

Supporting Information

Synthesis of Sucrose-Mimicking Disaccharide by Intramolecular Aglycone Delivery

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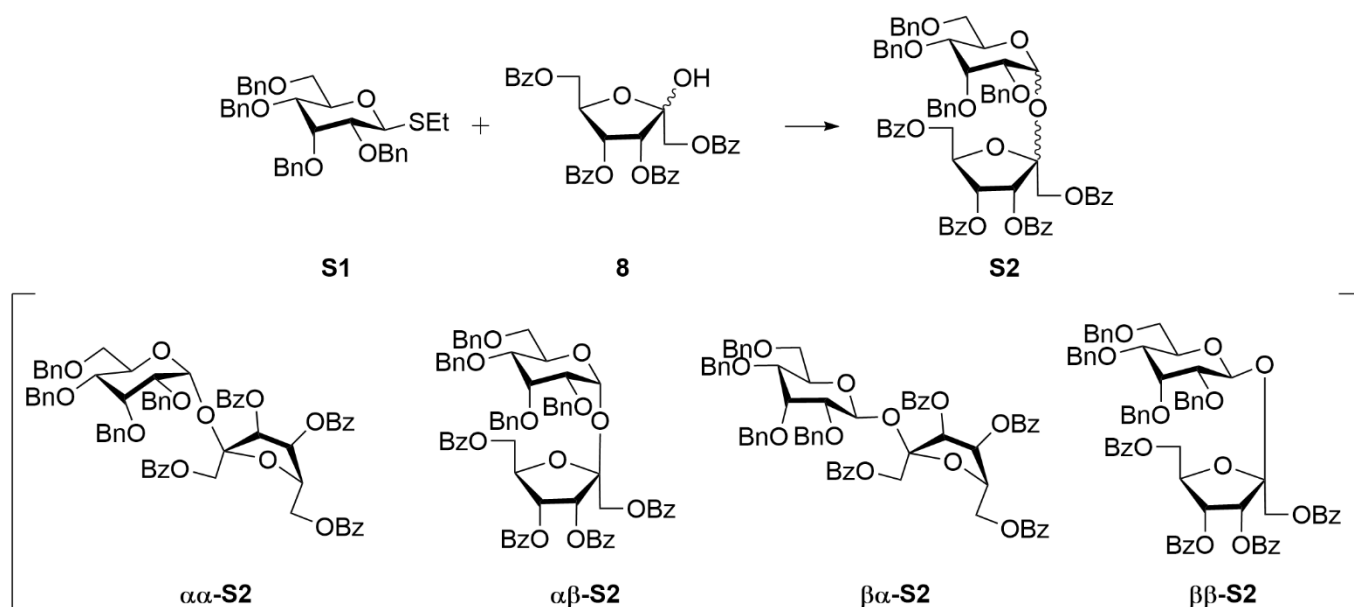
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Scheme S1. Synthesis of the stereoisomers of D-allosyl-D-psicofuranoside derivatives **S2** by the conventional glycosylation method.

Synthesis of allose donor **S1**

To a solution of compound **3** (1.7 g, 7.65 mmol) and benzyl bromide (7.3 mL, 61.2 mmol) in DMF (40 mL), 60% NaH (1.8 g, 45.9 mmol) was added at 0 °C. The reaction mixture was stirred at room temperature for 12 hours. The reaction was quenched by adding MeOH (excess) at 0 °C. After diluting the reaction mixture with ethyl acetate, it was washed with 1M HCl, brine, saturated aq. NaHCO₃, and brine, successively. The organic layer was dried with Na₂SO₄, filtered, and the solvent was removed. The resulting residue was purified by flash silica gel column chromatography using hexane–ethyl acetate (5/1, v/v) to afford compound **S1** (1.56 g, 21.4 mmol, 35%). *R*_f = 0.53 (hexane/ethyl acetate = 3/1, v/v); [α]²⁴_D –6.20 (*c* 1.00, CHCl₃), ¹H-NMR (400 MHz, CDCl₃) δ 7.38–7.25 (m, 20H, aromatic *H*), 5.02 (d, 1H, *J* = 10.4 Hz, H-1), 4.84 (d, 1H, *J* = 12.4 Hz, CH₂), 4.78 (d, 1H, *J* = 12.0 Hz, CH₂), 4.57 (m, 6H, CH₂), 4.40 (d, 1H, *J* = 12.0 Hz, CH₂), 4.11 (t, 1H, *J* = 2.4 Hz, H-3), 4.02 (ddd, 1H, *J* = 1.6 Hz, *J* = 4.4 Hz, *J* = 6.4 Hz, *J* = 10.0 Hz, H-5), 3.75 (dd, 1H, *J* = 2.0 Hz, *J* = 11.2 Hz, H-6), 3.67 (dd, 1H, *J* = 4.4 Hz, *J* = 11.2 Hz, H-6), 3.45 (dd, 1H, *J* = 2.4 Hz, *J* = 9.6 Hz, H-4), 3.27 (dd, 1H, *J* = 2.8 Hz, *J* = 10.0 Hz, H-2), 2.73 (m, 2H, SCH₂CH₃), 1.31 (t, 3H, *J* = 7.6 Hz, SCH₂CH₃). ¹³C-NMR (100 MHz, CDCl₃) δ 138.9–127.7 (aromatic *C*), 81.5 (C-1), 78.7 (C-2), 75.6 (C-4), 74.9 (C-5), 74.2 (CH₂), 73.3 (C-3, CH₂), 72.3 (CH₂), 71.5 (CH₂), 69.4 (C-6), 24.7 (SCH₂CH₃), 15.1 (SCH₂CH₃). HRMS ESI-TOF: calcd. for C₃₆H₄₀NaO₅S [M+Na]⁺ 607.2489; found 607.2490.

Synthesis of the stereoisomer of D-allosyl-D-psicofuranoside derivatives

To a solution of glycosyl donor **S1** (53 mg, 0.091 mmol), glycosyl acceptor (50 mg, 0.083 mmol), in CH₂Cl₂ (4.0 mL) in the presence of NIS (28 mg, 0.125 mmol) and MS4Å (300 mg) was added TfOH (1.5 μ L, 0.017 mmol) and stirred at –40 °C for 1.5 hours. The triethylamine (12 μ L, 0.083 mmol) was added at –40 °C. The reaction mixture was diluted with ethyl acetate, it was washed with 1 M HCl, brine, saturated aq. NaHCO₃ and brine, in that order. The organic layer was dried with MgSO₄, and the solvent was removed. The resulting residue was purified by gel filtration chromatography with chloroform. The di-saccharide fraction was purified by flash silica gel column chromatography with toluene–ethyl acetate (9/1, v/v) to afford compounds $\alpha\alpha$ -**S2** (α All- α Psi, 29 mg, 0.026 mmol, 31%) and mixture of $\alpha\beta$ -**S2** (α All- β Psi, 6.5 mg, 0.006 mmol, 7%), $\beta\alpha$ -**S2** (β All- α Psi, 21 mg, 0.019 mmol, 22%) and $\beta\beta$ -**S2** (β All- β Psi, 4.9 mg, 0.004 mmol, 5%). The yields of $\alpha\beta$ -**S2**, $\beta\alpha$ -**S2**, and $\beta\beta$ -**S2** were calculated from the amount of mixture and the integral value of the NMR spectra.

$\alpha\alpha$ -**S2**; ¹H-NMR (400 MHz, CDCl₃) δ 8.06–7.08 (aromatic *H*), 5.78 (d, 1H, *J* = 8.4 Hz, H-3^I), 5.72 (d, 1H, *J* = 4.0 Hz, H-1^{II}), 5.66 (dd, 1H, *J* = 6.0, 7.2 Hz, H-4^I), 4.94 (broad dd, *J* = 4.6, 8.2 Hz, H-5^I), 4.83 (d, 1H, *J* = 12.4 Hz, H-1^I), 4.77 (d, 1H, *J* = 13.6 Hz, CH₂), 4.70 (m, 3H, H-1^I, H-6^I, CH₂), 4.53 (d, 1H, *J* = 12.4 Hz, H-6^I), 4.56 (d, 2H, *J* = 13.6 Hz, CH₂, CH₂), 4.39 (d, 1H, *J* = 13.6 Hz, CH₂), 4.22 (broad s, 1H, H-3^{II}), 3.66 (dd, 1H, *J* = 2.0, 10.8 Hz, H-6^{II}), 3.57 (dd, 1H, *J* = 1.6, 10.0 Hz, H-4^{II}), 3.53 (d, 1H, *J* = 3.6 Hz, H-2^{II}), 3.48 (broad d, 1H, *J* = 10.4 Hz, H-6^{II}). ¹³C-NMR (100 MHz, CDCl₃) δ 166.1 (CO), 165.9 (CO), 165.6 (CO), 165.5 (CO), 139.7–127.4 (aromatic *C*), 104.1 (C-2^{II}), 90.4 (C-1^{II}), 79.4 (C-5^I), 76.9 (C-2^{II}), 74.6 (C-4^{II}), 73.5 (PhCH₂), 72.2 (PhCH₂, C-3^I), 71.6 (PhCH₂), 71.0 (C-3^{II}, PhCH₂), 70.1 (C-4^I), 68.5 (C-6^{II}), 67.3 (C-5^{II}), 64.7 (C-1^I), 63.4 (C-6^I).

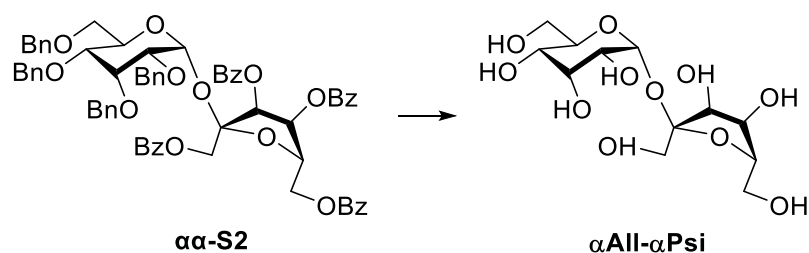
$\alpha\beta$ -**S2**; ¹H-NMR (400 MHz, CDCl₃) δ 8.13–6.97 (aromatic *H*), 6.24 (d, 1H, *J* = 5.2 Hz, H-3^I), 5.95 (dd, 1H, *J* = 5.2, 6.4 Hz, H-4^I), 5.58 (d, 1H, *J* = 3.6 Hz, H-1^{II}), 4.95 (d, 1H, *J* = 12.0 Hz, CH₂), 4.81–4.67 (m, 3H, H-5^I, H-6^I, H-6^I), 4.14 (t, 1H, *J* = 2.4 Hz, H-3^{II}), 3.80 (dd, 1H, *J* = 3.2, 10.4 Hz, H-6^{II}), 3.76 (dd, *J* = 2.0, 10.4 Hz, H-6^{II}), 3.53 (dd, 1H, *J* = 1.6, 10.4 Hz, H-4^{II}), 3.36 (dd, 1H, *J* = 3.6, 4.8 Hz, H-2^{II}). ¹³C-NMR (100 MHz, CDCl₃) δ 166.0–164.5 (CO), 138.7–126.8 (aromatic *C*), 107.3 (C-2^I), 90.9 (C-1^{II}), 79.7 (C-5^I), 75.6 (C-2^{II}), 74.9 (C-3^I), 74.4 (C-4^{II}), 74.3–72.8 (PhCH₂, PhCH₂, PhCH₂), 72.2 (C-4^I), 71.0 (C-3^{II}), 70.8 (PhCH₂), 67.6 (C-5^{II}), 65.5 (C-1^I), 62.8 (C-6^I).

$\beta\alpha$ -**S2**; ¹H-NMR (400 MHz, CDCl₃) δ 8.13–6.97 (aromatic *H*), 5.81 (d, 1H, *J* = 6.4 Hz, H-3^I), 5.79 (dd, 1H, *J* = 2.8, 6.4 Hz, H-4^I), 5.53 (d, 1H, *J* = 8.4 Hz, H-1^{II}), 5.19 (broad q, 1H, *J* = 2.8, 5.6 Hz, H-5^I), 4.72–4.27 (m, 4H, H-1^I, H-1^I, H-6^I, H-6^I), 4.16 (ddd, 1H, *J* = 1.6, 6.4, 8.0, 9.6 Hz, H-5^{II}), 4.05 (broad t, 1H, *J* = 2.4 Hz, H-3^{II}), 3.62 (dd, 1H, *J* = 1.6, 10.4 Hz, H-6^{II}), 3.28 (dd, 1H, *J* = 6.4, 10.8 Hz, H-6^{II}), 3.27–3.22 (m, 2H, H-2^{II}, H-4^{II}). ¹³C-NMR (100 MHz, CDCl₃) δ 166.0–164.5 (CO), 138.7–126.8 (aromatic *C*), 104.2 (C-2^I), 94.8 (C-1^{II}), 81.6 (C-5^I), 78.4 (C-2^{II}), 75.5 (C-4^{II}), 74.7 (C-3^{II}), 74.8 (C-3^I), 74.3–72.8 (PhCH₂, PhCH₂, PhCH₂), 72.0 (C-5^{II}), 71.2 (PhCH₂), 71.1 (C-4^I), 69.9 (C-6^{II}), 64.3 (C-1^I), 62.9 (C-6^I).

$\beta\beta$ -**S2**; ^1H -NMR (400 MHz, CDCl_3) δ 8.00–7.00 (aromatic H), 6.13 (d, 1H, $J = 4.4$ Hz, H-3^{I}), 5.96 (dd, $J = 4.8$ Hz, 8.8 Hz, H-4^{I}), 5.56 (d, $J = 8.0$ Hz, H-1^{II}), 4.86–4.37 (m, 4H, H-1^{I} , H-1^{I} , H-5^{I} , H-6^{I}), 4.08 (s, 1H, H-3^{II}), 4.02 (ddd, 1H, $J = 2.0, 5.0, 10.2$ Hz, H-5^{II}), 3.70 (dd, 1H, $J = 2.0, 11.2$ Hz, H-6^{II}), 3.57–3.53 (m, 2H, H-4^{II} , H-6^{II}), 3.38 (dd, $J = 2.4, 8.4$ Hz, H-2^{II}). ^{13}C -NMR (100 MHz, CDCl_3) δ 165.9 (CO), 165.7 (CO), 165.1 (CO), 164.6 (CO), 139.1–127.2 (aromatic C), 107.7 (C-2^{I}), 92.5 (C-1^{II}), 79.1 (C-5^{I}), 78.8 (C-2^{II}), 74.8 (C-3^{II} , C-4^{II}), 74.6 (C-3^{I} , PhCH_2), 73.4 (C-4^{I} , PhCH_2), 72.9 (C-5^{II}), 72.6 (PhCH_2), 71.8 (PhCH_2), 68.8 (C-6^{II}), 65.5 (C-1^{I}), 62.6 (C-6^{I}).

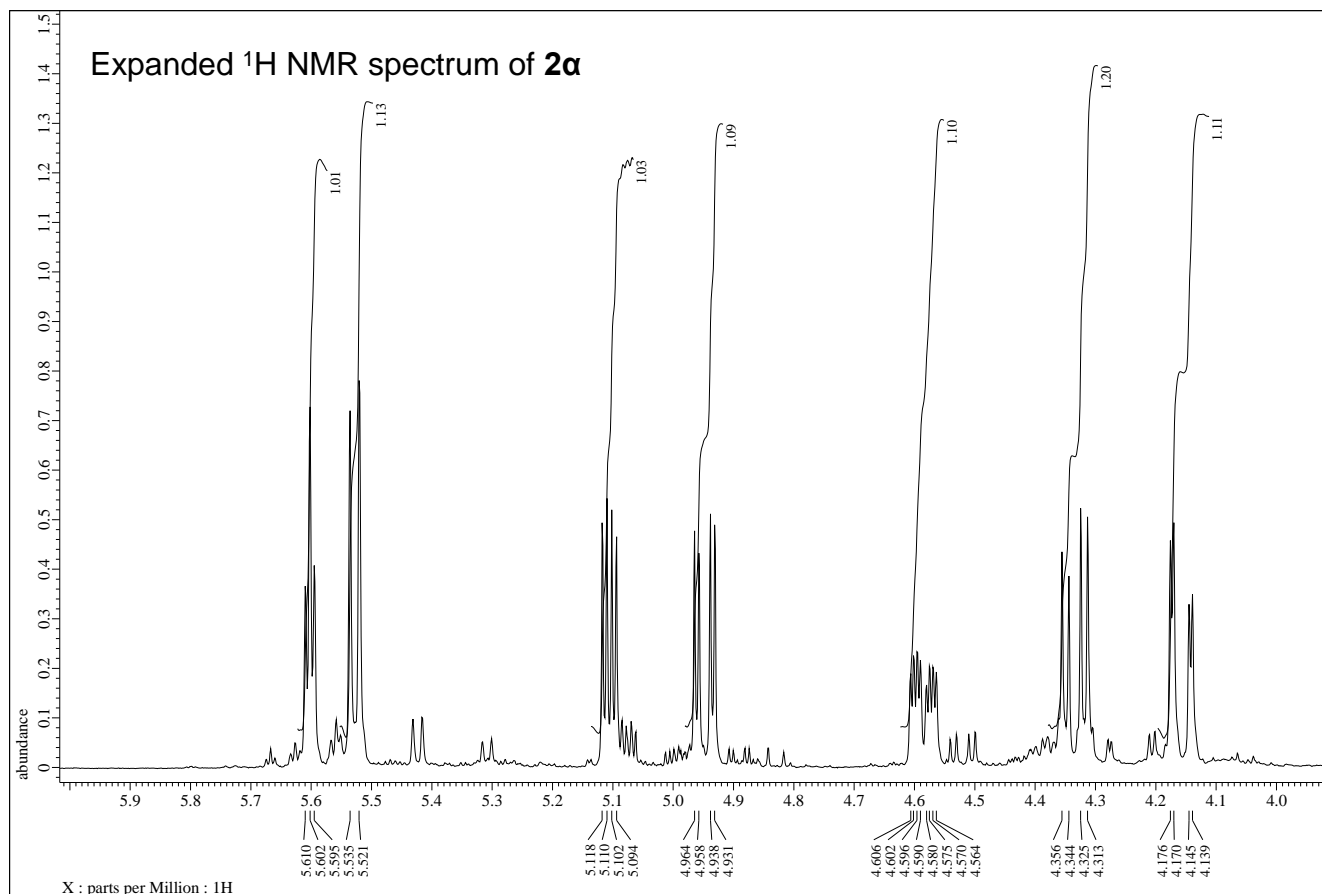
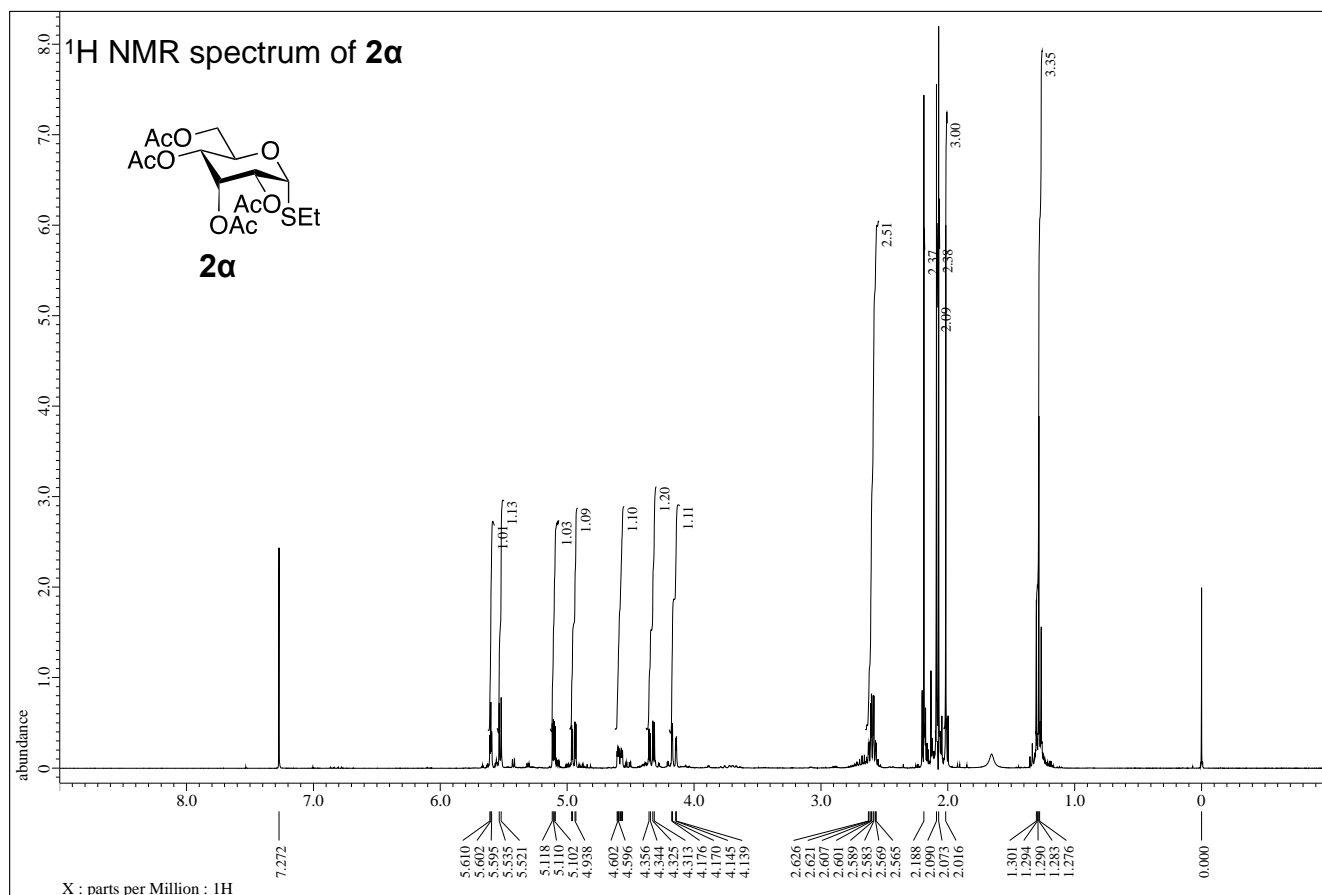
Table S1. ^{13}C -chemical shifts (ppm) and $^1J_{\text{C1-H1}}$ coupling constants (Hz) for stereoisomers of α/β -D-allopyranosyl-(1 \rightarrow 2)- α/β -D-psicofuranoside derivatives (CDCl_3).

