



# **Quercetin: A Potential Polydynamic Drug**

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**Abstract:** The study of natural products as potential drug leads has gained tremendous research interest. Quercetin is one of those natural products. It belongs to the family of flavonoids and, more specifically, flavonols. This review summarizes the beneficial pharmaceutical effects of quercetin, such as its anti-cancer, anti-inflammatory, and antimicrobial properties, which are some of the quercetin effects described in this review. Nevertheless, quercetin shows poor bioavailability and low solubility. For this reason, its encapsulation in macromolecules increases its bioavailability and therefore pharmaceutical efficiency. In this review, a brief description of the different forms of encapsulation of quercetin are described, and new ones are proposed. The beneficial effects of applying new pharmaceutical forms of nanotechnology are outlined.

Keywords: quercetin; flavonoids

## 1. Introduction

Flavonoids [1] (Scheme 1a) are a family of organic compounds found mostly in plants and in the food that humans consume. They exert many important biological actions, with favorable antioxidant effects. Flavonoids can be classified into different classes, depending on the substitutes of carbons of the rings [2]. One of these classes is flavonols. Flavonols are flavonoids with a keto group. Flavonols occur in vegetables and fruits such as onions, tomatoes, and apples. One of the most studied flavonols is quercetin (Scheme 1b).

Quercetin [3,4] is an organic compound that belongs to the family of flavonoids, with a wide range of medical properties [5,6]. Some of these include anti-allergy, antiinflammatory, anticancer, anti-tumor, and antiviral properties as well as cardiovascular protection. It has also been found that quercetin plays a vital role in plants [7]. Specifically, quercetin has antioxidant and antimicrobial activities, and as a result, it contributes to photosynthesis, growth, and seed germination. Moreover, the presence of quercetin in various regions of the brain contributes to combatting against various neurological diseases such as Alzheimer's and Parkinson's [8]. Until now, there is no specific treatment for these diseases, but flavonoids—and especially quercetin—have been used for treatment in animal models.



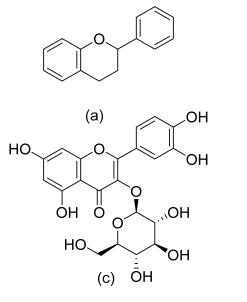
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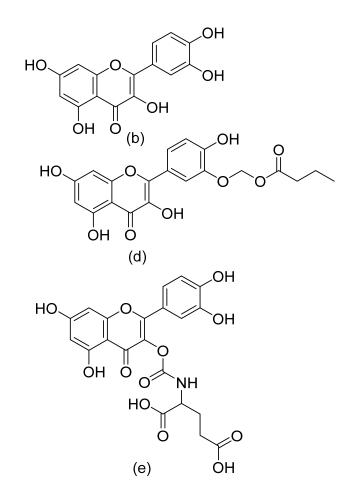
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**Scheme 1.** Basic skeleton using Chem Draw of (**a**) 2-phenyl chromane flavonoid (PubChemCID: 94156), (**b**) quercetin (PubChemCID: 5280343), (**c**) isoquercetin (PubChemCID: 5280804), (**d**) quercetin derivative that is effective against MDR cancer cells [1], and (**e**) quercetin-glutamic acid [2].

The skeletal formula of quercetin [9] was acquired using Chem Draw to find some physicochemical and toxicity properties through the SwissADME [10], pkCSM [11], and preADME [12] platforms. This procedure is very important for computational drug design. Some potential biological compounds fail to reach clinical trials due to their unfavorable (ADME) parameters [13–17].

Quercetin's molecular weight is <500 g/mol, its number of hydrogen-bonding donors is less than five, its number of hydrogen-bonding acceptors is less than 10, and its lipophilicity [18] is less than five. As a result, it obeys Lipinski's Rules of Five. Also, Veber's Rule [19] is qualified, because the number of rotatable bonds is less than seven. Quercetin has not been predicted to be hepatotoxic, and it has no skin sensitization [20]. Due to the fact that its blood brain barrier index (BBB) [21] is less than one, it is considered as inactive to the central nervous system (CNS). Also, it might be better absorbed from the intestinal tract on oral administration. It has very low solubility in water (about  $1 \mu g/mL$ ) and low bioavailability. Its low bioavailability has led researchers to synthesize various complexes with quercetin engulfed in transfer vehicles.

The SwissTarget platform was employed in order to assess quercetin's inhibitory activity. Inhibitory activity has been observed against a plethora of enzymes such as monoamine oxidase A, monoamine oxidase B [22], and thrombin or lipoxygenase [23]. Lipoxygenases belong to the category of oxidoreductases and are widely found in plant organisms, fungi, and animals. Such enzymes are not commonly found in yeasts and bacteria and are not elements of a typical prokaryotic cell.

Bioavailability is the ability of a compound to be active inside the organism and to enter systemic circulation. Quercetin is lipophilic, with poor water solubility. Therefore, its bioavailability is low, and for that reason, a common strategy to increase its bioavailability is for it to be engulfed in biomolecules and form soluble complexes.

#### 2. Polydynamic Biological Activity of Quercetin

#### 2.1. Mental Activity

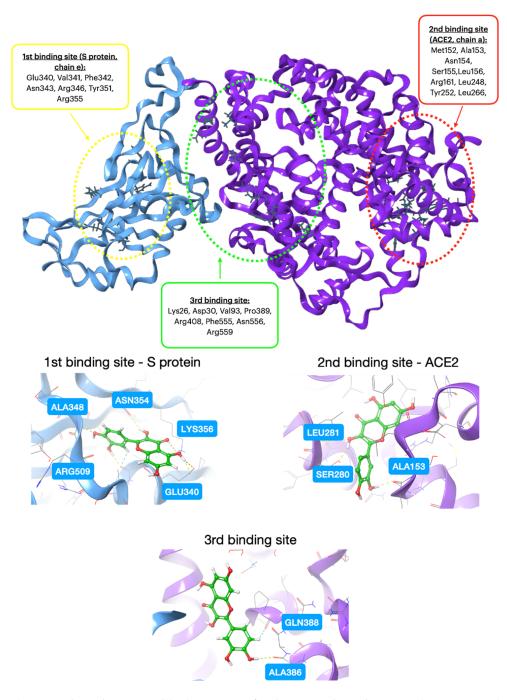
Quercetin can play a significant role in mental health diseases [24] such as depression and anxiety. Mice studies have demonstrated that some natural products including quercetin possess anxiolytic properties when administered orally. Moreover, they are unlikely to have side effects serious enough to prevent their pharmacological utility, so they could constitute the starting point for the development of more selective anxiolytic agents [25]. Due to its antioxidant activity, quercetin can lower nitric oxide and some other compounds that are vital for these diseases. According to SwissADME, quercetin can inhibit CYP isoenzymes. As a result, it can protect the organism from pathogenic factors.

#### 2.2. Ultraviolet (UV) Activity

One study has shown that quercetin encapsuled with polymer nanoparticles can be efficient for sun protection [26]. In recent years, ultraviolet (UV) radiation has been considered a public threat for health worldwide, as it is responsible for acute and chronic skin diseases, such as burns, premature aging skin, and carcinogenesis. Skin cancer is the most common type of cancer that is diagnosed worldwide. It can cause a high degree of mortality when it develops into its most severe form, that of melanoma. Thus, necessary protection is required during exposure to sunlight. Today, sunscreens are used to protect us from early photoaging, photosensitivity, skin cancer, and free radical damage. The main goal of sunscreens is to protect the human skin from UVA and UVB radiation. Recent studies have shown that compounds from natural plants may act as sun protectors [27]. Quercetin is one of those natural products that can reduce the damage from UV radiation.

#### 2.3. Antiviral Activity

Viral diseases are still a problem even after the discovery and use of antiviral drugs for more than 60 years now, due to the toxicity of some new antiviral preparations and the development of resistant viral strains. The human immunodeficiency virus (HIV) [28] is another disease that started to spread throughout the world. HIV has two categories, HIV type 1 and HIV type 2 (HIV-2). HIV was first recognized in 1981 in the USA. The origin of this virus is primate lentiviruses, which exist in chimpanzees. These animals became the host of the virus, which is then transmitted to humans after mutations [29]. Quercetin and isoquercetin (Scheme 1c) have antiviral activities against many types of viruses, including human immunodeficiency virus. Many scientists have suggested quercetin as an antiviral drug due to the fact that it can inhibit the first stages of the virus infection. Quercetin has also been found to exert important pharmacological activity against several other viruses [30]. One such activity is against the Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) [31,32], which recently emerged as a global threat to human health. It is the main cause of the COVID-19 pandemic that caused more than 6,000,000 deaths worldwide. Quercetin has been found to be able to interfere with SARS-CoV-2 and reduce the inflammation provoked by COVID-19 (Scheme 2). Also, blood tests have indicated that quercetin can reduce the time during which the molecular test appears positive by reducing the viral charge [33].



**Scheme 2.** Three-dimensional (3D) structure of spike protein bound to ACE2 (PDB ID: 6M0J). The dotted circle identifies the amino acids that constitute the respective binding sites for the Induced Fit Docking (IFD) experiments (**above**). The interactions were developed from the IFD experiments for quercetin with spike proteins of SARS-CoV-2 from the three studied binding sites (**bottom**). This image was sketched using Maestro software Version 10.2.

## 2.4. Anticancer Activity

Cancer is a serious disease that hurts many developed and developing countries. There are more than 100 types of cancer. Most often, a specific type of cancer is characterized by the type of cell in which it is formed. The most basic types of cancer are carcinoma, sarcoma, leukemia, lymphoma, multiple myeloma, melanoma, and brain and spinal cord tumors [34].

Quercetin has been found through in vitro experiments to exhibit anti-tumor activity against prostate [35,36], liver [37], breast [38], and pancreatic [39] cancer and melanoma [40]. Quercetin's anticancer effects on hepatocellular carcinoma [41] have been studied not only in vitro but also in vivo. Although the exact mechanism of action remains elusive, quercetin's anticancer effect may arise by regulating some enzymatic activities or also by modulating oxidative stress and some cellular pathways.

Chemoprevention involves treating cancer before it becomes aggressive, but it does have its downsides. This approach can potentially lead to side effects and toxicity. Quercetin has demonstrated synergistic effects in addressing tumors with multidrug resistance by blocking the expulsion of drugs facilitated by transporter proteins. It is used as a low-toxicity medicine. Studies have shown that the anticancer activity of quercetin can be improved by encapsulating quercetin inside nanoparticles. In vitro and in vivo studies have shown successful tumor treatment using quercetin nano-formulations. These approaches can reduce side effects. Some examples include polymeric nanoparticles, non-responsive polymeric nanoparticles, and stimuli-responsive polymeric nanoparticles. There are also examples of inorganic nanoparticles with quercetin—specifically, silica nanoparticles, gold nanoparticles, and metal oxide nanoparticles [42].

#### 2.5. Anti-Inflammatory Activity

Moreover, quercetin has been shown to exert anti-inflammatory activity. Inflammation is a multifactorial and complex biological response of body tissues to harmful stimuli, so as to restore the organism to homeostatic balance. Inflammation is found in some areas of the body and refers to the tissues of an organ or a tissue or a whole organ, etc. (e.g., arthritis, tendonitis, stomatitis, peritonitis, etc.). Rheumatoid arthritis is an example of an autoimmune inflammatory disease [43]. This disease affects more women than men, and it was discovered many years ago. The symptoms of this disease are stiffness and swelling that appear in the feet, fingers, and toes. In vitro [44] studies have shown that quercetin may be a good drug candidate for the treatment of this disease, because it can inhibit neutrophil activity. It can also inhibit the activation of NLRP3 inflammasomes [31]. Lipoxygenases (LOXs) are a group of monomeric oxidant metalloproteins, containing a non-heme-coordinated iron atom (non-heme ion Fe) [45,46]. Moreover, several in vitro experiments have shown that quercetin can inhibit soybean lipoxygenase [3], which is involved in inflammation.

## 2.6. Neurological Activity

Alzheimer's disease (AD) [47] is a fatal complex neurodegenerative disease that affects more than 24 million people worldwide [48]. The disease is characterized by multiple pathological features and is clinically associated with cognitive impairment, language loss capacity, and dementia. Current treatment options include results with moderate improvement of memory and cognitive function; however, they do not prevent progressive neurodegeneration. Multifunctional compounds capable of simultaneously interacting with the ingredients of many pathologies have been considered as a solution and are being researched for the treatment of complex pathologies of neurodegenerative diseases [49,50]. Quercetin is one of these compounds that can be used against Alzheimer's disease due to the fact that it has a neuroprotective effect against oxidative stress [51].

#### 2.7. Antioxidant Activity

When found in moderate concentrations, the active forms of oxygen (reactive oxygen species, ROS) participate in the normal processes of the organism, but their production in large concentrations leads to oxidative stress, disrupting the organism's cellular oxidation balance [52,52,53]. Antioxidants are substances that can protect cells against oxidation and the effects of free radicals, because they annihilate these radicals from the medium. They also constrain oxidation by oxidizing themselves. Antioxidants suppress various harmful activities of ROS, so they are used to prevent or treat such diseases. One health problem that oxidative stress is associated with is obesity. Obesity is one of the major health problems in the world, and it leads to increased amounts of fat cells. It is characterized by the overproduction of reactive oxygen stress. Quercetin, along with other natural products, has been shown to exert beneficial effects against obesity through different molecular pathways. In vivo experiments using obese rats have been shown to lose weight after treating with quercetin [54].

Moreover, Jose Angel Maranon Maroto proved that a combination of the polyphenols resveratrol, quercetin, and catechin has synergic antioxidant power. Polyphenolic compounds of natural origin are recognized as antioxidant agents, which act as free radical scavengers. Resveratrol is a polyphenolic compound that is present mostly in seeds and in the skin of grapes and other plant products [55].

## 2.8. Anti-Cardiovascular Disease

Heart diseases or cardiovascular diseases are those diseases that involve the heart or blood vessels (arteries and veins) [56]. Though the term technically it refers to any disease affecting the cardiovascular system, it is usually used to refer to those related to atherosclerosis (arterial diseases). These diseases present similar causes, mechanisms, and treatments. Most countries face high and increasing rates of cardiovascular diseases. It has been shown that they affect adolescents and kids, and for this reason, prevention against them is mandatory from childhood. When heart problems are diagnosed, the underlying cause (atherosclerosis) is usually quite advanced. Therefore, more emphasis is placed on the prevention of atherosclerosis through the modification of risk factors, such as healthy eating, exercise, and avoiding smoking. The protective effects of quercetin against cardiovascular diseases include the reduction of blood pressure and arterial pressure. Hypertension is the most common cause of cardiovascular diseases, such as in cardiac hypertrophy, responsible for abnormal cardiac growth, which leads to arrhythmia, myocardial infraction, and heart failure [57]. Many people currently suffer from hypertension, an alerting sign that pharmacological and natural interventions are needed in order to decrease blood pressure and inhibit various biochemical pathways that are involved in cardiovascular diseases. Studies have demonstrated that quercetin can decrease blood pressure via multiple mechanisms like inhibiting protein kinase C (PKC), a family of protein kinase enzymes implicated in governing heart failure [58], decreasing oxidative stress, inhibiting angiotensin, converting enzyme activity, or even modulating cell signaling and gene expression [59].

