

Review

Chemistry, Bioactivity, and Prediction of the Quality Marker (Q-Marker) of *Ferula* Plants in China: A Review

Yerlan Bahetjan ¹, Muguli Muhaxi ², Kejian Pang ², Murat Kizaibek ³ , Hui Tang ^{4,*}, Fatemeh Sefidkon ⁵ and Xinzhou Yang ^{1,*} 

¹ International Cooperation Base for Active Substances in Traditional Chinese Medicine in Hubei Province, School of Pharmaceutical Sciences, South-Central Minzu University, 182 Min-Zu Road, Wuhan 430074, China; yerlansb@163.com

² College of Biological and Geographical Sciences, Yili Normal University, Yining 835000, China; muguli@163.com (M.M.); arnebia@126.com (K.P.)

³ Traditional Kazakh Medicine Research Institute of Ili Kazakh Autonomous Prefecture, Yining 835000, China; murat_kizaibek@sina.com

⁴ Key Laboratory of Xinjiang Phytomedicine Resource and Utilization, Ministry of Education, Pharmacy School, Shihezi University, Shihezi 832003, China

⁵ Research Division of Medicinal Plants, Research Institute of Forests and Rangelands, Agricultural Research Education and Extension Organization (AREEO), Tehran P.O. Box 13185-116, Iran; sefidkon@rifr-ac.ir

* Correspondence: th_pha@shzu.edu.cn (H.T.); xzyang@mail.scuec.edu.cn (X.Y.)

Abstract: The genus of *Ferula* belongs to the family Apiaceae, and many *Ferula* plants are used as traditional Chinese medicines. *Ferula* plants were initially identified as early as the “Newly Revised Materia Medica” written in the Tang Dynasty (AD 659), and several of them are also recognized as the traditional medicines of the Uygur, Kazakh, and Mongolian. *Ferula* plants are distributed in China, Russia, India, Africa, Central Asia, and other places. Currently, the chemical components derived from *Ferula* plants are mainly coumarins, sesquiterpenes, and volatile oils. *Ferula* plants can exhibit diverse pharmacological activities such as anti-allergy, analgesia, relieving cough, anticoagulation, and anti-tumor. Therefore, this article summarized the domestic research conducted on the genus *Ferula*, appropriately combines the research status of the foreign genus *Ferula*, and describes the chemical composition, biological activity, toxicity issues, and Q-marker prediction. In addition, all the related studies about the genus *Ferula* are summarized by analyzing the various databases such as CNKI, Wanfang data, PubChem and SciFinder.

Keywords: *Ferula*; pharmacology; chemical composition; Apiaceae; Q-marker



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1. Introduction

Ferula is a tall perennial herb which belongs to the Apiaceae family, and these plants are commonly characterized by a peculiar pungent garlic odor. There are more than 180 species of *Ferula* found worldwide [1] and they are mainly located in Central Asia, the Middle East, Siberia, and South Asia [2]. In China, *Ferula* is mainly distributed in Xinjiang, and a small number of *Ferula* plants are also found in Gansu, Shanxi, Inner Mongolia, and Tibet [3,4]. The genus *Ferula* in China primarily comprises more than twenty species, with a few representative plants shown in Figure 1.

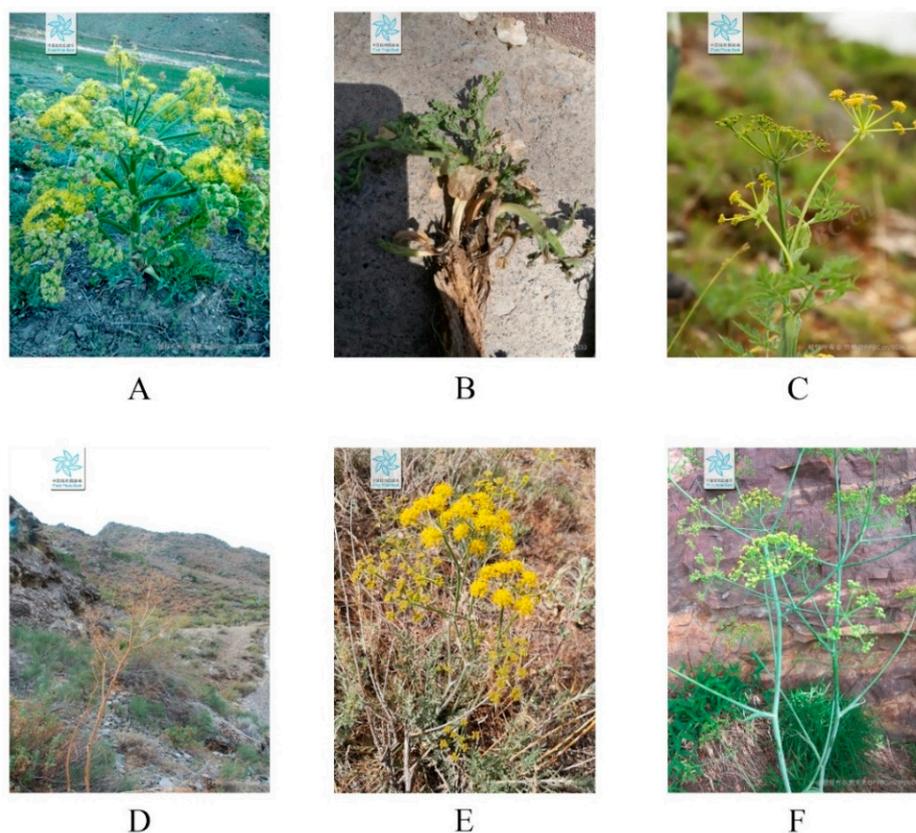


Figure 1. Field morphological pictures of several *Ferula* plants. (A). *Ferula sinkiangensis* [5]; (B). *F. fukanensis* [6]; (C). *F. kingdon-wardii* [7]; (D). *F. songarica* [8]; (E) *F. syreitschikowii* [9]; (F). *F. licentiana* [10]. ((A–F) are from the Plant Photo Bank of China, <http://www.iplant.cn/>, 16 June 2023).

The plants belonging to the genus *Ferula* (Apiaceae) have been implicated in the treatment of various ailments and have a long history of use in many traditional medicines. For instance, Nepali people use *Ferula* in their daily diets [11,12], and it is used locally as a traditional remedy in Saudi Arabia for the treatment of skin infections [13]. In addition, *Ferula* plants are widely used in India in food and as a medicine in Indian systems of medicine such as Ayurveda [14]. Interestingly, in China, *Ferula* plants are also used as an Uyghur medicinal herb, and have a long history of beneficial application [15]. *Ferula* has already been added to the Uyghur Medicine Criteria [16] and the calendar version of the Chinese Pharmacopoeia [17]. In other ethnic groups and regions of China, *Ferula* is often used as traditional medicines. The traditional uses of *Ferula* include beneficial effects against stomach pain, flatulence, intestinal parasites, indigestion, asthma, and flu [18]. For instance, the Kazakhs have used *F. soongarica* to treat headaches, colds and stomach aches, *F. caspica* can treat nervous breakdown, and *F. lehmannii* is used as an anti-parasitic, for anti-malnutrition and to cure cold and pain in the heart and abdomen [12,15]. A number of modern pharmacological and biological studies have indicated that *Ferula* has anticancer [19,20], anticarcinogenic [21], antimicrobial [22], antidiabetic [23], anti-flu [24], anti-inflammatory [25,26], and cardiovascular system effects [27], as well as use against Alzheimer's disease [28,29] and in anti-neuropathic pain [30] activities.

Ferula contains a variety of chemical compounds, primarily coumarins [31], particularly sesquiterpenes coumarins [32], volatile oils [33], sulfur-containing compounds [34], and aromatic compounds [35], all of which exhibit diverse biological activities. However, the Q-marker prediction studies of *Ferula* are relatively scarce. *Ferula* plants of China are mainly distributed in Xinjiang, and it is less commonly present in other regions. Thus, in addition to the over-exploitation and collection of *Ferula* resources, the quality of *Ferula* herbs available on the market is a matter of concern due to the prevalence of counterfeits

and inferior species that are accessible to consumers. Furthermore, the genus *Ferula* seed plant breeding has been continuously shrinking due to the rapid land development, irrigation, highway building, leading to significant decrease in *Ferula* resources. Therefore, novel strategies have to be developed to protect the resources of *Ferula* plants.

Traditional Chinese medicines (TCMs) play an important role in China's modern health care system and have been found to be efficacious in clinical practice. However, due to the lack of criteria for the selection of quality control indicators, both the certification and efficacy of herbal medicines remain unclear [36,37]. Therefore, new approaches are needed to establish Q-markers for determining the quality standards of traditional Chinese medicines, and for identifying the appropriate chemicals present in plants to use them as potential quality indicators [38,39].

There are quite a few Q-marker studies reported for the genus *Ferula*. Thus, the identification of the various markers for the genus *Ferula* by the Q-marker method is essential for the identification of its authenticity and the conservation of the plant resources.

2. Distribution of *Ferula* Plants in China

There are more than 180 species of genus *Ferula* in the worldwide, and more than 20 species have been recorded in China. The genus *Ferula* resources are mainly distributed in Xinjiang and a small number of other provinces in China (Table 1). Most *Ferula* plants are distributed at altitudes between 400 and 3500 m, and the distribution environment is mainly the desert and mountainous areas. However, in recent years, due to the destruction of the plant resources of the genus *Ferula*, wild *Ferula* is nearly extinct.

Table 1. Distribution table of domestic *Ferula* plants.

No	Plant Name	Major Origins	Growing Environment	Literature
1	<i>F. sinkiangensis</i> K.M. Shen	Yining, Xinjiang	750~1000 m in alpine meadows, stony slope areas	[40]
2	<i>F. fukanensis</i> K.M. Shen	Fukang, Xinjiang, Southern Gurbantunggut Desert, Xinjiang	The arid inland Gobi Desert	[41]
3	<i>F. ferulaeoides</i> (Steud.) Korovin	Junggar Basin and Tacheng Basin, Xinjiang	Growing in 430~1000 m sand dunes, sandy land, the environment is mostly desert land	[42]
4	<i>F. conocaula</i> Korov.	Ucha County, Xinjiang	Elevation 2700~3000 m in mountain valleys, environment is mostly mountainous yellow-brown desert soil	[42]
5	<i>F. syreitschikowii</i> K.-Pol.	Yili, Tacheng, Bole, Xinjiang	The environment is mostly low mountain brown calcareous soils, wastelands and gravelly slopes	[43]
6	<i>F. songarica</i> Pallas ex Sprengel	Tacheng and Altay, Xinjiang	The environment is mostly in mountainous grassy slopes and mountainous bushes	[42]
7	<i>F. krylovii</i> Korov.	Tori County, Xinjiang	The environment is mostly clayey saline grassland	[44]
8	<i>F. lehmannii</i> Boiss.	Manas County, Xinjiang	On low mountain slopes at 600~700 m elevation, the environment is mostly clayey gravelly sandy calcareous soil	[45]
9	<i>F. bungeana</i> Kitagawa	Heilongjiang, Jilin, Liaoning, Inner Mongolia, Hebei, Henan, etc.	The environment is mostly sandy and gravelly desert soil or near sandy areas	[46]
10	<i>F. dissecta</i> (Ledeb.) Ledeb	Tacheng Region, Altay Region, Junggar Basin, Xinjiang	The environment is mostly sandy and gravelly desert slopes, mostly in the mountains dominated by <i>Artemisia</i> spp.	[47]

Table 1. Cont.

No	Plant Name	Major Origins	Growing Environment	Literature
11	<i>F. jaeschkeana</i> Vatka	Ali, Zada, Tibet	The environment is mostly at 3600 m above sea level on mountain slopes	[48]
12	<i>F. akitschkensis</i> B. Fedtsch. ex K.-Pol.	Xinjiang Alatau Mountains, Altai Mountains and western Junggar Mountains	The environment is mostly mountain shrublands and gravelly slopes at 900~2100 m above sea level.	[3]
13	<i>F. ovina</i> (Boiss.) Boiss.	Altai, Tacheng, Xinjiang	The environment is mostly gravelly hillsides	[3]
14	<i>F. hexiensis</i> K. M. Shen	Southern Gansu	The environment is mostly hillside with low humidity	[3]
15	<i>F. olivacea</i> (Diels) Wolff ex Hand.-Mazz.	Lijiang, Yunnan	The environment is mostly in canyons and mountain gaps, woods and grasses	[49]
16	<i>F. caspica</i> M. Bieb.	Burqin and Tacheng, Xinjiang	The environment is mostly low mountain slopes and mountain gaps and desert areas	[50]
17	<i>F. subbul</i> (Kauffm.) Hook. f.	Zhaosu County, Xinjiang	The environment is mostly mountainous scrub and gravel slopes	[49]
18	<i>F. licentiana</i> Hand.-Mazz.	Taihang Mountains	The environment is mostly valley grassland at 400~600 m above sea level	[51]
19	<i>F. licentiana</i> Hand.-Mazz.	Jiangsu, Shandong, Anhui, etc.	The environment is mostly sunny slopes, mountain rock crevices and hillsides	[52]
20	<i>F. gracilis</i> (Ledeb.) Ledeb.	Xinjiang Altai Region	The environment is mostly meadows, riverside forest edges and gravelly mountain slopes	[3]
21	<i>F. karataviensis</i> (Regel et Schmaih.) Korov.	Xinyuan County, Xinjiang	The environment is mostly gravelly hillsides	[3]
22	<i>F. dubjanskyi</i> Korov. ex Pavlov	Gansu Province, Anxi and Su Bei, Mahaoshan area	The environment is mostly desert and Gobi desert in the sand and dunes	[53]
23	<i>F. canescens</i> (Ledeb.) Ledeb.	Fuyun County, Xinjiang	Gravelly hillsides in a mostly desert environment	[3]
24	<i>F. kingdon-wardii</i> Wolff	Northwest Yunnan	The environment is mostly grassy slopes and rock crevices at 2700~3200 m above sea level	[54]
25	<i>F. kirialovii</i> Pimenov	Tianshan Mountains and Junggar Basin, Xinjiang	The environment is mostly gravelly grassy slopes and shrubby places at an altitude of about 1500 m	[55]
26	<i>F. lapidosa</i> Korov.	Chabchal County, Xinjiang	The environment is mostly mountainous gravelly slopes and grassy areas	[3]

3. Research on the Chemical Composition of *Ferula* Plants in China

3.1. Coumarins

The full name of 7-hydroxycoumarin is 7-hydroxycoumarin O(7)-glucosiduronic acid. It is a beta-D-glucosiduronic acid. Coumarins are found in the genus *Ferula*, and most of them are derivatives with 7-hydroxycoumarin as the parent nucleus [56]. Coumarins in the genus *Ferula* can be further divided into different types of coumarins according to their substituents, mainly including sesquiterpene coumarins and monoterpene coumarins. Among them, sesquiterpene coumarins can be divided into bicyclic sesquiterpene coumarins (A), monocyclic sesquiterpene coumarins (B), and straight-chain sesquiterpene coumarins (C), in addition to some furan coumarins (D) [57] and other coumarins (E) (Table 2, Figures 2–4).

Researchers obtained a large number of coumarin-like substances from *Ferula sinkiangensis*, *F. lehmannii*, *F. feruloides* and *F. fukanensis* [58–66].

Table 2. Coumarin compounds in domestic *Ferula* plants.

No	Compound	Type	Literature
1	(3'S, 8'R, 9'S, 10'R)-sinkianol A	A	[52]
2	(5'S, 8'R, 9'S, 10'R)-ferukrinone	A	[52]
3	Ferukrin	A	[52]
4	(3'S, 5'S, 8'R, 9'S, 10'R)-kellerin	A	[52]
5	(3'S, 5'S, 8'R, 9'S, 10'R)-deacetylkellerin	A	[52]
6	Farnesiferol A	A	[52]
7	Farnesiferone A	A	[52]
8	Gummosin	A	[52]
9	Polyanthinin	A	[52]
10	(3'R, 5'R, 10'R)-sinkianol B	B	[52]
11	Farnesiferol B	B	[52]
12	Farnesiferol C	B	[52]
13	Galbanic acid	B	[52]
14	Methyl galbanate	B	[52]
15	Sinkiangenol A	A	[52]
16	Karatavicinol	C	[52]
17	Umbelliprenin	C	[52]
18	Sinkiangenol B	A	[53]
19	Sinkiangenol C	B	[53]
20	Sinkiangenol D	A	[53]
21	Sinkiangenol E	A	[53]
22	Sinkiangenol F	E	[53]
23	2,3-Dihydro-7-hydroxy-2R*,3S*-dimethyl-2-[4-methyl-5-(4-methyl-2-furanyl)-3(E)-pentenyl]-furan[3,2-c]coumarin	D	[53]
24	2,3-Dihydro-7-hydroxy-2R*,3R*-dimethyl-2-[4-methyl-5-(4-methyl-2-furanyl)-3(E)-pentenyl]-furan[3,2-c]coumarin	D	[53]
25	12'-hydroxy-karatavicinol	C	[53]
26	Fekrynol	B	[53]
27	Actylfekrynol	B	[53]
28	Ferocaulidin	A	[53]
29	Fekrol	B	[53]
30	Ferucrinone	A	[53]
31	Deacetylkellerin	A	[53]
32	Kellerin	A	[53]
33	Colladocin	A	[53]
34	Lehmannolol	A	[53]
35	Kamolone	A	[53]
36	Assafoetidnol B	A	[53]
37	Assafoetidnol A	A	[53]
38	Lehmannolone A	A	[54]
39	Assafoetidnol	B	[54]
40	Lehmannolone	B	[55]
41	Sinkianone	B	[55]
42	Colladonin	E	[56]
43	Episamarcandin	A	[57]
44	Sinkiangenorin D	A	[57]
45	Fekolone	B	[57]
46	Feselol	A	[57]
47	Ferulin A	D	[58]
48	Ferulin B	D	[58]

Table 2. Cont.

