c**–**g**,**i**, **3'j**, **8d**–**f**,**i**,**j**, signals of some aromatic carbons could not be found.

3.1.2. Procedure to Compounds 3a-m

A mixture of APBTT 1 (0.298 mmol) and Schiff base 2 (0.298 mmol) in benzene (5 mL) was heated for 3–24 h at 85 $^{\circ}$ C (until the dark violet color characteristic of APBTT 1 disappeared and a transparent yellow solution formed). Then, the reaction mixture was cooled to room temperature.

(3*R**,3*a*S*,11*aR**)-3*a*-Benzoyl-2,3-diphenyl-3,3*a*-dihydrobenzo[d]pyrrolo[3',4':2,3]pyrrolo[2,1-b] thiazole-1,4,5(2H)-trione (**3a**). The formed precipitate was filtered off. Then, the precipitate was stirred for 30 min at 40–45 °C in a mixture of toluene and ethanol (6:1 *v*/*v*, 3 mL). After that, the precipitate was filtered off and washed with a small amount of toluene (1 mL) and ethanol (1 mL) to produce compound **3a**. Yield: 80.1 mg (52%); yellow solid; mp 133–135 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.84 (m, 1H), 7.62 (m, 1H), 7.54–7.47 (m, 7H), 7.37 (m, 1H), 7.34 (m, 2H), 7.29 (m, 2H), 7.23 (m, 4H), 7.13 (m, 1H), 6.65 (s, 1H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 192.8, 191.3, 165.1, 155.6, 136.2, 135.4, 134.9, 134.0, 133.7, 130.3, 128.9 (2C), 128.7 (2C), 128.6 (2C), 128.5 (2C), 128.3 (2C), 128.3, 126.5, 126.2, 124.4 (2C), 123.1, 117.4, 79.5, 66.0, 65.5 ppm. IR (mineral oil): 1763, 1716, 1678 cm⁻¹. Anal. Calcd (%) for C₃₁H₂₀N₂O₄S: C 72.08; H 3.90; N 5.42. Found: C 72.23; H 3.98; N 5.43.

(3*R**,3*a*S*,11*aR**)-3*a*-(4-Methylbenzoyl)-2,3-diphenyl-3,3*a*-dihydrobenzo[d]pyrrolo[3',4':2,3]pyrrolo [2,1-*b*]thiazole-1,4,5(2*H*)-trione (3*b*). The solvent was evaporated to 1 mL. The resulting precipitate was filtered off, washed with benzene (0.5 mL), and recrystallized from benzene (2 mL) to produce compound 3*b*. Yield: 28 mg (18%); yellow solid; mp 180–182 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.85 (m, 1 H), 7.52 (m, 3H), 7.43 (m, 2H), 7.34 (m, 2H), 7.30 (m, 3H), 7.27–7.17 (m, 6H), 7.13 (m, 1H), 6.67 (s, 1H), 2.33 (s, 3H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 191.7, 191.4, 170.2, 165.1, 155.6, 144.7, 136.2, 134.8, 134.0, 132.5, 130.3, 129.5 (2C), 128.9 (2C), 128.7, 128.6 (2C), 128.5 (2C), 128.3, 126.5, 126.3, 124.4 (2C), 123.2, 117.4, 114.5, 79.4, 65.9, 65.4, 21.0 ppm. IR (mineral oil): 1785, 1716, 1672 cm⁻¹. Anal. Calcd (%) for C₃₂H₂₂N₂O₄S: C 72.44; H 4.18; N 5.28. Found: C 72.67; H 4.28; N 5.32.

(3*R**,3*a*S*,11*aR**)-3*a*-(4-Fluorobenzoyl)-2,3-diphenyl-3,3*a*-dihydrobenzo[d]pyrrolo[3',4':2,3]pyrrolo [2,1-*b*]thiazole-1,4,5(2*H*)-trione (**3c**). The solvent was evaporated to 2.5 mL. The obtained mixture was frozen. The resulting precipitate was filtered off, washed with benzene (0.5 mL), and recrystallized from toluene (2 mL) to produce compound **3c**. Yield: 40 mg (25%); yellow solid; mp 111–113 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.82 (m, 1H), 7.67 (m, 2H), 7.52 (m, 3H), 7.39 (m, 2H), 7.33 (m, 2H), 7.29 (m, 1H), 7.25 (m, 4H), 7.22 (m, 2H), 7.18–7.11 (m, 3H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 191.6, 190.7, 165.2, 164.9 (d, *J* = 254.5 Hz), 155.8, 136.2, 134.8, 134.1, 132.0 (d, *J* = 10.1 Hz, 2C), 131.8 (d, *J* = 3.0 Hz), 130.7, 128.6 (2C), 128.6 (2C), 126.6, 126.1, 125.3, 124.4 (2C), 123.0, 117.6, 116.0 (d, *J* = 22.2 Hz, 2C), 79.9, 65.7, 65.5 ppm. ¹⁹F NMR (376 MHz, DMSO-*d*₆): δ = -104.24 ppm. IR (mineral oil): 1735, 1697, 1658 cm⁻¹. Anal. Calcd (%) for $2C_{31}H_{19}FN_2O_4S\cdotC_7H_8$: C 71.37; H 3.99; N 4.82. Found: C 71.51; H 4.08; N 4.99.

 $(3R^*,3aS^*,11aR^*)$ -3*a*-(4-Bromobenzoyl)-2,3-diphenyl-3,3*a*-dihydrobenzo[d]pyrrolo[3',4':2,3]pyrrolo [2,1-b]thiazole-1,4,5(2H)-trione (**3d**). The solvent was evaporated to 2.5 mL, the reaction mass was frozen. The resulting precipitate was filtered off, washed with benzene (0.5 mL), and recrystallized from toluene (2 mL) to produce compound **3d**. Yield: 30 mg (17%); yellow solid; mp 207–209 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.81 (m, 1H), 7.75 (m, 2H), 7.53 (m, 5H), 7.33 (m, 1H), 7.29 (m, 2H), 7.26 (m, 4H), 7.22 (m, 2H), 7.13 (m, 1H), 6.57 (s, 1H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 192.2, 190.5, 165.08, 155.6, 136.1, 134.8, 134.2, 133.9, 131.8 (2C), 130.6 (2C), 128.8, 128.6 (2C), 128.5 (2C), 128.1, 127.7, 126.4, 126.0, 124.2 (2C), 122.8, 117.5, 79.8, 65.7, 65.4 ppm. IR (mineral oil): 1757, 1728, 1701 cm⁻¹. Anal. Calcd (%) for C₃₁H₁₉BrN₂O₄S: C 62.53; H 3.22; N 4.70. Found: C 62.64; H 3.35; N 4.60.