## 2.9. Skin Sensitivity

Numerous individuals experience skin wounds, which can be either chronic or temporary and may affect a substantial area of the skin. Healing processes typically fall into three categories: primary healing (also known as healing by first intention), which takes place within 12 to 24 h after the wound forms; secondary healing (or healing by second intention), observed in wounds with significant loss of soft tissue; and the healing of superficial wounds, such as those seen in superficial burns and abrasions, involving the epithelium and the papillary part of the dermis. Natural products, especially those from plants, are a new strategy for wound healing. Quercetin is used for the treatment for wounds because, as outlined, it shows anti-inflammatory activity. *Rubusniveus* [60] is one of the species against which quercetin was found to have some wound-healing activity [61]. There are a lot of examples of the biological activity of quercetin occurring in in vivo experiments. One example is in the species of *Bergia ammannioides* [62], against which quercetin was found to have antioxidant and anti-inflammatory abilities. Secondly, in *Melilotus officinalis* and in *Lespedeza capitata* [63], quercetin was found to increase the Ha-CaT human keratinocytes. Furthermore, in *Martynia annua* and *Tephrosia purpurea* [64], quercetin was found to have antioxidant activity. Also, quercetin has protection against endotoxin-induced inflammatory response [65], surgical-induced osteoarthritis [66], LPS-induced oxidative stress and inflammation [67], LPS/interferon c-induced nitric oxide production [68], TNF- $\alpha$  induced inflammation [69], and CCl4-induced inflammation [70].

## 2.10. Anti-Tuberculosis

Tuberculosis is a fatal infectious disease caused by the Mycobacterium tuberculosis (*Mycobacterium tuberculosis*). Despite the availability of effective treatment, tuberculosis is responsible for a million deaths worldwide per year. The bacterium has developed a resistance to the drugs on the market, and so the need arises to find other therapeutic compounds [71,72]. Quercetin can be a good inhibitor for the bacterium [73]. This was found through in vitro antituberculosis bioassays.

#### 2.11. Antidiabetic Activity

Insulin is a protein hormone that is necessary for the maintenance of normal blood glucose levels, either by increasing peripheral glucose uptake or by suppressing the production of hepatic glucose [74]. Quercetin might be a promising candidate that acts in many targets of diabetes, and it can regulate many pathways [75,76]. Furthermore, co-crystals comprised of quercetin and antidiabetic agents like metformin and DPP-IV inhibitors have been demonstrated to treat diabetes mellitus (DM) by reducing blood glucose levels and improving glucose tolerance. DM is a chronic disease that is diagnosed as a result of elevated blood glucose levels caused by inadequate insulin secretion, defective insulin action, or both [77,78].

## 2.12. Antimalaria Activity

Malaria is one of the most threatening tropical diseases that leads to millions of deaths every year. Almost all fatal cases are caused by *Plasmodium falciparum* and its strains, which have developed resistance to the drugs in circulation. Therefore, a need has arisen for new active compounds for the treatment of this disease. Quercetin is a potential antimalaria drug, as proven through in vitro experiments [79].

#### 2.13. Antichagas Activity

Chagas disease (CD) [80] is a disease that many scientists ignore, and its main bacteria is the *Trypanosoma cruzi* (TC). This disease appears mainly in Central and North America, but in recent decades, the number of CD cases has been increasing in other countries, such as in the south of the United States of America, in Canada, in the Western Mediterranean, and in the Western Pacific. It is estimated that about 6 to 7 million people are potentially infected by TC, which causes about 20,000 deaths per year and is the leading cause of infectious myocarditis. Quercetin and other flavonol derivatives can be antitrypanosomal candidates, showing IC<sub>50</sub>s of 0.6, 0.7, 0.8, and 1.0  $\mu$ g/mL [81].

#### 2.14. Antifungal Activity

Fungicides have often been observed to pose a risk to human health and can be harmful to the environment. Thus, there is a need to find alternative solutions to deal with fungi, with natural compounds that will not affect either human health or the environment. The *Candida parapsilosis* species is composed of three other species, i.e., *C. parapsilosis sensu lato*, *C. orthopsilosis*, and *C. metapsilosis*. These species are found in vegetables and fruits, and they are known to cause infections worldwide. Quercetin has been shown to have

antifungal activities through the determination of its minimum inhibitory concentration (MIC) [82].

#### 2.15. Combination of Quercetin with Other Drugs

There are several examples whereby quercetin and its derivatives have been combined with other compounds with biological interest and activity. One of these examples is sickle cell disease. Sickle cell disease and its variants constitute the most common blood disorders, affecting millions of individuals worldwide. Until now, there has been no treatment for this disease, and there is no acute method for prevention [83]. Another example is Fragile X Syndrome. This disease is the most common one implicated in intellectual disability. Someone who suffers from this disease has many serious medical problems [84]. Examples of the use of quercetin with other drugs will be described in the Section 3.

#### 2.16. Anti-Rhinitis Activity

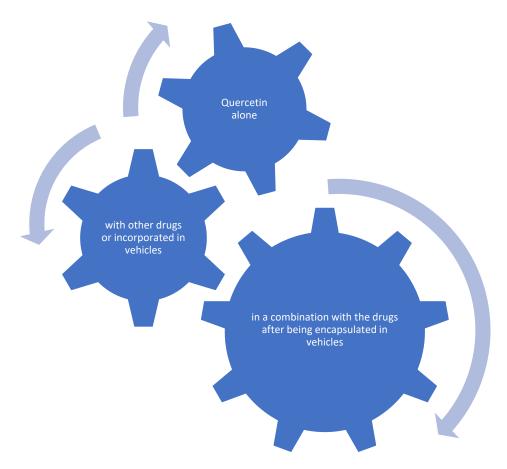
Acute rhinitis is one of the most common inflammatory diseases in Western countries. The major symptoms include nasal obstruction and nasal secretions. In the evolution of the disease, a frequent complication is acute rhinosinusitis, which can progress to chronic rhinosinusitis and then to intracranial complications, meaning that it is necessary to treat the disease as soon as possible. The cause of rhinosinusitis lies in the secretion of proinflammatory cytokines, key factors in initiating inflammation, consequently leading to local edema and swelling of the mucosa and an increase in nasal and sinus secretions. Quercetin has proven its antioxidant and anti-inflammatory properties against rhinosinusitis, both in rats and humans, by inhibiting the release of chemical mediators, such as histamines and leukotrienes, and reacting with relative oxygen species (ROS), which are also involved in rhinosinusitis [85,86].

## 2.17. Antidrug Resistance

Multidrug resistance (MDR) is defined as the ability of cancer cells to survive treatment with a variety of anticancer drugs, similar to the concept commonly applied to antibiotic treatment [87]. MDR is responsible for over 90% of deaths in cancer patients receiving chemotherapeutics or targeted drugs. Derivatives of quercetin (Scheme 1d) have been proven to be possible candidates in treating MDR cancer, as well as viral infections in humans [88].

Skeletal muscles are tissues that are involved not only in mobility and movement but also in glucose and lipid metabolism. Muscle atrophy is the loss of skeletal muscle mass due to increased myofibrillar protein degradation. It occurs under various circumstances such as injury and during side effects of pharmaceutical therapy and aging. Muscle atrophy causes falls, and therefore, it has become a serious problem, especially in aging society. Quercetin glucosides are proven to be perfect candidates in the treatment of muscle atrophy, since they play an important role in the downregulation of myostatin signaling via phosphorylation, a possible mechanism responsible for the inhibitory effects of quercetin glucosides [89,90].

All in all, quercetin is commercially available and is one of the most common natural products. Natural products have become more popular, and they have started to be used as lead compounds in medicine. They have a lot of advantages in contrast to common drugs. For instance, they have less side effects. In addition, flavonoids play a significant role in humans and plants. Efforts that have been made to increase the bioavailability and solubility of quercetin are outlined in the Results and Discussion section (Section 3). Basically, vehicles have been used, and quercetin is also administered with other drugs. We propose that a mixture of quercetin engulfed in vehicles along with other drugs should be tried (Scheme 3).

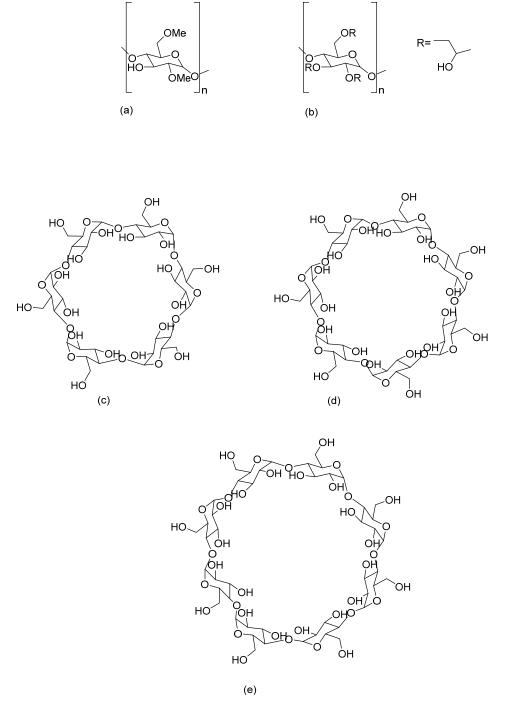


**Scheme 3.** Quercetin alone bears low bioavailability and solubility. In an attempt to increase both its bioavailability and solubility, it was encapsulated in vehicles using nanotechnology. In addition, it is administered with other drugs. We propose that it can be administered with drugs but also given as an encapsulated molecule in vehicles.

## 3. Results and Discussion

Quercetin interacts with various biomolecules, including cyclodextrin [91–95]. Cyclodextrins are macromolecules that are commonly employed in the food and pharmaceutical industries, serving diverse purposes within these fields [96].

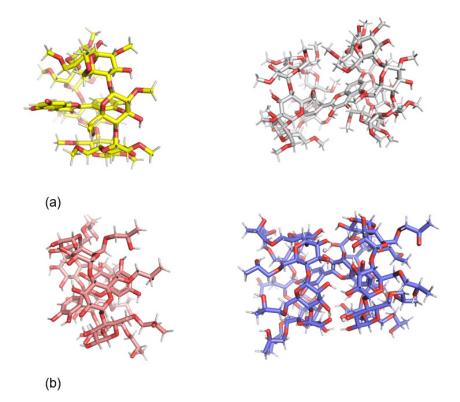
Cyclodextrins [97,98] are cyclic macromolecules composed of glucopyranose units [99,100]. The outer surface of cyclodextrins is hydrophilic, and the inner is hydrophobic [101]. They are soluble in water. The most common cyclodextrins are composed of six, seven, and eight glucopyranose units, which are  $\alpha$ -,  $\beta$ -, and  $\gamma$ -cyclodextrins (Scheme 4). Nevertheless, nowadays, a plethora of cyclodextrin derivatives have been synthesized that are comprised of less than six glucopyranose units. Larger cyclodextrins have also been achieved with more than eight glucopyranose units. Indeed, a wide range of cyclodextrins can be generated by employing different substitutes, leading to a diverse array of these molecules, with varied properties and applications. CDs can improve the bioavailability of drugs [99,102]. They play a vital role in computational drug design, and there are a lot of medicines with CD-drug complexes that are already in commercial use [103]. One example of CD-drug complexes is those with curcumin [101,104,105].



**Scheme 4.** Structure sketched in ChemDraw of (**a**) Me- $\beta$ -CD, (**b**) 2HP- $\beta$ -CD, (**c**)  $\alpha$ -cyclodextrin, and (**d**)  $\beta$ -cyclodextrin and (**e**)  $\gamma$ -cyclodextrin.

The synthesis of these complexes was performed by freeze-drying [106]. Moreover, 2D DOSY NMR experiments have shown the complexation between quercetin and 2HP- $\beta$ -CD and 2,6Me- $\beta$ -CD (Scheme 5). Quantum chemistry studies employing Density Functional Theory have shown that the binding between quercetin and dimeric assemblies of 2HP- $\beta$ -CD and 2,6Me- $\beta$ -CD (Scheme 5) cyclodextrins is relatively weak, leading to facile quercetin entrance and exit from the CD vehicle [92]. Fluorescence spectroscopy and molecular Dynamics experiments have shown that these dimeric assemblies remain stable. The quercetin-CD complex is stabilized through hydrophobic interactions. However, the 2HP- $\beta$ -CD<sub>2</sub> dimeric assembly is more stable than 2,6Me- $\beta$ -CD<sub>2</sub>, due to stronger binding [91].

The weak quercetin–CD binding allows quercetin to remain available at the intended target site, facilitating selective and safe action. Solubility experiments conducted on these complexes revealed an increase in the solubility of quercetin when encapsulated within cyclodextrins, particularly noticeable at pH 6.8.

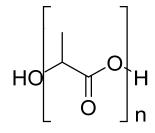


**Scheme 5.** Calculated minimum structure of complex (yellow and pink, right) and dimeric assembly (white and blue, left): (**a**) QUE with 2,6Me-β-CD and (**b**) QUE with 2HP-β-CD. This image was sketched in Pymol.

Furthermore, the systematic exploration and analysis of quercetin-based compounds that are chemically modified through the incorporation of amino acids was examined. In silico experiments, in vitro assays in different cancer cells, and NMR spectroscopy were used to reveal the interactions between these analogues with the Bcl-xl protein. It was shown that these analogues bind strongly in the protein and remain stable in the active center. Specifically, the conjugation of quercetin with amino acids, particularly Que-Glu (Scheme 1e), enhances quercetin's inhibitory effects on prostate cancer cells. This approach could offer a promising strategy to improve the therapeutic efficacy of these compounds [107].

One other approach to increase the bioavailability of quercetin is the use of nanoparticles [8,42]. The encapsulation of quercetin in nanoparticles has shown high solubility. This has led to the treatment of various diseases. The size of quercetin nanostructures is between 20 and 50 nm. Studies have shown that quercetin has more antioxidant activity inside nanostructures than free quercetin does. Even though there are treatments with quercetin against cancer, there is low availability of quercetin-involved nanoparticles to treat neurodegenerative diseases [108–112].

One other biomolecule that is used for the encapsulation of drugs is poly-d,l-lactide (PLA) (Scheme 6). PLA is often used to form complexes with other molecules due to its high hydrophobicity, biodegradability, and low toxicity. In vitro experiments and fluorescence experiments showed that the antioxidant activity of quercetin was retained inside the polymer. This may lead to the development of nanomedicine and to an antioxidant drug [113].



Scheme 6. Structure of Poly (D,L-lactide) sketched in ChemDraw.

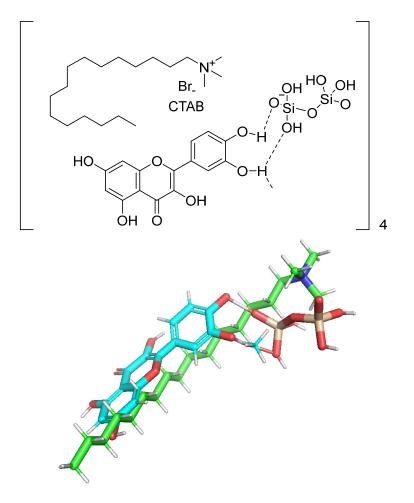
There are several examples in which quercetin and its derivatives are combined with other compounds with biological interest and activity (Table 1).

Table 1. Examples of quercetin in combination with other compounds in medicinal chemistry.