No	Compound	Type	Literature
49	Ferulin C	D	[58]
50	2,3-Dihydro-7-hydroxy-2 <i>S</i> *,3 <i>R</i> *-dimethyl-2-[4,8-dimethyl-3(<i>E</i>),7-nonadienyl]-furo[3,2- <i>c</i>] coumarin	D	[58]
51	2,3-Dihydro-7-hydroxy-2 <i>R</i> *,3 <i>R</i> *-dimethyl-2-[4,8-dimethyl-3(<i>E</i>),7-nonadienyl]-furo[3,2- <i>c</i>] coumarin (DAW22)	D	[58]
52	2,3-Dihydro-7-hydroxy-2 <i>S</i> *,3 <i>R</i> *-dimethyl-2-[4-methyl-5-(4-methyl-2-furanyl)-3(<i>E</i>)-pentenyl]-furo[3,2- <i>c</i>] coumarin	D	[58]
53	2,3-Dihydro-7-hydroxy-2 <i>R</i> *,3 <i>R</i> *-dimethyl-2-[4-methyl-5-(4-methyl-2-furanyl)-3(<i>E</i>)-pentenyl]-furo[3,2- <i>c</i>] coumarin	D	[58]
54	2,3-Dihydro-7-methoxy-2 <i>S</i> *,3 <i>R</i> *-dimethyl-2-[4,8-dimethyl-3(<i>E</i>),7-nonadienyl]-furo[3,2- <i>c</i>] coumarin	D	[58]
55	2,3-Dihydro-7-methoxy-2 <i>R</i> *,3 <i>R</i> *-dimethyl-2-[4,8-dimethyl-3(<i>E</i>),7-nonadienyl]-furo[3,2- <i>c</i>] coumarin	D	[58]
56	2,3-Dihydro-7-hydroxy-2 <i>S</i> *,3 <i>R</i> *-dimethyl-3-[4,8-dimethyl-3(<i>E</i>),7-nonadienyl]-furo[3,2- <i>c</i>] coumarin	D	[58]
57	2,3-Dihydro-7-hydroxy-2 <i>R</i> *,3 <i>R</i> *-dimethyl-3-[4,8-dimethyl-3(<i>E</i>),7-nonadienyl]-furo[3,2- <i>c</i>] coumarin	D	[58]
58	4,7-Dihydroxy-3-[3,7,11-trimethyl-2(<i>E</i>),6(<i>E</i>),10-dodecatrienyl] coumarin	E	[58]
59	2,3-Dihydro-7-hydroxy-2 <i>R</i> *,3 <i>R</i> *-dimethyl-2-[4,8-dimethyl-3(<i>E</i>),7-nonadiene-6-onyl] furo[3,2- <i>c</i>] coumarin	D	[59]
60	Fukanefuomarin A	D	[59]
61	Fukanefuomarin B	D	[59]
62	Fukanefuomarin C	D	[59]
63	Fukanefuomarin D	D	[59]
64	Fukanemarin A	E	[59]
65	Fukanefuomarin H	D	[60]
66	Fukanefuomarin I	D	[60]
67	Fukanefuomarin J	D	[60]
68	Fukanefuomarin K	D	[60]
69	Fukanefuomarin L	D	[60]
70	Fukanefuomarin M	E	[60]

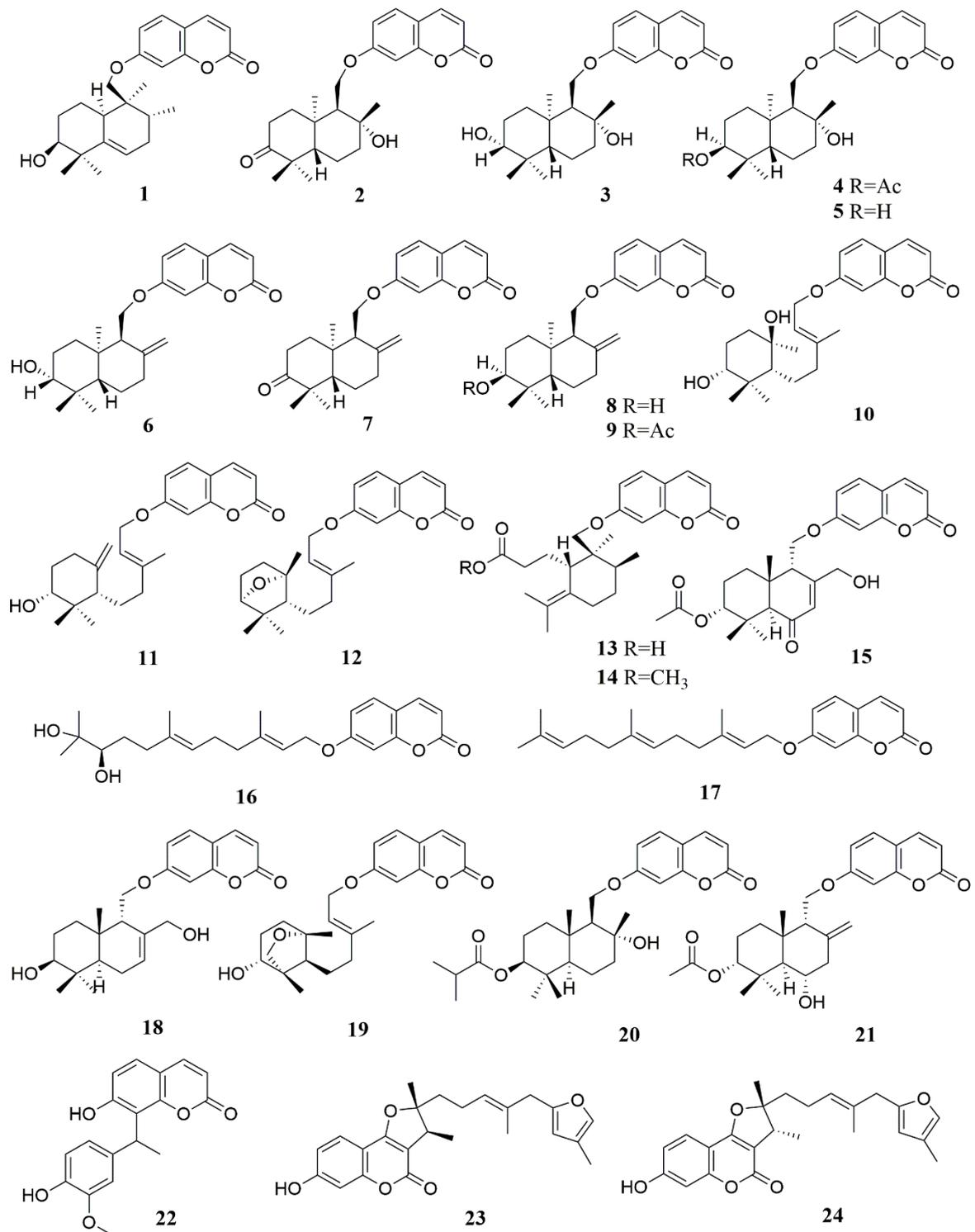


Figure 2. Chemical structures of compounds 1–24 from *Ferula* plants.

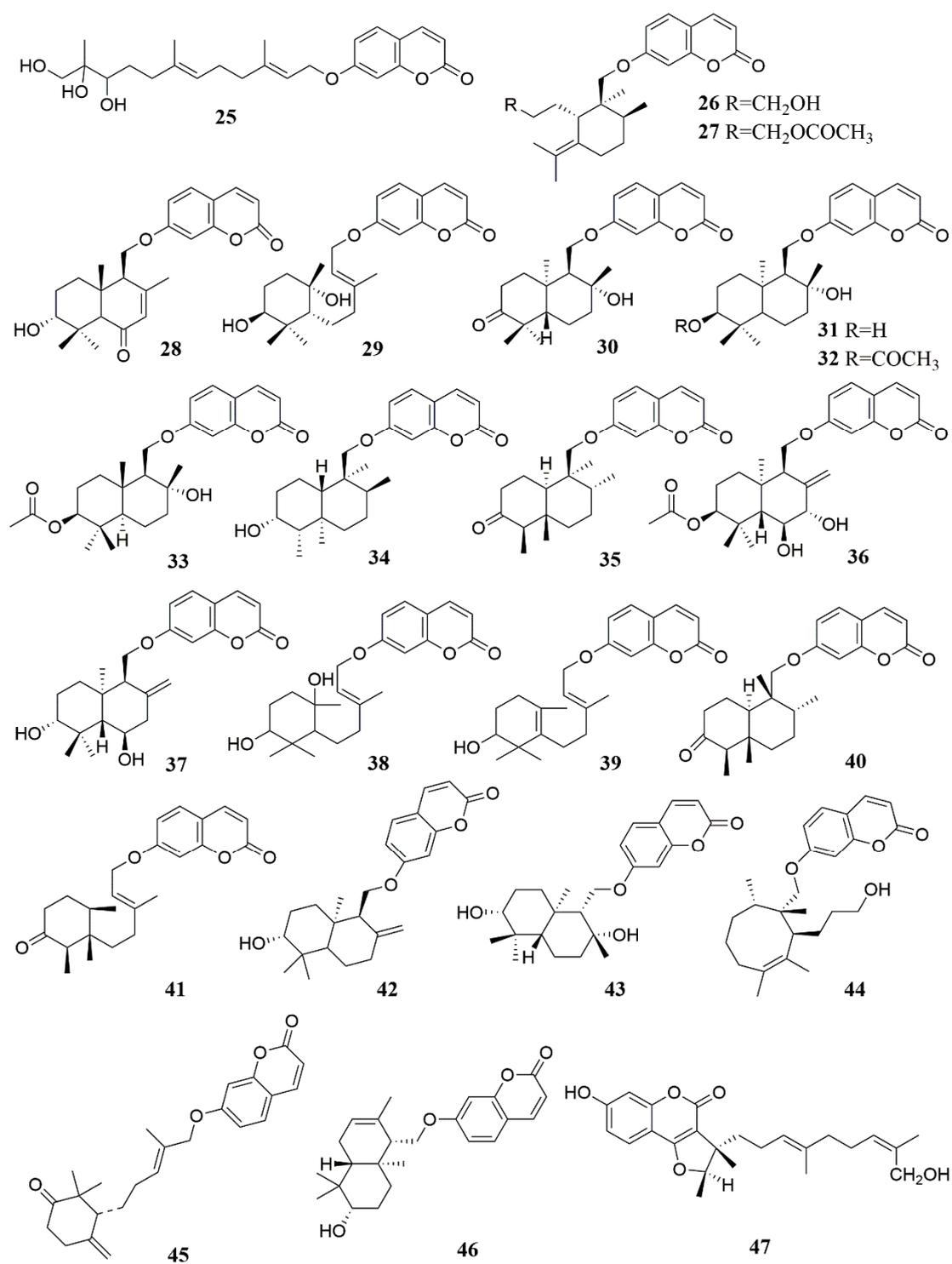


Figure 3. Chemical structures of compounds 25–47 from *Ferula* plants.

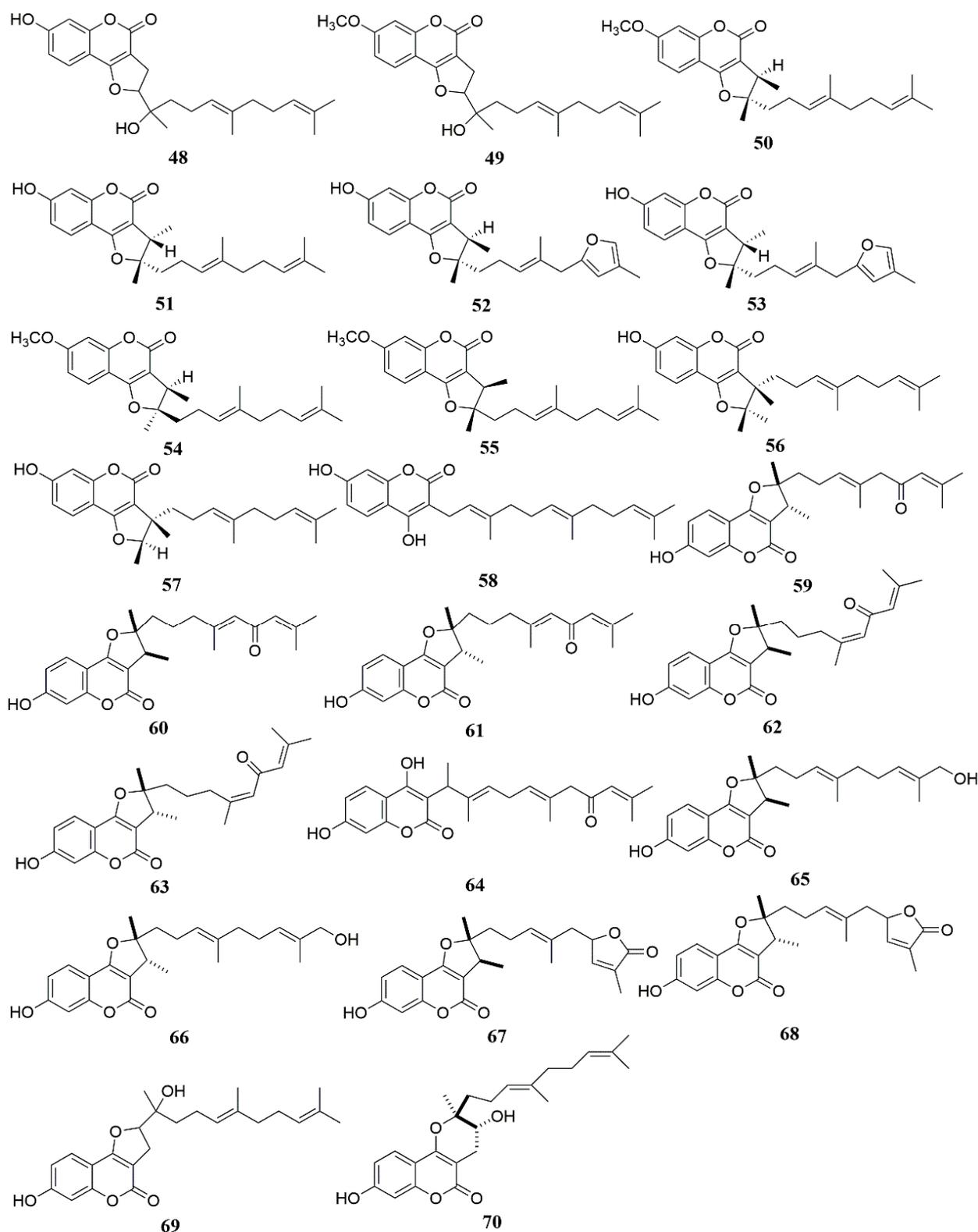


Figure 4. Chemical structures of compounds 48–70 from *Ferula* plants.

3.2. Sesquiterpenoids and Their Derivatives

In addition to the above-mentioned coumarins, sesquiterpenes are the second most abundant substances in *Ferula* plants. The sesquiterpenes of the genus *Ferula* are mainly carotene-type sesquiterpene parent nuclei, but carotene-type sesquiterpenes are rare in domestic *Ferula* plants and are mostly concentrated in foreign *Ferula* plants. The researchers

extracted the following sesquiterpenoids from *F. ferulaeoides*, *F. caspica*, *F. fukanensis* and *F. jaeschkeana* [67–71] (Table 3, Figures 5 and 6).

Table 3. Sesquiterpenoids in Domestic *Ferula* Plants.