(3*R**,3*a*S*,11*aR**)-3*a*-(*Furan*-2-*carbonyl*)-2,3-*diphenyl*-3,3*a*-*dihydrobenzo*[*d*]*pyrrolo*[3',4':2,3]*pyrrolo* [2,1-*b*]*thiazole*-1,4,5(2*H*)-*trione* (**3e**). The resulting precipitate was filtered off, washed with benzene (1 mL) and recrystallized from toluene (2–3 mL). Then, the obtained crystals were

stirred in a mixture of toluene and ethanol (5:1 v/v, 3 mL) at 50 °C for 10 min. Then, the precipitate was filtered off to produce compound **3e**. Yield: 71 mg (47%); yellow solid; mp 147–149 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.88 (m, 1H), 7.64 (m, 1H), 7.49 (m, 2H), 7.43 (m, 1H), 7.32 (m, 3H), 7.26 (m, 4H), 7.18 (m, 4H), 7.11 (m, 1H), 6.80 (s, 1H), 6.71 (m, 1H) ppm. ¹³C NMR (100 MHz, DMSO- d_6): δ = 191.3, 176.5, 165.4, 156.0, 149.7, 149.4, 136.4, 135.1, 133.9, 129.9, 129.3 (2C), 128.4, 128.3, 128.1, 127.7, 126.3, 126.1, 125.3, 124.7 (2C), 122.8, 121.8, 116.6, 114.3, 78.7, 65.5, 62.8 ppm. IR (mineral oil): 1770, 1732, 1716, 1674 cm⁻¹. Anal. Calcd (%) for 2C₂₉H₁₈N₂O₅S·C₇H₈: C 70.64; H 4.01; N 5.07. Found: C 70.82; H 4.11; N 5.00. (3R*,3aS*,11aR*)-2,3-Diphenyl-3a-(thiophene-2-carbonyl)-3,3a-dihydrobenzo[d]pyrrolo[3',4':2,3] pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (3f). The resulting precipitate was filtered off, washed with benzene (1 mL), and recrystallized from toluene (2–3 mL) to produce compound 3f. Yield: 70 mg (45%); yellow solid; mp 169–171 °C. ¹H NMR (400 MHz, DMSO- d_6): $\delta = 8.15$ (m, 1H), 7.87 (m, 1H), 7.51 (m, 3H), 7.40 (m, 1H), 7.35 (m, 1H), 7.31 (m, 4H), 7.26 (m, 2H), 7.23 (m, 2H), 7.21 (m, 2H), 7.17 (m, 1H), 7.12 (m, 1H), 6.75 (s, 1H) ppm. ¹³C NMR (100 MHz, DMSO- d_6): $\delta = 191.2, 182.1, 165.0, 155.2, 140.7, 137.9, 136.2, 134.7, 133.8, 130.3, 129.2, 128.9,$ 128.5, 128.4 (2C), 128.1 (2C), 126.3, 126.1, 125.2, 124.4 (2C), 123.0, 117.1, 79.1, 65.9, 64.6 ppm. IR (mineral oil): 1762, 1715, 1650 cm⁻¹. Anal. Calcd (%) for 2C₂₉H₁₈N₂O₄S₂·C₇H₈: C 68.64; H 3.90; N 4.93. Found: C 68.83; H 4.11; N 4.99.

 $(3R^*, 3aS^*, 11aR^*)$ -3a-Benzoyl-2-(3-nitrophenyl)-3-((E)-styryl)-3, 3a-dihydrobenzo[d]pyrrolo[3', 4': 2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**3g**). The solvent was evaporated to 1 mL. The resulting precipitate was filtered off, washed with benzene (0.5 mL), and recrystallized from acetonitrile (1 mL) to produce compound **3g**. Yield: 20 mg (12%); yellow solid; mp 212–214 °C. ¹H NMR (400 MHz, DMSO- d_6): δ = 8.51 (m, 1H), 8.07 (m, 1H), 7.96 (m, 1H), 7.87 (m, 1H), 7.69 (m, 1H), 7.62 (m, 1H), 7.50 (m, 5H), 7.36 (m, 2H), 7.29 (m, 2H), 7.24 (m, 3H), 6.73 (d, J 15.7 Hz, 1H), 6.18 (d, J 9.3 Hz, 1H), 6.02 (dd, J 15.7 Hz, J 9.8 Hz, 1H) ppm. ¹³C NMR (100 MHz, DMSO- d_6): δ = 193.3, 191.7, 165.4, 156.1, 147.9, 137.5, 137.4, 136.5, 136.0, 135.3, 134.1, 132.5, 130.4, 129.6 (2C), 129.2 (2C), 129.0, 128.6 (2C), 127.1 (2C), 126.9, 124.2, 123.6, 122.1, 121.6, 117.1, 78.9, 66.8, 64.2 ppm. IR (mineral oil): 1756, 1721, 1677 cm⁻¹. Anal. Calcd (%) for C₃₃H₂₁N₃O₆S: C 67.45; H 3.60; N 7.15. Found: C 67.62; H 3.71; N 7.10.

(3*R**,3*a*S*,11*a*R*)-3*a*-Benzoyl-3-(4-iodophenyl)-2-phenyl-3,3*a*-dihydrobenzo[d]pyrrolo[3',4':2,3] pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**3h**). The solvent was evaporated to 2.5 mL. The resulting precipitate was filtered off, washed with benzene (1 mL), and recrystallized from toluene (2–3 mL) to produce compound **3h**. Yield: 54 mg (28%); yellow solid; mp 170–172 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.85 (m, 1H), 7.59 (m, 3H), 7.51 (m, 3H), 7.47 (m, 4H), 7.36 (m, 1H), 7.30 (m, 3H), 7.14 (m, 3H), 6.68 (s, 1H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 192.4, 191.5, 165.0, 155.3, 137.0 (2C), 136.0, 135.6, 134.7, 133.8, 133.5, 131.0, 130.0, 128.9 (2C), 128.5 (2C), 128.2 (2C), 128.2, 127.8, 126.5, 126.2, 124.4 (2C), 123.0, 117.1, 95.2, 79.1, 66.3, 64.7 ppm. IR (mineral oil): 1762, 1715, 1691 cm⁻¹. Anal. Calcd (%) for C₃₁H₁₉IN₂O₄S: C 57.95; H 2.98; N 4.36. Found: C 58.16; H 3.10; N 4.28.