Additive	<b>Treatment of Disease</b>
Vitamin C, vitamin B3, folic acid	Sickle cell disease (SCD) [114,115]
Catechin	Synergistic inhibition of the platelet function and dietary use [116]
Kaempferol	Prevention and treatment of hereditary cardiomyopathy [117]
Astragalin	Treatment of atopic dermatitits [118]
Statins	Reducing cholesterol levels [119]
Doxorubicin	Inhibiting liver cancer [120]
Oleuropein	Preventing and treating joint disorders [121
Ibudilast	Treatment of Fragile X Syndrome [122]
Zafirlukast	Treating amyotrophic lateral sclerosis [123]
Rutin	Treating elevated blood lipid level-related diseases [124]
Polyphosphate	Treating osteoporosis [125]
Icaritin	Treatment of liver disease [126]
Vitamin D, retinol, and genistein	Improvement of skin conditions [127–129]
Maleic anhydride derivatives	Treatment of hepatocellular carcinoma [130
Haloperidol	Releaving neuropathic pain [131]
Metformin	Preventing against immune diseases [132]
Luteonil and delphinidin	Treatment of endometriosis [133]
Myrecetin	Curing adenocarcinoma, prostate carcinom and breast cancer [134]

Maroto et al. have developed a combination of resveratrol, quercetin, and catechin polyphenols in such proportions that it has a synergistic antioxidant power. They have shown that the preferred embodiment is the combination of antioxidants comprising resveratrol: quercetin: catechin in a 1:1:2 or 1:1:5 molar ratio. The results of the TOSC (Total Oxidant Scavenging Capacity) test showed that it was possible to obtain a potent antioxidant effect without the need to ingest large amounts of resveratrol or the other individual antioxidants, which is advantageous in order to minimize possible risks of side effects. The preferred combination is resveratrol: quercetin: catechin in a 1:1:2 molar ratio. The combination of resveratrol, quercetin, and catechin polyphenols can be used in different pharmaceutical forms, both solid and semi-solid, and these can be included in a variety of pharmaceutical, cosmetic, and foodstuff formulations [135].

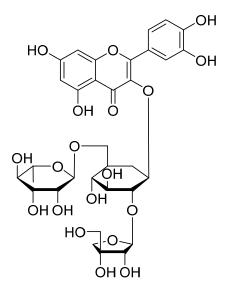
In another study, it was found that quercetin can be encapsulated in nanoparticles. Specifically, quercetin and silica nanogels were synthesized by aging and drying. The encapsulation was performed in PEGylated and CTAB-modified polymer nanomaterials (Scheme 7). An IR experiment confirmed the stability of quercetin inside the nanoparticles. The antioxidant activity of quercetin was tested in Cu(II), which is a metal that can induce oxidative stress. The entrapment of quercetin within nanoparticles demonstrated its ability to release its contents when exposed to conditions of Cu(II)-induced oxidative stress in neuronal and glial cultures. Such research works aim toward the development of flavonoids in nanomedicine and toward the treatment of Cu(II)-induced oxidative stress in neurodegenerative diseases [136].



**Scheme 7.** Structure of encapsulated quercetin (blue) inside CTAB nanomaterial (green) in 2D and 3D. This image was sketched in ChemDraw and Pymol.

As mentioned before, quercetin is being extensively studied as a potential cancer treatment because of its notable characteristics, such as its ability to regulate crucial molecular pathways linked to apoptosis and its effectiveness in inhibiting drug efflux in multidrugresistant tumors. To address the limitation of poor bioavailability, new formulations leveraging nanotechnology have emerged as a promising solution. Recent in vitro and in vivo studies have showcased successful tumor treatments using nano-formulations loaded with quercetin across various cancer models. These formulations exhibit high quercetin loading percentages in polymeric, lipid, and inorganic nanoparticles. Additionally, the co-delivery of different therapeutic agents has emerged as a promising strategy to elicit synergistic effects. Notably, quercetin has been shown to downregulate membrane transporter proteins, leading to increased intracellular concentrations of other chemotherapeutic compounds, and ultimately improving therapeutic outcomes. These studies are promising for the development and the augmentation of a new series of anti-tumor drugs [42].

Additionally, quercetin-sugar derivatives, which are depicted in Scheme 8, have been synthesized and used for the prevention and treatment of diseases related to the  $5HT_{1A}$  receptor by inhibiting it, or neuron cell damages, including drug or alcohol dependence, sleep disorders, panic state, delaying senility, and improving learning and memory [137].

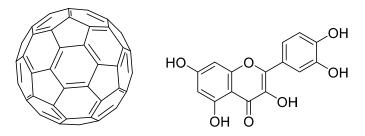


**Scheme 8.** A quercetin-sugar derivative that has been used for treating neurological disorders sketched in ChemDraw.

Another study showed that quercetin encapsulated in liposomes can be a candidate for the treatment of ischemia [138]. Liposomes can be used as drug carriers like cyclodextrins. Specifically, the synthesis of quercetin with liposomes was conducted with a thin-film hydration method. In vivo experiments with rats revealed a potent antioxidant activity of quercetin with this nano-formulation.

In the case of flavonoids, solvents play a leading role in their activity. The solvent seems to have an effect on hydrogen bonding through the available donor-acceptor sites in the flavonoid. Based on this, many studies focus on the solubility of flavonoids, especially quercetin, in different organic solvents.

Previous studies have shown that C60 fullerene leads to moderate toxicity because of its low water solubility. This may be harmful to aquatic organisms. One effective way to prevent this is the combination of C60-quercetin solutions (Scheme 9). As a result, the solubilization of C60 with quercetin leads to more biodegradable materials [139]. In recent years, scientists have achieved the synthesis of novel C60 fullerene-flavone derivatives, starting from quercetin, via cyclopropanation (Bingel reaction) of C60. The products of this reaction have antioxidant activity and may be used as novel drug leads [140].



Scheme 9. C60 fullerene with quercetin sketched in ChemDraw.

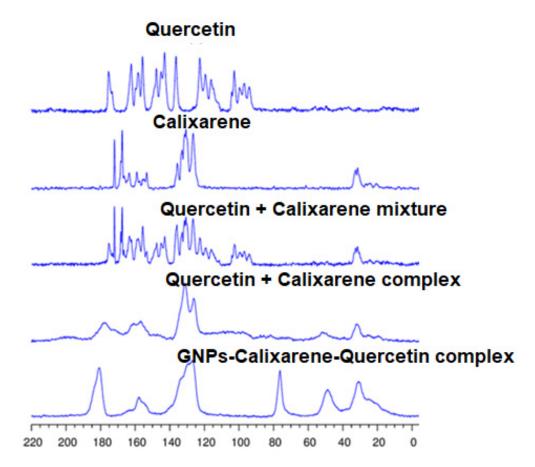
In 2023, Das Saha et al. [141] examined the anticancer potential of quercetin and 5fluorouracil-encapsulated (5-Fu) chitosan nanoparticles, since chitosan has been intensively investigated and used as a carrier in polymeric nanoparticles for drug delivery in both in vitro and in vivo models [142]. 5-Fu is already a chemotherapeutic drug approved by the FDA for treating various types of cancers [143], and quercetin has the ability to express its anticarcinogenic properties via the modification of intracellular signal transduction and the inhibition of cancer-activating enzymes [144]. These nanoparticles were synthesized using ionic gelation methods and were tested against cancer cell lines HepG2 (liver cancer), HCT116 (colorectal carcinoma), and HeLa (cervical cancer) and the normal cell line Hek293 (kidney cells). The results of the performed MTT assays indicated a higher cytotoxic potential of CS-5Fu-QCT nanoparticles in HCT116 cancer cells and no toxicity on the tested normal kidney cells compared to quercetin alone, meaning that this type of NP may be a very effective anticancer agent against colorectal carcinoma, with minimum to no side effects. The CS-5Fu-QCT NPs' possible mechanism of action works by causing G0/G1 phase cell-cycle arrest in HCT116 cells and altering the expression of pivotal proteins in the p53/p21 pathway, thus initiating cell apoptosis [141].

The field of nanotechnology, specifically nanomedicine, is an alternative and promising tool in the biomedical sciences. Nanomaterials, nanotubes, and nanoparticles offer a new perspective, and their use and application as carriers can contribute substantially to disease diagnosis, treatment, and monitoring [145–150]. The characteristics of these materials, such as their biocompatibility and economic viability, make them satisfactory carriers applied to multiple therapeutic and diagnostic agents [151,152]. Researchers who are motivated by the possibilities of nanomaterial, have found the effective interaction of a nanostructure and quercetin, and particularly the incorporation of a hybrid nanostructure with quercetin-coated titanate nanotube [153]. Studies report that a hybrid nanostructure, especially metal-based (High-Z elements) hybrid nanoparticles (MHNs), noble metals, and organic materials, offer an improvement in the efficacy of radiation therapy and have demonstrated cytotoxicity against tumor cells [154]. Moreover, there is evidence demonstrating that despite the unique and easy modification of a titanate nanostructure, the connection and incorporation of quercetin on it does not alter the morphology of the nanostructure, and its tubular structure is preserved [153]. Due to these facts, the investigation and development of incorporated molecules of quercetin in sodium (NaTNT) and zinc (ZnTNT) titanate nanotubes could interfere in cell proliferation and may be a powerful tool in a medical revolution.

Finally, the complexation of quercetin with the calixarene supramolecule was conducted (Scheme 10). To examine this complexation, several analytical techniques were used such as FTIR Analysis, Dynamic and Electrophoretic Light Scattering (DLS), Differential Scanning Calorimetry (DSC), and High-Resolution Transmission Electron Microscopy (HR-TEM). At first, quercetin was enclosed within a calixarene. This resulted in a significant increase of 62,000 times in aqueous solubility. Through solid-state NMR (Scheme 11) and in vitro and in silico experiments, it was found that the complex effectively hindered the growth of tumors, leading to a decrease in tumor volume. Next, a gold nanoparticle core was adorned with calixarene hosts to non-covalently accommodate nanoparticles (GNPs). The nanocarrier loaded with the NP quercetin significantly increased the cytotoxicity (more than 50-fold) compared to the original NP in colon cancer, and it also modified its cell membrane transport mechanism. This enhanced the tumor-targeting properties achieved through this innovative combination, shedding light on a promising avenue for advanced cancer therapies [155].



**Scheme 10.** Calixarene interacting with quercetin (purple with red) through DFT methodology. Sketched in Pymol.



**Scheme 11.** 1H–13C CPMAS NMR spectra of quercetin, calixarene mixture, quercetin, and calixarene complex and GNP-calixarene-quercetin complex sketched in TopSpin 4.2.0. software.

In Table 2, it is shown all the techniques that they were used for the encapsulation of quercetin in macromolecules.

**Table 2.** Overall table of techniques that were used for the encapsulation of quercetin in macro-molecules [156].

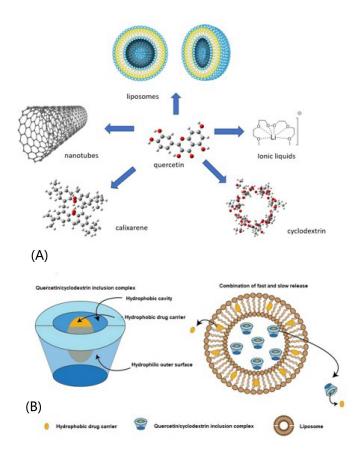
Technique	Reason That It Was Used
NMR spectroscopy	Structure elucidation of quercetin with cyclodextrins and observation of their complexation [91,157]
2D-DOSY NMR spectroscopy	Evaluation of the complex formation of quercetin with cyclodextrins [92,158]
Induced Fit Docking (IFD)	Evaluation of how effectively quercetin binds to essential vira components or enzymes. For instance, quercetin was used for IFD against acetylcholinesterase and butyrelcholinesterase [159–162].
Molecular dynamics	To obtain a deeper understanding of the stability and molecula interactions within the complexes formed by the "protein-ligand" pairs identified in the docking studies [92,163–167]
Molecular Mechanics Generalized Born Surface Area (MM GBSA)	To highlight the strongest binding capability of quercetin against different macromolecules [92,168]
Differential Scanning Calorimetry (DSC)	Validation of the formation of the complexes. For example, it was shown that quercetin was well distributed in the polyvinylpyrrolidone (PVP) matrix [169–173].
Fluorescence spectroscopic studies	Investigation of the interactions between quercetin and macromolecules. In particular, it was used in the formation of the dimeric assemblies of quercetin with cyclodextrins [91,174–180].
Solubility studies	Examination of the solubility of quercetin inside macromolecules in different pHs [155,181,182]
High-performance liquid chromatography (HPLC)	Validation of the purity and identification of the components. was used for the determination of quercetin in herbal extracts [183–186].
Gas chromatography (GC)	Analysis of quercetin and its separation from different plants materials, etc. [187–189]
UV/Vis spectroscopy	Quantification of quercetin in various contexts, encompassing pharmaceutical formulations [190], medicinal plants, beverage [191,192], and food.
Thin-layer chromatography (TLC)	Separation of quercetin from other flavonoids in a shared matr [193–196]
Electrophoresis	Analysis of quercetin [197–201]
Cyclic voltammetry (CV)	Determination of the antioxidant activity of quercetin in lyophilized onion tissue of onion var [202–204]
Pulse voltammetry (DPV)	Determination of the antioxidant activity and the electrochemical parameters of quercetin [205]
Raman spectroscopy	Quantitative analysis of quercetin in onion peels [206–210]
Limit of detection (LOD) and limit of quantitation (LOQ)	Validation of the analytical method by determining quercetin green tea [211]
Transmission Electron Microscopy (TEM)	Details for structural properties of quercetin in oil-in-water nanoemulsions [212,213]
Central Composite Design (CCD)	Evaluation of the effects of pH in determining quercetin in the presence of electroactive tannic acid [214,215]
Rheological measurements	Evaluation of the strength of the structure of quercetin with nanostructured lipid carriers in linseed oil [216]

Technique	<b>Reason That It Was Used</b>
Liquid Chromatography-Mass Spectroscopy (LC-MS)	Identification and quantification of quercetin in human hepatocytes as in vitro cell models [217]
Fourier-Transform Infrared Spectroscopy (FT-IR)	Analyzing the infrared absorption or emission of the molecule in buckwheat samples [218,219]
Capillary electrophoresis (CE)	Analysis of quercetin based on its electrophoretic mobility in red and white wine samples [197,220]
Enzyme-Linked Immunosorbent Assay (ELISA)	Quantitative analysis of quercetin to determine its anti-inflammatory effects in lipopolysaccharide stimulated cells [221–223]
Supercritical Fluid Chromatography (SFC)	Separation and extraction of quercetin from sumac fruits [224,225]
Flow Injection Analysis (FIA)	Subsequent detection of quercetin using normal and hot platinum microelectrode, showing the utility of Baranski's method [226,227]
Solid-Phase Microextraction (SPME)	Extraction and analysis of quercetin, combined with HPLC-UV detection method, in green and black tea and coffee samples [184,228,229]
X-ray crystallography	Determination of the three-dimensional structure of quercetin crystals existing as hydrogen-bonded dimers, contributing to its unique biological activities [230]
Matrix-Assisted Laser Desorption/Ionization Mass Spectrometry (MALDI-MS)	Analysis of quercetin utilizing MIL-101(Cr) as surface-assisted matrix for replacing traditional organic matrices [231–234]
Supercritical Fluid Extraction (SFE)	Extraction of quercetin from Hyperici herba [224,235]
Solid-Phase Extraction (SPE)	Preparation of samples for extracting and determining quercetin's and quercetin glucosides' concentration in food products [236–240]

 Table 2. Cont.