No	Compound	Literature
71	Ferulaeone A	[61]
72	Ferulaeone B	[61]
73	Ferulaeone C	[61]
74	Ferulaeone D	[61]
75	Ferulaeone E	[61]
76	Ferulaeone F	[61]
77	Ferulaeone G	[61]
78	Ferulaeone H	[61]
79	Dshamirone	[61]
80	(4 <i>E</i> ,8 <i>E</i>)-1-(2,4-dihydroxyphenyl)-2-hydroxy-5,9,13-trimethyl-tetra-deca-4,8,12-trien-1-one	[61]
81	(6 <i>E</i>)-1-(2,4-dihydroxyphenyl)-3,7,11-trimethyl-3-vinyl-6,10-dodecadien-1-one	[61]
82	(6 <i>E</i>)-1-(2,4-dihydroxyphenyl)-3,7-dimethyl-3-vinyl-8-(4-methyl-2-furyl)-6-octen-1-one	[61]
83	3-(2,4-dihydroxybenzoyl)-4 <i>S</i> *,5 <i>R</i> *-dimethyl-5-[4,8-dimethyl-3(<i>E</i>),7(<i>E</i>)-nonadien-1-yl] tetrahydro-2-furanone	[61]
84	3-(2-hydroxyl-4-methoxybenzoyl)-4 <i>S</i> *,5 <i>R</i> *-dimethyl-5-[4,8-dimethyl-3(<i>E</i>),7(<i>E</i>)-nonadien-1-yl] tetrahydro-2-furanone	[61]
85	3-(2,4-dihydroxybenzoyl)-4 <i>R</i> *,5 <i>R</i> *-dimethyl-5-[4,8-dimethyl-3(<i>E</i>),7(<i>E</i>)-nonadien-1-yl] tetrahydro-2-furanone	[61]
86	1-(2',4'-dihydroxyphenyl)-3,7,11-trimethyl-3-vinyl-6(<i>E</i>),10-dodecadien-1-one	[62]
87	2,3-dihydro-7-hydroxy-2,3-dimethyl-2-[4',8'-dimethyl-3',7'-nonadienyl]-furo [3,2,c] coumarin	[62]
88	2,3-dihydro-7-hydroxy-2,3-dimethyl-3-[4',8'-dimethyl-3',7'-nonadienyl]-furo [3,2,c] coumarin	[62]
89	Fukanefurochromones A	[63]
90	Fukanefurochromones B	[63]
91	Fukanefurochromones C	[63]
92	Fukanefurochromones D	[63]
93	Fukanefurochromones E	[63]
94	2,3-dihydro-7-hydroxy-2 <i>S</i> *,3 <i>R</i> *-dimethyl-2-[4,8-dimethyl-3(<i>E</i>),7-nonadienyl]-furo[2,3-b] chromone	[63]
95	Fukanedone A	[64]
96	Fukanedone B	[64]
97	Fukanedone C	[64]
98	Fukanedone D	[64]
99	Fukanedone E	[64]
100	Fukaneketoester A	[64]
101	Feruone	[5]
102	5α-(<i>p</i> -hydroxybenzyl) ester of ferutriol	[65]
103	Ferutriol	[65]
104	5α-isovalerate of ferutriol	[65]
105	Jaeschkeanol	[65]
106	Lapidin	[65]

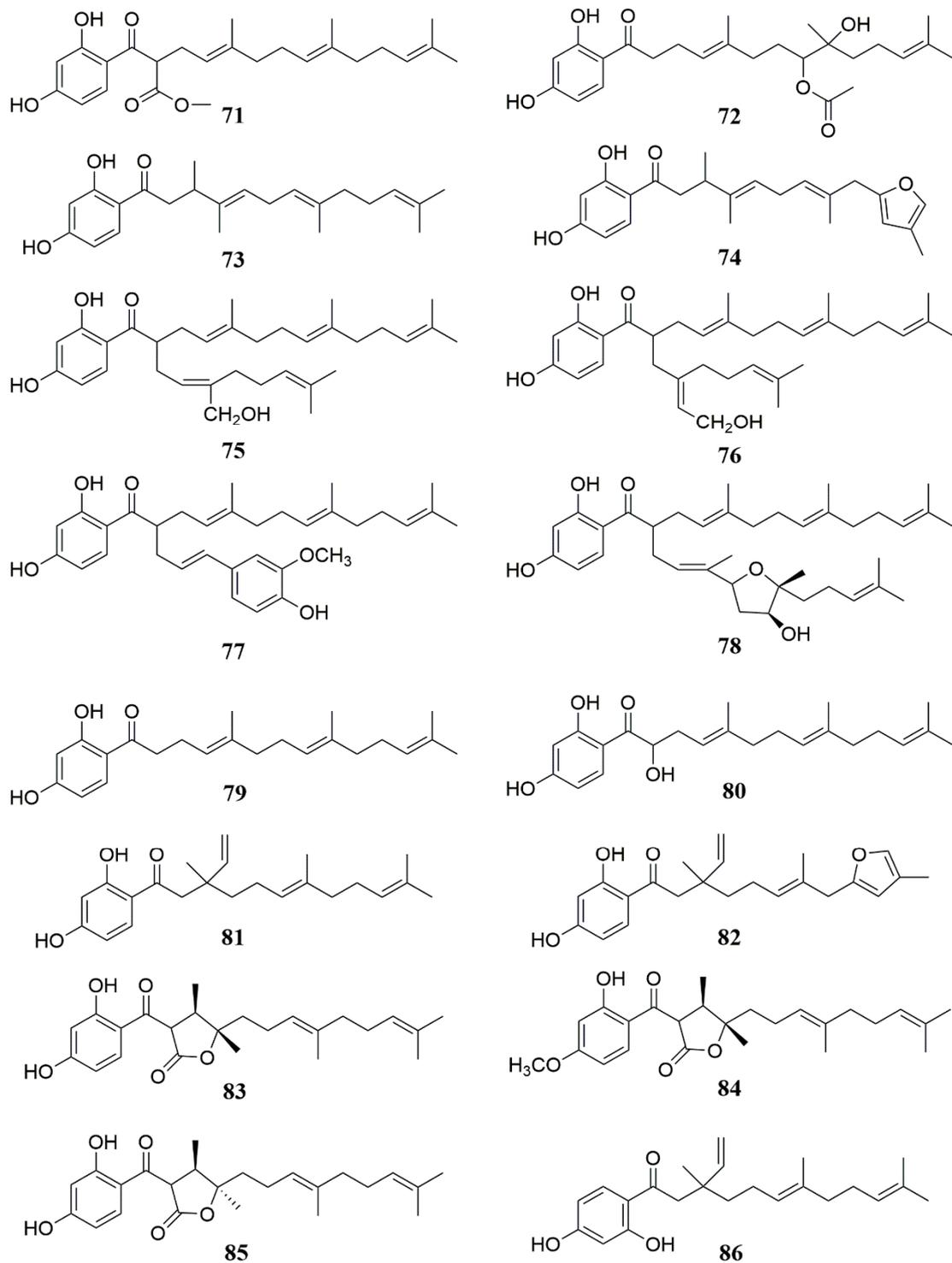


Figure 5. Chemical structures of compounds 71–86 from *Ferula* plants.

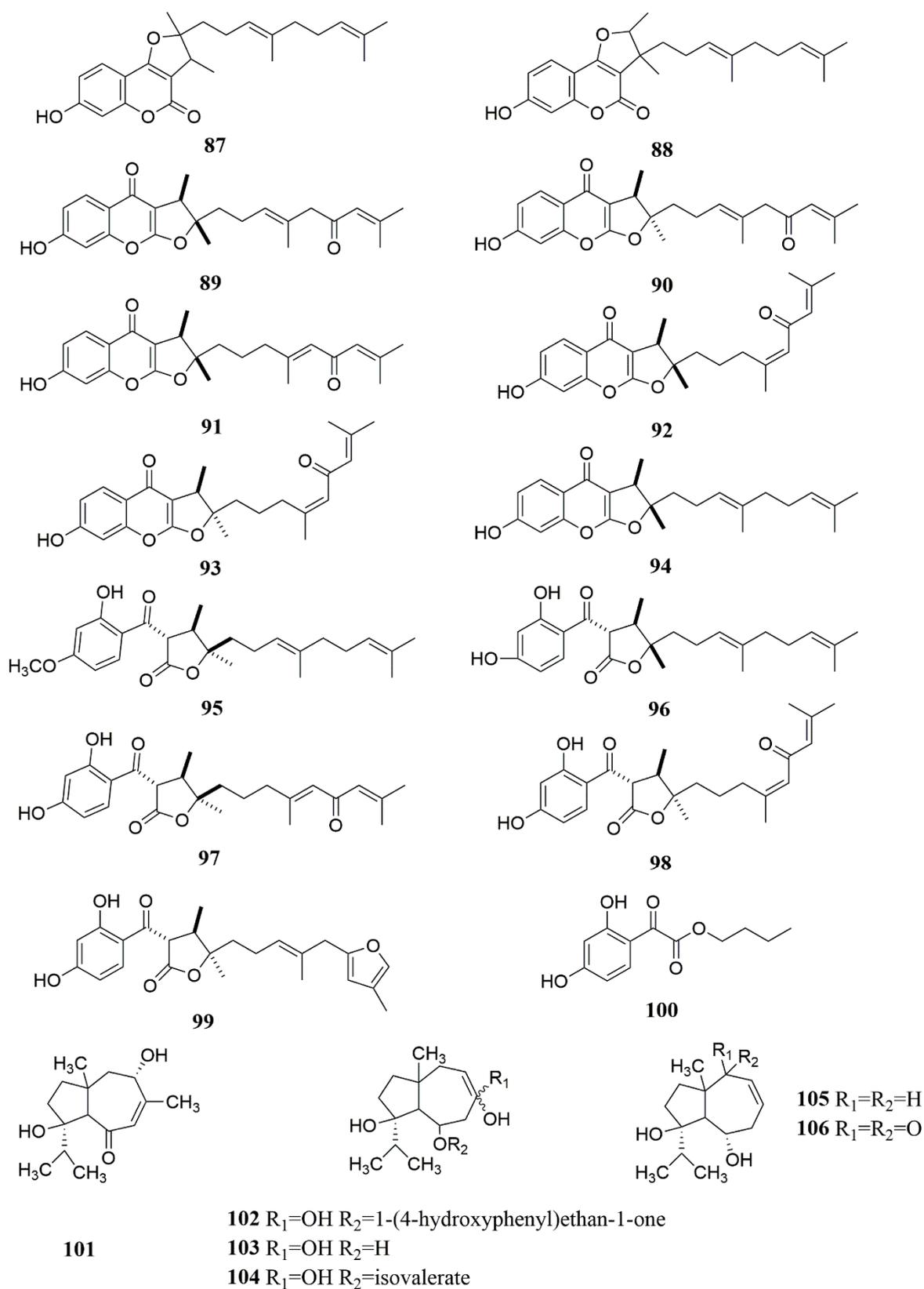


Figure 6. Chemical structures of compounds 87–106 from *Ferula* plants.

3.3. Volatile Oil

The volatile oils in genus *Ferula* are mainly terpenoids and polysulfide compounds [72]. Among the genus *Ferula* of China, the number of terpene components can reach more

than 80% in volatile oils, mostly are monoterpenes and sesquiterpenes, while polysulfide compounds are dominated by disulfides, trisulfides, bis-disulfides, and thio-disulfides. A large amount of volatile oil was extracted from *F. sinkiangensis*, *F. fukanensis*, *F. ferulaeoides*, and *F. ovina* [73–76] (Table 4). Min Zhi-da et al. [77] identified 26 polysulfides by gas chromatography-mass spectrometry (GC-MS (CI/EI)) in *F. sinkiangensis* and *F. fukanensis* (Figure 7).

Table 4. The composition of volatile oils of *F. sinkiangensis*, *F. fukanensis*, *F. ferulaeoides*, *F. ovina*.

No	Compound	Molecular Formula	Literature
107	(1S)- β -Pinene	C ₁₀ H ₁₆	[73]
108	1,7,7-Trimethyl-tricyclo[2.2.1.0 (2,6)] heptane	C ₁₀ H ₁₆	[73]
109	(1R)- α -Pinene	C ₁₀ H ₁₆	[73]
110	Camphene	C ₁₀ H ₁₆	[73]
111	6-Methyl-5-hepten-2-one	C ₈ H ₁₄ O	[73]
112	β -Pinene	C ₁₀ H ₁₆	[73]
113	α -Phellandrene	C ₁₀ H ₁₆	[73]
114	3-Carene	C ₁₀ H ₁₆	[73]
115	Terpinolene	C ₁₀ H ₁₆	[73]
116	1-Isopropyl-2-methylbenzene	C ₁₀ H ₁₆	[73]
117	D-Limonene	C ₁₀ H ₁₆	[73]
118	(E)-3,7-Dimethyl-1,3,6-octatriene	C ₁₀ H ₁₆	[73]
119	(Z)-3,7-Dimethyl-1,3,6-octatriene	C ₁₀ H ₁₆	[73]
120	γ -Terpinene	C ₁₀ H ₁₆	[73]
121	L-Camphor	C ₁₀ H ₁₆ O	[73]
122	Borneol	C ₁₀ H ₁₆ O	[73]
123	(-)- α -Copaene	C ₁₅ H ₂₄	[73]
124	2-Methoxy-4-methyl-1-(1-methylethyl)-benzene	C ₁₁ H ₁₆ O ₂	[73]
125	1-Methoxy-4-methyl-2-(1-methylethyl)-benzene	C ₁₁ H ₁₆ O	[73]
126	L-Bornyl acetate	C ₁₂ H ₂₀ O ₂	[73]
127	Lavandulol acetate	C ₁₂ H ₂₀ O ₂	[73]
128	(+)- α -Longipinene	C ₁₅ H ₂₄	[73]
129	(Z)-2,6,10-Trimethyl-1,5,9-undecatriene	C ₁₄ H ₂₄	[73]
130	Caryophyllene	C ₁₅ H ₂₄	[73]
131	(1S,2S,4R)-2-Acetate-1,3,3-trimethyl-bicyclo[2.2.1] hepten-2-ol	C ₁₂ H ₂₀ O ₂	[73]
132	1-Ethenyl-1-methyl-2-(1-methylethenyl)-4-(1-methylethylidene)-cyclohexane	C ₁₅ H ₂₄	[73]
133	1S-(1 α ,4 α ,7 α)-1,2,3,4,5,6,7,8-Octahydro-1,4-dimethyl-7-(1-methylethylidene)-azule	C ₁₅ H ₂₄	[73]
134	(4 α S,9 α R)-2,4 α ,5,6,7,8,9,9 α -Octahydro-3,5,5-trimethyl-9-methylidene-benzocycloheptene	C ₁₅ H ₂₄	[73]
135	(3Z,6E)-3,7,11-Trimethyl-1,3,6,10-dodecanetetraene	C ₁₅ H ₂₄	[73]
136	α -Farnesene	C ₁₅ H ₂₄	[73]
137	Isolongiflone-8-ol	C ₁₅ H ₂₆ O	[73]
138	Tran-nerolidol	C ₁₅ H ₂₆ O	[73]
139	Guaiol	C ₁₅ H ₂₆ O	[73]
140	α -Eudesmol	C ₁₅ H ₂₆ O	[73]
141	(2R,4 α R)-1,2,3,4,4 α ,5,6,7-Octahydro- α , α -3,8-tetramethyl-naphthalenemethanol	C ₁₅ H ₂₆ O	[73]
142	[3S-(3 α ,3 α β ,5 α)]-1,2,3,3 α ,4,5,6,7-Octahydro- α , α -3,8-tetramethyl-5-azulenemethanol	C ₁₅ H ₂₆ O	[73]

Table 4. Cont.

No	Compound	Molecular Formula	Literature
143	(1S)-1-[(1S)1,5-Dimethyl-4-hexen-1-4-methyl-3-cyclohexen-1-ol	C ₁₅ H ₂₆ O	[73]
144	Isoledene	C ₁₅ H ₂₄	[73]
145	Ethanol	C ₂ H ₆ O	[74]
146	Ethyl ester	C ₄ H ₈ O ₂	[74]
147	3-Methyl-butanal	C ₅ H ₁₀ O	[74]
148	2-Butanethiol	C ₄ H ₁₀ S	[74]
149	2,5-Dimethyl-furan	C ₆ H ₈ O	[74]
150	Furfural	C ₅ H ₄ O ₂	[74]
151	Methyl 1-propenyl disulfide	C ₄ H ₈ S ₂	[74]
152	Dimethyl trisulfide	C ₂ H ₆ S ₃	[74]
153	<i>n</i> -Propyl <i>sec</i> -butyl disulfide	C ₇ H ₁₆ S ₂	[74]
154	Propyl <i>n</i> -butyl disulfide	C ₇ H ₁₆ S ₂	[74]
155	1,2-Dithiacyclopentane	C ₃ H ₆ O ₂	[74]
156	2-Ethyl-hexanethiol	C ₈ H ₁₈ S	[74]
157	<i>bis</i> (1-Methylpropyl)-disulfide	C ₈ H ₁₈ S ₂	[74]
158	2,2'-Bioxirane	C ₄ H ₆ O ₂	[74]
159	3-Methyl-4-heptanol	C ₈ H ₁₈ O	[74]
160	1,1- <i>bis</i> (Methylthio)-ethane	C ₄ H ₁₀ O ₂	[74]
161	1,1-Dimethoxy-propane	C ₅ H ₁₂ O ₂	[74]
162	Heptadecane	C ₁₇ H ₃₆	[74]
163	2-Ethylthio-butane	C ₆ H ₁₄ S	[74]
164	α -Humulene	C ₁₅ H ₂₄	[74]
165	β -Selinene	C ₁₅ H ₂₄	[74]
166	Hexadecane	C ₁₆ H ₃₂	[74]
167	2-Methylthio butyrate methyl ester	C ₆ H ₁₂ OS	[74]
168	2,3-Dimethyl-3-hexanol	C ₈ H ₁₈ O	[74]
169	Hedycaryol	C ₁₅ H ₂₆ O	[74]
170	2,2- <i>bis</i> (Methylthio)-propane	C ₅ H ₁₂ S ₂	[74]
171	Octadecatriene	C ₁₈ H ₃₂	[74]
172	Palmitic acid	C ₁₆ H ₃₂ O ₂	[74]
173	Oleic acid	C ₁₈ H ₃₄ O ₂	[74]
174	α -Myrcene	C ₁₀ H ₁₆	[75]
175	<i>o</i> -Cymene	C ₁₀ H ₁₄	[75]
176	Enol	C ₁₀ H ₁₆ O	[75]
177	Limonene oxide	C ₁₀ H ₁₆ O	[75]
178	Camphor	C ₁₀ H ₁₆ O	[75]
179	Thymol	C ₁₀ H ₁₄ O	[75]
180	α -Terpineol	C ₁₀ H ₁₈ O	[75]
181	Anisyl acetate	C ₁₂ H ₂₀ O ₂	[75]
182	Carvacrol acetate	C ₁₀ H ₁₆ O	[75]
183	1-Methoxy-4-methyl-2-(1-ethyl)-benzene	C ₁₁ H ₁₆ O	[75]
184	Carvone	C ₁₀ H ₁₄ O	[75]
185	Bornyl acetate	C ₁₂ H ₂₀ O ₂	[75]
186	(-)-Verbenone	C ₁₀ H ₁₆ O	[75]
187	α -(E)-BETA-FARNESENE	C ₁₅ H ₂₄	[75]
188	1-Cycloethyl-1-pentyne	C ₁₁ H ₁₈	[75]
189	α -Bergamotene	C ₁₅ H ₂₄	[75]
190	1,11-Hexadecadiyne	C ₁₆ H ₂₆	[75]
191	α -Bisabolene	C ₁₅ H ₂₄	[75]
192	Asarone	C ₁₂ H ₁₆ O ₃	[75]
193	3,7,11-trimethyl(E)1,6,10-dodecatrien-3-ol	C ₁₅ H ₂₆ O	[75]
194	Farnesol	C ₁₅ H ₂₆ O	[75]
195	Graphene oxide	C ₁₅ H ₂₄ O	[75]
196	Disulfide bis(1-methylpropyl)	C ₈ H ₁₈ S ₂	[75]
197	<i>n</i> -Propyl <i>sec</i> -butyl disulfide	C ₇ H ₁₆ S ₂	[76]
198	Ocimene	C ₁₀ H ₁₆	[76]

Table 4. Cont.

