



Article Chemical Composition Analysis, Cytotoxic, Antimicrobial and Antioxidant Activities of *Physalis angulata* L.: A Comparative Study of Leaves and Fruit

Jayachithra Ramakrishna Pillai¹, Adil Farooq Wali^{1,*}, Godfred Antony Menezes², Muneeb U. Rehman³, Tanveer A. Wani⁴, Azher Arafah³, Seema Zargar⁵ and Tahir Maqbool Mir⁶

- ¹ Department of Pharmaceutical Chemistry, RAK College of Pharmaceutical Sciences, RAK Medical and Health Sciences University, Ras Al Khaimah 11172, United Arab Emirates; jayachithra@rakmhsu.ac.ae
- ² Department of Microbiology, RAKCOMS, RAK Medical and Health Sciences University, Ras Al Khaimah 11172, United Arab Emirates; godfred@rakmhsu.ac.ae
- ³ Department of Clinical Pharmacy, College of Pharmacy, King Saud University, Riyadh 11451, Saudi Arabia; muneebjh@gmail.com (M.U.R.); aazher@ksu.edu.sa (A.A.)
- ⁴ Department of Pharmaceutical Chemistry, College of Pharmacy, King Saud University, Riyadh 11451, Saudi Arabia; twani@ksu.edu.sa
- ⁵ Department of Biochemistry, College of Science, King Saud University, Riyadh 11451, Saudi Arabia; szargar@ksu.edu.sa
- National Center for Natural Products Research, Research Institute of Pharmaceutical Sciences, School of Pharmacy, University of Mississippi, Mississippi, MS 38677, USA; tmmir@olemiss.edu
- Correspondence: farooq@rakmhsu.ac.ae

Abstract: *Physalis angulata* L. belongs to the family Solanaceae and is distributed throughout the tropical and subtropical regions. *Physalis angulata* leaf and fruit extracts were assessed for in vitro anticancer, antioxidant activity, and total phenolic and flavonoid content. The GC-MS technique investigated the chemical composition and structure of bioactive chemicals reported in extracts. The anticancer activity results revealed a decrease in the percentage of anticancer cells' viability in a concentration- and time-dependent way. We also noticed morphological alterations in the cells, which we believe are related to *Physalis angulata* extracts. Under light microscopy, we observed that as the concentration of ethanolic extract (fruit and leaves) treated HeLa cells increased, the number of cells began to decrease.

Keywords: Physalis angulata; Solanaceae; antioxidant activity; cytotoxic activity; HeLa; MCF-7; DLD-1

1. Introduction

Plants with medicinal properties serve as the basic treatment for many ailments in cultural diversities worldwide. These medicinal plants exhibit various activities against many major diseases and their active constituents. In recent years, phytoscience has made significant contributions to modern medicine. As a result, medicinal plants are the primary source of treatment for a wide range of health issues. Another reason is that herbal treatments are readily available and accessible and are very inexpensive. In several nations, people in rural areas still rely on herbal medicines to treat all of their health problems. Phytomedicine is widely used in modern medicine to treat various disorders, including diabetes and cancer. Many studies have been published on the use of phytomedicine in cancer treatment. According to the World Health Organization (WHO), herbal medicine is used by 60% of the world's population, and about 80% of the people in emerging nations rely on it nearly entirely for their basic health care requirements. [1–3].

Physalis angulata (Figure 1) belongs to the family Solanaceae and includes more than 120 species with different herbal characteristics and perennial habits [4]. This plant is a bushy annual herb that grows to about 50 cm in height, and is glabrous or with minute



Citation: Ramakrishna Pillai, J.; Wali, A.F.; Menezes, G.A.; Rehman, M.U.; Wani, T.A.; Arafah, A.; Zargar, S.; Mir, T.M. Chemical Composition Analysis, Cytotoxic, Antimicrobial and Antioxidant Activities of *Physalis angulata* L.: A Comparative Study of Leaves and Fruit. *Molecules* 2022, 27, 1480. https://doi.org/10.3390/ molecules27051480

Academic Editors: Joanna Kapusta-Duch, Barbara Borczak and Teresa Leszczyńska

Received: 7 February 2022 Accepted: 17 February 2022 Published: 22 February 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). simple hairs [5]. It has bell-shaped flowers and balloon-like fruit which drops downwards. The fruit of *Physalis angulata* are edible and very tasty [6]. This herb is distributed widely throughout the tropical and subtropical regions. It is known by different names such as camapu; cut leaf groundcherry; wild tomato, winter cherry, etc. A literature survey revealed that *Physalis angulata* could be used as an herbal medicine for many ailments. It is widely known to be a stimulant for the immune system. It can be used in food, mainly in preparing sauces [7,8]. In several regions of the World, the extracts or infusions of this plant are used as anti-malarial, anti-asthmatic, and for dermatitis. The isolated phytoconstituents of *Physalis angulata* exhibited antitumor activities against many cancer cell lines [9,10] by in vitro studies. It has been traditionally used for antipyretic purposes in Japan. This plant has been used in intestinal and digestive problems and for many other ailments like sores, boils, cuts, etc. The leaves of the plant are used in salads. Studies proved that *Physalis* angulata exhibits many therapeutic activities like antiallergic, antiasthmatic, antileishmanial, antmalarial, and immunomodulatory activity [11–13]. The phytochemical investigation of Physalis angulata showed many primary and secondary metabolites such as carbohydrates, minerals, vitamins, lipids, and phytosterols. The entire plant contains various steroidal lactones which belong to physaline, and withanolide such as physalins A–I, physagulin A–G, withangulatin A, and withanolide T. Withanolides have a C-28 ergostane-type steroid structure with a δ -lactone group at C-22 and C-26. It also contains flavonol glycoside named myricetin 3-O-neohesperidoside [14–18].



Figure 1. *Physalis angulata* (fruit and leaves) belongs to the family Solanaceae, includes more than 120 species.

Cancer is the term used for diseases when abnormal cells multiply without any control in any body organ or tissue and attack the nearby body parts. If the spread is uncontrollable, it can cause death. It is considered the second foremost reason for death around the world. The common types of cancer found mostly in men are lung, liver, colorectal, prostate, and stomach. In women, breast, lung, colorectal, cervical, and thyroid cancers are mostly found [19].

Much recent research has focused on plant-derived chemoprotective constituents. These active constituents can act as target specific for malignant cells or can prevent cancerous cell multiplication [20]. The current cancer treatment is associated with severe side effects, including the damage of normal body cells and many others. Therefore, the need for safe drugs or methods for cancer treatment is in high demand. The current study on the leaves and fruit of *Physalis angulata* includes the cytotoxic activity on the breast cancer, colorectal and cervical cancer cell lines.

2. Results

2.1. Preliminary Phytochemical Screening

The preliminary phytochemical screening of the *Physalis angulata* fruit and leaves proved that it contains various secondary metabolites such as alkaloids, glycosides, flavonoids, tannins, and phenolics.

2.2. Anti-Oxidant Activities

The DPPH and hydrogen peroxide assays of all the extracts of the fruit and leaves of *Physalis angulata* exhibited remarkable radical scavenging activity. The standard used to compare the results was ascorbic acid.

Even though all the extracts showed activity, the ethanolic extracts of *Physalis angulata* leaves and fruit had very good activity for both the assays. The ethanolic extract of the leaves showed 85% and 81% radical scavenging activity upon a standard value of ascorbic acid with 98% in DPPH and hydrogen peroxide assays, respectively, as illustrated in Figure 2A,B.

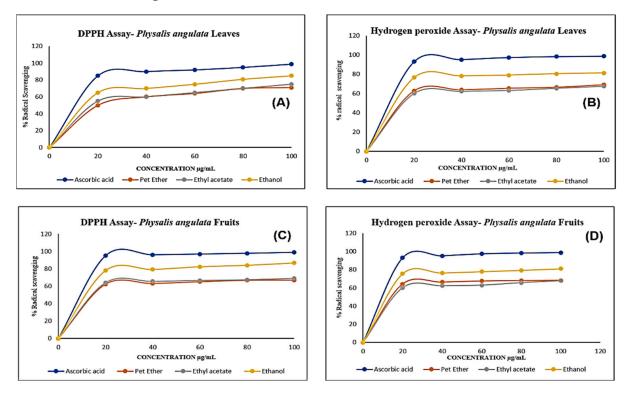


Figure 2. The antioxidant activity of Physalis angulata leaves and fruit extracts (**A**–**D**). The dosedependent DPPH and H_2O_2 scavenging activity of leaves and fruit extracts relative to that of L-ascorbic acid. All results are presented as the mean \pm SD (n = 3).