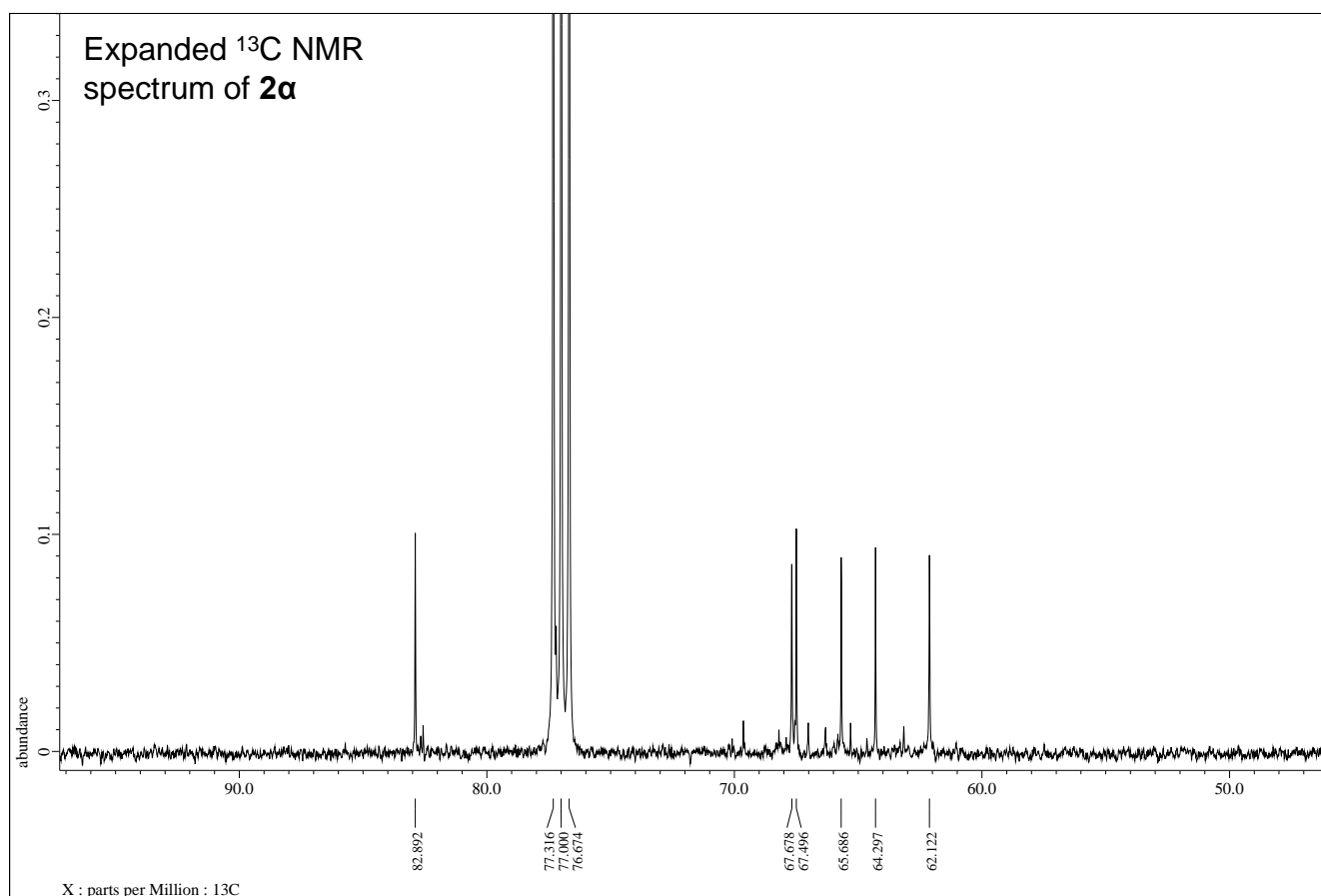
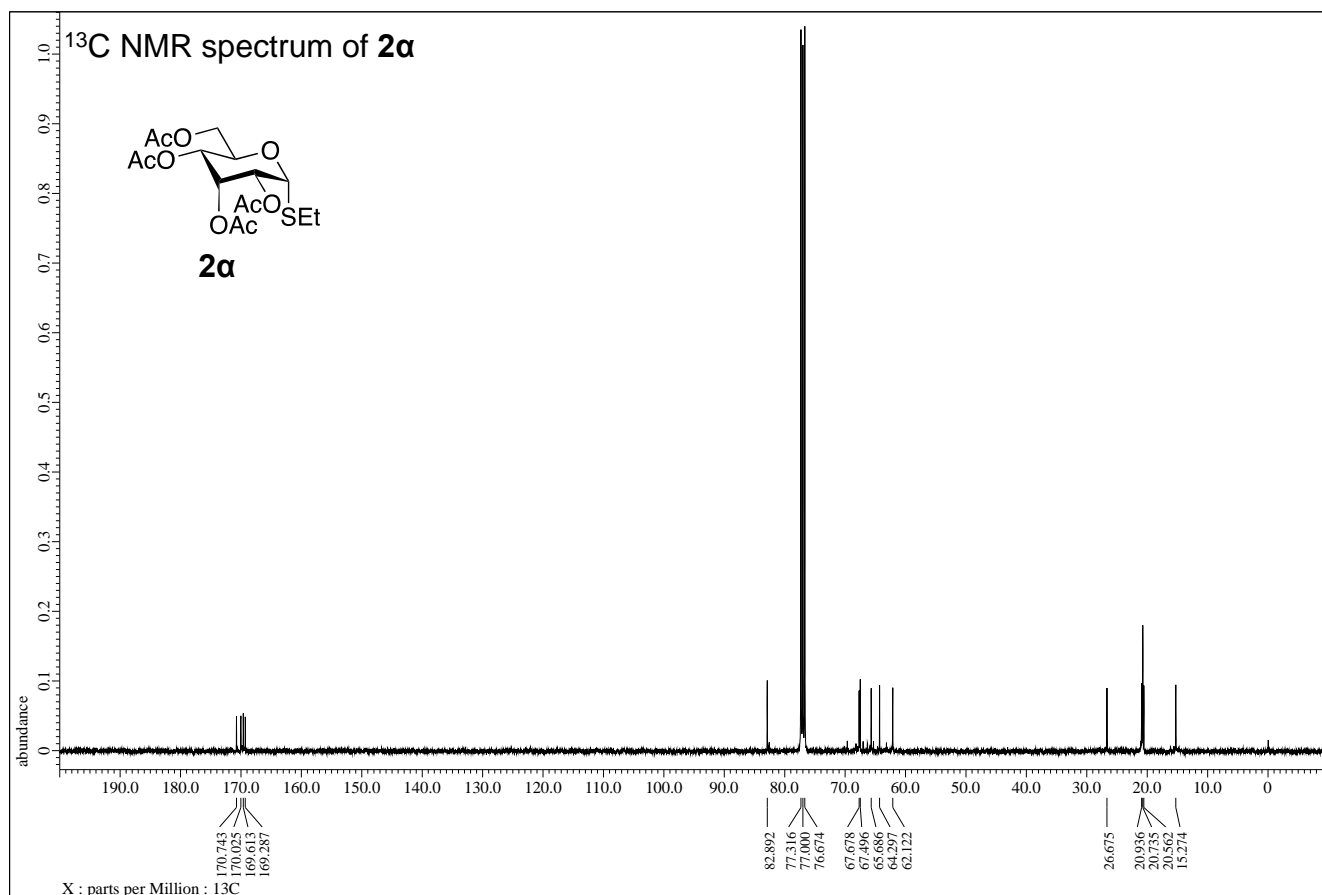
	$\alpha\alpha$ -S2	$\alpha\beta$ -S2	$\beta\alpha$ -S2	$\beta\beta$ -S2
δ_{C} D-allo 1	90.4	90.9	94.8	92.5
2	76.9	75.6	78.4	78.8
3	71.0	71.0	74.7	74.8
4	74.6	74.4	75.5	74.8
5	67.3	67.6	72.0	72.9
6	68.5	68.7	69.9	68.8
D-psico 1	64.7	65.5	64.3	65.5
2	104.1	107.3	104.2	107.7
3	72.2	74.9	74.8	74.6
4	70.1	72.2	71.1	73.4
5	79.4	79.7	81.6	79.1
6	63.4	62.8	62.9	62.6
$J_{\text{C1-H1}}$ (allo)	177.0	170.0	172.0	172–174

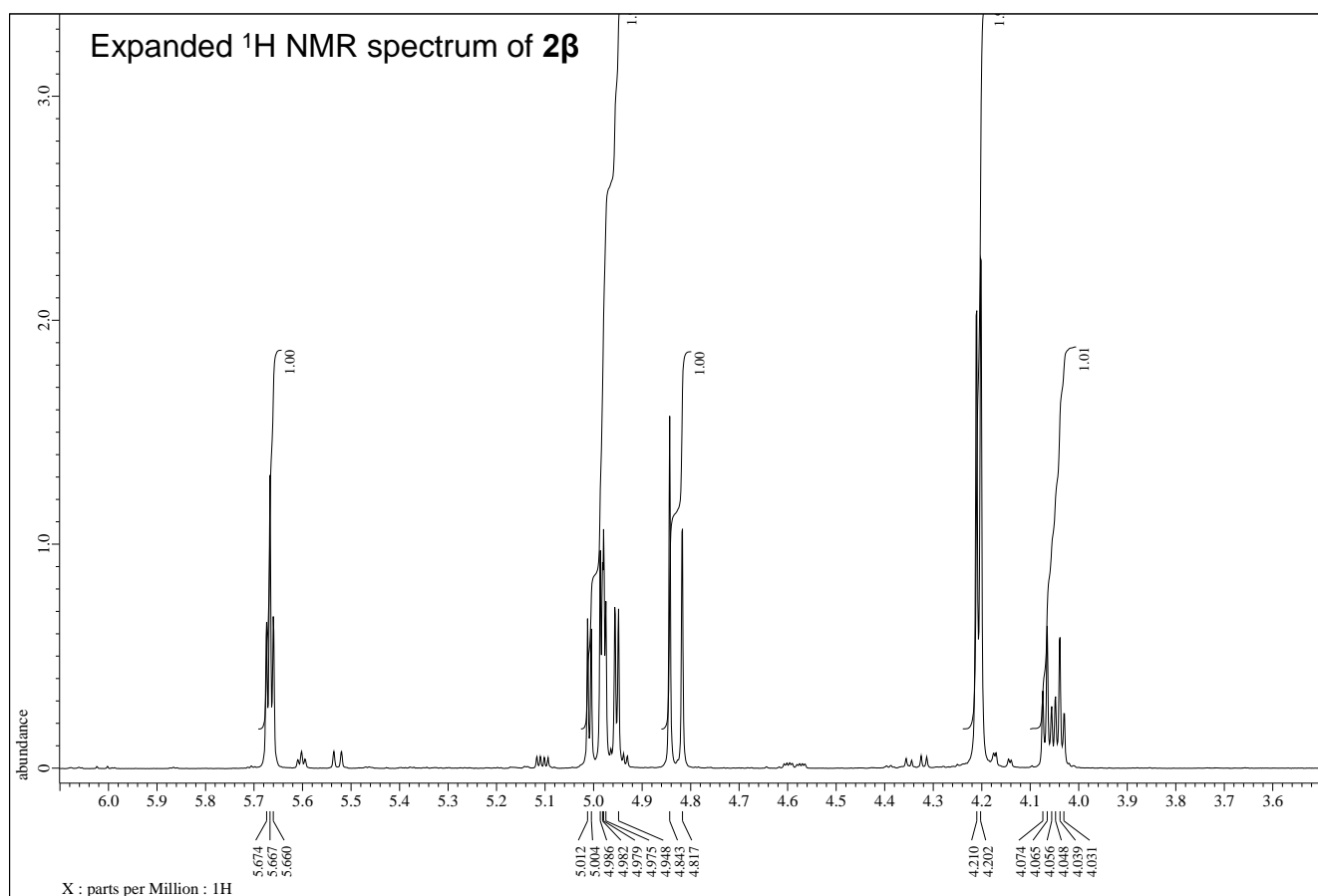
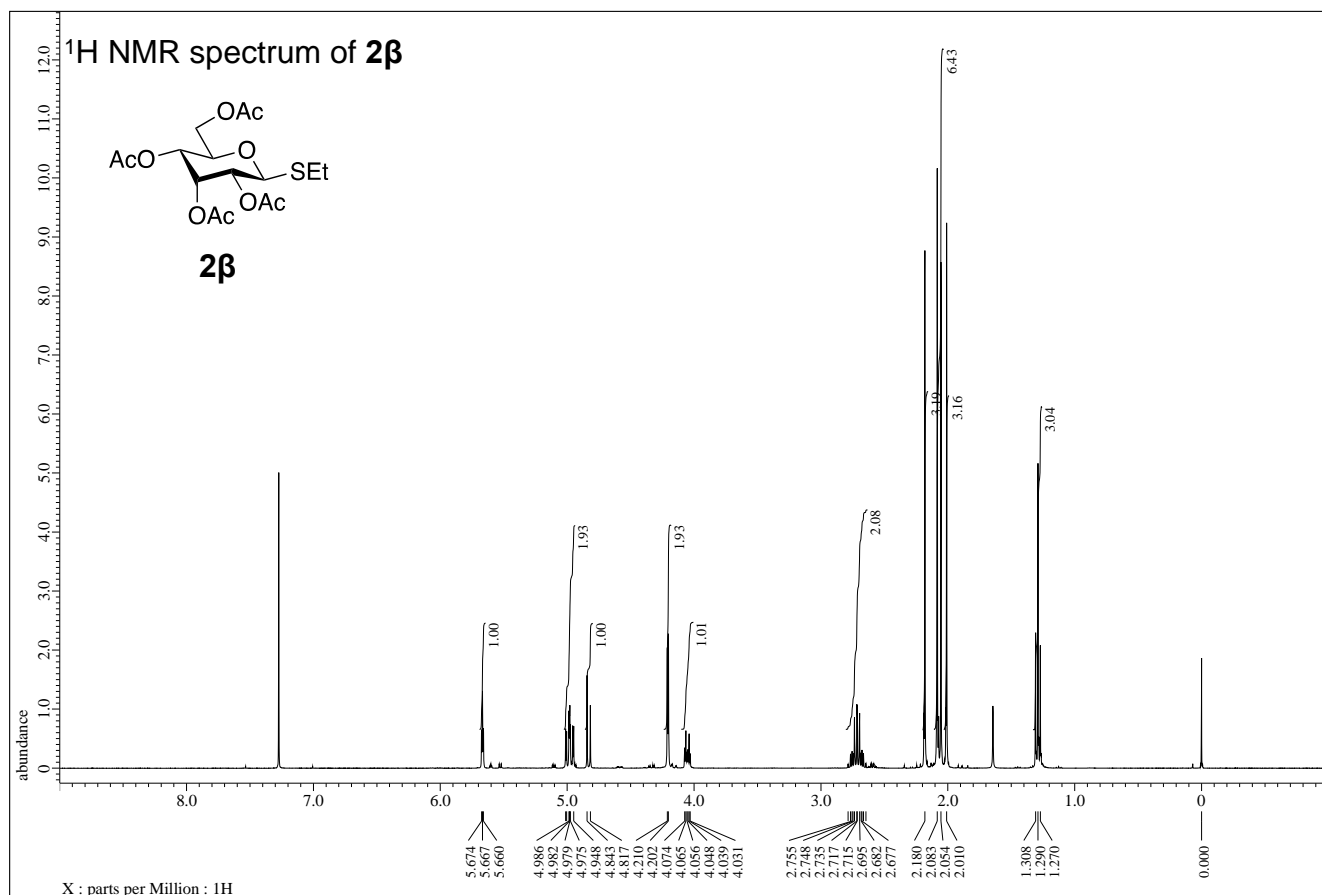


