



Review

Progress on the Extraction, Separation, Biological Activity, and Delivery of Natural Plant Pigments

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Abstract: Natural plant pigments are safe and have low toxicity, with various nutrients and biological activities. However, the extraction, preservation, and application of pigments are limited due to the instability of natural pigments. Therefore, it is necessary to examine the extraction and application processes of natural plant pigments in detail. This review discusses the classification, extraction methods, biological activities, and modification methods that could improve the stability of various pigments from plants, providing a reference for applying natural plant pigments in the industry and the cosmetics, food, and pharmaceutical industries.

Keywords: plant pigment; chemical classification; extraction method; biological activity; modification method

1. Introduction

The pigment is widely used as an additive in the food, beverage, and cosmetic industries, primarily due to consumers' pursuit of color. Pigments can be divided into synthetic and natural pigments. William discovered lavender, the first synthetic organic dye, in 1556. Afterward, numerous synthetic pigments were widely used in food, medicine, and cosmetics; however, many toxic substances, such as lead, arsenic, and mercury, have been detected in these synthetic pigments. Since then, the United States has promulgated the Food and Drug Act, which forbids poisonous synthetic pigments as food additives [1]. Natural pigments are safe and non-toxic, and recent research has demonstrated that natural pigments have different biological activities. Natural pigments can be derived from animals, plants, and microorganisms, with plant pigments being the most studied and utilized. Currently, FDA approved synthetic pigments in the United States include red moss red, fast green, bright blue, and sunset yellow; 32 kinds of natural pigments are allowed, such as chlorophyll, beet red, riboflavin, and β -carotene; six kinds of synthetic pigments are allowed in China, and 54 kinds of natural pigments are allowed, indicating that natural pigments continue to be the primary source of pigments. However, natural pigments are less stable than synthetic pigments; therefore, the pigment industry is implementing new technologies to ensure the stability of natural pigments during extraction and application [2].

Pigments extracted from plants are insoluble in water and unstable in light, heat, and at extreme pH levels [3]. Traditional natural pigment extraction techniques, such as solvent extraction, grinding, and pressing, these methods has long time, low yield, need numerous solvents, and some organic solvents are highly toxic [4]. Some green and efficient extraction methods, including (1) supercritical fluid extraction, (2) ultrasound-assisted extraction, (3) enzyme-assisted extraction, (4) microwave-assisted extraction, and (5) ultra high pressure-assisted extraction, have been developed to solve the above problems.



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Modern studies have confirmed that natural pigments have different biological activities, such as antioxidation, anti-inflammation, anti-cancer, neuro-protection, and cardiovascular protection [5]. Recently, a series of methods have been developed to improve the stability of natural pigments, including the synthesis of lipid-biological macromolecules [6], pigment–protein complexes [7], and pigment-metal nanoparticle synthesis [8].

This review focuses on the chemical structure of natural pigments used in the food and cosmetics industries and the research advancements and biological activity of natural pigment extraction technology.

2. Chemical Classification of Natural Pigments

According to the color classification, natural pigments can be primarily divided into six categories: red, orange, yellow, green, blue, and purple (Figure 1). According to the chemical structure classification, natural pigments can be primarily divided into five categories: carotenoids, polyphenols, quinones, pyrrole, and pyridines (Table 1).

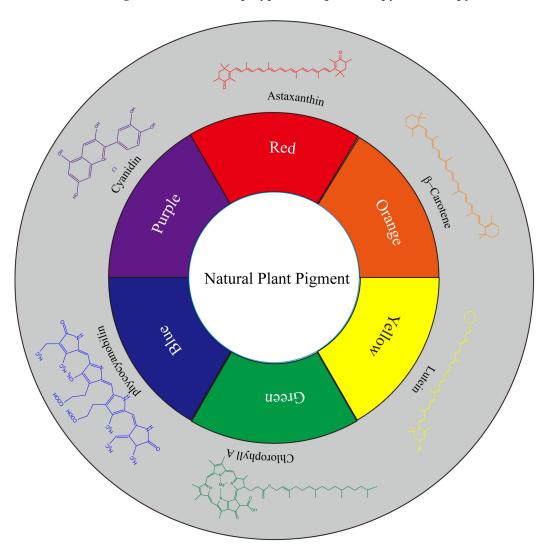


Figure 1. Representative natural plant pigments of different colors: Astaxanthin, β -carrot, Lutein, Chlorophyll A, Phycocyanobilin, and Cyanidin.

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Table 1. Chemical classification, chemical structure, and biological activity of natural plant pigments.