Similarly, the ethanolic extracts of the fruit exhibited 86% and 80.9% radical scavenging activity upon a standard value of ascorbic acid with 98% in DPPH and hydrogen peroxide assays, respectively, which is illustrated in Figure 2B,D.

2.3. Total Phenolic Content (TPC) and Total Flavonoid Content (TFC)

The TPC and TFC of all the leaves and fruit extracts were evaluated. The TPC of all the extracts of the leaves were 54.4 ± 3.4 , 96.7 ± 4.5 , and 140.65 ± 3.8 GAE/g respectively, with a calibration curve of R2 0.998, whereas the TFC with R2 0.998 was 210.32 ± 3.6 , 238.24 ± 4.4 , and 370.64 ± 4.33 QE/g for petroleum ether, ethyl acetate, and ethanolic extracts, respectively (Table 1).

| Extract | TPC (* mg GAE/g) | TFC (** mg QE/g) |
|-----------------|------------------|-------------------|
| Petroleum ether | 54.4 ± 3.4 | 210.32 ± 3.6 |
| Ethyl acetate | 96.7 ± 4.5 | 238.24 ± 4.3 |
| Ethanol | 140.65 ± 3.8 | 370.64 ± 4.33 |

Table 1. Total phenolics and flavonoids content of *Physalis angulata* Leaves.

* Total phenolics content is expressed in terms of Gallic acid equivalent (μg of GA/g). ** Total flavonoids content is expressed in terms of quercetin equivalent (μg of QE/g).

The TPC of all the three extracts of the fruit were 75.3 ± 4.6 , 69.4 ± 3.6 , and 106.5 ± 3.5 GAE/g, respectively, with a calibration curve of R2 0.998, whereas the TFC with R2 0.998 was 40.78 ± 4.3 , 100.83 ± 4.2 , and 130.4 ± 2.6 QE/g for petroleum ether, ethyl acetate, and ethanolic extracts, respectively (Table 2).

Table 2. Total phenolics and flavonoids content of *Physalis angulata* fruit.

| Extract | TPC (mg * GAE/g) | TFC (mg ** QE/g) |
|-----------------|------------------|------------------|
| Petroleum ether | 75.34 ± 4.6 | $40.\ 78\pm4.3$ |
| Ethyl acetate | 69.41 ± 3.6 | 100.83 ± 4.2 |
| Ethanol | $106.54\pm\!3.5$ | 130.48 ± 2.6 |

* Total phenolics content is expressed in terms of Gallic acid equivalent (µg of GA/g). ** Total flavonoids content is expressed in terms of quercetin equivalent (µg of QE/g).

2.4. Chemical Profiling of Ethanolic Extracts (Fruit and Leaves) of Physalis Angulata by Using GC-MS Analysis

The results of a GC-MS analysis of *Physalis angulata* L. ethanolic extracts (fruit and leaves) led to the identification of several chemicals. Table 3 (ethanolic extract fruit) and Table 4 (ethanolic extract leaves) compares the retention time, molecular formula, and molecular mass (m/z) of chemical compounds with various databases, literature surveys, and research publications. The chemical components found in the ethanolic extracts (fruit and leaves) of *Physalis angulata* L. presented in Tables 3 and 4 are based on the identification data. Supplementary Materials Figures S2 and S3 show a representative full scan chromatographic profile of *Physalis angulata* ethanolic extract (leaves and fruits) (Supplementary Materials file).

Table 3. Chemical profiling of ethanolic extract (fruit) of *Physalis angulata* by using GC-MS analysis.

| Retention Time (Min) | Molecular Formula | Molecular Mass (g/mol) | Peak Area (%) | Compound Name | Structure |
|-------------------------|--|---------------------------|------------------|----------------------|-----------|
| 37.743 | C ₁₂ H ₂₄ O ₂ | 200.3178 | 0.469 | Lauric acid | |
| 52.465 | C ₁₇ H ₃₄ O ₂ | 270.4504 | 0.116 | Methyl hexadecanoate | |
| 54.198 | C ₁₆ H ₃₂ O ₂ | 256.4009 | 0.134 | Palmitic acid | |

| Retention Time (Min) | Molecular Formula | Molecular Mass (g/mol) | Peak Area (%) | Compound Name | Structure |
|-------------------------|--|---------------------------|------------------|---|-----------|
| 56.552 | C ₁₉ H ₃₄ O ₂ | 294.4721 | 0.194 | Linoleic acid, methyl ester | |
| 57.885 | C ₁₈ H ₃₂ O ₂ | 280.444 | 1.774 | Linoleic acid | OH OH |
| 64.580 | C ₂₄ H ₃₈ O ₄ | 390.5566 | 0.072 | 1,2-Benzenedicarboxylic acid, diisooctyl ester | |
| 65.676 | C ₂₅ H ₃₄ O ₇ | 446.5181 | 0.203 | (22R)-6α,11β,21- Trihydroxy-16α,17α- propylmethylenedioxypregna- 1,4-diene-3,20-dione | |
| 68.131 | C ₂₀ H ₃₂ O | 288.4702 | 0.215 | 5-(7a-Isopropenyl-4,5- dimethyl-octahydroinden- 4-yl)-3-methyl-pent-2-enal | |
| 68.199 | C ₃₂ H ₅₄ O ₃ | 486.8045 | 0.134 | Acetic acid, 13-hydroxy- 4,4,6a,6b,8a,11,11,14b- octamethyldocosahydropicen- 3-yl ester | |

| Retention Time (Min) | Molecular Formula | Molecular Mass (g/mol) | Peak Area (%) | Compound Name | Structure |
|-------------------------|---|---------------------------|------------------|--|-----------|
| 69.396 | C ₂₈ H ₄₄ N ₂ O ₇ | 520.6599 | 0.220 | 1-Pyrrolidinebutanoic acid, 2-[(1,1- dimethylethoxy)carbonyl]- α-nitro-, 2,6-bis(1,1-dimethylethyl)- 4 -methoxyphenyl ester, [S-(R*,R*)]- | |
| 70.637 | C ₂₉ H ₄₄ O ₅ | 472.6572 | 2.763 | Cholest-5-ene-16,22-dione, 3β,26-dihydroxy-, 3-acetate, (20S,25R)- | |
| 71.299 | C ₃₀ H ₅₀ O ₂ | 442.7171 | 0.291 | Ergost-5-en-3-ol, acetate, (3β,24R)- | |
| 72.727 | C ₃₂ H ₅₂ O ₂ | 468.7550 | 2.244 | Lup-20(29)-en-3-ol, acetate, (3β)- | |
| 73.330 | $C_{29}H_{50}O_2$ | 430.7073 | 3.853 | α-Tocopherol | |

Table 4. Chemical profiling of ethanolic extract (leaves) of *Physalis angulata* by using GC-MS analysis.

| Retention Time | Molecular | Molecular Mass | Peak Area | Compound | Structure |
|----------------|-----------|----------------|-----------|---------------------------|-----------|
| (Min) | Formula | (g/mol) | (%) | Name | |
| 49.849 | C18H36O | 268.4780 | 3.51 | Hexahydrofarnesyl acetone | |

| Retention Time (Min) | Molecular Formula | Molecular Mass (g/mol) | Peak Area (%) | Compound Name | Structure |
|-------------------------|----------------------|---------------------------|------------------|--|---|
| 51.072 | C20H40O | 296.5315 | 2.055 | 3,7,11,15-Tetramethyl-2- hexadecen-1-ol | но |
| 57.095 | C20H40O | 296.5315 | 10.192 | Phytol | |
| 58.548 | C18H34O2 | 282.4628 | 35.017 | Oleic Acid | |
| 58.896 | C18H36O2 | 284.4772 | 5.908 | Octadecanoic acid | ~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~ |
| 58.998 | C23H39NO2 | 361.5627 | 0.747 | 3-[(1,5-Dimethyl- hexylamino)-methyl]-5,8a- dimethyl-3a,5,6,7,8,8a,9,9a- octahydro-3H-naphtho[2,3- b]furan-2-one | H H H H H H H H H H H H H H H H H H H |
| 59.848 | C18H32O2 | 280.4464 | 2.673 | 9,12-Octadecadienoic acid (Z,Z)- | O, OH |
| 60.442 | C14H24O2 | 224.3391 | 0.185 | Linalyl isobutyrate | |
| 62.796 | C21H38O4 | 354.5247 | 0.281 | 9,12-Octadecadienoic acid (Z,Z)-, 2,3-dihydroxypropyl ester | HO OH |