No	Compound	Molecular Formula	Literature
199	Ocimene (Mixture of isomers)	C ₁₀ H ₁₆	[76]
200	Disulfide, bis[1-(methylthio)ethyl]	C ₆ H ₁₄ S ₄	[76]
201	2-Methylbutyl benzene	C ₁₁ H ₁₆	[76]
202	Ethyl 1-methylpropyl disulfide	C ₆ H ₁₄ S ₂	[76]
203	(Z)-1,6,10-Dodecatriene-7,11-dimethyl-3-methylene	C ₁₅ H ₂₄	[76]
204	cis- α -Bisabolene	C ₁₅ H ₂₄	[76]
205	Dipropyl disulfide	C ₆ H ₁₄ S ₂	[76]
206	Hinesol	C ₁₅ H ₂₆ O	[76]
207	Neoisolongifolene	C ₁₅ H ₂₂	[76]
208	Bicyclo[4.4.0]dec-1-ene,2-isopropyl-5-methyl-9-methylene	C ₁₅ H ₂₄	[76]
209	2H-Pyran, tetrahydro-4-methyl-2-(2-methyl-1-propenyl)	C ₁₀ H ₁₈ O	[76]
210	2,4,6-Octatriene, 2,6-dimethyl-, (E,Z)-	C ₁₀ H ₁₆	[76]
211	Estragole	C ₁₀ H ₁₂ O	[76]
212	1-Methoxy-4-methyl-2-(1-methylethyl) Benzene	C ₁₁ H ₁₆ O	[76]
213	2,6-Dimethyl-2,6-octadiene	C ₁₀ H ₁₈	[76]
214	Methyl 2-(methylthio)butyrate	C ₆ H ₁₂ O ₂ S	[76]
215	1-Methyl-4-(1-methylethyl)-1,3-cyclohexadiene,	C ₁₀ H ₁₆	[76]
216	Methyl sec-butyl disulphide	C ₆ H ₁₂ O ₂ S	[76]
217	3-(Methylthio)-2-butanone	C ₅ H ₁₀ OS	[76]
218	Naphthalene,1,2,3,4,4a,5,6,8a-octahydro-7-methyl-4-methylene-1-(1-methyl-ethyl)-, (1 α ,4 α β ,8 α)-	C ₁₅ H ₂₂	[76]
219	1,4-Methanoazulene, decahydro-4,8,8-trimethyl-9-methylene-, [1S-(1.a.,3a. β ,4a.,8a. β)]-	C ₁₅ H ₂₆ O	[76]
220	Tetrahydro thiazole (4aS-cis)-2,4a,5,6,7,8,9,9a-octahydro-	C ₅ H ₁₂ S ₂	[76]
221	3,5,5-trimethyl-9-methylene -1H-Benzocycloheptene	C ₅ H ₁₀ OS	[76]
222	Thiopropionamide	C ₃ H ₇ NS	[76]
223	Longifolene-(V4)	C ₁₅ H ₂₄	[76]
224	Naphthalene, 1,2,3,4,4a,5,6,8a-octahydro -4a,8-dimethyl-2-(1-methyl ethenyl)-, [2R-(2 α ,4 α ,8 α β)]	C ₁₅ H ₂₄	[76]
225	Seychellene	C ₃ H ₇ NS	[76]
226	(R)-2,4a,5,6,7,8-hexahydro-3,5,5,9-tetramethyl-1H-benzocycloheptene	C ₁₅ H ₂₄	[76]
227	[S-(Z)]-3,7,11-trimethyl-1,6,10-Dodecatrien-3-ol	C ₁₅ H ₂₄	[76]
228	E-Famesene epoxide	C ₃ H ₇ NS	[76]
229	Di-epi- α -cedrene	C ₁₅ H ₂₄	[76]

3.4. Aromatic Compounds

Studies showed that ethyl p-hydroxybenzoate (**256**), emodin (**257**), and 1,3,7-trihydroxy-6-methyl-xanthone (**258**) were isolated from *F. fukanensis* [78]. Ferulic acid (**259**) was isolated for the first time from *F. sinkiangensis* [62]. 4,5-Dimethoxy-2,3-methylenedioxyphenylpropane-7-adamate was isolated from *F. licentiana* [79] and named as Tong Shan ferulic acid A (**260**). 2,4-Dihydroxy- α -oxo-phenylacetic acid (**261**), 3,3',4,4'-biphenyltetracarboxylic acid (**262**) and 2,4-dihydroxybenzophenone (**263**) were isolated from the rhizome of *F. songarica* [80]. 2,4-Dihydroxy- α -oxo-phenylacetic acid was first isolated from *F. songarica* and 3,3',4,4'-biphenyltetracarboxylic acid was a newly discovered compound (Figure 8).

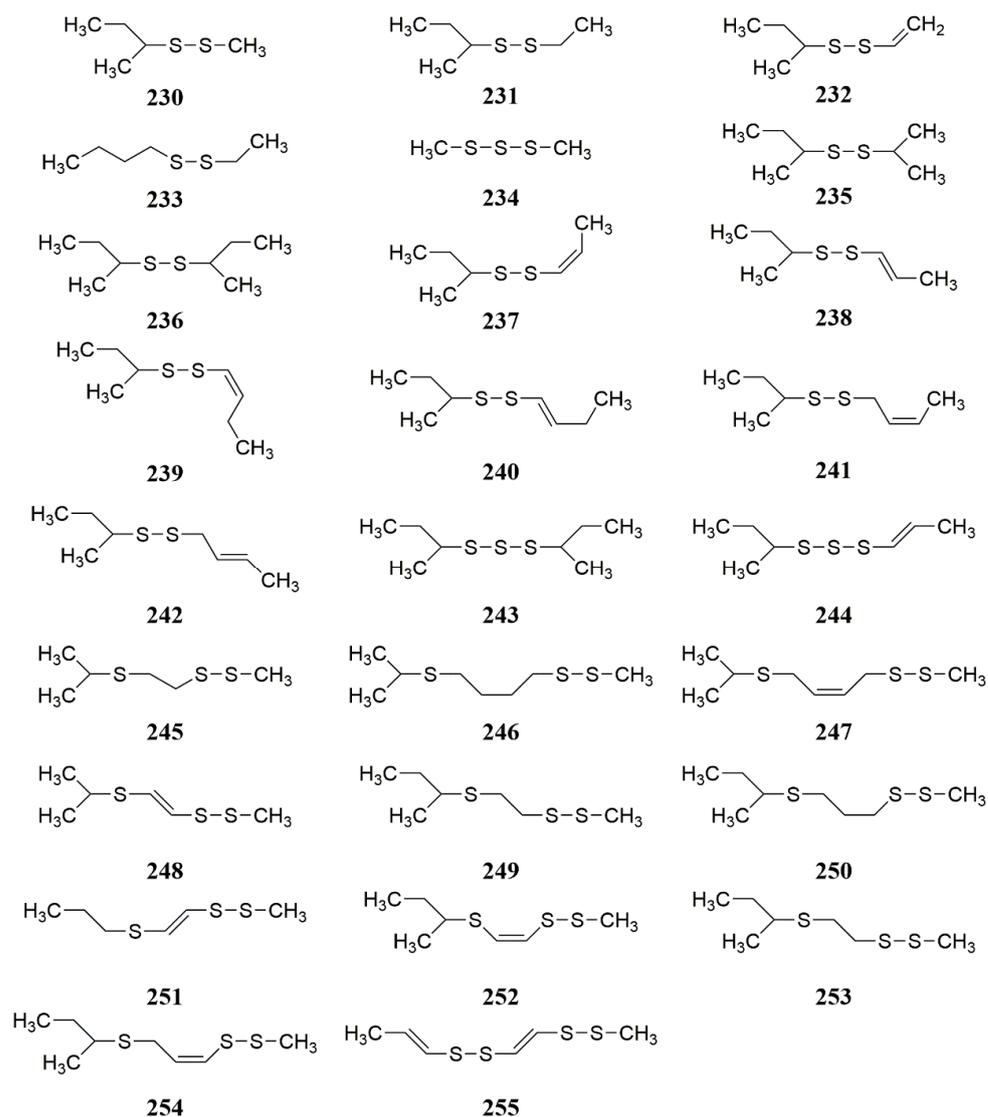


Figure 7. Chemical structures of compounds 230–255 from *Ferula* plants.

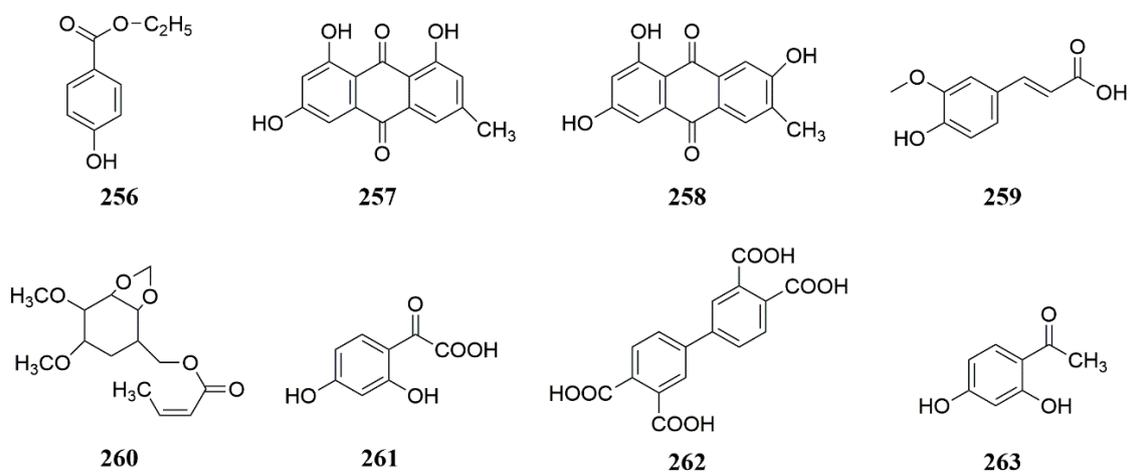


Figure 8. Chemical structures of compounds 256–263 from *Ferula* plants.

3.5. Other Substances

Veratric acid (**264**) and β -sitosterol (**265**) were isolated from *F. licentiana* [81]. Acetophenone compounds such as (1*S*,2*R*,4*S*)-(–)-borneol acetate (**266**), (5*Z*)-2,6,10-trimethyl-1,5,9-undecatriene (**267**), (*Z*)- β -farnesene (**268**), humulen-(v1) (**269**), \pm -trans-nerolidol (**270**), and carotene (**271**) were isolated from *F. ferulaeoides* [82,83] (Figure 9). Seventeen amino acids, including lysine, histidine, arginine, aspartic acid and threonine, were obtained from the roots and leaves of *F. lehmannii* Boiss [84]. Studies have shown that a series of trace elements including potassium, aluminum, calcium, copper, magnesium, barium, cadmium, and cobalt were isolated from *F. sinkiangensis* [85]. Radiatinol (**272**) and scopoletin (**273**) were isolated from *F. dissecta* [86].

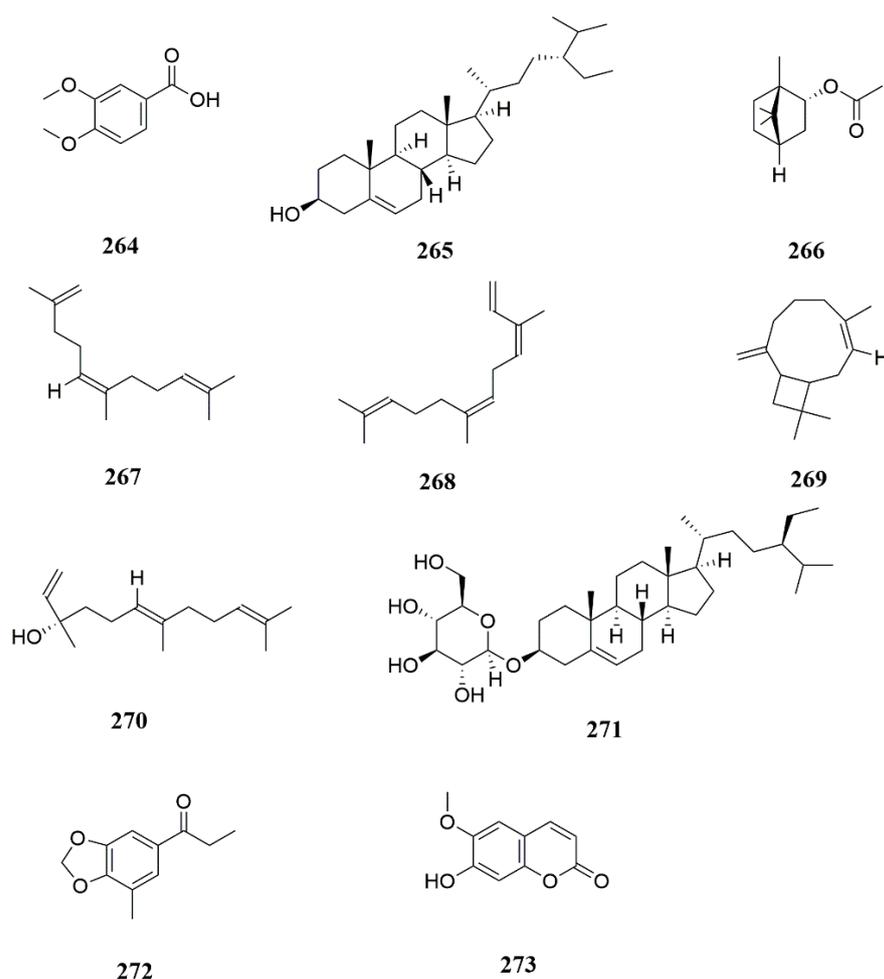


Figure 9. Chemical structures of compounds 264–273.

4. Advances in the Bioactivity of *Ferula* Plants in China

With the gradual increase in domestic research in the field of biological activity of the genus *Ferula*, the bioactive compounds were isolated from the *Ferula* plant by various applied extraction techniques [87], and a number of different compounds with anti-tumor [88], anti-allergy, anticoagulation [89], bacterial inhibition [90], action in the gastrointestinal system [91], nervous system [92], cardiovascular system [93,94], and other pharmacological activities have been found in the genus *Ferula* [95–97].

4.1. Anticancer Activity

The main chemical components of *Ferula* plants that exert anti-tumor effects are ferulic acids [98], sesquiterpenoids [99], and volatile oils [100]. For instance, Wu Jing [101] found that the inhibition rate of A549 lung cancer xenografts in nude mice gradually increased

with increase in dose of ferulic acid administration. The low (25 mg/L), medium (50 mg/L) and high (100 mg/L) dose groups inhibited the expression levels of both mTOR protein and mRNA in the tumor tissues of the nude mice, and decreased the expression levels of Ki-67 protein, which decreased with increasing dose, and also enhanced the expression levels of Caspase-3 protein. The investigators suggested that ferulic acid can attenuate growth of lung cancer probably by downregulating mTOR protein expression, inhibiting Ki-67 expression and promoting Caspase-3 levels. The petroleum ether, n-butanol, ethyl acetate, trichloromethane, and aqueous fractions, as well as volatile oil and 95% ethanol extract of *F. ferulaeoides* were screened for their pharmacological activities, among which trichloromethane and ethyl acetate fractions exhibited certain inhibitory effects on five distinct gastric cancer cell lines, whereas trichloromethane fractions showed the strongest inhibitory effect on gastric cancer cells SGC-7901 (IC_{50} : 7.98 ± 2.62 mg/L). In addition, volatile oil displayed strong inhibition of cell proliferation on gastric cancer cells AGS (IC_{50} : 8.73 ± 0.55 mg/L), but n-butanol fraction and aqueous fraction showed no significant inhibitory effects on these five gastric cancer cell lines (AGS, MKN-45, BGC-823, MGC-803, SGC-7901) [102]. A sesquiterpene coumarin-like substance, 2,3-dihydro-7-hydroxy-2R*,3R*-dimethyl-2-[4,8-dimethyl-3(E),7-nonadienyl]-furo[3,2-c]coumarin (DAW22), was isolated from *F. ferulaeoides* root, and was subsequently reported to induce apoptosis in C6 glioma cells through the mitochondria-mediated, endoplasmic reticulum stress and death receptor pathway. The dosing concentration is 20 μ M [103]. It also suppressed the proliferation of five different established human Malignant peripheral nerve sheath tumors (MPNST) cancer cell lines (STS-26T, S462, S462-TY, ST8814, T265) by promoting their apoptosis [104]. Moreover, experimental animal studies have revealed that in an immunocompromised nude mouse model with subcutaneous transplantation of STS-26T cells, when DAW22 was administered intraperitoneally at a daily dose of 60 mg/kg/day for 4 weeks, it significantly inhibited tumor xenografts compared to the control group. It was found by Western blotting analysis, that DAW22 inhibited phosphorylation of AKT and ERK and reduced the level of Non-phospho (Active) CTNNB1, thereby resulting in an inhibitory effect on cell proliferation and suppression of tumor growth in for MPNST cell lines. These results supported the use of DAW22 as an alternative therapeutic compound to MPNST which can effectively produce anticancer effects by affecting multiple signaling pathways in tumor disease progression. 8-*p*-Hydroxybenzoyl tovarol (TAW) was isolated from the roots of *F. dissecta*, and found to inhibit the growth of human cervical cancer HeLa cells by promoting protective autophagy, and in the future, advanced molecular dynamics techniques and free energy calculations and reverse molecular docking can be used to reveal the effects of the major coumarins, sesquiterpenoids, and terpenoids in the genus *Ferula* on the anticancer mechanism, and there are already documented studies of other plants using similar measurement techniques [105,106].

