

Article

Benzimidazole-Based Derivatives as Apoptotic Antiproliferative Agents: Design, Synthesis, Docking, and Mechanistic Studies

Bahaa G. M. Youssif ^{1,*} , Martha M. Morcoss ², Stefan Bräse ^{3,*} , Mohamed Abdel-Aziz ⁴,
Hamdy M. Abdel-Rahman ^{5,6} , Dalal A. Abou El-Ella ⁷ and El Shimaa M. N. Abdelhafez ⁴ 

¹ Pharmaceutical Organic Chemistry Department, Faculty of Pharmacy, Assiut University, Assiut 71526, Egypt

² Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Nahda University, Beni-Suef 62513, Egypt; martha.moheb@nub.edu.eg

³ Institute of Biological and Chemical Systems, IBCS-FMS, Karlsruhe Institute of Technology, 76131 Karlsruhe, Germany

⁴ Department of Medicinal Chemistry, Faculty of Pharmacy, Minia University, Minia 61519, Egypt; abulnil@hotmail.com (M.A.-A.); shimaanaguib_80@mu.edu.eg (E.S.M.N.A.)

⁵ Department of Medicinal Chemistry, Faculty of Pharmacy, Assiut University, Assiut 71526, Egypt; hamdym@aun.edu.eg

⁶ Pharmaceutical Chemistry Department, Faculty of Pharmacy, Badr University in Assiut (BUA), Assiut 71536, Egypt

⁷ Department of Pharmaceutical Chemistry, Faculty of Pharmacy Ain Shams University, Cairo 11566, Egypt; dalal@pharma.asu.edu.eg

* Correspondence: bahaa.youssif@pharm.aun.edu.eg (B.G.M.Y.); stefan.braese@kit.edu (S.B.)

Abstract: A new class of benzimidazole-based derivatives (**4a–j**, **5**, and **6**) with potential dual inhibition of EGFR and BRAF^{V600E} has been developed. The newly synthesized compounds were submitted for testing for antiproliferative activity against the NCI-60 cell line. All newly synthesized compounds **4a–j**, **5**, and **6** were selected for testing against a panel of sixty cancer cell lines at a single concentration of 10 μ M. Some compounds tested demonstrated remarkable antiproliferative activity against the cell lines tested. Compounds **4c**, **4e**, and **4g** were chosen for five-dose testing against 60 human tumor cell lines. Compound **4c** demonstrated strong selectivity against the leukemia subpanel, with a selectivity ratio of 5.96 at the GI₅₀ level. The most effective in vitro anti-cancer assay derivatives (**4c**, **4d**, **4e**, **4g**, and **4h**) were tested for EGFR and BRAF^{V600E} inhibition as potential targets for antiproliferative action. The results revealed that compounds **4c** and **4e** have significant antiproliferative activity as dual EGFR/BRAF^{V600E} inhibitors. Compounds **4c** and **4e** induced apoptosis by increasing caspase-3, caspase-8, and Bax levels while decreasing the anti-apoptotic Bcl2 protein. Moreover, molecular docking studies confirmed the potential of compounds **4c** and **4e** to act as dual EGFR/BRAF^{V600E} inhibitors.

Keywords: benzimidazole; hydrazone; thiosemicarbazide; antiproliferative; NCI



Citation: Youssif, B.G.M.; Morcoss, M.M.; Bräse, S.; Abdel-Aziz, M.; Abdel-Rahman, H.M.; Abou El-Ella, D.A.; Abdelhafez, E.S.M.N.

Benzimidazole-Based Derivatives as Apoptotic Antiproliferative Agents: Design, Synthesis, Docking, and Mechanistic Studies. *Molecules* **2024**, *29*, 446. <https://doi.org/10.3390/molecules29020446>

Academic Editor: Chiara Brullo

Received: 6 November 2023

Revised: 20 December 2023

Accepted: 22 December 2023

Published: 16 January 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Cancer has become a global burden due to the significant increase in cancer incidence and mortality rates [1,2]. Tumor resistance, drug toxicity, cancer recurrence, and the inadequate success rate of drug development reaching clinical trials are all limiting variables that exacerbate the difficulties in cancer treatment [3,4]. The search for novel classes of anti-cancer medications focuses on enhancing cancer patients' treatment efficacy and survival rates. The usual "one-size-fits-all" approach of standard non-targeting medicines damages healthy cells and may not benefit all patients [5,6]. Because cancer is a major focus of the precision medicine program, customized therapy approaches targeted at optimizing outcomes based on individual variability in genetic profile, lifestyle, and

environmental factors are gaining popularity [7]. As a result, targeted therapy is the cornerstone for precision medicine, allowing for individualized treatment targeting specific tumor oncogenic markers.

Due to their different biological properties and clinical uses, various benzimidazole compounds have recently attracted attention in anti-cancer agent research [8]. Because of its unique core structure and low toxicity, benzimidazole is an ideal scaffold in the development of anti-cancer drugs [9,10]. Benzimidazole is a bioactive heterocyclic molecule that is one of the top ten most commonly used five-membered nitrogen heterocycles in US Food and Drug Administration (FDA)-approved medications [11]. The electron-rich nitrogen heterocycles of benzimidazole may rapidly receive or donate protons and easily facilitate the creation of different weak contacts, giving it an advantage in binding with a wide range of therapeutic targets and demonstrating broad-ranging pharmacological effects [12–14]. A broad pharmacological profile of benzimidazole and its derivatives, substitution at the 1, 2, 5, and/or 6 positions, has been documented in numerous categories of medicinal agents with distinct features, with anti-cancer activity topping the list [8,15].

Benzimidazole and its derivatives have been reported to play important roles as topoisomerase inhibitors, DNA intercalation and alkylating agents, androgen receptor antagonists, poly (ADP-ribose) polymerase (PARP) inhibitors, protein kinase inhibitors, dihydrofolate reductase inhibitors, and microtubule inhibitors [16,17].

Kinases are enzymes that mediate the signaling cascade and regulate molecular functions and biological processes like growth, proliferation, differentiation, and apoptosis. The human kinome has roughly 535 protein kinases, which can be divided into tyrosine, serine/threonine, and tyrosine kinase-like enzymes [18,19]. Tyrosine kinases (TKs) include both receptor tyrosine kinases (RTKs) (such as EGFR, FGFR, and PDGFR) and non-receptor tyrosine kinases (NRTKs) (such as ABL, FBK, and SRC). Serine/threonine kinases include BRAF/MEK/ERK, CDKs, and PI3K/AKT/mTOR [20]. A deficiency in cellular kinase phosphorylation can eventually lead to constitutive activation of kinases in live cells, promoting the development of malignancy and tumor progression [18,21]. The benzimidazole scaffold is commonly used as a template for synthesizing kinase inhibitors, as shown in Figure 1.

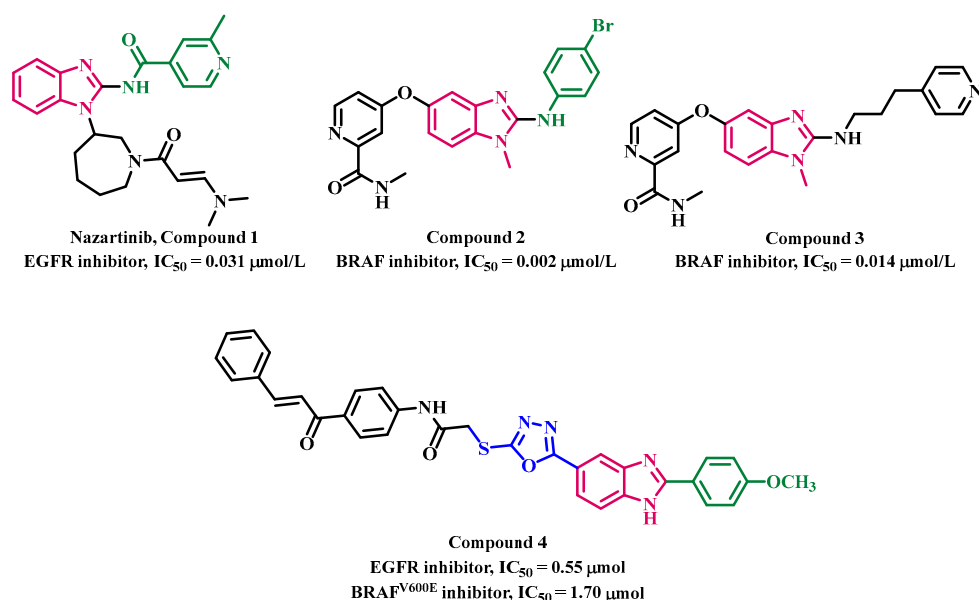


Figure 1. Structures of compounds 1–4.

Nazartinib (1, Figure 1) is an irreversible, third-generation EGFR TKI that targets EGFR-activating mutations [22,23]. Nazartinib contains dimethylamino crotonamide, which is known as the optimal group for several covalent pan-EGFR inhibitors, as well as a racemic 3-substituted azepane linker and a chloro substituent at the benzene ring of the benzimidazole nucleus, which contributes to its improved solubility, oral bioavailability, selectivity, and

high affinity for EGFR [24,25]. In xenograft models, nazartinib inhibits EGFR signaling and the MAPK pathway, inducing cell cycle arrest, apoptosis, and tumor regression [26]. Nazartinib is now being tested in a phase I/II clinical trial in patients with EGFR-mutant non-small cell lung cancer [22].

Ramurthy and colleagues reported the anti-BRAF^{V600E} activity of two benzimidazole-based compounds, **2** and **3** (Figure 1), with IC₅₀ values of 0.002 and 0.014 mmol/L, respectively [27]. The rapidly accelerated fibrosarcoma (RAF) oncogene is associated with cancer cellular processes such as proliferation, invasion, metastasis, and cell survival. Both **2** and **3** can decrease the proliferation of SK-MEL-28 melanoma cells containing B-RAF^{V600E} proteins [27]. Ramurthy and colleagues demonstrated that including a methyl group at the NH of benzimidazole increases the binding affinity to RAF kinase compared to the unmethylated homolog.

Recently [10], we reported on the design and synthesis of a novel series of 1,3,4-oxadiazole/benzimidazole chalcones as antiproliferative agents inducing apoptosis with dual inhibitory effect against EGFR and BRAF^{V600E}. Compound **4** (Figure 1) was shown to be the most potent antiproliferative agent, with a GI₅₀ value of 1.20 µM against four cancer cell lines, when compared to the reference doxorubicin, which had a GI₅₀ value of 1.10 µM. Compound **4** was tested for inhibitory efficacy against EGFR and BRAF^{V600E} as prospective antiproliferative targets. Compound **4** inhibited both targets significantly, with IC₅₀ values of 0.55 ± 0.10 µM and 1.70 ± 0.20 µM, respectively, when compared to the reference erlotinib IC₅₀ values of 0.08 ± 0.01 and 0.06 ± 0.01 µM, respectively.

Motivated by the promising anti-cancer activity of some benzimidazole-based derivatives and as part of our ongoing effort to develop a dual or multi-targeted anti-cancer agent [28–33], we present here the design, synthesis, and antiproliferative activity of a novel series of benzimidazole-based derivatives (**4a–j**, **5**, and **6**, Figure 2). NCI chose all newly synthesized compounds for an in vitro one-dose inhibitory experiment against a panel of 60 cancer cell lines. The most active compounds were chosen for five-dose experiments and mechanistic studies against two potential targets, EGFR and BRAF^{V600E}. Furthermore, the most effective derivatives were tested for apoptosis-inducing activity against caspase-3, caspase-8, Bax, and anti-apoptotic Bcl2. Finally, docking analyses were undertaken for the most potent derivatives to evaluate their binding mechanisms and interactions with the selected receptors.

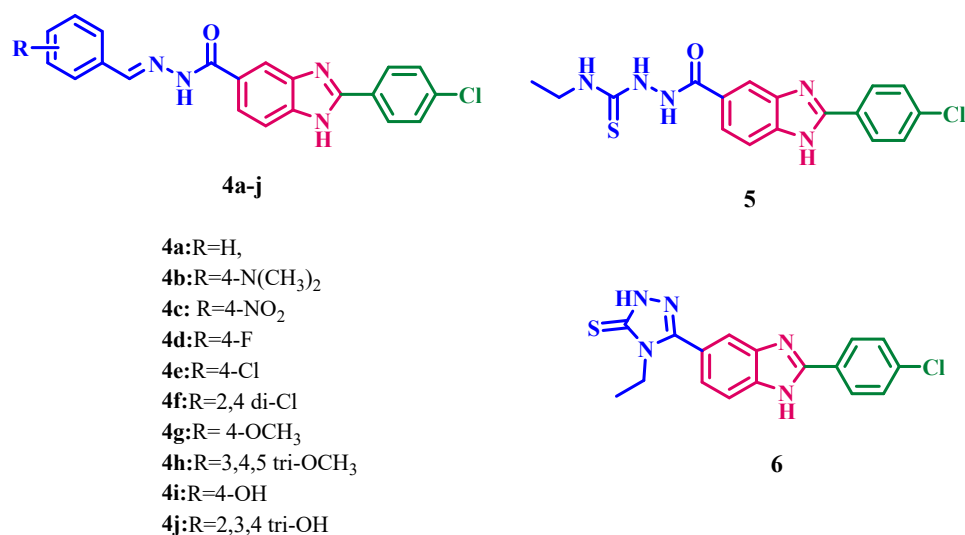


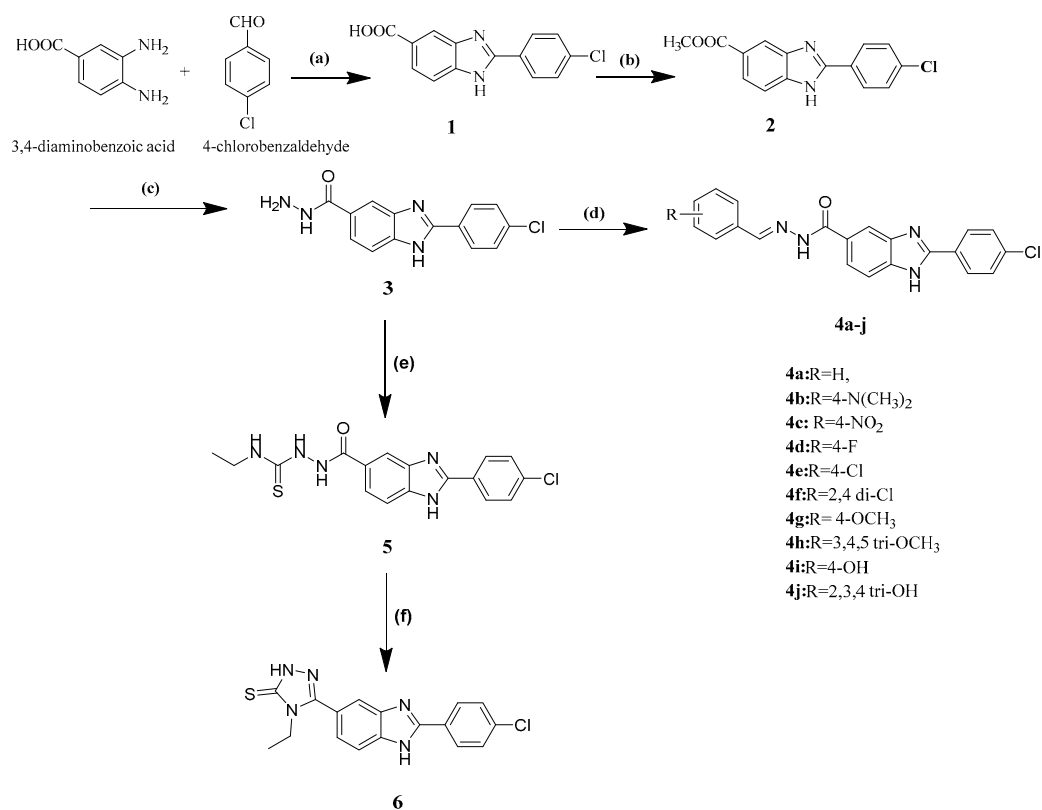
Figure 2. Structures of new targets **4a–j**, **5**, and **6**.