Classification	Natural Plant Pigment	Chemical Constitution	Bioactivity	Reference
			Anti-inflammation	[9]
	β-carotene		Anti-oxidation	[10]
		X	Anti-cancer	[11]
	Lycopene		Anti-oxidation	[12]
		<i></i>	Anti-inflammation	[13]
		\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	Anti-tumor	[14]
		Ţ	Anti-atherosclerosis	[15]
			Anti-inflammation	[16]
	Astaxanthin	Ů,	Neuro protection	[17]
Carotenoids		X-y-y-d-d-X	Anti-fibrosis	[18]
		7	Anti-cancer	[19]
			Anti-oxidation	[20]
			Cardiovascular protection	[21]
		All	Anti-oxidation	[22]
	Lutein		Neuro protection	[23]
			Anti-cancer	[24]
		. Он	Anti-inflammation	[25]
		X	Anti-oxidation	[26]
	Zeaxanthin		Anti-cancer	[27]
			Neuro protection	[28]
	Pelargonidin	ОН	Anti-obesity	[29]
		он	Anti-cancer	[30]
		HO	Anti-oxidation	[31]
		CI	Anti-inflammation	
				[32]
		OH	Anti-oxidation	[33]
	Cyanidin	OH OH	Bone protection	[34]
		HO	Anti-inflammation	[35]
		СГ	Anti-cancer	[36]
	Anthocyanins Delphindin	qн	Anti-cancer	[37]
		OH	Anti-inflammation	[38]
Polyphenols		но	Anti-oxidation	[39]
		CT OH	Neuro protection	[40]
		~~~	Bone protection	[41]
	Peonidin	10° (C) (O) (O) (O) (O) (O) (O) (O) (O) (O) (O	Anti-inflammation	[42]
	Petunidin		Anti-inflammation	[43]
		OH OH	Bone protection	[44]
			Cardiovascular protection	[45]
		CT OH	Cardiovascular protection	[40]
		l он	Anti-cancer	[46]
	Malvidin			
		ОН	Anti-inflammation	[47]
			Cardiovascular protection	[48]
		СГ	Anti-oxidation	[49]
		,	Anti-inflammation	[50]
			7 Hu Hilaminaton	[SO]
	Curcumin	NO.	Anti-cancer	[51]
			Neuro protection	[52]
		₩ ₩	Anti-bacterial	[53]
Quinone		он о он	Anti-cancer	[54]
			Anti-fibrosis	[55]
	Emodin	ОН	Anti-inflammation	[56]
		Ö	Neuro protection	[57]
	Alizarin	O II	Anti-coagulation	[58]
			Anti-bacterial	[59]
	Alizarin	<b>Г</b>		
		₿ Ын	Anti-cancer	[60]

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Table 1. Cont.

Classification	Natural Plant Pigment	Chemical Constitution	Bioactivity	Reference
	Chlorophyll A	Alfumm A	Antigen toxicity Anti-inflammation Anti-oxidation	[61] [62] [63]
Pyrrole	Chlorophyll B	A Thromas	Antigen toxicity Anti-oxidation	[61] [63]
	Phycocyanobilin		Anti-inflammation Anti-oxidation Neuro protection	[64] [65] [66]
	Phycoerythrobilin	ON HIND COOK	Anti-oxidation	[67]
Pyridines	Betacyanin	150 COO	Anti-hypersensitivity Anti-inflammation Anti-thrombotic	[68] [69] [70]
	Betaxanthin		Anti-oxidation Anti-tumor Anti-oxidation	[71] [72] [73]

#### 2.1. Carotenoid

Carotenoids, or isoprene derivatives, are conjugated polyene chains that are composed of eight isoprenes [74]. Carotenoids are abundant in animals, higher plants, fungi, and algae. The carotenoid database established by Yabuzaki et al. contains the chemical information of 1117 kinds of natural carrots and 683 kinds of organisms [75]. Carotenoids are primarily yellow, orange, or red. Carotenoids can be divided into two categories based on the different elements they contain: carotene has only carbon and hydrogen atoms, mainly  $\beta$ -carotene and lycopene. One is lutein containing carbon, hydrogen, oxygen, and other factors, primarily astaxanthin, lutein, and zeaxanthin [76]. Carotenoids have various biological activities, such as anti-oxidation, anti-cancer, and immune regulation. Some carotenoids can be used as the premise of vitamin A and have seen great potential for treating vitamin A deficiency [77]. The most important way for humans to intake carotenoids is through fruits and vegetables. The  $\beta$ -carotene mainly exists in green leafy vegetables and yellow or orange fruits or vegetables, such as broccoli, carrots, and mangoes. Lycopene is abundant in tomatoes, watermelons, grapes, and other fruits, but it does not belong to vitamin A and cannot be changed into vitamin A in the body [78].

## 2.2. Polyphenols

Polyphenols are also called plant tannins and can be divided primarily into flavonoids and non-flavonoids. Flavonoids mainly include anthocyanins, isoflavones, and flavonoids: the primary 2-phenylchromone structure as the skeleton. Most flavonoids are fat soluble compounds in water, whereas anthocyanins are water soluble. Non-flavonoids include stilbene compounds, phenolic acids, and lignans; the glycosylation state of polyphenols and its diversity has a significant impact on the immunomodulatory activity, the sugar moiety, type, position, and extent of glycosylation in determining the anti-oxidant, anti-inflammatory, and the immunomodulatory activities of polyphenols. [79]. Polyphenols are prone to denaturation and oxidation due to the influence of their chemical structure and the external environment. Modern studies have confirmed that hydroxylation reduces flavonoid stability, while glycosylation, methylation, and acylation improve flavonoid stability. Moreover, the external environment, including pH, temperature, light, and oxygen, may change polyphenol properties [80]. For example, anthocyanins extracted from grapes appear red when pH < 7, blue when pH = 7, and blue or light blue when pH > 10 [81]. Therefore, the effects of the external environment on polyphenols during the

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extraction should be considered. Polyphenols have various biological activities, including anti-allergic, anti-oxidation, anti-bacterial, and anti-inflammation [82]. Natural polyphenols are abundant in grapes, blueberries, mangoes, and other fruits and vegetables, such as pearl vegetables, Chinese toon buds, coriander, and other vegetables [83]; these fruits and vegetables are the primary sources of polyphenols for humans.

#### 2.3. Quinones

Quinnes share a cyclohexadienedione or cyclohexadiene dimethylene structure and are classified as quinones, naphthoquinones, and anthraquinones. Quinones are primarily derived from traditional Chinese medicines, such as the rhubarb of Polygonaceae, Polygonum multiflorum, Polygonum cuspidatum, Rubiaceae, Leguminosae cassia seeds, Senna, and Liliaceae aloe [84]. Emodin and alizarin are the most studied quinones in modern times. Emodin has an orange long needle like crystal with the chemical name 1-3-3-8-trihydroxy-6-methyl anthraquinone. Modern studies have confirmed that emodin has biological activities, such as hepato protection, anti-oxidation, and anti-bacterial [85], but it also has certain hepatotoxicity, nephrotoxicity, and reproductive toxicity. Emodin is a molecule that is fat soluble and water insoluble; thus, emodin-nicotinamide eutectic [86], emodin loaded nanoparticles [87], and emodin-PLGA film [88] can be developed to improve emodin bioavailability in vivo. Alizarin is an orange crystal or ochre yellow powder with the chemical name 1,2-dihydroxyanthraquinone. It has anticoagulants [58] and anti-cancer activities [89].

#### 2.4. Pyrrole

Pyrrole compounds are abundant in plants and animals. The most widely studied natural pyrrole pigments include plant derived chlorophyll, phycobilin, animal-derived heme, and bilirubin. The common characteristic of the above four pigments is that they are all connected by four pyrrole rings. Chlorophyll can be divided into Chlorophyll a, Chlorophyll b, Chlorophyll c, Chlorophyll d, Chlorophyll f, prochlorophyll, and bacterial chlorophyll. It has a core porphyrin ring and a leaf alcohol chain, with a magnesium atom in the middle of the porphyrin ring [90]. Chlorophyll absorbs blue and red light from sunlight and reflects scatters of green light, making the plant's leaves green. Chlorophyll is unstable in the external environment; a low pH, heat, light, oxygen, and enzyme reactions can also change the green properties. Therefore, the effects of the aforementioned conditions should be considered during the extraction and separation processes. Chlorophyll and its derivatives have anti-oxidation, anti-cancer, and anti-teratogenicity properties [91]. Phycobilin is a pigment in red algae and cyanobacteria that can be divided into phycocyanonbilin, phycoerythrobilin, and phycoviolobilin. Its common structure is linear tetrapyrrole, which differs from chlorophyll in that the leaf alcohol chain lacks a magnesium atom. Different phycobilins can exhibit red and blue colors due to differences in light absorption. The physical and chemical properties of phycobilin are easy to change under light conditions and an extreme pH. Phycobilin has anti-oxidant, anti-inflammatory, neuro-protective, and other biological activities [92].

# 2.5. Pyridines

Betaine is a water soluble pigment that contains a pyridine ring in *Caryophyllum*, including betaine and betaxanthin. Betaine is a reddish purple to dark purple: a pigment that causes plants to turn orange from yellow. Betaine has been widely used in food processing due to its good water solubility. More than 50 kinds of betaine and more than 30 kinds of betaxanthin have been identified [93]. Betaine is unstable at high temperatures, an extreme pH, with metal ions and light. Betaine has various biological activities, including anti-bacterial, anti-cancer, decrease blood lipid, liver protection, and neuroprotection [94].

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## 3. Extraction Methods of Natural Plant Pigments

Traditional pigment extraction methods include solvent extraction, pressing, and impregnation. However, these methods have the problem of consuming many organic solvents, taking a long time, and having low yields. Therefore, some new pigment extraction methods have been developed to improve the above problems.

# 3.1. Supercritical Fluid Extraction

Supercritical fluid extraction is a technology that uses a supercritical fluid as an extractant to separate a component (extract) from its mixture (matrix). Most supercritical solvents are hydrocarbons, aromatics, alcohols, and some gases. CO₂ is the most mainstream supercritical fluid because it is safe, lacks solvent residue after extraction, is cheap and easy to obtain, and can separate heat-sensitive compounds [95]. Co-solvent, pressure, and time effects on the extraction process should be considered in CO₂ supercritical fluid extraction. Abrahamsson et al. [96] extracted carotenoids and chlorophyll A from microalgae at a temperature of  $40\sim60$  °C, a pressure of  $15\sim30$  MPa, with a liquid CO₂ flow rate of 1–4 g/min, and co-solvent ethanol 0~0.2 mL/min. The extracted compounds were detected using a UV-vis spectrophotometer and modeled using the extractable amount and ethanol mole fraction. The carotenoid yield was 0.25 mg/g, the chlorophyll A yield was 0.96 mg/g, and the extraction amount depended on the co-solvent amount. DaPorto et al. [97] extracted polyphenols from white grape seeds at 40 °C, different pressures, CO₂ fringes, and co-solvent ratios. The optimal extraction conditions were as follows: the highest total polyphenol concentration was obtained at 80 bar pressure, 6 kg/h of the CO₂ flow rate, and 20% (w/w) of the co-solvent. Combined with previous research, increasing pressure can increase the yield, but not significantly.