4.2. Antibacterial Activity

Extraction studies on three distinct kinds of dry roots of *F. sinkiangensis*, *F. lehmannii* Boiss. and *F. ferulaeoides* revealed that the alcoholic and macerated extracts displayed antibacterial effects on *Staphylococcus aureus*, *Bacillus subtilis*, and *Sporosarcina*. The MIC about *F. sinkiangensis* root's alcoholic extracts for three bacteria were 31.25 mg/mL, 3.92 mg/mL, and 1000 mg/mL. The MIC about *F. sinkiangensis* root's macerated extracts for three bacteria were 1000 mg/mL, 1000 mg/mL, and 1000 mg/mL. The MIC about *F. ferulaeoides* root's alcoholic extracts for three bacteria were 3.91 mg/mL, 3.91 mg/mL, and 15.63 mg/mL. The MIC about *F. ferulaeoides* root's macerated extracts for three bacteria were 500 mg/mL, 500 mg/mL, and 1000 mg/mL. The MIC about *F. ferulaeoides* root's alcoholic extracts for three bacteria were 7.81 mg/mL, 3.91 mg/mL, and 15.63 mg/mL [107]. A total of 27 compounds were extracted and isolated from *F. ferulaeoides*, and 7 of the 27 compounds were found in the plant for the first time. These 27 compounds were screened for their potential antibacterial activity, and only a few coumarins among the 27 compounds were found to affect multi-drug resistant (MDR) and methicillin-resistant *Staphylococcus aureus* (MRSA)

bacteria, and six of them showed inhibition against tetracycline-resistant strain XU212 [108]. In addition, three new sesquiterpene derivatives were obtained from *F. ferulaeoides* [109] and it was observed that their minimum inhibitory concentrations (MIC) ranged from 0.5–128 µg/mL. It was found that the volatile oil of the seeds of *F. olivacea* exhibited an inhibitory effect on the Gram-positive bacteria such as *Enterococcus faecalis* and *S. aureus* [110,111].

4.3. Anti-Allergic, Anti-Inflammatory, and Immunosuppressive Effects

Using sodium cromoglycate (10 mg/kg) as the control group and volatile oil of *F. sinkiangensis* (50 mg/kg) as the experimental group, experiments were conducted to evaluate the potential effect of passive skin allergic reaction in mice, the effect of Arthus reaction in rabbits, and the antigen sensitization confrontation in the sensitized guinea pigs [112]. It was found that the *F. sinkiangensis* volatile oil (200 mg/kg) could significantly inhibit Ig E response to allergic reactions, and the degree of effect was similar to that of sodium cromoglycate. Ferulic acid can also reverse skin sensitization in rats and suppress the delayed hypersensitivity reactions in mice, thus indicating that ferulic acid has inhibitory effect on allergic reactions [113]. In another study, designed to examine the impact of the raw juices of *F. sinkiangensis*, *F. conocaula*, and *F. feruloides*, it was found that all three types of *Ferula* plants exhibited significant inhibitory effects on foot topography swelling (the dosing concentrations were *F. sinkiangensis*: 90 mg/kg, *F. conocaula*: 90 mg/kg, *F. feruloides*: 80 mg/kg) and capillary permeability (the dosing concentrations were *F. sinkiangensis*: 34 mg/kg, *F. conocaula*: 34 mg/kg, *F. feruloides*: 30 mg/kg) in rabbits induced by p-hornwort, and on the delayed hypersensitivity (the dosing concentrations were *F. sinkiangensis*: 98 mg/kg, *F. conocaula*: 97 mg/kg, *F. feruloides*: 78 mg/kg) and serum hemolysin formation (the dosing concentrations were *F. sinkiangensis*: 98 mg/kg, *F. conocaula*: 97 mg/kg, *F. feruloides*: 78 mg/kg) in mice induced by sheep erythrocytes or dinitrochlorobenzene [114].

4.4. Anticoagulant Effect

The extract of *F. lehmannii* was obtained by using 95% ethanol, and an anticoagulant assay was then performed. The experimental results revealed that *F. lehmannii* extract inhibited the endogenous coagulation pathway and showed an inhibitory effect on the activity of thrombinogen. The dosing concentrations were: the high-dose group (100 mg/kg), the medium-dose group (50 mg/kg), and the low-dose group (25 mg/kg) [115]. Radiatinol and scopoletin were isolated from *F. dissecta*, and both these compounds acted in the endogenous coagulation pathway and prolonged the prothrombin time as well as activated partial thromboplastin time at different concentrations [86].

4.5. Effects on the Cardiovascular System

Interestingly, another study showed that after ferulic acid was administered to C57 mice in the heart failure preclinical model, the mice in the heart failure administration group exhibited different degrees of improvement in cardiac left ventricular ejection fraction, end-systolic and end-diastolic internal diameter changes, collagen area percentage changes, and protein expression levels compared to the control mice. Ferulic acid could improve ventricular remodeling to some extent [116]. In addition, studies conducted with the hydroalcoholic extracted fractions and the aqueous decoction of *F. sinkiangensis* revealed that they could reduce the heart amplitude and enhance the heart rate in isolated frog hearts [117].

4.6. Effects on the Gastrointestinal Tract

It has been reported in the literature that *F. sinkiangensis* has been used to treat stomach disorders in Xinjiang [118]. The volatile oil and resin of *F. sinkiangensis* were found to be effective in three distinct gastric ulcer models, among which the volatile oil was found to be relatively better [119]. The rat model of acetic acid-injected gastric ulcer was established, and the model was tested by administering *F. sinkiangensis* original herbs and *F. sinkiangensis* prepared herbs by a folk concoction (0.048 g/mL), frying method (0.048 g/mL), a

vinegar moxibustion method (0.048 g/mL), and a boiling method (0.048 g/mL). It was found that the pH value of the rat stomach increased significantly, but the area of gastric ulcer decreased markedly after the administration of *F. sinkiangensis* original herbs and *F. sinkiangensis* prepared herbs [120]. In addition, studies on the raw juices of *F. feruloides* and *F. sinkiangensis* revealed that these juices can display inhibitory effects on gastric ulcer models in rats with acetylsalicylic acid and can effectively suppress the autonomous activity of smooth muscle in isolated intestinal tubes of rabbits [121]. The compounds were isolated from the *F. sinkiangensis* resin and they were analyzed in anti-ulcer vivo activity experiments. It was observed that compared with the positive control group famotidine, CHCl₃ extract (1.3515 g/kg) showed the best anti-ulcer activity [122].

4.7. Action on the Nervous System

A network pharmacological analysis of *Ferula* resulted in identification of 12 key active components against Alzheimer's disease, including Farnesiferol A, Farnesiferol B, conferol, and ferulic acid. It was predicted that *Ferula* can predominantly act through by modulating 14 key targets in the cholinergic synaptic signaling pathway and AD signaling pathway [123]. A bioactivity-oriented study on *F. sinkiangensis* showed that *F. sinkiangensis* significantly inhibited NO production induced by over-activated microglia and exhibited anti-neuroinflammatory effects [58]. Interestingly, a compound, Kellerin, extracted from *F. sinkiangensis*, was found to produce substantial effects in the rat middle cerebral artery occlusion (MCAO) model, lipopolysaccharide (LPS)-activated microglia model, mouse bilateral common carotid artery occlusion (BCCAO) model and lipopolysaccharide (LPS)-activated microglia model. Kellerin was found to markedly improve the neurological outcomes, reduce the size of cerebral infarcts and decrease brain edema in the rat MCAO model, and the dosing concentrations were: the low-dose group (3.5 mg/kg), the medium-dose group (7.0 mg/kg) and the high-dose group (14.0 mg/kg). Moreover, in the pathological conditions of focal cerebral ischemia, Kellerin could attenuate neuronal damage as well as microglial activation. In addition, in vitro study which LPS-stimulated BV2 cells, revealed that Kellerin protected the neuronal cells from injury by inhibiting microglia activation. Kellerin also reduced levels of pro-inflammatory cytokines, inhibited the NF- κ B signaling pathway, and decreased ROS production as well as NADPH oxidase activity, and the [124]. Kellerin was found to alleviate the cognitive impairment, reduce neuronal loss, inhibit microglia activation, and convert microglia from a pro-inflammatory M1 phenotype to an anti-inflammatory M2 phenotype in BCCAO mice. In addition, in another in vitro study, Kellerin was found to modulate the microglia polarization and inhibit NLRP3 and MAPK signaling pathways after LPS treatment [125].

4.8. Other Pharmacological Effects

Furthermore, studies have shown that *Ferula* resin can exhibit anti-pregnancy, anti-implantation and pregnancy termination effects [126,127]. It can also affect estrogen and progesterone levels in infertile mice and rabbits, thus modulating the smooth muscles of the uterus in mice and rabbits [128]. In another study, D-galactosamine/lipopolysaccharide acute liver injury model was established, and by comparing the D-galactosamine/lipopolysaccharide model group, the biphenyldiglyceride group and the high-, medium- and low-dose ferulic acid groups, it was observed that all the three ferulic acid groups could significantly decrease the activity of enzymes AST and ALT in mouse hepatocytes, increase the activity of antioxidant enzyme SOD as well as peroxidase GSH-Px in mice, and decrease the MDA content, thereby suggesting that ferulic acid exhibited a protective effect on acute liver injury [129]. The petroleum ether, ethyl acetate, n-butanol and methanol fractions of *F. sinkiangensis* displayed no substantial effect on the lipase activity, whereas the ethyl acetate, n-butanol and methanol fractions lowered total cholesterol in HepG-2 cells and triglycerides in non-alcoholic fatty liver model cells. Thus, it was hypothesized that *F. sinkiangensis* has certain lipid-regulating effects [130]. Four coumarin-like compounds were extracted from *F. moschata* showed anti-HIV activity [131].

4.9. Toxicological Studies

Acute toxicity experiments were conducted on the volatile oils of *F. feruloides* and *F. teterrima*, and the findings revealed that the LD50 of the volatile oil of *F. feruloides* was 10.24 g/kg and the LD50 of the volatile oil of *F. teterrima* was 491.61 mg/kg. After analyzing the volatile oils of both, it was found that the volatile oil of *F. teterrima* contained more polysulfides, accounting for 60% of the volatile oil, but no polysulfides were detected in the volatile oil of *F. feruloides*. Polysulfide can cause irritation and possess a garlic-like odor, so it was presumed that polysulfide was more likely to be a toxic component in *F. teterrima* [132]. However, the acute toxicity studies with polysulfide has not been reported previously in the literature. It was found that goats fed with *F. fukanensis* (2.5 g/kg) for 15 days showed obvious signs of toxicity, such as loss of appetite, anorexia, diarrhea, vitamin K-dependent reduction in coagulation factors, the toxic component of which may be possibly coumarins, but no obvious symptoms were found in these animals, such as impaired liver and platelet function [133]. Acute toxicity tests were conducted on *F. sinkiangensis* and *F. fukanensis* [134], and the LD50 values were calculated by Kou's method. The LD50 values of *F. sinkiangensis* volatile oil aqueous suspension and *F. fukanensi* volatile oil aqueous suspension were found to be 2.82 g/kg and 1.55 g/kg; and the LD50 values of *F. sinkiangensis* volatile oil emulsion and *F. fukanensi* volatile oil emulsion were separately 0.39 g/kg and 0.41 g/kg⁻¹. After analyzing ten batches of *F. sinkiangensis* test samples [135], two of them were observed to contain different ferulic acid contents. No. 1 *F. sinkiangensis* with (0.775 ± 1.44) mg/g and No. 5 *F. sinkiangensis* with (0.279 ± 1.63) mg/g, were selected and subjected to the toxicity studies by SRB method. The proliferation rates of No. 1 *F. sinkiangensis* and No. 5 *F. sinkiangensis* on the rat renal NRK cells was found to gradually increase with increasing drug concentrations, with IC50 values of No. 1 *F. sinkiangensis* at 60.60 mg/L and No. 5 *F. sinkiangensis* at 39.90 mg/L. The comparison of the IC50 values showed that the cytotoxicity of No. 1 *F. sinkiangensis* was significantly less than that of No. 5 *F. sinkiangensis*, but the ferulic acid content of No. 1 *F. sinkiangensis* was significantly higher than that of No. 5 *F. sinkiangensis*, thus indicating that the toxicity of *F. sinkiangensis* may be possibly related to its chemical composition.

5. Quality Marker (Q-Marker) Prediction Analysis

With the depletion of the domestic *Ferula* plant resources, the phenomenon of sub-standard and uneven quality of *Ferula* plants in the market needs to be urgently solved. Thus, the quality markers (Q-marker) [136] need be established for the effective protection of the domestic *Ferula* plant resources. At present, the Chinese Pharmacopoeia 2020 edition stipulates that the sources of medicinal *Ferula* plant is the resin of *F. sinkiangensis* and *F. fukanensi* and requires that their quality marker is ferulic acid. However, ferulic acid is widely present in different plants such as *Angelica sinensis* [137], *Ligusticum chuanxiong* [138], *Cimicifugafoetida* L. [139], *Ligusticum* [140] and *Fructus Toosendan* [141]. It is also an important active component of many traditional Chinese medicines [142]. A large amount of sesquiterpene coumarins are found in *Ferula* plants, and they may be used as unique chemical components which can effectively help to identify *Ferula* plants by the Q-marker method.

5.1. Q-Marker Prediction Analysis by Kinship and Chemical Composition Specificity of *Ferula* Plants

The global distribution of *Ferula* plants is widespread in Central Asia, Iran and Pakistan. It has been reported that during the course of foreign research on the chemical composition of *Ferula* plants, it was observed that genus *Ferula* contained a large number of sesquiterpene coumarins with umbelliferous lactones as the parent nucleus [143], and Ferulenol was the most abundant and the first isolated coumarin from *Ferula* [144]. Although various sesquiterpenes and coumarins are widely distributed in the Apiaceae family, they constitute the characteristic chemical constituents of genus *Ferula*, and can be used as important evidence of the affinities of *Ferula* plants. They could also form a sound

basis for the chemical taxonomy of genus *Ferula* [145], such as Farnesiferol A, Farnesiferol B, Farnesiferol C, and DAW22, because sesquiterpene coumarins are widespread in different plants of the genus *Ferula*. Additionally, the pharmacopoeia mentions ferulic acid as potentially another characteristic chemical constituent of the genus *Ferula*.

5.2. Q-Marker Prediction Analysis by Chemical Composition Validity

As an emerging method to evaluate and control the quality of Chinese herbal medicine Q-markers is essential to predict both the effectiveness and safety of Chinese herbal medicines. Therefore, Q-marker analysis of Chinese herbal medicines should be combined with the effectiveness of the target herbal medicines to facilitate better research on the effectiveness of Chinese herbal medicines [146,147]. Because of the presence of the different coumarin sesquiterpenoids in large quantities in the genus *Ferula*, most studies on the chemical composition effectiveness and safety of *Ferula* resources have primarily focused on these compounds, which are expected to become quality markers of *Ferula* plants.

5.2.1. Q-Marker Prediction Analysis by Traditional Drug Properties

The medicinal properties of *Ferula* plants have been recorded as Ku (bitter), Xing (pungent) and Wen (warm), as well as Guipi and Weijing (benefit for the spleen and stomach) in the Chinese Pharmacopoeia Edition (2020). The chemical compositions of the various bitter medicines in Chinese medicine are mainly alkaloids, glycosides, terpenoids and other bitter substances [148,149]. Terpenes have a bitter taste primarily due to their chemical structure with chelating structures such as lactones, endo-acetals, endo-hydrogen bonds and glycosidic groups [150]. The sesquiterpene coumarins, which are found in relatively large quantities in the genus *Ferula*, are based on 7-hydroxyumbelliferolactone as the parent nucleus. A number of prior studies have shown that terpenes and volatile oils are found in ample quantities in herbs with the Xing (pungent) taste [151], whereas coumarin sesquiterpenes are similarly abundant in the genus *Ferula*. In addition, the main chemical components of genus *Ferula*, such as Farnesiferol A, Farnesiferol B, Farnesiferol C and DAW22, are all coumarin sesquiterpenoids.