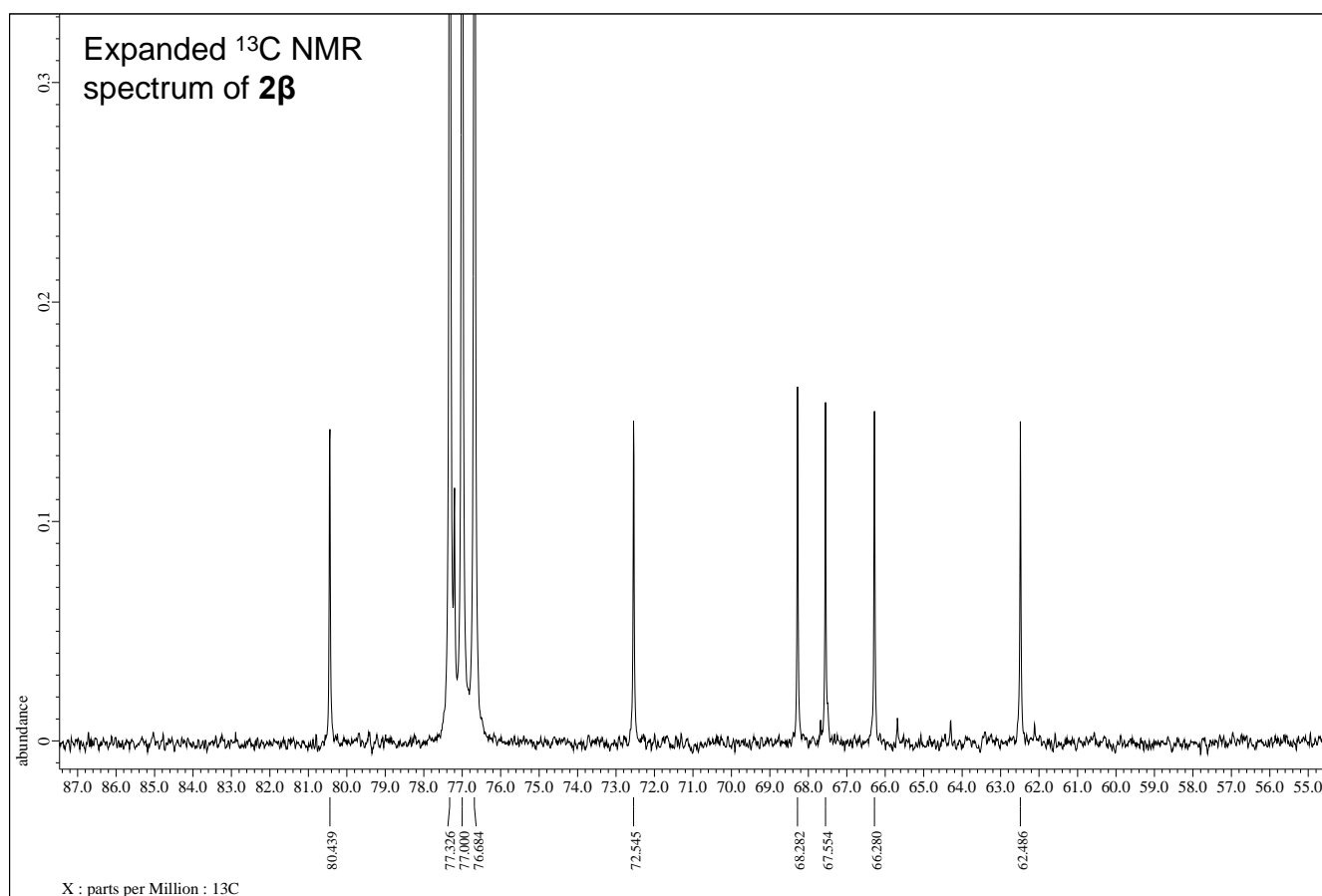
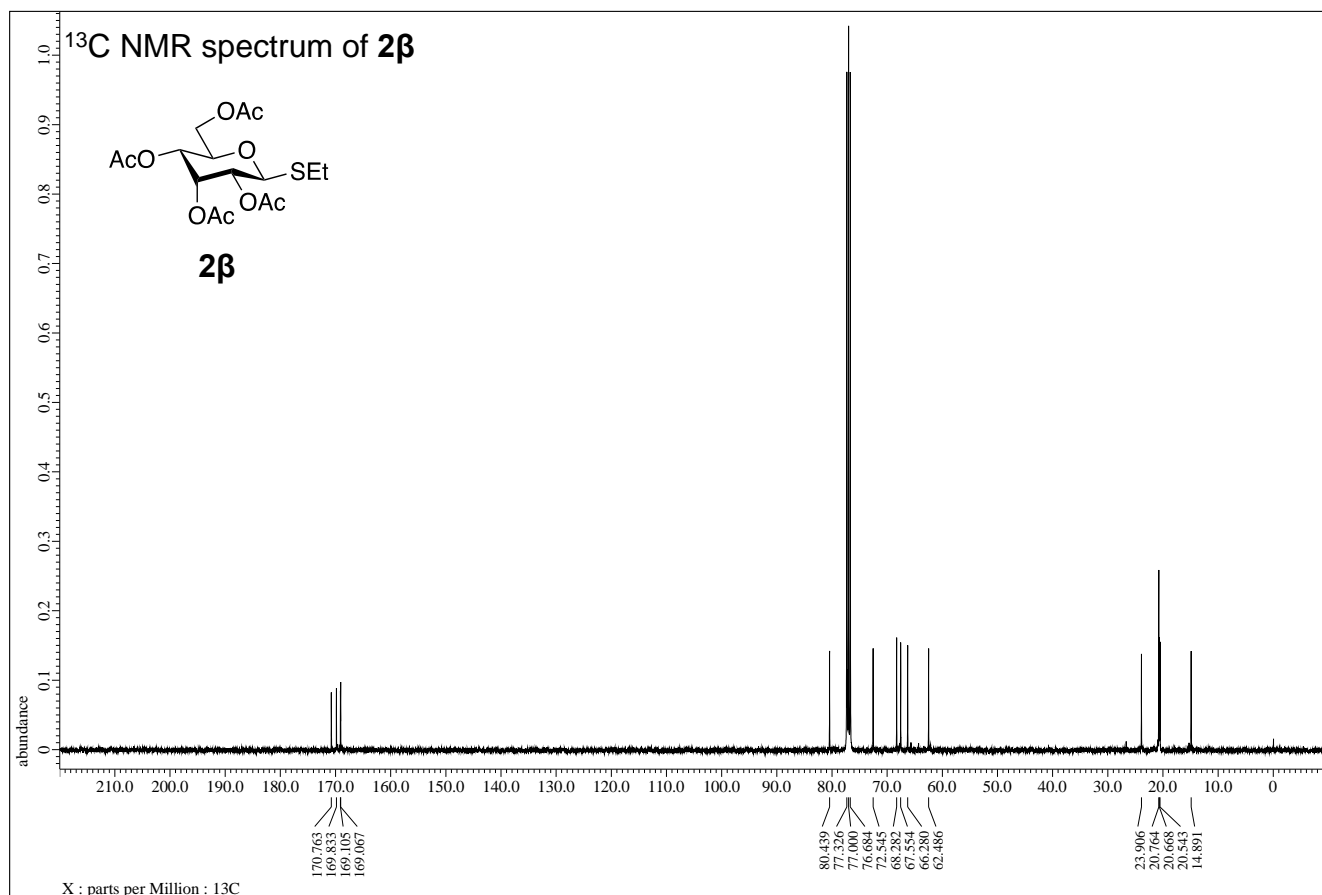
Scheme S2. Deprotection of α -D-allosyl-(1 \rightarrow 2)- α -D-psicofuranoside derivative.

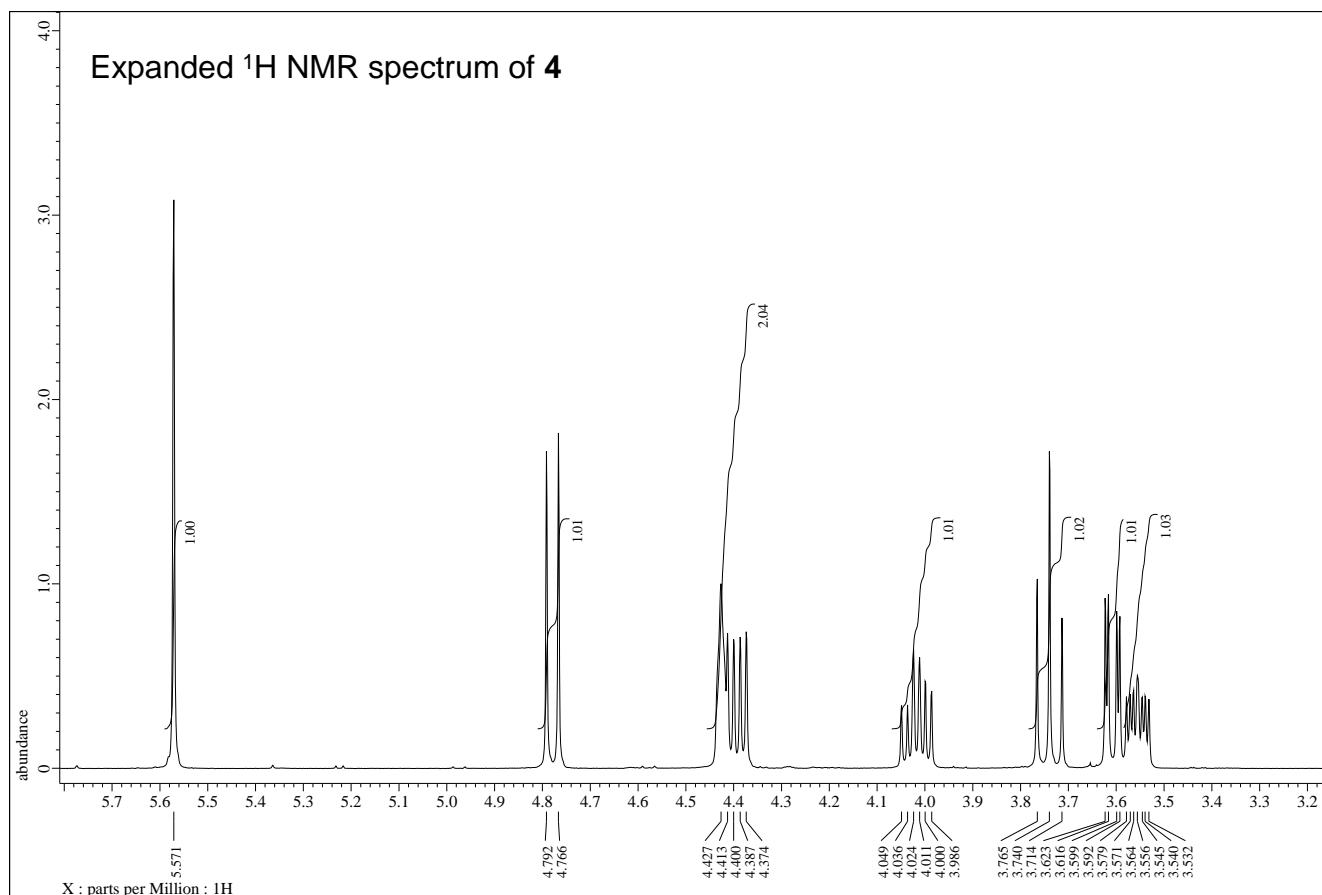
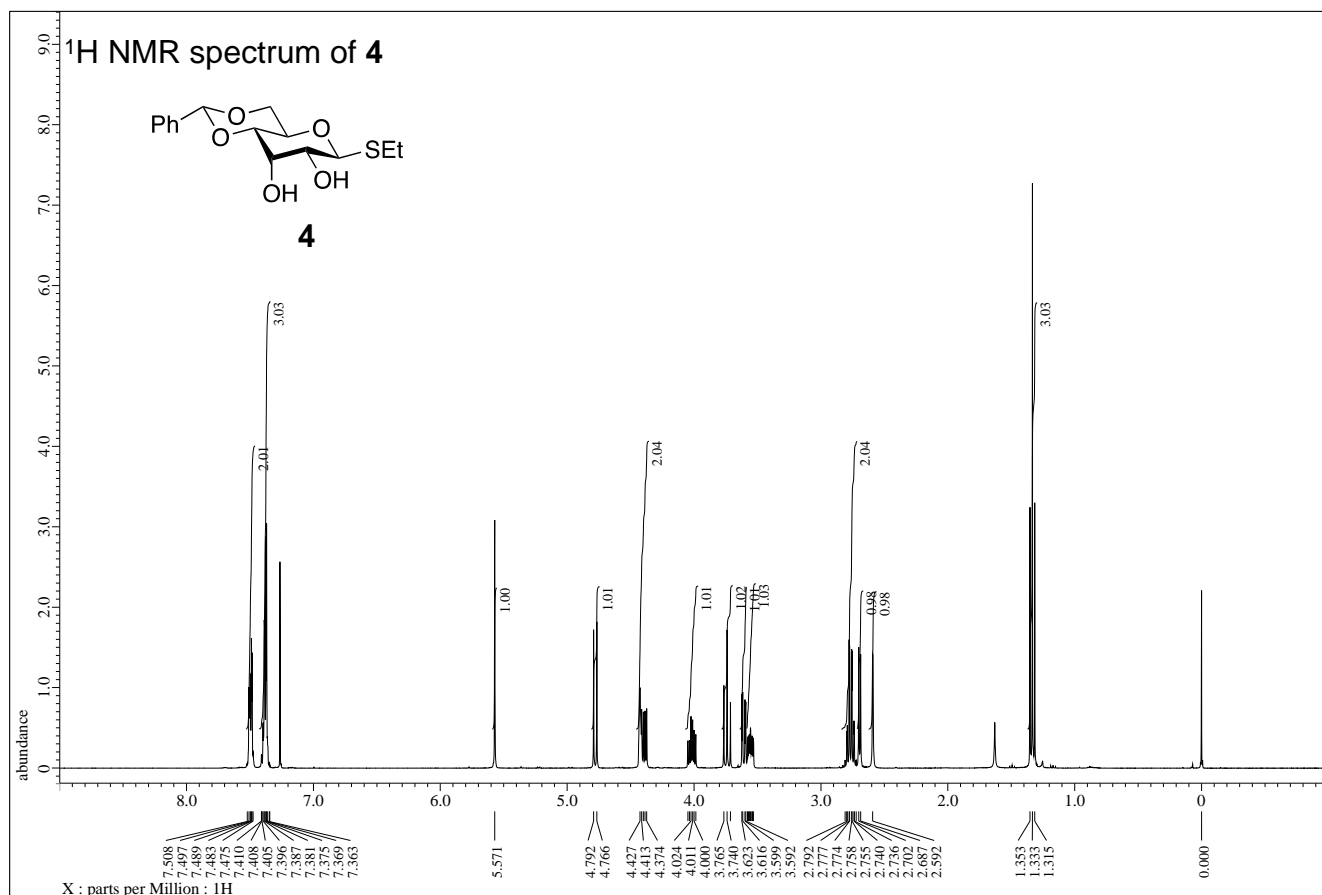
To a solution of the $\alpha\text{-S2}$ (29 mg, 0.026 mmol) in MeOH (1.0 mL) and THF (1.0 mL) was added 1 M NaOMe in MeOH (26 μ L) at 0 °C. The reaction mixture was stirred at room temperature for 12 hours, and neutralized with Amberlyst, filtered, and concentrated *in vacuo*. The resulting residue was purified by flash silica gel column chromatography with chloroform–MeOH (85/15, *v/v*) to afford intermediate (6.9 mg, 0.019 mmol, 73%). To a solution of the intermediate in H₂O (5.0 mL) was stirred in the presence of Pd(OH)₂/C (15 mg) at 40 °C under H₂ atmosphere for 1 day. The reaction mixture was filtered through celite. The filtrate was lyophilized. The residue was purified by C18 reversed-phase column chromatography (H₂O) to give deprotected product (6.9 mg, 0.019 mmol, 73%). ¹H-NMR (400 MHz, CDCl₃) δ 5.36 (d, 1H, J = 4.0 Hz, H-1^{II}), 4.48 (d, 1H, J = 6.4 Hz, H-3^I), 4.21–4.19 (m, 1H, H-5^I), 4.09 (t, 1H, J = 6.8 Hz, H-3^{II}), 4.05 (dd, 1H, J = 1.2, 6.0 Hz, H-4^I), 3.88 (td, 1H, J = 2.5, 5.6, 10.4 Hz, H-5^{II}), 3.91–3.82 (m, 2H, H-1^I, H-6^{II}), 3.77 (t, 1H, J = 3.6 Hz, H-2^{II}), 3.73–3.63 (m, 3H, H-1^I, H-6^I, H-6^{II}), 3.58 (dd, 1H, J = 2.8, 10.8 Hz, H-4^{II}). ¹³C-NMR (100 MHz, CDCl₃) δ 106.2 (C-2^I), 90.4 (C-1^I), 85.4 (C-5^I), 70.8 (C-3^I, C-3^{II}), 70.7 (C-4^I), 67.5 (C-5^{II}), 66.6 (C-2^{II}), 66.0 (C-4^{II}), 61.6 (C-6^I), 60.7 (C-1^I), 60.2 (C-6^{II}).