# 3.2. Ultrasound-Assisted Extraction

The purpose of ultrasonic-assisted extraction technology is to make the effective components of the extracted material quickly enter into the solvent under the action of an ultrasonic wave to obtain the multi-component mixed extract and then separate, refine, and purify the extract using appropriate methods. This is a new technology to obtain the required monomer chemical components. Ultrasonic waves can produce comprehensive effects, such as cavitation, vibration, crushing, and mixing, in the medium. Ultrasound can break the cell wall and accelerate the extract dissolution. Ultrasound-assisted extraction has often been used to extract thermally unstable active substances. The primary advantages are shortening the extraction time, improving extraction efficiency, a wide application, and simple operation [98]. Sahin et al. [99] used the ultrasound-assisted extraction of lutein and carotenoids from Chlorella, added them to chitosan to prepare chitosan films, and characterized them using Fourier transform infrared spectroscopy and SEM technology. The results display how 4.844  $\pm$  0.78 mg/g lutein and 0.533  $\pm$  0.06 mg/g carotenoids can be prepared using ultrasonic extraction. The pigment-chitosan complex has antiantioxidant activity and can be used as a wound-healing plaster. Zhong et al. [100] used the ultrasound-assisted extraction of diosgenin; the results presented that when the acetone volume fraction was 74%, the extraction time was 31 min and the temperature was 54  $^{\circ}$ C, while the dioscorea pigment yield was the highest (32.27%). The extracted dioscorea pigment exhibited good anti-oxidant activity.

## 3.3. Microwave-Assisted Extraction

Microwave-assisted extraction mainly depends on the heating effect of microwaves to obtain a higher temperature and mass transfer rate. The greater the solvent polarity, the faster the microwave energy absorption and heating up, accelerating the extraction rate. The primary advantages are speed and efficiency, uniform heating, solvent saving, and selectivity. The solvent used for the extraction, microwave time, stirring, and temperature influenced the microwave extraction. Long term microwave heating can change the extract's properties [101]. Sharma et al. [102] used the microwave-assisted extraction method to ex-

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tract betaine and betaaxanthin from tricolor amaranth leaves and characterized them using Fourier transform infrared spectroscopy. The highest betacyanin yield was 71.95 mg/g when the microwave power was 450 W, and the temperature was 90 °C for 15 min, while the beet flavin yield was the highest (42.30 mg/g) when the microwave power was 200 W, and the temperature was 60 °C. Fernandez-Aulis et al. compared the three methods of impregnation, ultrasonic extraction, and microwave-assisted extraction from corn. The results exhibited that the anthocyanin yield in the corn husk was  $21.89 \pm 1.23$  mg/g after 30 min,  $25.80 \pm 0.59$  mg/g using the ultrasonic method after 20 min, and  $24.47 \pm 0.85$  mg/g using the microwave-assisted method after 1 min. The microwave-assisted method greatly shortened the extraction time [103].

## 3.4. Enzyme-Assisted Extraction

The enzyme hydrolyzes the plant cell wall and destroys the cell matrix as a biological reaction catalyst, promoting the extraction material into the solvent, accelerating the extraction rate, and increasing the yield. The advantages of enzyme-assisted extraction included a short extraction time, strong specificity, mild reaction conditions, and high yield. Enzyme selection was the most important aspect of enzyme-assisted extraction. Enzymes, such as cellulase, hemicellulase, ligninase, pectinase, and amylase, could be utilized for enzyme-assisted extraction. One enzyme could be used for extraction, or a mixture of multiple enzymes could be used for the extraction [104]. Zhao et al. [105] investigated the optimal conditions for extracting astaxanthin from Haematococcus pluvialis using cellulase and Pectinase. The astaxanthin yield was 67.15% at 45 °C, pH 5.0, and 1.0% cellulase for 6 h. When the content of Pectinase was 0.08%, the astaxanthin yield was 75.30% at 55 °C, pH 4.5. Lombardelli et al. [106] extracted betaine from beetroots using an enzyme blend of cellulase, xylanase, and pectinase. After optimizing the conditions, such as the amount of mixed enzyme, the treatment time, and temperature, the optimum conditions for extracting betaine with mixed enzyme were determined: an enzyme amount of 25 U/g, a temperature of 25  $^{\circ}$ C, and a treatment time of 240 min.

## 3.5. Ultra High Pressure Assisted Extraction

The ultrahigh pressure extraction technology uses high pressure to penetrate the solvent into the raw material, allowing the target compound in the cell to dissolve in the solvent. After a period under high pressure, the target compound reached a dissolution equilibrium in the solvent. Afterward, the pressure returned to normal, and the target compound spread to the extract around the tissue. Simultaneously, ultrahigh pressure destroyed the tissues, promoting target compound release [107]. Nunes et al. [108] studied the extraction of the beet pigment from cactus by high pressure CO₂-acidified water extraction using the response surface optimization method. The results indicate a maximum yield of  $89 \pm 0.7$  mg/100 g of beet pigment at 100 bar, 40 °C, and a 20% solid–liquid mixture/pressurized CO₂. Ferrari et al. [109] compared anthocyanin and flavonoid extraction from new pigment rice Nerone Gold 26/6 using high pressure extraction, ultrasound-assisted extraction, and conventional solid–liquid extraction. High pressure extraction yielded the highest yield of the anthocyanins, and both methods produced higher yields than conventional solid–liquid extraction.

#### 3.6. Joint Application

Although the above extraction methods have advantages, they also have limitations, such as high cost and low equipment utilization, therefore, several studies used the above extraction methods in series to overcome these limitations. Shahram et al. [110] combined ultrasound-assisted and enzyme-assisted extraction and used the response surface optimization method to extract  $\beta$ -carotene from a citrus processing residue. The study determined that the  $\beta$ -carotene yield and the anti-antioxidant activity were the highest at a Pectinase concentration of 0.4%, an ultrasound at 115.5 min, and a pH of 5.11. Fu

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et al. [111] established a microwave-enzyme-assisted aqueous two phase extraction method to extract the total polyphenols and lutein from marigolds. The total polyphenols yield was 84.61 mg/g, the corresponding recovery rate was 95.35, the lutein yield was 7.32 mg/g, and the corresponding recovery rate was 99.85%. The yield was significantly increased compared to Soxhlet extraction, microwave-assisted extraction, and enzyme-assisted extraction. Therefore, pigment extraction cannot be limited to a single technique, and various techniques can be utilized.

# 4. Biological Activity of Natural Plant Pigments

# 4.1. Anti-Oxidation

Reactive oxygen species and reactive nitrogen play an important role in signal transduction and the maintenance of balance in human cells at medium/low concentrations. However, they also lead to oxidative stress at high concentrations, resulting in damage to the cellular structure, including lipids, membranes, proteins, DNA, cardiovascular diseases, cancer, nervous system disorders, diabetes, and stroke [112]. Modern studies have confirmed that natural plant pigments can scavenge reactive oxygen species and increase superoxide dismutase activity through Keap1/Nrf2, AMPK/Nrf2, and other signaling pathways to exert anti-antioxidant effects. Josson Akkara et al. [113] used  $\beta$ -carotene to treat liver injury induced by bromobenzene and compared its therapeutic effect with the standard hepatoprotective drug silymarin. The results indicated that β-carotene could reduce oxidative stress and bromobenzene induced liver injury by reducing peroxide and inflammatory cytokine levels while increasing oxidant levels. This therapeutic effect is similar to that of silymarin. Wang et al. [114] used anthocyanin in a carbon tetrachlorideinduced liver injury model in mice to study the anti-antioxidant effect of anthocyanin and its mechanism. The results exhibited how anthocyanins could reduce liver injury in mice by reducing malondialdehyde (MDA), interleukin-6 (IL-6), and interleukin-1 β (IL-1 β). It may be that in this mechanism, anthocyanins exert anti-antioxidant and anti-inflammatory effects via the Keap1/Nrf2 pathway. Yu et al. [115] confirmed that cyanidin-3-glucoside could scavenge reactive oxygen species via the AMPK/Nrf2 pathway, enhance the antioxidation defense ability of mice, weaken H₂O₂-induced apoptosis, and prevent liver injury. Brotosudarmo pointed out in their study that the hydroxyl and ketone groups in the astaxanthin structure can play a role in neutralizing reactive oxygen species, quenching harmful singlet oxygen, reducing the formation of free radicals, and thus exerting anti-antioxidant effects [116].

# 4.2. Anti-Inflammation

Inflammation is caused by tissue damage or pathogen activation by the innate immune system after the injury or infection of the human body, inducing leukocyte synthesis and removing inflamed sites and tissue regeneration. Inflammation is usually temporary; however, when it becomes chronic, it can lead to numerous diseases. Modern studies have confirmed that inflammation is associated with many diseases, such as asthma, cancer, and atherosclerosis [117]. Natural plant pigments can delay the inflammation process by reducing the expression of inflammatory factors such as tumor necrosis factor- $\alpha$ (TNF- $\alpha$ ) and interleukin. Takahashi et al. [118] investigated the anti-inflammatory effect of β-carotene in atopic dermatitis induced by a low zinc/magnesium diet. Studies have confirmed that β-carotene can reduce skin inflammation by inhibiting inflammatory factors such as IL-1  $\beta$ , reducing matrix metalloproteinases activity, and promoting fibroin expression. Blas-Valdivia et al. [119] studied the cardio-protection effect of C-phycocyanin and its mechanism using isoproterenol induced acute myocardial infarction in rats. The results displayed that C-phycocyanin could reduce cytokines in the contents, such as TNF-α, reactive oxygen species, IL-β, interferon-γ, myocardial enzymes, and creatine kinase, and play the role of anti-inflammation and anti-oxidation. Li et al. [120] studied the benefits of curcumin on traumatic brain injury and its mechanism. The results revealed that curcumin