5.2.2. Predictive Analysis by the Q-Marker of Traditional Efficacy

The effects of *Ferula* plants have been recorded in the Chinese Pharmacopoeia 2020 edition and found to be useful as digesting aid, relief of symptoms, dispersal of agglomerates as well as insecticide. They are also used for the treatment of excessive meat consumption, leading to stagnation and masses in the abdomen, and treating bruises, intestinal parasites and the abdominal pain [17]. Their effects are reported in the Xinxiubencao as an insecticide, deodorization, in elimination of intra-abdominal masses, relief of blood stasis, septicemia and pus, and alleviation of symptoms of poisoning [152]. *Ferula* plants have been also used to treat tuberculosis, eliminate lumps that have accumulated in the abdomen, cure cold, and treat malaria, which are effective against acute gastroenteritis as well as other intestinal diseases. They have a pain-relieving effect on the chest and stomach in addition to pain [153] and similar records have been found in other ethnomedicinal books [154]. Moreover, during the course of modern pharmacological research, the pharmacological effects of *Ferula* plants are similar to those recorded in ancient books, such as anti-allergy, action on the gastrointestinal tract, anti-bacterial as well as antiseptic effects, anticoagulant effects, and effects related to the cardiovascular system. The CHCl₃ extracted fraction of *F. sinkiangensis* resin can exhibit significant anti-gastric ulcer activity, and the sesquiterpene coumarins contained in this fraction were mainly Farnesiferol B and Farnesiferol C. In addition, prior studies have shown that the raw juices of *F. sinkiangensis* and *F. feruloides* have certain effects on gastrointestinal smooth muscle inhibition and anti-experimental gastric ulcer, whereas the chemical compositions of these three types mainly consist of the sesquiterpenes, Farnesiferol A [58], Farnesiferol B, Farnesiferol C. Interestingly, in vivo studies have shown that Farnesiferol B can protect the kidney from I/R-induced damage by reducing oxidative stress and inflammation. In vitro, Farnesiferol B was reported

to improve macrophage migration by activating TGR5 [155]. Furthermore, DAW22 was shown in the literature to inhibit five human MPNST cancer cell lines and C6 glioma cells to varying degrees. Overall, ferulic acid possesses numerous pharmacological activities, mainly related to anticancer, anti-bacterial, anti-inflammatory, antioxidant, anti-thrombotic actions, and can also exhibit hypolipidemic effects [156,157].

5.3. Q-Marker Predictive Analysis by Chemical Composition Measurability

The chemical compositions of Chinese medicines are complex, and therefore the chemical measurability of the subject is reported to be an essential element in the Q-marker study [158]. Thus, based on the previous summary study on the specificity and close linkage of the chemical compositions as well as effectiveness of the domestic plants of the genus *Ferula*, the sesquiterpene coumarins and ferulic acid were further identified as the quality markers of *Ferula* plants by measuring their chemical compositions. The contents of ferulic acid were determined by HPLC on the different parts of *F. sinkiangensis* and the roots and leaves of *F. fukanensis*, and the content of ferulic acid was found to be in the following descending order: gum > root > leaf > stem [159,160], and the content of ferulic acid was also determined by HPLC on the anti-ulcer extract of *F. sinkiangensis* [161]. In addition, the determination of Farnesiferol A, Farnesiferol C and ferulic acid in the ethyl acetate fraction of *F. sinkiangensis* was performed by HPLC [162]. The UPLC method was used to estimate the content of DAW22 in *F. feruloides*. It was observed that the methanolic extract of *F. feruloides* contained more DAW22, and DAW22 was most abundant in *F. feruloides* harvested in the first half of May [163].

6. Conclusions

Although the genus *Ferula* is widespread and diverse throughout the world, its distribution in China is concentrated mainly in Xinjiang and is less diverse. *Ferula* is native to Central Asia and Middle East, and especially found in eastern Iran and Afghanistan, and is exported to be present worldwide. Nevertheless, *Ferula* is not native to China, and it has been around for a long time but has become a traditional Chinese herb and ethnic medicine. It plays a vital role in the traditional Chinese medical system.

The document analysis of the China's *Ferula* resources revealed that there are several studies describing the chemical compositions and biological activities of *Ferula* plants. *Ferula* are mainly composed of coumarins, particularly sesquiterpenes coumarins, volatile oils, sulfur-containing compounds, and aromatic compounds. Interestingly, beneficial pharmacological activity of *Ferula* is not only traditionally used, but its various components have been reported to possess anticancer, anti-diabetic, antibacterial, anti-flu, anti-inflammatory and other effects.

The documented benefits of *Ferula* include the elimination of lumps, deworming, antibacterial and other traditional uses. In addition, some traditional uses of *Ferula* are closely related to the modern research and have clinical relevance. For example, modern phytochemical and pharmacological studies have revealed that DAW22 could target the main components in the MPNST tumorigenic pathways: namely, suppress the phosphorylation of AKT and ERK, and reduce the levels of non-phospho (active) CTNNB1. Moreover, other studies have reported that farnesylate A can facilitate the anticancer drugs to exert substantial cytotoxic effects by inhibiting P-glycoprotein. Another study showed that both Farnesiferol B and Farnesiferol C displayed the best inhibitory effects on P-gp pump efflux and they could be considered as lead scaffolds for further structure modifications. Currently, anticancer activity of *F. ferulaeoides* is being investigated by the author and his colleagues.

Because of the availability of a wide variety of TCM, their quality standards are not uniform, thereby resulting in the gradual increase in defective medicines, which can adversely the efficacy of TCM. Faced with this challenge, in 2016, Academician Changxiao Liu proposed the concept of the Q-marker. A Q-marker can better identify the various characteristic compounds present in TCM and can be used sequentially as a criterion to evaluate their quality and to better identify their authenticity, thus providing quality

assurance for the development of TCM. Additionally, it was found that there were only a few selected studies related to Q-markers of domestic *Ferula*. Q-marker studies can be of great help to the breeding, conservation and utilization of the domestic *Ferula*. However, it was observed that the few studies on the Q-markers of domestic *Ferula* were not conducive to the research development of domestic *Ferula* spp. in the context of mixed quality.

Moreover, there are only few toxicological studies on the genus *Ferula* in China. Toxicological studies could be of great help in the clinical analysis of the active components in the domestic *Ferula* plants, and hence toxicological studies about the chemical composition of domestic *Ferula* plants should also be strengthened in the future.

Eventually, *Ferula* plants have a long history of use in China, but *Ferula* plant resources have suffered great damage due to human activities in recent years, resulting in acute shortage of plant resources and protective measures are urgently needed. However, regarding the rich background of biological activities of *Ferula*, it appears that there are still a large number of unaccomplished investigations that need to be conducted in the future. Examples include research on the biological characteristics and genetic variation mechanisms of the genus *Ferula*, for better germplasm conservation and utilization; research on the medicinal value and pharmacological mechanisms of action of the genus *Ferula*, to explore its application prospects and value in the field of medicine; research on the cultivation and production techniques of *Ferula* spp. to improve its yield and quality and to meet market demand; analysis and research on the distribution status of the resources and ecological impact of the genus *Abies*, and the development of conservation strategies to effectively protect and manage these precious plant resources.

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References

1. Salehi, M.; Naghavi, M.R.; Bahmankar, M. A review of *Ferula* species: Biochemical characteristics, pharmaceutical and industrial applications, and suggestions for biotechnologists. *Ind. Crops Prod.* **2019**, *139*, 111511. [[CrossRef](#)]
2. Znati, M.; Hichem, B.J.; Cazaux, S.; Bouajila, J. Chemical composition, biological and cytotoxic activities of plant extracts and compounds isolated from *Ferula lutea*. *Molecules* **2014**, *19*, 2733–2747. [[CrossRef](#)] [[PubMed](#)]
3. Dan, R.H.; She, M.L. *Flora of China*; Science Press: Beijing, China, 1992; Volume 55, pp. 85–117.
4. He, S.; Tan, G.Y. Advances in Studies of *Ferula* L. *J. Xinjiang Agric. Univ.* **2002**, *25*, 1–7.
5. Eigner, D.; Scholz, D. The magic book of Gyani Dolma. *Pharm. Unserer. Zeit.* **1990**, *19*, 141–152. [[CrossRef](#)] [[PubMed](#)]
6. Bandyopadhyay, D.; Basak, B.; Chatterjee, A.; Lai, T.K.; Banerji, A.; Banerji, J.; Neuman, A.; Prange, T. Saradaferin, a new sesquiterpenoid coumarin from *Ferula assafoetida*. *Nat. Prod. Res.* **2006**, *20*, 961–965. [[CrossRef](#)]
7. Akaberi, M.; Iranshahy, M.; Iranshahi, M. Review of the traditional uses, phytochemistry, pharmacology and toxicology of giant fennel (*Ferula communis* L. subsp. *communis*). *Iran. J. Basic Med. Sci.* **2015**, *18*, 1050.
8. Mahendra, P.; Bisht, S. *Ferula asafoetida*: Traditional uses and pharmacological activity. *Pharmacogn. Rev.* **2012**, *6*, 141. [[CrossRef](#)]
9. Fan, C.; Li, X.; Zhu, J.; Song, J.; Yao, H. Endangered Uyghur Medicinal Plant *Ferula* Identification through the Second Internal Transcribed Spacer. *Evid. Based Complement. Altern. Med.* **2015**, *2015*, 479879. [[CrossRef](#)]
10. Xinjiang Food and Drug Administration. *The Xinjiang Uyghur Autonomous Region Traditional Chinese Medicine and Uyghur Medicine Pieces Concoct Standard*; Xinjiang People's Saitary Press: Urumqi, China, 2010.

11. Chinese Pharmacopoeia Commission. *Chinese Pharmacopoeia*; Chinese Medicine Science and Technology Press: Beijing, China, 2020; Volume 1, p. 198.
12. Iranshahy, M.; Iranshahi, M. Traditional uses, phytochemistry and pharmacology of asafoetida (*Ferula assa-foetida* oleo-gum-resin)—A review. *J. Ethnopharmacol.* **2011**, *134*, 1–10. [[CrossRef](#)]
13. Amalraj, A.; Gopi, S. Biological activities and medicinal properties of Asafoetida: A review. *J. Tradit. Complement. Med.* **2017**, *7*, 347–359. [[CrossRef](#)]
14. Yatham, P.; Shukla, D.; Srivastava, A.K.; Pragadheesh, V.S.; Kumar, D. Purification and identification of anticancer organosulfides from *Ferula assa-foetida* gum: Integrative analysis employing GC/GC-MS/RP-HPLC/NMR. *Nat. Prod. Res.* **2022**, *36*, 2869–2874. [[CrossRef](#)] [[PubMed](#)]
15. Saleem, M.; Alam, A.; Sultana, S. Asafoetida inhibits early events of carcinogenesis: A chemopreventive study. *Life Sci.* **2001**, *68*, 1913–1921. [[CrossRef](#)] [[PubMed](#)]
16. Divya, K.; Ramalakshmi, K.; Murthy, P.S.; Jagan Mohan Rao, L. Volatile oils from *Ferula asafoetida* varieties and their antimicrobial activity. *LWT Food Sci. Technol.* **2014**, *59*, 774–779. [[CrossRef](#)]
17. Javanshir, S.; Soukhtanloo, M.; Jalili-Nik, M.; Yazdi, A.J.; Amiri, M.S.; Ghorbani, A. Evaluation Potential Antidiabetic Effects of *Ferula latisecta* in Streptozotocin-Induced Diabetic Rats. *J. Pharmacopunct.* **2020**, *23*, 158–164. [[CrossRef](#)] [[PubMed](#)]
18. Lee, C.L.; Chiang, L.C.; Cheng, L.H.; Chuang, L.C.; Mohamed, H.A.E.-R.; Chang, F.R.; Wu, Y.C. Influenza A (H₁N₁) Antiviral and Cytotoxic Agents from *Ferula assa-foetida*. *J. Nat. Prod.* **2009**, *72*, 1568–1572. [[CrossRef](#)]
19. Ghasemi, Z.; Rezaee, R.; Aslani, M.R.; Boskabady, M.H. Anti-inflammatory, anti-oxidant, and immunomodulatory activities of the genus *Ferula* and their constituents: A review. *Iran J. Basic Med. Sci.* **2021**, *24*, 1613–1623.
20. Mobasheri, L.; Khorashadizadeh, M.; Safarpour, H.; Mohammadi, M.; Anani Sarab, G.; Askari, V.R. Anti-Inflammatory Activity of *Ferula assafoetida* Oleo-Gum-Resin (Asafoetida) against TNF-alpha-Stimulated Human Umbilical Vein Endothelial Cells (HUVECs). *Mediat. Inflamm.* **2022**, *2022*, 5171525. [[CrossRef](#)]
21. Safaeian, L.; Ghannadi, A.; Javanmard, S.H.; Vahidian, M.H. The effect of hydroalcoholic extract of *Ferula foetida* stems on blood pressure and oxidative stress in dexamethasone-induced hypertensive rats. *Res. Pharm. Sci.* **2014**, *10*, 326–334.
22. Ellman, G.L.; Courtney, K.D.; Andres, V., Jr.; Featherstone, R.M. A New and Rapid Colorimetric Determination of Acetylcholinesterase Activity. *Biochem. Pharmacol.* **1961**, *7*, 88–95. [[CrossRef](#)]
23. Karimi, G.; Iranshahi, M.; Hosseinalizadeh, F.; Riahi, B.; Sahebkar, A. Screening of acetylcholinesterase inhibitory activity of terpenoid and coumarin derivatives from the genus *Ferula*. *Pharmacol.* **2010**, *1*, 566–574.
24. Hashemzaei, M.; SadeghiBonjar, M.A.; Tabrizian, K.; Iranshahi, M.; Iranshahy, M.; Rezaee, R. Evaluation of the analgesic effect of Umbelliprenin and Umbelliprenin-morphine co-administration on the acute, chronic and neuropathic pain. *Indian J. Pharm. Educ. Res.* **2015**, *49*, 121–125. [[CrossRef](#)]
25. Dastan, D.; Salehi, P.; Aliahmadi, A.; Gohari, A.R.; Maroofi, H.; Ardalan, A. New coumarin derivatives from *Ferula pseudalliacea* with antibacterial activity. *Nat. Prod. Res.* **2016**, *30*, 2747–2753. [[CrossRef](#)]
26. Nazari, Z.E.; Iranshahi, M. Biologically active sesquiterpene coumarins from *Ferula* species. *Phytother. Res.* **2011**, *25*, 315–323. [[CrossRef](#)]
27. Pavela, R.; Morshedloo, M.R.; Lupidi, G.; Carolla, G.; Barboni, L.; Quassinti, L.; Bramucci, M.; Vitali, L.A.; Petrelli, D.; Kavallieratos, N.G.; et al. The volatile oils from the oleo-gum-resins of *Ferula assa-foetida* and *Ferula gummosa*: A comprehensive investigation of their insecticidal activity and eco-toxicological effects. *Food Chem. Toxicol.* **2020**, *140*, 111312. [[CrossRef](#)]
28. Soltani, S.; Amin, G.R.; Salehi-Sourmaghi, M.H.; Schneider, B.; Lorenz, S.; Iranshahi, M. Sulfur-containing compounds from the roots of *Ferula latisecta* and their cytotoxic activities. *Fitoterapia* **2018**, *124*, 108–112. [[CrossRef](#)]
29. Rasulev, B.F.; Saidkhodzhaev, A.I.; Nazrullaev, S.S.; Akhmedkhodzhaeva, K.S.; Khushbaktova, Z.A.; Leszczynski, J. Molecular modelling and QSAR analysis of the estrogenic activity of terpenoids isolated from *Ferula* plants. *SAR QSAR Environ. Res.* **2007**, *18*, 663–673. [[CrossRef](#)] [[PubMed](#)]
30. Guo, D.A.; Lu, A.; Liu, L. Modernization of traditional Chinese medicine. *J. Ethnopharmacol.* **2012**, *141*, 547–548. [[CrossRef](#)]
31. Guo, D.A.; Wu, Y.Y.; Ye, M.; Liu, X.; Cordell, G.A. A holistic approach to the quality control of traditional Chinese medicines. *Science* **2015**, *347*, 29–31.
32. Liu, C.-X.; Cheng, Y.-Y.; Guo, D.-A.; Zhang, T.-J.; Li, Y.-Z.; Hou, W.-B.; Huang, L.-Q.; Xu, H.-Y. A New Concept on Quality Marker for Quality Assessment and Process Control of Chinese Medicines. *Chin. Herb. Med.* **2017**, *9*, 3–13. [[CrossRef](#)]
33. Yang, W.; Zhang, Y.; Wu, W.; Huang, L.; Guo, D.; Liu, C. Approaches to establish Q-markers for the quality standards of traditional Chinese medicines. *Acta Pharm. Sin. B* **2017**, *7*, 439–446. [[CrossRef](#)] [[PubMed](#)]
34. State Key Laboratory of Systematic and Evolutionary Botany, I.o.B. The Chinese Academy of Sciences Angiospermae >> Apiaceae >> *Ferula* >> *Ferula sinkiangensis*. Available online: <http://ppbc.iplant.cn/tu/6312937> (accessed on 15 June 2023).
35. State Key Laboratory of Systematic and Evolutionary Botany, I.o.B. The Chinese Academy of Sciences Angiospermae >> Apiaceae >> *Ferula* >> *Ferula fukanensis*. Available online: <http://ppbc.iplant.cn/tu/4351033> (accessed on 15 June 2023).
36. State Key Laboratory of Systematic and Evolutionary Botany, I.o.B. The Chinese Academy of Sciences Angiospermae >> Apiaceae >> *Ferula* >> *Ferula kingdon-wardii*. Available online: <http://ppbc.iplant.cn/tu/8596963> (accessed on 15 June 2023).
37. State Key Laboratory of Systematic and Evolutionary Botany, I.o.B. The Chinese Academy of Sciences Angiospermae >> Apiaceae >> *Ferula* >> *Ferula songarica*. Available online: <http://ppbc.iplant.cn/tu/6473066> (accessed on 15 June 2023).