(3*R**,3*a*S*,11*aR**)-3*a*-Benzoyl-3-(4-bromophenyl)-2-(4-methoxyphenyl)-3,3*a*-dihydrobenzo[d]pyrrolo [3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**3i**). The reaction mixture was evaporated to dryness. Then, the residue was recrystallized from benzene (2–3 mL) to produce compound **3i**. Yield: 52 mg (28%); yellow solid; mp 221–223 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.84 (m, 1H), 7.61 (m, 1H), 7.52 (m, 1H), 7.46 (m, 4H), 7.42 (m, 4H), 7.33 (m, 2H), 7.25 (m, 2H), 6.84 (m, 2H), 6.61 (s, 1H), 3.69 (s, 3H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 192.5, 191.7, 165.0, 157.5, 155.4, 135.7, 134.8, 133.7 (2C), 133.5, 131.3, 131.2 (2C), 130.1, 129.0 (2C), 128.8, 128.3 (2C), 126.3, 126.2 (2C), 123.1, 122.0, 117.1, 113.8 (2C), 79.1, 66.5, 64.9, 55.1 ppm. IR (mineral oil): 1761, 1729, 1710, 1689 cm⁻¹. Anal. Calcd (%) for C₃₂H₂₁BrN₂O₅S: C 61.45; H 3.38; N 4.48. Found: C 61.63; H 3.50; N 4.40.

(3*R**,3*a*S*,11*aR**)-3*a*-Benzoyl-2-(4-chlorophenyl)-3-(3,4-dimethoxyphenyl)-3,3*a*-dihydrobenzo[d] pyrrolo[3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**3j**) and (3S*,3*a*S*,11*aR**)-3*a*-benzoyl-2-(4-chlorophenyl)-3-(3,4-dimethoxyphenyl)-3,3*a*-dihydrobenzo[d]pyrrolo[3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**3'j**). The reaction mixture was cooled to 10 °C. The resulting precipitate was filtered off (a mixture of products **3j** and **3'j**, 1:1). The mother liquor was evaporated

to dryness. Then, the residue was recrystallized from ethanol (2–3 mL) at 60–65 °C to afform product **3**j. Product **3**j: Yield: 50 mg (28%); yellow solid; mp 238–240 °C. ¹H NMR $(400 \text{ MHz}, \text{DMSO-}d_6): \delta = 7.84 \text{ (m, 1H)}, 7.62 \text{ (m, 1H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.34 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.34 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.54 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.54 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.54 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.54 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.54 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.54 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.54 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.54 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.54 \text{ (m, 4H)}, 7.58 \text{ (m, 2H)}, 7.50 \text{ (m, 5H)}, 7.54 \text{ (m, 4H)}, 7.58 \text{ (m, 5H)}, 7.54 \text{ (m, 5H$ 6.80 (m, 3H), 6.56 (s, 1H), 3.69 (s, 3H), 3.60 (s, 3H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 193.0, 190.7, 165.0, 155.8, 148.9, 148.4, 135.6, 135.2, 134.8, 133.4, 130.6, 130.2, 128.7 (2C), 128.4 (2C), 128.4 (2C), 128.1, 126.2 (2C), 126.1, 125.5, 123.0, 121.4, 117.2, 112.5, 111.3, 79.4, 66.0, 65.2, 55.5, 55.2 ppm. IR (mineral oil): 1761, 1726, 1713, 1682 cm⁻¹. Anal. Calcd (%) for C₃₃H₂₃ClN₂O₆S: C 64.86; H 3.79; N 4.58. Found: C 64.93; H 3.85; N 4.60. Product 3'j: Yield: 24 mg (13%, the mixture **3j**:**3**′**j**, 1:1), orange solid; mp 224–226 °C (mixture **3j**:**3**′**j**, 1:1). ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.80 (m, 1H), 7.62 (m, 2H), 7.56 (m, 2H), 7.36 (m, 5H), 7.30 (m, 2H), 6.77 (m, 1H), 6.71 (m, 3H), 6.47 (s, 1H), 3.59 (s, 3H), 3.54 (s, 3H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 191.0, 168.1, 165.7, 154.4, 148.7, 148.1, 135.7, 134.9, 134.4, 132.7, 130.7, 130.1, 129.0 (2C), 128.6 (2C), 127.7 (2C), 127.6, 125.9, 125.7 (2C), 122.43, 121.0, 115.8, 112.8, 111.4, 79.2, 70.6, 65.2, 55.3, 55.2 ppm. IR (mineral oil): 1761, 1713, 1680 cm⁻¹. Anal. Calcd. (%) for C₃₃H₂₃ClN₂O₆S: C 64.86; H 3.79; N 4.58. Found: C 64.12; H 3.94; N 4.71. (3R*,3aS*,11aR*)-3a-Benzoyl-3-(3,4-dimethoxyphenyl)-2-phenyl-3,3a-dihydrobenzo[d]pyrrolo [3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**3k**) and (3S*,3aS*,11aR*)-3a-benzoyl-3-(3,4dimethoxyphenyl)-2-phenyl-3,3a-dihydrobenzo[d]pyrrolo[3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)*trione* (3'k). The reaction mixture was cooled to 10 °C. The resulting precipitate was filtered off and recrystallized from benzene (2-3 mL) to give product 3'k. The mother liquor was evaporated to dryness, and the residue was recrystallized from toluene (2-3 mL) to produce product 3k. Product 3k: Yield: 40 mg (23%); yellow solid; mp 217–219 °C. ¹H NMR (400 MHz, DMSO- d_6): $\delta = 7.84$ (m, 1H), 7.64 (m, 1H), 7.54 (m, 7H), 7.33 (m, 4H), 7.16 (m, 1H), 6.86 (s, 1H), 6.81 (m, 2H), 6.54 (s, 1H), 3.68 (s, 3H), 3.61 (s, 3H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 193.1, 190.6, 165.0, 155.9, 148.9, 148.5, 136.3, 135.4, 134.9, 133.5, 130.3, 128.7 (2C), 128.6 (2C), 128.5 (2C), 128.1, 126.4, 126.1, 125.8, 124.3 (2C), 123.0, 121.1, 117.3, 112.2, 111.3, 79.6, 65.7, 65.5, 55.5, 55.2 ppm. IR (mineral oil): 1759, 1728, 1715, 1689 cm⁻¹. Anal. Calcd (%) for C₃₃H₂₄N₂O₆S: C 68.74; H 4.20; N 4.86. Found: C 68.90; H 4.27; N 4.79. Product **3'k**: Yield: 48 mg (28%); orange solid; mp 247–249 °C. ¹H NMR $(400 \text{ MHz}, \text{DMSO-}d_6): \delta = 7.82 \text{ (m, 3H)}, 7.62 \text{ (m, 1H)}, 7.52 \text{ (m, 3H)}, 7.40 \text{ (m, 2H)}, 7.32 \text{ (m, 2H$ 4H), 7.16 (m, 1H), 6.73 (m, 2H), 6.67 (m, 1H), 6.46 (s, 1H), 3.59 (s, 3H), 3.52 (s, 3H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 191.4, 188.0, 165.8, 154.5, 148.7, 148.0, 136.0, 135.8, 134.5, 132.9, 130.3, 129.2 (2C), 128.7 (2C), 127.9 (2C), 126.6, 126.1, 125.8, 124.1 (2C), 123.0, 122.5, 121.0, 115.9, 112.7, 111.2, 79.3, 70.8, 64.5, 55.3, 55.2 ppm. IR (mineral oil): 1762, 1709, 1669 cm⁻¹. Anal. Calcd (%) for C₃₃H₂₄N₂O₆S: C 68.74; H 4.20; N 4.86. Found: C 68.86; H 4.31; N 4.93.