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has an anti-inflammatory effect, inhibiting IL-1  $\beta$ , IL-6, TNF- $\alpha$ , and p38 expressions, thus alleviating traumatic brain injury.

## 4.3. Anti-Cancer

Current cancer treatments include chemotherapy and other induced irreversible metabolic damage to kill cancer cells; however, chemotherapy still has serious side effects. Therefore, it is necessary to identify new ways to treat cancer. Modern studies have confirmed that natural plant pigments can induce the apoptosis of cancer cells through signaling pathways such as P13K/AKT and ERK1/2MAPK, thereby exerting anti-cancer effects [121]. Cui et al. [122] investigated the anti-cancer effect of astaxanthin and established an N-nitroso-benzylamine induced rat esophageal cancer model. The results indicated that astaxanthin cereal significantly reduced the incidence of esophageal cancer by increasing glutathione peroxidase and superoxide dismutase activities and reducing NF-kB and COX2 protein expressions. Yeh et al. [123] confirmed that naphthoquinone shikonin extracted from shikonin could induce apoptosis and reduce human alveolar basal epithelial cells A549 proliferation in a dose dependent manner via a p53 mediated signal pathway. Lim et al. [124] studied the preventive effect of delphinin in anthocyanins on epithelial ovarian cancer. The results demonstrated that vermicellin induced the apoptosis of ovarian cancer cells by fragmenting DNA at the SKOV3 point and inhibited SKOV3 cell proliferation via two signaling pathways: P13K/AKT and ERK1/2MAPK line.

#### 4.4. Neuro Protection

The incidence of nervous system diseases is relatively high. Stroke, epilepsy, senile dementia, and other common nervous system diseases affect the health of tens of millions of people worldwide. Coupled with rare diseases, such as Wright syndrome, and Rasmussen encephalitis, it has become an increasingly severe medical burden and world problem [125]. Studies have confirmed that curcumin could inhibit the Wnt signaling pathway to inhibit apoptosis and the inflammation of neurons when exposed to hypoxia/oxygen reperfusion and play a neuroprotective role [126]. Jiang et al. [127] revealed that curcumin can inhibit nitric oxide synthase and nitrite/nitrate production, prevent the injury of cerebral capillary endothelial cells, reduce the infarct volume, and improve neurological function, confirming the protective effect of curcumin on cerebral ischemia/reperfusion injury. Gazzin et al. [128] examined the neuro-protection effect of curcumin in the spontaneous model of neonatal hyperbilirubinemia. The results illustrated that curcumin mediated multiple targets of neuro injury (inflammation, redox imbalance, and glutamate neurotoxicity), allowing the cerebellum to develop completely and recover abnormal rat behavior. Wang et al. [129] confirmed that shikonin could decrease the neurological deficit score, reduce the infarct size and MDA content, and increase superoxide dismutase and catalase contents in shikonin-treated mice, thus playing a neuroprotective role. Phycocyanin and its tetrapyrrole chromophore phycocyanin from spirulina improved multiple sclerosis by improving neuroinflammation and preventing demyelination and axonal loss [130]. Phycocyanin also has great potential in treating Alzheimer's disease [131].

# 4.5. Cardiovascular Protection

Cardiovascular disease is a circulatory system disease that affects the heart, arteries, and veins and is commonly related to arteriosclerosis. Coronary heart disease, hypertension, angina pectoris, and myocardial infarction are the major diseases. Recently, many natural products have been developed to treat vascular diseases [132]. Zhou et al. [133] studied the mechanism of the cardiovascular protective effect of curcumin. The results exhibited that curcumin could down regulate lncRNAH19 expression and inhibit Wnt/ $\beta$ -catenin expression, thus inhibiting vascular restenosis induced by intimal hyperplasia and protecting the cardiovascular system. Qian et al. [134] demonstrated that anthocyanins could alleviate cisplatin induced heart injury by inhibiting apoptosis when mediated by reactive oxygen species and the extracellular regulated kinase signal pathway. Thrombosis is partly

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attributed to the activation of endothelial tissue factors, leading to vascular thrombosis. Lee and others [135] have confirmed that carotenoids can inhibit the activity of tissue factors in endothelial cells by promoting protein kinase B (Akt) phosphorylation. Ferreira Santos et al. [136] studied the therapeutic effect of lycopene on angiotensin II induced hypertensive rats. The results revealed that lycopene could improve hypertension and cardiovascular remodeling caused by angiotensin II; however, it did not affect blood pressure in normal rats, confirming the therapeutic potential of lycopene in hypertension.

## 5. Modification of Natural Plant Pigments

Natural phytochromes have limited applications due to their instability. Lipid carriers, protein nanoparticles, chitosan nanoparticles, and metal ions have been used to modify and stabilize natural phytochromes; these methods can improve the stability of natural plant pigments, thereby increasing their application range. Table 2 lists the materials used in various methods and their advantages.

<b>Modification Method</b>	Type/Material	Merit	Reference	
Lipid carrier	Solid lipid nanoparticles	High biocompatibility; enhanced membrane permeability; enhanced solubility; biodegradability; increased intestinal	[137]	
	Nanostructured lipid carriers	drug dilution		
Ductain managertials	Soy isolate protein	Rich sources, biocompatibility, biodegradability; the synthesis	s [138]	
Protein nanoparticle	Albumin	process is simple		
Chitosan nanoparticles	Deacetylated chitin	Biodegradability, biocompatibility, bioactivity,	[139]	
Cintosari nanoparticies		non-toxicity, polycation		
	Au			
26.1	Ag	High biocompatibility, high biodegradability, high	[140]	
Metal	ZnO	anti-bacterial activity, targeted delivery, controlled release	[140]	
	Cu and oxides			

**Table 2.** Modification methods of natural plant pigments.

#### 5.1. Lipid Nanoparticles

Lipid-based colloidal carriers can improve the fat soluble natural compounds' solubility in water, bioavailability in vivo, and easily degradable compound stability. Therefore, they are widely used in the carrier research of natural compounds. Lipid nanoparticles can be divided into solid lipid nanoparticles with the solid lipid as the lipid core and nanostructured lipid carriers with a liquid lipid as the lipid core [141]. Osanlou et al. [142] prepared zeaxanthin solid lipid nanoparticles and nanostructured lipid carriers by combining high shear homogenization and an ultrasound. According to studies, zeaxanthin loaded lipid nanoparticles are spherical; zeaxanthin is wrapped in the lipid material but has no chemical interaction with the material. Tamjidi et al. [143] displayed that the nanostructured lipid carriers loaded with astaxanthin had good stability in simulated gastric juice and could increase astaxanthin's bioavailability. Pelissari et al. [144] used spray cooling technology to prepare solid lipid particles loaded with lycopene/sunflower oil and shortening as a carrier. Studies have revealed that combining Arabic gum and the carrier could reduce lycopene degradation in low temperature and vacuum environments [145]; prepared nanostructured lipid carriers loaded with curcumin and studied the anti-cancer effect of the nanosystems. The results indicate that these nanoparticles could achieve the sustained release of curcumin and have a more sustained anti-cancer effect than free curcumin.

#### 5.2. Protein Nanoparticles

Proteins have been widely used to study drug carriers due to their good biocompatibility and degradability. The proteins used in the delivery system include albumin, whey protein, and bean globulin. These proteins have a good loading capacity and sustained release [146]. Liu et al. [147] exposed that sodium caseinate, whey protein isolate, and soybean protein isolate could successfully load annatto and increase its stability and water solubility. In simulated cheese production, soybean protein isolate nanoparticles loaded with annatto could inhibit pigment penetration into whey, providing a theoretical basis

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for their application in food. Radomirovic et al. [148] have studied phycocyanin- $\beta$  lactoglobulin nanoparticle stability. Studies have revealed that  $\beta$ -lactoglobulin loaded with phycocyanin can reduce the trend of heat induced polymerization and the fibril formation of proteins, presenting higher stability. Gulsu et al. [149] demonstrated that the entrapment efficiency of bovine serum albumin particles, when loaded with the anti-cancer drug doxorubicin, was 75%, and the sustained release effect lasted for up to 96 h. The particles could reduce the cancer cell activity to a greater extent than free doxorubicin; thus, the nanoparticles were capable of delivering doxorubicin.

# 5.3. Chitosan Nanoparticles

Chitosan is a natural polymer with easy structure modification, low cytotoxicity, good biocompatibility, and biodegradability; therefore, it is a good choice as a carrier [150]. Chitosan can be used as a pigment carrier in pigment extraction due to its good adsorption. Tanabtabzadeh et al. [151] confirmed that chitosan adsorption could extract betaine from beetroot plants. Xue et al. [152] prepared chitosan microspheres loaded with R-phycocrythrin using the suspension crosslinking method and studied the effect of adding Agar to the microspheres on R-phycocrythrin release. The results displayed that R-phycocyanin has a maximum loading rate at pH 3.59, and this loading rate increases with an increase in the temperature and solution salt ion concentration. Chitosan can achieve the sustained release of R-phycocrythrin, and the release rate can be accelerated after adding agarose; therefore, the release rate of R-phycocrythrin can be adjusted according to the amount of agarose in the particles. Gandia-Herrero et al. [153] used spray drying procedures to encapsulate betaine into chitosan, which protected the anti-antioxidant activity of betaine for six months. The above studies confirmed the potential of chitosan for extraction, encapsulation, and preservation of pigments.