2. Results and Discussion

2.1. Chemistry

Scheme 1 depicts the synthesis method for compounds **4a–j**, **5**, and **6**. Condensation of 3,4-aminobenzoic acid with p-chlorobenzaldehyde in the presence of sodium metabisul-

phite and DMF resulted in the synthesis of benzimidazole-5-carboxylic acid (**1**) in a good yield [34,35]. Compound **1**'s infrared spectra showed that C=O was present at 1662 cm^{-1} . Furthermore, a prolonged peak at $2683\text{--}3358\text{ cm}^{-1}$ was identified as the COOH group's OH. Furthermore, the ^1H NMR spectra of sample **1** showed the existence of seven aromatic protons at δ 7.63–8.20 ppm and a singlet signal at δ 13.32 ppm that is attributable to the COOH proton, which is exchangeable with D_2O . The Fischer esterification of carboxylic acid **1** was accomplished using methanol and a few drops of conc. H_2SO_4 , and the mixture was heated at reflux for 17 h to obtain compound **2** in 60% yield [36,37]. Compound **2**'s IR spectra showed that the OH peak vanished and the C=O group shifted to 1716 cm^{-1} as a result of ester production. Compound **2**'s ^1H NMR spectra revealed a singlet peak associated with CH_3 at 3.87 ppm. The hydrazide **3** was prepared in 58% yield by heating the ester **2** at reflux with 99% hydrazine monohydrate in ethanol [38]. Compound **3**'s IR spectra revealed forked peaks in the infrared spectrum corresponding to the NH_2 and NH protons, respectively, at 3316, 3292, and 3423 cm^{-1} . In addition, ^1H NMR spectra revealed that the methyl protons signal vanished and singlet signals appeared at δ : 4.46 and 9.81 ppm, corresponding to NH_2 and NH, respectively. The target compounds **4a–j** were prepared in high yield by heating the hydrazide **3** at reflux with substituted benzaldehyde in absolute ethanol, catalyzed by a few drops of glacial acetic acid. The structure of hydrazone derivatives **4a–j** was confirmed using different spectroscopic techniques. The IR spectra of **4a–j** exhibited the disappearance of forked NH_2 and the presence of absorption bands at ν_{max} ; 3423–3225 and at ν_{max} ; 1441–1566 cm^{-1} , corresponding to NH and C=N, respectively. The ^1H NMR spectra of compounds **4a–j** revealed the disappearance of the NH_2 proton at δ : 4.46–5.66 ppm and the presence of two signals at δ : 8.32–8.86 and δ : 11.60–13.41 ppm attributed to azomethine proton $\text{CH}=\text{N}$ and hydrazone NH, respectively. Moreover, the ^{13}C NMR spectra of **4a–j** revealed the presence of a new signal at δ : 161.21–164.69 ppm related to hydrazone C=N carbon.



Scheme 1. Synthetic steps for compounds **4a–j**, **5**, and **6**.

Reagent and reaction conditions: (a) DMF, Na₂S₂O₈; (b) conc. H₂SO₄, methanol, reflux; (c) NH₂NH₂, H₂O; (d) aldehyde, CH₃COOH; (e) ethyl isothiocyanate, ethanol; (f), NaOH.

The thiosemicarbazide **5** was prepared via heating at reflux of the hydrazide **3** with ethyl isothiocyanate in absolute ethanol [39]. The 1,2,4-triazole-3-thione **6** was synthesized by cyclization of the 1,4-disubstituted thiosemicarbazide **5** in the presence of 2N sodium hydroxide solution followed by acidification with hydrochloric acid until pH = 3. The proposed mechanism for forming 1,2,4-triazole derivative **6** is through the nucleophilic addition of terminal nitrogen atoms to the carbonyl group; then, the species eliminate water to afford the triazole product **6**.

The chemical structure of compound **6** was elucidated by spectroscopic techniques and elemental analyses. The IR spectrum of **6** exhibited a band at $\nu_{\max}$: 3201–3361 cm^{−1} and $\nu_{\max}$: 1501–1537 cm^{−1}, corresponding to NH of triazole and C=S, respectively. The ¹H NMR spectrum of the target compound **6** revealed an exchangeable signal at δ : 13.96–14.36 ppm, attributed to NH triazole, and a triplet-quartet pattern was observed at δ : 1.16–1.20 and δ : 4.08–4.15 ppm, attributed to the ethyl protons. The ¹³C NMR spectrum of compound **6** revealed the presence of a signal at δ : 166.69 ppm related to C=S carbon and at δ : 13.91, 18.93 related to CH₃ and CH₂ moieties.

2.2. Biology

2.2.1. In Vitro Anti-Cancer Activity

The final compounds were submitted to the National Cancer Institute, Bethesda, USA, for screening in vitro for anti-cancer [40,41]. Eleven compounds, **4a–j**, **5**, and **6**, were selected according to the Development Therapeutic Program (DTP) protocol. The selected compounds were evaluated for their in vitro cytotoxic activity against a panel of sixty cancer cell lines (NCI-60 cell line panel) derived from six cell lines panel of leukemia, nine cell lines of lung cancer, six cell lines of CNS cancer, seven cell lines of colon cancer, eight cell lines of melanoma, six cell lines of ovarian cancer, eight cell lines of renal cancer, two cell lines of prostate cancer, and eight cell lines of breast cancer, representing the full nine human systems as leukemia, melanoma, and cancers of the lung, colon, brain, breast, ovary, kidney, and prostate.

In Vitro One-Dose Assay

The selected compounds **4a–j**, **5**, and **6** were tested at a single concentration of 10 μ M. The screening results for the selected compounds are illustrated in Table 1.

The screening results of the target compounds revealed that benzimidazole-bearing hydrazone derivatives **4a–j** have more potent anti-cancer activity than triazole derivatives **6**. Among hydrazone derivatives, compounds **4c**, **4d**, **4e**, **4f**, **4g**, **4h**, and **4j** showed promising anti-cancer activity in a one-dose assay at 10 μ M.

Compound **4c** revealed significant to complete cell death against the leukemia cell line with cell inhibition percentages of 90.13, 89.22, 93.42, 90.32, 95.76, and 108.49. Moreover, it showed remarkable anti-cancer activity against non-small cell lung cancer with cell inhibition percentages of 65.46, 50.97, 55.22, 40.32, 45.22, 83.75, 94.56, and 66.97. In addition, it displayed significant anti-cancer activity against colon cancer with cell inhibition percentages of 97.72, 85.09, 85.40, 70.12, 91.75, 86.97, and 87.34. In addition, it exhibited moderate to significant anti-cancer activity against the CNS cancer cell line with cell inhibition percentages of 51.50, 78.32, 58.34, 47.95, 30.71, and 77.67. It also showed moderate to complete cell death against melanoma with cell inhibition percentages of 100.91, 92.21, 74.09, 52.42, 33.28, 54.65, 97.42, 74.92, and 56.35. Likewise, it displayed moderate to significant anti-cancer activity against ovarian cancer with cell inhibition percentages of 80.88, 88.68, 76.99, 46.76, 78.40, and 39.50. Furthermore, it exhibited complete cell death against renal cancer with cell inhibition percentages of 50.97, 100.50, 67.71, 57.25, 68.40, 80.35, and 57.44. In addition, it displayed remarkable anti-cancer activity against prostate cancer with cell inhibition

percentages of 78.57 and 57.97. Finally, it exhibited significant to complete cell death against breast cancer with cell inhibition percentages of 99.28, 63.24, 85.96, 89.50, and 138.86.

Table 1. Cell inhibition percentages of NCI-60 cancer cell lines revealed by target compounds **4a–j**, **5**, and **6**.

Subpanel Cancer Cell Lines	Cell Inhibition Percent											
	4a	4b	4c	4d	4e	4f	4g	4h	4i	4j	5	6
Leukemia												
CCRF-CEM	11.20	6.50	90.13	78.63	84.49	48.15	34.69	63.44	13.58	45.33	86.90	0
RPMI-8226	13.83	5.27	89.22	32.55	97.17	15.43	40.78	80.13	1.52	5.54	95.85	0.68
HL-60(TB)	0	0	93.42	62.62	94.23	5.98	3.67	55.12	0	0	0	0
K-562	7.37	5.96	90.32	61.21	91.33	53.70	68.14	67.33	0.25	0	88.44	1.43
MOLT-4	12.90	3.18	95.76	76.45	95.55	72.64	31.79	58.45	15.60	17.16	90.05	2.85
SR	18.05	2.54	108.49	79.32	−124.1	54.59	99.35	54.32	2.26	51.12	88.91	17.53
Non-Small Cell Lung Cancer												
A549/ATCC	0	0	65.46	82.16	126.66	4.70	88.04	14.22	0	0	6.17	0
EKVX	1.38	2.85	50.97	26.79	91.77	13.67	70.56	4.23	0	82.89	18.21	0.44
HOP-62	0	2.51	55.22	39.48	91.95	5.89	69.68	18.33	0	59.21	0.87	1.71
HOP-92	0	0	40.32	15.44	71.87	0	64.03	28.33	0	92.98	1.09	0.32
NCI-H226	5.74	4.67	45.22	6.86	0.67	18.84	26.17	24.03	20.60	51.83	9.79	1.97
NCI-H23	1.85	1.51	83.75	38.62	100.37	4.26	58.45	46.27	4.44	93.41	5.79	4.53
NCI-H322M	0	0.24	29.93	22.45	72.31	0	44.09	12.45	0.47	59.69	3.36	0
NCI-H460	0	2.77	94.56	63.77	133.56	1.37	94.09	17.19	0	97.46	1.06	0
NCI-H522	1.00	3.02	66.97	3.87	36.76	7.79	17.38	21.72	9.85	88.43	5.4	1.85
Colon Cancer												
COLO 205	0.76	0	97.72	58.71	123.92	0	77.14	54.34	0.44	0	0	0
HCC-2998	0	6.31	85.09	20.98	96.31	3.25	48.89	20.02	0	14.96	5.54	0
HCT-116	0	4.70	85.40	51.35	101.88	8.63	81.02	63.83	0	5.85	0	0
HCT-15	4.81	0	70.12	33.19	95.31	0	42.48	3.98	0	4.51	5.90	1.62
HT29	3.67	2.26	91.75	95.67	98.33	2.18	78.10	60.07	0	0	2.33	2.07
KM12	3.80	2.41	86.97	35.11	129.12	1.75	82.72	38.17	2.85	13.12	7.97	0
SW-620	0	0	87.34	53.71	128.29	4.30	75.58	21.33	0	5.64	0	0
SF-268	0	0.73	51.50	21.80	62.03	3.63	80.07	20.31	13.65			
SF-295	13.18	1.06	78.32	68.72	149.27	1.19	138.85	55.97	0	56.38	1.33	1.04
Continued								56.23	1.95	12.11	4.31	0.56
SF-539	9.57	5.22	58.34	12.04	62.90	5.73	57.39			29.47	12.25	3.69
SNB-19	0	0	47.95	39.59	95.65	7.76	75.84	28.97	0	20.22	2.37	0
SNB-75	10.65	0.61	30.71	2.77	1.60	12.39	8.15	13.37	0	9.91	7.77	3.91
U251	4.35	0	77.67	98.38	115.25	41.02	92.04	46.76	0	43.95	2.36	0
Melanoma												
LOX IMVI	7.23	3.70	100.91	65.96	155.86	18.88	99.12	49.36	11.95	33.15	12.38	0
MALME-3M	1.28	12.86	92.21	88.22	158.65	9.55	93.47	71.57	21.36	19.90	1.54	2.41
M14	8.89	3.58	74.09	39.75	80.33	17.14	73.78	57.97	0.44	15.66	1.67	0
MDA-MB-435	0	0	52.42	6.12	61.19	0	31.49	35.98	0	1.11	0.91	0
SK-MEL-2	0	0	33.28	16.72	61.09	2.90	35.90	16.41	0	13.30	0	0
SK-MEL-28	0	0	54.65	28.10	109.23	0	87.53	53.48	0	20.39	0	0
SK-MEL-5	4.85	5.02	97.42	11.02	63.84	2.92	66.34	58.08	4.32	7.75	9.57	2.04
UACC-257	0	0	74.92	0	68.26	6.59	45.62	45.65	0	0	0	0
UACC-62	0	1.06	56.35	11.85	83.15	0	69.11	71.28	5.20	26.16	1.88	1.04

Table 1. Cont.

Subpanel Cancer Cell Lines	Cell Inhibition Percent											
	4a	4b	4c	4d	4e	4f	4g	4h	4i	4j	5	6
IGROV1	4.80	14.96	80.88	59.83	127.35	31.01	85.41	31.57	17.03			
OVCAR-3	0	0	88.68	29.34	152.88	0	79.87	38.44	0	26.51	14.29	0.16
OVCAR-4	0	0	76.99	63.60	117.88	0	130.63	0	0	32.23	0	0
OVCAR-5	0	0	46.76	10.84	76.28	0	22.66	0	0	25.74	5.13	0.60
OVCAR-8	1.37	5.81	78.40	48.73	85.77	17.10	68.70	39.48	0	11.87	3.26	0
NCI/ADR-RES	0	0.43	22.73	22.98	58.78	0	26.43	0	0	22.28	6.74	0
SK-OV-3	4.70	0	39.50	1.74	50.18	7.23	27.70	12.22	5.09	0	1.07	0
Renal Cancer										21.76	0.37	0
786-0	11.58	0	50.97	87.83	142.12	0	96.85	48.80	0			
A498	22.08	12.86	100.50	27.11	43.85	36.67	33.43	101.95	5.34	0	0	0
ACHN	3.26	1.36	67.71	64.33	120.18	9.01	78.11	32.27	4.33	22.48	57.69	56.86
CAKI-1	1.57	4.40	57.25	15.06	38.77	9.61	39.51	24.00	1.91	0.62	6.31	0
RXF 393	15.54	7.79	68.40	22.63	81.08	12.49	56.23	63.67	0	10.39	17.05	7.78
SN12C	3.99	1.47	80.35	58.81	136.90	1.23	93.76	31.47	0	7.62	7.61	13.89
TK-10	0	0	10.16	5.63	104.33	0	60.87	22.64	0	0	10.09	0
UO-31	8.73	13.31	57.44	24.61	78.42	27.84	18.07	11.66	15.86	0	0	0
Prostate Cancer										25.13	6.39	16.76
PC-3	0	4.81	78.57	53.21	124.90	7.03	104.40	40.06	0			
DU-145	3.83	0	57.97	14.62	69.46	0	20.74	2.57	0	18.38	7.83	7.76
Breast Cancer										5.98	0	0
MCF7	23.51	23.84	99.28	82.56	119.97	13.41	83.33	87.88	25.12			
MDA-MB 231	0	15.58	23.15	36.06	90.52	11.40	60.16	0	2.75	11.50	22.10	14.58
HS 578T	7.92	6.32	63.24	30.02	67.05	10.43	67.67	51.70	5.94	27.19	20.49	2.36
BT-549	4.12	0	85.96	0	0	0	43.07	29.28	0	16.99	9.75	1.05
T-47D	18.22	14.13	89.50	40.63	113.10	12.30	76.71	84.11	24.20	15.72	0	7.91
MDA-MB-468	0	0	138.86	23.87	144.13	0	75.82	0	0	19.77	18.43	0.30
										90.44	0	0

Compound **4d** exhibited moderate to significant cytotoxic activity with cell inhibition percentages of 78.63, 32.55, 62.62, 61.21, 76.45, 79.32, 82.16, 39.48, 38.62, 63.77, 58.71, 51.35, 95.67, 35.11, 53.71, 68.72, 39.59, 98.38, 65.96, 88.22, 39.75, 59.83, 63.60, 48.73, 87.83, 64.33, 58.81, 53.21, 82.56, 36.06, and 40.63 against CCRF-CEM, RPMI-8226, HL-60, K-562, MOLT-4, SR, A549, HOP-62, NCI-H23, NCI-H460, COLO 205, HCT-16, HT 29, KM12, SW-620, SF-295, SNB-19, U251, LOXIMVI, MALME-3M, M14, IGROV1, OVCAR-4, OVCAR-8, 786-O, ACHN, SN12C, PC-3, MCF7, MDA-MB-231, and T-47D.