| Retention Time (Min) | Molecular Formula | Molecular Mass (g/mol) | Peak Area (%) | Compound Name | Structure |
|-------------------------|----------------------|---------------------------|------------------|--|---|
| 62.906 | C21H40O3 | 340.5419 | 0.370 | Oleic acid, 3-hydroxypropyl ester | к, j, |
| 63.586 | C22H40O2 | 336.5524 | 0.765 | Butyl 9,12-octadecadienoate | |
| 63.688 | C21H40O4 | 356.5406 | 2.726 | 9-Octadecenoic acid (Z)-, 2-hydroxy-1- (hydroxymethyl)ethyl ester | |
| 64.095 | C21H40O3 | 340.5415 | 0.605 | Glycidyl stearate | |
| 64.554 | C24H38O4 | 390.5574 | 0.205 | 1,2-Benzenedicarboxylic acid, diisooctyl ester | |
| 66.304 | C42H88O5Si5 | 813.5739 | 0.607 | 5β-Cholestane- 3α,7α,12α,24α,25-pentol TMS | HO HO HO HO HO HO HO HO HO HO HO HO HO H |
| 68.708 | C30H50 | 410.7198 | 0.940 | Squalene | |

| Retention Time (Min) | Molecular Formula | Molecular Mass (g/mol) | Peak Area (%) | Compound Name | Structure |
|-------------------------|----------------------|---------------------------|------------------|--|-----------|
| 69.040 | C40H66 | 546.9531 | 0.639 | Lycopersene | |
| 69.413 | C28H44N2O7 | 520.6594 | 0.750 | 1-Pyrrolidinebutanoic acid, 2-[(1,1- dimethylethoxy)carbonyl]- α-nitro-, 2,6-bis(1,1-dimethylethyl)- 4-methoxyphenyl ester, [S-(R*,R*)]- | |
| 70.654 | C30H44O2 | 436.6709 | 0.180 | Anthiaergosatn-5,7,9,22- tetraen, 3-acetoxy- | |
| 71.180 | C29H48O | 412.6919 | 0.325 | Stigmasterol | HO |
| 71.308 | C30H50O2 | 442.7172 | 1.029 | Ergost-5-en-3-ol, acetate, (3β,24R)- | |
| 71.716 | C31H50O2 | 454.7287 | 0.240 | Stigmasta-5,22-dien-3-ol, acetate, (3β,22Z)- | |

| Retention Time (Min) | Molecular Formula | Molecular Mass (g/mol) | Peak Area (%) | Compound Name | Structure |
|-------------------------|----------------------|---------------------------|------------------|----------------------|-----------|
| 72.744 | C29H48 | 396.6925 | 0.699 | Stigmastan-3,5-diene | |
| 73.695 | C29H50O2 | 430.7075 | 0.716 | α-Tocopherol | |

2.5. Elemental Analysis

The dried leaf and fruit samples of *Physalis angulata* were analyzed by ICP OES. The relative standard deviation for the different samples was determined for the precision of the procedure. *Physalis angulata* leaf and fruit contains various macro and micronutrients. The leaf is rich in calcium and potassium when compared to fruit. The leaf contains 24,503 mg/Kg calcium, iron 3338.8 mg/Kg, and 35,725 mg/Kg potassium. In comparison, the fruit contains 4143 mg/Kg calcium, 478.9 mg/Kg iron, and 32,856 mg/Kg potassium (Table 5).

Table 5. Elemental analyses of Physalis angulata leaves and fruit by ICP-OES.

| | Mac | roelements | |
|----------|------------|--------------------------------|--------------------------------|
| SI No | Elements | Physalis Angulata Leaf (mg/kg) | Physalis Angulata Fruit (mg/kg |
| 1 | Calcium | 24,503 | 4143 |
| 2 | Chromium | 4.0 | 1.3 |
| 3 | Potassium | 35,725 | 32,856 |
| 4 | Magnesium | 4551 | 2823 |
| 5 | Phosphorus | 4642.5 | 5836.4 |
| | Mic | roelements | |
| | Elements | Physalis Angulata Leaf (mg/kg) | Physalis Angulata Fruit (mg/kg |
| 1 | Boron | 30.0 | 20.1 |
| 2 | Cobalt | <0.1 | < 0.1 |
| 3 | Copper | 27.7 | 9.8 |
| 4 | Iron | 3338.8 | 478.9 |
| 5 | Manganese | 48.6 | 15.7 |
| 6 | Molybdenum | 2.3 | 1.2 |
| 7 | Sodium | 1213 | 398.6 |
| 8 | Nickel | 4.1 | 2.5 |
| 9 | Vanadium | 1.5 | 1.2 |
| 10 | Zinc | 17.5 | 17.5 |
| 11 | Aluminum | 4713.8 | 716.2 |
| | Oth | er elements | |
| 1 | Silver | 0.3 | 0.5 |
| 2 | Arsenic | <0.1 | <0.1 |
| 3 | Barium | 76.1 | 10.0 |
| 4 | Beryllium | <0.1 | <0.1 |
| 5 | Cadmium | 0.4 | 0.4 |
| 6 | Lead | 1.4 | <0.1 |
| 7 | Tin | <0.1 | <0.1 |
| 8 | Strontium | 59.8 | 11.5 |

2.6. Cytotoxic Studies

The cytotoxic activity of ethanolic extracts of leaves and fruit of *Physalis angulata* was determined by the MTT cell viability assay using HeLa, DLD-1, and MCF-7 cell lines. The cell viability was decreased with both extracts depending on the dose. The MTT cell viability assay was carried out in different fruit and leaf extracts (6.25, 12.5, 25. 50, and 100 μ g/mL) for all three cell lines. The calculated cell viability was decreased with an increase in extract concentration. The percentage viability of *Physalis angulata* leaf extracts at 100 μ g/mL was observed at 46.23, 33.66, and 51.54 for DLD-1, HeLa, and MCF-7 cell lines, respectively (Figure 3). The percentage viability of *Physalis angulata* fruit extracts at 100 μ g/mL was observed 70, 69.41, and 65.27 for DLD-1, HeLa, and MCF-7 cell lines, respectively. The fruit extracts' LC50 value (calculated using ED50 PLUS V1.0 software) were 188, 167, and 157 μ g/mL for DLD-1, HeLa, and MCF-7 cell lines, respectively. The fruit extracts of *Physalis angulata* were 90, 44, and 100 μ g/mL for DLD-1, Hela, and MCF-7 cell lines, respectively. The leaf extracts of *Physalis angulata* were 90, 44, and 100 μ g/mL for DLD-1, Hela, and MCF-7 cell lines, respectively. The same for the leaf extracts of *Physalis angulata* were 90, 44, and 100 μ g/mL for DLD-1, Hela, and MCF-7 cell lines, respectively. The MTT assay of the ethanolic extracts of *Physalis angulata* fruit and leaves revealed that the leaf extracts have significant cytotoxic activity on different cell lines.

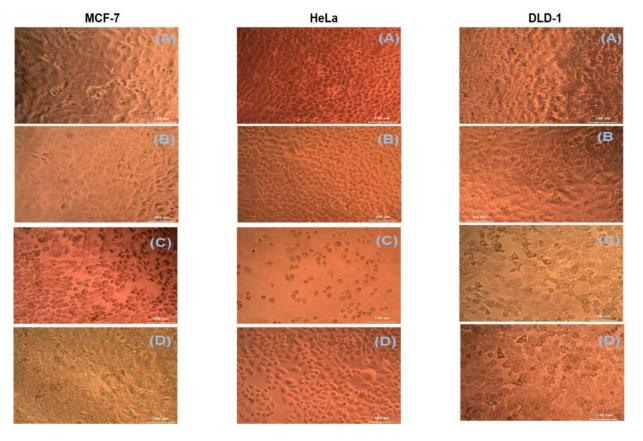


Figure 3. Effect of ethanolic extract of *Physalis angulata* leaves and fruit on different cancer cell lines. Impact of *Physalis angulata* extracts on morphological changes and viability of human cancer cells MCF-7, HeLa, and DLD-1. After 72 h of incubation, the photomicrograph shows the effect of different concentrations of extract of *Physalis angulata* leaf and fruit on MCF-7, HeLa, and DLD-1 cells. Photomicrograph (**A**): Blank control; (**B**): Positive control; (**C**): Low concentration; (**D**): High concentration. The data display the mean of the test as measured in triplicate. One-way ANOVA analysis to compare between control, 6.25 and 100 μg/mL of the extract.