## 5.4. Metal Ions

Dietary or intestinal microorganisms produce metal nanoparticles in human or animal blood and tissues. These are primarily single atoms of metals, such as gold, silver, iron, and platinum, or protein nanoparticles combined with proteins. These endogenous metal nanoparticles have been found to inhibit cancer cell growth and reproduction. Therefore, metal nanoparticles as transport carriers have unique physical and chemical properties [154]. Venil et al. [155] treated keraxanthin extracted from Sanguisorba officinalis with a silver nitrate solution to prepare silver nanoparticles loaded with cornaxanthin. The nanoparticles demonstrated significant cytotoxicity to the human keratinocyte line with potential wound-healing properties. Sigurdson et al. [156] confirmed that the stability of acylated anthocyanins was significantly improved after forming chelates with aluminum and iron ions. Davaeifar et al. [157] synthesized phycocyanin–zinc oxide nanorods. The phycocyanin-coated zinc oxide nanorods had good thermal stability and less cytotoxicity to fibroblasts L929. In summary, metal nanoparticles have great potential for improving pigment stability and as carriers for pigment transport.

# 6. Prospect

Natural pigments are widely used in cosmetics and food industries due to their wide variety, different colors, and high safety; for example, Pepha®-Ctive containing  $\beta$ -carotene was launched by Pentapharm Ltd., Linablue® containing phycocyanin was launched by Dainippon Ink and Chemicals Inc., AstaPure® containing astaxanthin was launched by Algatech and FucoVital TM containing fucoxanthin [158].

However, natural plant pigments have problems, such as poor stability and solubility. Additionally, the practical application of natural pigments has numerous problems. First, the number of natural colorants approved by authoritative national organizations for food and beverages is limited. Second, there are no clear standards for using some pigments. The scope of application and dosage without detection standards could lead to the abuse and overuse of pigments, resulting in harmful events. Finally, a natural pigment yield

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was low due to the limitations of extraction technology, resulting in a large waste of resources, and its practical application was limited. Modern pharmacological studies have confirmed that natural pigments have various biological activities and can be used for treating different diseases. However, their application is now limited to the food and cosmetic industries, and few of them have been used in the pharmaceutical industry. Therefore, its pharmacological effect cannot be fully realized. This study reviewed the chemical classification, extraction methods, biological activities, and modification methods of natural plant pigments to expand the natural plant pigment application in medicine, food, and cosmetics. These different extraction technologies increased the natural pigment yield while preventing degradation and oxidation during extraction. These technologies reduced the use of organic solvents while increasing the environmental protection and safety of the extraction process. Additionally, more mature studies confirmed that pigment stability could be improved by wrapping pigments into different nanoparticles in order to prolong the service life of natural pigments, which helps to increase the application of the natural pigment in different ways.

The rich and diverse plant resources in nature provide a large number of natural pigments. In recent years, people's demand for natural plant pigments has increased. With the development of production technology, the largescale preparation of natural plant pigments tends to mature. The commercial application of products based on natural plant pigments is becoming more promising with potential economic benefits.

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#### References

1. Simon, J.E.; Decker, E.A.; Ferruzzi, M.G.; Giusti, M.M.; Mejia, C.D.; Goldschmidt, M.; Talcott, S.T. Establishing Standards on Colors from Natural Sources. *J. Food Sci.* **2017**, *82*, 2539–2553. [CrossRef] [PubMed]

- 2. De Mejia, E.G.; Zhang, Q.; Penta, K.; Eroglu, A.; Lila, M.A. The Colors of Health: Chemistry, Bioactivity, and Market Demand for Colorful Foods and Natural Food Sources of Colorants. *Annu. Rev. Food Sci. Technol.* **2020**, *11*, 145–182. [CrossRef] [PubMed]
- 3. Paul Tania, B.T.K.; Mondal, A.; Tiwari, O.N.; Muthuraj, M.; Bhunia, B. A comprehensive review on recent trends in production, purification, and applications of prodigiosin. In *Biomass Conversion and Biore-Finery*; Springer: Berlin/Heidelberg, Germany, 2022. [CrossRef]
- 4. Nirmal, N.P.; Mereddy, R.; Maqsood, S. Recent developments in emerging technologies for beetroot pigment extraction and its food applications. *Food Chem.* **2021**, *356*, 129611. [CrossRef] [PubMed]
- 5. Shen, N.; Ren, J.; Liu, Y.; Sun, W.; Li, Y.; Xin, H.; Cui, Y. Natural edible pigments: A comprehensive review of resource, chemical classification, biosynthesis pathway, separated methods and application. *Food Chem.* **2023**, *403*, 134422. [CrossRef]
- 6. Liu, Y.; Liu, Y. Construction of lipid-biomacromolecular compounds for loading and delivery of carotenoids: Preparation methods, structural properties, and absorption-enhancing mechanisms. *Crit. Rev. Food Sci. Nutr.* **2022**, *5*, 1–24. [CrossRef]
- 7. Slonimskiy, Y.B.; Egorkin, N.A.; Friedrich, T.; Maksimov, E.G.; Sluchanko, N.N. Microalgal protein AstaP is a potent carotenoid solubilizer and delivery module with a broad carotenoid binding repertoire. *FEBS J.* **2022**, *289*, 999–1022. [CrossRef]
- 8. Guo, Y.; Sun, Q.; Wu, F.G.; Dai, Y.; Chen, X. Polyphenol-Containing Nanoparticles: Synthesis, Properties, and Therapeutic Delivery. *Adv. Mater.* **2021**, *33*, e2007356. [CrossRef]
- 9. Bai, S.K.; Lee, S.J.; Na, H.J.; Ha, K.S.; Han, J.A.; Lee, H.; Kwon, Y.G.; Chung, C.K.; Kim, Y.M. β-Carotene inhibits inflam-matory gene expression in lipopolysaccharide-stimulated macrophages by suppressing redox-based NF-kappa B activation. *Exp. Mol. Med.* 2005, *37*, 323–334. [CrossRef]

Molecules **2023**, 28, 5364 13 of 18

10. Ha, D.-O.; Park, C.U.; Kim, M.-J.; Lee, J. Antioxidant and prooxidant activities of β-carotene in accelerated autoxidation and photosensitized model systems. *Food Sci. Biotechnol.* **2012**, *21*, 607–611. [CrossRef]

- 11. Yang, S.-S.; Kim, S.-G.; Park, B.-S.; Go, D.-S.; Yu, S.-K.; Kim, C.S.; Kim, J.; Kim, D.K. Effect of β-carotene on Cell Growth In-hibition of KB Human Oral Cancer Cells. *Int. J. Oral. Biol.* **2016**, *41*, 105–111. [CrossRef]
- 12. Ou, S.; Fang, Y.; Tang, H.; Wu, T.; Chen, L.; Jiang, M.; Zhou, L.; Xu, J.; Guo, K. Lycopene protects neuroblastoma cells against oxidative damage via depression of ER stress. *J. Food Sci.* **2020**, *85*, 3552–3561. [CrossRef]
- 13. Polat, H.; Sagit, M.; Gurgen, S.G.; Yasar, M.; Ozcan, I. Protective role of lycopene in experimental allergic rhinitis in rats. *Int. J. Pediatr. Otorhinolaryngol.* **2021**, *150*, 110905. [CrossRef]
- Marzocco, S.; Singla, R.K.; Capasso, A. Multifaceted Effects of Lycopene: A Boulevard to the Multitarget-Based Treatment for Cancer. Molecules 2021, 26, 5333. [CrossRef]
- Liu, H.; Liu, J.; Liu, Z.; Wang, Q.; Liu, J.; Feng, D.; Zou, J. Lycopene Reduces Cholesterol Absorption and Prevents Athero-sclerosis in ApoE(-/-) Mice by Downregulating HNF-1alpha and NPC1L1 Expression. J. Agric. Food Chem. 2021, 69, 10114–10120. [CrossRef]
- Chang, M.X.; Xiong, F. Astaxanthin and its Effects in Inflammatory Responses and Inflammation-Associated Diseases: Recent Advances and Future Directions. *Molecules* 2020, 25, 5342. [CrossRef]
- 17. Wang, Y.; Liu, Y.; Liu, B.; Wu, P.; Xu, S.; Shi, H. Protective effects of astaxanthin on subarachnoid hemorrhage-induced early brain injury: Reduction of cerebral vasospasm and improvement of neuron survival and mitochondrial function. *Acta Histochem.* **2019**, *121*, 56–63. [CrossRef]
- 18. Diao, W.; Chen, W.; Cao, W.; Yuan, H.; Ji, H.; Wang, T.; Chen, W.; Zhu, X.; Zhou, H.; Guo, H.; et al. Astaxanthin protects against renal fibrosis through inhibiting myofibroblast activation and promoting CD8(+) T cell recruitment. *Biochim. Biophys. Acta Gen. Subj.* 2019, 1863, 1360–1370. [CrossRef]
- 19. Karimian, A.; Mir Mohammadrezaei, F.; Hajizadeh Moghadam, A.; Bahadori, M.H.; Ghorbani-Anarkooli, M.; Asadi, A.; Abdolmaleki, A. Effect of astaxanthin and melatonin on cell viability and DNA damage in human breast cancer cell lines. *Acta Histochem.* 2022, 124, 151832. [CrossRef]
- 20. Zheng, X.; Huang, Q. Assessment of the antioxidant activities of representative optical and geometric isomers of astaxanthin against singlet oxygen in solution by a spectroscopic approach. *Food Chem.* **2022**, *395*, 133584. [CrossRef]
- 21. Chen, Y.; Wang, L.; Huang, S.; Ke, J.; Wang, Q.; Zhou, Z.; Chang, W. Lutein attenuates angiotensin II- induced cardiac re-modeling by inhibiting AP-1/IL-11 signaling. *Redox. Biol.* **2021**, *44*, 102020. [CrossRef]
- 22. Jv, D.J.; Ji, T.H.; Xu, Z.; Li, A.; Chen, Z.Y. The remarkable enhancement of photo-stability and antioxidant protection of lutein coupled with carbon-dot. *Food Chem.* **2023**, *405*, 134551. [CrossRef] [PubMed]
- 23. Mohammad Pour, M.; Farjah, G.H.; Karimipour, M.; Pourheidar, B.; Khadem Ansari, M.H. Protective effect of lutein on spinal cord ischemia-reperfusion injury in rats. *Iran. J. Basic Med. Sci.* **2019**, 22, 412–417. [CrossRef] [PubMed]
- 24. Han, L.; Song, X. Lutein induces an inhibitory effect on the malignant progression of pancreatic adenocarcinoma by targeting BAG3/cholesterol homeostasis. *J. Biochem. Mol. Toxicol.* **2022**, *36*, e22958. [CrossRef] [PubMed]
- 25. El-Akabawy, G.; El-Sherif, N.M. Zeaxanthin exerts protective effects on acetic acid-induced colitis in rats via modulation of pro-inflammatory cytokines and oxidative stress. *Biomed. Pharmacother.* **2019**, *111*, 841–851. [CrossRef] [PubMed]
- 26. Huang, Y.; Shi, C.; Li, J. The protective effect of zeaxanthin on human limbal and conjunctival epithelial cells against UV-induced cell death and oxidative stress. *Int. J. Ophthalmol.* **2019**, *12*, 369–374. [CrossRef]
- 27. Sheng, Y.N.; Luo, Y.H.; Liu, S.B.; Xu, W.T.; Zhang, Y.; Zhang, T.; Xue, H.; Zuo, W.B.; Li, Y.N.; Wang, C.Y.; et al. Zeaxanthin Induces Apoptosis via ROS-Regulated MAPK and AKT Signaling Pathway in Human Gastric Cancer Cells. *Onco. Targets Ther.* **2020**, *13*, 10995–11006. [CrossRef]