Compound **4e** revealed significant to complete cell death against leukemia with cell inhibition percentages of 84.49, 97.17, 94.23, 91.33, 95.55, and 124.18. In addition, it exhibited moderate to complete cell death against non-small cell lung cancer with cell inhibition percentages of 126.66, 91.77, 91.95, 71.87, 100.37, 72.31, 133.56, and 36.76. Moreover, it displayed significant to complete cell death against colon cancer with cell inhibition percentages of 123.92, 96.31, 101.88, 95.31, 98.33, 129.12, and 128.29. Furthermore, it showed remarkable complete cell death against CNS cancer with cell inhibition percentages of 62.03, 149.27, 62.90, 95.65, and 115.25. Additionally, it exhibited remarkable complete cell death against melanoma with cell inhibition percentages of 155.86, 158.65, 80.33, 61.19, 61.09, 109.23, 63.84, 68.26, and 83.15. It also revealed remarkable complete cell death against ovarian cancer with cell inhibition percentages of 127.35, 152.88, 117.88, 76.28, 85.77, 58.78, and 50.18. In addition, it showed moderate to complete cell death against renal cancer with cell inhibition percentages of 142.12, 43.85, 120.18, 38.77, 81.08, 136.90, 104.33, and 78.42.

Finally, it displayed remarkable complete cell death against prostate and breast cancers with cell inhibition percentages of 124.90, 69.46, 119.97, 90.52, 67.05, 113.10, and 144.13.

Compound **4f** exhibited remarkable anti-cancer activity against leukemia cell lines such as CCRF-CEM, K-562, MOLT-4, and SR with cell inhibition percentages of 48.15, 53.70, 72.64, and 54.59. In addition, it showed moderate cytotoxic activity with cell inhibition percentages of 41.02, 31.01, and 36.67 against U251, IGROV1, and A498.

Compound **4g** induced moderate to significant cytotoxic activity with cell inhibition percentages of 34.69, 40.78, 68.14, 31.79, and 99.35 against different leukemia cell lines. Furthermore, it displayed remarkable anti-cancer activity against different cell lines of non-small cell lung cancer with cell inhibition percentages of 88.04, 70.56, 69.68, 64.03, 58.45, 44.09, and 94.09. In addition, it showed moderate to significant anti-cancer activity against colon cancer with cell inhibition percentages of 77.14, 48.89, 81.02, 42.48, 78.10, 82.72, and 75.58. Moreover, it exhibited a remarkable complete cell death against CNS cancer with cell inhibition percentages of 80.07, 138.85, 57.39, 75.84, and 92.04. Likewise, it revealed moderate to significant cytotoxic activity with cell inhibition percentages of 99.12, 93.47, 73.78, 31.49, 35.90, 87.53, 66.34, 45.62, and 69.11 against different melanoma cell lines. It also exhibited significant to complete cell death against ovarian cancer with cell inhibition percentages of 85.41, 79.87, 130.63, and 68.70. Additionally, it showed moderate to significant cytotoxic activity with cell inhibition percentages of 96.85, 33.43, 78.11, 39.51, 56.23, 93.76, and 60.87 against different renal cancer cell lines. Finally, it displayed remarkable complete cell death against prostate and breast cancer with cell inhibition percentages of 104.40, 83.33, 67.67, 43.07, 76.71, and 75.82. Compound **4h** induced moderate cytotoxic activity with cell inhibition percentages of 44.21 and 44.91 against MALME-3M and T-47D.

In the same context, compound **4h** exhibited remarkable anti-cancer activity against leukemia with cell inhibition percentages of 63.44, 80.13, 55.12, 67.33, 58.45, and 54.32. Furthermore, it showed moderate to remarkable cytotoxic activity with cell inhibition percentages of 46.27, 54.34, 63.83, 60.07, and 38.17 against non-small cell lung and colon cancer. Additionally, it induced moderate to remarkable anti-cancer activity against CNS cancer with cell inhibition percentages of 55.97, 56.23, and 46.76. Furthermore, it displayed moderate to significant cytotoxic activity with cell inhibition percentages of 49.36, 71.57, 57.97, 35.98, 53.48, 58.08, 45.65, and 71.28 against different melanoma cell lines. Moreover, it showed moderate to complete cell death against different cell lines of ovarian, renal, prostate, and breast cancer with cell inhibition percentages of 31.57, 38.44, 39.48, 48.80, 101.95, 32.27, 63.67, 31.47, 40.06, 87.88, 51.70, and 84.11.

In the same context, compound **4j** exhibited moderate to significant cytotoxic activity with cell inhibition percentages of 45.33, 51.12, 59.21, 51.83, 59.69, 56.38, 43.95, 32.23, and 90.44 against CCRF-CEM, SR, HOP-62, NCI-H226, NCI-H322M, SF-268, U251, OVCAR-3, and MDA-MB-468. In conclusion, different substitutions of aldehydes such as Cl, NO₂, and OCH₃ have significant anti-cancer activity.

In Vitro Five-Dose Assay

Compounds **4c**, **4e**, and **4g** were selected for advanced five-dose testing against the full panel of 60 human tumor cell lines [42,43], which represents nine tumor subpanels screened at five concentrations (0.01, 0.1, 1, 10, and 100 µM). The results are used to construct a dose-response curve log of concentration versus percentage growth inhibition (Figures 3–5) and calculate three response parameters for each cell line (GI₅₀, TGI, and LC₅₀). The GI₅₀ value (concentration of maximum 50% cell proliferation inhibition) relates to compound concentration causing inhibition by 50% in total cell growth, the TGI value (cytostatic activity) refers to the concentration of the compound resulting in a reduction of total growth, and the LC₅₀ value (cytotoxic activity) indicates the concentration of the compound causing a net 50% inhibition of initial cells at the end of the incubation period of 48 h.

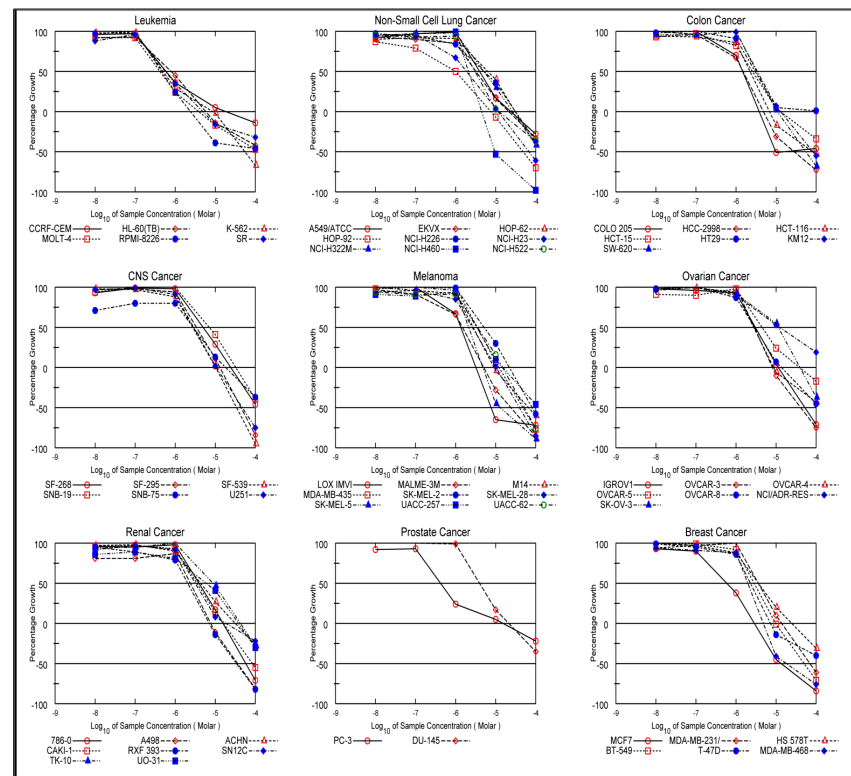


Figure 3. Dose–response curves for all cell lines for compound 4c.

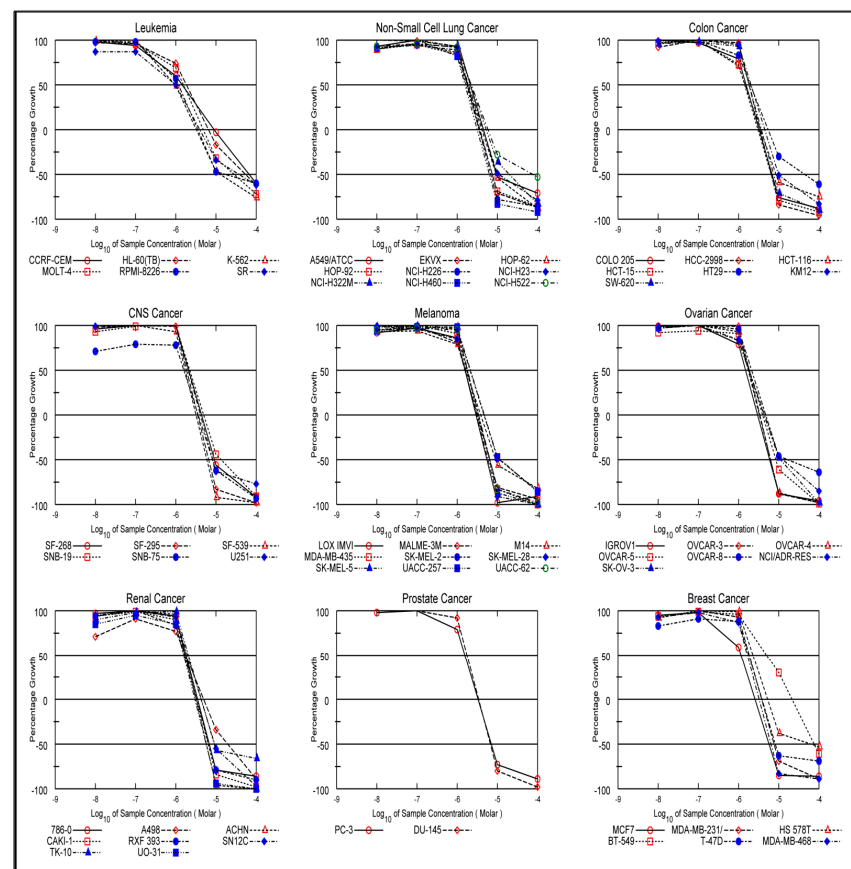


Figure 4. Dose–response curves for all cell lines for compound 4e.

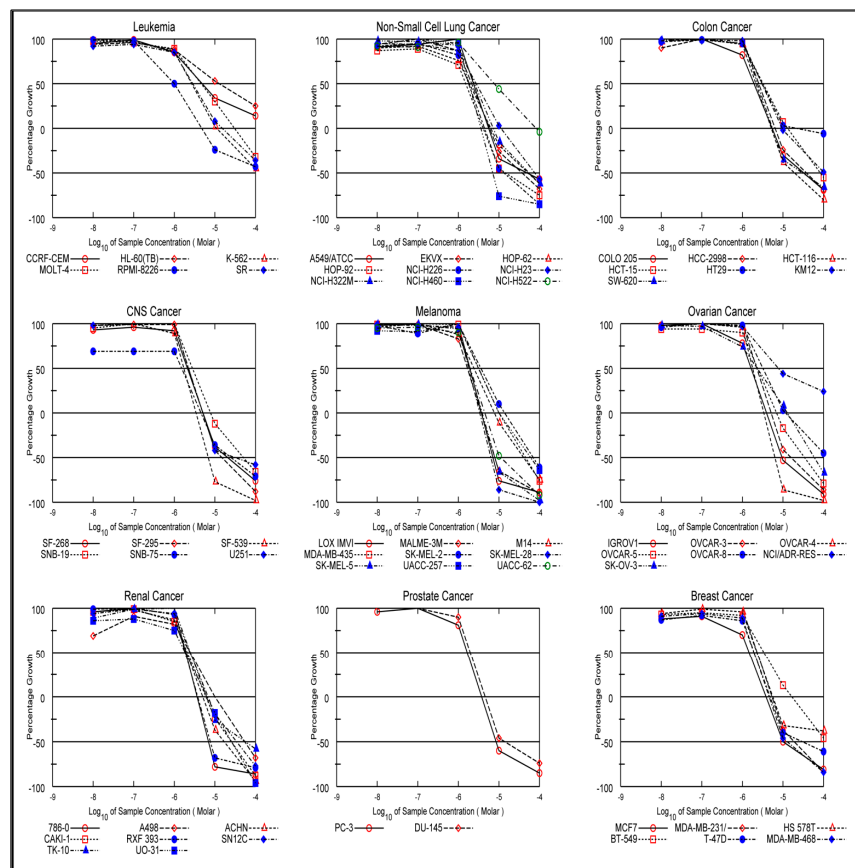


Figure 5. Dose–response curves for all cell lines for compound 4g.

The standard for a compound's selectivity depends on the ratio resulting from dividing the entire MID panel (the average sensitivity of all cell lines to the test agent) (μM) by its individual MID (μM) subpanel. Ratios between 3 and 6 indicate moderate selectivity; ratios >6 refer to high selectivity towards the respective cell line, whereas compounds that do not meet either of these criteria are rated nonselective [44]. In vitro five-dose assay screening results are illustrated in Tables 2–4.

Compounds 4c, 4e, and 4g achieved complete cell death for different cancer cell lines. These compounds were thus selected for five-dose testing to screen their activity at five different concentrations. In addition, these compounds have a broad and potent spectrum of anti-cancer activity without selectivity toward any of the tested cancer cell lines.

Compound 4c revealed a GI_{50} value ranging from 0.420 to 8.99 μM toward the majority of the examined cell lines, a selectivity ratio from 0.57 to 5.96, a TGI from 2.41 to >100 μM , and a LC_{50} from 9.61 to >100 μM . Compound 4c showed moderate selectivity toward leukemia cell lines as the ratio between selectivity towards cancer subpanels = 5.96 (Table 2 and Figure 3).

In the same context, compound 4e was found to have broad-spectrum cell growth inhibition activity against most of the tested tumor subpanels, with GI_{50} values ranging from 0.97 to 4.93 μM , selectivity ratio ranging from 0.79 to 1.35, TGI from 2.13 to 9.03 μM , and LC_{50} from 5.73 to 77.90 μM (Table 3 and Figure 4).

Finally, compound 4g exhibited potent antitumor activity toward all the examined cell lines, with GI_{50} values ranging between 0.997 and 7.81 μM , selectivity ratio ranging from 0.87 to 1.39, TGI from 1.64 to >100 μM , and LC_{50} between 6.33 and >100 μM (Table 4 and Figure 5).

Table 2. Screening results of in vitro five doses in μM for compound **4c**.