(3*R**,3*a*S*,11*aR**)-3*a*-Benzoyl-2-benzyl-3-(4-bromophenyl)-3,3*a*-dihydrobenzo[*d*]pyrrolo[3',4':2,3] pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**3l**). The solvent was evaporated. The resulting mass was stirred in acetonitrile (2 mL) at 80 °C for 5 min. Then, the obtained mixture was cooled to room temperature, and the formed precipitate was filtered off to produce compound **3l**. Yield: 60 mg (33%); yellow solid; mp 204–206 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.79 (m, 1H), 7.59 (m, 3H), 7.54 (m, 1H), 7.37 (m, 9H), 7.16 (m, 2H), 7.09 (m, 2H), 5.58 (s, 1H), 4.95 (d, J 15.2 Hz, 1H), 3.71 (d, J 14.7 Hz, 1H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 192.7, 191.2, 166.1, 155.1, 134.7, 134.6, 134.0, 133.8, 132.6, 131.9 (2C), 130.7, 129.8, 128.8 (2C), 128.7 (2C), 128.3 (2C), 128.3, 128.1 (2C), 128.0, 127.6, 126.3, 123.2, 122.6, 117.3, 78.9, 65.6, 63.9, 45.4 ppm. IR (mineral oil): 1760, 1719, 1672 cm⁻¹. Anal. Calcd (%) for C₃₂H₂₁BrN₂O₄S: C 63.06; H 3.47; N 4.60. Found: C 63.14; H 3.58; N 4.63.

(3*R**,3*a*S*,11*a*R*)-3*a*-Benzoyl-2-(benzylideneamino)-3-phenyl-3,3*a*-dihydrobenzo[d]pyrrolo[3',4': 2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**3m**). The solvent was evaporated to 1 mL. The resulting precipitate was filtered off, stirred in benzene (2–3 mL) at 85 °C for 10 min, and then filtered off to produce compound **3m**. Yield: 42 mg (26%); yellow solid; mp 224–226 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 8.63 (s, 1H), 7.81 (m, 1H), 7.65 (m, 1H), 7.55 (m, 7H), 7.43 (m, 3H), 7.35 (m, 5H), 7.29 (2H), 6.54 (s, 1H) ppm. ¹³C NMR (100 MHz, DMSO-*d*₆): δ = 192.4, 190.4, 162.3, 156.6, 155.5, 134.9, 134.6, 133.7, 133.5, 132.9, 131.3, 130.4, 129.0, 128.9

(2C), 128.8 (2C), 128.8 (2C), 128.7 (2C), 128.3, 128.0 (2C), 127.5 (2C), 126.1, 123.0, 117.5, 78.3, 65.3, 64.7 ppm. IR (mineral oil): 1757, 1725, 1686 cm⁻¹. Anal. Calcd (%) for C₃₂H₂₁N₃O₄S: C 70.71; H 3.89; N 7.73. Found: C 70.93; H 3.97; N 7.81.

3.1.3. Procedure to Compounds 8a-j

A mixture of APBTT **1** (0.298 mmol) and carbodiimide **7** (0.313 mmol) in toluene (5 mL) was heated for 1 h at 115 °C (until the dark violet color characteristic of APBTT **1** disappeared and a transparent yellow solution formed). Then, the reaction mixture was cooled to room temperature. The resulting precipitate was filtered off and stirred in ethanol (2 mL) at 45–50 °C for 30 min. Then, the precipitate was filtered off and recrystallized from toluene (5 mL) to produce the corresponding compound **8**.

(3*a*S*,11*a*R*)-3*a*-Benzoyl-2-cyclohexyl-3-(cyclohexylimino)-3,3*a*-dihydrobenzo[*d*]pyrrolo[3',4':2,3] pyrrolo[2,1-*b*]thiazole-1,4,5(2H)-trione (**8a**). Yield: 69 mg (44%); yellow solid; mp 264–266 °C. ¹H NMR (400 MHz, CDCl₃): δ = 7.87 (m, 1H), 7.68 (m, 3H), 7.49 (m, 2H), 7.25 (m, 3H), 4.31 (m, 1H), 3.41 (m, 1H), 2.32 (m, 2H), 1.86 (s, 3H), 1.73 (m, 4H), 1.51 (m, 1H), 1.42 (m, 3H), 1.36 (m, 2H), 1.21 (m, 2H), 0.91 (m, 2H), 0.38 (m, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 190.7, 187.7, 168.0, 154.1, 141.6, 135.1, 134.8, 133.8, 129.3, 129.1 (2C), 128.7 (2C), 128.5, 126.9, 122.6, 118.0, 77.6, 65.4, 60.6, 54.0, 34.1, 32.1, 28.2, 27.8, 25.9, 25.8, 25.7, 25.2, 23.9, 23.8 ppm. IR (mineral oil): 1779, 1749, 1722, 1677 cm⁻¹. Anal. Calcd (%) for C₃₁H₃₁N₃O₄S: C 68.74; H 5.77; N 7.76. Found: C 68.85; H 5.83; N 7.71.