- 28. Zhang, L.N.; Li, M.J.; Shang, Y.H.; Liu, Y.R.; Han-Chang, H.; Lao, F.X. Zeaxanthin Attenuates the Vicious Circle Between Endoplasmic Reticulum Stress and Tau Phosphorylation: Involvement of GSK-3 beta Activation. *J. Alzheimers Dis.* 2022, 86, 191–204. [CrossRef]
- 29. Guo, L.; Kang, J.S.; Kang, N.J.; Je, B.I.; Lee, Y.J.; Park, Y.H.; Choi, Y.W. Pelargonidin suppresses adipogenesis in 3T3-L1 cells through inhibition of PPAR-gamma signaling pathway. *Arch. Biochem. Biophys.* **2020**, *686*, 108365. [CrossRef]
- 30. Tian, Z.; Sun, C.; Liu, J. Pelargonidin inhibits vascularization and metastasis of brain gliomas by blocking the PI3K/AKT/mTOR pathway. *J. Biosci.* **2022**, *47*, 64. [CrossRef]
- 31. Lee, I.C.; Bae, J.S. Pelargonidin Protects Against Renal Injury in a Mouse Model of Sepsis. J. Med. Food 2019, 22, 57–61. [CrossRef]
- 32. Seo, M.; Kim, H.; Lee, J.H.; Park, J.W. Pelargonidin ameliorates acetaminophen-induced hepatotoxicity in mice by inhibiting the ROS-induced inflammatory apoptotic response. *Biochimie* **2020**, *168*, 10–16. [CrossRef]
- 33. Suantawee, T.; Thilavech, T.; Cheng, H.; Adisakwattana, S. Cyanidin Attenuates Methylglyoxal-Induced Oxidative Stress and Apoptosis in INS-1 Pancreatic beta-Cells by Increasing Glyoxalase-1 Activity. *Nutrients* **2020**, *12*, 1319. [CrossRef]
- 34. Samarpita, S.; Rasool, M. Cyanidin attenuates IL-17A cytokine signaling mediated monocyte migration and differentiation into mature osteoclasts in rheumatoid arthritis. *Cytokine* **2021**, *142*, 155502. [CrossRef]
- 35. Gan, Y.; Fu, Y.; Yang, L.; Chen, J.; Lei, H.; Liu, Q. Cyanidin-3-O-Glucoside and Cyanidin Protect Against Intestinal Barrier Damage and 2,4,6-Trinitrobenzenesulfonic Acid-Induced Colitis. *J. Med. Food* **2020**, 23, 90–99. [CrossRef]
- 36. Yue, H.; Xu, Q.; Lv, L.; Xu, J.; Fan, H.; Wang, W. Cyanidin and peonidin inhibit SPCA-1 growth in vitro via inducing Cell Cycle Arrest and Apoptosis. *Acta Pol. Pharm. Drug Res.* **2019**, *76*, 503–509. [CrossRef]

Molecules **2023**, 28, 5364 14 of 18

37. Yun, J.M.; Afaq, F.; Khan, N.; Mukhtar, H. Delphinidin, an anthocyanidin in pigmented fruits and vegetables, induces apoptosis and cell cycle arrest in human colon cancer HCT116 cells. *Mol. Carcinog.* **2009**, *48*, 260–270. [CrossRef]

- 38. Tsiogkas, S.G.; Mavropoulos, A.; Skyvalidas, D.N.; Patrikiou, E.; Ntavari, N.; Daponte, A.I.; Grammatikopoulou, M.G.; Dar-diotis, E.; Roussaki-Schulze, A.V.; Sakkas, L.I.; et al. Delphinidin diminishes in vitro interferon-gamma and interleukin-17 producing cells in patients with psoriatic disease. *Immunol. Res.* **2022**, *70*, 161–173. [CrossRef]
- 39. Chen, J.; Li, H.Y.; Wang, D.; Guo, X.Z. Delphinidin protects beta2m-/Thy1+ bone marrow-derived hepatocyte stem cells against TGF-beta1-induced oxidative stress and apoptosis through the PI3K/Akt pathway in vitro. *Chem. Biol. Interact.* **2019**, 297, 109–118. [CrossRef]
- 40. Heysieattalab, S.; Sadeghi, L. Effects of Delphinidin on Pathophysiological Signs of Nucleus Basalis of Meynert Lesioned Rats as Animal Model of Alzheimer Disease. *Neurochem. Res.* **2020**, *45*, 1636–1646. [CrossRef]
- 41. Ren, Z.T.; Raut, N.A.; Lawal, T.O.; Patel, S.R.; Lee, S.M.; Mahady, G.B. Peonidin-3-O-glucoside and cyanidin increase osteoblast differentiation and reduce RANKL-induced bone resorption in transgenic medaka. *Phytother. Res.* **2021**, *35*, 6255–6269. [CrossRef]
- 42. Kwon, J.Y.; Lee, K.W.; Hur, H.J.; Lee, H.J. Peonidin inhibits phorbol-ester-induced COX-2 expression and transformation in JB6 P+ cells by blocking phosphorylation of ERK-1 and -2. *Ann. N. Y. Acad. Sci.* **2007**, *1095*, 513–520. [CrossRef] [PubMed]
- 43. Zhang, G.; Chen, S.; Zhou, W.; Meng, J.; Deng, K.; Zhou, H.; Hu, N.; Suo, Y. Anthocyanin composition of fruit extracts from Lycium ruthenicum and their protective effect for gouty arthritis. *Ind. Crop. Prod.* **2019**, 129, 414–423. [CrossRef]
- 44. Nagaoka, M.; Maeda, T.; Moriwaki, S.; Nomura, A.; Kato, Y.; Niida, S.; Kruger, M.C.; Suzuki, K. Petunidin, a B-ring 5'-O-Methylated Derivative of Delphinidin, Stimulates Osteoblastogenesis and Reduces sRANKL-Induced Bone Loss. *Int. J. Mol. Sci.* **2019**, 20, 2795. [CrossRef]
- 45. Cai, X.; Yang, C.; Shao, L.; Zhu, H.; Wang, Y.; Huang, X.; Wang, S.; Hong, L. Targeting NOX 4 by petunidin improves anoxia/reoxygenation-induced myocardium injury. *Eur. J. Pharmacol.* **2020**, *888*, 173414. [CrossRef] [PubMed]
- 46. Shih, P.H.; Yeh, C.T.; Yen, G.C. Effects of anthocyanidin on the inhibition of proliferation and induction of apoptosis in human gastric adenocarcinoma cells. *Food Chem. Toxicol.* **2005**, *43*, 1557–1566. [CrossRef] [PubMed]
- 47. Dai, T.; Shi, K.; Chen, G.; Shen, Y.; Pan, T. Malvidin attenuates pain and inflammation in rats with osteoarthritis by suppressing NF-kappaB signaling pathway. *Inflamm. Res.* **2017**, *66*, 1075–1084. [CrossRef]
- 48. Wei, H.; Li, H.; Wan, S.P.; Zeng, Q.T.; Cheng, L.X.; Jiang, L.L.; Peng, Y.D. Cardioprotective Effects of Malvidin Against Isoproterenol-Induced Myocardial Infarction in Rats: A Mechanistic Study. *Med. Sci. Monit.* 2017, 23, 2007–2016. [CrossRef]
- 49. Gilani, S.J.; Bin-Jumah, M.N.; Al-Abbasi, F.A.; Imam, S.S.; Alshehri, S.; Ghoneim, M.M.; Shahid Nadeem, M.; Afzal, M.; Alzarea, S.I.; Sayyed, N.; et al. Antiamnesic Potential of Malvidin on Aluminum Chloride Activated by the Free Radical Scavenging Property. ACS Omega 2022, 7, 24231–24240. [CrossRef]
- 50. Shehzad, A.; Rehman, G.; Lee, Y.S. Curcumin in inflammatory diseases. Biofactors 2013, 39, 69–77. [CrossRef]
- 51. Agrawal, D.K.; Mishra, P.K. Curcumin and its analogues: Potential anticancer agents. Med. Res. Rev. 2010, 30, 818–860. [CrossRef]
- 52. Monroy, A.; Lithgow, G.J.; Alavez, S. Curcumin and neurodegenerative diseases. Biofactors 2013, 39, 122–132. [CrossRef]
- 53. Zheng, D.; Huang, C.; Huang, H.; Zhao, Y.; Khan, M.R.U.; Zhao, H.; Huang, L. Antibacterial Mechanism of Curcumin: A Review. *Chem. Biodivers.* **2020**, *17*, e2000171. [CrossRef]
- 54. Zhang, F.Y.; Li, R.Z.; Xu, C.; Fan, X.X.; Li, J.X.; Meng, W.Y.; Wang, X.R.; Liang, T.L.; Guan, X.X.; Pan, H.D.; et al. Emodin induces apoptosis and suppresses non-small-cell lung cancer growth via downregulation of sPLA2-IIa. *Phytomedicine* **2022**, *95*, 153786. [CrossRef]
- 55. Xiao, D.; Zhang, Y.; Wang, R.; Fu, Y.; Zhou, T.; Diao, H.; Wang, Z.; Lin, Y.; Li, Z.; Wen, L.; et al. Emodin alleviates cardiac fibrosis by suppressing activation of cardiac fibroblasts via upregulating metastasis associated protein 3. *Acta Pharm. Sin. B* **2019**, *9*, 724–733. [CrossRef]
- 56. Song, Y.; Cui, X.; Zhao, R.; Hu, L.; Li, Y.; Liu, C. Emodin protects against lipopolysaccharide-induced inflammatory injury in HaCaT cells through upregulation of miR-21. *Artif. Cells Nanomed. Biotechnol.* **2019**, 47, 2654–2661. [CrossRef]
- 57. Leung, S.W.; Lai, J.H.; Wu, J.C.; Tsai, Y.R.; Chen, Y.H.; Kang, S.J.; Chiang, Y.H.; Chang, C.F.; Chen, K.Y. Neuroprotective Effects of Emodin against Ischemia/Reperfusion Injury through Activating ERK-1/2 Signaling Pathway. *Int. J. Mol. Sci.* 2020, 21, 2899. [CrossRef]
- 58. Jeon, J.-H.; Song, H.-Y.; Kim, M.-G.; Lee, H.-S. Anticoagulant Properties of Alizarin and Its Derivatives Derived from the Seed Extract of Cassia obtusifolia. *J. Korean Soc. Appl. Biol. Chem.* **2009**, *52*, 163–167. [CrossRef]
- 59. Lee, J.H.; Kim, Y.G.; Park, S.; Hu, L.; Lee, J. Phytopigment Alizarin Inhibits Multispecies Biofilm Development by Cutibacterium acnes, Staphylococcus aureus, and Candida albicans. *Pharmaceutics* **2022**, *14*, 1047. [CrossRef]
- 60. Saha, M.; Singha, S.; Ghosh, D.; Kumar, S.; Karmakar, P.; Das, S. A Cobalt(II)/Cobalt(III) complex of alizarin that was analyzed from the stand point of binding with DNA, for ROS generation and anticancer drug prospecting was identified as an analogue of anthracyclines. *J. Mol. Struct.* **2022**, 1262, 16. [CrossRef]
- 61. Demir, E.; Kaya, B.; Kocaoglu Cenkci, S. Antigenotoxic Activities of Ascorbic acid, Chlorophyll a, and Chlorophyll b in Acrolein and Malondialdehyde-Induced Genotoxicity in Drosophila melanogaster. *Ekoloji* **2013**, 22, 36–42. [CrossRef]
- 62. Subramoniam, A.; Asha, V.V.; Nair, S.A.; Sasidharan, S.P.; Sureshkumar, P.K.; Rajendran, K.N.; Karunagaran, D.; Rama-lingam, K. Chlorophyll revisited: Anti-inflammatory activities of chlorophyll a and inhibition of expression of TNF-alpha gene by the same. *Inflammation* 2012, 35, 959–966. [CrossRef] [PubMed]

Molecules **2023**, 28, 5364 15 of 18

63. Lanfer-Marquez, U.M.; Barros, R.M.C.; Sinnecker, P. Antioxidant activity of chlorophylls and their derivatives. *Food Res. Int.* **2005**, 38, 885–891. [CrossRef]

- 64. Guo, W.; Zeng, M.; Zhu, S.; Li, S.; Qian, Y.; Wu, H. Phycocyanin ameliorates mouse colitis via phycocyanobilin-dependent antioxidant and anti-inflammatory protection of the intestinal epithelial barrier. *Food Funct.* **2022**, *13*, 3294–3307. [CrossRef] [PubMed]
- 65. Garcia-Pliego, E.; Franco-Colin, M.; Rojas-Franco, P.; Blas-Valdivia, V.; Serrano-Contreras, J.I.; Penton-Rol, G.; Cano-Europa, E. Phycocyanobilin is the molecule responsible for the nephroprotective action of phycocyanin in acute kidney injury caused by mercury. *Food Funct.* **2021**, *12*, 2985–2994. [CrossRef] [PubMed]
- 66. Penton-Rol, G.; Marin-Prida, J.; McCarty, M.F. C-Phycocyanin-derived Phycocyanobilin as a Potential Nutraceutical Approach for Major Neurodegenerative Disorders and COVID-19- induced Damage to the Nervous System. *Curr. Neuropharmacol.* **2021**, *19*, 2250–2275. [CrossRef]
- 67. Yabuta, Y.; Fujimura, H.; Kwak, C.S.; Enomoto, T.; Watanabe, F. Antioxidant Activity of the Phycoerythrobilin Compound Formed from a Dried Korean Purple Laver (*Porphyra* sp.) during in Vitro Digestion. *Food Sci. Technol. Res.* **2010**, *16*, 347–351. [CrossRef]
- 68. Kwankaew, N.; Okuda, H.; Aye-Mon, A.; Ishikawa, T.; Hori, K.; Sonthi, P.; Kozakai, Y.; Ozaki, N. Antihypersensitivity effect of betanin (red beetroot extract) via modulation of microglial activation in a mouse model of neuropathic pain. *Eur. J. Pain.* **2021**, 25, 1788–1803. [CrossRef]
- 69. Zielinska-Przyjemska, M.; Olejnik, A.; Dobrowolska-Zachwieja, A.; Luczak, M.; Baer-Dubowska, W. DNA damage and apoptosis in blood neutrophils of inflammatory bowel disease patients and in Caco-2 cells in vitro exposed to betanin. *Postep. Hig. Med. Dosw.* **2016**, 70, 265–271. [CrossRef]
- 70. Song, F.; Zuo, X.; Zhao, Y.; Li, Q.; Tian, Z.; Yang, Y. Betanin-enriched red beet extract attenuated platelet activation and aggregation by suppressing Akt and P38 Mitogen-activated protein kinases phosphorylation. *J. Funct. Foods.* **2019**, *61*, 103491. [CrossRef]