Panel Name	Cell Name	GI ₅₀			TGI ^b	LC ₅₀
		Conc. per Cell Line	Subpanel MID ^b	Selectivity Ratio (MID ^a /MID ^b)		
Leukemia	CCRF-CEM	0.591	0.55	5.96	17.70	>100
	HL-60(TB)	0.797			5.73	>100
	K-562	0.499			8.44	54.60
	MOLT-4	0.421			3.85	>100
	RPMI-8226	0.461			2.41	>100
	SR	0.566			4.87	>100
Non-Small Cell Lung Cancer	A549/ATCC	3.98	3.52	0.93	24.10	>100
	EKVX	3.19			21.20	>100
	HOP-62	6.42			30.80	>100
	HOP-92	0.985			7.52	47.60
	NCI-H226	5.03			31.20	>100
	NCI-H23	1.85			11.50	67.60
	NCI-H322M	5.04			26.40	>100
	NCI-H460	2.10			4.49	9.61
	NCI-H522	3.05			11.70	>100
Colon Cancer	COLO 205	1.46	2.53	1.30	3.76	-
	HCC-2998	1.48			4.81	28.10
	HCT-116	2.25			6.90	82.30
	HCT-15	2.60			13.30	>100
	HT29	3.02			>100	>100
	KM12	3.43			13.00	82.60
	SW-620	3.44			10.80	55.60
CNS Cancer	SF-268	4.97	4.05	0.81	24.30	>100
	SF-295	3.40			13.10	43.80
	SF-539	2.73			10.10	33.80
	SNB-19	7.48			32.40	>100
	SNB-75	2.78			18.00	>100
	U251	2.92			10.80	48.00
Melanoma	LOX IMVI	1.34	2.87	1.14	3.22	7.74
	MALME-3M	1.48			5.00	23.60
	M14	2.73			9.13	67.10
	MDA-MB-435	3.57			12.20	47.60
	SK-MEL-2	5.10			21.90	81.30
	SK-MEL-28	2.60			10.30	39.00
	SK-MEL-5	2.11			4.78	12.80
	UACC-257	3.37			15.40	>100
	UACC-62	3.51			14.90	52.20

Table 2. Cont.

Panel Name	Cell Name	GI ₅₀		TGI ^b	LC ₅₀	
		Conc. per Cell Line	Subpanel MID ^b			Selectivity Ratio (MID ^a /MID ^b)
Ovarian Cancer	IGROV1	3.04	5.77	0.57	11.40	52.40
	OVCAR-3	3.25			8.23	40.90
	OVCAR-4	2.75			8.64	>100
	OVCAR-5	4.43			37.80	>100
	OVCAR-8	2.89			13.70	>100
	NCI/ADR-RES	12.60			>100	>100
	SK-OV-3	11.40			39.60	>100
Renal Cancer	786-0	3.92	4.30	0.76	15.70	58.20
	A498	2.40			7.75	36.40
	ACHN	4.36			31.90	>100
	CAKI-1	3.49			16.10	85.40
	RXF 393	2.03			7.07	34.00
	SN12C	3.14			18.10	>100
	TK-10	8.99			40.23	>100
	UO-31	6.05			38.30	>100
Prostate Cancer	PC-3	0.420	2.20	1.49	15.30	>100
	DU-145	3.99			21.40	>100
Breast Cancer	MCF7	0.582	2.50	1.31	2.85	13.30
	MDA-MB-231	3.75			13.90	69.10
	HS 578T	3.58			24.40	>100
	BT-549	2.84			9.70	50.50
	T-47D	2.29			7.21	>100
	MDA-MB-468	1.97			4.79	17.70
MID ^a		3.28				

MID^a: Average sensitivity of all cell lines in μM . MID^b: Average sensitivity of all subpanel cell lines in μM .

Table 3. Screening results of in vitro five doses in μM for compound 4e.

Panel Name	Cell Name	GI ₅₀		TGI ^b	LC ₅₀
		Conc. per Cell Line	Subpanel MID ^b		
Leukemia	CCRF-CEM	1.43	1.33	1.35	9.03
	HL-60(TB)	1.85			6.53
	K-562	0.969			3.23
	MOLT-4	1.55			4.81
	RPMI-8226	1.16			3.52
	SR	1.01			3.95

Table 3. Cont.

Panel Name	Cell Name	GI ₅₀			TGI ^b	LC ₅₀
		Conc. per Cell Line	Subpanel MID ^b	Selectivity Ratio (MID ^a /MID ^b)		
Non-Small Cell Lung Cancer	A549/ATCC	2.16	1.9	0.94	4.53	9.52
	EKVX	1.82			3.68	7.43
	HOP-62	1.80			4.10	9.32
	HOP-92	1.67			3.53	7.47
	NCI-H226	1.77			3.48	6.86
	NCI-H23	1.93			4.43	10.80
	NCI-H322M	2.15			5.26	19.80
	NCI-H460	1.56			3.13	6.30
	NCI-H522	2.24			5.85	77.90
Colon Cancer	COLO 205	1.55	1.72	1.04	3.25	8.82
	HCC-2998	1.82			3.44	6.50
	HCT-116	1.45			3.51	8.52
	HCT-15	1.42			3.03	6.46
	HT29	1.97			5.47	44.10
	KM12	2.03			4.46	9.80
	SW-620	1.83			3.69	7.43
	CNS Cancer	SF-268			2.08	1.94
SF-295		1.86	3.50	6.56		
SF-539		1.70	3.17	5.93		
SNB-19		2.34	5.06	13.30		
SNB-75		1.59	3.62	8.22		
U251		2.10	4.18	8.35		
Melanoma		LOX IMVI	1.57	1.77	1.01	
	MALME-3M	1.55	3.15			6.41
	M14	1.65	3.84			8.96
	MDA-MB-435	1.73	3.35			6.49
	SK-MEL-2	2.09	4.64			11.40
	SK-MEL-28	1.58	3.04			5.86
	SK-MEL-5	1.81	3.39			6.33
	UACC-257	2.14	4.72			11.60
	UACC-62	1.78	3.40			6.49
Ovarian Cancer	IGROV1	1.50	1.81	0.99	2.98	5.92
	OVCAR-3	1.84			3.39	6.28
	OVCAR-4	1.74			3.29	6.23
	OVCAR-5	1.85			3.97	8.52
	OVCAR-8	2.11			4.74	16.50
	NCI/ADR-RES	1.83			4.43	12.70
	SK-OV-3	1.79			4.31	11.00

Table 3. Cont.

Panel Name	Cell Name	GI ₅₀		TGI ^b	LC ₅₀	
		Conc. per Cell Line	Subpanel MID ^b			Selectivity Ratio (MID ^a /MID ^b)
Renal Cancer	786-0	1.79	1.77	1.01	3.48	6.76
	A498	1.74			4.94	19.60
	ACHN	1.73			3.17	5.78
	CAKI-1	1.70			3.29	6.37
	RXF 393	1.61			3.27	6.64
	SN12C	1.96			4.26	9.26
	TK-10	2.06			4.30	8.98
	UO-31	1.57			2.99	5.67
Prostate Cancer	PC-3	1.55	1.65	1.08	3.30	7.05
	DU-145	1.76			3.44	6.70
Breast Cancer	MCF7	1.15	2.27	0.79	2.56	5.73
	MDA-MB-231	1.85			3.77	7.67
	HS 578T	2.28			5.27	63.20
	BT-549	4.93			2.13	75.80
	T-47D	1.79			3.83	8.21
	MDA-MB-468	1.64			3.23	6.36
MID ^a		1.79				

MID^a: Average sensitivity of all cell lines in μM . MID^b: Average sensitivity of all subpanel cell lines in μM .

Table 4. Screening results of five doses in μM for compound 4g.

Panel Name	Cell Name	GI ₅₀		TGI ^b	LC ₅₀	
		Conc. per Cell Line	Subpanel MID ^b			Selectivity Ratio (MID ^a /MID ^b)
Leukemia	CCRF-CEM	4.90	2.88	0.87	>100	>100
	HL-60(TB)	1.30			>100	>100
	K-562	2.69			11.20	>100
	MOLT-4	4.53			30.40	>100
	RPMI-8226	0.997			4.77	>100
	SR	2.87			15.40	>100
Non-Small Cell Lung Cancer	A549/ATCC	2.46	2.71	0.93	5.65	48.10
	EKVX	2.11			5.84	36.60
	HOP-62	1.88			6.37	53.60
	HOP-92	1.52			4.11	15.10
	NCI-H226	1.79			4.43	13.50
	NCI-H23	2.74			11.20	75.90
	NCI-H322M	2.49			7.20	55.40
	NCI-H460	1.83			3.59	7.05
	NCI-H522	7.60			81.50	>100

Table 4. Cont.

Panel Name	Cell Name	GI ₅₀			TGI ^b	LC ₅₀
		Conc. per Cell Line	Subpanel MID ^b	Selectivity Ratio (MID ^a /MID ^b)		
Colon Cancer	COLO 205	1.94	2.63	0.96	5.43	33.00
	HCC-2998	2.57			6.44	37.80
	HCT-116	2.14			5.12	18.90
	HCT-15	3.24			12.80	82.00
	HT29	3.09			22.10	>100
	KM12	3.03			9.58	>100
	SW-620	2.41			5.58	31.00
CNS Cancer	SF-268	2.09	2.19	1.15	5.10	20.80
	SF-295	2.23			5.05	15.00
	SF-539	1.73			3.46	6.92
	SNB-19	3.13			7.95	49.60
	SNB-75	1.53			4.57	25.70
	U251	2.43			5.22	30.40
Melanoma	LOX IMVI	2.19	2.43	1.04	4.01	7.34
	MALME-3M	1.67			3.63	7.90
	M14	2.41			7.76	40.50
	MDA-MB-435	3.11			9.84	45.50
	SK-MEL-2	3.59			13.80	69.70
	SK-MEL-28	1.79			3.37	6.35
	SK-MEL-5	1.89			3.88	7.95
	UACC-257	3.27			10.10	60.00
UACC-62	1.94	4.47	10.90			
Ovarian Cancer	IGROV1	1.65	3.05	0.83	3.97	9.57
	OVCAR-3	2.28			5.14	16.00
	OVCAR-4	1.79			3.36	6.33
	OVCAR-5	2.35			6.89	33.50
	OVCAR-8	3.19			11.50	>100
	NCI/ADR-RES	7.81			>100	>100
	SK-OV-3	2.30			12.80	59.10
Renal Cancer	786-0	1.93	2.20	1.14	3.67	6.98
	A498	2.45			9.87	54.60
	ACHN	2.00			5.04	16.90
	CAKI-1	2.10			6.03	25.60
	RXF 393	1.86			3.80	7.76
	SN12C	2.46			6.79	33.90
	TK-10	2.95			6.59	56.70
	UO-31	1.86			6.38	25.70

Table 4. Cont.

Panel Name	Cell Name	GI ₅₀			TGI ^b	LC ₅₀
		Conc. per Cell Line	Subpanel MID ^b	Selectivity Ratio (MID ^a /MID ^b)		
Prostate Cancer	PC-3	1.65	1.81	1.39	3.76	8.55
	DU-145	1.98			4.61	14.00
Breast Cancer	MCF7	1.47	2.24	1.12	3.84	10.00
	MDA-MB-231	2.42			5.45	18.80
	HS 578T	2.30			5.64	>100
	BT-549	3.36			1.64	>100
	T-47D	1.94			4.83	30.00
	MDA-MB-468	1.94			4.55	12.60
	MID ^a				2.52	

MID^a: Average sensitivity of all cell lines in μM . MID^b: Average sensitivity of all subpanel cell lines in μM .

2.2.2. EGFR Inhibitory Assay

The most effective derivatives from in vitro anti-cancer assay (**4c**, **4d**, **4e**, **4g**, and **4h**) were tested for EGFR inhibition as a potential target for antiproliferative action [45,46]. Table 5 shows the results as IC₅₀ values versus erlotinib as the control drug.

Table 5. IC₅₀ values of compounds **4c**, **4d**, **4e**, **4g**, and **4h** against EGFR and BRAF^{V600E}.

Compd. No.	EGFR Inhibition IC ₅₀ $\pm$ SEM (μM)	BRAF ^{V600E} Inhibition IC ₅₀ $\pm$ SEM (μM)
4c	0.11 $\pm$ 0.01	0.31 $\pm$ 0.07
4d	0.68 $\pm$ 0.05	0.95 $\pm$ 0.07
4e	0.09 $\pm$ 0.01	0.20 $\pm$ 0.02
4g	0.27 $\pm$ 0.03	0.60 $\pm$ 0.05
4h	0.56 $\pm$ 0.05	0.82 $\pm$ 0.06
Erlotinib	0.08 $\pm$ 0.04	0.06 $\pm$ 0.02
Vemurafenib	--	0.03 $\pm$ 0.01

The results showed that the investigated compounds had potential EGFR inhibitory action, with IC₅₀ values ranging from 0.09 to 0.68 μM compared to erlotinib, which has an IC₅₀ value of 0.08 μM . Furthermore, the findings of this investigation are consistent with those of an in vitro five-dose inhibition assay in which compound **4e** was the most potent EGFR inhibitor with an IC₅₀ value of 0.09 μM , equivalent to the reference erlotinib. Compound **4c** was 1.2-fold less potent than **4e**, having an IC₅₀ value of 0.11 μM . Compounds **4d**, **4g**, and **4h** inhibited EGFR in a weak to moderate approach, with IC₅₀ values of 0.68 μM , 0.27 μM , and 0.56 μM , respectively, being seven-fold, three-fold, and six-fold less effective than **4e**. These findings suggested that compounds **4c** and **4e** could be promising antiproliferative agents that target the EGFR inhibitory pathway.

2.2.3. BRAF^{V600E} Inhibitory Assay

Compounds **4c**, **4d**, **4e**, **4g**, and **4h** were investigated further as possible BRAF^{V600E} inhibitors [47]. Table 2 displays the IC₅₀ values compared to erlotinib and vemurafenib, which were used as controls. According to Table 5, the compounds tested showed considerable BRAF^{V600E} suppressive action, with IC₅₀ values ranging from 0.20 to 0.85 μM .

In all cases, the tested derivatives were less potent than erlotinib ($IC_{50} = 0.06 \mu M$) and vemurafenib ($IC_{50} = 0.03 \mu M$).

Compounds **4c** and **4e**, which were the most potent derivatives in the antiproliferative and EGFR suppressive assays, were also the most effective derivatives as anti-BRAF^{V600E}, with IC_{50} values of $0.31 \pm 0.07 \mu M$ and $0.20 \pm 0.02 \mu M$, respectively. These findings suggest that compounds **4c** and **4e** have significant antiproliferative activity as dual EGFR/BRAFV^{600E} inhibitors, implying that additional structural modifications may be required to develop a more potent lead molecule for future development.

2.2.4. Apoptosis Induction Assays

One method of treating cancer is to regulate or stop the uncontrolled proliferation of cancer cells. Using the natural dying process of the cell is a highly successful strategy. Targeting apoptosis is effective for many types of cancer because apoptosis evasion is a hallmark of cancer that is not particular to the etiology or type of cancer. Many anti-cancer drugs target distinct stages of the intrinsic and extrinsic apoptotic pathways [48–50]. Compounds **4c** and **4e**, the most effective derivatives in all in vitro investigations, were investigated for their capacity to initiate apoptosis.

Assay for Caspase-3 Activation

Caspase-3 is a key caspase that cleaves numerous cell proteins, resulting in apoptosis [51]. Compounds **4c** and **4e** were tested as activators of caspase-3 against the prostatic cancer (PC-3) cell line, compared to staurosporine as the control drug [52], with the results presented in Table 6. Compounds **4c** and **4e** showed high caspase-3 protein overexpression levels of 365 ± 12 and 470 ± 15 Pg/mL, respectively. They elevated the protein caspase-3 in the PC-3 cancer cell line by around seven to nine times compared to untreated control cells. Compared to reference staurosporine, which had a caspase-3 level overexpression of 260 ± 5 pg/mL, compounds **4c** and **4e** were more active as caspase-3 activators. These findings suggested that the compounds under investigation have apoptotic potential, which could provide additional support for their antiproliferative impact.

Table 6. Caspase-3, caspase-8, Bax, and Bcl-2 levels for compounds **4c**, **4e**, and staurosporine on prostatic cancer (PC-3) cell line.