(3*a*S*,11*a*R*)-2-*Cyclohexyl*-3-(*cyclohexylimino*)-3*a*-(4-*methylbenzoyl*)-3,3*a*-dihydrobenzo[*d*]*pyrrolo* [3',4':2,3]*pyrrolo*[2,1-*b*]*thiazole*-1,4,5(2*H*)-*trione* (**8b**). Yield: 25 mg (15%); yellow solid; mp 230–232 °C. ¹H NMR (400 MHz, CDCl₃): δ = 7.87 (m, 1H), 7.59 (m, 2H), 7.29 (m, 2H), 7.24 (m, 3H), 4.33 (m, 1H), 3.45 (m, 1H), 2.46 (s, 3H), 2.31 (m, 2H), 1.86 (m, 3H), 1.75 (m, 1H), 1.69 (m, 4H), 1.53 (m, 1H), 1.44 (m, 3H), 1.39 (m, 1H), 1.35 (m, 1H), 1.29 (m, 1H), 1.20 (m, 2H), 0.93 (m, 2H), 0.47 (m, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 190.1, 187.8, 168.1, 154.1, 146.6, 141.8, 134.8, 131.2, 129.8, 129.4, 129.0, 128.9, 128.5, 128.2, 126.8, 125.3, 122.5, 118.0, 77.7, 60.5, 54.0, 34.1, 32.2, 28.2, 27.8, 25.9, 25.8, 25.7, 25.2, 24.0, 23.7, 21.8 ppm. IR (mineral oil): 1788, 1744, 1729, 1674 cm⁻¹. Anal. Calcd (%) for C₃₂H₃₃N₃O₄S: C 69.17; H 5.99; N 7.56. Found: C 69.55; H 6.12; N 7.66.

(3*a*S*,11*a*R*)-2-*Cyclohexyl*-3-(*cyclohexylimino*)-3*a*-(4-fluorobenzoyl)-3,3*a*-dihydrobenzo[d]pyrrolo [3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**8c**). Yield: 18 mg (11%); yellow solid; mp 228–230 °C. ¹H NMR (400 MHz, CDCl₃): δ = 7.92 (m, 1H), 7.79 (m, 2H), 7.33 (m, 2H), 7.23 (m, 3H), 4.36 (m, 1H), 3.47 (m, 1H), 2.36 (m, 2H), 1.92 (m, 3H), 1.80 (m, 1H), 1.74 (m, 3H), 1.60–1.47 (m, 4H), 1.45–1.37 (m, 2H), 1.29–1.19 (m, 2H), 1.00 (m, 2H), 0.52 (m, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 189.1, 187.4, 168.0, 166.7 (d, *J* = 261.6 Hz), 154.0, 141.5, 134.7, 131.5 (d, *J* = 10.1 Hz, 2C), 130.2 (d, *J* = 3.0 Hz), 129.1, 128.6, 127.0, 122.6, 118.1, 116.5 (d, *J* = 24.2 Hz, 2C), 77.5, 65.3, 60.6, 54.1, 34.1, 32.3, 28.3, 27.8, 25.9, 25.8, 25.6, 25.1, 23.9, 23.8 ppm. ¹⁹F NMR (376 MHz, DMSO-*d*₆): δ = -100.03 ppm. IR (mineral oil): 1789, 1749, 1726, 1673 cm⁻¹. Anal. Calcd (%) for C₃₁H₃₀FN₃O₄S: C 66.53; H 5.40; N 7.51. Found: C 66.68; H 5.51; N 7.60.

(3*a*S*,11*a*R*)-3*a*-(4-Bromobenzoyl)-2-cyclohexyl-3-(cyclohexylimino)-3,3*a*-dihydrobenzo[d]pyrrolo [3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**8d**). Yield: 57 mg (31%); yellow solid; mp 180–182 °C. ¹H NMR (400 MHz, CDCl₃): δ = 7.92 (m, 1H), 7.71 (m, 2H), 7.61 (m, 2H), 7.32 (m, 1H), 7.26 (m, 2H), 4.35 (m, 1H), 3.46 (m, 1H), 2.36 (m, 2H), 1.91 (m, 3H), 1.80 (m, 1H), 1.74 (m, 2H), 1.60 (m, 3H), 1.48 (m, 1H), 1.46–1.31 (m, 3H), 1.25 (m, 2H), 1.02 (m, 2H), 0.55 (m, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 189.6, 187.3, 167.9, 153.9, 141.3, 134.7, 132.5 (2C), 130.7, 130.0 (2C), 129.0, 128.6, 127.0, 122.6, 118.1, 77.4, 65.2, 60.5, 54.1, 34.1, 32.3, 28.2, 27.8, 25.8, 25.6, 25.1, 23.9, 23.8 ppm. IR (mineral oil): 1791, 1789, 1752, 1726, 1672 cm⁻¹. Anal. Calcd (%) for C₃₁H₃₀BrN₃O₄S: C 60.00; H 4.87; N 6.77. Found: C 60.14; H 4.97; N 6.65. (3*a*S*,11*a*R*)-2-Cyclohexyl-3-(cyclohexylimino)-3*a*-(furan-2-carbonyl)-3,3*a*-dihydrobenzo[d]pyrrolo [3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**8e**). Yield: 71 mg (45%); yellow solid; mp 251–253 °C. ¹H NMR (400 MHz, CDCl₃): δ = 7.91 (m, 1H), 7.60 (s, 1H), 7.51 (m, 1H), 7.28 (m, 3H), 6.72 (s, 1H), 4.34 (m, 1H), 3.60 (br.s, 1H), 2.34 (m, 2H), 1.85 (m, 4H), 1.73 (d, J = 12, 3H), 1.63 (m, 2H), 1.52 (m, 2H), 1.38 (m, 3H), 1.22 (m, 3H), 0.95–0.62 (m, 1H) ppm. 13 C NMR (100 MHz, CDCl₃): δ = 186.6, 168.2, 149.4 (2C), 140.5, 134.9, 129.3, 128.4, 126.7, 122.6, 120.9, 118.0, 114.2, 77.7, 77.2, 54.1, 34.0, 28.3, 27.8, 25.9, 25.9, 25.7, 25.2, 24.0, 23.8 ppm. IR (mineral oil): 1775, 1745, 1721, 1667 cm⁻¹. Anal. Calcd (%) for C₂₉H₂₉N₃O₅S: C 65.52; H 5.50; N 7.90. Found: C 65.71; H 5.58; N 8.03.