- Esatbeyoglu, T.; Wagner, A.E.; Motafakkerazad, R.; Nakajima, Y.; Matsugo, S.; Rimbach, G. Free radical scavenging and antioxidant activity of betanin: Electron spin resonance spectroscopy studies and studies in cultured cells. *Food Chem. Toxicol.* 2014, 73, 119–126. [CrossRef]
- 72. Henarejos-Escudero, P.; Hernandez-Garcia, S.; Guerrero-Rubio, M.A.; Garcia-Carmona, F.; Gandia-Herrero, F. Antitumoral Drug Potential of Tryptophan-Betaxanthin and Related Plant Betalains in the Caenorhabditis elegans Tumoral Model. *Anti. Oxid.* **2020**, 9, 646. [CrossRef]
- 73. Martinez-Rodriguez, P.; Guerrero-Rubio, M.A.; Hernandez-Garcia, S.; Henarejos-Escudero, P.; Garcia-Carmona, F.; Gan-dia-Herrero, F. Characterization of betalain-loaded liposomes and its bioactive potential in vivo after ingestion. *Food Chem.* **2023**, 407, 135180. [CrossRef]
- 74. Lakey-Beitia, J.; Kumar, D.J.; Hegde, M.L.; Rao, K.S. Carotenoids as Novel Therapeutic Molecules Against Neurodegenerative Disorders: Chemistry and Molecular Docking Analysis. *Int. J. Mol. Sci.* **2019**, *20*, 5553. [CrossRef]
- 75. Yabuzaki, J. Carotenoids Database: Structures, chemical fingerprints and distribution among organisms. *Database* **2017**, 2017, bax004. [CrossRef]
- 76. Liu, C.; Hu, B.; Cheng, Y.; Guo, Y.; Yao, W.; Qian, H. Carotenoids from fungi and microalgae: A review on their recent pro-duction, extraction, and developments. *Bioresour. Technol.* **2021**, *337*, 125398. [CrossRef]
- 77. Priyadarshani, A.M. A review on factors influencing bioaccessibility and bioefficacy of carotenoids. *Crit. Rev. Food Sci. Nutr.* **2017**, 57, 1710–1717. [CrossRef]
- 78. Melendez-Martinez, A.J.; Mandic, A.I.; Bantis, F.; Bohm, V.; Borge, G.I.A.; Brncic, M.; Bysted, A.; Cano, M.P.; Dias, M.G.; Elgersma, A.; et al. A comprehensive review on carotenoids in foods and feeds: Status quo, applications, patents, and research needs. *Crit. Rev. Food Sci. Nutr.* **2022**, *62*, 1999–2049. [CrossRef]
- 79. Mahsa Sobhani, M.H.F.; Sarah, K.; Reza, K. Immunomodulatory; Anti-inflammatory antioxidant Effects of Polyphenols A Comparative Review on the Parental Compounds and Their Metabolites. *Food Rev. Int.* **2021**, *37*, 759–811. [CrossRef]
- 80. Zhang, H.; Wang, M.; Xiao, J. Stability of polyphenols in food processing. Adv. Food Nutr. Res. 2022, 102, 1–45. [CrossRef]
- 81. Roy, S.; Rhim, J.W. Anthocyanin food colorant and its application in pH-responsive color change indicator films. *Crit. Rev. Food Sci. Nutr.* **2021**, *61*, 2297–2325. [CrossRef]
- 82. Silva, A.S.; Reboredo-Rodriguez, P.; Suntar, I.; Sureda, A.; Belwal, T.; Loizzo, M.R.; Tundis, R.; Sobarzo-Sanchez, E.; Rastrelli, L.; Forbes-Hernandez, T.Y.; et al. Evaluation of the status quo of polyphenols analysis: Part I-phytochemistry, bioactivity, inter-actions, and industrial uses. *Compr. Rev. Food Sci. Food Saf.* 2020, 19, 3191–3218. [CrossRef] [PubMed]
- 83. Li, A.N.; Li, S.; Zhang, Y.J.; Xu, X.R.; Chen, Y.M.; Li, H.B. Resources and biological activities of natural polyphenols. *Nutrients* **2014**, *6*, 6020–6047. [CrossRef] [PubMed]
- 84. Dulo, B.; Phan, K.; Githaiga, J.; Raes, K.; Meester, S.D. Natural Quinone Dyes: A Review on Structure, Extraction Techniques, Analysis and Application Potential. *Waste Biomass Valorization* **2020**, *12*, 6339–6374. [CrossRef]
- 85. Dong, X.; Fu, J.; Yin, X.; Cao, S.; Li, X.; Lin, L.; Huyiligeqi; Ni, J. Emodin: A Review of its Pharmacology, Toxicity and Phar-macokinetics. *Phytother. Res.* **2016**, *30*, 1207–1218. [CrossRef] [PubMed]
- 86. Ban, E.; An, S.H.; Park, B.; Park, M.; Yoon, N.E.; Jung, B.H.; Kim, A. Improved Solubility and Oral Absorption of Emodin-Nicotinamide Cocrystal Over Emodin with PVP as a Solubility Enhancer and Crystallization Inhibitor. *J. Pharm. Sci.* **2020**, *109*, 3660–3667. [CrossRef]

Molecules **2023**, 28, 5364 16 of 18

87. Lu, Z.; Ji, C.; Luo, X.; Lan, Y.; Han, L.; Chen, Y.; Liu, X.; Lin, Q.; Lu, F.; Wu, X.; et al. Nanoparticle-Mediated Delivery of Emodin via Colonic Irrigation Attenuates Renal Injury in 5/6 Nephrectomized Rats. *Front. Pharmacol.* **2020**, *11*, 606227. [CrossRef]

- 88. Pan, C.J.; Wang, J.; Huang, N. Preparation, characterization and in vitro anticoagulation of emodin-eluting controlled biodegradable stent coatings. *Colloids Surf. B Biointerfaces* **2010**, 77, 155–160. [CrossRef]
- 89. Xu, Z.; Hou, Y.; Zou, C.; Liang, H.; Mu, J.; Jiao, X.; Zhu, Y.; Su, L.; Liu, M.; Chen, X.; et al. Alizarin, a nature compound, inhibits the growth of pancreatic cancer cells by abrogating NF-kappaB activation. *Int. J. Biol. Sci.* **2022**, *18*, 2759–2774. [CrossRef]
- 90. Senge, M.O.; Ryan, A.A.; Letchford, K.A.; MacGowan, S.A.; Mielke, T. Chlorophylls, Symmetry, Chirality, and Photosyn-thesis. *Symmetry* **2014**, *6*, 781–843. [CrossRef]
- 91. Solymosi, K.; Mysliwa-Kurdziel, B. Chlorophylls and their Derivatives Used in Food Industry and Medicine. *Mini. Rev. Med. Chem.* **2017**, 17, 1194–1222. [CrossRef]
- 92. Mysliwa-Kurdziel, B.; Solymosi, K. Phycobilins and Phycobiliproteins Used in Food Industry and Medicine. *Mini. Rev. Med. Chem.* **2017**, 17, 1173–1193. [CrossRef]
- 93. Chen, L.; Zhu, Y.; Hu, Z.; Wu, S.; Jin, C. Beetroot as a functional food with huge health benefits: Antioxidant, antitumor, physical function, and chronic metabolomics activity. *Food Sci. Nutr.* **2021**, *9*, 6406–6420. [CrossRef]
- 94. Sadowska-Bartosz, I.; Bartosz, G. Biological Properties and Applications of Betalains. Molecules 2021, 26, 2520. [CrossRef]
- 95. Uwineza, P.A.; Waskiewicz, A. Recent Advances in Supercritical Fluid Extraction of Natural Bioactive Compounds from Natural Plant Materials. *Molecules* **2020**, 25, 3847. [CrossRef]
- 96. Abrahamsson, V.; Cunico, L.P.; Andersson, N.; Nilsson, B.; Turner, C. Multicomponent inverse modeling of supercritical fluid extraction of carotenoids, chlorophyll A, ergosterol and lipids from microalgae. *J. Supercrit. Fluids* **2018**, *139*, *53*–61. [CrossRef]
- 97. Da Porto, C.; Natolino, A. Supercritical fluid extraction of polyphenols from grape seed (Vitis vinifera): Study on process variables and kinetics. *J. Supercrit. Fluids* **2017**, *130*, 239–245. [CrossRef]
- 98. Wen, C.; Zhang, J.; Zhang, H.; Dzah, C.S.; Zandile, M.; Duan, Y.; Ma, H.; Luo, X. Advances in ultrasound assisted extraction of bioactive compounds from cash crops-A review. *Ultrason. Sonochem.* **2018**, *48*, 538–549. [CrossRef]
- 99. Sahin, S.; Nasir, N.T.B.M.; Erken, I.; Cakmak, Z.E.; Cakmak, T. Antioxidant composite films with chitosan and carotenoid extract from Chlorella vulgaris: Optimization of ultrasonic-assisted extraction of carotenoids and surface characterization of chitosan films. *Mater. Res. Express* **2019**, *6*, 095404. [CrossRef]
- 100. Zhong, M.; Huang, S.; Wang, H.; Huang, Y.; Xu, J.; Zhang, L. Optimization of ultrasonic-assisted extraction of pigment from Dioscorea cirrhosa by response surface methodology and evaluation of its stability. *RSC Adv.* **2019**, *9*, 1576–1585. [CrossRef]
- 101. Das, A.; Basak, S.; Chakraborty, S.; Dhibar, M. Microwave: An Ecologically Innovative, Green Extraction Technology. *Curr. Anal. Chem.* **2022**, *18*, 858–866. [CrossRef]
- 102. Sharma, A.; Mazumdar, B.; Keshav, A. Valorization of unsalable Amaranthus tricolour leaves by microwave-assisted extraction of betacyanin and betaxanthin. *Biomass Convers. Bioref.* **2020**, *17*, 1–17. [CrossRef]
- 103. Fernandez-Aulis, F.; Hernandez-Vazquez, L.; Aguilar-Osorio, G.; Arrieta-Baez, D.; Navarro-Ocana, A. Extraction and Identification of Anthocyanins in Corn Cob and Corn Husk from Cacahuacintle Maize. *J. Food Sci.* **2019**, *84*, 954–962. [CrossRef] [PubMed]
- 104. Das, S.; Nadar, S.S.; Rathod, V.K. Integrated strategies for enzyme assisted extraction of bioactive molecules: A review. *Int. J. Biol. Macromol.* **2021**, 191, 899–917. [CrossRef] [PubMed]
- 105. Zhao, X.; Zhang, X.; Liu, H.; Zhu, Y. Enzyme-assisted extraction of astaxanthin from Haematococcus pluvialis and its stability and antioxidant activity. *Food Sci. Biotechnol.* **2019**, 28, 1637–1647. [CrossRef]
- 106. Lombardelli, C.; Benucci, I.; Mazzocchi, C.; Esti, M. A Novel Process for the Recovery of Betalains from Unsold Red Beets by Low-Temperature Enzyme-Assisted Extraction. *Foods* **2021**, *10*, 236. [CrossRef]
- 107. Xi, J. Ultrahigh pressure extraction of bioactive compounds from plants-A review. *Crit. Rev. Food Sci. Nutr.* **2017**, *57*, 1097–1106. [CrossRef]
- 108. Nunes, A.N.; do Carmo, C.S.; Duarte, C.M.M. Production of a natural red pigment derived from Opuntia spp. using a novel high pressure CO₂ assisted-process. *RSC Adv.* **2015**, *5*, 83106–83114. [CrossRef]
- 109. Ferrari, P.F.; Pettinato, M.; Casazza, A.A.; Atanasio, G.D.; Palombo, D.; Perego, P. Polyphenols from Nerone Gold 26/6, a new pigmented rice, via non-conventional extractions: Antioxidant properties and biological validation. *J. Chem. Technol. Biotechnol.* **2021**, *96*, 1691–1699. [CrossRef]
- 110. Shahram, H.; Dinani, S.T. Optimization of ultrasonic-assisted enzymatic extraction of beta-carotene from orange processing waste. *J. Food Process. Eng.* **2019**, 42, 16. [CrossRef]
- 111. Fu, X.Q.; Ma, N.; Sun, W.P.; Dang, Y.Y. Microwave and enzyme co-assisted aqueous two-phase extraction of polyphenol and lutein from marigold (*Tagetes erecta* L.) flower. *Ind. Crop. Prod.* **2018**, *123*, 296–302. [CrossRef]
- 112. Valko, M.; Leibfritz, D.; Moncol, J.; Cronin, M.T.; Mazur, M.; Telser, J. Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.* **2007**, *39*, 44–84. [CrossRef]
- 113. Josson Akkara, P.; Sabina, E.P. A biochemical approach to the anti-inflammatory, antioxidant and antiapoptotic potential of beta-carotene as a protective agent against bromobenzene-induced hepatotoxicity in female Wistar albino rats. *J. Appl. Biomed.* **2020**, *18*, 87–95. [CrossRef]
- 114. Wang, B.; Cui, S.; Mao, B.; Zhang, Q.; Tian, F.; Zhao, J.; Tang, X.; Chen, W. Cyanidin Alleviated CCl(4)-Induced Acute Liver Injury by Regulating the Nrf2 and NF-kappaB Signaling Pathways. *Antioxidants* **2022**, *11*, 2383. [CrossRef]

Molecules **2023**, 28, 5364 17 of 18

115. Yu, L.; Zhang, S.D.; Zhao, X.L.; Ni, H.Y.; Song, X.R.; Wang, W.; Yao, L.P.; Zhao, X.H.; Fu, Y.J. Cyanidin-3-glucoside protects liver from oxidative damage through AMPK/Nrf2 mediated signaling pathway in vivo and in vitro. *J. Funct. Foods* **2020**, *73*, 12. [CrossRef]

- 116. Brotosudarmo, T.H.P.; Limantara, L.; Setiyono, E.; Heriyanto. Structures of Astaxanthin and Their Consequences for Thera-peutic Application. *Int. J. Food Sci.* **2020**, 2020, 2156582. [CrossRef]