Compd. No.	Caspase-3		Caspase-8		Bax		Bcl-2	
	Conc (Pg/mL)	Fold Change	Conc (ng/mL)	Fold Change	Conc (Pg/mL)	Fold Change	Conc (ng/mL)	Fold Reduction
4c	365 ± 12	7	1.90	16	185	26	0.80	4
4e	470 ± 15	9	2.20	18	220	31	0.75	4
Staurosporine	260 ± 5	5	1.65	14	170	24	1.00	3
Control	52	1	0.12	1	7	1	3.00	1

Assays for Caspase-8, Bax, and Bcl2 Levels

Using staurosporine as a control, compounds **4c** and **4e** were investigated further for their impact on caspase-8, Bax, and anti-apoptotic Bcl-2 levels against the prostate cancer (PC-3) cell line. Table 3 lists the results. Compound **4e** (2.20 ng/mL) had the highest level of caspase-8 overexpression, followed by compound **4c** (1.90 ng/mL) and the reference drug staurosporine (1.65 ng/mL). Compounds **4c** and **4e**, increased caspase-8 levels by 16 and 18 times, respectively, compared to the untreated control cell.

Compared to untreated PC-3 cancer cells, compounds **4c** and **4e** increased Bax levels by 26- and 31-fold (185 pg/mL and 220 pg/mL, respectively). Both are more active than staurosporine, which induces Bax levels up to 170 pg/mL, and a 24-fold increase over untreated cells. Finally, compounds **4c** and **4e** induced equipotent down-regulation of anti-apoptotic Bcl-2 protein levels in the PC-3 cancer cell line when compared to stau-

rosporine. These findings suggest that compounds **4c** and **4e** act as caspase-3, caspase-8, and Bax activators and down-regulators of the anti-apoptotic Bcl-2, classifying them as apoptotic triggers.

3. Docking Study

The most active compounds, **4c** and **4e**, have multiple structural options regarding their probable binding mechanism and interactions with EGFR and BRAF active sites. These possibilities were investigated by molecular modeling studies using the Molecular Operating Environment (MOE) software (version 09.2022) [53]. Furthermore, the docking methodology was validated, and the co-crystallized ligand erlotinib, GW572016, was re-docked into the active site using the EGFR domain (PDB ID: 1 M17). On the other hand, the structure of BRAF kinase in a complex with the inhibitor vemurafenib was used to validate the docking protocol for BRAF^{V600} (PDB ID: 3OG7) [54]. With a docking score (S) of -13.4442 , the re-docked ligand displayed an RMSD of 0.8429 between the docked pose and the co-crystallized ligand for EGFR. When the co-crystallized ligand was docked, the RMSD between the two was 0.2808 (docking score (S) = -10.2469) for the BRAF^{V600}.

The most potent molecules, **4c** and **4e**, were docked into the EGFR's active site along with erlotinib (as a positive control). Tables 7 and 8 present the results of the docking experiments together with the kind of interactions, the distances (in Å) between the interacting residues, and the binding affinity (in kcal mol⁻¹).

Table 7. The target compound's binding energy score throughout the EGFR docking study.

EGFR (PDB: 1 M17)				
Compound	Binding Affinity (kcal mol ⁻¹)	Distance from the Main Residue (in Å)	Interaction Type	Amino Acids Residue
4c	−0.5	3.60	H-donor	MET 742
		3.30	H-acceptor	MET 769
		4.09	pi-H	LEU 694
		4.36	pi-H	LEU 694
4e	−1.5	3.35	H-acceptor	MET 769
		3.69	pi-H	LEU 694
Erlotinib	−1.9	3.35	H-acceptor	MET 769
		3.69	pi-H	LEU 694

Table 8. The target compounds' binding energy scores throughout the BRAF^{V600} docking study.

BRAF ^{V600E} (PDB: 3OG7)				
Compd.	Binding Affinity (kcal mol ⁻¹)	Distance from the Main Residue (Å)	Interaction Type	Amino Acids Residue
4c	−5.4	2.92	H-donor	CYS 532
		3.12	H-acceptor	LYS 483
		3.66	pi-H	LYS 483
4e	−1.9	3.07	H-acceptor	CYS 532
		3.52	pi-H	CYS 532
		3.54	pi-H	SER 535
Vemurafenib	−3.5	3.15	H-donor	GLN 530
		3.14	H-acceptor	CYS 532
		3.15	H-acceptor	ASP 594
		3.08	H-acceptor	LYS 483
		2.98	H-acceptor	GLY 596
		3.23	Ionic	LYS 483
		3.08	Ionic	LYS 483
		3.88	Ionic	LYS 483

Compounds **4c** and **4e** demonstrated substantial interaction with Met793 and other important interacting amino acids required for inhibition of EGFR when compared to co-docked ligand erlotinib as a positive control. Compared to erlotinib, which has a binding affinity of $-1.9 \text{ kcal mol}^{-1}$, compounds **4c** and **4e** had strong interaction scores of -0.5 and $-1.5 \text{ kcal mol}^{-1}$, respectively. The many binding interactions with the critical amino acid within the EGFR active site, particularly the gatekeeper Met793 and LEU 694 residues, can account for these significant binding affinities (Figures 6–8).

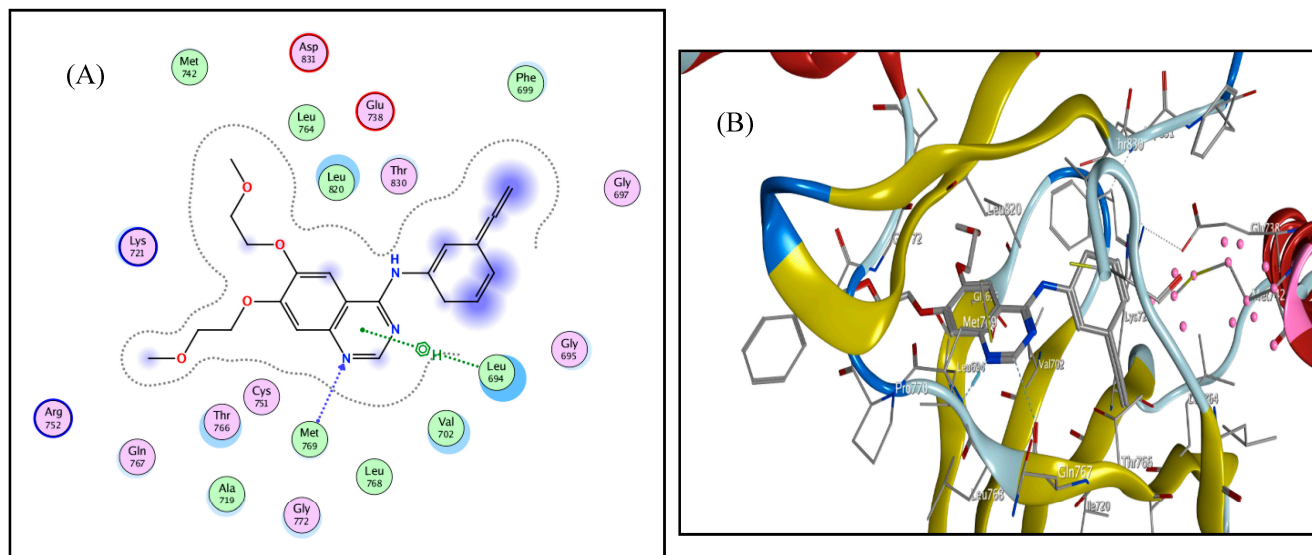


Figure 6. (A) 2D interaction, (B) 3D interaction of erlotinib inside EGFR binding pocket.

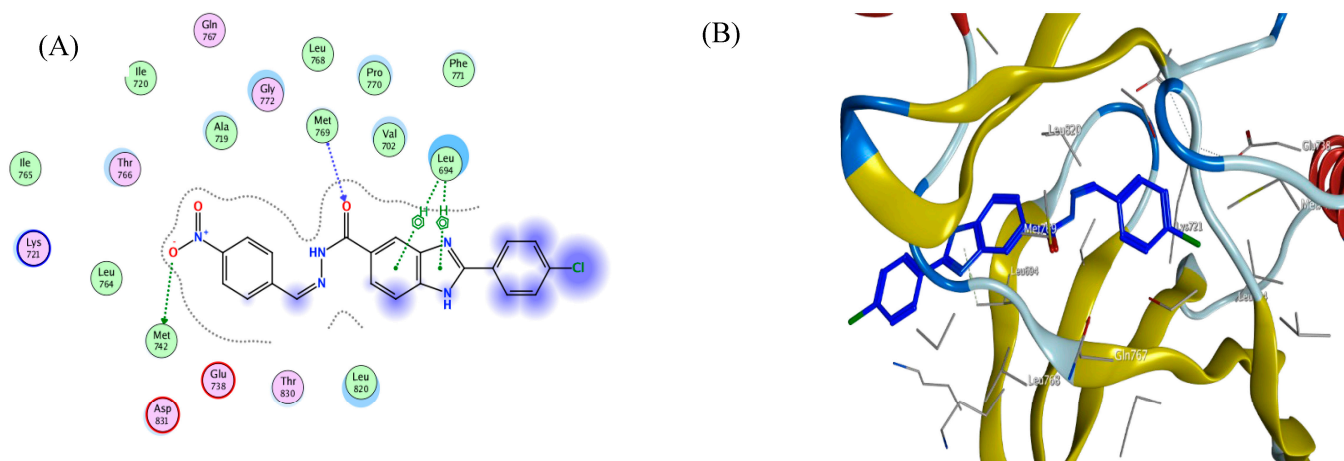


Figure 7. (A) 2D interaction, (B) 3D interaction of **4c** inside EGFR binding pocket.

Additionally, vemurafenib was docked into the BRAF^{V600E} active site among the three most active molecules, **4c** and **4e**. The docking study data are compiled in Table 8, including information on the kind of interactions, binding affinity (in kcal mol^{-1}), and distances (in Å) from the interacting residues. A co-docked ligand, vemurafenib, was a positive control in comparing molecules **4c** and **4e**. Strong interactions between these drugs and CYS 532 and other essential interacting amino acids required for BRAF^{V600} were shown. With binding affinities of -5.4 and $-1.9 \text{ kcal mol}^{-1}$, respectively, compounds **4c** and **4e** showed considerable interaction scores compared to Vemurafenib, which had a binding affinity of $-3.5 \text{ kcal mol}^{-1}$. The high binding scores of these compounds were established by a significant interaction between the synthesized compounds and the essential amino acid in the BRAF active site, with CYS 532 (Figures 9–11).

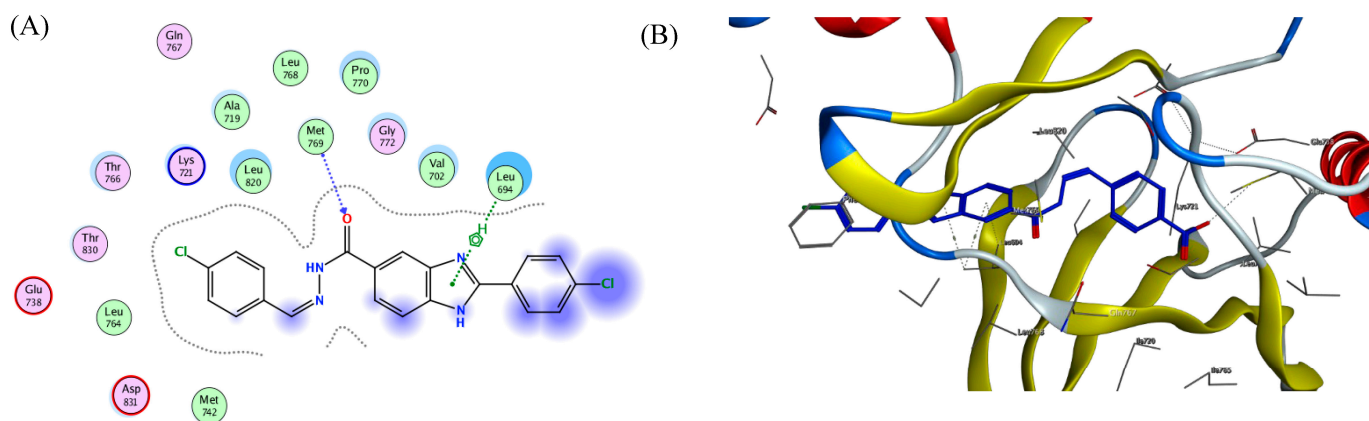


Figure 8. (A) 2D interaction, (B) 3D interaction of 4e inside EGFR binding pocket.

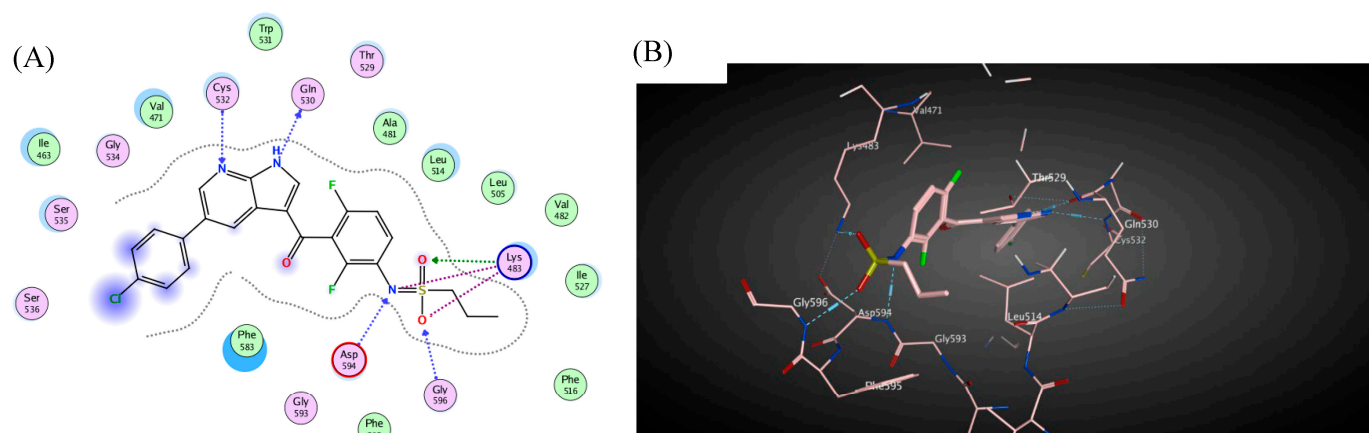


Figure 9. (A) 2D interaction, (B) 3D interaction of vemurafenib inside BRAF^{V600} binding pocket.

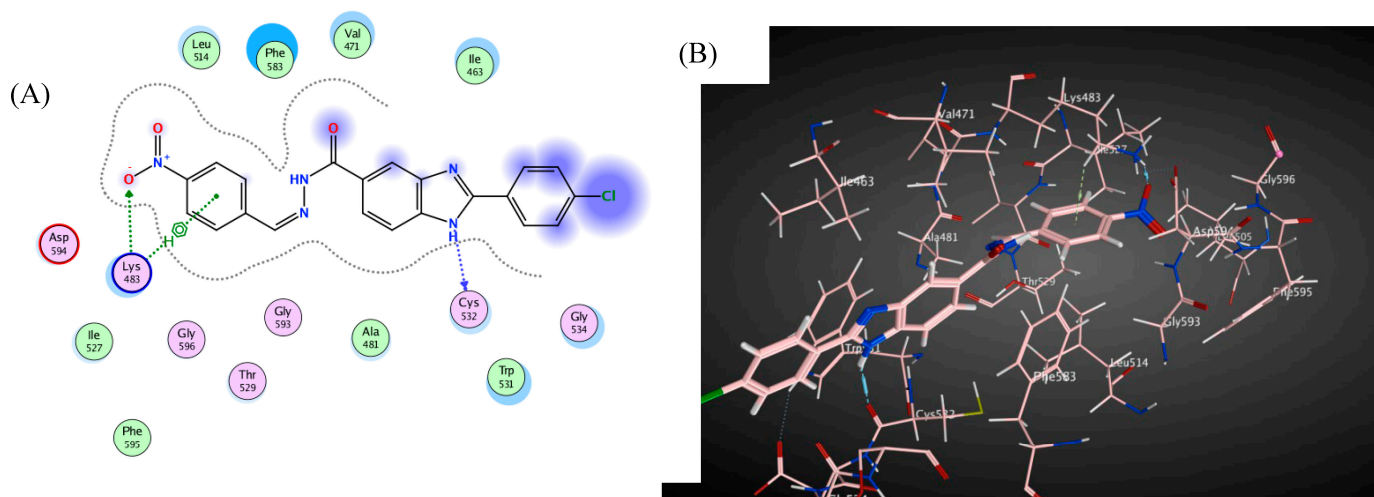


Figure 10. (A) 2D interaction, (B) 3D interaction of 4c inside BRAF^{V600} binding pocket.