## 4. Conclusions

In conclusion, this review delves into the diverse biological and medicinal implications of quercetin, especially its anti-inflammatory, antioxidant, anti-tumor, and antiviral properties. The exploration of quercetin's synergistic potential when combined with other drugs and natural products holds promise for the development of innovative medications. Such combinations not only offer the prospect of novel drugs but also present an avenue for mitigating side effects and toxicity. Addressing the challenge of quercetin's poor bioavailability, researchers have successfully conducted complexation within various macromolecules, including cyclodextrins, polymers, liposomes, and nanomaterials. Both experimental and computational efforts have resulted in stable complexes, demonstrating enhanced bioavailability in vitro. Future applications may involve quercetin complexes with cyclodextrins inside the liposomes, suggesting a wide spectrum of medicinal possibilities (Scheme 12A). In addition, combined macromolecules like cyclodextrins and calixarenes can be used to engulf quercetin (Scheme 12B). All these suggested future studies will designate the capability of quercetin to serve as a lead compound.



**Scheme 12.** Future formulations of quercetin in various macromolecules sketched in PowerPoint installed on Windows 10. Quercetin can be incorporated into various macromolecules such as nanotubes, fullerenes, etc., along with their combinations (**A**). In addition, quercetin can be incorporated into liposomes in a simple form or complexed in CDs or calixarenes or other forms in which slow or fast release can be controlled (**B**).

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#### References

- 1. Pallag, A.; Bungau, S.; Tit, D.M.; Jurca, T.; Sirbu, V.; Honiges, A.; Horhogea, C. Comparative Study of Polyphenols, Flavonoids and Chlorophylls in *Equisetum arvense* L. Populations. *Rev. Chim.* **2016**, *67*, 530–533.
- 2. Panche, A.N.; Diwan, A.D.; Chandra, S.R. Flavonoids: An Overview. J. Nutr. Sci. 2016, 5, e47. [CrossRef] [PubMed]
- Li, Y.; Yao, J.; Han, C.; Yang, J.; Chaudhry, M.T.; Wang, S.; Liu, H.; Yin, Y. Quercetin, Inflammation and Immunity. *Nutrients* 2016, 8, 167. [CrossRef] [PubMed]
- Formica, J.V.; Regelson, W. Review of the Biology of Quercetin and Related Bioflavonoids. *Food Chem. Toxicol.* 1995, 33, 1061–1080. [CrossRef] [PubMed]
- Yang, D.; Wang, T.; Long, M.; Li, P. Quercetin: Its Main Pharmacological Activity and Potential Application in Clinical Medicine. Oxid. Med. Cell. Longev. 2020, 2020, 8825387. [CrossRef] [PubMed]
- 6. Kim, J.K.; Park, S.U. Quercetin and Its Role in Biological Functions: An Updated Review. EXCLI J. 2018, 17, 856–863. [CrossRef]
- 7. Singh, P.; Arif, Y.; Bajguz, A.; Hayat, S. The Role of Quercetin in Plants. Plant Physiol. Biochem. 2021, 166, 10–19. [CrossRef]

- 8. Amanzadeh, E.; Esmaeili, A.; Rahgozar, S.; Nourbakhshnia, M. Application of Quercetin in Neurological Disorders: From Nutrition to Nanomedicine. *Rev. Neurosci.* **2019**, *30*, 555–572. [CrossRef]
- 9. E Mendoza, E.; Burd, R. Quercetin as a Systemic Chemopreventative Agent: Structural and Functional Mechanisms. *Mini-Rev. Med. Chem.* **2012**, *11*, 1216–1221. [CrossRef]
- 10. Daina, A.; Michielin, O.; Zoete, V. SwissADME: A Free Web Tool to Evaluate Pharmacokinetics, Drug-Likeness and Medicinal Chemistry Friendliness of Small Molecules. *Sci. Rep.* 2017, 7, 42717. [CrossRef]
- 11. Pires, D.E.V.; Blundell, T.L.; Ascher, D.B. PkCSM: Predicting Small-Molecule Pharmacokinetic and Toxicity Properties Using Graph-Based Signatures. J. Med. Chem. 2015, 58, 4066–4072. [CrossRef] [PubMed]
- Viana Nunes, A.M.; das Chagas Pereira de Andrade, F.; Filgueiras, L.A.; de Carvalho Maia, O.A.; Cunha, R.L.O.R.; Rodezno, S.V.A.; Maia Filho, A.L.M.; de Amorim Carvalho, F.A.; Braz, D.C.; Mendes, A.N. PreADMET Analysis and Clinical Aspects of Dogs Treated with the Organotellurium Compound RF07: A Possible Control for Canine Visceral Leishmaniasis? *Environ. Toxicol. Pharmacol.* 2020, *80*, 103470. [CrossRef] [PubMed]
- 13. Guan, L.; Yang, H.; Cai, Y.; Sun, L.; Di, P.; Li, W.; Liu, G.; Tang, Y. ADMET-Score—A Comprehensive Scoring Function for Evaluation of Chemical Drug-Likeness. *Medchemcomm* **2019**, *10*, 148–157. [CrossRef] [PubMed]
- Georgiou, N.; Chontzopoulou, E.; Cheilari, A.; Katsogiannou, A.; Karta, D.; Vavougyiou, K.; Hadjipavlou-Litina, D.; Javornik, U.; Plavec, J.; Tzeli, D.; et al. Thiocarbohydrazone and Chalcone-Derived 3,4-Dihydropyrimidinethione as Lipid Peroxidation and Soybean Lipoxygenase Inhibitors. ACS Omega 2022, 8, 11966–11977. [CrossRef] [PubMed]
- 15. Georgiou, N.; Gouleni, N.; Chontzopoulou, E.; Skoufas, G.S.; Gkionis, A.; Tzeli, D.; Vassiliou, S.; Mavromoustakos, T. Structure Assignment, Conformational Properties and Discovery of Potential Targets of the Ugi Cinnamic Adduct NGI25. *J. Biomol. Struct. Dyn.* **2021**, *41*, 1253–1266. [CrossRef] [PubMed]
- 16. Georgiou, N.; Cheilari, A.; Karta, D.; Chontzopoulou, E.; Plavec, J.; Tzeli, D.; Vassiliou, S.; Mavromoustakos, T. Conformational Properties and Putative Bioactive Targets for Novel Thiosemicarbazone Derivatives. *Molecules* **2022**, *27*, 4548. [CrossRef]
- Georgiou, N.; Katsogiannou, A.; Skourtis, D.; Iatrou, H.; Tzeli, D.; Vassiliou, S.; Javornik, U.; Plavec, J.; Mavromoustakos, T. Conformational Properties of New Thiosemicarbazone and Thiocarbohydrazone Derivatives and Their Possible Targets. *Molecules* 2022, 27, 2537. [CrossRef]
- 18. Ginex, T.; Vazquez, J.; Gilbert, E.; Herrero, E.; Luque, F.J. Lipophilicity in Drug Design: An Overview of Lipophilicity Descriptors in 3D-QSAR Studies. *Future Med. Chem.* **2019**, *11*, 1177–1193. [CrossRef]
- Veber, D.F.; Johnson, S.R.; Cheng, H.-Y.; Smith, B.R.; Ward, K.W.; Kopple, K.D. Molecular Properties That Influence the Oral Bioavailability of Drug Candidates. J. Med. Chem. 2002, 45, 2615–2623. [CrossRef]
- 20. Li, G.; Wang, Y.; Fei, T.; Wu, D.; Tao, L. Application of Quercetin or Quercetin Derivative for Relieving Smoke Harm.
- Daina, A.; Zoete, V. A BOILED-Egg to Predict Gastrointestinal Absorption and Brain Penetration of Small Molecules. *ChemMed-Chem* 2016, 11, 1117–1121. [CrossRef] [PubMed]
- 22. Mathew, B.; Carradori, S.; Guglielmi, P.; Uddin, M.S.; Kim, H. New Aspects of Monoamine Oxidase B Inhibitors: The Key Role of Halogens to Open the Golden Door. *Curr. Med. Chem.* **2020**, *28*, 266–283. [CrossRef] [PubMed]
- 23. Zhao, L.; Funk, C.D. Lipoxygenase Pathways in Atherogenesis. Trends Cardiovasc. Med. 2004, 14, 191–195. [CrossRef] [PubMed]
- 24. Hosseini, A.; Razavi, B.M.; Banach, M.; Hosseinzadeh, H. Quercetin and Metabolic Syndrome: A Review. *Phyther. Res.* 2021, *35*, 5352–5364. [CrossRef] [PubMed]
- Yoon, B.H.; Jung, J.W.; Lee, J.-J.; Cho, Y.-W.; Jang, C.-G.; Jin, C.; Oh, T.H.; Ryu, J.H. Anxiolytic-like Effects of Sinapic Acid in Mice. Life Sci. 2007, 81, 234–240. [CrossRef] [PubMed]
- Nunes, A.R.; Vieira, Í.G.P.; Queiroz, D.B.; Leal, A.L.A.B.; Maia Morais, S.; Muniz, D.F.; Calixto-Junior, J.T.; Coutinho, H.D.M. Use of Flavonoids and Cinnamates, the Main Photoprotectors with Natural Origin. *Adv. Pharmacol. Sci.* 2018, 2018, 5341487. [CrossRef]
- Fogaça, L.A.; Feuser, P.E.; Ricci-Júnior, E.; Hermes de Araújo, P.H.; Sayer, C.; da Costa, C. ZnO and Quercetin Encapsulated Nanoparticles for Sun Protection Obtained by Miniemulsion Polymerization Using Alternative Co-Stabilizers. *Mater. Res. Express* 2020, 7, 015096. [CrossRef]
- 28. Reeves, J.D.; Doms, R.W. Human Immunodeficiency Virus Type 2. J. Gen. Virol. 2002, 83, 1253–1265. [CrossRef]
- 29. Sharp, P.M.; Hahn, B.H. The Evolution of HIV-1 and the Origin of AIDS. *Philos. Trans. R. Soc. B Biol. Sci.* 2010, 365, 2487–2494. [CrossRef]
- 30. Di Petrillo, A.; Orrù, G.; Fais, A.; Fantini, M.C. Quercetin and Its Derivates as Antiviral Potentials: A Comprehensive Review. *Phyther. Res.* **2022**, *36*, 266–278. [CrossRef]
- 31. Wang, G.; Wang, Y.; Yao, L.; Gu, W.; Zhao, S.; Shen, Z.; Lin, Z.; Liu, W.; Yan, T. Pharmacological Activity of Quercetin: An Updated Review. *Evid.-Based Complement. Altern. Med.* **2022**, 2022, 3997190. [CrossRef] [PubMed]
- 32. Agrawal, P.K.; Agrawal, C.; Blunden, G. Quercetin: Antiviral Significance and Possible COVID-19 Integrative Considerations. *Nat. Prod. Commun.* **2020**, *15*, 1934578X20976293. [CrossRef]
- Moschovou, K.; Antoniou, M.; Chontzopoulou, E.; Papavasileiou, K.D.; Melagraki, G.; Afantitis, A.; Mavromoustakos, T. Exploring the Binding Effects of Natural Products and Antihypertensive Drugs on SARS-CoV-2: An In Silico Investigation of Main Protease and Spike Protein. *Int. J. Mol. Sci.* 2023, 24, 15894. [CrossRef] [PubMed]
- 34. Oz, M.; Selcuk, I.; Arik, Z.; Gungor, T. Targeted Agents in Ovarian Carcinoma. Med. Sci. 2016, 5, 547. [CrossRef]