2.7. Antibacterial Potential

Using the serial dilution technique, *Physalis angulata* leaves and fruit extracts were evaluated for their antibacterial activity. The results show that *Physalis angulata* leaves and fruit extracts possess moderate activity on these pathogens. Among the MIC/MBC

of the leaf extracts against *E. coli* (ATCC 25922), the ethyl acetoacetate extracts possess moderate activity with less concentration. Similar results were observed for fruit extracts also. Among the MIC/MBC of the leaf and fruit extracts against *S. aureus* (ATCC 25923), the ethyl acetoacetate extracts also showed moderate activity against this pathogen (Table 6). Figures S4 and S5 represents the MIC values plates (Supplementary Materials file). All the results were compared against the standard gentamycin.

| | Physalis Ang | ulata Leaves | Physalis An | gulata Fruit |
|--------------------------|--------------|--------------|--------------|--------------|
| | | E. coli | | |
| Extract | MBC (mg) | MIC (mg) | MBC (mg) | MIC (mg) |
| Petroleum ether Ethyl | 5 1.25 | 10 2.5 | 1.25 1.25 | 2.5 2.5 |
| acetoacetate Ethanol | 5 | 10 | 5 | 10 |
| | | S. aureus | | |
| Petroleum ether | 5 | 10 | 5 | 10 |
| Ethyl acetoacetate | 1.25 | 2.5 | 5 | 10 |
| Ethanol | 2.5 | 5 | 5 | 10 |

Table 6. Antibacterial activity of *Physalis angulata* leaves and fruit.

MIC-Minimum inhibitory concentration (mg/mL), MBC-Minimum bactericidal concentration (mg/mL).

3. Discussion

The application of medicinal plants for managing various ailments is worldwide now. It varies from traditional to popular medicines of each country using standardized medicinal plant extracts. Natural medicines possess a very high therapeutic index against different tumor cells, and they have been proved to control the abnormal multiplication of cells [21,22]. This study gives information about the phytoconstituents, TFC, and TPC content of *Physalis angulata* leaf and fruit extracts with the in-vitro antioxidant, antimicrobial and cytotoxic activities.

Free radicals are the major reason for many malignancies, diabetes, degenerative diseases, etc. [23]. These free radicals are produced due to the lack of a natural antioxidant defense mechanism [24]. Many phytonutrients from plants have shown radical scavenging activity against these free radicals [25]. These phytonutrients are in high demand now [24]; the phytonutrient flavonoids are polyphenols. Almost all types of plants contain flavonoids [23]. Recent studies show that flavonoids and phenolics are powerful antioxidants that improve the immune system. Diet, rich in flavonoids, can prevent cancer, neurodegenerative, and cardiovascular diseases [18,26]. Many studies show that women with higher levels of flavonoid intake through diet will have a lower risk of developing breast cancer [27]. Many studies have proved that the flavonoids and phenolic content in the plant material play an important role against oxidative stress [26]. Scavenging activities are facilitated by the presence of the number and position of hydroxyl and methoxy groups in the phenolic rings [28].

DPPH and hydrogen peroxide assays were used in this study to assess the antioxidant activities of the various extracts. These tests are very common and accurate to determine the radical scavenging activity due to the hydrogen donating capacity to the free radicals. The ethanolic extracts of *Physalis angulata* leaves and fruit showed significant antioxidant activity. The total phenolic content and flavonoid content of the leaf extracts of *Physalis angulata* was higher than that of the fruit extracts. In the leaf extracts, ethanolic extracts showed the maximum TPC and TFC. The significant antioxidant activity shown by the leaf extracts may be because of the higher levels of phenolics and flavonoids present in them [29,30].

Bioactive components of medicinal plants have been used to manage cancer for many years [31]. The cytotoxic activity of the herbal drugs against various types of cancer has

been established by several clinical studies and phytochemical screening [32,33]. The main concern with modern cancer chemotherapy is the drug resistance that develops during the treatment [34]. Many studies have shown that natural compounds separated or derived from plants have a significant potential for cancer prevention [27]. The cytotoxic activities of polyphenols have been reported in various studies against different cancer cells [35]. The polyphenols are capable of initiating apoptosis, which is the reason for their anticancer activities [36].

Since the TPC and TFC were relatively high in the ethanolic extracts, and due to this extract's significantly high antioxidant activity, we chose the ethanolic extract for the cytotoxic studies. The ethanolic extracts of *Physalis angulata* fruit and leaves were evaluated against three different cell lines such as HeLa, DLD-1, and MCF-7. The MTT assay method was used to detect the cytotoxic activity of the plant extracts. The percentage cell viability decreased significantly with increased sample concentration with the ethanolic extracts of fruit and leaves of *Physalis angulata*. When comparing the results of fruit and leaves, the ethanolic extracts of the leaves of Physalis angulata showed more significant cytotoxic activity than fruit extracts. When tested against HeLa, DLD-1 and MCF-7 cell lines, the ethanolic extracts of the leaf showed excellent activity with HeLa with a low LC₅₀ ($44 \pm 10\mu$ g/mL) value.

Our study shows that the leaves and fruit of *Physalis angulata* are rich with micro and macro elements, which are highly important for the body as medicine and food supplements. Many studies have shown that these nutrients can help the body balance blood pressure, normalize increased cholesterol levels, protect the liver, promote immunity, etc. [37,38]. Along with the phytoconstituents present in the leaves and the fruit, this also might be the reason for the therapeutic activities of *Physalis angulata*.

The estimation of *Physalis angulata* leaves' antibacterial activity and fruit extracts demonstrated reasonable activity against both Gram-negative and Gram-positive bacteria [39]. Many studies showed that the polyphenols present in the plant possess antibacterial activity. *Physalis angulata* plant also contains a good amount of polyphenols [40]. These reports correlate with the current findings. The polyphenols present in *Physalis angulata* can cause its antibacterial activity [41].

Various chemical compounds were identified using GC-MS in the methanolic extract of Physalis angulata leaves and fruit. The major chemical compounds were identified from the fruit extract linoleic acid, such as Cho-lest-5-ene-16,22-dione, 3β,26-dihydroxy-, 3-acetate, (20S, 25R)-, Lup-20(29)-en-3-ol, acetate, (3β) - and α -Tocopherol, whereas Hexahydrofarnesyl acetone, 3,7,11,15-Tetramethyl-2-hexadecen-1-ol, Phytol, Oleic Acid, Octadecanoic acid, 9,12-Octadecadienoic acid (Z,Z)-, 9-Octadecenoic acid (Z)-, 2-hydroxy-1-(hydroxymethyl) ethyl ester, and Ergost-5-en-3-ol, acetate, $(3\beta, 24R)$ were identified from the leaves extract. In this study, the higher antioxidant and antimicrobial activities of the methanol extract could be correlated to a higher number of bioactive compounds, including Lauric acid, Oleic acid, Hexahydrofarnesyl acetone, Oleic acid, n-hexadecanoic acid, Phytol, and α -Tocopherol. Similarly, the occurrence of these compounds is also evidenced in many medicinal plant leaves extracted with methanol [42]. Lauric acid is a saturated fatty acid reported to have antimicrobial activity against various bacterial strains [42]. Linoleic acid is polyunsaturated fatty acid reported in many studies to inhibit the growth of all the strains of Gram-positive bacteria [43]. One of the most naturally occurring biologically active forms of Vitamin E is alpha-tocopherol. It's a known antioxidant. The antibacterial and cytotoxic activities of α tocopherol are reported in research studies [44,45]. Hexadecanoic acid and its derivatives exhibit strong antimicrobial and anti-inflammatory activity [46,47]. Phytol is a natural antioxidant and possesses various pharmacological importance. Phytol is an important diterpene that possesses antimicrobial, antioxidant, and anticancer activities [48]. Oleic acid is an omega-9 fatty acid produced in the body. It has been reported that the oleic acid present in various plants can exhibit apoptotic and antibacterial activity [49,50]. The compound stigmasterol in hexane extract is also reported to have various biological activities [51].