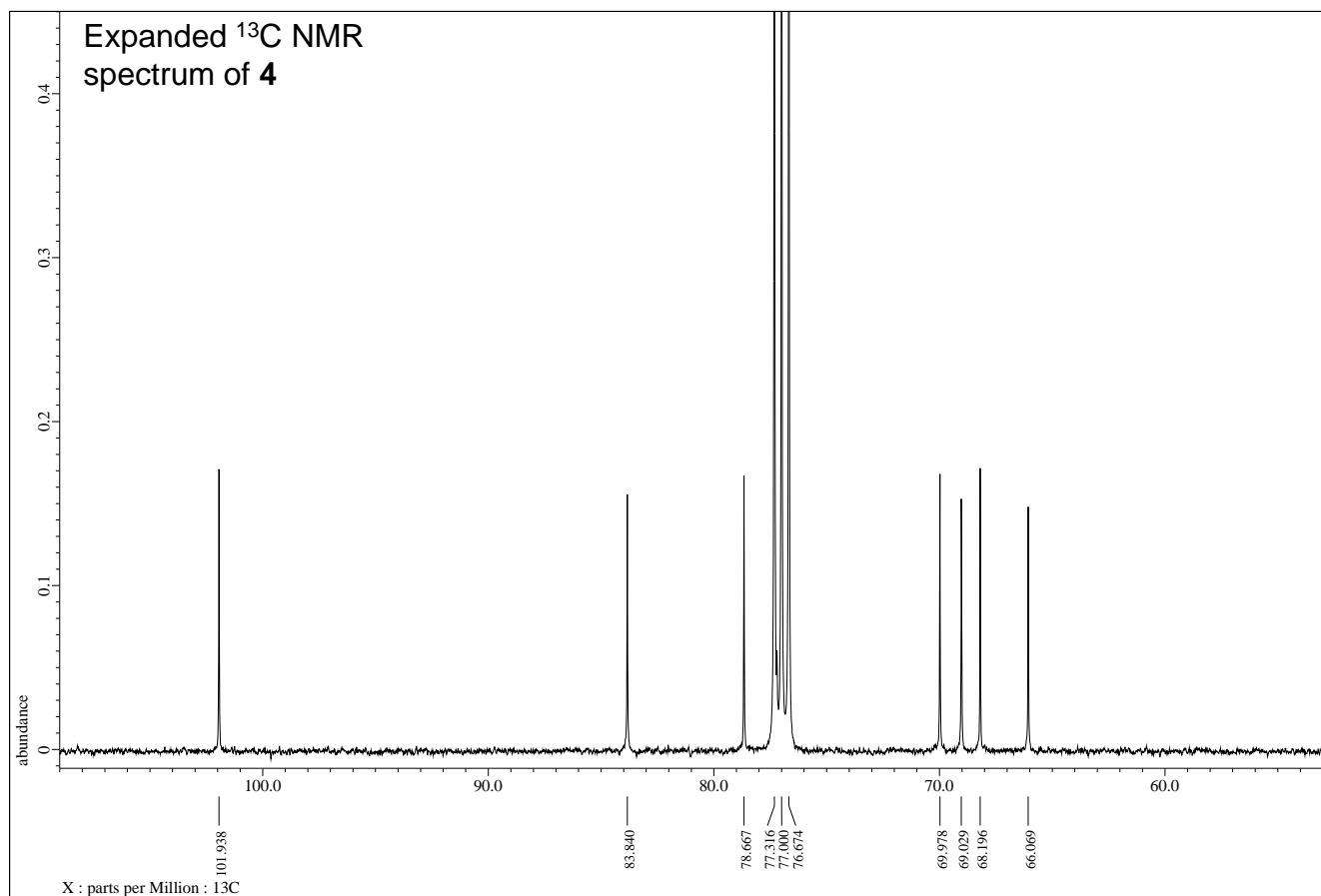
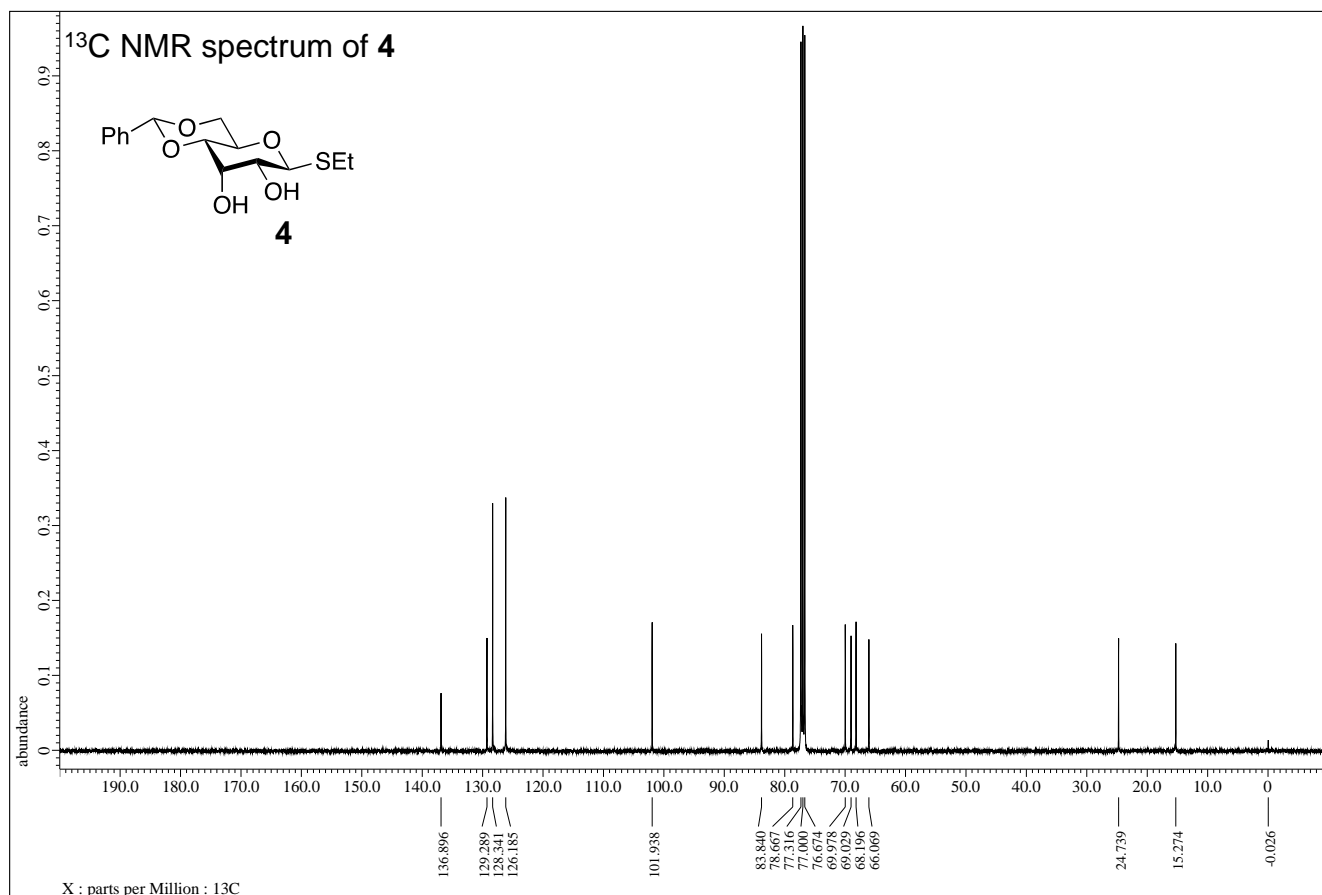


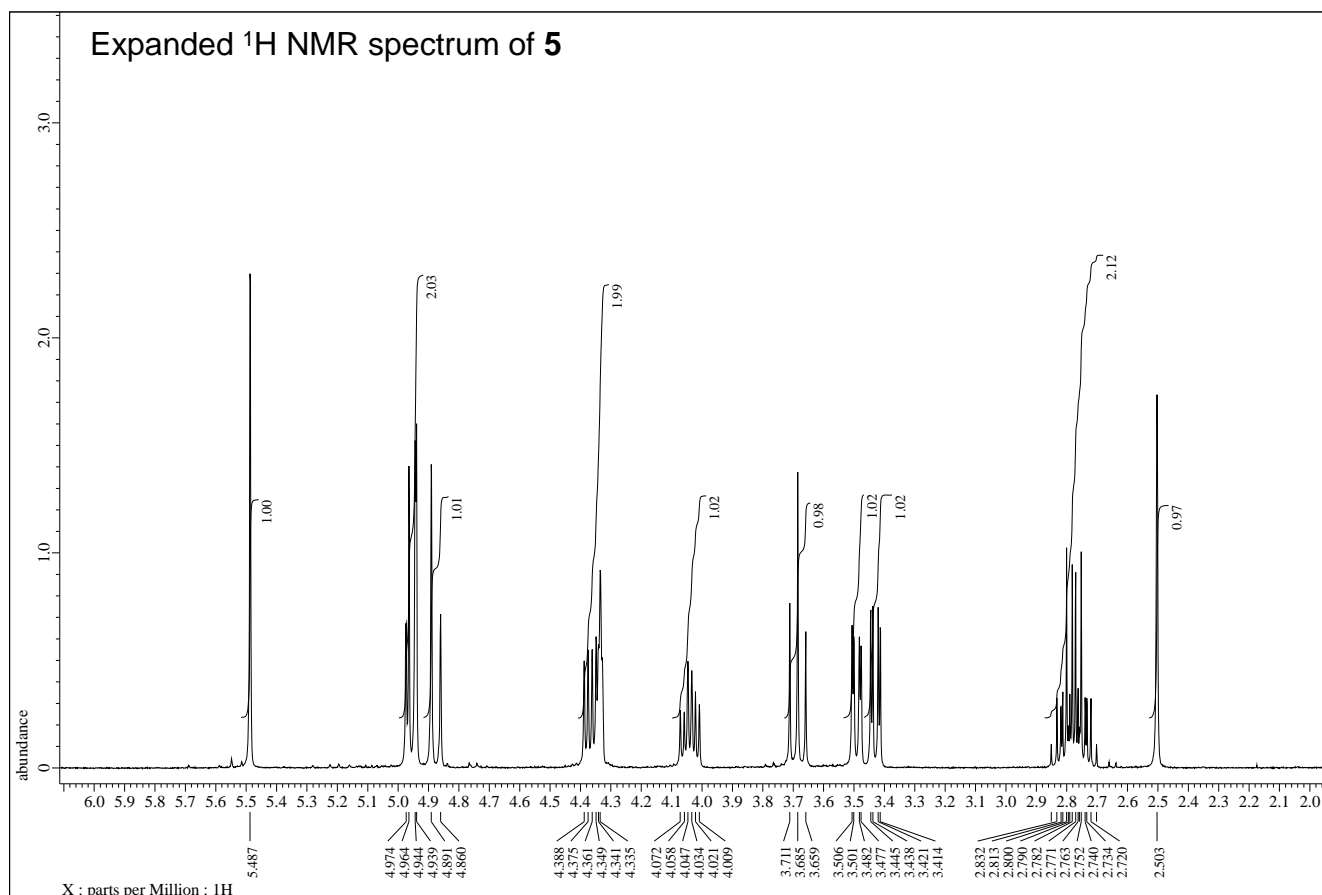
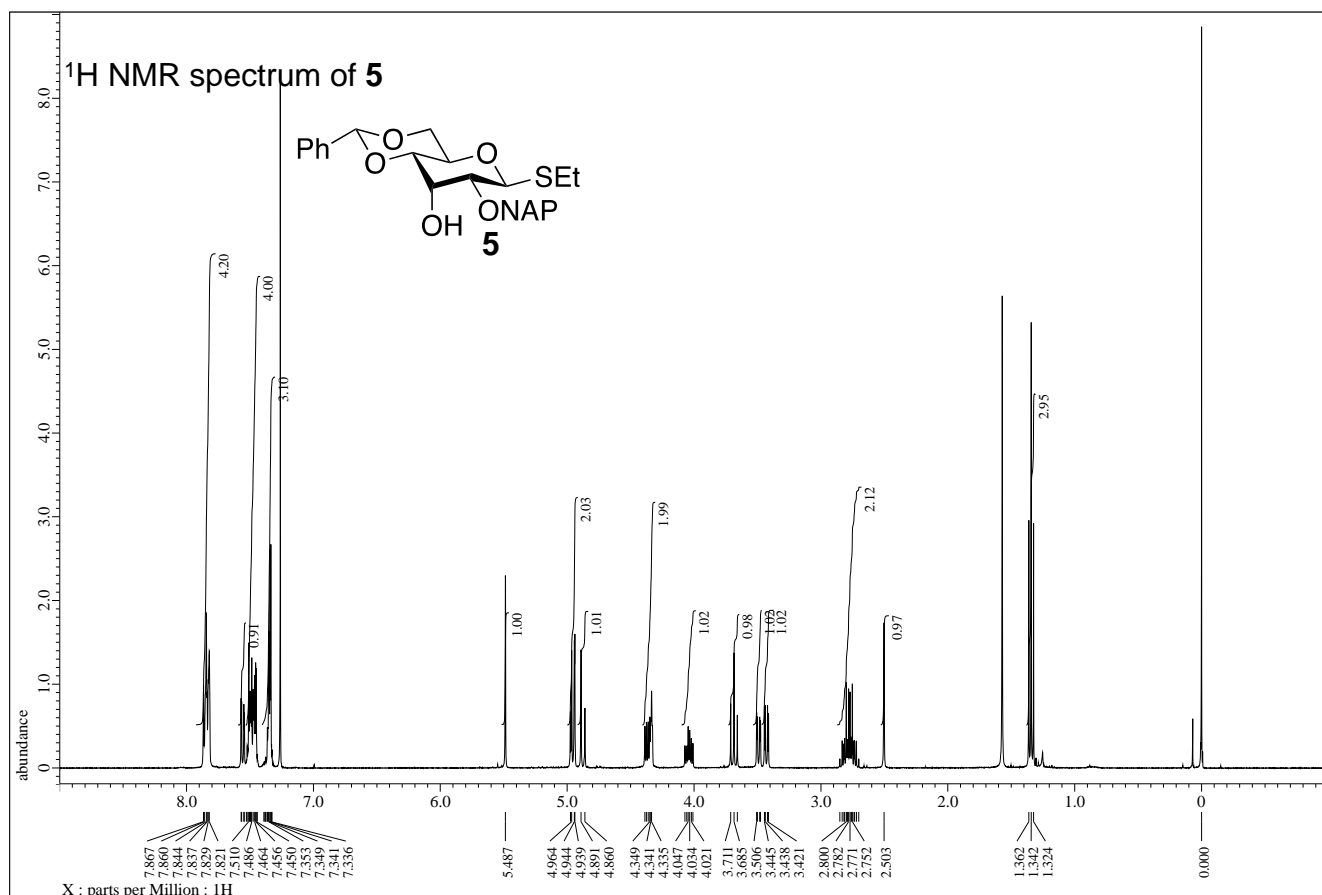


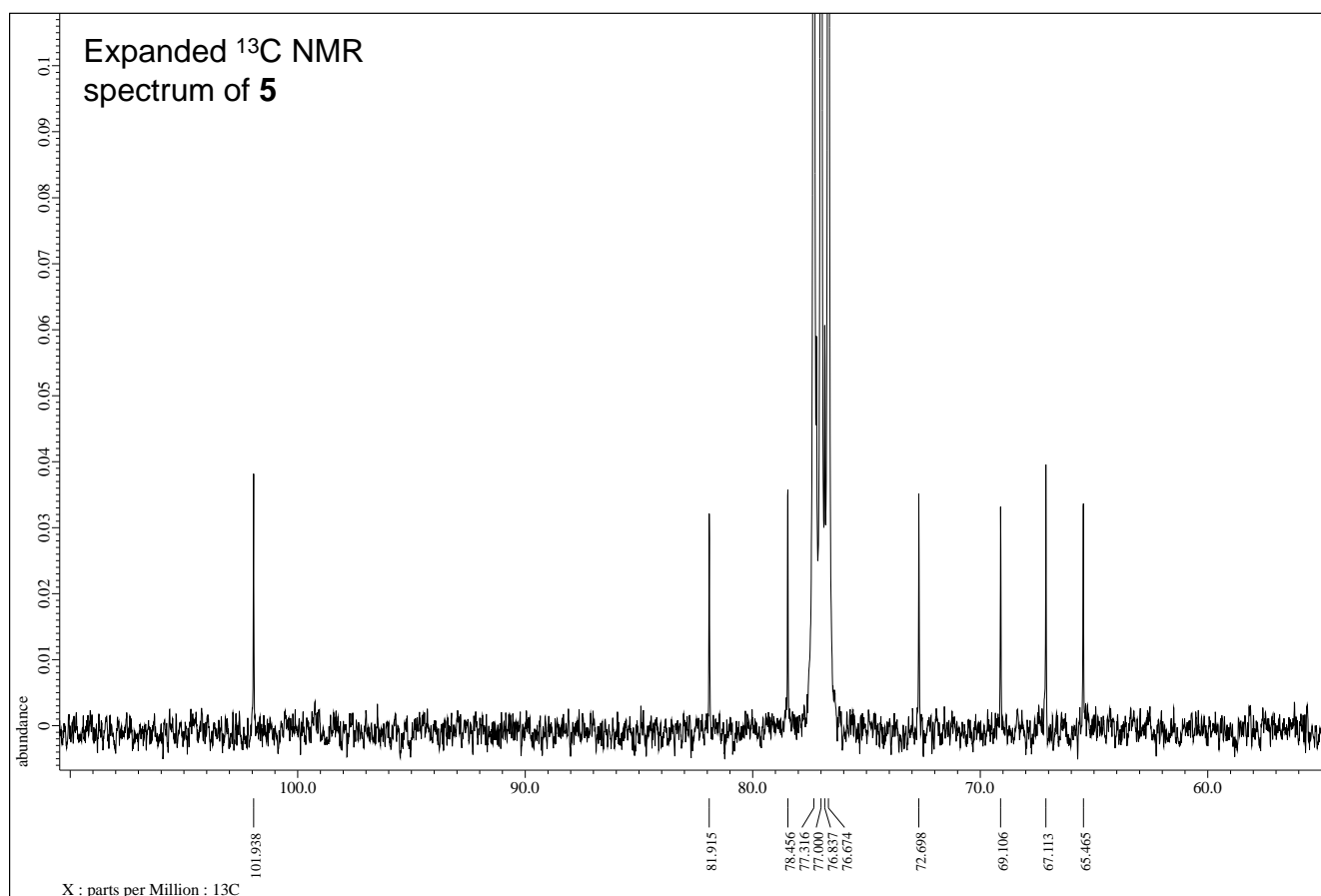
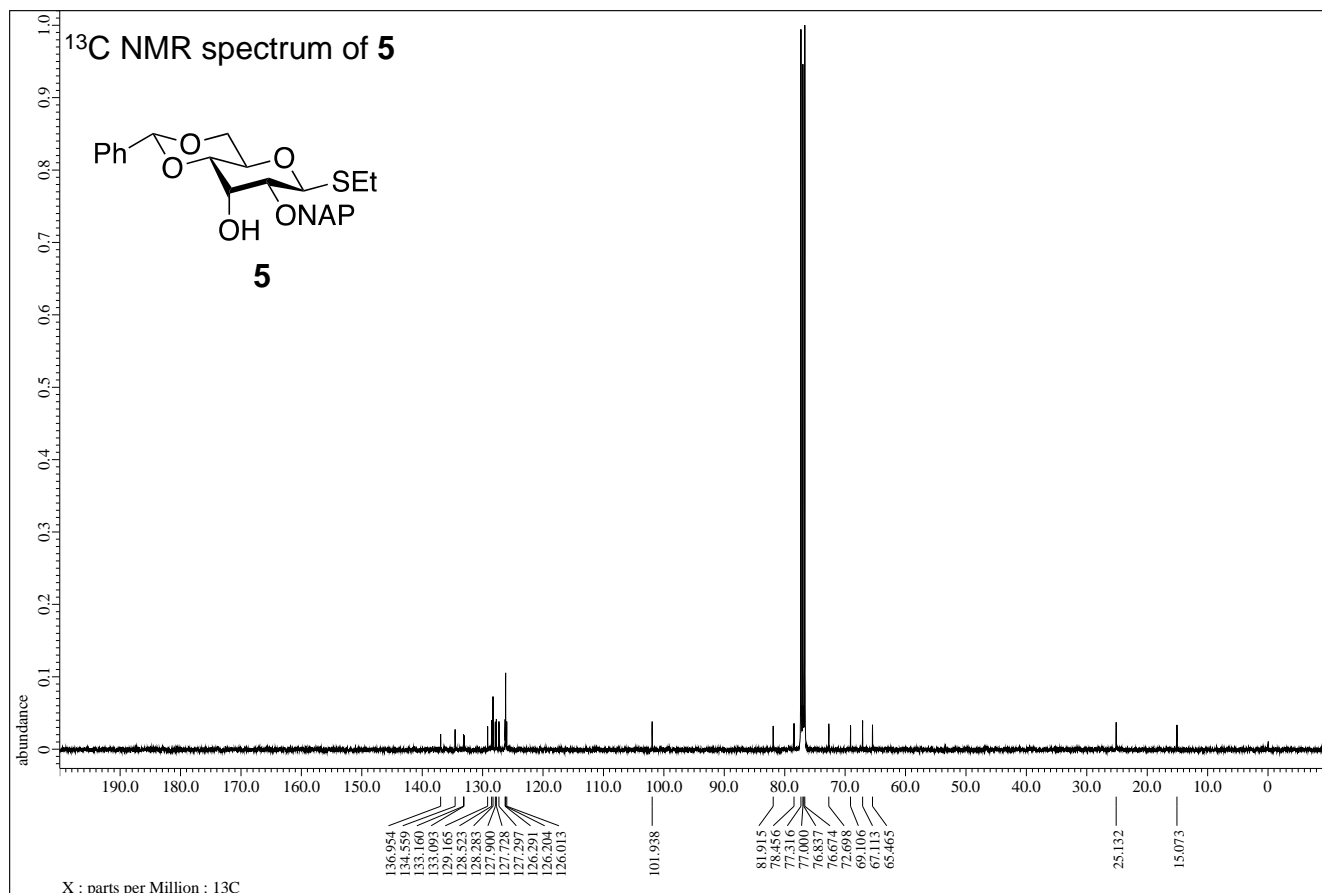