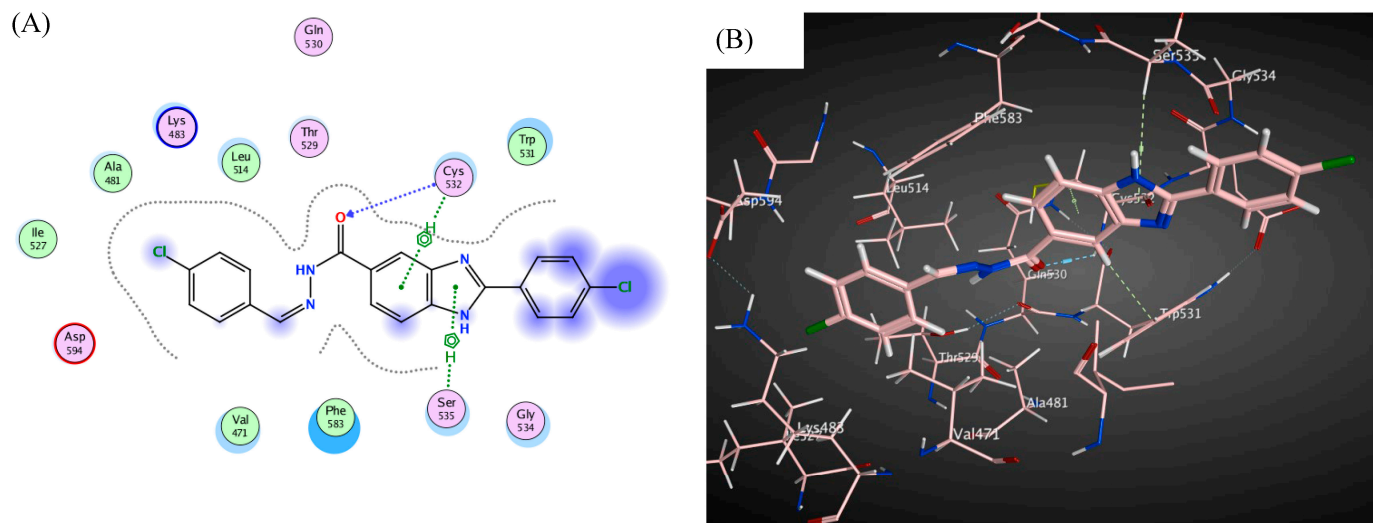


Figure 11. (A) 2D interaction, (B) 3D interaction of **4e** inside BRAF^{V600} binding pocket.

4. Conclusions

Finally, a new class of novel compounds with benzimidazole scaffolds (**4a–j**, **5**, and **6**) was synthesized and tested for anticancer activity. NCI chose compounds **4c**, **4e**, and **4g** for five dose tests, with selectivity ratios ranging from 0.60 to 5.80 at the GI₅₀ levels. Compounds **4c** and **4e** showed significant EGFR and BRAF^{V600E} inhibition. Apoptotic marker assay results indicate that compounds **4c** and **4e** act as caspase-3, caspase-8, and Bax activators as well as down-regulators of the anti-apoptotic Bcl-2, classifying them as apoptotic triggers. Molecular docking simulations of **4c** and **4e** also revealed good binding modes within the EGFR and BRAF^{V600} active sites. To obtain a more effective lead compound for future development, compounds **4c** and **4e** require further structural modifications as well as the synthesis of more derivatives.

5. Experimental

5.1. Chemistry

The melting point was obtained utilizing a Thomas-Hoover capillary device and is uncorrected. The infrared (IR) spectra were recorded as films on KBr plates using the FT-IR spectrometer. Thin-layer chromatography was used to evaluate the reaction mixture, homogeneity, and purity of the synthesized compounds (Merck, Darmstadt, Germany).

General Details: See Supplementary Materials.

The key intermediates **1–3** were prepared according to previously reported procedures [34,36,38].

5.1.1. General Procedure for Synthesis of N'-(substituted benzylidene)-2-(4-chlorophenyl)-1H-benzimidazole-5-carbohydrazide (**4a–j**)

Equimolar quantities of the appropriate compound **3** (2 mmol, 0.50 g) and the appropriate substituted benzaldehyde in absolute ethanol (25 mL) were refluxed for 6 h in the presence of a catalytic amount of glacial acetic acid. The resulting crude solid was filtered off and recrystallized from absolute ethanol to afford compounds **4a–j** in a good yield.

N'-Benzylidene-2-(4-chlorophenyl)-1H-benzimidazole-5-carbohydrazide (**4a**)

Yellow powder, 80% yield, m.p. 207–209 °C, IR (KBr, $\nu_{\max}$ cm^{−1}): 3426, 3393 (NH benzimidazole, NH hydrazone), 3073 (CH aromatic), 1621 (C=O), 1606 (C=C), 1455 (C=N), 1373–1502 (N-O); ¹H NMR (DMSO-*d*₆, 400 MHz, δ ppm): 6.70 (d, 2H, *J* = 8.00 Hz, Ar-2H), 7.45–7.50 (m, 4H, Ar-H), 7.62 (d, 1H, *J* = 8.00 Hz, Ar-H), 7.76 (d, 2H, *J* = 8.00 Hz, Ar-H), 7.88 (d, 2H, *J* = 8.00 Hz, Ar-H), 7.95 (d, 1H, *J* = 8.00 Hz, Ar-H), 8.48 (s, 1H, CH=NH), 11.90 (s, 1H, NH, D₂O exchangeable), 12.82 (s, 1H, NH benzimidazole, D₂O exchangeable); ¹³C NMR

(DMSO-*d*₆, 100 MHz, δ ppm): 113.65, 116.48, 121.99, 126.37, 127.27, 128.28, 129.15, 129.79, 130.06, 130.98, 133.14, 134.69, 147.10, 151.39, 164.40 (aromatic carbons), 168.14 (C=N), 172.76 (C=O); elemental analysis calculated for C₂₁H₁₅N₅O₃ (M.Wt. 385.38): C, 65.45; H, 3.92; N, 18.17; found C, 65.71; H, 4.13; N, 18.42.

N'-(4-(Dimethylamino)benzylidene)-2-(4-chlorophenyl)-1H-benzimidazole-5-carbohydrazide (4b)

Buff powder, 80% yield, m.p. 283–285 °C, IR (KBr, $\nu_{\max}$, cm^{−1}): 3385–3226 (NH benzimidazole, NH hydrazone), 3029 (CH aromatic), 1663 (C=O), 1626 (C=C), 1470 (C=N); ¹H NMR (DMSO-*d*₆, 500 MHz, δ ppm): 2.96 (s, 6H, 2 CH₃), 6.76 (d, 2H, *J* = 8.50 Hz, Ar-H), 7.57 (d, 2H, *J* = 8.50 Hz, Ar-H), 7.66 (d, 3H, *J* = 8.50 Hz, Ar-H), 7.81 (d, 1H, *J* = 8.00 Hz, Ar-H), 8.21 (d, 2H, *J* = 8.50 Hz, Ar-H), 8.33 (s, 1H, CH=NH), 11.65 (s, 1H, NH hydrazone D₂O exchangeable), 13.42 (s, 1H, NH benzimidazole, D₂O exchangeable); ¹³C NMR (DMSO-*d*₆, 125 MHz, δ ppm): 56.54 (CH₃), 110.95, 111.51, 111.54, 112.27, 122.09, 128.88, 128.94, 129.03, 129.68, 135.33, 135.55, 148.83, 151.38, 151.95, 152.92 (aromatic carbons), 163.43 (C=N), 163.73 (C=O); elemental analysis calculated for C₂₃H₂₀ClN₅O (M.Wt. 417.90): C, 66.11; H, 4.82; N, 16.76; found C, 66.32; H, 4.52; N, 16.99.

N'-(4-Nitrobenzylidene)-2-(4-chlorophenyl)-1H-benzimidazole-5-carbohydrazide (4c)

Brown powder, 73% yield, m.p. 297–299 °C, IR (KBr, $\nu_{\max}$, cm^{−1}): 3427–3327 (NH benzimidazole, NH hydrazone), 3054 (CH aromatic), 1652 (C=O), 1614 (C=C), 1467 (C=N), 1384–1514 (N-O); ¹H NMR (DMSO-*d*₆, 500 MHz, δ ppm): 7.66 (d, 2H, *J* = 8.50 Hz, Ar-H), 7.82–7.85 (m, 1H, Ar-H), 8.01 (d, 2H, *J* = 8.50 Hz, Ar-H), 8.21 (d, 3H, *J* = 9.00 Hz, Ar-H), 8.31 (d, 3H, *J* = 8.50 Hz, Ar-H), 8.57 (s, 1H, CH=NH), 12.25 (s, 1H, NH hydrazone, D₂O exchangeable), 13.44 (s, 1H, benzimidazole NH, D₂O exchangeable); ¹³C NMR (DMSO-*d*₆, 125 MHz, δ ppm): 123.89, 124.18, 124.56, 124.82, 127.90, 128.26, 128.44, 128.85, 128.92, 129.45, 129.70, 135.63, 141.21, 145.36, 148.02 (aromatic carbons), 148.24 (C=N), 164.40 (C=O); elemental analysis calculated for C₂₁H₁₄ClN₅O₃ (M.Wt. 419.83): C, 60.08; H, 3.36; N, 16.68; found C, 60.28; H, 3.54; N, 16.90.

N'-(4-Fluorobenzylidene)-2-(4-chlorophenyl)-1H-benzimidazole-5-carbohydrazide (4d)

Brown powder, 60% yield, m.p. 285–287 °C, IR (KBr, $\nu_{\max}$, cm^{−1}): 3423–3228 (NH benzimidazole, NH hydrazone), 3061 (CH aromatic), 1663 (C=O), 1625 (C=C), 1470 (C=N); ¹H NMR (DMSO-*d*₆, 400 MHz, δ ppm): 7.31 (t, *J* = 7.20 Hz, 2H, Ar-H), 7.66 (d, 3H, *J* = 6.80 Hz, Ar-H), 7.81–7.84 (m, 3H, Ar-H), 8.21 (d, 3H, *J* = 6.80 Hz, Ar-H), 8.48 (s, 1H, CH=NH), 11.97 (s, 1H, NH hydrazone, D₂O exchangeable), 13.41 (s, 1H, benzimidazole NH, D₂O exchangeable); ¹³C NMR (DMSO-*d*₆, 100 MHz, δ ppm): 111.84, 116.31, 116.48, 116.82, 128.53, 128.89, 129.06, 129.70, 129.78, 131.47, 135.60, 146.47, 146.81, 152.91, 162.57 (aromatic carbons), 164.18 (C=N), 164.54 (C=O); elemental analysis calculated for C₂₁H₁₄ClFN₄O (M.Wt. 392.82): C, 64.21; H, 3.59; N, 14.26; found C, 64.42; H, 3.73; N, 14.39.

N'-(4-Chlorobenzylidene)-2-(4-chlorophenyl)-1H-benzimidazole-5-carbohydrazide (4e)

Brown powder, 85% yield, m.p. 265–267 °C, IR (KBr, $\nu_{\max}$, cm^{−1}): 3434–3393 (NH benzimidazole, NH hydrazone), 3086 (CH aromatic), 1640 (C=O), 1604 (C=C), 1489 (C=N); ¹H NMR (DMSO-*d*₆, 500 MHz, δ ppm): 7.53 (d, 2H, *J* = 8.50 Hz, Ar-H), 7.66 (d, 2H, *J* = 8.50 Hz, Ar-H), 7.71–7.73 (m, 1H, Ar-H), 7.78 (d, 2H, *J* = 8.50 Hz, Ar-H), 7.83–7.84 (m, 1H, Ar-H), 8.21 (d, 3H, *J* = 8.50 Hz, Ar-H), 8.47 (s, 1H, CH=NH), 12.02 (s, 1H, NH hydrazone, D₂O exchangeable), 13.40 (s, 1H, benzimidazole NH, D₂O exchangeable); ¹³C NMR (DMSO-*d*₆, 125 MHz, δ ppm): 128.25, 128.90, 129.19, 129.42, 129.69, 129.78, 131.60, 131.67, 132.98, 133.80, 134.70, 134.93, 135.60, 146.59, 152.65 (aromatic carbons), 164.21 (C=N), 172.72 (C=O); elemental analysis calculated for C₂₁H₁₄Cl₂N₄O (M.Wt. 409.27): C, 61.63; H, 3.45; N, 13.69; found C, 61.91; H, 3.32; N, 13.70.

N'-(2,4-Dichlorobenzylidene)-2-(4-chlorophenyl)-1H-benzimidazole-5-carbohydrazide (**4f**)

Yellow powder, 79% yield, m.p. 294–297 °C, IR (KBr, $\nu_{\max}$, cm^{-1}): 3459–3393 (NH benzimidazole, NH hydrazone), 3025 (CH aromatic), 1663 (C=O), 1620 (C=C), 1467 (C=N); ^1H NMR (DMSO- d_6 , 500 MHz, δ ppm): 7.52 (d, 1H, J = 8.50 Hz, Ar-H), 7.64–7.70 (m, 4H, Ar-H), 7.85 (s, 1H, Ar-H), 8.06 (d, 1H, J = 8.50 Hz, Ar-H), 8.20 (d, 3H, J = 8.00 Hz, Ar-H), 8.84 (s, 1H, CH=NH), 12.22 (s, 1H, NH hydrazone, D₂O exchangeable), 13.41 (s, 1H, benzimidazole NH, D₂O exchangeable); ^{13}C NMR (DMSO- d_6 , 125 MHz, δ ppm): 111.75, 127.19, 128.48, 128.56, 128.86, 128.91, 129.29, 129.59, 129.68, 129.81, 131.23, 134.29, 135.36, 135.49, 135.61, 138.10, 142.74 (aromatic carbons), 164.20 (C=N), 172.42 (C=O); elemental analysis calculated for C₂₁H₁₃Cl₃N₄O (M.Wt. 443.71): C, 56.85; H, 2.95; N, 12.63; found C, 56.99; H, 3.19; N, 12.91.

N'-(4-Methoxybenzylidene)-2-(4-chlorophenyl)-1H-benzimidazole-5-carbohydrazide (**4g**)

Yellow powder, 82% yield, m.p. 273–275 °C, IR (KBr, $\nu_{\max}$, cm^{-1}): 3449–3390 (NH benzimidazole, NH hydrazone), 3029 (CH aromatic), 2951 (CH aliphatic), 1661 (C=O), 1605 (C=C), 1466 (C=N); ^1H NMR (DMSO- d_6 , 500 MHz, δ ppm): 3.85 (s, 3H, OCH₃), 7.02 (d, 2H, J = 8.00 Hz, Ar-H), 7.65 (d, 2H, J = 8.50 Hz, Ar-H), 7.69–7.72 (m, 3H, Ar-H), 7.81–7.83 (m, 1H, Ar-H), 8.21 (d, 3H, J = 8.50 Hz, Ar-H), 8.42 (s, 1H, CH=NH), 11.87 (s, 1H, NH hydrazone, D₂O exchangeable); ^{13}C NMR (DMSO- d_6 , 125 MHz, δ ppm): 55.91 (OCH₃), 114.42, 114.66, 123.25, 126.83, 127.17, 128.21, 128.79, 128.98, 129.58, 130.72, 132.33, 135.42, 147.61, 152.57, 161.43 (aromatic carbons), 163.93 (C=N), 172.79 (C=O); elemental analysis calculated for C₂₂H₁₇ClN₄O₂ (M.Wt. 404.85): C, 65.27; H, 4.23; N, 13.84; found C, 65.00; H, 4.03; N, 13.98.