4. Materials and Methods

4.1. Plant Collection and Preparation of the Plant Extract

The *Physalis angulata* L. (Syn.: P. minima L.; P. longifolia) plant was collected from Kerala, India. The plant was then authenticated by Dr. T. Sabu, JNTBGRI, Thiruvananthapuram, India. A voucher herbarium specimen (No. 852016) was deposited in JNTBGRI herbarium (TBGT). The leaf and fruit of *Physalis angulata* were used for the current study. The leaves and fruit were separated from the plant, shade dried, and powdered. 60 g each of leaf and fruit powder were taken in two different containers, and 500 mL of petroleum ether was added to each. The extraction was done by the ultra-sonication method. After sonication for 1 h, the extract was collected and filtered. The leaf and fruit powders were dried and extracted again with 500 mL of ethyl acetate under sonication. After extraction with ethyl acetate, it was filtered again, and the extracts were collected. The dried plant parts were extracted again with ethanol and collected the extract. After the solvent evaporation, the leaf extracts weighed around 5 g, and the fruit extracts weighed around 4 g. These extracts were used for various activities and phytochemical screening. The flowchart of Section 4 is represented by Figure S1, (Supplementary Materials file).

4.2. Preliminary Phytochemical Evaluation

The extracts of *Physalis angulata* fruit and leaves were subjected to preliminary phytochemical screening using the standard protocol [52]. It was found that various major secondary metabolites are present in the leaf and fruit.

4.3. In Vitro Anti-Oxidant Screening

The in vitro antioxidant screening of the leaf and fruit extracts of *Physalis angulata* was done by spectrophotometric methods such as DPPH and Hydrogen peroxide assays. The spectrophotometric assays depend on the reaction between the free radical and that of the antioxidant moiety that can donate a hydrogen atom. Ascorbic acid was used as the standard.

4.3.1. DPPH Assay

The DPPH assay for the radical scavenging activity of both the leaves and the fruit extracts of Physalis angulata was done using the method described by Adil Wali et al. [53]. When the DPPH radical reacts with the hydrogen donor from the extracts, its color disappears. The absorbance was measured spectrophotometrically at 517 nm (Shimadzu, UV-1800). For each sample, the readings were repeated three times. A comparison of all these results was done with that of standard.

4.3.2. Hydrogen Peroxide Assay

The hydrogen peroxide assay for the radical scavenging ability of both the leaves and the fruit extracts of *Physalis angulata* was done based on the method of Marchelak et al. [54]. The absorbance was measured spectrophotometrically at 230 nm (Shimadzu, UV-1800), and the blank solution was prepared with phosphate buffer without hydrogen peroxide. The readings were taken three times for each sample. A comparison of all these results was done with that of standard.

4.4. Determination of TPC

The leaves and the fruit extracts of Physalis angulata were tested for TPC (Total Phenolic Content) by the Folin- Ciocalteau method [55]. The calibration curve was plotted by mixing 1 mL of 0.5, 0.75, 1.0, 1.25, and 1.5 mg/mL Gallic acid solution with 2.5 mL of Folin- Ciocalteau reagent and 2.5 mL of sodium carbonate solution. The solutions were mixed thoroughly and kept for 30 min. The absorbance was measured at 725 nm using UV-Spectrophotometer (Shimadzu, UV-1800). 0.5 mL of all the extracts of both leaf and fruit of Physalis angulata (5 mg/mL) was mixed with the same reagents. After keeping for 1 h, the absorbance was measured. The TPC is calculated as follows:

Total phenolic content = cV/m

The total phenolic content is expressed as Gallic Acid Equivalent or GAE (mg/g),

c = gallic acid concentration calculated from the calibration curve

V = extract volume

m = mass of the plant extract (g).

Three readings were taken for each sample.

4.5. Determination of TFC

The TFC (the Total Flavonoid Content) of the leaves and the fruit extracts of *Physalis angulata* was determined spectrophotometrically by following the aluminum chloride method [56]. The calibration curve was generated by using the standard quercetin. The standard was prepared by mixing 1 mL each of 0.5, 0.75, 1.0, 1.25, and 1.5 mg/mL quercetin solution with ethanol and 10% aluminum chloride solution. This was mixed with 1 M potassium acetate and distilled water. The absorbance of these samples was taken after incubation for 3 min using a Shimadzu UV spectrophotometerUV-1800 at 415 nm. All the extracts (0.5 mL) of both the leaf and fruit of Physalis angulata were taken, and absorbance was measured. The TFC is calculated as follows:

Total flavonoid content (TFC) = cV/m

The total phenolic content is expressed as Quercetin equivalent (QE),

c = Quercetin concentration calculated from the calibration curve

V = extract volume

m = mass of the plant extract (g).

All the readings were taken in triplicates

4.6. Chemical Profiling of Ethanolic Extracts (Fruit and Leaves) of Physalis Angulata by Using GC-MS Analysis

The phytochemical investigation of ethanoic extracts (fruit and leaves) were performed on GC-MS equipment and tgas chromatography (Agilent7890 A GC) was done with a DB 5MS 30 m \times 0.250 mm Diameter \times 0.25 Micro Meter Thickness) interfaced with a 5675C Inert MSD with a Triple-Axis detector used for the GC-MS analysis. The carrier gas was helium, set to a column velocity flow of 1.0 mL/min.

Other GC-MS conditions include a 150 °C ion source, a 280 °C interface, a 7.0699 psi pressure, a 1.8 mm out time, and a 3 μ L injector in split mode with a split ratio 1:50 and a 280 °C injection temperature. The column temperature was set to 50 °C for 5 min before gradually increasing to 80 e V at a rate of 4 °C/min. The temperature was raised to 280 °C at a 20 °C/min rate for 5 min. It took 75 min to complete the elution. All information, acquisitions, and evaluations were regulated using the National Institute of Standards and Technology (NIST 08) Spectral Database Library to validate the chemical compounds.

4.7. Elemental Analysis by ICP-OES

The elemental analysis of the dried leaf and fruit of Physalis angulata was done by a Spectrogenesis ICP-OES (Inductively coupled Plasma-Optical Emission spectrometer) After weighing, around 3 g of the dried fruit and leaf powder of Physalis angulata were taken for the analysis. These samples were digested using concentrated nitric acid (3 mL). It was then heated on a hot plate to digest the organic matter. After cooling, the samples were treated with concentrated hydrochloric acid (1 mL) and all of the samples were heated again and then and then cooled to room temperature. After filtration, these samples were transferred to a volumetric flask and, with distilled water, made up to the volume. This sample is used as the stock to evaluate the micro and macronutrients by ICP-OES. Proper dilutions were made, and a calibration curve was plotted accordingly. By free aspiration, the diluted samples were introduced into the ICP-OES. By associating with that of the prepared standard, the intensities of the micro and macro elements present in the test samples were calculated.

4.8. Evaluation of In-Vitro Cytotoxic Activity

The cytotoxic evaluation of the ethanolic extracts of the fruit and leaves of *Physalis* angulata was done by MTT assay. HeLa (Cervical carcinoma), DLD-1 (Human Colorectal Adeno carcinoma) and MCF-7 (Human Breast Adeno carcinoma) cells were procured from National Centre for Cell Sciences (NCCS), Pune, India and maintained Dulbecco's modified Eagles medium, DMEM (Sigma Aldrich, St. Louis, MO, USA). This method uses a colorimetric method to assess the metabolic activity of cells [57]. This allows us to determine the number of live cells, their growth, and cytotoxicity. This method is based on metabolically active cells reducing tetrazolium salt, yellow color, to purple color crystals. The ethanolic extract of the leaf and fruit of *Physalis angulata* and the HeLa cells were tested on MCF-7, DLD-1. All the cell lines were cultured with DMEM, L-glutamine, 10% FBS, sodium bicarbonate, and an antibiotic solution with penicillin, streptomycin, and amphotericin B with the concentration of 100 U/mL, 100 μ g/mL, and $2.5 \,\mu$ g/mL, respectively. All these cell lines were kept in a humidified incubator at 37 °C. An inverted phase-contrast microscope was used to evaluate the viability of cells and the MTT method of assay. The compound stock solution was prepared by mixing 1 mg of sample in 1 mL DMEM.