- 35. Erdogan, S.; Turkekul, K.; Dibirdik, I.; Doganlar, O.; Doganlar, Z.B.; Bilir, A.; Oktem, G. Midkine Downregulation Increases the Efficacy of Quercetin on Prostate Cancer Stem Cell Survival and Migration through PI3K/AKT and MAPK/ERK Pathway. *Biomed. Pharmacother.* **2018**, *107*, 793–805. [CrossRef]
- Ward, A.B.; Mir, H.; Kapur, N.; Gales, D.N.; Carriere, P.P.; Singh, S. Quercetin Inhibits Prostate Cancer by Attenuating Cell Survival and Inhibiting Anti-Apoptotic Pathways. World J. Surg. Oncol. 2018, 16, 108. [CrossRef]
- 37. Hisaka, T.; Sakai, H.; Sato, T.; Goto, Y.; Nomura, Y.; Fukutomi, S.; Fujita, F.; Mizobe, T.; Nakashima, O.; Tanigawa, M.; et al. Quercetin Suppresses Proliferation of Liver Cancer Cell Lines In Vitro. *Anticancer Res.* **2020**, *40*, 4695–4700. [CrossRef]
- Niazvand, F.; Orazizadeh, M.; Khorsandi, L.; Abbaspour, M.; Mansouri, E.; Khodadadi, A. Effects of Quercetin-Loaded Nanoparticles on MCF-7 Human Breast Cancer Cells. *Medicina (B. Aires)* 2019, 55, 114. [CrossRef]
- 39. Pham, T.N.D.; Stempel, S.; Shields, M.A.; Spaulding, C.; Kumar, K.; Bentrem, D.J.; Matsangou, M.; Munshi, H.G. Quercetin Enhances the Anti-Tumor Effects of BET Inhibitors by Suppressing HnRNPA1. *Int. J. Mol. Sci.* **2019**, *20*, 4293. [CrossRef]
- 40. Sturza, A.; Pavel, I.; Ancușa, S.; Danciu, C.; Dehelean, C.; Duicu, O.; Muntean, D. Quercetin Exerts an Inhibitory Effect on Cellular Bioenergetics of the B164A5 Murine Melanoma Cell Line. *Mol. Cell. Biochem.* **2018**, 447, 103–109. [CrossRef]
- Wu, L.; Li, J.; Liu, T.; Li, S.; Feng, J.; Yu, Q.; Zhang, J.; Chen, J.; Zhou, Y.; Ji, J.; et al. Quercetin Shows Anti-tumor Effect in Hepatocellular Carcinoma LM3 Cells by Abrogating JAK2/STAT3 Signaling Pathway. *Cancer Med.* 2019, *8*, 4806–4820. [CrossRef] [PubMed]
- Caro, C.; Pourmadadi, M.; Eshaghi, M.M.; Rahmani, E.; Shojaei, S.; Paiva-Santos, A.C.; Rahdar, A.; Behzadmehr, R.; García-Martín, M.L.; Díez-Pascual, A.M. Nanomaterials Loaded with Quercetin as an Advanced Tool for Cancer Treatment. *J. Drug Deliv. Sci. Technol.* 2022, 78, 103938. [CrossRef]
- 43. Tanaka, Y. Inflammation and Regeneration Rheumatoid Arthritis. BioMed Cent. 2020, 40, 1-8.
- 44. Yuan, K.; Zhu, Q.; Lu, Q.; Jiang, H.; Zhu, M.; Li, X.; Huang, G.; Xu, A. Quercetin Alleviates Rheumatoid Arthritis by Inhibiting Neutrophil Inflammatory Activities. J. Nutr. Biochem. 2020, 84, 108454. [CrossRef] [PubMed]
- Chontzopoulou, E.; Papaemmanouil, C.D.; Chatziathanasiadou, M.V.; Kolokouris, D.; Kiriakidi, S.; Konstantinidi, A.; Gerogianni, I.; Tselios, T.; Kostakis, I.K.; Chrysina, E.D.; et al. Molecular Investigation of Artificial and Natural Sweeteners as Potential Anti-Inflammatory Agents. J. Biomol. Struct. Dyn. 2021, 40, 12608–12620. [CrossRef] [PubMed]
- Smirnova, E.O.; Egorova, A.M.; Lantsova, N.V.; Chechetkin, I.R.; Toporkova, Y.Y.; Grechkin, A.N. Recombinant Soybean Lipoxygenase 2 (GmLOX2) Acts Primarily as a ω 6 (S)-Lipoxygenase. *Curr. Issues Mol. Biol.* 2023, 2, 6283–6295. [CrossRef] [PubMed]
- 47. Scheltens, P.; De Strooper, B.; Kivipelto, M.; Holstege, H.; Chételat, G.; Teunissen, C.E.; Cummings, J.; van der Flier, W.M. Alzheimer's Disease. *Lancet* 2021, 397, 1577–1590. [CrossRef]
- 48. Ruwizhi, N.; Aderibigbe, B.A. Cinnamic Acid Derivatives and Their Biological Efficacy. Int. J. Mol. Sci. 2020, 21, 5712. [CrossRef]
- Liao, Q.; Li, Q.; Zhao, Y.; Jiang, P.; Yan, Y.; Sun, H.; Liu, W.; Feng, F.; Qu, W. Design, Synthesis and Biological Evaluation of Novel Carboline-Cinnamic Acid Hybrids as Multifunctional Agents for Treatment of Alzheimer's Disease. *Bioorg. Chem.* 2020, 99, 103844. [CrossRef]
- Lan, J.S.; Hou, J.W.; Liu, Y.; Ding, Y.; Zhang, Y.; Li, L.; Zhang, T. Design, Synthesis and Evaluation of Novel Cinnamic Acid Derivatives Bearing N-Benzyl Pyridinium Moiety as Multifunctional Cholinesterase Inhibitors for Alzheimer's Disease. J. Enzym. Inhib. Med. Chem. 2017, 32, 776–788. [CrossRef]
- Salehi, B.; Machin, L.; Monzote, L.; Sharifi-Rad, J.; Ezzat, S.M.; Salem, M.A.; Merghany, R.M.; El Mahdy, N.M.; Klllç, C.S.; Sytar, O.; et al. Therapeutic Potential of Quercetin: New Insights and Perspectives for Human Health. ACS Omega 2020, 5, 11849–11872. [CrossRef] [PubMed]
- Xu, D.; Hu, M.-J.; Wang, Y.-Q.; Cui, Y.-L. Antioxidant Activities of Quercetin and Its Complexes for Medicinal Application. Molecules 2019, 24, 1123. [CrossRef] [PubMed]
- Tsiailanis, A.D.; Renziehausen, A.; Kiriakidi, S.; Vrettos, E.I.; Markopoulos, G.S.; Sayyad, N.; Hirmiz, B.; Aguilar, M.-I.; Del Borgo, M.P.; Kolettas, E.; et al. Enhancement of Glioblastoma Multiforme Therapy through a Novel Quercetin-Losartan Hybrid. *Free Radic. Biol. Med.* 2020, 160, 391–402. [CrossRef] [PubMed]
- 54. Ulusoy, H.G.; Sanlier, N. A Minireview of Quercetin: From Its Metabolism to Possible Mechanisms of Its Biological Activities. *Crit. Rev. Food Sci. Nutr.* **2020**, *60*, 3290–3303. [CrossRef] [PubMed]
- 55. Maroto, J.Á.M. Synergic Polyphenol Combination ES2391211B1. ES2391211B1, 2 October 2013.
- 56. De, P.; Bedos-Belval, F.; Vanucci-Bacque, C.; Baltas, M. Cinnamic Acid Derivatives in Tuberculosis, Malaria and Cardiovascular Diseases—A Review. *Curr. Org. Chem.* **2012**, *16*, 747–768. [CrossRef]
- 57. Ivanov, V.; Ivanova, S.; Roomi, W.; Niedzwicki, A.; Rath, M. Novel Composition and Method for the Treatment of Hypertension. US2004242504A1, 30 May 2003.
- 58. Jalili, T. Quercetin Supplementation to Treat Hypertenstion. US2004258674A1, 12 April 2004.
- Larson, A.J.; Symons, J.D.; Jalili, T. Quercetin: A Treatment for Hypertension?—A Review of Efficacy and Mechanisms. *Pharmaceuticals* 2010, *3*, 237–250. [CrossRef] [PubMed]
- 60. George, B.P.; Parimelazhagan, T.; Sajeesh, T.; Saravanan, S. Antitumor and Wound Healing Properties of Rubus Niveus Thunb. Root. J. Environ. Pathol. Toxicol. Oncol. 2014, 33, 145–158. [CrossRef]
- 61. Polera, N.; Badolato, M.; Perri, F.; Carullo, G.; Aiello, F. Quercetin and Its Natural Sources in Wound Healing Management. *Curr. Med. Chem.* **2019**, *26*, 5825–5848. [CrossRef]

- 62. Ezzat, S.M.; Choucry, M.A.; Kandil, Z.A. Antibacterial, Antioxidant, and Topical Anti-Inflammatory Activities of Bergia Ammannioides: A Wound-Healing Plant. *Pharm. Biol.* **2016**, *54*, 215–224. [CrossRef]
- 63. Pastorino, G.; Marchetti, C.; Borghesi, B.; Cornara, L.; Ribulla, S.; Burlando, B. Biological Activities of the Legume Crops Melilotus Officinalis and Lespedeza Capitata for Skin Care and Pharmaceutical Applications. *Ind. Crops Prod.* 2017, *96*, 158–164. [CrossRef]
- 64. Lodhi, S.; Jain, A.; Jain, A.P.; Pawar, R.S.; Singhai, A.K. Effects of Flavonoids from Martynia Annua and Tephrosia Purpurea on Cutaneous Wound Healing. *Avicenna J. Phytomed.* **2016**, *6*, 578–591. [PubMed]
- Tang, J.; Diao, P.; Shu, X.; Li, L.; Xiong, L. Quercetin and Quercitrin Attenuates the Inflammatory Response and Oxidative Stress in LPS-Induced RAW264.7 Cells: In Vitro Assessment and a Theoretical Model. *BioMed Res. Int.* 2019, 2019, 7039802. [CrossRef] [PubMed]
- Wei, B.; Zhang, Y.; Tang, L.; Ji, Y.; Yan, C.; Zhang, X. Protective Effects of Quercetin against Inflammation and Oxidative Stress in a Rabbit Model of Knee Osteoarthritis. *Drug Dev. Res.* 2019, *80*, 360–367. [CrossRef] [PubMed]
- 67. Sul, O.-J.; Ra, S.W. Quercetin Prevents LPS-Induced Oxidative Stress and Inflammation by Modulating NOX2/ROS/NF-KB in Lung Epithelial Cells. *Molecules* **2021**, *26*, 6949. [CrossRef]
- 68. Nakamura, M.; Fukuma, Y.; Notsu, K.; Kono, M. Quercetin and HSC70 Coregulate the Anti-Inflammatory Action of the Ubiquitin-like Protein MNSFβ. *Mol. Biol. Rep.* **2022**, *49*, 1213–1222. [CrossRef] [PubMed]
- Chen, T.; Zhang, X.; Zhu, G.; Liu, H.; Chen, J.; Wang, Y.; He, X. Quercetin Inhibits TNF-α Induced HUVECs Apoptosis and Inflammation via Downregulating NF-KB and AP-1 Signaling Pathway in vitro. *Medicine* 2020, 99, e22241. [CrossRef] [PubMed]
- 70. Ma, J.-Q.; Li, Z.; Xie, W.-R.; Liu, C.-M.; Liu, S.-S. Quercetin Protects Mouse Liver against CCl4-Induced Inflammation by the TLR2/4 and MAPK/NF-KB Pathway. *Int. Immunopharmacol.* **2015**, *28*, 531–539. [CrossRef]
- Bairwa, R.; Kakwani, M.; Tawari, N.R.; Lalchandani, J.; Ray, M.K.; Rajan, M.G.R.; Degani, M.S. Novel Molecular Hybrids of Cinnamic Acids and Guanylhydrazones as Potential Antitubercular Agents. *Bioorg. Med. Chem. Lett.* 2010, 20, 1623–1625. [CrossRef]
- 72. Sotgiu, G.; Centis, R.; D'Ambrosio, L.; Tadolini, M.; Castiglia, P.; Migliori, G.B. Do We Need a New Fleming Époque: The Nightmare of Drug-Resistant Tuberculosis. *Int. J. Mycobacteriol.* **2013**, *2*, 123–125. [CrossRef]
- 73. Sasikumar, K.; Ghosh, A.R.; Dusthackeer, A. Antimycobacterial Potentials of Quercetin and Rutin against Mycobacterium Tuberculosis H37Rv. *3 Biotech* 2018, *8*, 1–6. [CrossRef]
- 74. Huang, S.; Czech, M.P. The GLUT4 Glucose Transporter. Cell Metab. 2007, 5, 237–252. [CrossRef] [PubMed]
- 75. Dhanya, R. Quercetin for Managing Type 2 Diabetes and Its Complications, an Insight into Multitarget Therapy. *Biomed. Pharmacother.* **2022**, *146*, 112560. [CrossRef] [PubMed]
- Ahrens, M.J.; Thompson, D.L.; Atm Metabolics Lllp. Composition for Treating Diabetes and Metabolic Disorders with Quercetin, Myrcetin and Chlorogenic Acid. EP2129371B1, 11 March 2008.
- 77. Kruthiventi, A.; Javed, I. Pharmaceutical Co-Crystals of Quercetin. US20120258170A1, 30 October 2009.
- 78. Ansari, P.; Choudhury, S.T.; Seidel, V.; Rahman, A.B.; Aziz, M.A.; Richi, A.E.; Rahman, A.; Jafrin, U.H.; Hannan, J.M.A.; Abdel-Wahab, Y.H.A. Therapeutic Potential of Quercetin in the Management of Type-2 Diabetes Mellitus. *Life* 2022, 12, 1146. [CrossRef] [PubMed]
- 79. Ali, A.H.; Sudi, S.; Shi-Jing, N.; Rozianoor, W.; Hassan, M.; Basir, R.; Agustar, H.K.; Embi, N.; Sidek, H.M.; Latip, J. Dual Anti-Malarial and GSK3 β-Mediated Cytokine-Modulating Activities of Quercetin Are Requisite of Its Potential as a Plant-Derived Therapeutic in Malaria. *Pharmaceuticals* 2021, 14, 248. [CrossRef] [PubMed]
- 80. da Silva, A.A.; Maia, P.I.d.S.; Lopes, C.D.; de Albuquerque, S.; Valle, M.S. Synthesis, Characterization and Antichagasic Evaluation of Thiosemicarbazones Prepared from Chalcones and Dibenzalacetones. J. Mol. Struct. 2021, 1232, 130014. [CrossRef]
- Tasdemir, D.; Kaiser, M.; Brun, R.; Yardley, V.; Schmidt, T.J.; Tosun, F.; Ru, P. Antitrypanosomal and Antileishmanial Activities of Flavonoids and Their Analogues: In Vitro, In Vivo, Structure-Activity Relationship, and Quantitative Structure-Activity Relationship Studies. *Antimicrob. Agents Chemother.* 2006, 50, 1352–1364. [CrossRef] [PubMed]
- Fábio, M.; Rocha, G.; Sales, J.A.; Gleiciane, M.; Galdino, L.M.; Aguiar, L.D.; Pereira-Neto, W.D.A.; Cordeiro, R.D.A.; Souza, D.D.; Maia, C. Antifungal Effects of the Flavonoids Kaempferol and Quercetin: A Possible Alternative for the Control of Fungal Biofilms. *Biofouling* 2019, 35, 320–328. [CrossRef]
- 83. Tebbi, C.K. Sickle Cell Disease: A Review. Hemato 2022, 3, 341-366. [CrossRef]
- 84. Lozano, R.; Azarang, A.; Wilaisakditipakorn, T.; Hagerman, R.J. Fragile X Syndrome: A Review of Clinical Management. *Intractable Rare Dis. Res.* **2016**, *5*, 145–157. [CrossRef]
- 85. Chodoeva, R. Quercetin-Based Composition for Treating Rhinosinusitis. US2021000787A1, 15 February 2021.
- Tiboc-Schnell, C.N.; Filip, G.A.; Man, S.C.; Decea, N.; Moldovan, R.; Opris, R.; Sas, V.; Tabaran, F. Quercetin Attenuates Naso-Sinusal Inflammation and Inflammatory Response in Lungs and Brain on an Experimental Model of Acute Rhinosinusitis in Rats. J. Physiol. Pharmacol. 2020, 71, 479–490. [CrossRef]
- Zahedipour, F.; Kesharwani, P.; Sahebkar, A. Mechanisms of Multidrug Resistance in Cancer. In *Aptamers Engineered Nanocarriers* for Cancer Therapy; Elsevier: Amsterdam, The Netherlands, 2022; pp. 51–83. [CrossRef]
- Joshi, N.S.; Aggarwal, P.; Hiprara, V.K.; Jaggi, M.; Singh, A.; Awasthi, A.; Verma, R. Novel Quercetin Derivatives as Anti-Cancer Agents. US2011034413A1, 8 August 2008.
- 89. Otsuka, Y.; Egawa, K.; Kanzaki, N.; Izumo, T.; Rogi, T.; Shibata, H. Quercetin Glycosides Prevent Dexamethasone-Induced Muscle Atrophy in Mice. *Biochem. Biophys. Rep.* **2019**, *18*, 100618. [CrossRef] [PubMed]