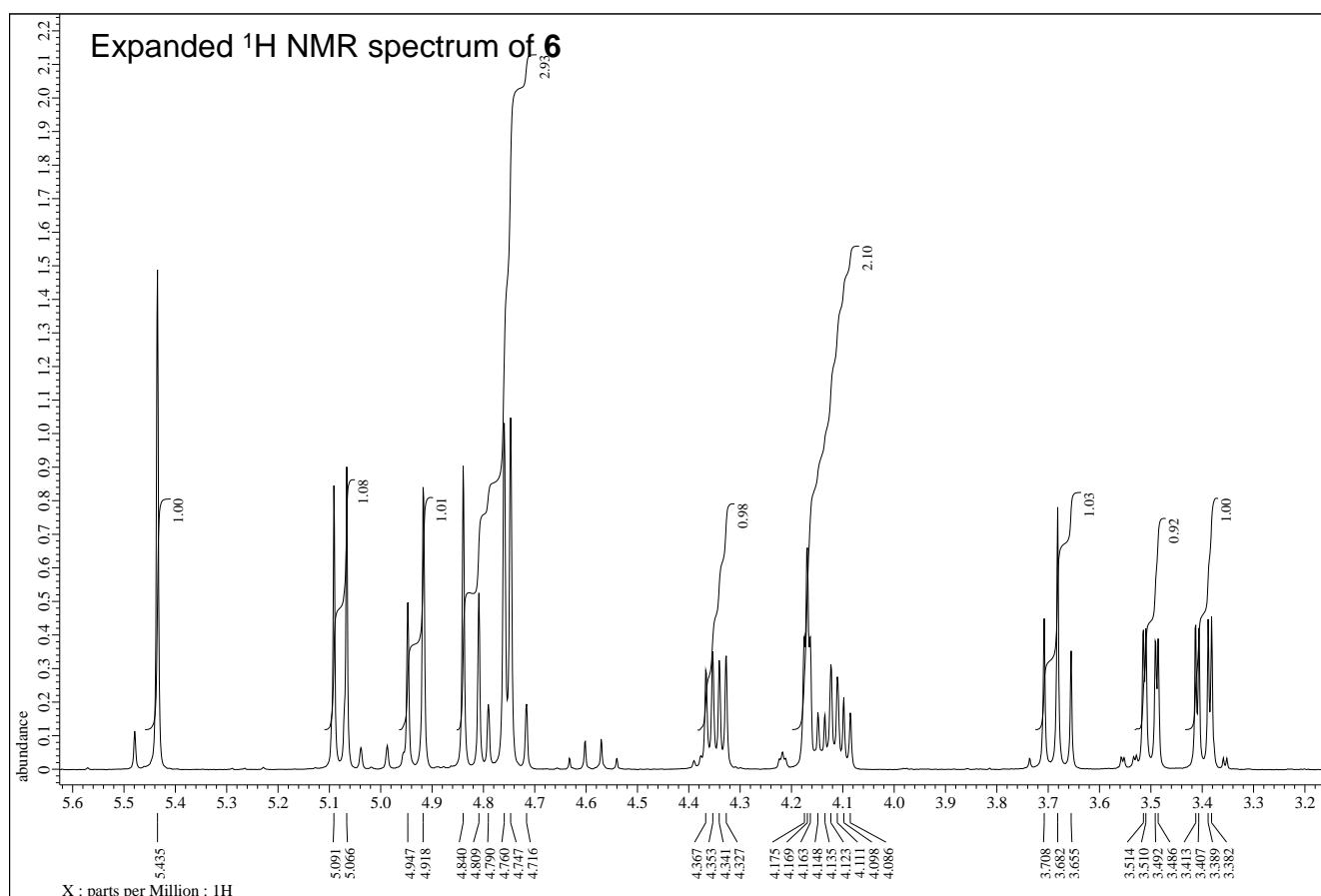
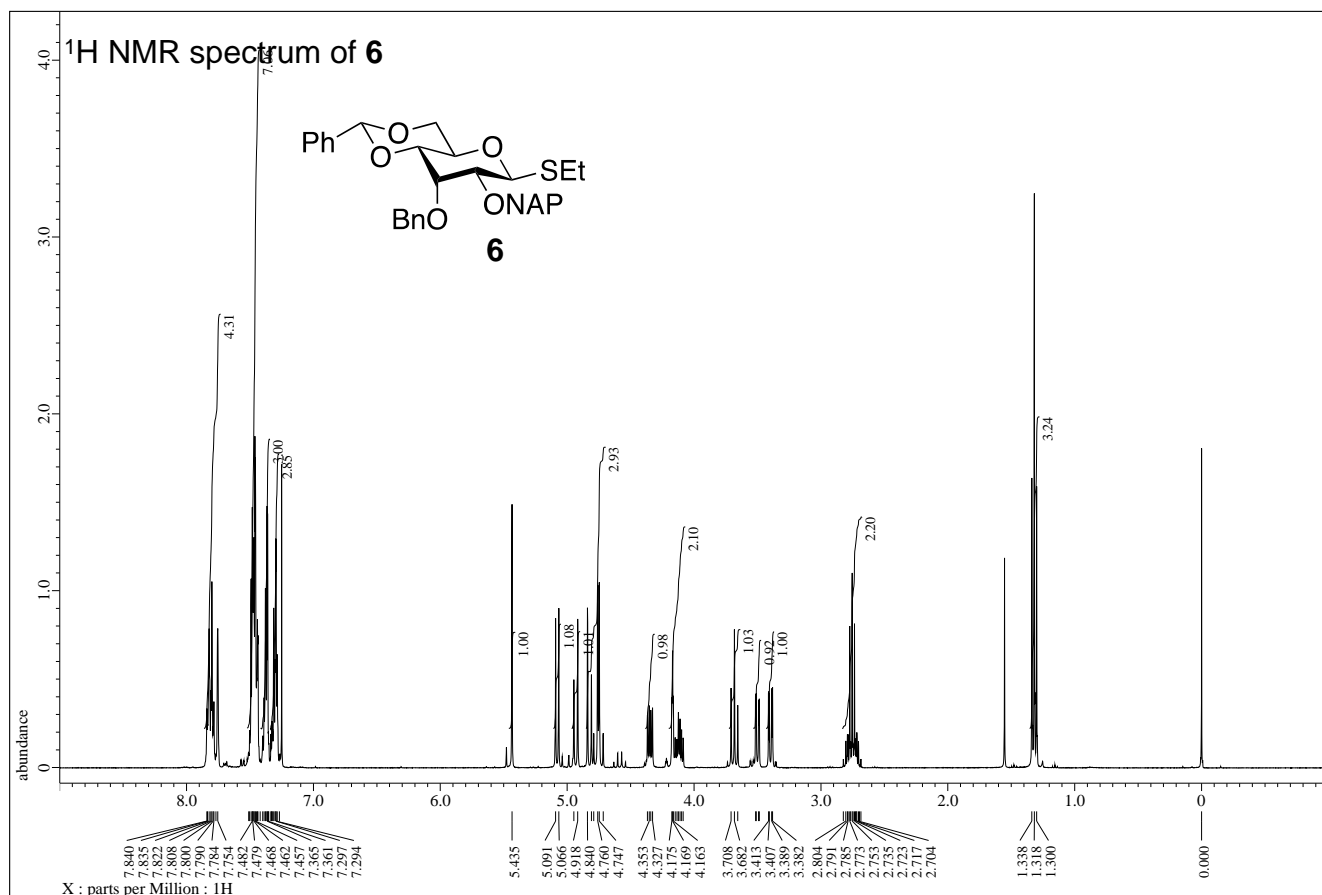


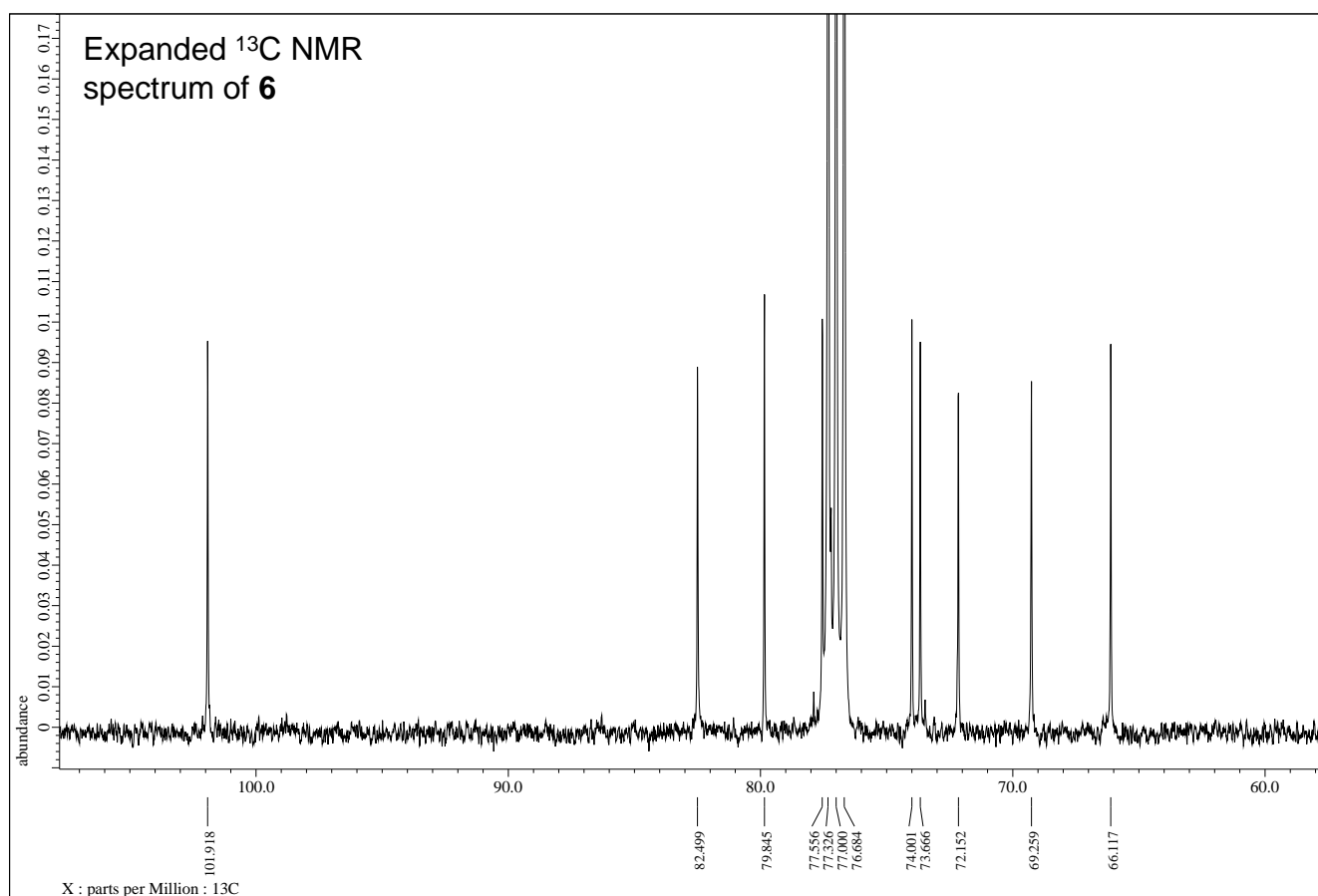
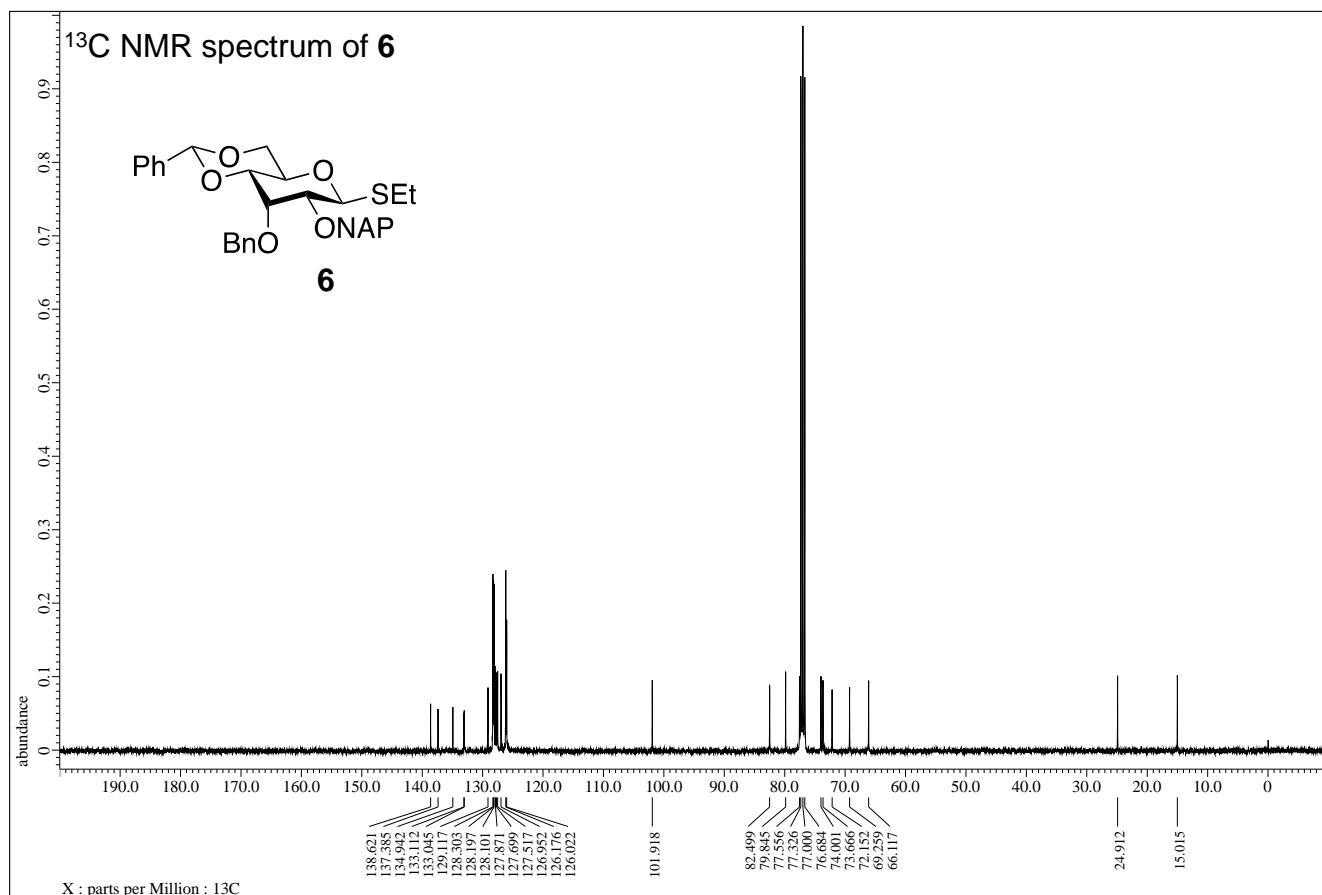