Cytotoxic Estimation

The growth medium was removed after 24 h. Each compound was freshly prepared and diluted five times. 100 μ L concentrations were added three times to the respective wells and incubated and humidified. Control cells were also maintained untreated.

4.9. Estimation of Anti-Bacterial Activity

According to the standard procedure, *Physalis angulata* leaves and fruit extracts were evaluated for their antibacterial activity [58]. Inoculum containing 106 CFU/mL of Gramnegative and Gram-positive bacterial culture such as Escherichia coli and Staphylococcus aureus (ATCC 25922 and ATCC 25923, respectively) were used for the evaluation. All these strains were isolated from Ras Al Khaimah Medical and Health Sciences University's clinical microbiology and immunology division. The diffusion method was used for the estimation. Wells (8 mm diameter) were holed into the Muller Hinton agar medium. These wells are then packed with *Physalis angulata* leaves and fruit extract (100 μ L–25 mg/mL). Plates were diffused for 2 h at room temperature. A 1% DMSO (5 mg/500 μ L) solution was used to prepare the stock solution of all the extracts of *Physalis angulata*. All their plates were kept vertically and incubated for twenty-four hours at 37 °C. Ethanol and gentamycin (10 μ g) were used as negative control and standard/positive control, respectively. The growth inhibition zones of each extract were calculated after the incubation period. The test was repeated against both pathogens. Mean \pm standard deviation was used to represent the data.

The broth microdilution method (BMM) was used to determine the MIC (minimum inhibitory concentration) and MBC (minimum bactericidal concentration). BMM is a method used to quantify the in-vitro activity of an antibacterial agent. For BMM, the requirements are a sterile tray with various concentrations of antimicrobial agents of interest tested against a standardized number of microbial cells [59]. MICs were determined by observing the lowest concentration of an antimicrobial agent that would inhibit the visible growth of a microorganism after overnight incubation. The lowest concentration of an antimicrobial that prevents the growth of an organism after subculture onto antibiotic-free media gives the value for MBC [50].

5. Conclusions

The present study discussed the cytotoxic activity of the ethanolic extracts of *Physalis angulata* leaves and fruit. The various extracts of the leaf and fruit of *Physalis angulata* showed significant antioxidant activity. The cytotoxic activity was evaluated by the MTT assay method against various cell lines such as HeLa (cervical), DLD-1 (colon), and MCF-

7 (breast). When comparing the results of the antioxidant activity of various extracts of *Physalis angulata* leaves and fruit, the ethanolic extract showed better activity than that of ascorbic acid. The TPC and TFC values were also high in ethanolic extracts of *Physalis angulata* compared with other extracts. When comparing the results of fruit and leaf extracts, the leaf extracts of *Physalis angulata* showed better antioxidant activity and TFC and TPC. The cytotoxic activity shown by the ethanolic extracts of *Physalis angulata* leaf was significantly better than that of fruit extracts of the plant. The ethanolic extract of the leaf exhibited very good anticancer activity against human HeLa cell lines through its low IC₅₀ (44 \pm 10 µg/mL) values. Thus, further investigations and isolation can be done for the leaf of *Physalis angulata*.

Supplementary Materials: Figure S1. The transparent flowchart for summarizing Section 4, Figure S2. Representative full scan chromatographic profile of ethanolic extract (leaves) of *Physalis angulata*, Figure S3. Representative full scan chromatographic profile of ethanolic extract (fruits) of *Physalis angulata*. Figure S4. Antibacterial activity of *Physalis angulata* fruit extracts. Plate A: petroleum ether, Plate B: ethyl acetate, and Plate C: ethanol extracts. Figure S5. Antibacterial activity of *Physalis angulata* leaves extracts. Plate A: petroleum ether, Plate B: ethyl acetate, and Plate C: ethanol extracts. Figure S5. Antibacterial activity of *Physalis angulata* leaves extracts. Plate A: petroleum ether, Plate B: ethyl acetate, and Plate C: ethanol extracts.

Author Contributions: Conceptualization, J.R.P. and A.F.W.; Data curation, M.U.R., G.A.M. and A.F.W.; formal analysis, G.A.M. and M.U.R.; funding acquisition, T.A.W. and J.R.P.; investigation, J.R.P., A.F.W., G.A.M., S.Z., A.A., T.A.W. and T.M.M.; methodology, T.A.W., A.A. and T.M.M.; writing—original draft, J.R.P., A.F.W. and M.U.R.; writing—review and editing, T.A.W., S.Z., A.A. and T.M.M. All authors have read and agreed to the published version of the manuscript.

Funding: The authors extend their appreciation to Researchers Supporting Project number (RSP-2021/357), King Saud University, Riyadh, Saudi Arabia, for funding this work.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors wish to thank RAK Medical and Health Sciences University, Ras Al Khaimah, United Arab Emirates, for their research support for the RAKMHSU-REC-124-2018-F-P research project. The authors extend their appreciation to Researchers Supporting Project number (RSP-2021/357), King Saud University, Riyadh, Saudi Arabia, for funding this work.

Conflicts of Interest: The authors declare that they have no conflicts of interest.

Sample Availability: Samples of the compounds are available from the authors.

References

- Sagbo, I.J.; Otang-Mbeng, W. Plants Used for the Traditional Management of Cancer in the Eastern Cape Province of South Africa: A Review of Ethnobotanical Surveys, Ethnopharmacological Studies and Active Phytochemicals. *Molecules* 2021, 26, 4639. [CrossRef]
- 2. Twilley, D.; Rademan, S.; Lall, N. A review on traditionally used South African medicinal plants, their secondary metabolites and their potential development into anticancer agents. *J. Ethnopharmacol.* **2020**, *261*, 113101. [CrossRef]
- Kausar, F.; Kim, K.-H.; Farooqi, H.M.U.; Farooqi, M.A.; Kaleem, M.; Waqar, R.; Khalil, A.A.K.; Khuda, F.; Abdul Rahim, C.S.; Hyun, K.; et al. Evaluation of Antimicrobial and Anticancer Activities of Selected Medicinal Plants of Himalayas, Pakistan. *Plants* 2022, 11, 48. [CrossRef] [PubMed]
- 4. Cumali, O.; Shahid, F.; Huseyin, O.; Selcuk, O.; Bekir, B.; Hikmet, G. Germination Biology of Two Invasive Physalis Species and Implications for Their Management in Arid and Semi-arid Regions. *Sci. Rep.* **2017**, *7*, 16960. [CrossRef]
- Rivera, D.; Ocampo, Y.; Franco, L.A. *Physalis angulata* Calyces Modulate Macrophage Polarization and Alleviate Chemically Induced Intestinal Inflammation in Mice. *Biomedicines* 2020, *8*, 24. [CrossRef] [PubMed]
- Domingues, L.A.; Quaglio, A.E.V.; de Almeida Costa, C.A.R.; Di Stasi, L.C. Intestinal anti-inflammatory activity of Ground Cherry (*Physalis angulata* L.) standardized CO₂ phytopharmaceutical preparation. *World J. Gastroenterol.* 2017, 23, 4369–4380.
- Medina-Medrano, J.R.; Almaraz-Abarca, N.; González-Elizondo, M.S.; Uribe-Soto, J.N.; González-Valdez, L.S.; Herrera-Arrieta, Y. Phenolic constituents and antioxidant properties of five wild species of *Physalis* (Solanaceae). *Bot. Stud.* 2015, *56*, 24. [CrossRef]