- 117. Alfaro, S.; Acuna, V.; Ceriani, R.; Cavieres, M.F.; Weinstein-Oppenheimer, C.R.; Campos-Estrada, C. Involvement of In-flammation and Its Resolution in Disease and Therapeutics. *Int. J. Mol. Sci.* **2022**, 23, 10719. [CrossRef]
- 118. Takahashi, N.; Kake, T.; Hasegawa, S.; Imai, M. Effects of Post-administration of beta-Carotene on Diet-induced Atopic Dermatitis in Hairless Mice. *J. Oleo. Sci.* **2019**, *68*, 793–802. [CrossRef]
- 119. Blas-Valdivia, V.; Moran-Dorantes, D.N.; Rojas-Franco, P.; Franco-Colin, M.; Mirhosseini, N.; Davarnejad, R.; Halajisani, A.; Tavakoli, O.; Cano-Europa, E. C-Phycocyanin prevents acute myocardial infarction-induced oxidative stress, inflammation and cardiac damage. *Pharm. Biol.* **2022**, *60*, 755–763. [CrossRef]
- 120. Li, G.; Duan, L.; Yang, F.; Yang, L.; Deng, Y.; Yu, Y.; Xu, Y.; Zhang, Y. Curcumin suppress inflammatory response in traumatic brain injury via p38/MAPK signaling pathway. *Phytother. Res.* **2022**, *36*, 1326–1337. [CrossRef]
- 121. Manochkumar, J.; Doss, C.G.P.; Efferth, T.; Ramamoorthy, S. Tumor preventive properties of selected marine pigments against colon and breast cancer. *Algal Res.* **2022**, *61*, 102594. [CrossRef]
- 122. Cui, L.; Xu, F.; Wang, M.; Li, L.; Qiao, T.; Cui, H.; Li, Z.; Sun, C. Dietary natural astaxanthin at an early stage inhibits N-nitrosomethylbenzylamine-induced esophageal cancer oxidative stress and inflammation via downregulation of NFkappaB and COX2 in F344 rats. *Onco. Targets Ther.* 2019, 12, 5087–5096. [CrossRef] [PubMed]
- 123. Yeh, Y.C.; Liu, T.J.; Lai, H.C. Shikonin Induces Apoptosis, Necrosis, and Premature Senescence of Human A549 Lung Cancer Cells through Upregulation of p53 Expression. *Evid. Based Complement Altern. Med.* **2015**, 2015, 620383. [CrossRef] [PubMed]
- 124. Lim, W.; Song, G. Inhibitory effects of delphinidin on the proliferation of ovarian cancer cells via PI3K/AKT and ERK 1/2 MAPK signal transduction. *Oncol. Lett.* **2017**, *14*, 810–818. [CrossRef] [PubMed]
- 125. Wittchen, H.U.; Jacobi, F.; Rehm, J.; Gustavsson, A.; Svensson, M.; Jonsson, B.; Olesen, J.; Allgulander, C.; Alonso, J.; Faravelli, C.; et al. The size and burden of mental disorders and other disorders of the brain in Europe 2010. *Eur. Neuropsychopharmacol.* 2011, 21, 655–679. [CrossRef] [PubMed]
- 126. Zhou, J.; Wu, N.; Lin, L. Curcumin Suppresses Apoptosis and Inflammation in Hypoxia/Reperfusion-Exposed Neurons via Wnt Signaling Pathway. *Med. Sci. Monit.* 2020, 26, e920445. [CrossRef]
- 127. Jiang, J.; Wang, W.; Sun, Y.J.; Hu, M.; Li, F.; Zhu, D.Y. Neuroprotective effect of curcumin on focal cerebral ischemic rats by preventing blood-brain barrier damage. *Eur. J. Pharmacol.* **2007**, *561*, 54–62. [CrossRef]
- 128. Gazzin, S.; Dal Ben, M.; Montrone, M.; Jayanti, S.; Lorenzon, A.; Bramante, A.; Bottin, C.; Moretti, R.; Tiribelli, C. Curcumin Prevents Cerebellar Hypoplasia and Restores the Behavior in Hyperbilirubinemic Gunn Rat by a Pleiotropic Effect on the Molecular Effectors of Brain Damage. *Int. J. Mol. Sci.* 2020, 22, 299. [CrossRef]
- 129. Wang, Z.; Liu, T.; Gan, L.; Wang, T.; Yuan, X.; Zhang, B.; Chen, H.; Zheng, Q. Shikonin protects mouse brain against cerebral ischemia/reperfusion injury through its antioxidant activity. *Eur. J. Pharmacol.* **2010**, *643*, 211–217. [CrossRef]
- 130. Cervantes-Llanos, M.; Lagumersindez-Denis, N.; Marin-Prida, J.; Pavon-Fuentes, N.; Falcon-Cama, V.; Piniella-Matamoros, B.; Camacho-Rodriguez, H.; Fernandez-Masso, J.R.; Valenzuela-Silva, C.; Raices-Cruz, I.; et al. Beneficial effects of oral administration of C-Phycocyanin and Phycocyanobilin in rodent models of experimental autoimmune encephalomyelitis. *Life Sci.* 2018, 194, 130–138. [CrossRef]
- 131. Matamoros, B.P.; Prida, J.M.; Rol, G.P. Nutraceutical and therapeutic potential of Phycocyanobilin for treating Alzheimer's disease. *J. Biosci.* **2021**, *46*, 42. [CrossRef]
- 132. Gu, S.; Li, X. Regulation of Autophagy in Cardiovascular Diseases by Natural Products. *Adv. Exp. Med. Biol.* **2020**, 1207, 731–736. [CrossRef]
- 133. Zhou, F.; Hu, X.; Feng, W.; Li, M.; Yu, B.; Fu, C.; Ou, C. LncRNA H19 abrogates the protective effects of curcumin on rat carotid balloon injury via activating Wnt/beta-catenin signaling pathway. *Eur. J. Pharmacol.* **2021**, *910*, 174485. [CrossRef]
- 134. Qian, P.; Yan, L.J.; Li, Y.Q.; Yang, H.T.; Duan, H.Y.; Wu, J.T.; Fan, X.W.; Wang, S.L. Cyanidin ameliorates cisplatin-induced cardiotoxicity via inhibition of ROS-mediated apoptosis. *Exp. Ther. Med.* **2018**, *15*, 1959–1965. [CrossRef]
- 135. Lee, D.K.; Grantham, R.N.; Mannion, J.D.; Trachte, A.L. Carotenoids enhance phosphorylation of Akt and suppress tissue factor activity in human endothelial cells. *J. Nutr. Biochem.* **2006**, *17*, 780–786. [CrossRef]
- 136. Ferreira-Santos, P.; Aparicio, R.; Carron, R.; Sevilla, M.A.; Monroy-Ruiz, J.; Montero, M.J. Lycopene-supplemented diet ameliorates cardiovascular remodeling and oxidative stress in rats with hypertension induced by Angiotensin II. *J. Funct. Foods* **2018**, 47, 279–287. [CrossRef]
- 137. Akbari, J.; Saeedi, M.; Ahmadi, F.; Hashemi, S.M.H.; Babaei, A.; Yaddollahi, S.; Rostamkalaei, S.S.; Asare-Addo, K.; Nokhodchi, A. Solid lipid nanoparticles and nanostructured lipid carriers: A review of the methods of manufacture and routes of admin-istration. *Pharm. Dev. Technol.* **2022**, 27, 525–544. [CrossRef]
- 138. Jain, A.; Singh, S.K.; Arya, S.K.; Kundu, S.C.; Kapoor, S. Protein Nanoparticles: Promising Platforms for Drug Delivery Applications. *ACS Biomater. Sci. Eng.* **2018**, *4*, 3939–3961. [CrossRef]
- 139. Divya, K.; Jisha, M.S. Chitosan nanoparticles preparation and applications. *Environ. Chem. Lett.* **2017**, *16*, 101–112. [CrossRef]

Molecules **2023**, 28, 5364 18 of 18

140. Shabatina, T.; Vernaya, O.; Shumilkin, A.; Semenov, A.; Melnikov, M. Nanoparticles of Bioactive Metals/Metal Oxides and Their Nanocomposites with Antibacterial Drugs for Biomedical Applications. *Materials* **2022**, *15*, 3602. [CrossRef]

- 141. Sakellari, G.I.; Zafeiri, I.; Batchelor, H.; Spyropoulos, F. Formulation design, production and characterisation of solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) for the encapsulation of a model hydrophobic active. *Food Hy-Drocoll. Health* **2021**, *1*, 100024. [CrossRef]
- 142. Osanlou, R.; Emtyazjoo, M.; Banaei, A.; Hesarinejad, M.A.; Ashrafi, F. Preparation of solid lipid nanoparticles and nanostructured lipid carriers containing zeaxanthin and evaluation of physicochemical properties. *Colloid Surf. A-Physicochem. Eng. Asp.* **2022**, 641, 9. [CrossRef]
- 143. Tamjidi, F.; Shahedi, M.; Varshosaz, J.; Nasirpour, A. Stability of astaxanthin-loaded nanostructured lipid carriers as affected by pH, ionic strength, heat treatment, simulated gastric juice and freeze-thawing. *J. Food Sci. Technol.* **2017**, *54*, 3132–3141. [CrossRef] [PubMed]
- 144. Pelissari, J.R.; Souza, V.B.; Pigoso, A.A.; Tulini, F.L.; Thomazini, M.; Rodrigues, C.E.C.; Urbano, A.; Favaro-Trindade, C.S. Production of solid lipid microparticles loaded with lycopene by spray chilling: Structural characteristics of particles and ly-copene stability. *Food Bioprod. Process.* **2016**, *98*, 86–94. [CrossRef]
- 145. Bondi, M.L.; Emma, M.R.; Botto, C.; Augello, G.; Azzolina, A.; Di Gaudio, F.; Craparo, E.F.; Cavallaro, G.; Bachvarov, D.; Cervello, M. Biocompatible Lipid Nanoparticles as Carriers To Improve Curcumin Efficacy in Ovarian Cancer Treatment. *J. Agric. Food Chem.* 2017, 65, 1342–1352. [CrossRef] [PubMed]
- 146. Verma, D.; Gulati, N.; Kaul, S.; Mukherjee, S.; Nagaich, U. Protein Based Nanostructures for Drug Delivery. *J. Pharm.* **2018**, 2018, 9285854. [CrossRef]
- 147. Liu, H.; Zhang, Y.; Zhang, J.B.; Xiong, Y.; Peng, S.F.; McClements, D.J.; Zou, L.Q.; Liang, R.H.; Liu, W. Utilization of protein nanoparticles to improve the dispersibility, stability, and functionality of a natural pigment: Norbixin. *Food Hydrocoll.* **2022**, 124, 10. [CrossRef]
- 148. Radomirovic, M.; Minic, S.; Stanic-Vucinic, D.; Nikolic, M.; Van Haute, S.; Rajkovic, A.; Velickovic, T.C. Phycocyanobil-in-modified beta-lactoglobulin exhibits increased antioxidant properties and stability to digestion and heating. *Food Hydrocol-Loids* **2022**, 123, 10. [CrossRef]
- 149. Gulsu, A.; Aslanpay, M.C.; Alper, M.; Gunes, H. Doxorubicin Release from Bovine Serum Albumin Microparticles. *Pharm. Chem. J.* 2022, *55*, 1157–1162. [CrossRef]
- 150. Yan, X.T.; Song, G.L.; Wang, X.; Zhao, Y.; Chen, Y. The Preparation and Medical Applications of Chitosan Mterospheres. *Curr. Org. Chem.* **2018**, 22, 720–733. [CrossRef]
- 151. Tanabtabzadeh, M.S.; Javanbakht, V.; Golshirazi, A.H. Extraction of Betacyanin and Betaxanthin Pigments from Red Beetroots by Chitosan Extracted from Shrimp Wastes. *Waste Biomass Valorization* **2019**, *10*, 641–653. [CrossRef]
- 152. Xue, Z.X.; Yang, G.P.; Wang, G.C.; Niu, J.F.; Cao, X.Y. Preparation of porous chitosan/agarose microsphere and its R-phycoerythrin release properties. *J. Appl. Polym. Sci.* **2007**, *103*, 2759–2766. [CrossRef]
- 153. Gandia-Herrero, F.; Cabanes, J.; Escribano, J.; Garcia-Carmona, F.; Jimenez-Atienzar, M. Encapsulation of the most potent antioxidant betalains in edible matrixes as powders of different colors. *J. Agric. Food Chem.* **2013**, *61*, 4294–4302. [CrossRef]
- 154. Vodyanoy, V. The Role of Endogenous Metal Nanoparticles in Biological Systems. Biomolecules 2021, 11, 1574. [CrossRef]
- 155. Venil, C.K.; Malathi, M.; Velmurugan, P.; Renuka Devi, P. Green synthesis of silver nanoparticles using canthaxanthin from Dietzia maris AURCCBT01 and their cytotoxic properties against human keratinocyte cell line. *J. Appl. Microbiol.* **2021**, *130*, 1730–1744. [CrossRef]
- 156. Sigurdson, G.T.; Robbins, R.J.; Collins, T.M.; Giusti, M.M. Spectral and colorimetric characteristics of metal chelates of acylated cyanidin derivatives. *Food Chem.* **2017**, 221, 1088–1095. [CrossRef]
- 157. Davaeifar, S.; Modarresi, M.H.; Mohammadi, M.; Hashemi, E.; Shafiei, M.; Maleki, H.; Vali, H.; Zahiri, H.S.; Noghabi, K.A. Synthesizing, characterizing, and toxicity evaluating of Phycocyanin-ZnO nanorod composites: A back to nature approaches. *Colloids Surf. B Biointerfaces* **2019**, 175, 221–230. [CrossRef]
- 158. Morocho-Jacome, A.L.; Ruscinc, N.; Martinez, R.M.; de Carvalho, J.C.M.; Santos de Almeida, T.; Rosado, C.; Costa, J.G.; Ve-lasco, M.V.R.; Baby, A.R. (Bio)Technological aspects of microalgae pigments for cosmetics. *Appl. Microbiol. Biotechnol.* **2020**, 104, 9513–9522. [CrossRef]

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