- 90. Karaboga, A.S.; Perez-Neuno, V.I.; Souchet, M.; Decaudin, D. Muscle Atrophy Inhibitor Containing Quercetin Glycoside. CN106255500A, 2 April 2015.
- Leonis, G.; Vakali, V.; Zoupanou, N.; Georgiou, N.; Diamantis, D.A.; Tzakos, A.G.; Mavromoustakos, T.; Tzeli, D. Computational and Spectroscopic Analysis of the Quercetin Encapsulation in (2HP-β-CD)2 and (2,6Me-β-CD)2 Complexes. *J. Mol. Struct.* 2023, 1294, 136430. [CrossRef]
- Vakali, V.; Papadourakis, M.; Georgiou, N.; Zoupanou, N.; Diamantis, D.A.; Javornik, U.; Papakyriakopoulou, P.; Plavec, J.; Valsami, G.; Tzakos, A.G.; et al. Comparative Interaction Studies of Quercetin with 2-Hydroxyl-Propyl-β-Cyclodextrin and 2,6-Methylated-β-Cyclodextrin. *Molecules* 2022, 27, 5490. [CrossRef] [PubMed]
- 93. Manta, K.; Papakyriakopoulou, P.; Nikolidaki, A.; Balafas, E.; Kostomitsopoulos, N.; Banella, S.; Colombo, G.; Valsami, G. Comparative Serum and Brain Pharmacokinetics of Quercetin after Oral and Nasal Administration to Rats as Lyophilized Complexes with β-Cyclodextrin Derivatives and Their Blends with Mannitol/Lecithin Microparticles. *Pharmaceutics* 2023, 15, 2036. [CrossRef] [PubMed]
- Savic, I.M.; Nikolic, V.D.; Savic-Gajic, I.; Nikolic, L.B.; Radovanovic, B.C.; Mladenovic, J.D. Investigation of Properties and Structural Characterization of the Quercetin Inclusion Complex with (2-Hydroxypropyl)-β-Cyclodextrin. *J. Incl. Phenom. Macrocycl. Chem.* 2015, *82*, 383–394. [CrossRef]
- 95. Manta, K.; Papakyriakopoulou, P.; Chountoulesi, M.; Diamantis, D.A.; Spaneas, D.; Vakali, V.; Naziris, N.; Chatziathanasiadou, M.V.; Andreadelis, I.; Moschovou, K.; et al. Preparation and Biophysical Characterization of Quercetin Inclusion Complexes with β-Cyclodextrin Derivatives to Be Formulated as Possible Nose-to-Brain Quercetin Delivery Systems. *Mol. Pharm.* 2020, 17, 4241–4255. [CrossRef]
- 96. Palli, V.; Leonis, G.; Zoupanou, N.; Georgiou, N.; Chountoulesi, M.; Naziris, N.; Tzeli, D.; Demetzos, C.; Valsami, G.; Marousis, K.D.; et al. Losartan Interactions with 2-Hydroxypropyl-β-CD. *Molecules* 2022, 27, 2421. [CrossRef]
- Kfoury, M.; Landy, D.; Ruellan, S.; Auezova, L.; Greige-Gerges, H.; Fourmentin, S. Determination of Formation Constants and Structural Characterization of Cyclodextrin Inclusion Complexes with Two Phenolic Isomers: Carvacrol and Thymol. *Beilstein J. Org. Chem.* 2016, 12, 29–42. [CrossRef]
- Caira, M.R.; Bourne, S.A.; Samsodien, H.; Smith, V.J. Inclusion Complexes of 2-Methoxyestradiol with Dimethylated and Permethylated β-Cyclodextrins: Models for Cyclodextrin-Steroid Interaction. *Beilstein J. Org. Chem.* 2015, *11*, 2616–2630. [CrossRef]
- Haimhoffer, Á.; Rusznyák, Á.; Réti-Nagy, K.; Vasvári, G.; Váradi, J.; Vecsernyés, M.; Bácskay, I.; Fehér, P.; Ujhelyi, Z.; Fenyvesi, F. Cyclodextrins in Drug Delivery Systems and Their Effects on Biological Barriers. *Sci. Pharm.* 2019, 87, 33. [CrossRef]
- 100. Tiwari, G.; Tiwari, R.; Rai, A. Cyclodextrins in Delivery Systems: Applications. J. Pharm. Bioallied Sci. 2010, 2, 72. [CrossRef] [PubMed]
- Wüpper, S.; Lüersen, K.; Rimbach, G. Cyclodextrins, Natural Compounds, and Plant Bioactives—A Nutritional Perspective. Biomolecules 2021, 11, 401. [CrossRef] [PubMed]
- Chen, W.; Chang, C.E.; Gilson, M.K. Calculation of Cyclodextrin Binding Affinities: Energy, Entropy, and Implications for Drug Design. *Biophys. J.* 2004, 87, 3035–3049. [CrossRef] [PubMed]
- 103. Liossi, A.S.; Ntountaniotis, D.; Kellici, T.F.; Chatziathanasiadou, M.V.; Megariotis, G.; Mania, M.; Becker-Baldus, J.; Kriechbaum, M.; Krajnc, A.; Christodoulou, E.; et al. Exploring the Interactions of Irbesartan and Irbesartan–2-Hydroxypropyl-β-Cyclodextrin Complex with Model Membranes. *Biochim. Biophys. Acta-Biomembr.* 2017, 1859, 1089–1098. [CrossRef]
- 104. Diamantis, D.A.; Ramesova, S.; Chatzigiannis, C.M.; Degano, I.; Gerogianni, P.S.; Karadima, K.E.; Perikleous, S.; Rekkas, D.; Gerothanassis, I.P.; Galaris, D.; et al. Exploring the Oxidation and Iron Binding Profile of a Cyclodextrin Encapsulated Quercetin Complex Unveiled a Controlled Complex Dissociation through a Chemical Stimulus. *Biochim. Biophys. Acta-Gen. Subj.* 2018, 1862, 1913–1924. [CrossRef]
- Castro, E.; Barbiric, D. Molecular Modeling and Cyclodextrins: A Relationship Strengthened By Complexes. *Curr. Org. Chem.* 2006, 10, 715–729. [CrossRef]
- 106. Kellici, T.F.; Chatziathanasiadou, M.V.; Diamantis, D.; Chatzikonstantinou, A.V.; Andreadelis, I.; Christodoulou, E.; Valsami, G.; Mavromoustakos, T.; Tzakos, A.G. Mapping the Interactions and Bioactivity of Quercetin(2-Hydroxypropyl)-β-Cyclodextrin Complex. *Int. J. Pharm.* 2016, 511, 303–311. [CrossRef]
- 107. Kellici, T.F.; Chatziathanasiadou, M.V.; Lee, M.S.; Sayyad, N.; Geromichalou, E.G.; Vrettos, E.I.; Tsiailanis, A.D.; Chi, S.W.; Geromichalos, G.D.; Mavromoustakos, T.; et al. Rational Design and Structure-Activity Relationship Studies of Quercetin-Amino Acid Hybrids Targeting the Anti-Apoptotic Protein Bcl-XL. Org. Biomol. Chem. 2017, 15, 7956–7976. [CrossRef]
- 108. Debnath, K.; Jana, N.R.; Jana, N.R. Quercetin Encapsulated Polymer Nanoparticle for Inhibiting Intracellular Polyglutamine Aggregation. *ACS Appl. Bio Mater.* **2019**, *2*, 5298–5305. [CrossRef]
- Jullian, C.; Moyano, L.; Yañez, C.; Olea-Azar, C. Complexation of Quercetin with Three Kinds of Cyclodextrins: An Antioxidant Study. Spectrochim. Acta-Part A Mol. Biomol. Spectrosc. 2007, 67, 230–234. [CrossRef]
- Liu, M.; Dong, L.; Chen, A.; Zheng, Y.; Sun, D.; Wang, X.; Wang, B. Inclusion Complexes of Quercetin with Three β-Cyclodextrins Derivatives at Physiological PH: Spectroscopic Study and Antioxidant Activity. *Spectrochim. Acta-Part A Mol. Biomol. Spectrosc.* 2013, 115, 854–860. [CrossRef] [PubMed]

- 111. Sangpheak, W.; Kicuntod, J.; Schuster, R.; Rungrotmongkol, T.; Wolschann, P.; Kungwan, N.; Viernstein, H.; Mueller, M.; Pongsawasdi, P. Physical Properties and Biological Activities of Hesperetin and Naringenin in Complex with Methylated P-Cyclodextrin. *Beilstein J. Org. Chem.* 2015, *11*, 2763–2773. [CrossRef] [PubMed]
- 112. Kicuntod, J.; Khuntawee, W.; Wolschann, P.; Pongsawasdi, P.; Chavasiri, W.; Kungwan, N.; Rungrotmongkol, T. Inclusion Complexation of Pinostrobin with Various Cyclodextrin Derivatives. J. Mol. Graph. Model. 2016, 63, 91–98. [CrossRef] [PubMed]
- 113. Kumari, A.; Yadav, S.K.; Pakade, Y.B.; Singh, B.; Yadav, S.C. Development of Biodegradable Nanoparticles for Delivery of Quercetin. *Colloids Surf. B Biointerfaces* **2010**, *80*, 184–192. [CrossRef] [PubMed]
- 114. Zwicker, J.I.; Furie, B.; Flaumenhaft, R. Method for Treating Sickle Cell Disease Using Quercetin-Containing Compositions. WO 2023/288044 A1, 2023.
- 115. Renjit, S. Sickle Cell Anemia Treatment. US 2006/0115459 A1, 26 November 2005.
- 116. Trimboli, D.; Gatti, V.; Naccari, G.C. Combination of Catechin and Quercetin for Pharmaceutical or Dietary Use. WO 02/34262 A1, 11 October 2001.
- 117. Zhang, J.; Li, B. New Application of Quercetin and Kaempferol. CN113018293A, 10 March 2021.
- 118. Antipruritic Composition Containing Astragalin and Quercetin. KR20120121684A, 27 April 2011.
- 119. Lines, T. Reducing Cholesterol Levels with Combined Use of Quercetin and Statin. WO2010027572A2, 23 July 2009.
- 120. Won, L.K.; Joo, L.H.; Jun, L.S.; Young, C.J.; Ju, K.N.; Hoon, K.J.; Hyuk, L.J. Composition for Inhibiting Liver Cancer Containing Doxorubicin and Quercetin. KR100553266B1, 20 February 2006.
- 121. Marchi, D.; Feige, J.; Horcajada, M. Compositions and Methods Using a Combination of Oleuropein and Quercetin for Use in Cartilage Degeneration. WO2022106410A1, 11 November 2021.
- 122. Brown, D. Treatment of Fragile X Syndrome with Ibudilast in Combination with Metformin, Cannbidiol, Sertraline or Quercetin. WO2021044158A1, 4 September 2020.
- 123. Lines, T. Quercetin-Containing Compositions for Use in Treating Amyotrophic Lateral Sclerosis. WO2022243942A1, 19 May 2022.
- 124. Song, H.B.; Tae, S.J.; Ki, H.B.; Yong, B.P.; Myung, S.C.; Sik, M.S.; Yong, K.K.; Lee, E.S.; Byung, H.H.; Yang, K.C.; et al. Composition Containing Rutin and Quercetin for Preventing or Treating Elevated Blood Lipid Level-Related Diseases. WO0015237A1, 15 September 1999.
- 125. Muller, W.; Ernst, L.G.; Schroder, H.-C.; Wilhelm, F.; Wang, X. Synergistic Composition Comprising Quercetin and Polyphosphate for Treatment of Bone Disorders. WO2015132304A1, 4 March 2015.
- 126. Sang-Chan, K.I.M.; Park, S.M.; Kim, J.K.; Kim, E.O.; PARK, C.A.; Sung-Hui, B.Y.U.N.; Sook-Jahr, P.A.R.K. Composition for Preventing or Treating Liver Disease, Comprising Icaritin and Quercetin. WO2022065550A1, 25 September 2020.
- 127. Jeung-Hye, P. Composition Containing Quercetin and Vitamin D for Alleviation of Acnegenic Skin. WO2018230824A1, 9 April 2018.
- 128. Burger, A.R.; Granger, S.P.; Scott, I.R. Skin Care Compositions Containing Naringenin and/or Quercetin and a Retinoid. US5665367A, 27 September 1996.
- Jeung-Hye, P. Composition, Containing Quercetin, Genistein, and Alpha-Lipoic Acid, for Relieving Acne Skin. WO2020111757A1, 27 November 2019.
- 130. Vasquez Garzon, V.R.; Carrasco Torres, G.; Andrade Jorge, E.; Trujillo Ferrara, J.G.; Trevino Villa, S. Quercetin and Maleic Anhydride Derivatives for The Treatment of Hepatocellular Carcinoma. MX2018008239A, 3 July 2018.
- 131. Lopez Munoz, F.J.; Espinosa Juarez, J.V.; Jaramilo Morales, O.A. Pharmaceutical Composition of Haloperidol and Quercetin with Analgesic Effect In Neuropathic Pain. MX2017007166A, 5 June 2017.
- 132. Mi-La, C.; Min-Jung, P.; Seon-Yeong, L.; Sung-Hee, L.; Eun-Ji, Y.; Hye-Jin, S. Composition for Preventing or Treating Immune Disease Comprising Metformin and Quercetin as Active Ingredients. KR20140132932A, 9 May 2013.
- 133. Gwonhwa, S.; Whasun, L.; Sunwoo, P. Pharmaceutical Composition for Preventing or Treating Endometriosis Comprising Quercetin Luteolin Delphinidin or Mixture Thereof. KR20210044409A, 15 October 2019.
- 134. Kuebler, U. Medicament, Useful to Treat or Prevent Malignantly Transformed Cells, e.g., Adeno-Carcinoma, Prostate Carcinoma and Breast Carcinoma, Comprises a Mixture of Quercetin and Myrecetin and/or Anisomycin and Rapamycin as Kinase Inhibitors. DE102006036307A1, 3 August 2006.
- 135. Polifenoles, C, Combinación Sinérgica de Polifenoles. WO2012150370A1, 2 May 2012.
- 136. Nday, C.M.; Halevas, E.; Jackson, G.E.; Salifoglou, A. Quercetin Encapsulation in Modified Silica Nanoparticles: Potential Use against Cu(II)-Induced Oxidative Stress in Neurodegeneration. *J. Inorg. Biochem.* **2015**, *145*, 51–64. [CrossRef] [PubMed]
- 137. Zhao, Y.; Yang, M.; Li, Y.; Luan, X.; Luo, Z. Quercetin Derivatives and Their Medical Usages. US2004132671A1, 26 September 2003.
- Ferreira-Silva, M.; Faria-Silva, C.; Carvalheiro, M.C.; Simões, S.; Marinho, H.S.; Marcelino, P.; Campos, M.C.; Metselaar, J.M.; Fernandes, E.; Baptista, P.V.; et al. Quercetin Liposomal Nanoformulation for Ischemia and Reperfusion Injury Treatment. *Pharmaceutics* 2022, 14, 104. [CrossRef] [PubMed]
- 139. Antonio Tamayo-Ramos, J.; Martel, S.; Barros, R.; Bol, A.; Atilhan, M.; Aparicio, S. On the Behavior of Quercetin + Organic Solvent Solutions and Their Role for C60 Fullerene Solubilization. *J. Mol. Liq.* **2022**, *345*, 117714. [CrossRef]
- de la Torre, M.D.L.; Tomé, A.C.; Silva, A.M.S.; Cavaleiro, J.A.S. Synthesis of [60]Fullerene–Quercetin Dyads. *Tetrahedron Lett.* 2002, 43, 4617–4620. [CrossRef]