- Fan, Y.; Mao, Y.; Cao, S.; Xia, G.; Zhang, Q.; Zhang, H.; Qiu, F.; Kang, N. S5, a Withanolide Isolated from *Physalis Pubescens*, L., Induces G2/M Cell Cycle Arrest via the EGFR/P38 Pathway in Human Melanoma A375 Cells. *Molecules* 2018, 23, 3175. [CrossRef] [PubMed]
- 9. Khan, F.A.; Zahoor, M.; Ullah, N.; Khan, S.; Khurram, M.; Khan, S.; Ali, J. A general introduction to medicinal plants and silybum marianum. *Life Sci. J.* 2014, *11*, 471–481.
- Marsha, D.C.; Wulandari, L.R.; Sujuti, H. Pro-apoptotic and anti-proliferative effects of *Physalis angulata* leaf extract on retinoblastoma cells. *Int J. Ophthalmol.* 2019, 12, 1402–1407.
- 11. Jiangjie, L.; Min, X.; Jiahui, C.; Dongliang, Y.; Yijun, M.; Huizhong, W. Transcriptome-wide identification of microRNAs and functional insights inferred from microRNA—Target pairs in *Physalis angulata* L. *Plant Signal. Behav.* **2019**, *14*, e1629267. [CrossRef]
- 12. Qinghong, M.; Jiajia, F.; Zhiguo, L.; Xiwen, L.; Fangbo, Z.; Yanlin, Z.; Yi, S.; Li, L.; Liu, X.; Erbing, H. Cytotoxic Withanolides from the Whole Herb of *Physalis angulata* L. *Molecules* **2019**, *24*, 1608. [CrossRef]
- Shangguo, F.; Kaixin, Z.; Kaili, J.; Yuchen, C.; Chuanlan, C.; Yanyan, M.; Lingyan, W.; Xiaori, Z.; Qicai, Y.; Wang, H. Complete chloroplast genomes of four *Physalis* species (Solanaceae): Lights into genome structure, comparative analysis, and phylogenetic relationships. *BMC Plant Biol.* 2020, 20, 242. [CrossRef]
- 14. Cheng-Peng, S.; Chong-Yue, Q.; Feng, Z.; Ning, K.; Li-Xia, C.; Feng, Q. Physalins V-IX, 16,24-cyclo-13,14-seco withanolides from *Physalis angulata* and their antiproliferative and anti-inflammatory activities. *Sci. Rep.* **2017**, *7*, 4057. [CrossRef]
- 15. David, E.R.; Yanet, C.O.; Jenny, P.C.; Lía, B.; Fredyc, D.; Luis, A.F. A screening of plants used in Colombian traditional medicine revealed the anti-inflammatory potential of *Physalis angulata* calyces. *Saudi J. Biol. Sci.* **2019**, *26*, 1758–1766.
- Kindscher, K.; Long, Q.; Corbett, S.; Bosnak, K.; Loring, H.; Cohen, M.; Timmermann, B.N. The Ethnobotany and Ethnopharmacology of Wild Tomatillos, *Physalis longifolia* Nutt, and Relted Physalis Species: A Review. *Econ. Bot.* 2012, 66, 298–310. [CrossRef]
- 17. Jyothibasu, T.; Venkata, R.K.; Sreenu, T.; Subba, R.T. Anti-asthmatic activity of Alcoholic Extract of *Physalis angulata* induced by Ovalbumin. *Am. J. Pharm. Tech. Res.* **2012**, *2*, 892–897.
- 18. Mazumder, K.; Biswas, B.; Raja, I.M.; Fukase, K. Review of Cytotoxic Plants of the Indian Subcontinent and a Broad-Spectrum Analysis of Their Bioactive Compounds. *Molecules* 2020, 25, 1904. [CrossRef] [PubMed]
- 19. Mehrdad, I.; Ramin, R.; Mona, N.N.; Ali, H.; Jamal, K. Cytotoxic activity of the genus Ferula (*Apiaceae*) and its bioactive constituents. *AJP* **2018**, *8*, 296–312.
- Jayachithra, R.; Adil, F.W.; Fatima, E.M.; Aisha, O.E.; Aya, A. In vitro antioxidant activity and quantitative elemental analysis of Adansonia digitata L. fruit using inductively coupled plasma optical emission spectroscopy. Ann. Phytomed. 2019, 8, 127–133.
- 21. Lu, D.Y.; Lu, T.R.; Yarla, N.S.; Lu, Y.; Che, J.Y.; Ding, J.; Xu, B.; Zhu, H.; Shen, Y.; Wu, H.Y. Natural drug cancer treatments, strategies from herbal medicine to chemical or biological drugs. In *Studies in Natural Products Chemistry*; Chapter 4; Atta-ur-Rahman, Ed.; Elsevier: Amsterdam, The Netherlands, 2020; Volume 66, pp. 91–115. ISSN 1572-5995.
- 22. Esmail Al-Snafi, A. Phenolics and flavonoids contents of medicinal plants, as natural ingredients for many therapeutic purposes— A review. *IOSR J. Pharm.* 2020, *10*, 42–81.
- Yu, X.; Yang, T.; Qi, Q.; Du, Y.; Shi, J.; Liu, X.; Liu, Y.; Zhang, H.; Zhang, Z.; Yan, N. Comparison of the contents of phenolic compounds including flavonoids and antioxidant activity of rice (*Oryza sativa*) and Chinese wild rice (*Zizania latifolia*). *Food Chem.* 2021, 344, 128600. [CrossRef]
- 24. Patricia, C.; Rodríguez, A.B.; Javier, E.; María, G. Plant Phenolics: Bioavailability as a Key Determinant of Their Potential Health-Promoting Applications. *Antioxidants* **2020**, *9*, 1263. [CrossRef]
- Dimkić, I.; Petrović, M.; Gavrilović, M.; Gašić, U.; Ristivojević, P.; Stanković, S.; Janaćković, P. New perspectives of purple starthistle (Centaurea calcitrapa) leaf extracts: Phytochemical analysis, cytotoxicity and antimicrobial activity. *AMB Expr.* 2020, 10, 183. [CrossRef]
- Neupane, P.; Lamichhane, J. Estimation of total phenolic content, total flavonoid content and antioxidant capacities of five medicinal plants from Nepal. Vegetos 2020, 33, 360–366. [CrossRef]
- Tonisi, S.; Okaiyeto, K.; Hoppe, H.; Mabinya, L.V.; Nwodo, U.U.; Okoh, A.I. Chemical constituents, antioxidant and cytotoxicity properties of Leonotisleonurus used in the folklore management of neurological disorders in the Eastern Cape, South Africa. 3 *Biotech.* 2020, 10, 141. [CrossRef]
- Alara, O.R.; Abdurahman, N.H.; Olalere, O.A. Ethanolic extraction of flavonoids, phenolics and antioxidants from Vernonia amygdalina leaf using two-level factorial design. J. King Saud Univ. Sci. 2020, 32, 7–16. [CrossRef]
- 29. Harpreet, K.; Tanu, B. Role Of Flavonoids In Cancer Prevention: Chemistry And Mode Of Action. *Eur. J. Mol. Clin. Med.* **2020**, *7*, 3608–3625.
- 30. Lombrea, A.; Scurtu, A.D.; Avram, S.; Pavel, I.Z.; Turks, M.; Lugiņina, J.; Peipiņš, U.; Dehelean, C.A.; Soica, C.; Danciu, C. Anticancer Potential of Betulonic Acid Derivatives. *Int. J. Mol. Sci.* **2021**, *22*, 3676. [CrossRef]
- 31. Ohiagu, F.O.; Chikezie, P.C.; Chikezie, C.M.; Enyoh, C.E. Anticancer activity of Nigerian medicinal plants: A review. *Futur. J. Pharm. Sci.* **2021**, *7*, 70. [CrossRef]
- 32. Ketha, A.; Vedula, G.S.; Sastry, A.V.S. In vitro antioxidant, anti-inflammatory, and anticancer activities of methanolic extract and its metabolites of whole plant Cardiospermum canescens Wall. *Futur. J. Pharm. Sci.* **2020**, *6*, 11. [CrossRef]
- Lakshmi, M. Plant-Based Drugs as an Adjuvant to Cancer Chemotherapy. In *Alternative Medicine*; IntechOpen: London, UK, 2020. [CrossRef]