 $(3aS^*,11aR^*)-2-Cyclohexyl-3-(cyclohexylimino)-3a-(thiophene-2-carbonyl)-3,3a-dihydrobenzo[d] pyrrolo[3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione ($ **8f** $). Yield: 96 mg (59%); yellow solid; mp 274–276 °C. ¹H NMR (400 MHz, CDCl₃): <math>\delta$ = 7.91 (m, 2H), 7.55 (m, 1H), 7.32 (m, 2H), 7.23 (m, 2H), 4.39 (m, 1H), 3.70 (m, 1H), 2.38 (m, 2H), 1.94 (m, 3H), 1.83 (m, 1H), 1.76 (m, 3H), 1.60 (m, 2H), 1.54 (m, 2H), 1.42 (m, 2H), 1.29 (m, 2H), 1.13 (m, 2H), 0.75 (m, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 170.7, 168.0, 154.0, 141.3, 137.4 (2C), 134.8, 133.1, 129.2, 128.6, 128.5, 126.9, 122.6, 118.0, 77.8, 77.2, 60.4, 54.1, 34.2, 32.3, 28.3, 27.8, 25.9, 25.8, 25.7, 25.2, 24.0, 23.8 ppm. IR (mineral oil): 1779, 1747, 1724, 1682, 1657 cm⁻¹. Anal. Calcd (%) for C₂₉H₂₉N₃O₄S₂: C 63.60; H 5.34; N 7.67. Found: C 63.83; H 5.41; N 7.54.

 $(3aS^*,11aR^*)$ -3a-(4-Chlorobenzoyl)-2-cyclohexyl-3-(cyclohexylimino)-3,3a-dihydrobenzo[d]pyrrolo[3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**8g**). Yield: 43 mg (25%); yellow solid; mp 248–250 °C. ¹H NMR (400 MHz, CDCl₃): δ = 7.92 (m, 1H), 7.69 (m, 2H), 7.53 (m, 2H), 7.30 (m, 3H), 4.35 (m, 1H), 3.46 (m, 1H), 2.36 (m, 2H), 1.92 (m, 3H), 1.80 (m, 1H), 1.74 (m, 3H), 1.58 (m, 2H), 1.48 (m, 2H), 1.39 (m, 2H), 1.26 (m, 2H), 1.01 (m, 2H), 0.53 (m, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 189.4, 187.4, 167.9, 153.9, 142.0, 141.3, 134.7, 132.1, 130.0 (2C), 129.5 (2C), 129.1, 128.6, 127.0, 122.6, 118.1, 77.5, 65.3, 60.5, 54.1, 34.1, 32.3, 28.3, 27.8, 25.9, 25.8, 25.6, 25.1, 23.9, 23.8 ppm. IR (mineral oil): 1791, 1789, 1751, 1726, 1674 cm⁻¹. Anal. Calcd (%) for C₃₁H₃₀ClN₃O₄S: C 64.63; H 5.25; N 7.29. Found: C 64.69; H 5.19; N 7.37. ($3aS^*,11aR^*$)-3a-Benzoyl-2-isopropyl-3-(isopropylinino)-3,3a-dihydrobenzo[d]pyrrolo[3',4':2,3]

pyrrolo[2,1-*b*]*thiazole*-1,4,5(2*H*)-*trione* (**8**h). Yield: 89 mg (65%); yellow solid; mp 207–207 °C. ¹H NMR (400 MHz, CDCl₃): δ = 7.94 (m, 1H), 7.74 (m, 3H), 7.56 (m, 2H), 7.35–7.25 (m, 3H), 4.78 (m, 1H), 3.79 (m, 1H), 1.54 (m, 6H), 1.30 (m, 3H), 0.50 (d, J = 8 Hz, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 190.3, 187.8, 167.9, 154.1, 141.4, 135.2, 134.8, 133.8, 129.2, 129.2 (2C), 128.8 (2C), 128.5, 126.9, 122.6, 118.1, 77.6, 66.7, 52.8, 46.1, 24.4, 22.0, 18.8, 18.4 ppm. IR (mineral oil): 1786, 1749, 1724, 1669 cm⁻¹. Anal. Calcd (%) for C₂₅H₂₃N₃O₄S: C 65.06; H 5.02; N 9.10. Found: C 65.17; H 5.09; N 9.18.

(3*a*S*,11*a*R*)-2-*Isopropyl*-3-(*isopropylimino*)-3*a*-(4-*methylbenzoyl*)-3,3*a*-*dihydrobenzo*[*d*]*pyrrolo* [3',4':2,3]*pyrrolo*[2,1-*b*]*thiazole*-1,4,5(2*H*)-*trione* (**8i**). Yield: 106 mg (75%); yellow solid; mp 216–218 °C. ¹H NMR (400 MHz, CDCl₃): δ = 7.94 (m, 1H), 7.67 (m, 2H), 7.37 (m, 2H), 7.35–7.25 (m, 3H), 4.79 (m, 1H), 3.84 (m, 1H), 2.52 (s, 3H), 1.55 (m, 6H), 1.31 (m, 3H), 0.53 (m, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 189.8, 187.8, 167.9, 154.1, 146.8, 141.6, 134.8, 131.2, 129.9, 129.3, 129.0, 128.5, 126.9 (2C), 122.6, 118.0 (2C), 115.0, 52.7, 46.1, 24.4, 22.1, 21.8, 18.8, 18.4 ppm. IR (mineral oil): 1786, 1751, 1727 cm⁻¹. Anal. Calcd (%) for C₂₆H₂₅N₃O₄S: C 65.67; H 5.30; N 8.84. Found: C 65.81; H 5.26; N 8.80.