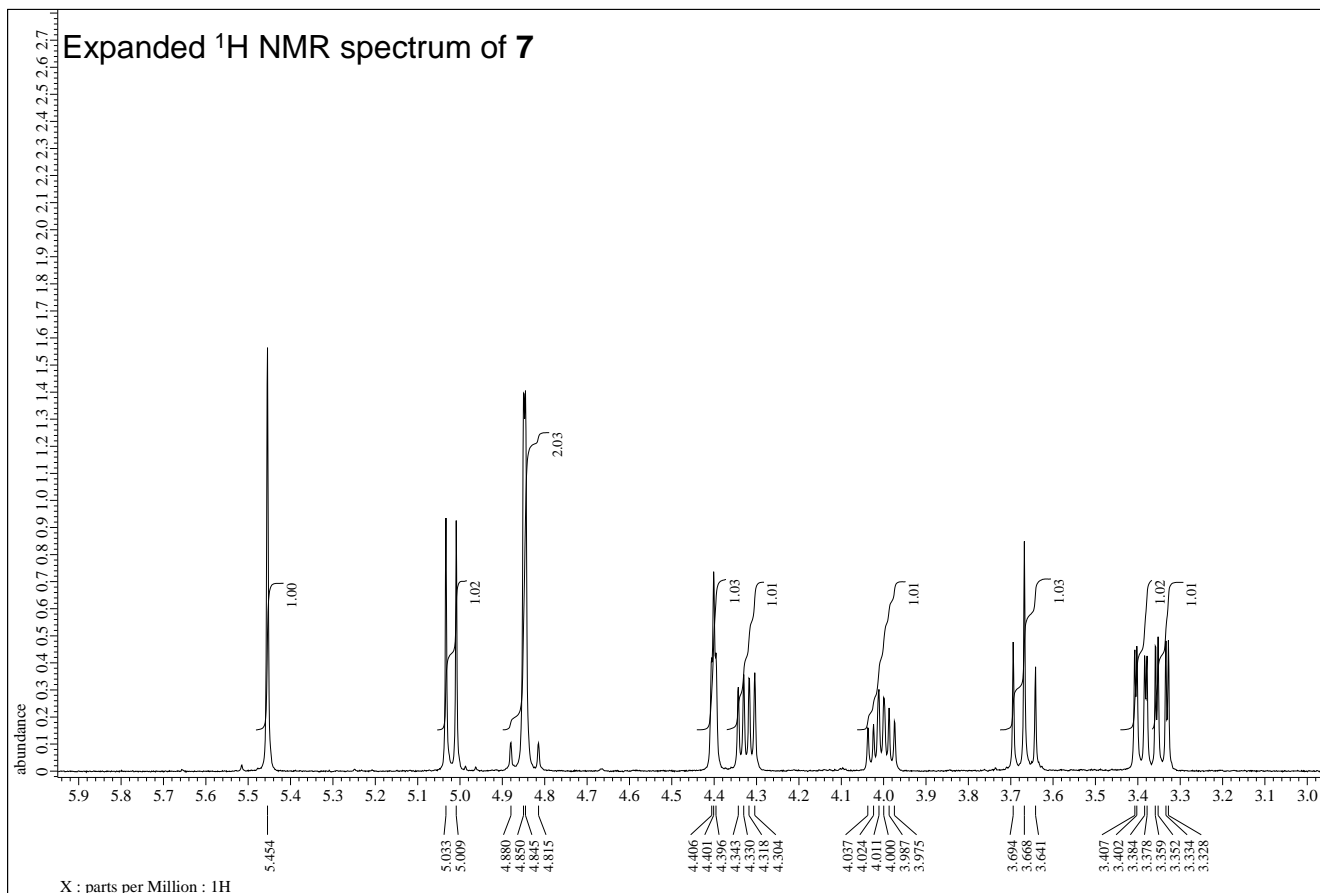
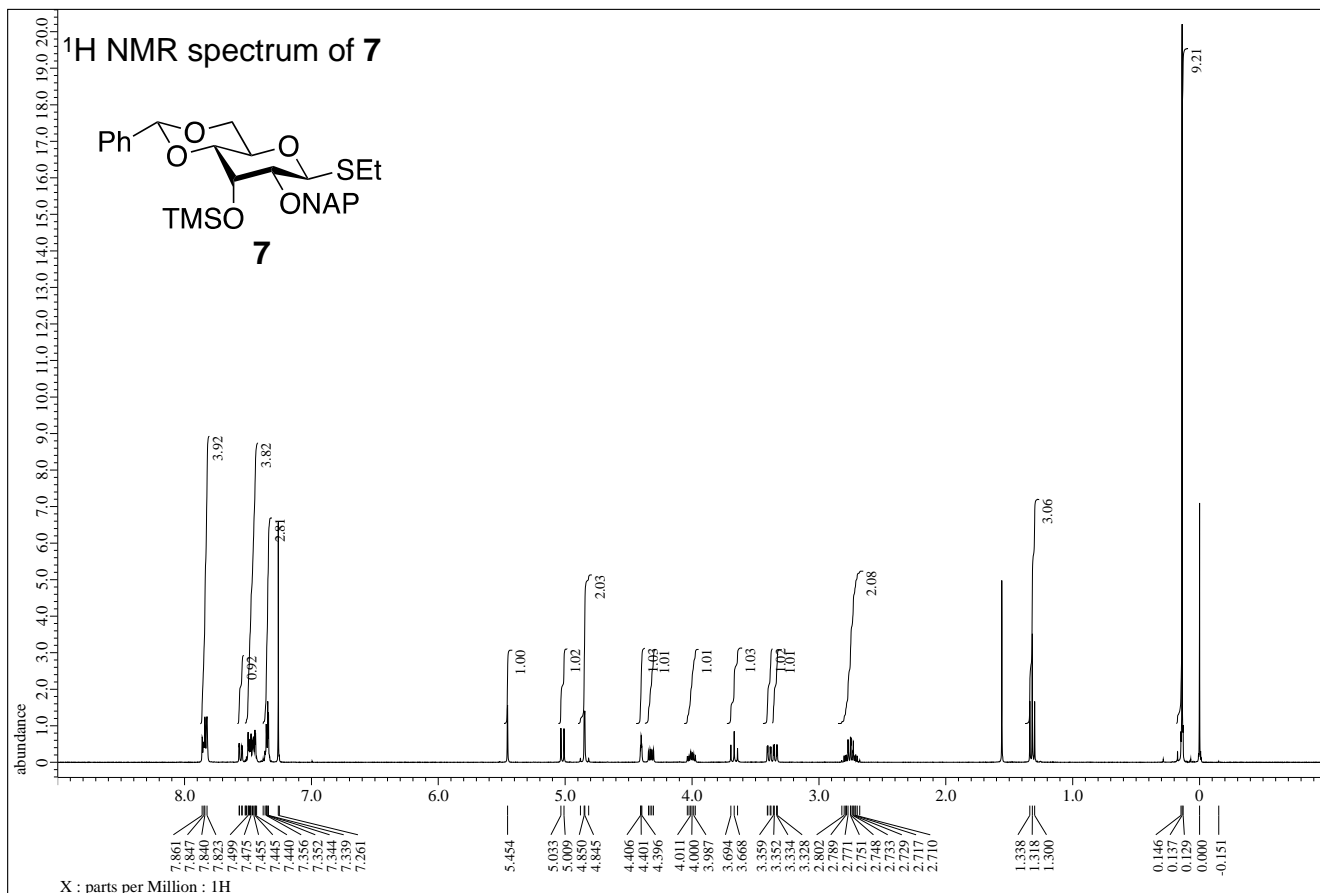


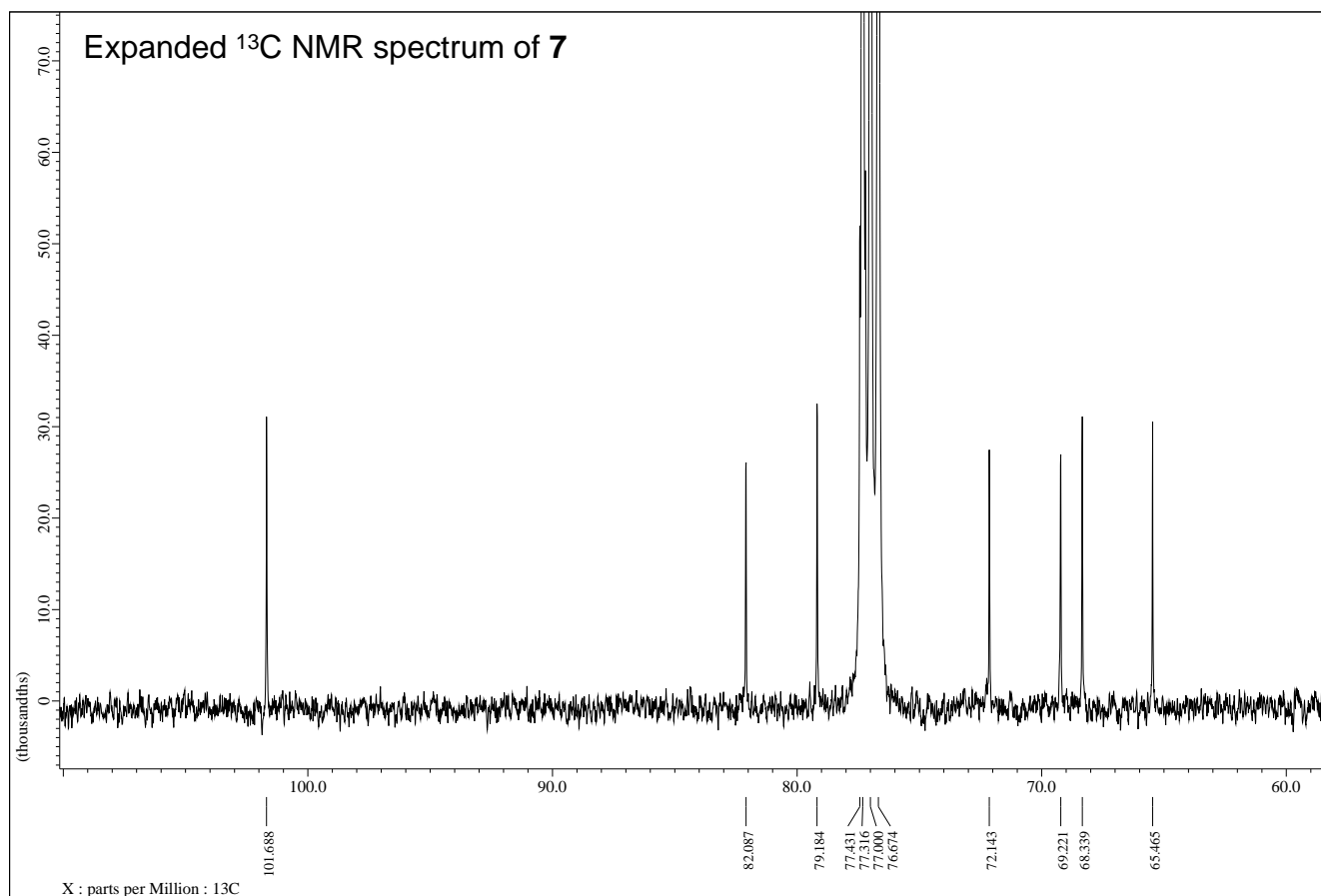
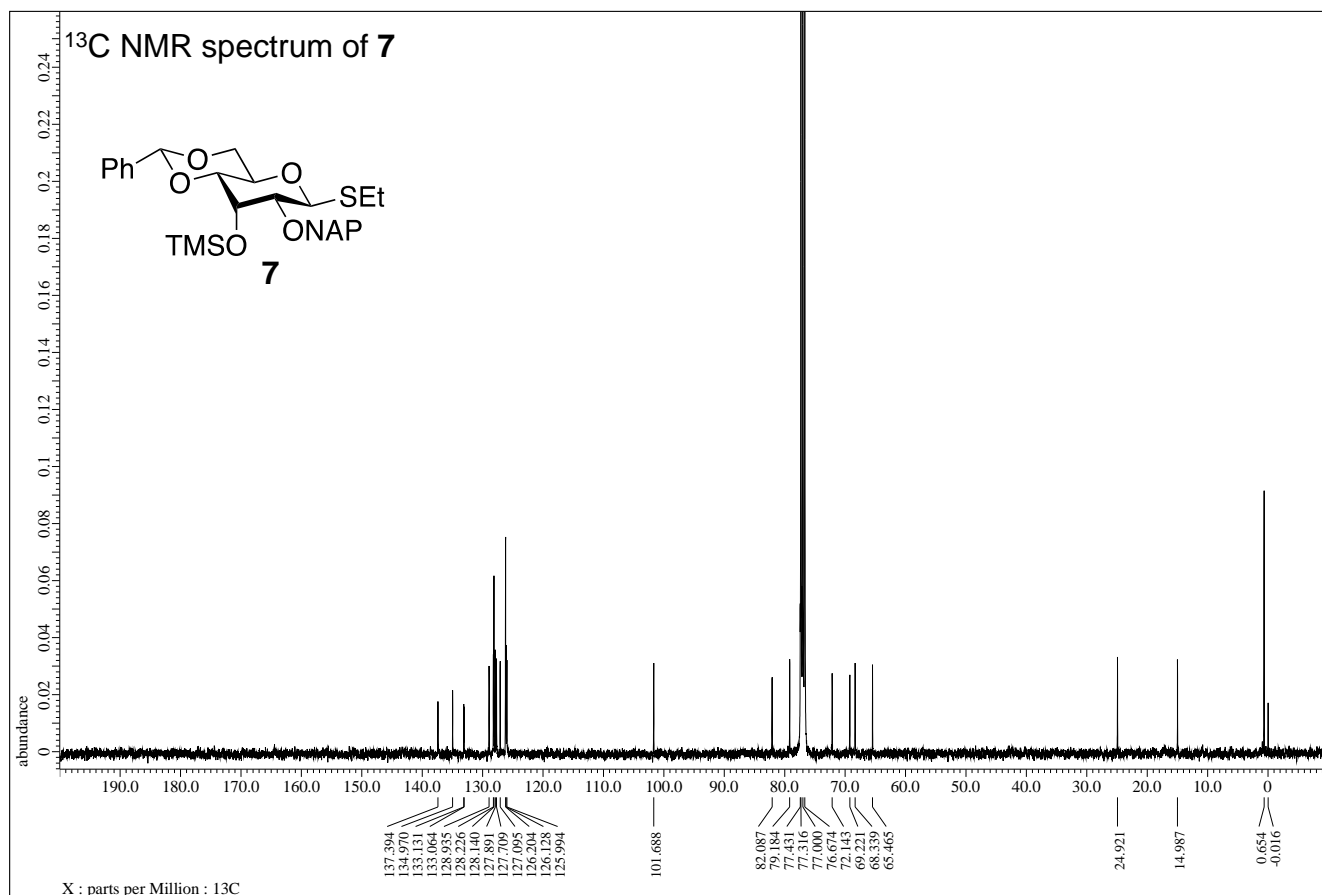


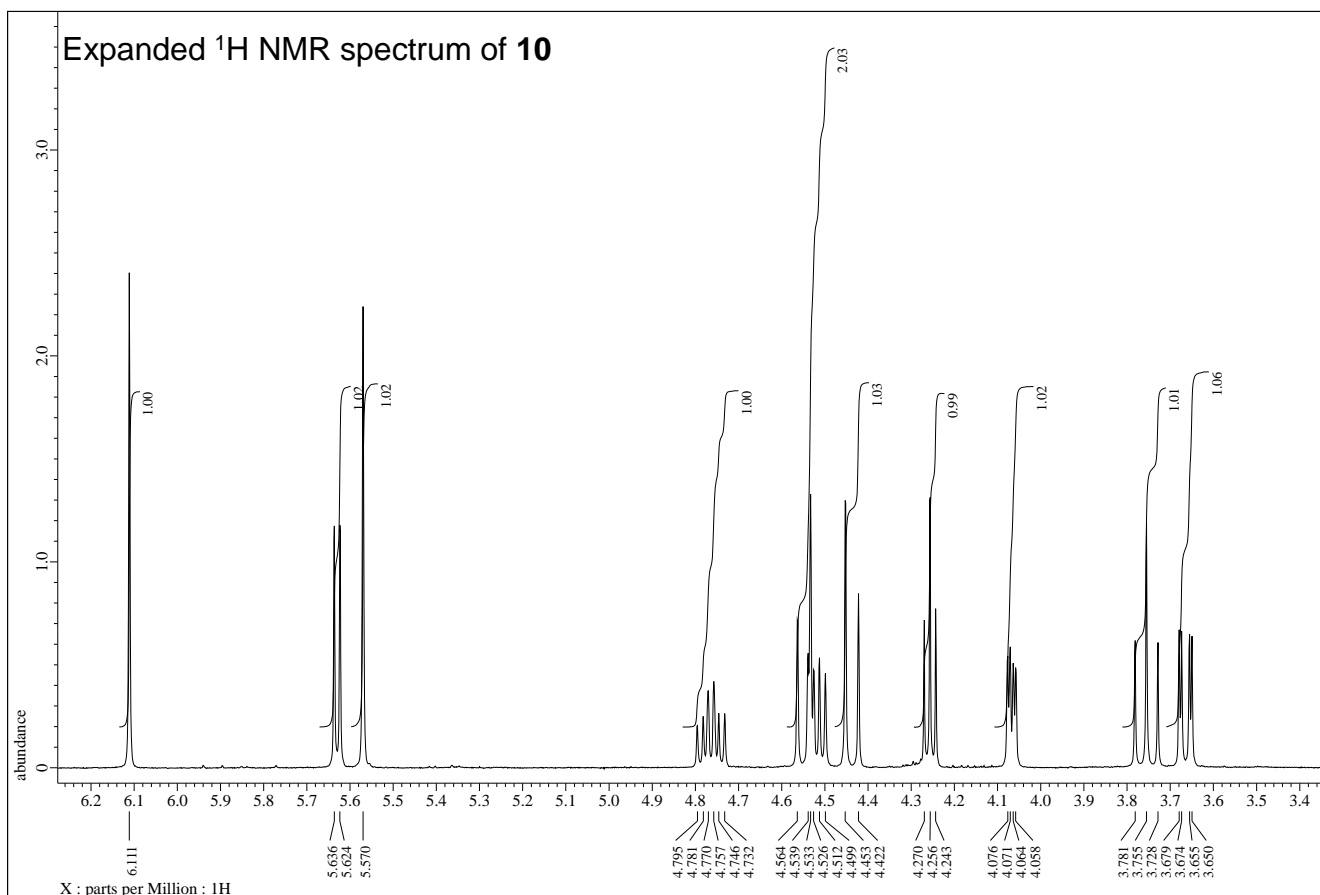
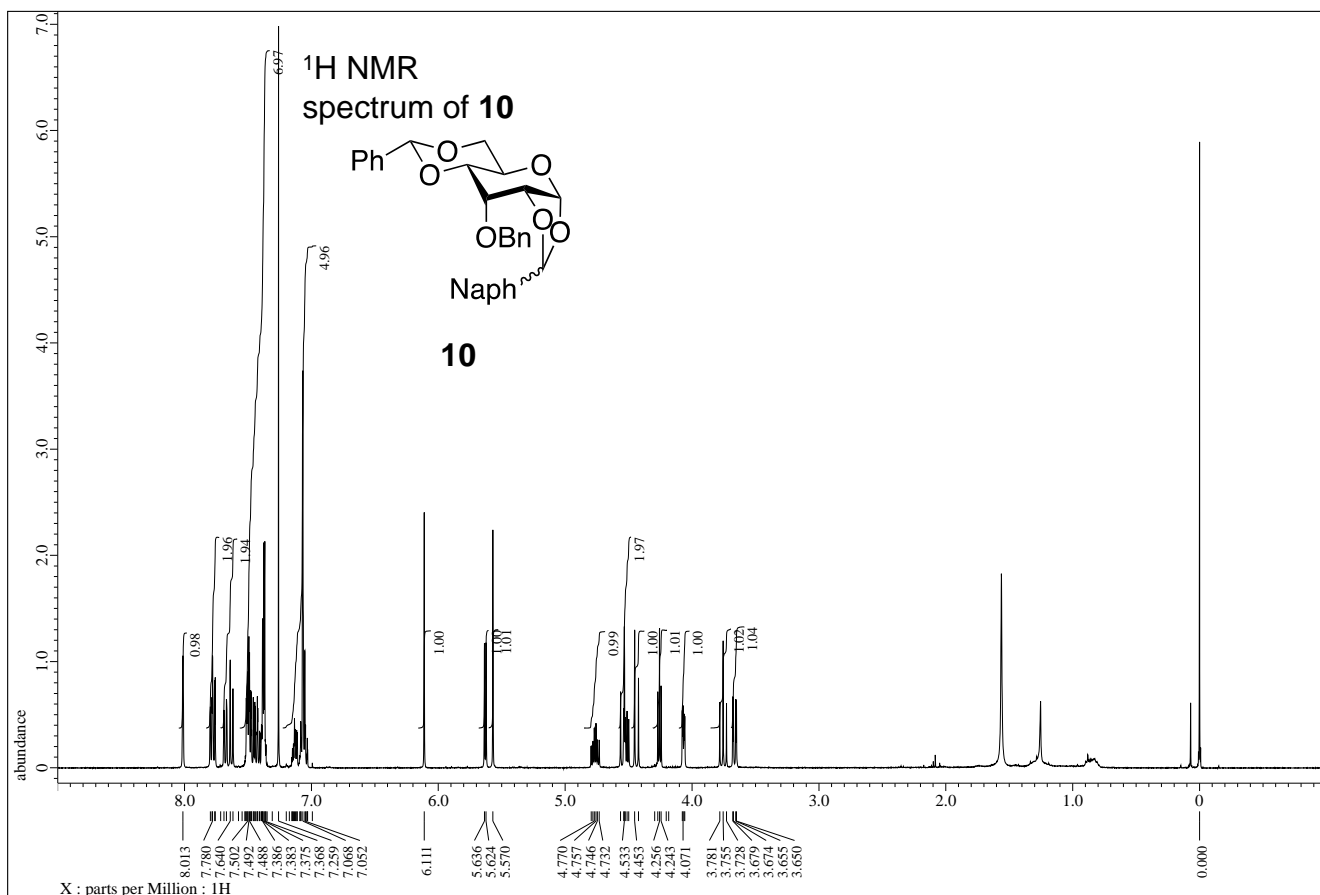