N'-(3,4,5-Trimethoxybenzylidene)-2-(4-chlorophenyl)-1H-benzimidazole-5-carbohydrazide (**4h**)

Yellow powder, 84% yield, m.p. 288–290 °C, IR (KBr, $\nu_{\max}$, cm^{-1}): 3440–3318 (NH benzimidazole, NH hydrazone), 3023 (CH aromatic), 2993 (CH aliphatic), 1644 (C=O), 1600 (C=C), 1451 (C=N); ^1H NMR (DMSO- d_6 , 500 MHz, δ ppm): 3.72 (s, 3H, OCH₃), 3.84 (s, 6H, 2OCH₃), 7.08 (s, 2H, Ar-H), 7.65 (d, 2H, J = 8.50 Hz, Ar-H), 7.71–7.73 (m, 1H, Ar-H), 7.82–7.84 (m, 1H, Ar-H), 8.20–8.23 (m, 3H, Ar-H), 8.41 (s, 1H, CH=NH), 11.98 (s, 1H, NH hydrazone, D₂O exchangeable); ^{13}C NMR (DMSO- d_6 , 125 MHz, δ ppm): 56.39 (OCH₃), 60.60 (OCH₃), 104.75, 105.98, 122.82, 122.99, 127.84, 128.89, 128.93, 129.68, 130.42, 135.57, 139.52, 147.95, 152.83, 153.63, 153.90 (aromatic carbons), 161.80 (C=N), 164.25 (C=O); elemental analysis calculated for C₂₄H₂₁ClN₄O₄ (M.Wt. 464.91): C, 62.00; H, 4.55; N, 12.05; found C, 62.25; H, 4.82; N, 12.29.

N'-(4-Hydroxybenzylidene)-2-(4-chlorophenyl)-1H-benzimidazole-5-carbohydrazide (**4i**)

Yellow powder, 80% yield, m.p. 263–265 °C, IR (KBr, $\nu_{\max}$, cm^{-1}): 3403–3344 (NH benzimidazole, NH hydrazone), 3188 (OH), 3073 (CH aromatic), 1645 (C=O), 1613 (C=C), 1447 (C=N); ^1H NMR (DMSO- d_6 , 500 MHz, δ ppm): 6.85 (d, 2H, J = 8.50 Hz, Ar-H), 7.59 (d, 3H, J = 8.50 Hz, Ar-H), 7.66 (d, 3H, J = 9.00 Hz, Ar-H), 8.21 (d, 3H, J = 8.50 Hz, Ar-H), 8.38 (s, 1H, CH=NH), 10.02 (s, 1H, OH, D₂O exchangeable), 11.72 (s, 1H, NH hydrazone, D₂O exchangeable), 13.37 (s, 1H, benzimidazole NH, D₂O exchangeable); ^{13}C NMR (DMSO- d_6 , 125 MHz, δ ppm): 112.11, 115.56, 116.19, 116.36, 125.87, 127.09, 128.62, 128.89, 129.37, 129.69, 130.02, 130.14, 135.56, 148.29, 159.79 (aromatic carbons), 159.99 (C=N), 163.92 (C=O); elemental analysis calculated for C₂₁H₁₅ClN₄O₂ (M.Wt. 390.83): C, 64.54; H, 3.87; N, 14.34; found C, 64.24; H, 3.67; N, 14.08.

N'-(3,4,5-Trihydroxybenzylidene)-2-(4-nitrophenyl)-1H-benzimidazole-5-carbohydrazide (**4j**)

Yellow powder, 84% yield, m.p. 295–297 °C, IR (KBr, $\nu_{\max}$, cm^{-1}): 3462–3423 (NH benzimidazole, NH hydrazone), 3218 (OH), 3054 (CH aromatic), 1650 (C=O), 1615 (C=C), 1480 (C=N); ^1H NMR (DMSO- d_6 , 500 MHz, δ ppm): 6.42 (d, 1H, J = 8.50 Hz, Ar-H), 6.80 (d, 1H, J = 8.50 Hz, Ar-H), 7.66 (d, 3H, J = 8.50 Hz, Ar-H), 7.84 (s, 1H, Ar-H), 8.21 (d,

3H, $J = 8.50$ Hz, Ar-H), 8.49 (s, 1H, CH=NH), 8.58 (s, 1H, OH, D₂O exchangeable), 9.61 (s, 1H, OH, D₂O exchangeable), 11.65 (s, 1H, OH, D₂O exchangeable), 12.06 (s, 1H, NH hydrazone, D₂O exchangeable), 13.39 (s, 1H, benzimidazole NH, D₂O exchangeable); ¹³C NMR (DMSO-*d*₆, 125 MHz, δ ppm): 108.09, 111.36, 111.88, 121.74, 127.17, 128.40, 128.87, 128.91, 129.17, 129.60, 129.69, 133.15, 133.78, 135.62, 147.93, 149.10, 150.38 (aromatic carbons), 163.51 (C=N), 172.71 (C=O); elemental analysis calculated for C₂₁H₁₅ClN₄O₄ (M.Wt. 422.83): C, 59.65; H, 3.58; N, 13.25; found C, 60.01; H, 3.43; N, 13.51.

5.1.2. General procedure for the synthesis of 2-aryl-1H-benzimidazole-5-carbonyl hydrazine-1-carbothioamide (5a)

Equimolar quantities (2 mmol, 0.50 g) of compound **3** and appropriate isothiocyanate in absolute ethanol (25 mL) were heated under reflux for 6 h. The resulting crude solid was filtered off, dried, and used for the following step without further purification.

5.1.3. General procedure for the synthesis of 4-ethyl-5-(2-(4-chlorophenyl)-1H-benzimidazol-5-yl)-2,4-dihydro-3H-1,2,4-triazole-3-thione (6a)

Appropriate thiosemicarbazide **5a** (10 mmol) was dissolved in 2N sodium hydroxide solution (20 mL), and the resulting solution was stirred under reflux for 4 h. The solution was acidified with dilute HCl until (pH 3); the formed precipitate was filtered off, washed with distilled water, dried, and then recrystallized from ethanol to afford corresponding 1,2,4-triazole **6a** in a good yield.

Yellow powder, 63% yield, m.p. 292–294 °C, IR (KBr, $\nu_{\max}$, cm^{−1}): 3456, 3329 (NH benzimidazole, NH hydrazone), 3133 (CH aromatic), 2863 (CH aliphatic), 1502 (C=S), 1144 (C-N); ¹H NMR (DMSO-*d*₆, 500 MHz, δ ppm): 1.16 (t, 3H, $J = 7.00$ Hz, CH₃), 4.08 (q, 2H, $J = 7.50$ Hz, CH₂), 6.48 (d, 1H, $J = 8.50$ Hz, Ar-H), 7.64 (d, 2H, $J = 8.50$ Hz, Ar-H), 7.78 (d, 1H, $J = 8.50$ Hz, Ar-H), 7.91 (s, 1H, Ar-H), 8.27 (d, 2H, $J = 8.50$ Hz, Ar-H), 13.96 (s, 1H, NH triazole, D₂O exchangeable); ¹³C NMR (DMSO-*d*₆, 125 MHz, δ ppm): 13.91 (CH₃), 18.93 (CH₂), 56.50, 120.31, 123.38, 127.26, 128.83, 129.01, 129.48, 129.57, 129.66, 135.57, 152.34, 152.60 (aromatic carbon), 166.96 (C=S); elemental analysis calculated for C₁₇H₁₄ClN₅S (M.Wt. 355.84): C, 57.38; H, 3.97; N, 19.68; S, 9.01; found C, 57.42; H, 4.23; N, 19.74; S, 9.25.

5.2. Biology

5.2.1. In Vitro Antiproliferative Activity of Compounds Selected by the NCI

The NCI anti-cancer screening methodology has been described in detail elsewhere (<http://www.dtp.nci.nih.gov>, accessed on 20 February 2021). Following the Drug Evaluation Branch protocol, National Cancer Institute, Bethesda, USA, a primary anti-cancer assay was performed on approximately 60 human tumor cell line panels derived from nine neoplastic diseases.

5.2.2. Assay for EGFR Inhibitory Effect

Compounds **4c**, **4d**, **4e**, **4g**, and **4h** were investigated for EGFR inhibitory action using erlotinib as the control drug [45]. The detailed methods are described in the Supplementary Materials to this article.

5.2.3. Assay for BRAF^{V600E} Inhibitory Action

Compounds **4c**, **4d**, **4e**, **4g**, and **4h** were tested as possible BRAF^{V600E} inhibitors, compared to erlotinib and vemurafenib as control drugs [47]. Refer to the Supplementary Materials for more details.

5.2.4. Apoptotic Markers Assays

Compounds **4c** and **4e**, the most effective derivatives in all in vitro investigations, were investigated for their capacity to initiate apoptosis through their effect on caspase-3, caspase-8, Bax, and anti-apoptotic Bcl2 [52]. For more details, see Supplementary Materials.

5.3. Docking Study

Molecular modelling investigations were conducted using the Molecular Operating Environment (MOE) software version 2022. Compounds 4c and 4e structures were put together in MOE. Furthermore, the protein data bank at the Research Collaboration for Structural Bioinformatics (RSCB) protein database [PDB] (PDB: ID1M17 and 3OG7) provided the X-ray crystal structure of erlotinib and vemurafenib coupled to the EGFR and BRAF^{V600} active site, respectively. Erlotinib and vemurafenib were redocked into the 1M17 and 3OG7 active site to validate the docking strategy. After the co-crystallized ligand and water molecules were eliminated, the enzyme underwent a 3D protonation process in which hydrogen atoms were added to restore its normal geometry, making it ready for docking. Using the GW572016 scoring function, the conformers that were produced were docked into the EGFR and BRAF^{V600} receptor using MOE-dock. The best 50 poses underwent a force field refinement using molecular mechanics [55,56].

Supplementary Materials: The following supporting information can be downloaded at <https://www.mdpi.com/article/10.3390/molecules29020446/s1>, Figures S1–S47.

Author Contributions: Conceptualization, M.A.-A., H.M.A.-R., D.A.A.E.-E. and E.S.M.N.A.; methodology, B.G.M.Y. and M.M.M.; software, M.M.M.; validation, M.A.-A., H.M.A.-R., D.A.A.E.-E. and E.S.M.N.A.; formal analysis, B.G.M.Y.; data curation, M.A.-A.; writing—original draft preparation, B.G.M.Y. and M.A.-A.; writing—review and editing, B.G.M.Y., S.B. and M.A.-A.; supervision, H.M.A.-R., D.A.A.E.-E. and E.S.M.N.A. All authors have read and agreed to the published version of the manuscript.

Funding: The authors acknowledge support from the KIT-Publication Fund of the Karlsruhe Institute of Technology.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data will be provided upon request.

Conflicts of Interest: The authors reported no potential conflicts of interest.

References

- Ma, X.; Yu, H. Cancer issue: Global burden of cancer. *Yale J. Biol. Med.* **2006**, *79*, 85.
- Arnold, M.; Abnet, C.C.; Neale, R.E.; Vignat, J.; Giovannucci, E.L.; McGlynn, K.A.; Bray, F. Global Burden of 5 Major Types of Gastrointestinal Cancer. *Gastroenterology* **2020**, *159*, 335–349.e15. [\[CrossRef\]](#)
- Maeda, H.; Khatami, M. Analyses of repeated failures in cancer therapy for solid tumors: Poor tumor-selective drug delivery, low therapeutic efficacy and unsustainable costs. *Clin. Transl. Med.* **2018**, *7*, 11. [\[CrossRef\]](#)
- Kuczyński, E.A.; Sargent, D.J.; Grothey, A.; Kerbel, R.S. Drug rechallenge and treatment beyond progression—Implications for drug resistance. *Nat. Rev. Clin. Oncol.* **2013**, *10*, 571–587. [\[CrossRef\]](#)
- Hoelder, S.; Clarke, P.A.; Workman, P. Discovery of small molecule cancer drugs: Successes, challenges and opportunities. *Mol. Oncol.* **2012**, *6*, 155–176. [\[CrossRef\]](#)
- Kong, X.; Xu, J.; Yang, X.; Zhai, Y.; Ji, J.; Zhai, G. Progress in tumour-targeted drug delivery based on cell-penetrating peptides. *J. Drug Target.* **2021**, *30*, 46–60. [\[CrossRef\]](#)
- Hodson, R. Precision medicine. *Nature* **2016**, *537*, S49. [\[CrossRef\]](#)
- Tan, Y.J.; Lee, Y.T.; Petersen, S.H.; Kaur, G.; Kono, K.; Tan, S.C.; Majid, A.M.S.A.; Oon, C.E. BZD9L1 sirtuin inhibitor as a potential adjuvant for sensitization of colorectal cancer cells to 5-fluorouracil. *Ther. Adv. Med. Oncol.* **2019**, *11*, 1758835919878977. [\[CrossRef\]](#)
- Hagar, F.F.; Abbas, S.H.; Abdelhamid, D.; Gomaa, H.A.M.; Youssif, B.G.M.; Abdel-Aziz, M. New 1,3,4-oxadiazole-chalcone/benzimidazole hybrids as potent antiproliferative agents. *Arch. Pharm.* **2022**, *356*, e2200357. [\[CrossRef\]](#)
- Hagar, F.F.; Abbas, S.H.; Gomaa, H.A.M.; Youssif, B.G.M.; Sayed, A.M.; Abdelhamid, D.; Abdel-Aziz, M. Chalcone/1,3,4-Oxadiazole/Benzimidazole hybrids as novel anti-proliferative agents inducing apoptosis and inhibiting EGFR & BRAFV600E. *BMC Chem.* **2023**, *17*, 116. [\[CrossRef\]](#)
- Vitaku, E.; Smith, D.T.; Njardarson, J.T. Analysis of the structural diversity, substitution patterns, and frequency of nitrogen heterocycles among US FDA approved pharmaceuticals: Miniperspective. *J. Med. Chem.* **2014**, *57*, 10257–10274. [\[CrossRef\]](#)
- Gaba, M.; Mohan, C. Development of drugs based on imidazole and benzimidazole bioactive heterocycles: Recent advances and future directions. *Med. Chem. Res.* **2015**, *25*, 173–210. [\[CrossRef\]](#)