 $(3aS^*,11aR^*)$ -3a-(4-Chlorobenzoyl)-2-isopropyl-3-(isopropylimino)-3,3a-dihydrobenzo[d]pyrrolo [3',4':2,3]pyrrolo[2,1-b]thiazole-1,4,5(2H)-trione (**8j**). Yield: 106 mg (72%); yellow solid; mp 228–230 °C. ¹H NMR (400 MHz, CDCl₃): δ = 7.64 (m, 1H), 7.41 (m, 2H), 7.30 (m, 1H), 7.06–6.96 (m, 4H), 4.48 (m, 1H), 3.49 (m, 1H), 1.25 (d, J = 4 Hz, 6H), 1.02 (d, J = 8 Hz, 3H), 0.29 (d, J = 8 Hz, 3H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 189.1, 187.4, 167.7, 153.9, 142.1, 141.2, 134.7, 132.0, 131.3, 130.1 (2C), 129.6, 129.0, 128.6 (2C), 127.0, 122.6, 118.1, 52.8, 46.2, 24.4, 22.2, 18.8, 18.4 ppm. IR (mineral oil): 1786, 1750, 1726, 1664 cm⁻¹. Anal. Calcd (%) for C₂₅H₂₂ClN₃O₄S: C 60.54; H 4.47; N 8.47. Found: C 60.76; H 4.58; N 8.55.

3.1.4. Procedures to Compounds 4a,b,6d

5-Hydroxy-4-(2-oxo-2H-benzo[b][1,4]thiazin-3(4H)-ylidene)-1,5-diphenylpyrrolidine-2,3-dione (4a). Method 1. Acetic acid (3 mL) was added to the mixture of APBTT 1a (100 mg, 0.298 mmol) and *N*-benzylideneaniline 2a (54 mg, 0.298 mmol). The mixture was stirred for 24 h at room temperature. The resulting precipitate was filtered off and washed with acetone (2 mL) to produce compound 4a. Yield: 110 mg (86%); red solid; mp 179–181 °C. Method 2. Aniline

5a (27.2 μL, 0.298 mmol) was added to the mixture of acetic acid (3 mL) and APBTT **1a** (100 mg, 0.298 mmol). The mixture was stirred for 24 h at room temperature. The resulting precipitate was filtered off and washed with acetone (2 mL) to produce compound **4a**. Yield: 121 mg (95%); red solid; mp 179–181 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 14.05 (s, 1H), 7.60 (m, 1H), 7.52 (m, 2H), 7.38 (m, 1H), 7.34 (m, 2H), 7.22 (m, 3H), 7.18 (m, 1H), 7.16 (s, 1H), 7.13 (m, 1H), 7.11 (s, 1H), 7.04 (m, 2H) ppm. IR (mineral oil): 3467, 3169, 3083, 1723, 1684, 1632 cm⁻¹. Anal. Calcd (%) for C₂₄H₁₆N₂O₄S: C 67.28; H 3.76; N 6.54. Found: C 67.39; H 3.81; N 6.47.

1-Benzyl-5-hydroxy-4-(2-oxo-2H-benzo[b][1,4]thiazin-3(4H)-ylidene)-5-(4-methylphenyl)pyrrolidine -2,3-dione (**4b**). Benzylamine **5b** (15.6 μL, 0.143 mmol) was added to the mixture of acetic acid (3 mL) and APBTT **1a** (50 mg, 0.143 mmol). The mixture was stirred for 24 h at room temperature. The resulting precipitate was filtered off and washed with acetone (1 mL) to produce compound **4b**. Yield: 63 mg (46%); orange solid; mp 162–164 °C. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 14.24 (s, 1H), 7.55 (m, 1H), 7.46 (m, 2H), 7.31 (m, 3H), 7.12 (m, 3H), 7.03 (m, 4H), 6.67 (s, 1H), 4.20 (dd, *J* 64.1 Hz, *J* 15.2 Hz, 2H), 2.24 (s, 3H) ppm. ¹³C NMR (400 MHz, DMSO-*d*₆): δ = 183.1, 177.4, 160.4, 138.8, 137.8, 137.0, 136.3, 129.5, 128.6, 128.1 (2C), 127.8 (2C), 127.5 (2C), 126.3, 126.1, 125.9 (2C), 125.5, 120.5, 119.4, 109.9, 88.6, 42.5, 20.5 ppm. IR (mineral oil): 3474, 3192, 3058, 1720, 1640 cm⁻¹. Anal. Calcd (%) for C₂₆H₂₀N₂O₄S: C 68.41; H 4.42; N 6.14. Found: C 68.56; H 4.47; N 6.20.

(Z)-3-(*Benzo*[*d*]*thiazo*1-2(3*H*)-*y*l*idene*)-4-(4-*bromopheny*])-2,4-*dioxo*-*N*-*pheny*]*butanamide* (6*d*). This product was a by-product in the synthesis of compound 3d. Product 6d was precipitated in the first fraction upon recrystallization of the main product from toluene. Yield: 3 mg (2%); colorless crystals; mp 179–181 °C. ¹H NMR (400 MHz, CDCl₃): δ = 13.76 (br.s, 1H), 10.39 (s, 1H), 8.07 (m, 1H), 7.53 (m, 3H), 7.43 (m, 3H), 7.29 (m, 2H), 7.21 (m, 3H), 7.02 (m, 1H) ppm. IR (mineral oil): 3278, 3136, 1718, 1672, 1635 cm⁻¹. Anal. Calcd (%) for C₂₃H₁₅BrN₂O₃S: C 57.63; H 3.15; N 5.84. Found: C 57.86; H 3.24; N 5.91.

3.2. Computational Details

The DFT calculations for all model structures were carried out at the M06-2X/6-31G^{*} level of theory with the help of the Gaussian-09 program package [81]. No symmetry restrictions have been applied during the geometry optimization procedure. The Hessian matrices were calculated analytically for all optimized model structures to prove the location of correct minima or saddle points (transition states) on the potential energy surface. The Cartesian atomic coordinates for all model structures are presented in attached xyz-files (Supplementary Materials).

3.3. Biology

3.3.1. Screening of Substances in 96-Well Plates

Chlorella vulgaris (strain IMBR-19, obtained from the A.O. Kovalevsky Institute of Biology of the Southern Seas of RAS, Sevastopol, Russia) was cultivated in BG-11 medium. BG-11 medium was prepared using following solutions: Solution #1: Na₂-EDTA—20 mg in 20 mL; Solution #2: citric acid—120 mg and iron(III) citrate—120 mg in 20 mL; Solution #3: K₂HPO₄·3H₂O—800 mg in 20 mL; Solution #4. MgSO₄·7H₂O—1.5 g in 20 mL; Solution #5: CaCl₂·2H₂O—720 mg in 20 mL; Solution #6: Na₂CO₃—400 mg in 20 mL; Solution #7: NaNO₃—15 g in 100mL; Solution #8: H₃BO₃—57.2 mg, MnCl₂·4H₂O—36.2 mg, ZnSO₄·7H₂O—4.4 mg, CuSO₄·5H₂O—1.58 mg, Na₂MoO₄·2H₂O—7.8 mg, and 1 mL of 0.988 g/L Co(NO₃)₂·6H₂O in 20 mL. All of the solutions were prepared using Milli-Q water. To obtain 500 mL of BG-11 medium, 500 µL of solutions #1–6 and #8 and 5 mL of solution #7 were added to 400 mL of water. Then, the volume was adjusted to 500 mL. A solution of D(+)-glucose in BG-11 (6 g/L) was used for the preparation of positive controls.