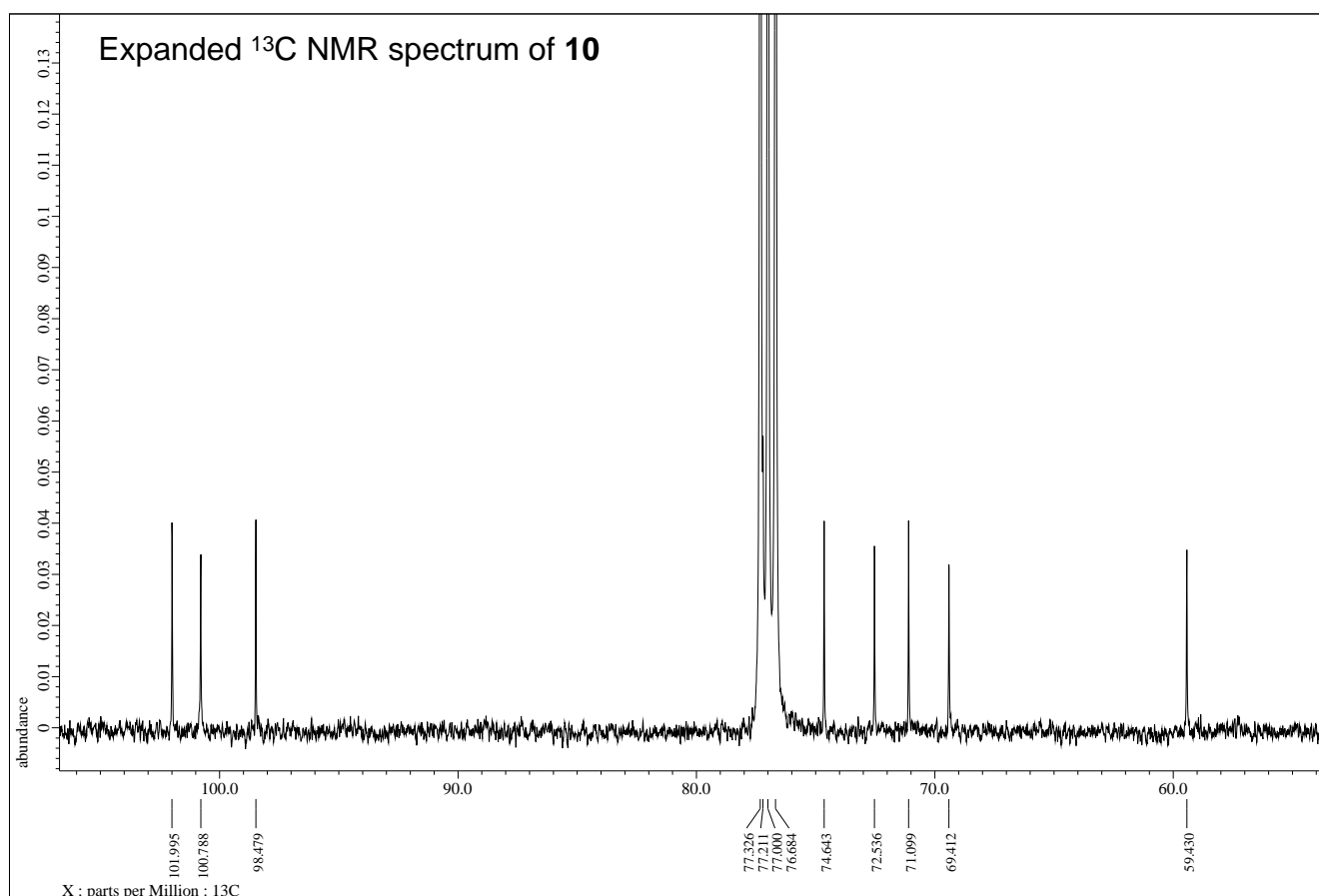
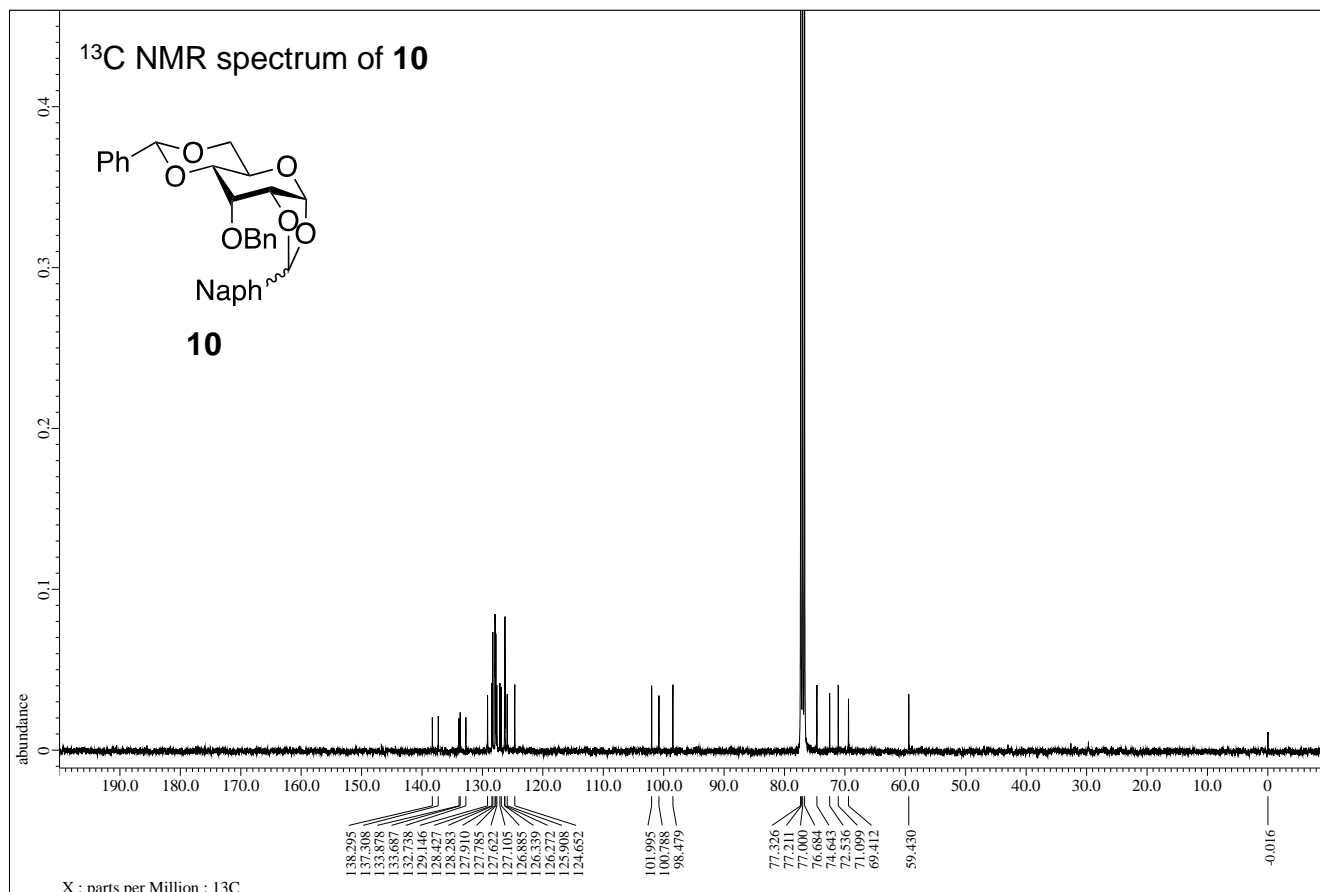


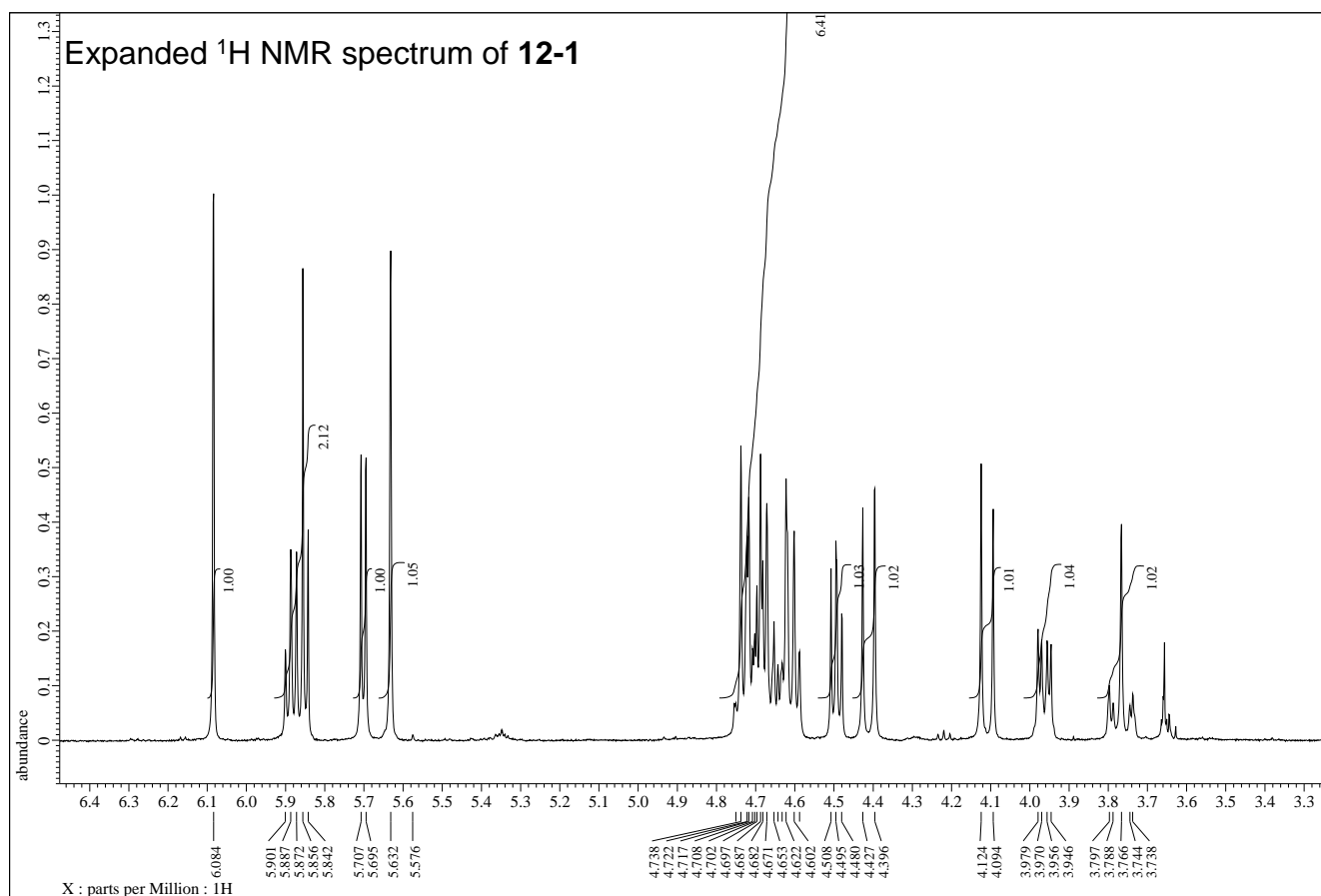
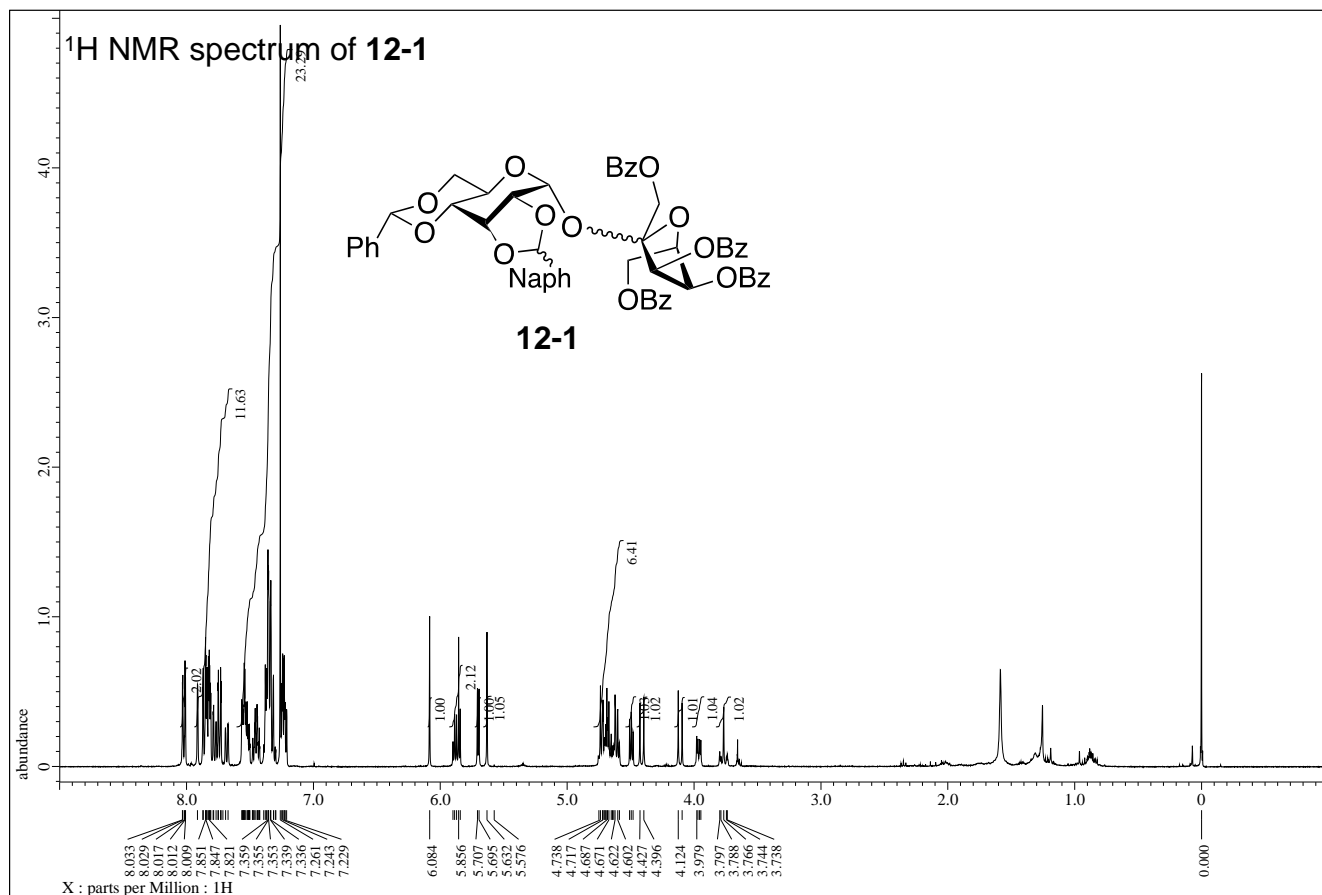