13. Kerru, N.; Gummidi, L.; Maddila, S.; Gangu, K.K.; Jonnalagadda, S.B. A Review on Recent Advances in Nitrogen-Containing Molecules and Their Biological Applications. *Molecules* **2020**, *25*, 1909. [\[CrossRef\]](#)
14. Subramanian, P.; Rudolf, G.C.; Kaliappan, K.P. Recent Trends in Copper-Catalyzed C–H Amination Routes to Biologically Important Nitrogen Scaffolds. *Chem. Asian J.* **2015**, *11*, 168–192. [\[CrossRef\]](#)
15. Tan, Y.J.; Lee, Y.T.; Yeong, K.Y.; Petersen, S.H.; Kono, K.; Tan, S.C.; Oon, C.E. Anti-cancer activities of a benzimidazole compound through sirtuin inhibition in colorectal cancer. *Future Med. Chem.* **2018**, *10*, 2039–2057. [\[CrossRef\]](#)
16. Shrivastava, N.; Naim, M.J.; Alam, M.J.; Nawaz, F.; Ahmed, S.; Alam, O. Benzimidazole scaffold as anti-cancer agent: Synthetic approaches and structure–activity relationship. *Arch. Pharm.* **2017**, *350*, e201700040. [\[CrossRef\]](#)
17. Shimomura, I.; Yokoi, A.; Kohama, I.; Kumazaki, M.; Tada, Y.; Tatsumi, K.; Ochiya, T.; Yamamoto, Y. Drug library screen reveals benzimidazole derivatives as selective cytotoxic agents for KRAS-mutant lung cancer. *Cancer Lett.* **2019**, *451*, 11–22. [\[CrossRef\]](#)
18. Zhong, L.; Li, Y.; Xiong, L.; Wang, W.; Wu, M.; Yuan, T.; Yang, W.; Tian, C.; Miao, Z.; Wang, T. Small molecules in targeted cancer therapy: Advances, challenges, and future perspectives. *Signal Transduct. Target. Ther.* **2021**, *6*, 201. [\[CrossRef\]](#)
19. Ardito, F.; Giuliani, M.; Perrone, D.; Troiano, G.; Lo Muzio, L. The crucial role of protein phosphorylation in cell signaling and its use as targeted therapy (Review). *Int. J. Mol. Med.* **2017**, *40*, 271–280. [\[CrossRef\]](#)
20. Lee, Y.T.; Tan, Y.J.; Oon, C.E. Benzimidazole and its derivatives as cancer therapeutics: The potential role from traditional to precision medicine. *Acta Pharm. Sin. B* **2023**, *13*, 478–497. [\[CrossRef\]](#)
21. Tsatsanis, C.; Spandidos, D.A. The role of oncogenic kinases in human cancer. *Int. J. Mol. Med.* **2000**, *5*, 583–673. [\[CrossRef\]](#)
22. Tan, D.S.-W.; Leighl, N.B.; Riely, G.J.; Yang, J.C.-H.; Sequist, L.V.; Wolf, J.; Seto, T.; Felip, E.; Aix, S.P.; Jonnaert, M.; et al. Safety and efficacy of nazartinib (EGF816) in adults with EGFR-mutant non-small-cell lung carcinoma: A multicentre, open-label, phase 1 study. *Lancet Respir. Med.* **2020**, *8*, 561–572. [\[CrossRef\]](#)
23. Masuzawa, K.; Yasuda, H.; Hamamoto, J.; Nukaga, S.; Hirano, T.; Kawada, I.; Naoki, K.; Soejima, K.; Betsuyaku, T. Characterization of the efficacies of osimertinib and nazartinib against cells expressing clinically relevant epidermal growth factor receptor mutations. *Oncotarget* **2017**, *8*, 105479–105491. [\[CrossRef\]](#)

24. Lelais, G.; Epple, R.; Marsilje, T.H.; Long, Y.O.; McNeill, M.; Chen, B.; Lu, W.; Anumolu, J.; Badiger, S.; Bursulaya, B. Discovery of (R, E)-N-(7-Chloro-1-(1-[4-(dimethylamino) but-2-enoyl] azepan-3-yl)-1 H-benzo [d] imidazol-2-yl)-2-methylisonicotinamide (EGF816), a Novel, Potent, and WT Sparing Covalent Inhibitor of Oncogenic (L858R, ex19del) and Resistant (T790M) EGFR Mutants for the Treatment of EGFR Mutant Non-Small-Cell Lung Cancers. *J. Med. Chem.* **2016**, *59*, 6671–6689. [[PubMed](#)]
25. Kim, D.-W.; Tan, D.S.-W.; Aix, S.P.; Sequist, L.V.; Smit, E.F.; Hida, T.; Yang, J.C.-H.; Felip, E.; Seto, T.; Grohé, C. Preliminary Phase II results of a multicenter, open-label study of nazartinib (EGF816) in adult patients with treatment-naïve EGFR-mutant non-small cell lung cancer (NSCLC). *Am. Soc. Clin. Oncol.* **2018**, *36*, 9094. [[CrossRef](#)]
26. Jia, Y.; Juarez, J.; Li, J.; Manuia, M.; Niederst, M.J.; Tompkins, C.; Timple, N.; Vaillancourt, M.-T.; Pferdekamper, A.C.; Lockerman, E.L. EGF816 exerts anti-cancer effects in non-small cell lung cancer by irreversibly and selectively targeting primary and acquired activating mutations in the EGF receptor. *Cancer Res.* **2016**, *76*, 1591–1602. [[CrossRef](#)]
27. Ramurthy, S.; Subramanian, S.; Aikawa, M.; Amiri, P.; Costales, A.; Dove, J.; Fong, S.; Jansen, J.M.; Levine, B.; Ma, S.; et al. Design and Synthesis of Orally Bioavailable Benzimidazoles as Raf Kinase Inhibitors. *J. Med. Chem.* **2008**, *51*, 7049–7052. [[CrossRef](#)] [[PubMed](#)]
28. Alshammari, M.B.; Aly, A.A.; Youssif, B.G.M.; Bräse, S.; Ahmad, A.; Brown, A.B.; Ibrahim, M.A.A.; Mohamed, A.H. Design and synthesis of new thiazolidinone/uracil derivatives as antiproliferative agents targeting EGFR and/or BRAFV600E. *Front. Chem.* **2022**, *10*, 1076383. [[CrossRef](#)]
29. Al-Wahaibi, L.H.; Abou-Zied, H.A.; Beshr, E.A.; Youssif, B.G.; Hayallah, A.M.; Abdel-Aziz, M. Design, Synthesis, Antiproliferative Actions, and DFT Studies of New Bis-Pyrazoline Derivatives as Dual EGFR/BRAFV600E Inhibitors. *Int. J. Mol. Sci.* **2023**, *24*, 9104. [[CrossRef](#)] [[PubMed](#)]
30. Al-Wahaibi, L.H.; El-Sheref, E.M.; Hammouda, M.M.; Youssif, B.G. One-Pot Synthesis of 1-Thia-4-azaspiro [4.4/5] alkan-3-ones via Schiff Base: Design, Synthesis, and Apoptotic Antiproliferative Properties of Dual EGFR/BRAFV600E Inhibitors. *Pharmaceuticals* **2023**, *16*, 467. [[CrossRef](#)]
31. Al-Wahaibi, L.H.; Gouda, A.M.; Abou-Ghadi, O.F.; Salem, O.I.; Ali, A.T.; Farghaly, H.S.; Abdelrahman, M.H.; Trembleau, L.; Abdu-Allah, H.H.; Youssif, B.G. Design and synthesis of novel 2,3-dihydropyrazino[1,2-a]indole-1,4-dione derivatives as antiproliferative EGFR and BRAFV600E dual inhibitors. *Bioorg. Chem.* **2020**, *104*, 104260. [[CrossRef](#)] [[PubMed](#)]
32. Al-Wahaibi, L.H.; Mahmoud, M.A.; Mostafa, Y.A.; Raslan, A.E.; Youssif, B.G. Novel piperine-carboximidamide hybrids: Design, synthesis, and antiproliferative activity via a multi-targeted inhibitory pathway. *J. Enzym. Inhib. Med. Chem.* **2023**, *38*, 376–386. [[CrossRef](#)] [[PubMed](#)]
33. Aly, A.A.; Alshammari, M.B.; Ahmad, A.; Gomaa, H.A.M.; Youssif, B.G.M.; Bräse, S.; Ibrahim, M.A.A.; Mohamed, A.H. Design, synthesis, docking and mechanistic studies of new thiazolyl/thiazolidinylpyrimidine-2,4-dione antiproliferative agents. *Arab. J. Chem.* **2023**, *16*, 104612. [[CrossRef](#)]
34. El-Lateef, H.M.A.; Elbastawes, M.A.; Ibrahim, T.M.A.; Khalaf, M.M.; Gouda, M.; Wahba, M.G.; Zaki, I.; Morcoss, M.M. Design, Synthesis, Docking Study, and Antiproliferative Evaluation of Novel Schiff Base-Benzimidazole Hybrids with VEGFR-2 Inhibitory Activity. *Molecules* **2023**, *28*, 481. [[CrossRef](#)] [[PubMed](#)]
35. Galal, S.A.; Hegab, K.H.; Kassab, A.S.; Rodriguez, M.L.; Kerwin, S.M.; El-Khamry, A.-M.A.; El Diwani, H.I. New transition metal ion complexes with benzimidazole-5-carboxylic acid hydrazides with antitumor activity. *Eur. J. Med. Chem.* **2009**, *44*, 1500–1508. [[CrossRef](#)] [[PubMed](#)]
36. Morcoss, M.M.; Abdelhafez, E.S.; Abdel-Rahman, H.M.; Abdel-Aziz, M.; El-Ella, A.; Dalal, A. Novel Benzimidazole/Hydrazone Derivatives as Promising Anti-cancer Lead Compounds: Design, Synthesis, and Molecular Docking Study. *J. Adv. Biomed. Pharm. Sci.* **2020**, *3*, 45–52.
37. Abuilaiwi, F.A.; Laoui, T.; Al-Harathi, M.; Atieh, M. Modification and functionalization of multiwalled carbon nanotube (MWCNT) via fischer esterification. *Arab. J. Sci. Eng.* **2010**, *35*, 37–48.
38. Morcoss, M.M.; Abdelhafez, E.S.M.; Ibrahim, R.A.; Abdel-Rahman, H.M.; Abdel-Aziz, M.; El-Ella, D.A.A. Design, synthesis, mechanistic studies and in silico ADME predictions of benzimidazole derivatives as novel antifungal agents. *Bioorg. Chem.* **2020**, *101*, 103956. [[CrossRef](#)]
39. Al-Warhi, T.; Alqahtani, L.S.; Alsharif, G.; Abualnaja, M.; Ali, O.A.A.; Qahl, S.H.; Althagafi, H.A.E.; Alharathi, F.; Jafri, I.; Elsaid, F.G. Design, Synthesis, and Investigation of Cytotoxic Activity of cis-Vinylamide-Linked Combretastatin Analogues as Potential Anti-cancer Agents. *Symmetry* **2022**, *14*, 2088. [[CrossRef](#)]
40. El-Sherief, H.A.; Youssif, B.G.; Bukhari, S.N.A.; Abdelazeem, A.H.; Abdel-Aziz, M.; Abdel-Rahman, H.M. Synthesis, anti-cancer activity and molecular modeling studies of 1, 2, 4-triazole derivatives as EGFR inhibitors. *Eur. J. Med. Chem.* **2018**, *156*, 774–789. [[CrossRef](#)]
41. Wong, C.C.; Sagineedu, S.R.; Sumon, S.H.; Sidik, S.M.; Phillips, R.; Lajis, N.H.; Stanslas, J. NCI in vitro and in silico anticancer screen, cell cycle perturbation and apoptosis-inducing potential of new acylated, benzyldiene and isopropylidene derivatives of andrographolide. *Environ. Toxicol. Pharmacol.* **2014**, *38*, 489–501. [[CrossRef](#)]
42. Ali, A.R.; El-Bendary, E.R.; Ghaly, M.A.; Shehata, I.A. Synthesis, in vitro anti-cancer evaluation and in silico studies of novel imidazo [2,1-b] thiazole derivatives bearing pyrazole moieties. *Eur. J. Med. Chem.* **2014**, *75*, 492–500. [[CrossRef](#)] [[PubMed](#)]
43. Ali, A.R.; El-Bendary, E.R.; Ghaly, M.A.; Shehata, I.A. Novel acetamidothiazole derivatives: Synthesis and in vitro anti-cancer evaluation. *Eur. J. Med. Chem.* **2013**, *69*, 908–919. [[CrossRef](#)] [[PubMed](#)]

44. Kryshchysyn, A.; Atamanyuk, D.; Lesyk, R. Fused Thiopyrano[2,3-d]thiazole Derivatives as Potential Anti-cancer Agents. *Sci. Pharm.* **2012**, *80*, 509–530. [\[CrossRef\]](#)
45. Mahmoud, M.A.; Mohammed, A.F.; Salem, O.I.; Gomaa, H.A.; Youssif, B.G. New 1, 3, 4-oxadiazoles linked with the 1, 2, 3-triazole moiety as antiproliferative agents targeting the EGFR tyrosine kinase. *Arch. Pharm.* **2022**, *355*, 2200009. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Zubair, T.; Bandyopadhyay, D. Small Molecule EGFR Inhibitors as Anti-Cancer Agents: Discovery, Mechanisms of Action, and Opportunities. *Int. J. Mol. Sci.* **2023**, *24*, 2651. [\[CrossRef\]](#)
47. Youssif, B.G.; Gouda, A.M.; Moustafa, A.H.; Abdelhamid, A.A.; Gomaa, H.A.; Kamal, I.; Marzouk, A.A. Design and synthesis of new triarylimidazole derivatives as dual inhibitors of BRAFV600E/p38 α with potential antiproliferative activity. *J. Mol. Struct.* **2021**, *1253*, 132218. [\[CrossRef\]](#)
48. Liu, Y.; Zhu, X. Endoplasmic reticulum-mitochondria tethering in neurodegenerative diseases. *Transl. Neurodegener.* **2017**, *6*, 21. [\[CrossRef\]](#)
49. Villa-Pulgarin, J.A.; Gajate, C.; Botet, J.; Jimenez, A.; Justies, N.; Varela-M, R.E.; Cuesta-Marban, A.; Müller, I.; Modolell, M.; Revuelta, J.L. Mitochondria and lipid raft-located FOF1-ATP synthase as major therapeutic targets in the antileishmanial and anti-cancer activities of ether lipid edelfosine. *PLoS Neglected Trop. Dis.* **2017**, *11*, e0005805. [\[CrossRef\]](#)
50. Bao, H.; Zhang, Q.; Zhu, Z.; Xu, H.; Ding, F.; Wang, M.; Du, S.; Du, Y.; Yan, Z. BHX, a novel pyrazoline derivative, inhibits breast cancer cell invasion by reversing the epithelial-mesenchymal transition and down-regulating Wnt/ β -catenin signalling. *Sci. Rep.* **2017**, *7*, 9153. [\[CrossRef\]](#)
51. Martin, S. Caspases: Executioners of apoptosis. In *Pathobiology of Human Disease*; Elsevier: Amsterdam, The Netherlands, 2014.
52. Youssif, B.G.; Mohamed, A.M.; Osman, E.E.A.; Abou-Ghadi, O.F.; Elnaggar, D.H.; Abdelrahman, M.H.; Treamblu, L.; Gomaa, H.A. 5-Chlorobenzofuran-2-carboxamides: From allosteric CB1 modulators to potential apoptotic antitumor agents. *Eur. J. Med. Chem.* **2019**, *177*, 1–11. [\[CrossRef\]](#) [\[PubMed\]](#)
53. AboulWafa, O.M.; Daabees, H.M.; Badawi, W.A. 2-Anilinopyrimidine derivatives: Design, synthesis, in vitro anti-proliferative activity, EGFR and ARO inhibitory activity, cell cycle analysis and molecular docking study. *Bioorg. Chem.* **2020**, *99*, 103798. [\[CrossRef\]](#) [\[PubMed\]](#)
54. Gomaa, H.A.; Shaker, M.E.; Alzarea, S.I.; Hendawy, O.; Mohamed, F.A.; Gouda, A.M.; Ali, A.T.; Morcoss, M.M.; Abdelrahman, M.H.; Trembleau, L.; et al. Optimization and SAR investigation of novel 2,3-dihydropyrazino[1,2-a]indole-1,4-dione derivatives as EGFR and BRAFV600E dual inhibitors with potent antiproliferative and antioxidant activities. *Bioorg. Chem.* **2022**, *120*, 105616. [\[CrossRef\]](#) [\[PubMed\]](#)
55. Ibrahim, T.S.; Bokhtia, R.M.; Al-Mahmoudy, A.M.; Taher, E.S.; AlAwadh, M.A.; Elagawany, M.; Abdel-Aal, E.H.; Panda, S.; Gouda, A.M.; Asfour, H.Z.; et al. Design, synthesis and biological evaluation of novel 5-((substituted quinolin-3-yl/1-naphthyl)methylene)-3-substituted imidazolidin-2,4-dione as HIV-1 fusion inhibitors. *Bioorg. Chem.* **2020**, *99*, 103782. [\[CrossRef\]](#)
56. Shaykoon, M.S.; Marzouk, A.A.; Soltan, O.M.; Wanas, A.S.; Radwan, M.M.; Gouda, A.M.; Youssif, B.G.; Abdel-Aziz, M. Design, synthesis and antitrypanosomal activity of heteroaryl-based 1,2,4-triazole and 1,3,4-oxadiazole derivatives. *Bioorg. Chem.* **2020**, *100*, 103933. [\[CrossRef\]](#)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.