Cultures of *C. vulgaris* were maintained and cultivated in aseptic conditions, with the only exception being DMSO solutions of tested substances that were not sterilized upon dilution.

Substances to be tested were diluted to 1×10^{-3} , 1×10^{-4} , and 1×10^{-5} mol/L in 99% DMSO before the experiment. Poorly soluble substances were either kept for 16 h on a rotator or treated by ultrasound using an ultrasound homogenizer equipped with a 3 mm probe (VCX-130, Sonics and Materials, Newtown, CT, USA).

Starter cultures of *C. vulgaris* were prepared as follows: stock culture cells (2 mL, in exponential growth phase) were washed two times with BG-11 medium by centrifugation (20 min, 350 g). The cells were then diluted in 10 mL of BG-11, and the cell concentration was measured using a hemocytometer.

In the wells of 96-well culture plates, BG-11 medium, starter culture of *C. vulgaris*, and tested substances diluted in DMSO were combined, resulting in a total volume of 300 µL. The resulting number of cells was 5×10^4 cells/well, with a resulting volume fraction of DMSO at 1%. The resulting concentrations of tested substances were 1×10^{-5} , 1×10^{-6} , and 1×10^{-7} mol/L. In negative control and positive control wells, pure DMSO was added. BG-11 with glucose was added to the positive control wells, resulting in a concentration of glucose of 2 g/L. All substances were tested in duplicate (2 wells for each concentration). In the edge and corner wells of the plates, sterile distilled water was added. The plates were sealed with a gas-permeable film.

All cultures were maintained for 5 days in a humid chamber at +28 °C at 150 rpm under cyclic illumination consisting of 12 h on: 12 h off. The light intensity was 100 μ mol·m⁻²·s⁻¹. The lighting unit was an array of evenly distributed white LEDs with a cooling device preventing well heating, positioned below the culture plates (bottom illumination).

After the end of cultivation, the contents of the wells were mixed using a multichannel pipette, and the cell concentration was assessed by measuring the absorbance at 750 nm.

3.3.2. Evaluation of Lead Substances in 50-mL Flasks

Substances to be tested were diluted to 1×10^{-2} , 1×10^{-3} , 1×10^{-4} , and 1×10^{-5} mol/L in 99% DMSO before the experiment. Starter cultures of *C. vulgaris* were prepared as follows: stock culture cells (12 mL, in exponential growth phase) were washed twice with BG-11 medium by centrifugation (15 min, 450 g). The cells were then diluted in 10 mL of BG-11, and the cell concentration was measured using a hemocytometer.

In the 50-mL Erlenmeyer flasks, BG-11 medium, the starter culture of *C. vulgaris*, and tested substances diluted in DMSO were combined, resulting in a total volume of 30 mL. The resulting number of cells was 1×10^7 cells/flask, with a resulting volume fraction of DMSO at 1%. The resulting concentrations of tested substances were 1×10^{-4} , 1×10^{-5} , 1×10^{-6} , and 1×10^{-7} mol/L. In the negative control and positive control flasks, pure DMSO was added. BG-11 with glucose was added to the positive control flasks, resulting in a concentration of glucose of 2 g/L. All substances were tested in triplicate (three flasks for each concentration). The flasks were sealed with gas-permeable cellulose caps. All cultures were maintained for 5 days in a humid chamber at +28 °C at 150 rpm under cyclic illumination consisting of 12 h on: 12 h off. The light intensity was 100 µmol·m⁻²·s⁻¹. The lighting unit was an array of evenly distributed white LEDs with a cooling device preventing overheating, positioned below the culture plates (bottom illumination).

3.3.3. Cell Count and Pigments Analysis

The flask contents were carefully mixed, and then 10 mL of cell culture was transferred into 15-mL centrifuge tubes. The cells were washed twice with 10 mL of water by centrifugation (15 min, 450 g). The washed cells were diluted in 10 mL of water, and the cell concentration was measured using a hemocytometer.

Pigment extraction was carried out as follows: two milliliters of cell culture were transferred to centrifuge tubes. The cells were washed by centrifugation two times at $7000 \times g$ for 10 min and concentrated two-fold. After the second wash, the sediment was vortexed for 1 min, and then 90% methanol was added. The tubes were heated at +60 °C for 30 min in a solid-state thermostat. Then, the samples were cooled to room temperature and centrifuged at $10,000 \times g$ for 10 min. The absorbance of the supernatant containing extracted

pigments was measured at 665, 652, and 470 nm. The concentrations of chlorophylls and carotenoids were calculated as described in the papers [82,83]. After that, the concentration of pigments in micrograms per 1×10^7 cells was calculated.

4. Conclusions

An approach to a new 6/5/5/5-tetracyclic alkaloid-like spiroheterocyclic system of benzo[*d*]pyrrolo[3',4':2,3]pyrrolo[2,1-*b*]thiazole **3**, **8** was developed on the basis of a reaction of 3-aroylpyrrolo[2,1-*c*][1,4]benzothiazine-1,2,4-triones **1** with Schiff bases **2** and carbodiimides **7**. This reaction proceeded as a nucleophile-induced ring contraction—intramolecular cyclization cascade. The formation of the benzo[*d*]pyrrolo[3',4':2,3]pyrrolo[2,1-*b*]thiazoles **3**, **8** was found to be diastereoselective, with the exception of compounds **3k**,**j**, **3'k**,**j**. Compounds **3a**, **8j** were found to promote the growth of *Chlorella vulgaris* and to increase the chlorophyll content in its cells.

5. Patents

The method for preparing products 8 has been patented [84].

Supplementary Materials: The following supporting information can be downloaded at: https://www. mdpi.com/article/10.3390/molecules29092089/s1, The following are available online, copies of NMR spectra for new compounds **3a–m**, **3'**j,**k**, **4a**,**b**, **6d**, **8a–j**, details of DFT calculations, Cartesian atomic coordinates for all model structures, ORTEP images of X-ray crystal structures.

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