- 141. Das, S.; Saha, M.; Mahata, L.C.; China, A.; Chatterjee, N.; Das Saha, K. Quercetin and 5-Fu Loaded Chitosan Nanoparticles Trigger Cell-Cycle Arrest and Induce Apoptosis in HCT116 Cells via Modulation of the P53/P21 Axis. ACS Omega 2023, 8, 36893–36905. [CrossRef]
- 142. Mohammed, M.A.; Syeda, J.T.M.; Wasan, K.M.; Wasan, E.K. An Overview of Chitosan Nanoparticles and Its Application in Non-Parenteral Drug Delivery. *Pharmaceutics* **2017**, *9*, 53. [CrossRef]
- 143. Valencia-Lazcano, A.A.; Hassan, D.; Pourmadadi, M.; Shamsabadipour, A.; Behzadmehr, R.; Rahdar, A.; Medina, D.I.; Díez-Pascual, A.M. 5-Fluorouracil Nano-Delivery Systems as a Cutting-Edge for Cancer Therapy. *Eur. J. Med. Chem.* 2023, 246, 114995. [CrossRef] [PubMed]
- 144. Moon, Y.J.; Wang, X.; Morris, M.E. Dietary Flavonoids: Effects on Xenobiotic and Carcinogen Metabolism. *Toxicol. Vitr.* 2006, 20, 187–210. [CrossRef] [PubMed]
- 145. Li, R.; Liu, B.; Gao, J. The Application of Nanoparticles in Diagnosis and Theranostics of Gastric Cancer. *Cancer Lett.* **2017**, *386*, 123–130. [CrossRef] [PubMed]
- Pugazhendhi, A.; Edison, T.N.J.I.; Karuppusamy, I.; Kathirvel, B. Inorganic Nanoparticles: A Potential Cancer Therapy for Human Welfare. *Int. J. Pharm.* 2018, 539, 104–111. [CrossRef] [PubMed]
- Khan, A.; Khan, A.A.P.; Asiri, A.M.; Rub, M.A.; Rahman, M.M.; Ghani, S.A. In Vitro Studies of Carbon Fiber Microbiosensor for Dopamine Neurotransmitter Supported by Copper-Graphene Oxide Composite. *Microchim. Acta* 2014, 181, 1049–1057. [CrossRef]
- 148. Kratz, J.M.; Teixeira, M.R.; Ferronato, K.; Teixeira, H.F.; Koester, L.S.; Simões, C.M.O. Preparation, Characterization, and In Vitro Intestinal Permeability Evaluation of Thalidomide–Hydroxypropyl-β-Cyclodextrin Complexes. AAPS PharmSciTech 2012, 13, 118–124. [CrossRef]
- Khan, A.; Khan, A.A.P.; Asiri, A.M. Toward Design and Measurement of Electrical Conductivity and Thermal Properties of Silver Nanoparticle Embedded Poly(o-anisidine) Molybdophosphate Nanocomposite and Its Application as Microbiosensor. *Polym. Compos.* 2017, 38, E237–E245. [CrossRef]
- Khan, A.; Khan, A.A.P.; Asiri, A.M.; Rub, M.A.; Azum, N.; Rahman, M.M.; Khan, S.B.; Ghani, S.A. A New Trend on Biosensor for Neurotransmitter Choline/Acetylcholine—An Overview. *Appl. Biochem. Biotechnol.* 2013, 169, 1927–1939. [CrossRef]
- 151. Li, M.; Zhang, F.; Su, Y.; Zhou, J.; Wang, W. Nanoparticles Designed to Regulate Tumor Microenvironment for Cancer Therapy. *Life Sci.* 2018, 201, 37–44. [CrossRef]
- 152. Baskar, G.; Garrick, B.G.; Lalitha, K.; Chamundeeswari, M. Gold Nanoparticle Mediated Delivery of Fungal Asparaginase against Cancer Cells. J. Drug Deliv. Sci. Technol. 2018, 44, 498–504. [CrossRef]
- 153. Alban, L.; Monteiro, W.F.; Diz, F.M.; Miranda, G.M.; Scheid, C.M.; Zotti, E.R.; Morrone, F.B.; Ligabue, R. New Quercetin-Coated Titanate Nanotubes and Their Radiosensitization Effect on Human Bladder Cancer. *Mater. Sci. Eng. C* 2020, 110, 110662. [CrossRef] [PubMed]
- 154. Down, C.J.; Nair, R.; Thurairaja, R. Bladder Cancer. Surgery 2016, 34, 532–539. [CrossRef]
- 155. Yilmaz, M.; Karanastasis, A.A.; Chatziathanasiadou, M.V.; Oguz, M.; Kougioumtzi, A.; Clemente, N.; Kellici, T.F.; Zafeiropoulos, N.E.; Avgeropoulos, A.; Mavromoustakos, T.; et al. Inclusion of Quercetin in Gold Nanoparticles Decorated with Supramolecular Hosts Amplifies Its Tumor Targeting Properties. ACS Appl. Bio Mater. 2019, 2, 2715–2725. [CrossRef] [PubMed]
- 156. Mansour, F.R.; Abdallah, I.A.; Bedair, A.; Hamed, M. Analytical Methods for the Determination of Quercetin and Quercetin Glycosides in Pharmaceuticals and Biological Samples. *Crit. Rev. Anal. Chem.* **2023**, 1–26. [CrossRef]
- 157. Ahmedova, A.; Paradowska, K.; Wawer, I. 1H, 13C MAS NMR and DFT GIAO Study of Quercetin and Its Complex with Al(III) in Solid State. *J. Inorg. Biochem.* 2012, *110*, 27–35. [CrossRef] [PubMed]
- 158. Mehranfar, F.; Bordbar, A.-K.; Parastar, H. A Combined Spectroscopic, Molecular Docking and Molecular Dynamic Simulation Study on the Interaction of Quercetin with β-Casein Nanoparticles. *J. Photochem. Photobiol. B Biol.* **2013**, *127*, 100–107. [CrossRef]
- Nazir, N.; Karim, N.; Abdel-Halim, H.; Khan, I.; Wadood, S.F.; Nisar, M. Phytochemical Analysis, Molecular Docking and Antiamnesic Effects of Methanolic Extract of *Silybum marianum* (L.) Gaertn Seeds in Scopolamine Induced Memory Impairment in Mice. J. Ethnopharmacol. 2018, 210, 198–208. [CrossRef]
- 160. Malkhasian, A.Y.S.; Howlin, B.J. Docking and DFT Studies on Ligand Binding to Quercetin 2,3-Dioxygenase. *J. Biomol. Struct. Dyn.* **2016**, *34*, 2453–2461. [CrossRef]
- 161. Xiao, Z.-P.; Wang, X.-D.; Peng, Z.-Y.; Huang, S.; Yang, P.; Li, Q.-S.; Zhou, L.-H.; Hu, X.-J.; Wu, L.-J.; Zhou, Y.; et al. Molecular Docking, Kinetics Study, and Structure–Activity Relationship Analysis of Quercetin and Its Analogous as Helicobacter Pylori Urease Inhibitors. J. Agric. Food Chem. 2012, 60, 10572–10577. [CrossRef]
- 162. Singh, S.P.; Selvaraj, C.; Knowar, B.K.; Singh, S.K.; Singh, C.B.; Sahoo, D. Competitive Inhibition of Quercetin and Apigenin at the ATP Binding Site of D-Alanine-D-Alanine Ligase of Helicobacter Pylori—A Molecular Modeling Approach. *Curr. Biotechnol.* 2019, 7, 340–348. [CrossRef]
- 163. Ghosh, A.; Sarmah, P.; Patel, H.; Mukerjee, N.; Mishra, R.; Alkahtani, S.; Varma, R.S.; Baishya, D. Nonlinear Molecular Dynamics of Quercetin in Gynocardia Odorata and Diospyros Malabarica Fruits: Its Mechanistic Role in Hepatoprotection. *PLoS ONE* 2022, 17, e0263917. [CrossRef] [PubMed]
- 164. Kikiowo, B.; Ahmad, I.; Alade, A.A.; Ijatuyi, T.T.; Iwaloye, O.; Patel, H.M. Molecular Dynamics Simulation and Pharmacokinetics Studies of Ombuin and Quercetin against Human Pancreatic α-Amylase. J. Biomol. Struct. Dyn. 2023, 41, 10388–10395. [CrossRef] [PubMed]

- 165. Maran, M.; Gangadharan, S.; Emerson, I.A. Molecular Dynamics Study of Quercetin Families and Its Derivative Compounds from Carica Papaya Leaf as Breast Cancer Inhibitors. *Chem. Phys. Lett.* **2022**, *793*, 139470. [CrossRef]
- 166. Omirin, E.S.; Omotuyi, O.; Afokhume, O.G.; Okoh, E.F.; Boboye, S.O.; Olugbogi, E.A.; Adelegan, O.O.; Ibitoye, B.O.; Aderiye, M.A.; Agosile, O.O. Molecular Dynamics Simulations on Quercetin-3-(6-Malonylglucoside) From Morus Alba Shows Optimal Inhibition of Bcl-2 with Favorable Anti-Tumor Activities. *bioRxiv* 2015, 1–15. [CrossRef]
- Anusuya, S.; Gromiha, M.M. Quercetin Derivatives as Non-Nucleoside Inhibitors for Dengue Polymerase: Molecular Docking, Molecular Dynamics Simulation, and Binding Free Energy Calculation. J. Biomol. Struct. Dyn. 2017, 35, 2895–2909. [CrossRef] [PubMed]
- 168. Lokhande, K.B.; Ballav, S.; Yadav, R.S.; Swamy, K.V.; Basu, S. Probing Intermolecular Interactions and Binding Stability of Kaempferol, Quercetin and Resveratrol Derivatives with PPAR-γ: Docking, Molecular Dynamics and MM/GBSA Approach to Reveal Potent PPAR-γ Agonist against Cancer. J. Biomol. Struct. Dyn. 2022, 40, 971–981. [CrossRef] [PubMed]
- Li, X.-Y.; Li, Y.-C.; Yu, D.-G.; Liao, Y.-Z.; Wang, X. Fast Disintegrating Quercetin-Loaded Drug Delivery Systems Fabricated Using Coaxial Electrospinning. Int. J. Mol. Sci. 2013, 14, 21647–21659. [CrossRef]
- Sahoo, N.G.; Kakran, M.; Shaal, L.A.; Li, L.; Müller, R.H.; Pal, M.; Tan, L.P. Preparation and Characterization of Quercetin Nanocrystals. J. Pharm. Sci. 2011, 100, 2379–2390. [CrossRef]
- 171. Zhang, L.; Yang, X.; Li, S.; Gao, W. Preparation, Physicochemical Characterization and in Vitro Digestibility on Solid Complex of Maize Starches with Quercetin. *LWT-Food Sci. Technol.* **2011**, *44*, 787–792. [CrossRef]
- 172. Koontz, J.L.; Marcy, J.E.; O'Keefe, S.F.; Duncan, S.E. Cyclodextrin Inclusion Complex Formation and Solid-State Characterization of the Natural Antioxidants α-Tocopherol and Quercetin. *J. Agric. Food Chem.* **2009**, *57*, 1162–1171. [CrossRef]
- 173. Sengupta, B.; Sengupta, P.K. The Interaction of Quercetin with Human Serum Albumin: A Fluorescence Spectroscopic Study. *Biochem. Biophys. Res. Commun.* 2002, 299, 400–403. [CrossRef] [PubMed]
- Gutierrez, A.C.; Gehlen, M.H. Time Resolved Fluorescence Spectroscopy of Quercetin and Morin Complexes with Al3+. Spectrochim. Acta Part A Mol. Biomol. Spectrosc. 2002, 58, 83–89. [CrossRef] [PubMed]
- 175. Poór, M.; Boda, G.; Kunsági-Máté, S.; Needs, P.W.; Kroon, P.A.; Lemli, B. Fluorescence Spectroscopic Evaluation of the Interactions of Quercetin, Isorhamnetin, and Quercetin-3'-Sulfate with Different Albumins. J. Lumin. 2018, 194, 156–163. [CrossRef]
- 176. Alabri, Z.K.; Hussain, J.; Mabood, F.; Rehman, N.U.; Ali, L.; Al-Harrasi, A.; Hamaed, A.; Khan, A.L.; Rizvi, T.S.; Jabeen, F.; et al. Fluorescence Spectroscopy-Partial Least Square Regression Method for the Quantification of Quercetin in Euphorbia Masirahensis. *Measurement* 2018, 121, 355–359. [CrossRef]
- 177. Yang, X.; Wu, D.; Du, Z.; Li, R.; Chen, X.; Li, X. Spectroscopy Study on the Interaction of Quercetin with Collagen. J. Agric. Food Chem. 2009, 57, 3431–3435. [CrossRef] [PubMed]
- 178. Wybranowski, T.; Kruszewski, S. Optical Spectroscopy Study of the Interaction between Quercetin and Human Serum Albumin. *Acta Phys. Pol. A* 2014, 125, 57–60. [CrossRef]
- 179. Pham-Hoang, B.; Winckler, P.; Waché, Y. Fluorescence Lifetime and UV-Vis Spectroscopy to Evaluate the Interactions between Quercetin and Its Yeast Microcapsule. *Biotechnol. J.* 2018, *13*, 1700389. [CrossRef]
- Hussain, J.; Rehman, N.U.; Mabood, F.; Al-Harrasi, A.; Ali, L.; Rizvi, T.S.; Khan, A.; Rafiq, K.; Al-Rabaani, H.; Jabeen, F. Application of Fluorescence Spectroscopy Coupled with PLSR for the Estimation of Quercetin in Four Medicinal Plants. *Chem. Data Collect.* 2019, 21, 100228. [CrossRef]
- Verma, V.; Sharma, P.; Sharma, J.; Kaur Lamba, A.; Lamba, H.S. Development, Characterization and Solubility Study of Solid Dispersion of Quercetin by Solvent Evaporation Method. *Mater. Today Proc.* 2017, *4*, 10128–10133. [CrossRef]
- 182. Verma, N.; Trehan, N. HPLC Analysis of Methanolic Extract of Herbs for Quercetin Content. J. Pharmacogn. Phytochem. 2013, 2, 159–162.
- 183. D'Mello, P.M.; Joshi, U.J.; Shetgiri, P.P.; Dasgupta, T.K.; Darji, K.K. A Simple HPLC Method for Quantitation of Quercetin in Herbal Extracts. *J. AOAC Int.* 2011, *94*, 100–105. [CrossRef] [PubMed]
- Rahimi, M.; Bahar, S.; Heydari, R.; Amininasab, S.M. Determination of Quercetin Using a Molecularly Imprinted Polymer as Solid-Phase Microextraction Sorbent and High-Performance Liquid Chromatography. *Microchem. J.* 2019, 148, 433–441. [CrossRef]
- Stefova, M.; Kulevanova, S.; Stafilov, T. Assay of Flavonols and Quantification of Quercetin in Medicinal Plants by Hplc with Uv-Diode Array Detection. J. Liq. Chromatogr. Relat. Technol. 2001, 24, 2283–2292. [CrossRef]
- 186. Olszewska, M. Separation of Quercetin, Sexangularetin, Kaempferol and Isorhamnetin for Simultaneous HPLC Determination of Flavonoid Aglycones in Inflorescences, Leaves and Fruits of Three Sorbus Species. J. Pharm. Biomed. Anal. 2008, 48, 629–635. [CrossRef] [PubMed]
- Canini, A.; Alesiani, D.; D'Arcangelo, G.; Tagliatesta, P. Gas Chromatography–Mass Spectrometry Analysis of Phenolic Compounds from *Carica papaya* L. Leaf. J. Food Compos. Anal. 2007, 20, 584–590. [CrossRef]
- 188. Deng, F.; Zito, S.W. Development and Validation of a Gas Chromatographic–Mass Spectrometric Method for Simultaneous Identification and Quantification of Marker Compounds Including Bilobalide, Ginkgolides and Flavonoids in *Ginkgo biloba* L. Extract and Pharmaceutical Preparatio. J. Chromatogr. A 2003, 986, 121–127. [CrossRef]
- 189. Gross, M.; Pfeiffer, M.; Martini, M.; Campbell, D.; Slavin, J.; Potter, J. The Quantitation of Metabolites of Quercetin Flavonols in Human Urine. *Cancer Epidemiol. Biomark. Prev.* **1996**, *5*, 711–720.
- 190. Pejic, N.; Kuntic, V.; Vujic, Z.; Micic, S. Direct Spectrophotometric Determination of Quercetin in the Presence of Ascorbic Acid. *Il Farm.* 2004, 59, 21–24. [CrossRef]