38. State Key Laboratory of Systematic and Evolutionary Botany, I.o.B. The Chinese Academy of Sciences Angiospermae >> Apiaceae >> Ferula >> *Ferula syreitschikowii*. Available online: <http://ppbc.iplant.cn/tu/6509768> (accessed on 15 June 2023).
39. State Key Laboratory of Systematic and Evolutionary Botany, I.o.B. The Chinese Academy of Sciences Angiospermae >> Apiaceae >> Ferula >> *Ferula licentiana*. Available online: <http://ppbc.iplant.cn/tu/4286927> (accessed on 15 June 2023).
40. Li, Y.D.; Fu, S.Y.; He, J.; Fan, C.Z.; Li, X.J. Analysis of Specific Medicinal Plants Resources *Ferula sinkiangensis* in Xinjiang. *Mod. Chin. Med.* **2016**, *18*, 714–717.
41. Yan, X.H.; Zhou, B.; Liu, H.B.; Xu, H.Y.; Yang, W.X.; Jiao, X.G.; Tian, S.G. Investigation and protection for endangered *Ferula fukanensis* K.M. Shen. *J. Xinjiang Med. Univ.* **2012**, *35*, 1155–1158.
42. Xinjiang Institute of Biological Soil and Desert Research. *Flora of Xinjiang Medical Plants*; Xinjiang People's Publishing House: Urumqi, China, 1977; Volume 1.
43. Tan, Y.; Gao, T.T.; Ma, Y.; Ding, X.; Chen, Y.H. Pharmacognostic Identification of *Ferula syreitschikowii*. *J. Chin. Med. Mater.* **2011**, *34*, 1694–1696.
44. Editorial Committee of the Chinese Medical Encyclopedia. *Uyghur Medicine*; Shanghai Scientific & Technical Publishers: Shanghai, China, 2005.
45. Zhou, R.H. *Resource of Chinese Medicinal Materials*; China Medico-Pharmaceutical Science & Technology Publishing House: Beijing, China, 1993; Volume 05.
46. Editorial Board of "Primary colour mapping of weeds in Chinese farmland". *Primary Colour Mapping of Weeds in Chinese Farmland*; China Agriculture Press: Beijing, China, 1990.
47. Ding, X.; Tan, Y.; Liu, J.S.; Zhao, W.B. Pharmacognostical Study on *Ferula dissecta* (Ledeb.) Ledeb. *Lishizhen Med. Mater. Med.* **2011**, *22*, 1555–1556.
48. Science and Technology Commission of Tibet Autonomous Region. *Tibetan Plant List*; Science and Technology Commission of Tibet Autonomous Region: Lhasa, China, 1980.
49. Jiang, J.W. *Quick-Consultative Dictionary of World Medicinal Plants*; China Medico-Pharmaceutical Science & Technology Publishing House: Beijing, China, 2015.
50. Xinjiang Bayi Agricultural College. *Search List of Plants of Xinjiang*; Xinjiang People's Publishing House: Urumqi, China, 1983.
51. Shangguan, T.L.; Ma, Z.Q.; Xie, S.L. *Rare and Endangered Plants in Shanxi Province*; China Science and Technology Press: Beijing, China, 1998.
52. He, J.Q. *Flora of Northern Anhui Resources*; China Agriculture Press: Beijing, China, 2001.
53. Liu, N.F. *Scientific Expedition to Dunhuang Nature Reserve, Gansu*; Chinese Forestry Publishing House: Beijing, China, 2001.
54. Southwest Forestry College; Yunnan Forestry Department; Zhongdian County Forestry Bureau. *Report on a Comprehensive Scientific Study of the Bitahai Nature Reserve in Yunnan*; Southwest Forestry College, Yunnan Forestry Department, Zhongdian County Forestry Bureau: Kunming, China, 2002; Volume 2.
55. Executive Committee of the First Annual Youth Academic Conference of Jiangsu Province. In Proceedings of the First Annual Youth Academic Conference of Jiangsu Province (Science Fascicle), Nanjing, China, 17–19 November 1992; China Science and Technology Press: Beijing, China, 1992.
56. Lin, J.R.; Jin, M.; Wu, C.M. Advances in studies on chemical constituents and pharmacological effects of *Ferula*. *Strait Pharm. J.* **2014**, *26*, 1–3.
57. El-Razek, M.A.; Ohta, S.; Hirata, T. Terpenoid Coumarins of the Genus *Ferula*. *Heterocycles* **2003**, *34*, 68971. [[CrossRef](#)]
58. Xing, Y.; Li, N.; Zhou, D.; Chen, G.; Jiao, K.; Wang, W.; Si, Y.; Hou, Y. Sesquiterpene Coumarins from *Ferula sinkiangensis* Act as Neuroinflammation Inhibitors. *Planta Med.* **2017**, *83*, 135–142. [[CrossRef](#)]
59. Wang, J.; Wang, H.; Zhang, M.; Li, X.; Zhao, Y.; Chen, G.; Si, J.; Jiang, L. Sesquiterpene coumarins from *Ferula sinkiangensis* KM Shen and their cytotoxic activities. *Phytochemistry* **2020**, *180*, 112531. [[CrossRef](#)] [[PubMed](#)]
60. Li, X.Y.; Li, G.Y.; Wang, H.Y.; Wang, Y.; Wang, J.H. Chemical Constituents from *Ferula Lehmannii* Boiss. *Mod. Chin. Med.* **2010**, *12*, 17–20.
61. Yang, J.R.; An, Z.; Li, Z.H.; Jing, S.; Qina, H.L. Sesquiterpene coumarins from the roots of *Ferula sinkiangensis* and *Ferula teterrima*. *Chem. Pharm. Bull.* **2006**, *54*, 1595–1598. [[CrossRef](#)]
62. Wang, Y.E.; Si, J.Y.; Li, X.J.; Jiang, L.; Zhu, J. Chemical Constituents from the Seeds of *Ferula sinkiangensis*. *Mod. Chin. Med.* **2011**, *13*, 26–28.
63. Li, G.; Li, X.; Cao, L.; Zhang, L.; Shen, L.; Zhu, J.; Wang, J.; Si, J. Sesquiterpene coumarins from seeds of *Ferula sinkiangensis*. *Fitoterapia* **2015**, *103*, 222–226. [[CrossRef](#)]
64. Meng, H.; Li, G.; Huang, J.; Zhang, K.; Wang, H.; Wang, J. Sesquiterpene coumarin and sesquiterpene chromone derivatives from *Ferula ferulaeoides* (Steud.) Korov. *Fitoterapia* **2013**, *86*, 70–77. [[CrossRef](#)]
65. Motai, T.; Kitanaka, S. Sesquiterpene coumarins from *Ferula fukanensis* and nitric oxide production inhibitory effects. *Chem. Pharm. Bull. (Tokyo)* **2004**, *52*, 1215–1218. [[CrossRef](#)] [[PubMed](#)]
66. Motai, T.; Daikonya, A.; Kitanaka, S. Sesquiterpene coumarins from *Ferula fukanensis* and their pro-inflammatory cytokine gene expression inhibitory effects. *Chem. Pharm. Bull.* **2013**, *61*, 618–623. [[CrossRef](#)] [[PubMed](#)]
67. Meng, H.; Li, G.; Huang, J.; Zhang, K.; Wei, X.; Ma, Y.; Zhang, C.; Wang, J. Sesquiterpenoid derivatives from *Ferula ferulaeoides* (Steud.) Korov. *Phytochemistry* **2013**, *86*, 151–158. [[CrossRef](#)] [[PubMed](#)]

68. Kahraman, C.; Topcu, G.; Bedir, E.; Tatli, I.I.; Ekizoglu, M.; Akdemir, Z.S. Phytochemical screening and evaluation of the antimicrobial and antioxidant activities of *Ferula caspica* M. Bieb. extracts. *Saudi Pharm. J.* **2019**, *27*, 525–531. [[CrossRef](#)]
69. Motai, T.; Kitanaka, S. Sesquiterpene Chromones from *Ferula fukanensis* and Their Nitric Oxide Production Inhibitory Effects. *J. Nat. Prod.* **2005**, *68*, 1732–1735. [[CrossRef](#)]
70. Motai, T.; Kitanaka, S. Sesquiterpene Phenylpropanoids from *Ferula fukanensis* and Their Nitric Oxide Production Inhibitory Effects. *J. Nat. Prod.* **2005**, *68*, 365–368. [[CrossRef](#)]
71. Garg, S.N.; Agarwal, S.K. New Sesquiterpenes from *Ferula jaeschkeana*. *Planta Med.* **1986**, *53*, 341–342. [[CrossRef](#)]
72. Liu, Q.X.; Hui, H. The chemical constituents of volatile oil from *Ferula* L. in China and its taxonomical significance. *J. Plant Resour. Environ.* **1997**, *6*, 27–32.
73. Lei, L.J.; Teng, L.; Zhao, X.; Yu, F.S.; Wang, C.H. Extraction and assay of volatile oil from *Ferula ferulaeoidis* (Sted.) Korov. *Chin. Tradit. Pat. Med.* **2013**, *35*, 1251–1256.
74. Deng, W.P.; Xie, C.X.; Fu, J.H. Analysis of Volatile Oil from *Ferula sinkiangensis* by GC/MS. *J. Chin. Mass Spectrom. Soc.* **2007**, *28*, 114–116+121.
75. Yang, M.H.; Luo, J.Y.; Qiao, M.L.; Yang, M.H.; Sheng, P. GC-MS Analysis of Volatile Oil from *Ferula ferulaeoides* and Anti-gastric Cancer Activity of D-Limonene in vitro. *Chin. J. Mod. Appl. Pharm.* **2020**, *37*, 806–813.
76. Li, X.; Wang, Y.; Zhu, J.; Xiao, Q. Essential oil composition analysis of three cultivars seeds of Resina *Ferulae* from Xinjiang, China. *Pharm. Mag.* **2011**, *7*, 116–120. [[CrossRef](#)]
77. Zhi-da, M.; Qi-fi, M.; Mizuno, M.; Tanaka, T.; Munekazu, I. Polysulfanes in the volatile oils of *Ferula* species. *Planta Med.* **1986**, *53*, 300–302. [[CrossRef](#)]
78. Xu, Q.M. Studies of the Chemical Constituents of *Varia kweichowensis* and *Ferula fukanensis* KM Shen. Ph.D. Thesis, Peking Union Medical College, Tsinghua University, Beijing, China, 2005.
79. Zhu, G.X.; Zhang, H.Q. A New Compound from the Root of *Ferula tunshanica* Su. *J. China Pharm. Univ.* **1998**, *29*, 19–20.
80. Hu, Y.; Li, X.D.; Li, G.Y.; Li, X.Y.; Zhang, K.; Zuo, W.J.; Zeng, Y.N.; Wang, J.H. Studies on the Chemical Constituents of *Ferula songorica*. *Mod. Chin. Med.* **2009**, *11*, 18–19.
81. Zhu, G.X.; Zhang, H.Q. Studies on the Chemical Constituents in the Root of *Ferula tunshanica* Su. *J. China Pharm. Univ.* **1996**, *27*, 585–588.
82. Liu, T. Chemical Constituents from Two Plants of *Apiaceae* and Their Biological Activities. Ph.D. Thesis, Fudan University, Shanghai, China, 2013.
83. Liu, J. Study on Chemical Constituents and Quality Control Method of *Ferula ferulaeoides*. Master's Thesis, Inner Mongolia Medical University, Hohhot, China, 2019.
84. Zhao, W.B.; Tan, Y.; Xiang, Y.; Zhu, Y. Determination of Amino Acids and Mineral Elements Content in *Ferulaeoides korov* Boiss. *Res. Pract. Chin. Med.* **2009**, *23*, 67–68.
85. Deng, W.P. Extract and Study of the Chemical Constituents of *Ferula Sinkiangensis*. Master's Thesis, Xinjiang University, Urumqi, China, 2008.
86. Han, H.Y.; Li, G.Y.; Wang, H.Y.; Ma, Q.D.; Gao, J.B.; Wang, J.H. Anticoagulant activity of radiatinol and scopoletin from *Ferula dissecta* (Ledeb.) Ledeb. in vitro. *J. Nongken Med.* **2010**, *32*, 112–114.
87. Furlan, V.; Bren, U. *Helichrysum italicum*: From Extraction, Distillation, and Encapsulation Techniques to Beneficial Health Effects. *Foods* **2023**, *12*, 802. [[CrossRef](#)]
88. Iranshahi, M.; Kalategi, F.; Rezaee, R.; Shahverdi, A.R.; Ito, C.; Furukawa, H.; Tokuda, H.; Itoigawa, M. Cancer chemopreventive activity of terpenoid coumarins from *Ferula* species. *Planta Med.* **2008**, *74*, 147–150. [[CrossRef](#)] [[PubMed](#)]
89. Monti, M.; Pinotti, M.; Appendino, G.; Dallochio, F.; Bellini, T.; Antognoni, F.; Poli, F.; Bernardi, F. Characterization of anti-coagulant properties of prenylated coumarin ferulenol. *Biochim. Biophys. Acta* **2007**, *1770*, 1437–1440. [[CrossRef](#)] [[PubMed](#)]
90. Ozkan, H.; Yanmis, D.; Karadayi, M.; Bal, T.; Baris, O.; Gulluce, M. Determination of genotoxic and antigenotoxic properties of essential oil from *Ferula orientalis* L. using Ames/Salmonella and *E. coli* WP2 bacterial test systems. *Toxicol. Ind. Health* **2014**, *30*, 714–723. [[CrossRef](#)]
91. Illuri, R.; Venkataramana, S.H.; Daguet, D.; Kodimule, S. Sub-acute and acute toxicity of *Ferula asafoetida* and *Silybum marianum* formulation and effect of the formulation on delaying gastric emptying. *BMC Complement. Altern. Med.* **2019**, *19*, 159. [[CrossRef](#)] [[PubMed](#)]
92. Maiuolo, J.; Bava, I.; Carresi, C.; Gliozzi, M.; Musolino, V.; Scicchitano, M.; Macri, R.; Oppedisano, F.; Scarano, F.; Caterina Zito, M.; et al. The Effect of *Ferula communis* Extract in *Escherichia coli* Lipopolysaccharide-Induced Neuroinflammation in Cultured Neurons and Oligodendrocytes. *Int. J. Mol. Sci.* **2021**, *22*, 7910. [[CrossRef](#)]
93. Ghollitabar, S.; Roshan, V.D. Effect of treadmill exercise and *Ferula gummosa* on myocardial HSP72, vascular function, and antioxidant defenses in spontaneously hypertensive rats. *Clin. Exp. Hypertens.* **2013**, *35*, 347–354. [[CrossRef](#)]
94. Esmaeili, H.; Hafezimoghadam, Z.; Esmailidehaj, M.; Rezvani, M.E.; Hafizibarjin, Z. The effect of asafoetida essential oil on myocardial ischemic-reperfusion injury in isolated rat hearts. *Avicenna J. Phytomed.* **2017**, *8*, 338–349.
95. Zhou, Y.; Xin, F.; Zhang, G.; Qu, H.; Yang, D.; Han, X. Recent Advances on Bioactive Constituents in *Ferula*. *Drug Dev. Res.* **2017**, *78*, 321–331. [[CrossRef](#)]
96. Xing, Y.C.; Li, N.; Xue, J. Progress on chemical constituents of *Ferula* genus. *J. Shenyang Pharm. Univ.* **2012**, *29*, 730–741.