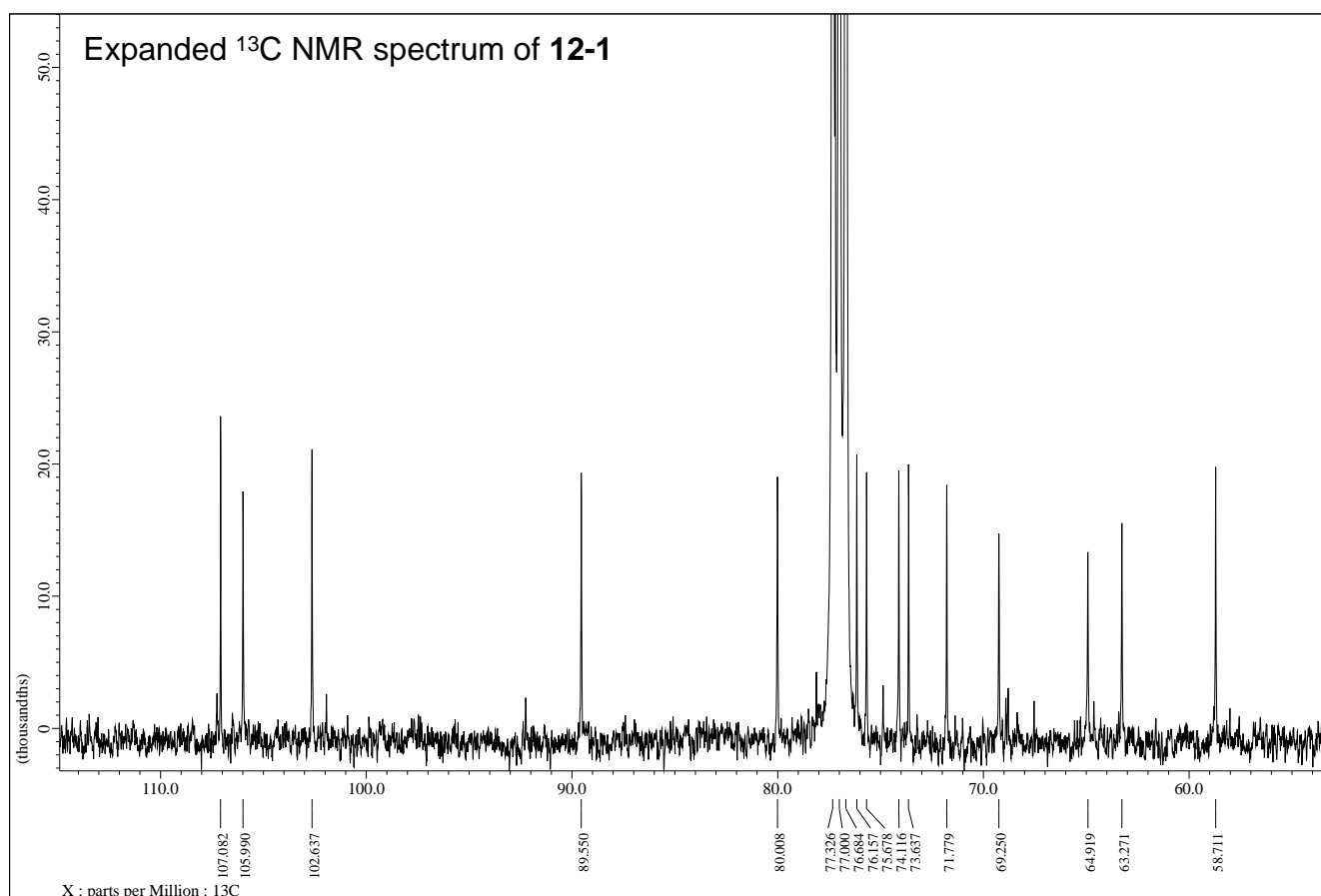
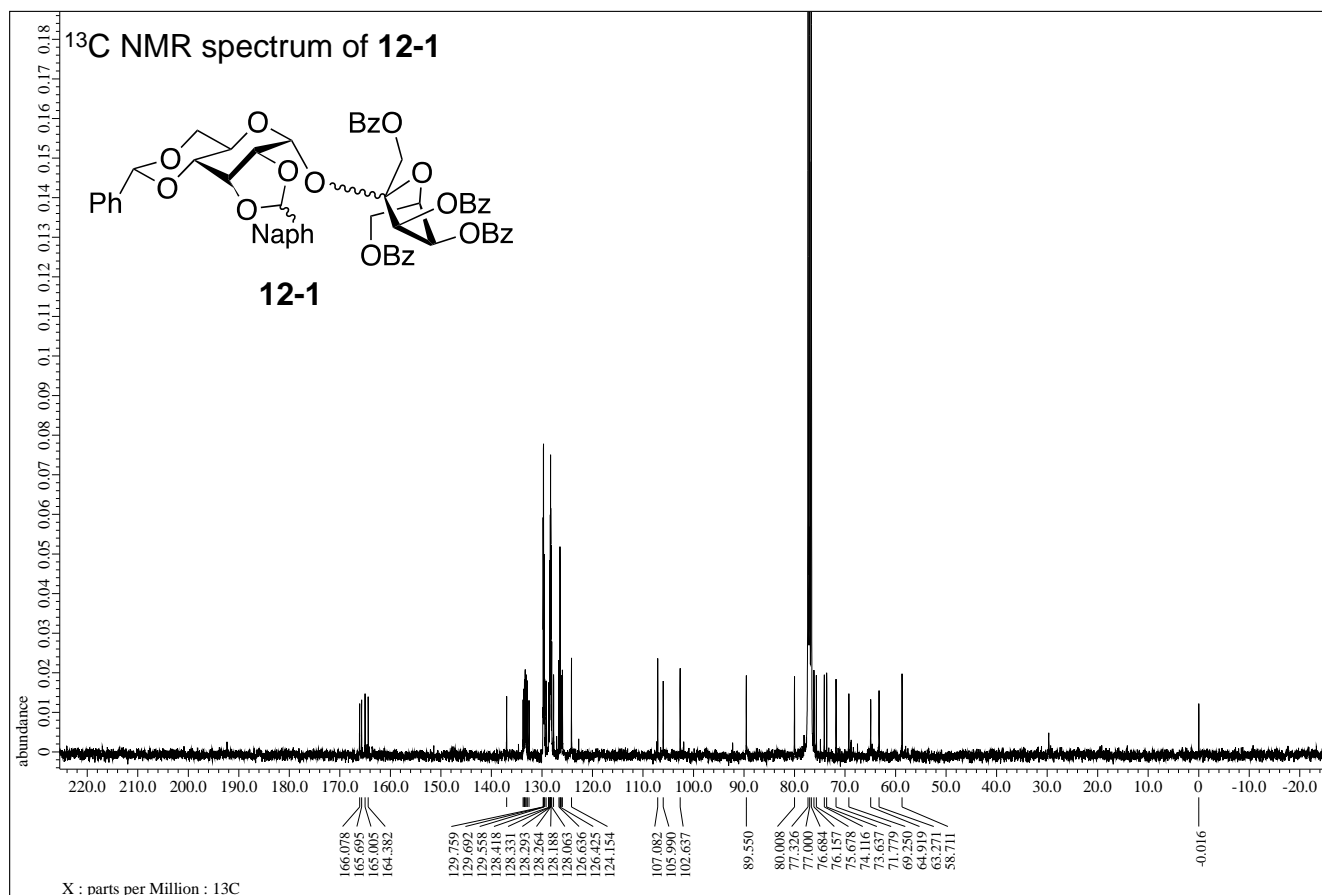


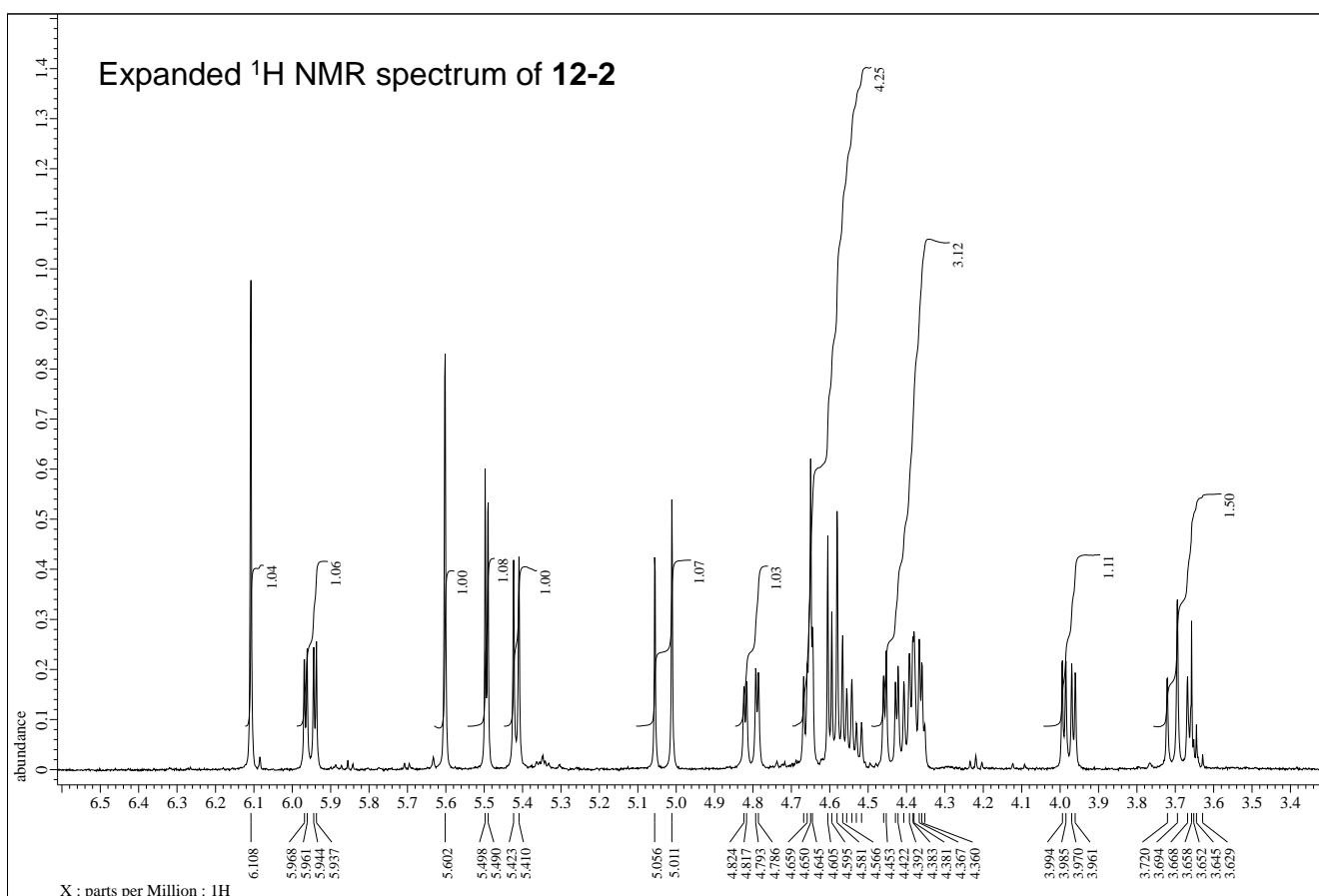
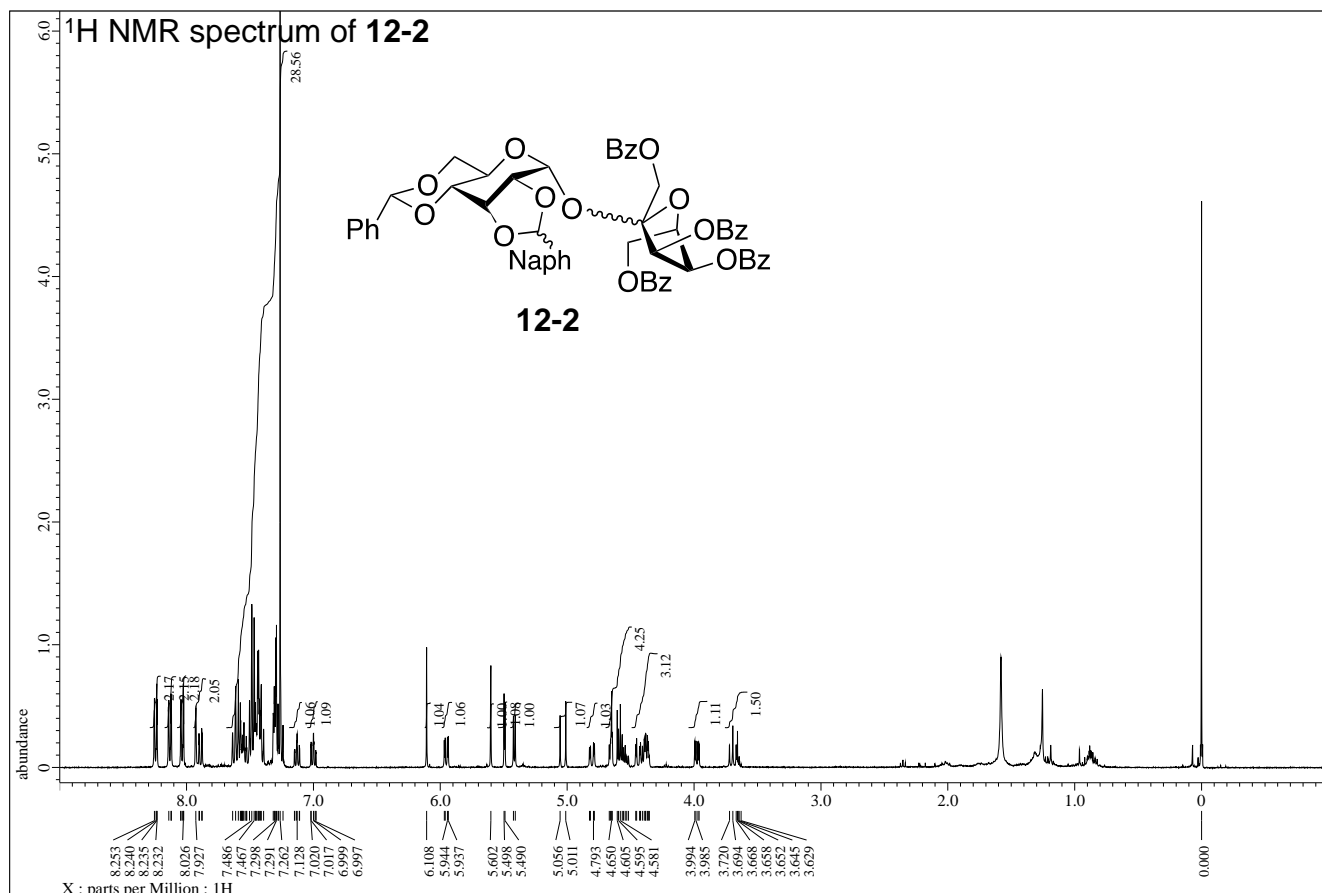


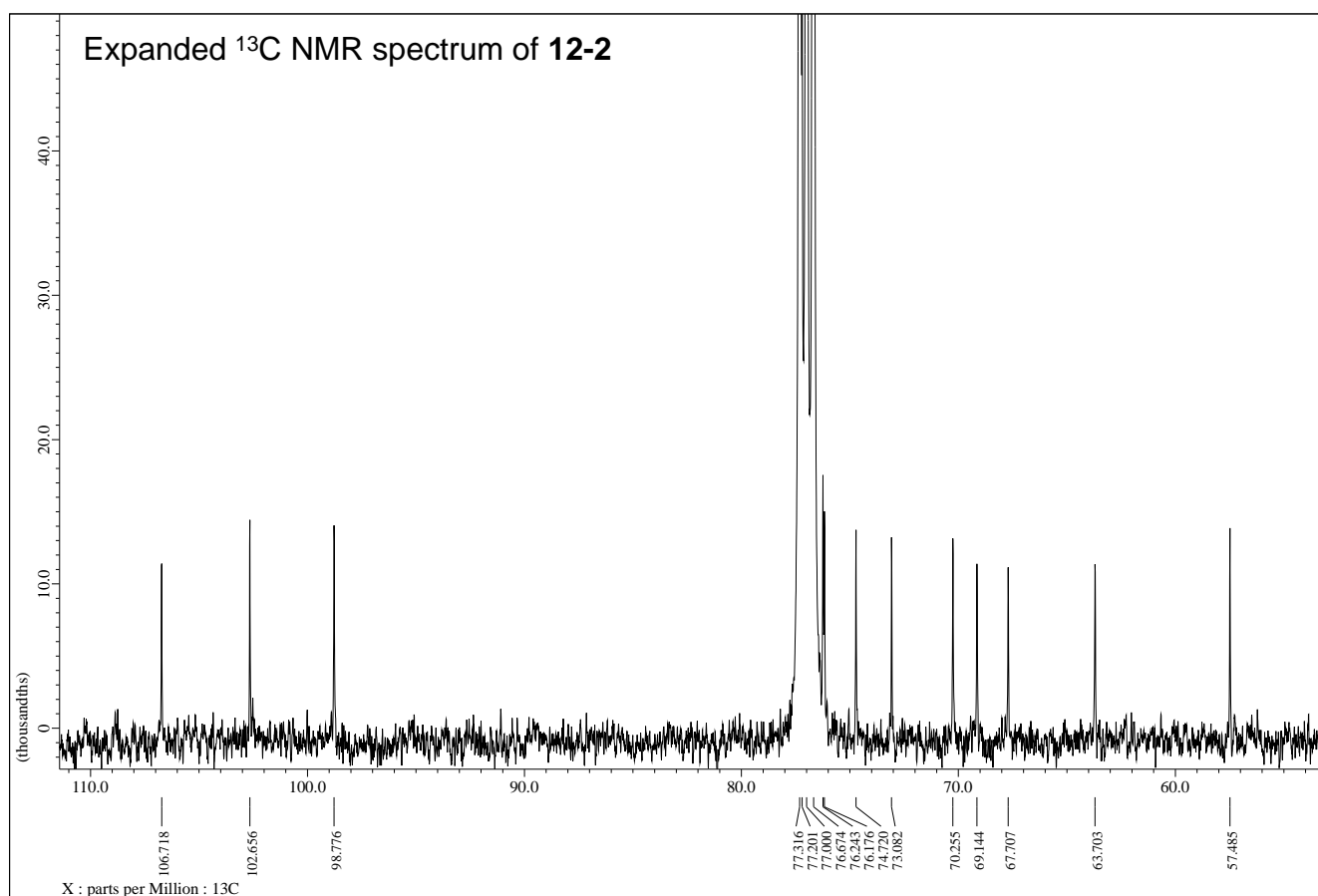
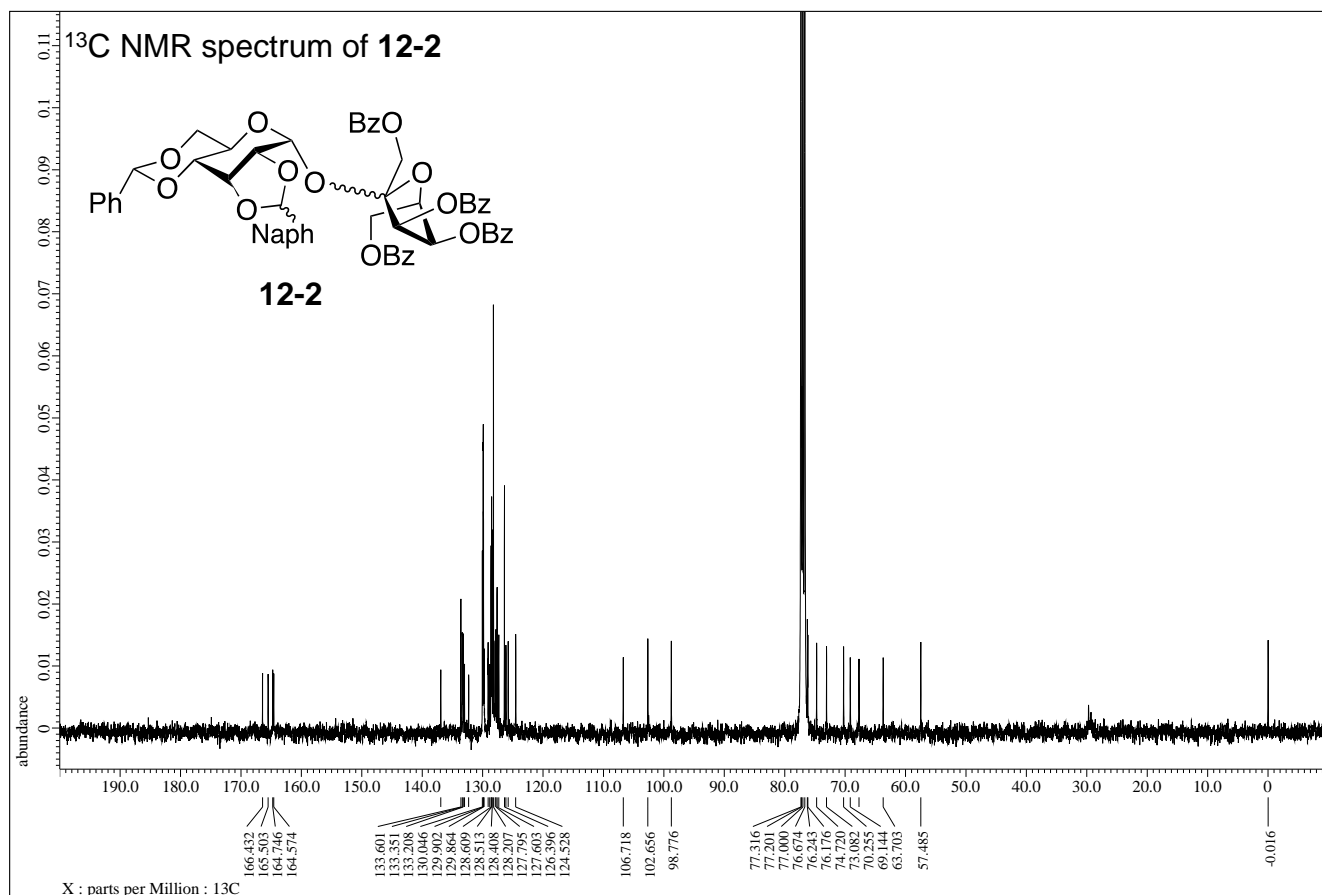












¹H NMR spectrum of [1·BzONa]