- Belščak-Cvitanović, A.; Valinger, D.; Benković, M.; Tušek, A.J.; Jurina, T.; Komes, D.; Gajdoš Kljusurić, J. Integrated Approach for Bioactive Quality Evaluation of Medicinal Plant Extracts Using HPLC-DAD, Spectrophotometric, near Infrared Spectroscopy and Chemometric Techniques. Int. J. Food Prop. 2017, 20, S2463–S2480. [CrossRef]
- 192. Kurzawa, M. Determination of Quercetin and Rutin in Selected Herbs and Pharmaceutical Preparations. *Anal. Lett.* **2010**, *43*, 993–1002. [CrossRef]
- 193. Ligor, M.; Kornyšova, O.; Maruška, A.; Buszewski, B. Determination of Flavonoids in Tea and Rooibos Extracts by TLC and HPLC. J. Planar Chromatogr.—Mod. TLC 2008, 21, 355–360. [CrossRef]
- 194. Randhawa, K.; Kumar, D.; Jamwal, A.; Kumar, S. Screening of Antidepressant Activity and Estimation of Quercetin from Coccinia Indica Using TLC Densitometry. *Pharm. Biol.* **2015**, *53*, 1867–1874. [CrossRef] [PubMed]
- 195. Reed, G.A. Stability of Drugs, Drug Candidates, and Metabolites in Blood and Plasma. *Curr. Protoc. Pharmacol.* 2016, 2016, 7.6.1–7.6.12. [CrossRef] [PubMed]
- Patel, A.A.; Amin, A.A.; Patwari, A.H.; Shah, M.B. Validated High Performance Thin Layer Chromatography Method for Simultaneous Determination of Quercetin and Gallic Acid in Leea Indica. *Rev. Bras. Farmacogn.* 2017, 27, 50–53. [CrossRef]
- 197. Chen, G.; Zhang, H.; Ye, J. Determination of Rutin and Quercetin in Plants by Capillary Electrophoresis with Electrochemical Detection. *Anal. Chim. Acta* 2000, 423, 69–76. [CrossRef]
- 198. Li, X.; Zhang, Y.; Yuan, Z. Separation and Determination of Rutin and Quercetin in the Flowers of *Sophora japonica* L. by Capillary Electrophoresis with Electrochemical Detection. *Chromatographia* **2002**, *55*, 243–246. [CrossRef]
- 199. Suntornsuk, L.; Kasemsook, S.; Wongyai, S. Quantitative Analysis of Aglycone Quercetin in Mulberry Leaves (*Morus alba* L.) by Capillary Zone Electrophoresis. *Electrophoresis* **2003**, *24*, 1236–1241. [CrossRef]
- 200. Wang, J.; Wang, H.; Han, S. Ultrasensitive Determination of Epicatechin, Rutin, and Quercetin by Capillary Electrophoresis Chemiluminescence. *Acta Chromatogr.* 2012, 24, 679–688. [CrossRef]
- Memon, A.F.; Solangi, A.R.; Memon, S.Q.; Mallah, A.; Memon, N.; Memon, A.A. Simultaneous Determination of Quercetin, Rutin, Naringin, and Naringenin in Different Fruits by Capillary Zone Electrophoresis. *Food Anal. Methods* 2017, 10, 83–91. [CrossRef]
- Zielinska, D.; Wiczkowski, W.; Piskula, M.K. Determination of the Relative Contribution of Quercetin and Its Glucosides to the Antioxidant Capacity of Onion by Cyclic Voltammetry and Spectrophotometric Methods. J. Agric. Food Chem. 2008, 56, 3524–3531. [CrossRef]
- Reddaiah, K.; Reddy, T.M.; Swamy, K. Electrochemical Determination of Quercetin at β–Cyclodextrin Modified Chemical Sensor: A Voltammetric Study. *Anal. Bioanal. Electrochem.* 2012, 4, 122.
- Guss, E.V.; Ziyatdinova, G.K.; Zhupanova, A.S.; Budnikov, H.C. Voltammetric Determination of Quercetin and Rutin on Their Simultaneous Presence on an Electrode Modified with Polythymolphthalein. J. Anal. Chem. 2020, 75, 526–535. [CrossRef]
- Korotkova, E.I.; Voronova, O.A.; Dorozhko, E.V. Study of Antioxidant Properties of Flavonoids by Voltammetry. J. Solid State Electrochem. 2012, 16, 2435–2440. [CrossRef]
- 206. Numata, Y.; Tanaka, H. Quantitative Analysis of Quercetin Using Raman Spectroscopy. Food Chem. 2011, 126, 751–755. [CrossRef]
- Jurasekova, Z.; Domingo, C.; Garcia-Ramos, J.V.; Sanchez-Cortes, S. Effect of PH on the Chemical Modification of Quercetin and Structurally Related Flavonoids Characterized by Optical (UV-Visible and Raman) Spectroscopy. *Phys. Chem. Chem. Phys.* 2014, 16, 12802–12811. [CrossRef] [PubMed]
- 208. Paczkowska, M.; Lewandowska, K.; Bednarski, W.; Mizera, M.; Podborska, A.; Krause, A.; Cielecka-Piontek, J. Application of Spectroscopic Methods for Identification (FT-IR, Raman Spectroscopy) and Determination (UV, EPR) of Quercetin-3-O-Rutinoside. Experimental and DFT Based Approach. Spectrochim. Acta Part A Mol. Biomol. Spectrosc. 2015, 140, 132–139. [CrossRef]
- Jurasekova, Z.; Torreggiani, A.; Tamba, M.; Sanchez-Cortes, S.; Garcia-Ramos, J.V. Raman and Surface-Enhanced Raman Scattering (SERS) Investigation of the Quercetin Interaction with Metals: Evidence of Structural Changing Processes in Aqueous Solution and on Metal Nanoparticles. J. Mol. Struct. 2009, 918, 129–137. [CrossRef]
- Cornard, J.P.; Merlin, J.C.; Boudet, A.C.; Vrielynck, L. Structural Study of Quercetin by Vibrational and Electronic Spectroscopies Combined with Semiempirical Calculations. *Biospectroscopy* 1997, *3*, 183–193. [CrossRef]
- Savic, I.M.; Nikolic, V.D.; Savic, I.M.; Nikolic, L.B.; Stankovic, M.Z. Development and Validation of a New RP-HPLC Method for Determination of Quercetin in Green Tea. J. Anal. Chem. 2013, 68, 906–911. [CrossRef]
- 212. Sah, M.K.; Gautam, B.; Pokhrel, K.P.; Ghani, L.; Bhattarai, A. Quantification of the Quercetin Nanoemulsion Technique Using Various Parameters. *Molecules* 2023, *28*, 2540. [CrossRef]
- 213. Dinesh Kumar, V.; Verma, P.R.P.; Singh, S.K. Morphological and in vitro Antibacterial Efficacy of Quercetin Loaded Nanoparticles against Food-Borne Microorganisms. *LWT-Food Sci. Technol.* **2016**, *66*, 638–650. [CrossRef]
- Mosleh, M.; Ghoreishi, S.M.; Masoum, S.; Khoobi, A. Determination of Quercetin in the Presence of Tannic Acid in Soft Drinks Based on Carbon Nanotubes Modified Electrode Using Chemometric Approaches. *Sens. Actuators B Chem.* 2018, 272, 605–611. [CrossRef]
- Savic-Gajic, I.M.; Savic, I.M.; Nikolic, V.D. Modelling and Optimization of Quercetin Extraction and Biological Activity of Quercetin-Rich Red Onion Skin Extract from Southeastern Serbia. J. Food Nutr. Res. 2018, 57, 15–26.
- Huang, J.; Wang, Q.; Li, T.; Xia, N.; Xia, Q. Nanostructured Lipid Carrier (NLC) as a Strategy for Encapsulation of Quercetin and Linseed Oil: Preparation and in Vitro Characterization Studies. J. Food Eng. 2017, 215, 1–12. [CrossRef]

- Vacek, J.; Papoušková, B.; Vrba, J.; Zatloukalová, M.; Křen, V.; Ulrichová, J. LC–MS Metabolic Study on Quercetin and Taxifolin Galloyl Esters Using Human Hepatocytes as Toxicity and Biotransformation in vitro Cell Model. *J. Pharm. Biomed. Anal.* 2013, 86, 135–142. [CrossRef] [PubMed]
- 218. Kokalj Ladan, M.; Straus, J.; Tavčar Benković, E.; Kreft, S. FT-IR-Based Method for Rutin, Quercetin and Quercitrin Quantification in Different Buckwheat (Fagopyrum) Species. *Sci. Rep.* **2017**, *7*, 7226. [CrossRef] [PubMed]
- Pralhad, T.; Rajendrakumar, K. Study of Freeze-Dried Quercetin–Cyclodextrin Binary Systems by DSC, FT-IR, X-ray Diffraction and SEM Analysis. J. Pharm. Biomed. Anal. 2004, 34, 333–339. [CrossRef]
- Prasongsidh, B.C.; Skurray, G.R. Capillary Electrophoresis Analysis of Trans- and Cis-Resveratrol, Quercetin, Catechin and Gallic Acid in Wine. *Food Chem.* 1998, 62, 355–358. [CrossRef]
- 221. Zhang, J.; Li, H.; Wang, W.; Li, H. Assessing the Anti-inflammatory Effects of Quercetin Using Network Pharmacology and in Vitro Experiments. *Exp. Ther. Med.* **2022**, *23*, 301. [CrossRef]
- Zhang, X.-W.; Chen, J.-Y.; Ouyang, D.; Lu, J.-H. Quercetin in Animal Models of Alzheimer's Disease: A Systematic Review of Preclinical Studies. Int. J. Mol. Sci. 2020, 21, 493. [CrossRef]
- 223. Nickel, T.; Hanssen, H.; Sisic, Z.; Pfeiler, S.; Summo, C.; Schmauss, D.; Hoster, E.; Weis, M. Immunoregulatory Effects of the Flavonol Quercetin in vitro and in vivo. *Eur. J. Nutr.* **2011**, *50*, 163–172. [CrossRef] [PubMed]
- 224. Ekinci, M.S. Supercritical Fluid Extraction of Quercetin from Sumac (*Rhus coriaria* L.): Effects of Supercritical Extraction Parameters. *Sep. Sci. Technol.* 2022, 57, 256–262. [CrossRef]
- 225. Huang, Y.; Feng, Y.; Tang, G.; Li, M.; Zhang, T.; Fillet, M.; Crommen, J.; Jiang, Z. Development and Validation of a Fast SFC Method for the Analysis of Flavonoids in Plant Extracts. *J. Pharm. Biomed. Anal.* **2017**, *140*, 384–391. [CrossRef] [PubMed]
- 226. Magnuszewska, J.; Krogulec, T. Analytica Chimica Acta Application of Hot Platinum Microelectrodes for Determination of Flavonoids in Flow Injection Analysis and Capillary Electrophoresis. *Anal. Chim. Acta* **2013**, *786*, 39–46. [CrossRef] [PubMed]
- 227. Saldanha, L.; Vilegas, W.; Dokkedal, A. Characterization of Flavonoids and Phenolic Acids in Myrcia Bella Cambess. Using FIA-ESI-IT-MSn and HPLC-PAD-ESI-IT-MS Combined with NMR. *Molecules* 2013, 18, 8402–8416. [CrossRef] [PubMed]
- Kumar, A.; Malik, A.K.; Tewary, D.K. A New Method for Determination of Myricetin and Quercetin Using Solid Phase Microextraction–High Performance Liquid Chromatography–Ultra Violet/Visible System in Grapes, Vegetables and Red Wine Samples. Anal. Chim. Acta 2009, 631, 177–181. [CrossRef]
- Cecchi, L.; Ieri, F.; Vignolini, P.; Mulinacci, N.; Romani, A. Characterization of Volatile and Flavonoid Composition of Different Cuts of Dried Onion (*Allium cepa* L.) by HS-SPME-GC-MS, HS-SPME-GC×GC-TOF and HPLC-DAD. *Molecules* 2020, 25, 408. [CrossRef]
- 230. York, N.; May, R. The Crystal and Molecular Structure of Quercetin: A Biologically Active and Naturally Occurring Flavonoid. *Bioorg. Chem.* **1986**, *69*, 55–69.
- 231. Han, G.; Zeng, Q.; Jiang, Z.; Xing, T.; Huang, C. Talanta MIL-101 (Cr) as Matrix for Sensitive Detection of Quercetin by Matrix-Assisted Laser Desorption/Ionization Mass Spectrometry. *Talanta* **2017**, *164*, 355–361. [CrossRef]
- 232. Wang, J.; Sporns, P. MALDI-TOF MS Analysis of Food Flavonol Glycosides. J. Agric. Food Chem. 2000, 48, 1657–1662. [CrossRef]
- Frison-Norrie, S.; Sporns, P. Identification and Quantification of Flavonol Glycosides in Almond Seedcoats Using MALDI-TOF MS. J. Agric. Food Chem. 2002, 50, 2782–2787. [CrossRef] [PubMed]
- Kroslakova, I.; Pedrussio, S.; Wolfram, E. Direct Coupling of HPTLC with MALDI-TOF MS for Qualitative Detection of Flavonoids on Phytochemical Fingerprints. *Phytochem. Anal.* 2016, 27, 222–228. [CrossRef] [PubMed]
- 235. Stojković, E.D.; Zdravkovski, Z. Supercritical Fluid Extraction of Quercetin and Rutin from Hyperici Herba Supercritical Fluid Extraction of Quercetin and Rutin from Hyperici Herba. J. Liq. Chromatogr. Relat. Technol. 2007, 26, 2517–2533. [CrossRef]
- Hamed, M.; Abdallah, I.A.; Bedair, A.; Mansour, F.R. Sample Preparation Methods for Determination of Quercetin and Quercetin Glycosides in Diverse Matrices. *Microchem. J.* 2023, 194, 109233. [CrossRef]
- 237. Molinelli, A.; Weiss, R.; Mizaikoff, B. Advanced Solid Phase Extraction Using Molecularly Imprinted Polymers for the Determination of Quercetin in Red Wine. *J. Agric. Food Chem.* 2002, *50*, 1804–1808. [CrossRef]
- 238. Song, X.; Li, J.; Wang, J.; Chen, L. Quercetin Molecularly Imprinted Polymers: Preparation, Recognition Characteristics and Properties as Sorbent for Solid-Phase Extraction. *Talanta* **2009**, *80*, 694–702. [CrossRef]
- 239. Braga, L.R.; Rosa, A.A.; Dias, A.C.B. Synthesis and Characterization of Molecularly Imprinted Silica Mediated by Al for Solid Phase Extraction of Quercetin in *Ginkgo biloba* L. *Anal. Methods* **2014**, *6*, 4029–4037. [CrossRef]
- Asfaram, A.; Arabi, M.; Ostovan, A.; Sadeghi, H.; Ghaedi, M. Simple and Selective Detection of Quercetin in Extracts of Plants and Food Samples by Dispersive-Micro-Solid Phase Extraction Based on Core–Shell Magnetic Molecularly Imprinted Polymers. *New J. Chem.* 2018, 42, 16144–16153. [CrossRef]

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