97. Guo, T.T.; Zhou, Y.P.; Dang, W.; Meng, C.R.; Zhou, D.; Chen, G.; Li, N. "Preexistence" and "present life" of traditional Chinese medicine *Ferulae Resina*. *Chin. Tradit. Herb. Drugs* **2021**, *52*, 5401–5413.
98. Ying, F.H.; Qian, X.Q.; Liu, B.R. Progress in the study of the anti-tumor mechanism of action of ferulic acid. *Mod. J. Integr. Tradit. Chin. West. Med.* **2010**, *19*, 4238–4240.
99. Suzuki, K.; Okasaka, M.; Kashiwada, Y.; Takaishi, Y.; Honda, G.; Ito, M.; Takeda, Y.; Kodzhimatov, O.K.; Ashurmetov, O.; Sekiya, M.; et al. Sesquiterpene Lactones from the Roots of *Ferula Waria* and Their Cytotoxic Activity. *J. Nat. Prod.* **2007**, *70*, 1915–1918. [[CrossRef](#)] [[PubMed](#)]
100. Wang, S.; Sheng, P.; Yao, L.; Du, B.J. GC-MS fingerprint of in vitro anti-gastric cancer active parts from roots of Uyghur medicine *Ferula ferulaeoides*. *Chin. Tradit. Herb. Drugs* **2015**, *46*, 2874–2879.
101. Wu, J.; Wang, Y.X.; Wei, N.; Wu, X.S.; Liu, X. Ferulic acid inhibits lung cancer growth and its mechanism. *Zhejiang Med. J.* **2018**, *40*, 1303–1306+1311+1414.
102. Yang, M.H.; Luo, J.Y.; Qiao, M.L.; Sheng, P.; Yang, M.H. Mechanism of Uyghur Medicine Root of *Ferula ferulaeoides* in Resisting Gastric Cancer Activity and Inducing Cell Apoptosis and Cell Cycle Arrest in vitro. *Chin. J. Exp. Tradit. Med. Formulae* **2018**, *24*, 112–122.
103. Zhang, L.; Tong, X.; Zhang, J.; Huang, J.; Wang, J. DAW22, a natural sesquiterpene coumarin isolated from *Ferula ferulaeoides* (Steud.) Korov. that induces C6 glioma cell apoptosis and endoplasmic reticulum (ER) stress. *Fitoterapia* **2015**, *103*, 46–54. [[CrossRef](#)]
104. Li, X.X.; Zhang, S.J.; Chiu, A.P.; Lo, L.H.; Huang, J.; Rowlands, D.K.; Wang, J.; Keng, V.W. Targeting of AKT/ERK/CTNBN1 by DAW22 as a potential therapeutic compound for malignant peripheral nerve sheath tumor. *Cancer Med.* **2018**, *7*, 4791–4800. [[CrossRef](#)]
105. Zhang, C.; Jiang, Y.; Zhang, J.; Huang, J.; Wang, J. 8-p-Hdroxybenzoyl Tovarol Induces Paraptosis Like Cell Death and Protective Autophagy in Human Cervical Cancer HeLa Cells. *Int. J. Mol. Sci.* **2015**, *16*, 14979–14996. [[CrossRef](#)]
106. Kores, K.; Kolenc, Z.; Furlan, V.; Bren, U. Inverse Molecular Docking Elucidating the Anticarcinogenic Potential of the Hop Natural Product Xanthohumol and Its Metabolites. *Foods* **2022**, *11*, 1253. [[CrossRef](#)] [[PubMed](#)]
107. Gao, T.T.; Yu, F.H.; Tan, Y.; Liu, W.X. Study on Antimicrobial Activity in vitro of Extracts from Three Species of *Ferula* Root. *North. Hortic.* **2013**, *24*, 156–158.
108. Liu, T.; Wang, S.; Fu, W.; Gibbons, S.; Mu, Q. Sesquiterpenoids with Anti-MDR *Staphylococcus aureus* Activities from *Ferula feruloides*. *Chem. Biodivers.* **2015**, *12*, 599–614. [[CrossRef](#)] [[PubMed](#)]
109. Liu, T.; Osman, K.; Kaatz, G.W.; Gibbons, S.; Mu, Q. Antibacterial Sesquiterpenoid Derivatives from *Ferula ferulaeoides*. *Planta Med.* **2013**, *79*, 701–706. [[CrossRef](#)]
110. Nazemisalman, B.; Vahabi, S.; Yazdinejad, A.; Haghghi, F.; Jam, M.S.; Heydari, F. Comparison of antimicrobial effect of *Ziziphora tenuior*, *Dracocephalum moldavica*, *Ferula gummosa*, and *Prangos ferulacea* essential oil with chlorhexidine on *Enterococcus faecalis*: An in vitro study. *Dent. Res. J.* **2018**, *15*, 111–116.
111. Sonigra, P.; Meena, M. Metabolic Profile, Bioactivities, and Variations in the Chemical Constituents of Essential Oils of the *Ferula* Genus (Apiaceae). *Front. Pharm.* **2020**, *11*, 608649. [[CrossRef](#)]
112. Zhang, H.Q.; Hu, J. Anti-allergic pharmacological action of *Ferula sinkiangensis*. *China J. Chin. Mater. Med.* **1986**, *08*, 49–52.
113. Hu, H.J.; Hu, J. Effect of ferulic acid on metabolic reactions. *J. China Pharm. Univ.* **1991**, *12*, 49–52.
114. Ye, E.B.; Liu, F.; Xiong, Y.J.; Cheng, G.; Yang, X.Z.; Gzo, Y. Anti-inflammatory and immunosuppressive effects of three types of *Ferula* from Xinjiang. *Northwest Pharm. J.* **1993**, *8*, 72–75.
115. Li, X.Y.; Li, G.Y.; Wang, H.Y.; Lou, M.M.; Han, H.Y.; Zhang, K.; Ma, Q.D.; Wang, J.H. Study on Anticoagulation Effects of *Ferula lehmannii* Boiss. *China Pharm.* **2010**, *13*, 1626–1628.
116. Cai, S.S.; Deng, S.Z.; Liu, L.; Sun, D.M. Effects of Ferulic Acid on Ventricular Collagen Remodeling in Mice with Chronic Heart Failure. *Acta Med. Univ. Sci. Technol. Huazhong* **2012**, *41*, 684–687.
117. Jiangsu New Medical College. *Dictionary of Chinese Medicines (Upper Volume)*; Shanghai Scientific & Technical Publishers: Shanghai, China, 1977.
118. Pategule, Y.; Aierxili, T.; Gulishan, M.; Silapu, A. Clinical application and research progress of Uyghur medicine with the Uyghur drug *Ferula* in Uyghur medicine. *J. Med. Pharm. Chin. Minor.* **2011**, *17*, 61–64.
119. Li, X.J.; Jiang, L.; Da, P. Preparation and Investigation of the Pharmacodynamics of Effective Antiulcerative Composition in *Ferula sinkiangensis* KM Shen. *Mod. Chin. Med.* **2007**, *9*, 8–10. [[CrossRef](#)] [[PubMed](#)]
120. Guo, T.T.; Lu, J.; Jiang, L.; Kang, X.L. The effect of *Ferula sinkiangensis* K.M. Shen and its processed products on gastric ulcer model in rats. *J. Xinjiang Med. Univ.* **2013**, *36*, 1463–1466.
121. Xiong, Y.J.; Liu, F.; Ye, E.B.; Hu, J.; Yng, X.Z. Comparison of the gastrointestinal effects of three types of Xinjiang *Ferula*. *J. Xinjiang Med. Univ.* **1993**, *16*, 300–302.
122. Teng, L.; Ma, G.; Li, L.; Ma, L.; Xu, X. Karatavicinol a, a new anti-ulcer sesquiterpene coumarin from *Ferula sinkiangensis*. *Chem. Nat. Compd.* **2013**, *49*, 606–609. [[CrossRef](#)]
123. Xu, M.; Zhang, L.; Li, P.; Wang, C.; Shi, Y. Network pharmacology used to decode potential active ingredients in *Ferula assafoetida* and mechanisms for the application to Alzheimer's disease. *J. Tradit. Chin. Med. Sci.* **2020**, *7*, 199–209. [[CrossRef](#)]

124. Mi, Y.; Jiao, K.; Xu, J.K.; Wei, K.; Liu, J.Y.; Meng, Q.Q.; Guo, T.T.; Zhang, X.N.; Zhou, D.; Qing, D.G.; et al. Kelllerin from *Ferula sinkiangensis* exerts neuroprotective effects after focal cerebral ischemia in rats by inhibiting microglia-mediated inflammatory responses. *J. Ethnopharmacol.* **2021**, *269*, 113718. [[CrossRef](#)]
125. Zhang, W.; Mi, Y.; Jiao, K.; Xu, J.; Guo, T.; Zhou, D.; Zhang, X.; Ni, H.; Sun, Y.; Wei, K.; et al. Kelllerin alleviates cognitive impairment in mice after ischemic stroke by multiple mechanisms. *Phytother. Res.* **2020**, *34*, 2258–2274. [[CrossRef](#)]
126. Yang, Z.M.; Wang, Y.T.; Wu, C.Q.; Li, C.G.; Zhang, H.Y.; Xing, B. A short summary of animal experimental observations on the anti-fertility of the Chinese medicine *Ferula*. *Jilin J. Chin. Med.* **1982**, *04*, 42–43.
127. Zhang, S.; Zhang, Z.Y.; Zhang, Y.; He, W. Termination of pregnancy by *Ferula*. *Chin. Tradit. Herb. Drugs* **1991**, *22*, 458–459.
128. Wang, D.M.; Wang, D.B. The effect of *Ferula* on the isolated uterus of mice and rabbits and its relationship with estrogen and progesterone levels in vivo. *Chin. Tradit. Herb. Drugs* **1986**, *17*, 28–30.
129. Zhong, Z.L.; Zhang, F.; Jiang, Y.H.; Zhang, W.T.; Tong, J.C.; Li, J.; Chu, J.R.; Xie, H.T. Protective effects of Ferulic acid on acute liver injury induced by D-GalN /LPS in mice. *Chin. J. Clin. Pharmacol. Ther.* **2018**, *23*, 1335–1339.
130. Zhang, H.Y.; Shi, H.; Jiang, L.; Zhou, L.L.; Zhou, M.X. Study on Screening of Active Parts of Xinjiang *Ferulae* Resina for Regulating Blood Lipid. *Chin. J. Inf. Tradit. Chin. Med.* **2015**, *22*, 43–46.
131. Ping, Z.; Yoshihisa, T.; Hongquan, D.; Bei, C.; Gisho, H.; Michiho, I.; Yoshio, T.; Olimjon, K.K.; Kuo-Hsing, L. Coumarins and bicoumarin from *Ferula sumbul*: Anti-HIV activity and inhibition of cytokine release. *Phytochemistry* **2000**, *53*, 689–697.
132. Luo, X.; Ma, G.Z.; Dan, M.; Wang, J.; Yu, F.S.; Teng, L.; Wang, C.H. A comparative study of the acute toxicity of two volatile oils of *Ferula alba* and their chemical composition. *Chin. Tradit. Pat. Med.* **2014**, *37*, 1130–1135.
133. Li, J.; Xu, H.Y.; Jia, X.G.; Tian, S.G. Progress in the study of the chemical composition and biological activity of *Ferula fukangensis* KM Shen. *J. Xinjiang Med. Univ.* **2012**, *35*, 1159–1161.
134. Mai, Q.F.; Chen, C.L. Studies on the volatile oils of two types of *Ferula* from Xinjiang. *Chin. Tradit. Herb. Drugs* **1983**, *14*, 10.
135. Ren, Y.; Zhang, H.; Zhou, L. Experimental study on the toxicity of *Ferula sinkiangensis*. *Xinjiang J. Tradit. Chin. Med.* **2014**, *32*, 28–31.
136. Liu, C.X.; Chen, S.L.; Xiao, X.H.; Zhang, T.J.; Hou, W.B.; Liao, M.L. A new concept on quality marker of Chinese materia medica: Quality control for Chinese medicinal products. *Chin. Tradit. Herb. Drugs* **2016**, *47*, 1443–1457.
137. Wang, Z.H. Determination of ferulic acid in *Angelica* and research of quality standards. *Hebei J. Tradit. Chin. Med.* **2012**, *34*, 1058–1063.
138. Zhang, X.L. Determination of ferulic acid in rhizoma chuanxiong by HPLC, China. *Chin. Pharm. Aff.* **2013**, *23*, 469–471.
139. Zhao, X.D.; Chen, D.H.; Si, J.Y.; Shen, L.G. Studies on the phenolic acid constituents from Chinese medicine “Sheng-Ma”, rhizome of *Cimicifuga foetida* L. *Acta Pharm. Sin.* **2002**, *37*, 535–538.
140. Zhang, J.L.; Zhou, Z.H.; Chen, R.Y.; Xie, F.Z.; Yu, D.Q.; Zhou, T.H. Study on chemistry and pharmacology of genus *Ligusticum*. *Chin. Pharm. J.* **2002**, *37*, 14–17.
141. Li, F.; Zhu, X.; Chen, M.; Hou, Y. Study on the Chemical Constituents from Fructus Toosendan. *J. Chin. Med. Mater.* **2010**, *33*, 910–912.
142. Li, D.; Rui, Y.X.; Guo, S.D.; Luan, F.; Liu, R.; Zeng, N. Ferulic acid: A review of its pharmacology, pharmacokinetics and derivatives. *Life Sci.* **2021**, *284*, 119921. [[CrossRef](#)]
143. Rahman, A.-U. *Studies in Natural Products Chemistry*; Elsevier: Amsterdam, The Netherlands, 2018.
144. Appendino, G.; Mercalli, E.; Fuzzati, N.; Arnoldi, L.; Stavri, M.; Gibbons, S.; Ballero, M.; Maxia, A. Antimycobacterial Coumarins from the Sardinian Giant Fennel (*Ferula communis*). *J. Nat. Prod.* **2004**, *67*, 2108–2110. [[CrossRef](#)]
145. Yang, X.W. Bioactive Material Basis of Medicinal Plants in Genus *Ferula*. *Mod. Chin. Med.* **2018**, *20*, 123–144.
146. Liu, C.X. Five-year review on development of quality markers of traditional Chinese medicine. *Chin. Tradit. Herb. Drugs* **2021**, *52*, 2511–2518.
147. Zhou, K.; Xie, J.L.; Li, Y.J.; Wang, Y.L.; Zhang, X.; Liu, C.H. Advances in the study of the chemical composition and pharmacological effects and quality marker prediction of *lysiontus pauciflorus*. *China Pharm.* **2021**, *32*, 1391–1396.
148. Lian, X.Y.; Wu, Y.L.; Feng, H.S. A brief discussion on the medicinal characteristics of bitter medicines and their combinatory effects. *Heilongjiang J. Tradit. Chin. Med.* **1999**, *1*, 52–53.
149. Hao, X.X. Overview of bitter substance research. *J. Huanggang Norm. Univ.* **2008**, *28*, 90–92.
150. Meng, Z.H.; Sun, W.N.; Fu, X.Y.; Zhou, X.; Wan, J. Progress on the mechanism of bitterness and evaluation methods of traditional Chinese medicine. *J. Guangdong Pharm. Univ.* **2016**, *32*, 537–540.
151. Zhou, F.H.; Yi, Z.X.; Luo, X.F. Analysis of the chemical composition of xing(sin) taste herbs. *J. Anhui Agric. Sci.* **2006**, *12*, 2760–2782.
152. China Herb Company. *Commonly Used Chinese Herbs in China*; Science Press: Beijing, China, 1995.
153. Liang, Y.H.; Liu, W.X.; Wang, X.F.; Zhu, Y. Materia medica evidence of the efficacy of Uyghur medicine *Ferula*. *J. Chin. Med. Mater.* **2017**, *40*, 1478–1481.
154. Liu, Y.M.; Shawuti, Y. *Uyghur Pharmacopoeia (First Book)*; Xinjiang People’s Publishing House: Xinjiang, China, 1999; pp. 250–255.
155. Zhang, L.; Fu, X.; Gui, T.; Wang, T.; Wang, Z.; Kullak-Ublick, G.A.; Gai, Z. Effects of Farnesiferol B on Ischemia-Reperfusion-Induced Renal Damage, Inflammation, and NF-kappaB Signaling. *Int. J. Mol. Sci.* **2019**, *20*, 6280. [[CrossRef](#)]
156. Liang, N.; Sun, S.P.; Luo, Y.E.; Qiu, L.Z.; Liu, B. Advances in ferulic acid research. *Heilongjiang J. Tradit. Chin. Med.* **2009**, *38*, 39–40.
157. Hu, Y.Y.; Xu, X.Y. Advances in the chemical and pharmacological study of ferulic acid. *Chin. Tradit. Pat. Med.* **2006**, *28*, 253–255.

158. Xu, J.X.; Xu, J.; Cao, Y.; Zhu, Y.J.; Li, X.Y.; Ge, D.Z.; Ma, L.; Zhang, T.J.; Liu, C.X. Modern research progress on Chinese medicine *Cynanchum otophyllum* Schneid and its predictive analysis of quality markers. *China J. Chin. Mater. Med.* **2021**, *46*, 5486–5495.
159. Gao, T.T.; Ding, X.; Tan, Y.; Wang, H.; Wang, X.F. Determination of Ferulic Acid in Different Parts of *Ferula sinkiangensis* Using HPLC. *Nat. Prod. Res. Dev.* **2013**, *25*, 1237–1239.
160. Wang, D.D.; Liu, H.B.; Song, H.L.; Yang, W.X.; Jia, X.G.; Tian, S.G. Determination of Ferulic acid content in roots and leaves of *Ferula fukanensis* KM Shen. by HPLC. *J. Xinjiang Med. Univ.* **2012**, *35*, 1139–1142.
161. Ainiwaer, A.; Ma, L.Y.; Ma, G.Z. HPLC determination on ferulic acid content of Xinjiang asafetida antiulcer extract. *J. Xinjiang Med. Univ.* **2009**, *32*, 1436–1438.
162. Zhang, H.Y.; Li, W.; Zhou, L.L.; Jiang, L.; Zhou, M.X. Study on Quality Control of Ethyl-acetate Parts of *Ferula sinkiangensis*. *Chin. J. Inf. Tradit. Chin. Med.* **2016**, *23*, 83–86.
163. Zhu, Y.; Zhang, K.; Liang, X.; Tan, Y.; Huang, J.; Wang, J.H. Content Determination of Sesquiterpene Coumarin (DAW22) in *Ferula ferulaeoides* by UPLC. *Chin. J. Exp. Tradit. Med. Formulae* **2016**, *22*, 63–65.

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