- 34. Lee, J.J.; Saiful Yazan, L.; Kassim, N.K.; Che Abdullah, C.A.; Esa, N.; Lim, P.C.; Tan, D.C. Cytotoxic Activity of Christia vespertilionis Root and Leaf Extracts and Fractions against Breast Cancer Cell Lines. *Molecules* **2020**, *25*, 2610. [CrossRef]
- 35. Fabiani, R. Antitumoral Properties of Natural Products. *Molecules* 2020, 25, 650. [CrossRef]
- 36. Sanger, G.; Rarung, L.K.; Wonggo, D.; Dotulong, V.; Damongilala, L.J.; Tallei, T.E. Cytotoxic activity of seaweeds from North Sulawesi marine waters against cervical cancer. *J. Appl. Pharm. Sci.* **2021**, *11*, 66–73.
- Aldakheel, R.K.; Rehman, S.; Almessiere, M.A.; Khan, F.A.; Gondal, M.A.; Mostafa, A.; Baykal, A. Bactericidal and In Vitro Cytotoxicity of Moringa oleifera Seed Extract and Its Elemental Analysis Using Laser-Induced Breakdown Spectroscopy. *Pharmaceuticals* 2020, 13, 193. [CrossRef]
- 38. Almessiere, M.A.; Altuwiriqi, R.; Gondal, M.A.; AlDakheel, R.K.; Alotaibi, H.F. Qualitative and quantitative analysis of human nails to find correlation between nutrients and vitamin D deficiency using LIBS and ICP-AES. *Talanta* **2018**, *185*, 61–70. [CrossRef]
- Álvarez-Martínez, F.J.; Barrajón-Catalán, E.; Encinar, J.A.; Rodríguez-Díaz, J.C.; Micol, V. Antimicrobial Capacity of Plant Polyphenols against Gram-positive Bacteria: A Comprehensive Review. *Curr. Med. Chem.* 2020, 27, 2576–2606. [CrossRef] [PubMed]
- 40. Ushie, O.A.; Neji, P.A.; Abeng, F.E.; Okpashi, V.E.; Baba, N.H.; Azuaga, T.I. Phytochemical Screening and Antimicrobial Activities of Chloroform and Ethyl Acetate Extracts of *Physalis angulata. J. Chem. Soc. Niger.* **2019**, *44*, 1062–1069.
- Fratianni, F.; Cozzolino, A.; De Feo, V.; Coppola, R.; Ombra, M.N.; Nazzaro, F. Polyphenols, Antioxidant, Antibacterial, and Biofilm Inhibitory Activities of Peel and Pulp of *Citrus medica* L., Citrus bergamia, and *Citrus medica* cv. Salò Cultivated in Southern Italy. *Molecules* 2019, 24, 4577. [CrossRef]
- Matsue, M.; Mori, Y.; Nagase, S.; Sugiyama, Y.; Hirano, R.; Ogai, K.; Ogura, K.; Kurihara, S.; Okamoto, S. Measuring the Antimicrobial Activity of Lauric Acid against Various Bacteria in Human Gut Microbiota Using a New Method. *Cell Transplant*. 2019, 28, 1528–1541. [CrossRef] [PubMed]
- 43. Dilika, F.; Bremner, P.D.; Meyer, J.J.M. Antibacterial activity of linoleic and oleic acids isolated from Helichrysum pedunculatum: A plant used during circumcision rites. *Fitoterapia* **2000**, *71*, 450–452. [CrossRef]
- Wagener, B.M.; Anjum, N.; Evans, C.; Brandon, A.; Honavar, J.; Creighton, J.; Traber, M.G.; Stuart, R.L.; Stevens, T.; Pittet, J.F. α-Tocopherol Attenuates the Severity of Pseudomonas aeruginosa-induced Pneumonia. *Am. J. Respir. Cell Mol. Biol.* 2020, 63, 234–243. [CrossRef] [PubMed]
- 45. Ghzaiel, I.; Zarrouk, A.; Nury, T.; Libergoli, M.; Florio, F.; Hammouda, S.; Ménétrier, F.; Avoscan, L.; Yammine, A.; Samadi, M.; et al. Antioxidant Properties and Cytoprotective Effect of *Pistacia lentiscus* L. Seed Oil against 7β-Hydroxycholesterol-Induced Toxicity in C2C12 Myoblasts: Reduction in Oxidative Stress, Mitochondrial and Peroxisomal Dysfunctions and Attenuation of Cell Death. *Antioxidants* 2021, 10, 1772. [CrossRef]
- Abd-ElGawad, A.M.; Elgamal, A.M.; El-Amier, Y.A.; Mohamed, T.A.; El Gendy, A.E.-N.G.; Elshamy, A.I. Chemical Composition, Allelopathic, Antioxidant, and Anti-Inflammatory Activities of Sesquiterpenes Rich Essential Oil of Cleome amblyocarpa Barratte & Murb. *Plants* 2021, 10, 1294. [CrossRef]
- 47. Tian, M.; Hong, Y.; Wu, X.; Zhang, M.; Lin, B.; Zhou, Y. Chemical constituents and cytotoxic activities of essential oils from the flowers, leaves and stems of Zingiber striolatum diels. *Rec. Nat. Prod.* **2020**, *14*, 144–149. [CrossRef]
- Srinivasan, R.; Mohankumar, R.; Kannappan, A.; Karthick Raja, V.; Archunan, G.; Karutha Pandian, S.; Ruckmani, K.; Veera Ravi, A. Exploring the Anti-quorum Sensing and Antibiofilm Efficacy of Phytol against Serratia marcescens Associated Acute Pyelonephritis Infection in Wistar Rats. *Front. Cell. Infect. Microbiol.* 2017, 7, 498. [CrossRef] [PubMed]
- 49. Abubakar, M.N.; Majinda, R.R.T. GC-MS Analysis and Preliminary Antimicrobial Activity of *Albizia adianthifolia* (Schumach) and *Pterocarpus angolensis* (DC). *Medicines* **2016**, *3*, 3. [CrossRef]
- Elmi, A.; Spina, R.; Risler, A.; Philippot, S.; Mérito, A.; Duval, R.E.; Abdoul-latif, F.M.; Laurain-Mattar, D. Evaluation of Antioxidant and Antibacterial Activities, Cytotoxicity of Acacia seyal Del Bark Extracts and Isolated Compounds. *Molecules* 2020, 25, 2392. [CrossRef]
- Wali, A.F.; Avula, B.; Ali, Z.; Khan, I.A.; Mushtaq, A.; Rehman, M.U.; Akbar, S.; Masoodi, M.H. Antioxidant, Hepatoprotective Potential and Chemical Profiling of Propolis Ethanolic Extract from Kashmir Himalaya Region Using UHPLC-DAD-QToF-MS. *BioMed Res. Int.* 2015, 2015, 393462. [CrossRef]
- Marchelak, A.; Owczarek, A.; Rutkowska, M.; Michel, P.; Kolodziejczyk-Czepas, J.; Nowak, P.; Olszewska, M.A. New insights into antioxidant activity of Prunus spinosa flowers: Extracts, model polyphenols and their phenolic metabolites in plasma towards multiple in vivo-relevant oxidants. *Phytochem. Lett.* 2019, *30*, 288–295. [CrossRef]
- Mas, A.J.; Heng, Y.K. Total Phenolic Content and Antioxidant and Antibacterial Activities of Pereskia bleo. *Adv. Pharmacol. Sci.* 2019, 2019, 7428593. [CrossRef]
- 54. Wali, A.F.; Mushtaq, A.; Rehman, M.U.; Akbar, S.; Masoodi, M.H. In vitro antioxidant and antimicrobial activities of propolis from Kashmir Himalaya region. *Free Radic. Antioxid.* **2016**, *6*, 51–57. [CrossRef]
- 55. Mishra, T.; Arya, R.K.; Meena, S.; Joshi, P.; Pal, M.; Meena, B.; Upreti, D.K.; Rana, T.S.; Datta, D. Isolation, Characterization and Anticancer Potential of Cytotoxic Triterpenes from Betula utilis Bark. *PLoS ONE* **2016**, *11*, e0159430. [CrossRef] [PubMed]
- 56. Fabio, F.; Luigi, G. Herbal Medicine Today: Clinical and Research Issues. *Evid. Based Complement. Alternat. Med.* **2007**, *4*, 37–40. [CrossRef]

- 57. Wali, A.F.; Al Dhaheri, Y.; Ramakrishna Pillai, J.; Mushtaq, A.; Rao, P.G.; Rabbani, S.A.; Firdous, A.; Elshikh, M.S.; Farraj, D.A.A. LC-MS Phytochemical Screening, In Vitro Antioxidant, Antimicrobial and Anticancer Activity of Microalgae Nannochloropsis oculata Extract. *Separations* **2017**, *7*, 54. [CrossRef]
- 58. Garcia, L. Broth Microdilution MIC Test. In *Clinical Microbiology Procedures Handbook*, 3rd ed.; Chapter 2; ASM Press: Washington, DC, USA, 2010; pp. 25–41. [CrossRef]
- 59. Dahiya, P.; Purkayastha, S. Phytochemical screening and antimicrobial activity of some medicinal plants against multi-drug resistant bacteria from clinical isolates. *Indian J. Pharm. Sci.* **2012**, *74*, 443–450.